

1 ***P. falciparum* gametocyte density and infectivity in peripheral blood and**
2 **skin tissue of naturally infected parasite carriers**

3

4 **Running title:** Gametocytes in the skin of malaria patients

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36 **ABSTRACT**

37 Transmission of *Plasmodium falciparum* depends on the presence of mature gametocytes that
38 can be ingested by mosquitoes taking a bloodmeal when feeding on human skin. It has long
39 been hypothesised that skin sequestration contributes to efficient transmission. Although skin
40 sequestration would have major implications for our understanding of transmission biology
41 and the suitability of mosquito feeding methodologies to measure the human infectious
42 reservoir, it has never been formally tested. In two populations of naturally infected
43 gametocyte carriers from Burkina Faso, we assessed transmission potential to mosquitoes and
44 directly quantified male and female gametocytes and asexual parasites in: i) finger prick
45 blood, ii) venous blood, iii) skin biopsies, and in pools of mosquitoes that fed iv) on venous
46 blood or, v) directly on the skin. Whilst more mosquitoes became infected when feeding
47 directly on the skin compared to venous blood, concentrations of gametocytes in the
48 subdermal skin vasculature were identical to that in other blood compartments. Asexual
49 parasite densities, gametocyte densities and sex ratios were identical in the mosquito blood
50 meals taken directly from the skin of parasite carriers and their venous blood.
51 We also observed sparse gametocytes in skin biopsies from legs and arms of gametocyte
52 carriers by microscopy. Taken together, we provide conclusive evidence for the absence of
53 significant skin sequestration of *P. falciparum* gametocytes. Gametocyte densities in
54 peripheral blood are thus informative for predicting onward transmission potential to
55 mosquitoes. Quantifying this human malaria transmission potential is of pivotal importance
56 for the deployment and monitoring of malaria elimination initiatives.

57

58 **IMPORTANCE**

59 Our observations settle a long-standing question in the malaria field and close a major
60 knowledge gap in the parasite cycle. By deploying mosquito feeding experiments and stage-

61 specific molecular and immunofluorescence parasite detection methodologies in two
62 populations of naturally infected parasite carriers, we conclusively reject the hypothesis of
63 gametocyte skin sequestration. Our findings provide novel insights in parasite stage
64 composition in human blood compartments, mosquito bloodmeals and their implications for
65 transmission potential. We demonstrate that gametocyte levels in venous or finger prick blood
66 can be used to predict onward transmission potential to mosquitoes. Our findings thus pave
67 the way for methodologies to quantify the human infectious reservoir based on conventional
68 blood sampling approaches to support the deployment and monitoring of malaria elimination
69 efforts for maximum public health impact.

70

71

72 INTRODUCTION

73 Significant reductions in malaria burden in recent decades have stimulated malaria
74 elimination initiatives (1). It is widely accepted that malaria elimination with current tools is
75 unlikely for the majority of African settings (2). Therefore, novel interventions are needed and
76 approaches that specifically reduce malaria transmission may be of key importance (3).
77 Transmission of malaria depends on the presence of mature male and female gametocytes that
78 circulate in the bloodstream and may be ingested by mosquitoes from the subdermal
79 capillaries upon blood feeding. For *P. falciparum*, these circulating mature gametocytes are
80 the product of a prolonged developmental process that starts with commitment of asexual
81 parasites to the sexual pathway upon activation of AP2-G(4, 5). Developing gametocytes are
82 sequestered for 10-12 days, primarily to the bone marrow and spleen (6), until their release
83 into the blood circulation as mature gametocytes. Mosquitoes may become infected when
84 feeding and ingesting mature male and female gametocytes, even if their densities in the
85 peripheral blood are low (7). Interestingly, mosquito infections have been observed from
86 gametocyte donors whose low gametocyte density appears incompatible with transmission
87 (8). Mosquito infection rates are typically higher when mosquitoes feed directly on the skin of
88 gametocyte carriers, as compared to feeding on venous blood through an artificial membrane
89 (9, 10). In addition to a strategic adjustment of gametocyte sex-ratio to maximize transmission
90 success (7, 11, 12), gametocyte aggregation and sequestration may facilitate mosquito
91 infections from low gametocyte densities. Aggregation of gametocytes in blood meals has
92 been observed (13) and may increase the chance that both male and female gametocytes are
93 ingested. Gametocyte sequestration in the skin tissue may further increase transmission rates
94 and would parallel sequestration patterns for other human parasites. The importance of skin
95 sequestration for transmission to invertebrate vectors was recently demonstrated for skin-
96 dwelling *Trypanosoma brucei* (14), as was previously reported for *Onchocerca volvulus*,

97 different species of *Mansonella*, *Leishmania infantum* and *L. donovani*, where parasite burden
98 in the skin is the best predictor of infectiousness (15-18).

99 Indirect evidence for skin sequestration of mature gametocytes in the microvasculature
100 of the skin was first described following surveys conducted in the 1940s and 1950s in DR
101 Congo: gametocyte prevalence in a survey using skin scarification was 3-fold higher
102 compared to a survey 5 years earlier using finger prick blood (19). In a follow up study with
103 1243 paired samples, a more modest 13.4% increase in *P. falciparum* parasite prevalence and
104 15.6% increase in gametocyte prevalence was observed when blood and dermal fluids from
105 skin scarification were used for sample preparation instead of finger prick blood (20). The
106 hypothesized skin sequestration of intra-erythrocytic *P. falciparum* gametocytes may be
107 related to mechanical retention in cutaneous capillaries (21, 22), analogous to the reversible
108 rigidity that likely prevents immature gametocytes from entering circulation (23, 24).
109 Alternatively, sequestration may be related to gametocyte cytoadhesive properties (25)
110 mediated by parasite proteins that are present on the infected red blood cell (iRBC) surface,
111 analogous to adhesion of asexual *P. falciparum* parasites to receptors on human vascular
112 endothelial cells by *P. falciparum* erythrocyte membrane-1 (PfEMP1)(26).

113 Whilst sequestration of mature gametocytes in the skin of naturally infected
114 individuals remains speculative, it may play an important role in determining *Plasmodium*
115 transmission efficiency (8, 22). Here, we report on two independent studies in naturally
116 infected gametocyte carriers from Burkina Faso where we quantified mature *P. falciparum*
117 gametocytes in skin tissue, blood samples and mosquito blood meals in association with
118 onward transmission to *Anopheles* mosquitoes.

119

120 **RESULTS**

121 A total of 31 individuals aged 15-48 (median 29) participated in experiments with paired skin
122 feeding (27) and membrane feeding (28). The median number of dissected mosquitoes per
123 experiment was 35 (interquartile range (IQR) 33-37) for direct skin feeding and 73 (IQR 69-
124 82) for membrane feeding. Of 31 paired experiments, 18 (58.1%) direct skin feeding and 22
125 (71.0%) membrane feeding experiments resulted in at least one infected mosquito ($p=0.289$).
126 Total gametocyte density, quantified in venous blood by quantitative reverse transcriptase
127 PCR (qRT-PCR) targeting female-specific *Pfs25* mRNA and male-specific *Pfmget* mRNA
128 (29), was positively associated with the proportion of mosquitoes that became infected
129 following direct skin feeding (Spearman $\rho=0.415$, $p=0.0204$) or membrane feeding (Spearman
130 $\rho=0.596$, $p = 0.0004$) (Figure 1A). The proportion of infected mosquitoes was higher by direct
131 skin feeding as compared to membrane feeding assays (odds ratio 2.01; 95% CI 1.21 – 3.33, p
132 = 0.007), in line with previous studies (9, 10, 30). The median number of oocysts was 4 (IQR
133 2-7.5; maximum 38) for mosquitoes that became infected after feeding directly on the skin
134 and 2 (IQR 1-5; maximum 24) for mosquitoes that became infected after feeding on venous
135 blood through a membrane feeder.

136 To examine whether this higher infectivity in direct skin feeding assays was related to
137 higher ingested gametocyte densities, or to a higher gametocyte fraction in the blood meal, we
138 directly quantified gametocytes and asexual parasites in mosquito blood meals. The blood
139 content of individually fed mosquitoes was released into an RNA preservative 15 minutes
140 after starting the feeding; RNA was then extracted and quantified from pools of 4 mosquitoes.
141 We quantified asexual parasites by skeleton-binding protein 1 *sbp1* qRT-PCR (31) and
142 gametocytes (*Pfs25* and *Pfmget* qRT-PCR) in a median of 3 mosquito pools per participant,
143 each containing 4 individual mosquitoes, from skin-feeding (range=2-3) and 4 pools per
144 participant, each containing 4 individual mosquitoes, from membrane feeding (range=2-4).
145 We observed strong correlations between parasite quantities in pools of mosquitoes that fed

146 on skin or venous blood through artificial membranes for asexual ring-stage parasites
147 ($r=0.921$, $p<0.0001$), male ($r=0.790$, $p<0.0001$) and female gametocytes ($r=0.655$, $p=0.0001$)
148 (Figure 1B). We also expressed gametocytes as a fraction of the total parasite biomass. This
149 fraction ranged from very low ($<1\%$ gametocytes in an individual with 21,086 ring-stage
150 asexual parasites/ μL and 179 gametocytes/ μL) to 100% in 3 individuals without asexual
151 parasites detected by qRT-PCR (Figure 1C). We observed no tendency towards a higher
152 fraction of gametocytes in skin-fed mosquitoes or capillary blood compared to venous blood
153 (Figure 1D).

154 In a complementary study, 9 adult gametocyte carriers participated in skin biopsy
155 sampling. After a screening visit, participants were seen on 2 occasions spaced 4 days apart.
156 One participant came on day 5 for the return visit instead of day 4; one other participant
157 withdrew consent prior to the second visit. On each occasion, venous blood, finger prick
158 blood and 4 small skin biopsy punches were taken from the leg ($n=2$) and arm ($n=2$). Half of
159 these biopsies were used for RNA extraction; the other half for histological assessments. Male
160 and female gametocytes and ring-stage asexual parasites were quantified by qRT-PCR to
161 calculate the gametocyte fraction in finger prick blood (16 observations; 9 donors), venous
162 blood ($n=16$; 9 donors), as well as skin tissue from the arm ($n=13$; 7 donors) and leg ($n=12$; 8
163 donors). Gametocytes were detected in all tissue and all blood samples by qRT-PCR; asexual
164 parasites were detected in 17/25 tissue and in 30/32 blood samples. The gametocyte fraction
165 was highly variable between donors (and between time-points) whilst estimates from the
166 different compartments from the same donor and time-point showed strong correlation: the
167 gametocyte fraction in venous blood was strongly associated with that in finger prick blood
168 (Spearman $\rho = 0.947$, $p<0.0001$), arm skin tissue (Spearman $\rho = 0.928$, $p < 0.0001$) and leg
169 skin tissue (Spearman $\rho = 0.870$, $p=0.0002$) (Figure 2A). Parasite density estimates per
170 microliter of blood or tissue were generally lower in the skin tissue compared to blood

171 samples (Figure 2B) and not significantly different between venous or finger prick blood
172 ($p \geq 0.121$) or between leg skin tissue or arm skin tissue ($p \geq 0.116$). The same RNA aliquots
173 were also processed for analysis by Nanostring expression array, a highly sensitive probe-
174 based expression platform that we have optimized for use in *P. falciparum* (32, 33). Using a
175 previously defined stage-specific marker set for asexual rings and mature gametocytes (33,
176 34), there was no evidence for higher gametocyte transcripts in skin samples compared to
177 blood samples (Figure 2C). The two approaches to quantify gene expression also showed a
178 strong correlation for *sbp1* and *Pfs25* (Figure 2D).

179 To directly detect gametocytes in subcutaneous tissue, skin biopsy samples that were
180 stored in formalin were processed for imaging. Given the low densities of gametocytes
181 predicted based on the qRT-PCR quantification (estimated median of 55.0 gametocytes in arm
182 tissue samples (IQR 28.2-153.0) and 36.9 gametocytes in leg tissue samples (IQR 11.6-98.3);
183 we established a protocol to image 10 μ m sections by confocal microscopy, hence maximizing
184 the detectability of sparse gametocytes (Figure 3A). Skin sections were initially analysed by
185 haematoxylin and eosin staining and labelled with the endothelial marker CD31 (Figures 3B)
186 to confirm integrity of the tissue. Evaluation of gametocyte markers identified Pfs16
187 antibodies (6, 35) as highly specific and sensitive using the confocal imaging protocol (Figure
188 3C), while antibodies against Pfs48/45 and Pfs230 were unable to detect gametocytes in
189 formalin fixed parasites and therefore not evaluated further. Screening of at least 12 sections
190 per skin snip in arm and leg samples from each participant identified several putative
191 gametocytes. A Pfs16 positive cell with a characteristic crescent shape, three-dimensional
192 structure and nuclear stain is shown in close association with a vessel (Figure 3D and
193 Supplementary movies 1 and 2). Based on these results, with low success gametocyte
194 detection rates by this highly sensitive fluorescence microscopy protocol, no further
195 gametocyte carriers were recruited as tissue donors.

196

197 **DISCUSSION**

198 Here, we tested a long-standing hypothesis of *P. falciparum* gametocyte sequestration in skin
199 tissue in two populations of naturally infected individuals in Burkina Faso. By combining
200 mosquito feeding assays and direct quantification of parasite populations in skin tissue,
201 mosquito blood meals and blood compartments, we conclude that there is no evidence for
202 significant skin sequestration of mature gametocytes.

203 Parasite sequestration in skin tissue is an intuitive explanation for how vector-borne
204 parasites can maximize the likelihood of update by blood-feeding insects. This phenomenon,
205 well demonstrated for a range of helminths (15-18) and protozoic trypanosomes (14), has
206 remained speculative for *Plasmodium* parasites (22). Two recent studies in Cameroonian
207 parasite carriers that used microscopy as diagnostic tool yielded conflicting results: one
208 observed higher *P. falciparum* parasite prevalence in finger prick capillary blood compared to
209 venous blood from hospital patients (36), the other found no differences for asexual parasites
210 or gametocytes in gametocyte carriers (37). The utility of finger prick blood to estimate
211 parasite biomass in skin tissue is uncertain. Studies published in the 1940s and 50s reported
212 superiority of skin scarification as compared to finger prick blood samples for parasite
213 detection (19, 20, 38). In the most extensive of these studies, in 1243 natural infections, 1 cm²
214 skin of the scapular region was very slightly scarified with 4-5 light incisions, expressing a
215 mixture of dermal fluids and capillary blood, with the first drop appearing richest in parasites
216 (20). This study demonstrated a 10-20% increase in prevalence of asexual parasites and
217 gametocytes of *P. vivax*, *P. malariae* and *P. falciparum* but not *P. ovale*. Also parasite
218 density, expressed as parasites per 15,000 examined white blood cells, appeared increased
219 (20). In the current study, we therefore not only collected venous blood and finger prick blood
220 but we also directly quantified parasite stage composition in skin tissue of naturally infected

221 donors and in blood meals of mosquitoes that naturally fed on the skin of the corresponding
222 donor. We used the absolute quantity of gametocytes and the fraction of the total parasite
223 biomass that is gametocyte as indicators of sequestration. In skin biopsy samples, we only
224 sporadically encountered gametocytes by histology. We chose a fluorescence imaging
225 protocol to image thick sections by confocal microscopy. This method allowed capturing of
226 entire parasites and three-dimensional reconstruction of parasite and surrounding tissues.
227 Using Pfs16 labelling we classified gametocytes by crescent shape, three-dimensional
228 structure (as opposed to non-specific speckles and autofluorescence, which is an inherent
229 issue of this approach), nuclear stain and presence of a surrounding red blood cell. The
230 frequency of immunofluorescence-detected gametocytes in our tissue samples was lower than
231 that by molecular methods in a tissue sample taken during the same visit. The quality of the
232 skin tissue, tested by analysing the tissue sections by haematoxylin and eosin staining, as well
233 as by labelling for endothelial cells, clearly indicates they were processed and preserved well.

234 In contrast, molecular detection of gametocytes was successful for all tissue samples
235 by qRT-PCR and for the majority of samples by Nanostring. Because the volume of blood is
236 unknown in tissue samples and specifically gametocytes are hypothesized to be enriched in
237 skin tissue (19, 20, 22), we compared the gametocyte fraction between different blood
238 compartments and found no evidence for a biased gametocyte fraction. Gametocyte
239 quantification in mosquito blood meals corroborated this finding and allowed a direct
240 comparison of parasite densities. Again, we observed no evidence for higher concentrations of
241 gametocytes in mosquitoes that fed directly on the skin of gametocyte donors compared to
242 venous blood and observed a very strong association between gametocyte fractions from the
243 different blood compartments. There must therefore be an alternative explanation for the
244 higher infection rates that we, in line with other studies (9, 10), observed in direct skin feeding
245 experiments compared to membrane feeding experiments using venous blood. Gametocyte

246 activation may occur following phlebotomy and may reduce infection rates observed
247 following membrane feeding. In addition, anticoagulants used in phlebotomy can have a
248 pronounced effect on mosquito infection rates (39). Although heparin is the preferred
249 anticoagulant (39), it may still have a disadvantageous impact on sporogonic development. In
250 malaria-naïve individuals in whom *P. falciparum* gametocytes were induced during controlled
251 human malaria infection studies, replacement of heparin plasma by serum resulted in
252 increased mosquito infection rates (10). Since human immune responses are unlikely to be of
253 relevance in these gametocytaemic volunteers, this observation provides additional indirect
254 evidence for a transmission modulatory effect of heparin.

255 We conclude that there is no evidence for gametocyte sequestration in skin tissue. Our
256 findings argue against a long-standing hypothesis that never had a solid evidence base or
257 proposed mechanism. Since the deformability of erythrocytes infected with mature
258 gametocytes is similar to that of uninfected erythrocytes (23, 40) and there is no evidence for
259 antigens on the surface of mature gametocyte-infected erythrocytes (41, 42), it is perhaps
260 unsurprising that gametocyte concentrations are similar in the different blood compartments.
261 While direct skin-feeding assays tend to result in higher infectivity compared that observed in
262 indirect feeding procedures using venous blood, our data demonstrate that any differences
263 observed are based on technical rather than biological differences in the feeding procedure.
264 Our findings also indicate that gametocyte levels in venous or finger prick blood can be used
265 to predict onward transmission potential to mosquitoes. Our findings thus pave the way for
266 methodologies to quantify the human infectious reservoir based on conventional blood
267 sampling approaches to support the deployment and monitoring of malaria elimination efforts
268 for maximum public health impact.

269

270 **MATERIALS AND METHODS**

271 **Ethics statement**

272 Ethical approval for the studies was granted by the Ethical Review Committee of the Ministry
273 of Health of Burkina Faso (Deliberation numbers 2016-03-033 and 2017-02-018) and the
274 Ethics Committee of the London School of Hygiene and Tropical Medicine (#10489 and
275 #11962). Individual written informed consent was obtained from each participant prior to
276 enrolment. Malaria cases were treated according to the National guidelines in Burkina Faso
277 (43).

278

279 **Study site and population**

280 Study participants were recruited in the village of Balonghin, located in Saponé district, in
281 Burkina Faso. Malaria transmission is seasonal and intense. The main malaria vectors are
282 *Anophele gambiae s.s.*, *An. coluzzii*, *An. arabiensis* and *An. funestus*. *P. falciparum* parasite
283 carriage and gametocyte carriage by molecular methods in the study area are 51-84% and 49-
284 75%, respectively (44).

285

286 **Study design**

287 **Paired skin feeding and membrane feeding study**

288 This study was conducted in October-December 2017. Individuals from the eligible age range
289 (15-50 years) in the study area were invited to study information meetings based on a village
290 census list and, if expressing an interest to participate, invited for screening at Balonghin
291 health facility. Eligible participants had *P. falciparum* gametocyte densities ≥ 1
292 gametocyte/500 leucocytes by microscopy (≥ 16 gametocytes/ μL when assuming 8000
293 leucocytes/ μL). Exclusion criteria were: signs of acute or chronic disease that required
294 immediate clinical care; haemoglobin concentration < 8 g/dL; current or previous participation
295 in malaria vaccine trials; recent blood transfusion or administration of blood products; use of

296 antimalarials in the last 2 weeks; co-infection with *P. malariae* or *P. ovale*. Eligible
297 participants were provided transport to the Centre National de Recherche et de Formation sur
298 le Paludisme (CNRFP) in Ouagadougou for membrane feeding and skin feeding. Immediately
299 after venipuncture in lithium heparin and EDTA tubes (BD Vacutainer™), 400-500µL of
300 heparinized blood in duplicate (for infectivity) and 400-500µL EDTA blood (for gametocyte
301 quantification in blood meals) was offered to 60 starved 4–5-day-old female *An. coluzzii*
302 mosquitoes via an artificial membrane attached to a water-jacketed glass feeder maintained at
303 37°C (28). After exactly 15 minutes of feeding in the dark, fully fed mosquitoes from heparin
304 blood were transferred to storage cups by aspiration and maintained with glucose solution at
305 27-29°C for 6-8 days before dissection with 1% mercurochrome staining and examination for
306 oocysts by two independent microscopists. From mosquitoes that fed on EDTA blood, 16
307 fully fed mosquitoes were sacrificed after feeding for exactly 15 minutes by sharp needle
308 puncture of their midguts to release the blood contents into 50µl of RNAprotect cell reagent;
309 blood meal material was stored for individual mosquitoes at -80°C. Immediately following
310 membrane feeding, direct skin feeding took place. The participant's calves were exposed to
311 60 mosquitoes distributed over 2 paper cups that were allowed to feed for exactly 15 minutes.
312 From this group, 12 fully fed mosquitoes were immediately sacrificed and their midguts
313 punctured as described above. Remaining mosquitoes were maintained on glucose solution
314 before dissections for oocyst presence, as above. In addition to the membrane and direct skin
315 feeding assays, K2EDTA blood was collected by venipuncture (BD Vacutainer™) and finger
316 prick (BD Microtainer®).

317

318 **Skin biopsy study**

319 In the period September 2016-March 2017, adults (aged 18-50 years) were invited for study
320 participation as described above. Participants were eligible if they had *P. falciparum*

321 gametocyte densities ≥ 1 gametocyte/500 leucocytes by microscopy (≥ 16 gametocytes/ μL).
322 For skin biopsy sampling, exclusion criteria were signs of acute or chronic disease that
323 requires immediate clinical care; haemoglobin concentration < 11 g/dL; skin infections or
324 conditions; history of vasovagal responses to blood sampling or biopsies; allergy to lidocaine/
325 prilocaine. Eligible individuals were invited to the CNRFP central lab in Ouagadougou on
326 two occasions, 4 days apart for sample collection. At each occasion, skin biopsy samples
327 including the dermis and hypodermis were taken from under the arm ($n=2$) and leg ($n=2$)
328 using single use punchers (4mm Biopsy Punch; Miltex Inc. York, US). This procedure was
329 performed 1 hour after applying local anaesthetic by means of a xylocaine-adrenaline by a
330 qualified dermatologist. Half of the biopsy samples (one each from arm and leg) were
331 immediately immersed in 2 mL of 10% formalin and placed at 4°C overnight; following
332 washing, samples were stored in 2 mL of 70% ethanol and stored at 4°C until further
333 processing. Other biopsy samples were transferred to 1000 μL RNALater stabilization
334 reagent (Qiagen), incubated overnight at $2-8^{\circ}\text{C}$ and then transferred to -80°C . Finger prick
335 and venous blood samples were collected in EDTA-coated tubes, as above.

336

337 **Molecular analysis**

338 Mosquito homogenates were pooled (4 mosquitoes in a total of $200\mu\text{L}$ of RNAProtect per
339 pool) with 4 pools (16 mosquitoes) for membrane feeding experiments and 3 pools per skin
340 feeding experiment (12 mosquitoes). Mosquitoes where no blood was released into
341 RNAProtect (upon visual expectation upon thawing) were not used for extraction and, as a
342 result, fewer pools of mosquitoes were extracted. Nucleic acids from these $200\mu\text{L}$ mosquito
343 pools and from $100\mu\text{L}$ venous and finger prick whole blood samples in RNAProtect Cell
344 Reagent were isolated using the bead-based MagNAPure LC automatic extractor (Total
345 Nucleic Acid Isolation Kit—High Performance, Roche Applied Science) and eluted in $50\mu\text{L}$

346 of water. In these samples, ring-stage asexual parasites, female gametocytes and male
347 gametocytes were quantified by individual quantitative reverse-transcription PCR (qRT-PCR)
348 assays targeting *sbp1* (31); *Pfs25* (45) and *PfMGET* (29), respectively. Skin biopsy samples
349 were immediately stored in RNAlater solution after collection. RNA extraction from skin
350 tissue was performed using the Qiagen RNeasy Plus Mini kit (Qiagen). First, the tissue
351 samples were removed from RNAlater solution and then homogenized in RLT lysis buffer
352 (Qiagen) using Polytron Homogenizer (Kinematica). The homogenized lysate was passed
353 through genomic DNA eliminator columns (Qiagen) and subsequently applied to RNeasy spin
354 columns. Following several washes, RNA was eluted in nuclease-free water according to the
355 manufacturer's instructions.

356 The NanoString nCounter custom code set included differentially expressed genes to
357 distinguish specific *P. falciparum* parasite stages as defined from our previous study (34). A
358 total of 456 parasite genes were included in the custom probe set including housekeeping
359 genes. 161 genes representing asexual circulating stages, 147 genes representing asexual
360 sequestering stages, 26 genes representing gametocyte rings, 27 immature gametocytes and 29
361 mature gametocyte genes. The remaining set was not annotated for any of these parasite
362 stages. For NanoString analysis, 5 µl of purified total RNA was used for initial hybridization
363 reaction. RNA from each sample was allowed to hybridize with reporter and capture probes at
364 65°C for 20 hours according to the nCounter gene expression assay protocol (NanoString
365 Technologies). RNA-probe complexes were immobilized to nCounter cartridge followed by
366 scanning in the nCounter Digital Analyzer. Data was first normalized by applying background
367 subtraction and then normalized to expression of housekeeping genes using the R package
368 "NanoStringNorm". The dataset was then quantile normalized using the R package
369 "aroma.light" and rank scaled. Mature gametocyte and asexual marker genes, as defined in³³,
370 were then averaged per patient, per tissue and per visit.

371

372 **Histological analysis of skin samples**

373 Skin biopsies were processed by passing through an increasing alcohol gradient and xylene
374 before embedded in paraffin wax. 10 μ m sections of biopsy samples were cut on a microtome
375 and placed on adhesion slides (SuperFrost® Plus Gold, VWR). Slides were dried at room
376 temperature for at least one hour then baked overnight at 42°C. The slides were allowed to
377 reach room temperature before proceeding with the staining protocol. Slides were incubated at
378 60°C to melt the wax around the section; sections were cleared with xylene and rehydrated by
379 passing through a decreasing alcohol gradient (xylene: 5 minutes twice; 100% ethanol: 3
380 minutes, twice; 90% ethanol: 3 minutes, twice; 70% ethanol: 3 minutes, twice). After
381 incubation in distilled water for 3 minutes, heat induced antigen retrieval was performed using
382 citrate buffer pH 6.0 (TCS Biosciences) in a table top autoclave. Slides were immersed in
383 buffer using a metal rack in an empty tip box (without lid) and autoclave initiated until it
384 reached 126°C, at which point the autoclave was unplugged and slides allowed to incubate in
385 the autoclave for a further 10 minutes. Subsequently, the slides were removed and cooled in
386 their buffer in a running water bath. Once at room temperature, slides were transferred to
387 distilled water and then TBST (Tris Buffered Saline with 0.05% Tween 20) for 3 minutes
388 each. Slides were then blocked with goat block containing 2.5% normal goat serum (Vector
389 Laboratories) complemented with 2.5% normal human serum (ThermoFisher Scientific). All
390 blocking and staining were performed in a humidified chamber. All staining solutions were
391 removed by tapping the side of the slide gently on tissue paper. Excess liquid was removed by
392 wicking away with tissue paper, being careful not to touch the sections. This was done to
393 maintain intact, well-formed skin sections which are particularly delicate. After 30-60 minutes
394 blocking at room temperature, the slides were incubated in primary antibodies diluted in goat
395 block. Sections were stained with 1:20 (1.12 μ g/ml) mouse anti-CD31 (Cell Marque: clone

396 JC70) at 4°C overnight or 1:1250 (1.04µg/ml) rabbit anti-Pfs16 (6) at room temperature for
397 one hour. The slides were then washed with TBST for 3 minutes thrice before adding 1:100
398 goat anti-mouse IgG-AlexaFluor488 (ThermoFisher, A-11029) or 1:250 goat anti-rabbit IgG-
399 AlexaFluor647 (ThermoFisher, A-21245) secondary antibody diluted in goat block and
400 incubated at room temperature for 30 minutes. Following secondary antibody staining, the
401 sections were washed twice with TBST and then once with TBS for 3 minutes each, before
402 incubation with 2.5nM final concentration of DAPI diluted in TBS for 10 mins at room
403 temperature. Sections were washed twice more in TBS for 3 minutes, before addition of
404 TrueView autofluorescence quenching reagent (Vector Laboratories) and incubation for 3
405 minutes at room temperature. Sections were washed once more in TBS for 5 minutes before
406 mounting with Vectashield Vibrance mountant (Vector Laboratories). Slides were viewed on
407 a Nikon A1R inverted confocal microscope with Piezo Z-drive to acquire z-stacks. In addition
408 to skin biopsies, clots of cultured *P. falciparum* parasites (strains Pf2004, 3D7 and NF54)
409 were generated to act as positive and negative controls. Asexual and mixed asexual-immature
410 gametocyte clots and mature gametocyte clots were generated as described previously (6).
411 Sections of formalin fixed paraffin embedded blocks were used to optimise Pfs16 antibody
412 and DAPI staining and determine the staining of mature gametocytes. Using these controls
413 gametocytes in the skin were determined by their circumferential staining with Pfs16 and
414 obvious outline of a red blood cell. Red blood cells were determined by their bright
415 autofluorescence under 488nm laser light. Images and movies were generated using Image J
416 software.

417

418 **Sample size justification**

419 For the paired skin feeding-membrane feeding study, we assumed an average of 15% infected
420 mosquitoes in patent gametocyte carriers with a standard deviation of 20% and a within

421 subject correlation of the outcome of 0.5 (9, 46, 47). If we then expected two-fold higher
422 mosquito infection rates in direct skin feeding, 17 paired membrane feeding and skin-feeding
423 experiments on patent gametocyte carriers would give 80% power to detect this difference at
424 an alpha of 0.05. Sample size justification for skin-biopsy sampling was based on a paired
425 comparison of the proportion of the total parasite population that is mature gametocyte. We
426 expected that 73% of the skin snip biopsy samples had higher gametocyte concentrations,
427 based on a meta-analysis that demonstrated enhanced infectivity following skin feeding
428 compared to venous blood membrane-feeding (9). When assuming that 70% of infected adults
429 have detectable malaria parasites in skin tissue and allow quantification of the proportion of
430 parasites that is gametocyte, and a lower limit of the 95%-CI >50%, 45 paired skin snip
431 samples and venous/finger prick blood samples would give 83% power with an alpha of 0.05
432 to detect a different in parasite stage composition. A go/no-go criterion was defined where an
433 initial 10 gametocyte carriers were recruited for biopsy samples and additional participants
434 would only be recruited if gametocytes were detected in $\geq 50\%$ of all samples.

435

436 **Statistical analysis**

437 All statistical analyses were performed in STATA version 15.0 (Statacorp; College Station,
438 TX, US). The proportion of infectious gametocyte carriers was compared between paired
439 feeding experiments using McNemar's test; the proportion of infected mosquitoes was
440 compared between direct skin feeding and membrane feeding using logistic regression
441 controlling for study participant as a fixed effect. Spearman non-parametric correlation
442 coefficients were calculated to assess associations between continuous variables; the paired
443 Wilcoxon rank-sum test was used to compare parasite densities between blood or tissue
444 samples from the same participants. The gametocyte fraction was calculated as the sum of

445 male and female gametocytes, expressed as a proportion of the total parasite biomass of
446 asexual ring-stage parasites and gametocytes.

447

448 **Supplemental data**

449 **Supplementary movie 1** (3D movie):

450 3D projection of Z-stack of mature gametocyte in skin snip. This movie shows the 3D
451 reconstruction of the z-stack (step-size 0.2 micron) to illustrate the localisation of a mature
452 gametocyte. The gametocyte is stained with Pfs16 (magenta), denoted by DAPI staining
453 (cyan), and within an RBC (yellow). It is in close proximity to skin vasculature. Movie was
454 generated using Image J software.

455

456 **Supplementary Movie 2** (Z stack):

457 Z-stack of mature gametocyte in skin snip. Confocal z-stack of mature gametocyte taken
458 across the whole thickness of the section (step-size 0.2 micron). Gametocyte stained with
459 Pfs16 (magenta), with DAPI (cyan) nuclear staining. Movie generated using Image J
460 software.

461

462 **Data availability**

463 Data underlying this manuscript are available through
464 https://datadryad.org/stash/share/_Di1z3S3jl2ahewKXHAXHfAtI7slSBGNAZmgueslqbl.

465

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474

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479

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656

657 **FIGURE LEGENDS**

658 **Figure 1. The density and infectivity of gametocytes in different blood compartments. A.**

659 Gametocyte density in venous blood in association with the proportion of mosquitoes that
660 become infected when feeding directly on the skin of the blood donor (blue) or on venous
661 blood offered through an artificial membrane feeder (red). Size indicates the number of
662 examined mosquitoes; error bars indicate the 95% confidence interval around the proportion
663 of infected mosquitoes. **B.** The density of ring stage asexual parasites (green), male
664 gametocytes (blue) and female gametocytes (red) in mosquito blood meals when feeding
665 directly on the skin (X-axis) versus venous blood offered through an artificial membrane

666 feeder (Y-axis). Error bars indicate the standard error of density estimates in pools of
667 mosquitoes fed directly on the skin (median 3 pools) or venous blood (median 4 pools). **C.**
668 The fraction of the total parasite biomass that is gametocyte in finger prick capillary blood
669 (red), mosquitoes that fed directly on the skin (green), mosquitoes that fed on venous blood
670 (blue) or venous blood (purple). The box plot indicates median, interquartile range and range;
671 dots indicate individual samples. **D.** The gametocyte fraction in venous blood (X-axis)
672 versus on the Y-axis finger prick capillary blood (red; Spearman $\rho=0.970$; $p<0.0001$),
673 mosquitoes that fed directly on the skin (green; Spearman $\rho=0.916$; $p<0.0001$), mosquitoes
674 that fed on venous blood (green; Spearman $\rho=0.912$; $p<0.0001$).

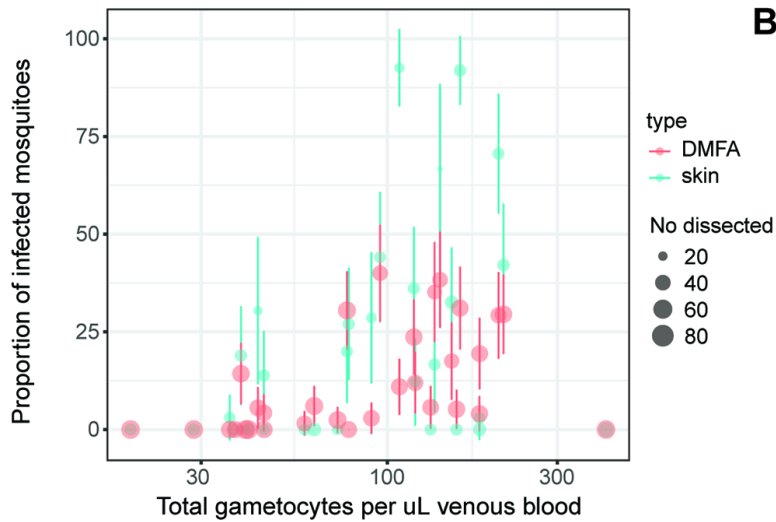
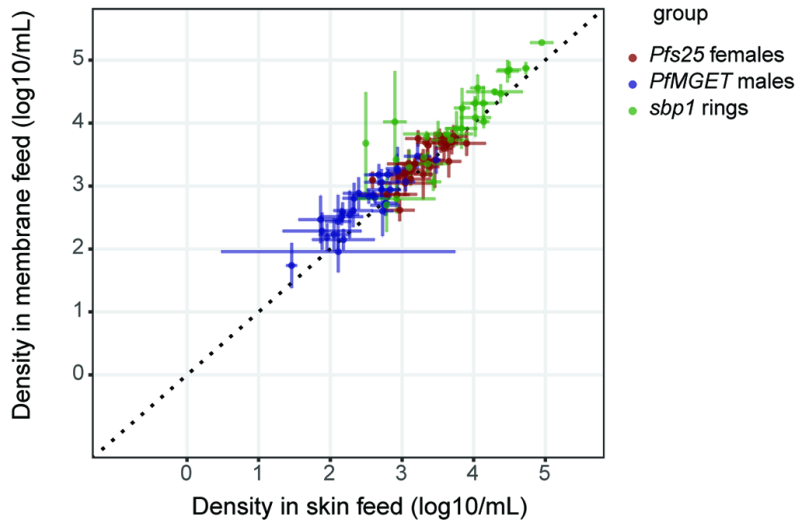
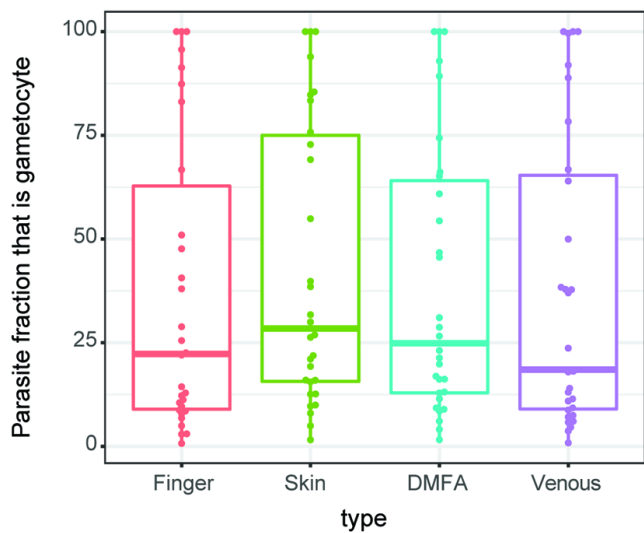
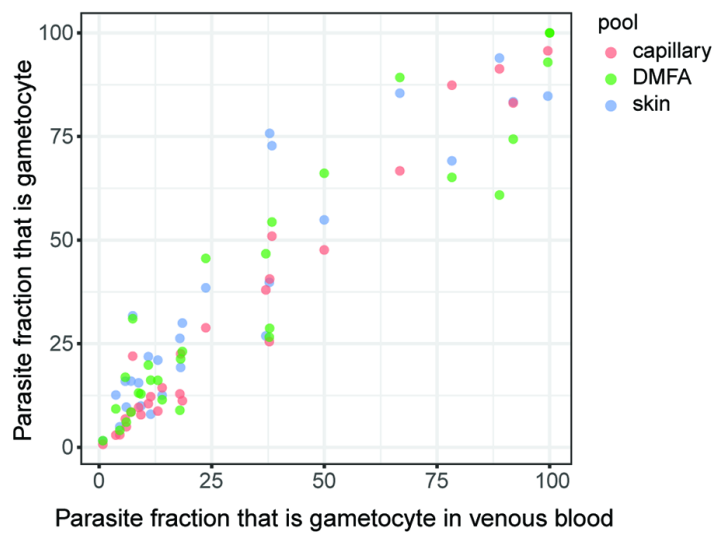
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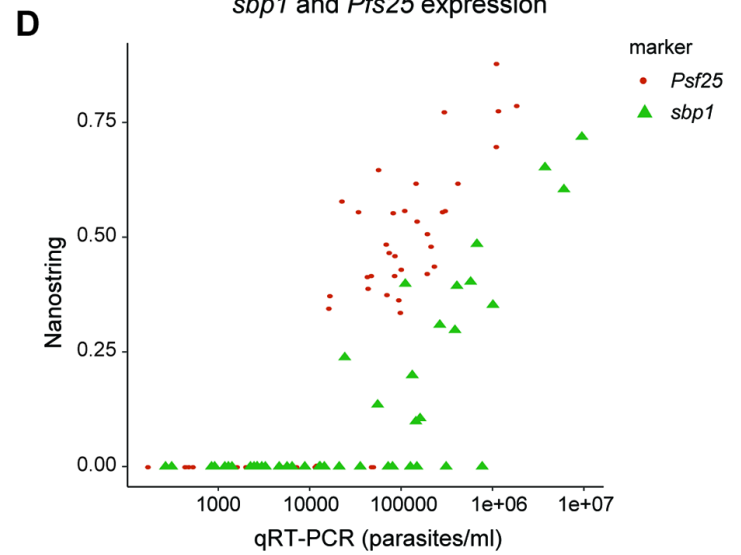
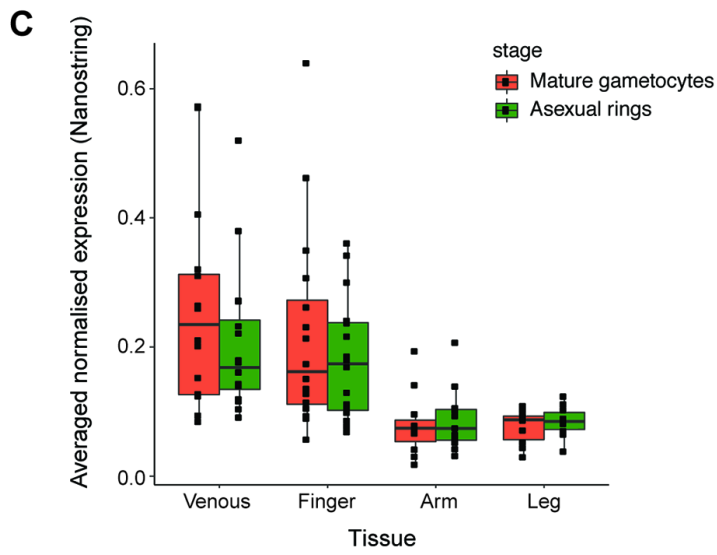
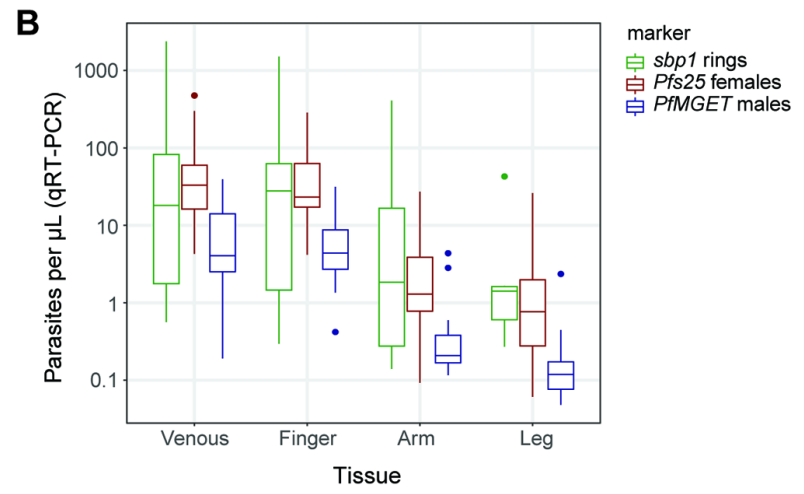
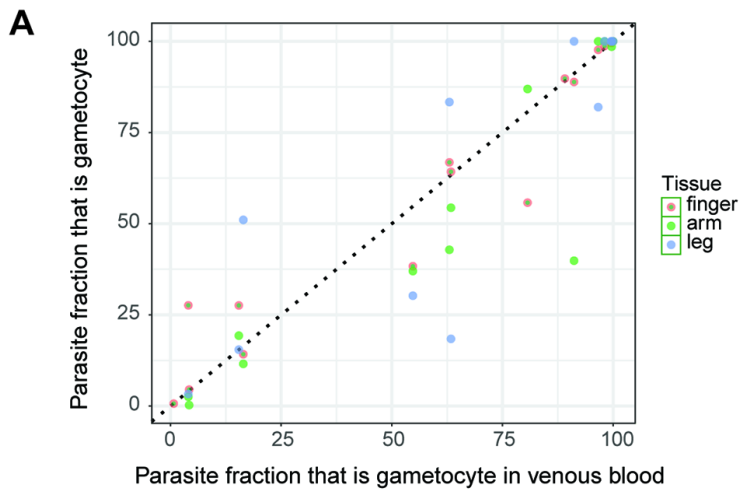
676 **Figure 2. qRT-PCR and Nanostring comparison of parasite densities in skin biopsy**
677 **samples and blood samples. A.** Gametocyte fractions (the proportion of gametocytes in the
678 total parasite biomass assessed by *sbp1*, *Pfs25* and *PfMGET* qRT-PCR) across compartments.
679 **B-D.** Relative numbers of asexual parasites and gametocytes in skin tissue from the arm, skin
680 tissue from the leg, finger prick and venous blood based on qRT-PCR (**B**) and Nanostring (**C**).
681 Nanostring data were normalized on the basis of background subtraction and expression of
682 housekeeping genes. **D.** Correlation between estimates of ring-stage asexual parasites by *sbp1*
683 and female gametocytes by *Pfs25* for qRT-PCR (X-axis) and Nanostring (Y-axis) showing
684 good agreement but higher sensitivity of qRT-PCR.

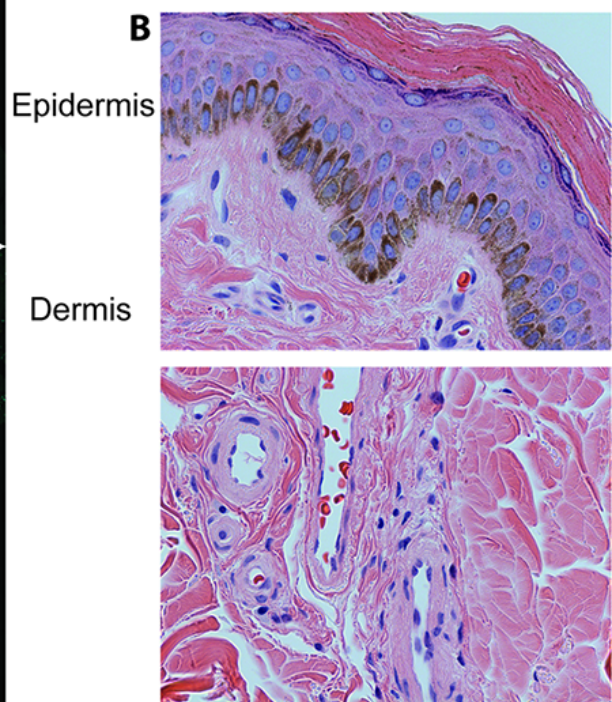
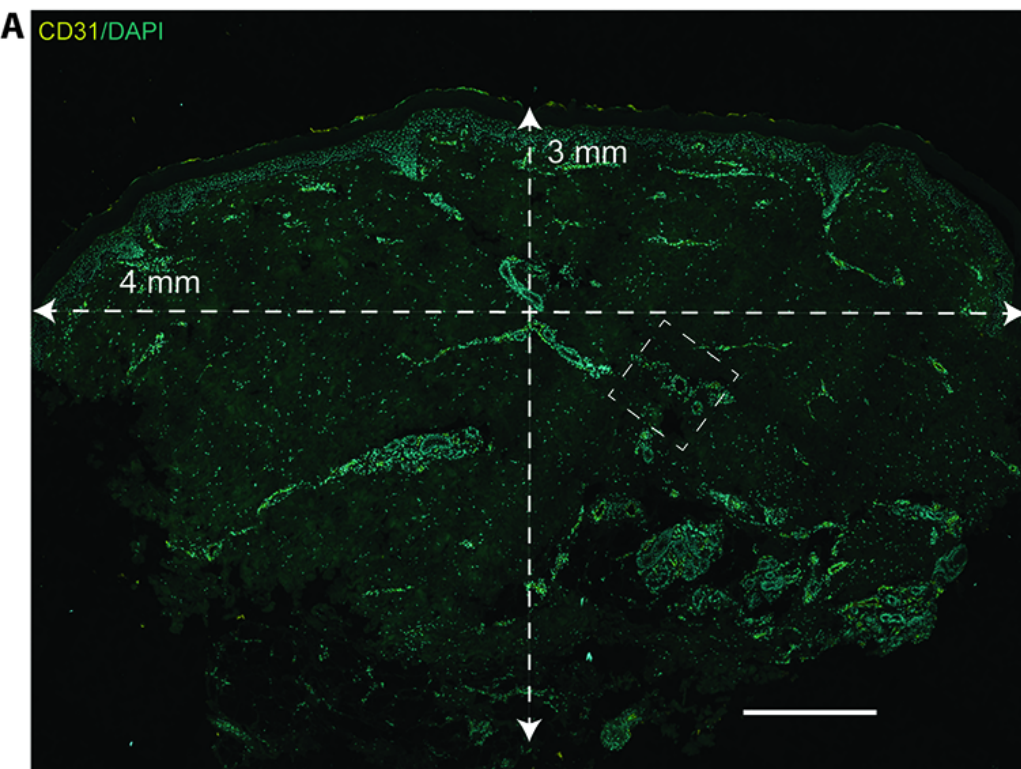
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686 **Figure 3. Histological analysis of skin samples. A.** 10 μ m cross section of a skin snip from
687 leg with dimensions indicated. Sample was stained with CD31 and DAPI and a maximum
688 projection across the depth of the section is shown. The insert represents a small section
689 including several vessels stained with CD31. Scale bar = 500 μ m, insert = 10 μ m. **B.** 3 μ m
690 section of a skin snip from arm stained with haematoxylin and eosin. Sections in A and B

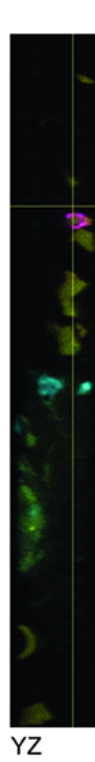
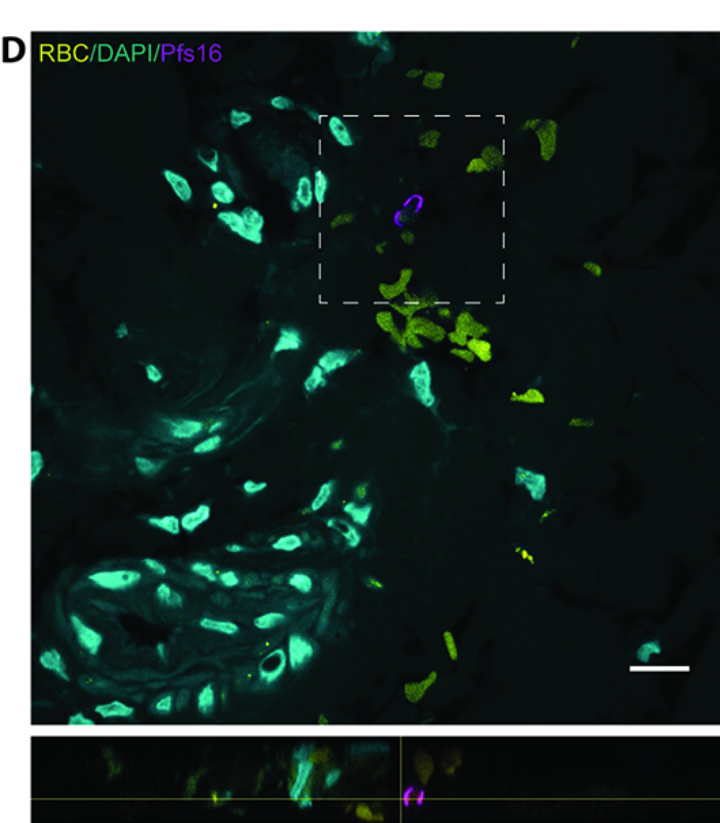
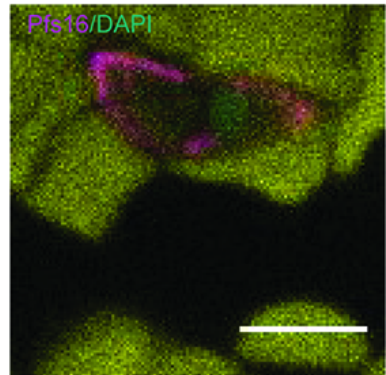
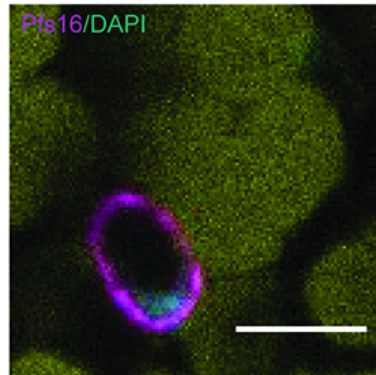
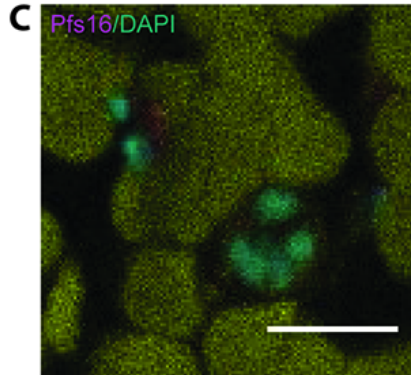
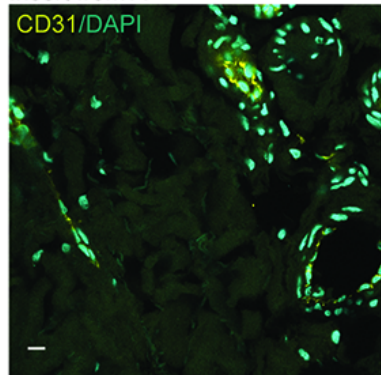
691 show the different layers of the epidermis on top, followed by the dermis with multiple
692 vessels. **C.** Samples were stained with DAPI (cyan) and Pfs16 (magenta) for gametocytes.
693 Representative images of asexual parasite (left), an immature (middle) and mature (right)
694 gametocyte images from control blood clots. Scale bar = 10 μ m. **D.** Representative image of a
695 gametocyte in skin samples from arm. DAPI staining indicates several vessels in the vicinity
696 of a gametocyte stained with Pfs16. XZ and YZ orientations are included to demonstrate the
697 three-dimensional nature of the tissue section and the gametocyte. Scale bar = 10 μ m.
698

A**B****C****D**

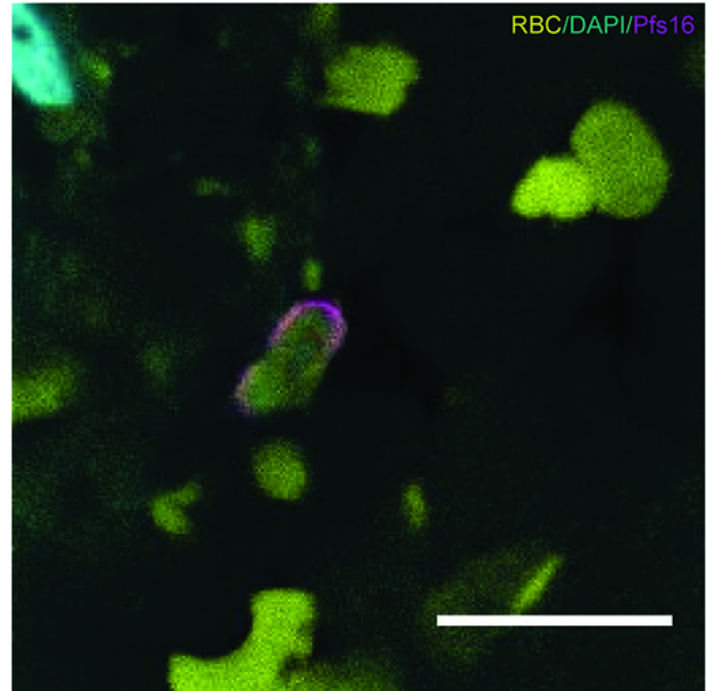




Insert from A



Insert from D



XZ