1	Xanthomonas sontii sp. nov., a non-pathogenic bacterium isolated from healthy basmati rice
2	(Oryza sativa) seeds from India.
3	Kanika Bansal ^{1,¶} , Amandeep Kaur ^{1,¶} , Samriti Midha ^{1,§} , Sanjeet Kumar ^{1,#} , Suresh Korpole ¹
4	and Prabhu B. Patil ^{1,*}
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6	¹ Bacterial Genomics and Evolution Laboratory, CSIR-Institute of Microbial Technology,
7	Chandigarh, India.
8	[§] Present address: Institute of Infection and Global Health, University of Liverpool, Liverpool, UK.
9	[#] Present address: Department of Archaeogenetics, Max Planck Institute for the Science of Human
10	History, Jena, Germany.
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12	[¶] Equal contribution
13	*Corresponding author
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17	Running title: Non-pathogenic Xanthomonas from rice plants.
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26	Data submission: Whole genome sequences of PPL1, PPL2 and PPL3 strains are submitted to
27	NCBI with accession numbers NQYO, NQYP, NMPO respectively.
28	Address correspondence to Prabhu B. Patil, pbpatil@imtech.res.in

29 Abstract

30 Three yellow pigmented, Gram negative, aerobic, rod shaped, motile bacterial strains designated as PPL1, PPL2 and PPL3 were isolated from healthy basmati rice seeds. Phenotypic, biochemical 31 and 16S rRNA gene sequence analysis assigned these strains to the genus Xanthomonas. The 16S 32 rRNA gene sequence was having 99.59% similarity with X. sacchari CFBP4641^T. However, 33 whole genome based phylogenomic analysis revealed that these strains formed a distinct 34 monophyletic clade with X. sacchari CFBP4641^T as their closest neighbour. Taxonogenomic 35 studies based on average nucleotide identity (orthoANI) and digital DNA-DNA hybridization 36 (dDDH) values of these strains with type strains (or representative strains) of different 37 Xanthomonas species including X. sacchari showed below recommended threshold values of ANI 38 (<96%) and dDDH (70%) for species delineation. Furthermore, at the whole genome level, PPL1 39 and PPL2 were found to be clonal, while PPL3 was not a clonal, but belonging to the same 40 species. Our in planta pathogenicity studies revealed that the strains PPL1, PPL2 and PPL3 are 41 non-pathogenic to rice plants. Hence, based on the present study, they form a novel lineage and 42 species associated with rice seeds for which the name Xanthomonas sontii sp. nov. is proposed. 43 The type strain for the X. sontii sp. nov. is $PPL1^{T}$ (CFBP8688^T = ICMP23426^T = MTCC12491^T) 44 and strains PPL2 (CFBP8689 = ICMP23427 = MTCC12492) and PPL3 (CFBP8690 = ICMP23428 45 = MTCC12492) as other strains of the species. 46

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Abbreviations: OrthoANI, Orthologous average nucleotide identity; dDDH, digtal DNA-DNA
hybridization; NA, nutrient agar; PSA, peptone sucrose agar; GYCA, glucose yeast extract
calcium carbonate agar; PBS, phosphate buffer saline; TSBA, tryptic soy agar; MCS, MiSeq
control software.

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- Keywords: Xanthomonas sontii, non-pathogenic, healthy basmati rice seeds, orthoANI, dDDH
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60 Introduction

Xanthomonas is a Gram-negative, yellow-pigmented plant associated bacterium that infects 61 62 economically important crops such as rice [1], pomegranate [2], citrus [3], pepper, cabbage [4], banana [5] etc. It is a complex genus comprising of 33 species (http://www.bacterio.net/), which 63 64 are further classified into 150 pathovars [6]. In 1997, Vauterin et al., divided these Xanthomonas species into three clusters based on 16S rRNA gene sequence phylogenetic analysis [7]. Besides 65 main core Xanthomonas cluster, X. albilineans, X. hyacinthi, X. theicola, and X. translucens 66 grouped as second cluster, whereas X. sacchari formed a distinct phylogenetic cluster. However, 67 advent of next generation sequencing technology and introduction of robust whole genome based 68 69 tools like orthologous average nucleotide identity (orthoANI) and digital DNA-DNA hybridisation 70 (dDDH) have revolutionized the field of bacterial taxonomy [8-10]. Infact, these methods are refining comprehensive taxonomic framework, which are essential for developing diagnostic 71 72 strategies and understanding host-pathogen relationships in management of crops [8].

Xanthomonas is emerging as a serious threat for economically important crops. X. oryzae pv. oryzae, X. campestris, X. axonopodis pv. manihotis were considered in the top 10 plant pathogenic bacteria [11]. Among these, X. oryzae causes bacterial blight disease to rice plants resulting in 30-50% decrease in rice yield every year [12], [13], [14]. Other than pathogenic Xanthomonas strains that cause disease in rice plants, some of the non-pathogenic Xanthomonas strains have also been identified from rice plants [15], [16]. Most of these non-pathogenic strains reported were largely characterized based on phenotypic and biochemical analysis providing limited information.

80 In the present study, we report isolation and characterization of three creamish-yellow pigmented 81 bacterial strains from healthy basmati rice (Oryza sativa) seeds. Genome based polyphasic analysis supported with pathogenicity tests revealed that these strains are non-pathogenic to rice 82 and belong to a novel species, for which we propose Xanthomonas sontii sp. nov. These non-83 pathogenic stains are widely over-looked due to their less economic importance. However, these 84 non-pathogenic isolates were isolated from rice plant, where their pathogenic counterparts (X. 85 oryzae) causes infection and are devastating worldwide. Hence, identification and detailed 86 analysis of these non-pathogenic strains can provide important insights into the lifestyle adapted 87 by these strains and virulence mechanisms of their pathogenic counterparts. 88

90 Materials and methods

91 Bacterial strain isolation and culture conditions

Strains were isolated from surface sterilized healthy rice seeds (Pusa basmati 1121 variety) that 92 were collected from Fazilka, Punjab, India (30.4036° N, 74.0280° E). For bacterial strain isolation, 93 surface sterilized seeds were partially crushed in 0.85 % NaCl (normal saline) using sterile mortar 94 95 and pestle. The mixture was then suspended in 50 ml of solution [17]. The solution was incubated for 2 h at 28°C and serial dilutions performed up to 10⁻⁶ and different dilutions (100µl) were 96 plated on media like nutrient agar (NA), peptone sucrose agar (PSA) and glucose yeast extract 97 calcium carbonate agar (GYCA). Plates were incubated at 28°C up to 6 days. Bacterial colonies 98 99 isolated were grown and maintained on PSA medium.

100 Phenotypic and biochemical characterisation

We analysed the morphology of strains by observing presence of flagella using transmission 101 electron microscopy. PPL1 strain was grown in nutrient broth and incubated at 28 °C for 20 h. 102 Subsequently, cells were harvested by centrifugation at 2000 rpm for 10 minutes. Cell pellet 103 104 was washed twice with phosphate buffer saline (1X PBS) and finally resuspended in PBS. Bacterial suspension was placed on carbon-coated copper grid (300 mesh, Nisshin EM Co., Ltd.) 105 106 for 15 minutes. The grid was then negatively stained for 30 seconds with 2% phosphotungstic 107 acid, dried and examined under JEM 2100 transmission electron microscope (JEOL, Tokyo, Japan) operating at 200 kV. 108

Further, biochemical characterization such as carbohydrate utilization, acid production and various
enzymatic activities were performed using OMNILOG GEN III system (BIOLOG) according to
manufacturer's instructions.

112 DNA extraction, genome sequencing, assembly and annotation

Genomic DNA extraction was carried out using ZR Fungal/Bacterial DNA MiniPrep kit (Zymo 113 Research, Irvine, CA, USA). Qualitative assessment of DNA was performed using NanoDrop 114 1000 (Thermo Fisher Scientific, Wilmington, DE, USA) and agarose gel electrophoresis. 115 Quantitative test was performed using Qubit 2.0 fluorometer (Life Technologies). Nextera XT 116 sample preparation kits (Illumina, Inc., San Diego, CA, USA) were used to prepare Illumina 117 paired-end sequencing libraries (250 x 2 read length) with dual indexing adapters. In-house 118 sequencing of the Illumina libraries was carried out on Illumina MiSeq platform (Illumina, Inc., 119 San Diego, CA, USA). Adapter trimming was performed automatically by MiSeq control software 120 121 (MCS), and remaining adapters were detected by NCBI server and were removed by manual

trimming. Sequencing reads were *de novo* assembled into high quality draft genome on CLC
Genomics Workbench v7.5 (CLC bio, Aarhus, Denmark) using default settings. Genome
annotation was performed by NCBI PGAP pipeline
(http://www.ncbi.nlm.nih.gov/genome/annotation_prok).

126 **Phylogenomic and taxonogenomic analysis:**

127 The 16S rRNA sequence of the strains was fetched from genome sequence and its comparision 128 with validly published reference bacteria was carried out using Eztaxon (https://www.ezbiocloud.net/). Core genome tree was constructed using PhyML[18]. Core genome 129 alignment was obtained using Roary v3.11.2 [19] with identity cutoff 60%. The core gene 130 alignment was converted into phylip format using SeaView v4.4.2-1 [20] and then, newick tree 131 132 was obtained using PhyML. Stenotrophomonas maltophilia ATCC13637 was used as an outgroup. 133 Taxonogenomic analysis of all type strains or representative strains of Xanthomonas was performed using OrthoANI v1.2 [21] values calculated by using USEARCH v5.2.32 [22] and 134 dDDH were calculated using Web tool GGDC 2.0 (http://ggdc.dsmz.de/distcalc2.php) 135

136 In planta pathogenicity test

PPL1, PPL2, PPL3 and BXO1 were grown to saturation in PSA media and inoculated on 30 days old rice plant (PUSA-basmati 1121) variety. Inoculation was performed by dipping scissors in bacterial culture and clipping tips of rice leaves. After 14 days, infection was assessed by measuring length of lesions on leaves. Here, BXO1 was positive control and PBS was used as negative control. Pathogenicity data of each isolate was obtained from 10 inoculated leaves based on two independent experiments.

143 **Results and discussion:**

144 Phenotypic and biochemical characterisation of PPL1, PPL2 and PPL3

All strains PPL1, PPL2 and PPL3 were isolated form glucose yeast extract calcium carbonate agar
(GYCA) media after 24 h of incubation at 28 °C. Colonies appeared as yellow, round, smooth,
convex and circular. All strains were Gram-negative, rod shaped bacteria with monopolar flagella,
as shown in figure 1.

- 149 Major biochemical characteristics of PPL1, PPL2 and PPL3 strains along with their closest
- 150 neighbour *X. sacchari* NCPPB 4341^T were determined using BIOLOG GEN III MICROPLATETM
- and compared with X. albiliniens LMG 494^{T} [23] (table 1). All three strains grew well between
- 152 20°C to $37 \square$ °C with optimum temperature 28 \square °C. No growth observed at 50 \square °C. Further,
- strains were able to grow at pH 6.0 and up to 4% NaCl whereas no growth observed at pH 5.0

154 and 8% NaCl. In BIOLOG results, all strains PPL1, PPL2 and PPL3 were positive for utilization of D-maltose, D-trehalose, D-cellobiose, gentiobiose, sucrose, D-turanose, α-D-lactose, D-155 melibiose, β -methyl-D-glucoside, D-salicin, N-acetyl-D-glucosamine, α -D-glucose, D-mannose, 156 D-fructose, D-galactose, L-fucose, glycerol, gelatin, L-alanine, L-aspartic Acid, L-glutamic Acid, 157 pectin, quinic acid, methyl pyruvate, L-lactic acid, citric acid, L-malic acid, tween 40, propionic 158 acid, acetic acid. Strains were resistant to rifamycin SV, lincomycin, vancomycin, tetrazolium 159 violet, tetrazolium blue, lithium chloride. Overall, biochemical characteristics of PPL1, PPL2 and 160 PPL3 strains were in accordance with its close relative X. sacchari NCPPB 4341^T. Biochemical 161 characteristics of X. albilineans LMG 494^T relative species of X. sacchari was taken from 162 literature and included in table 1 [23]. 163

164 **Pathogenicity test:**

Pathogenicity was checked by *in planta* inoculation studies. After 14 days of infection, BXO1 infected leaves showed approx. 13 cm lesion while, PPL1, PPL2, PPL3 and negative control did not show significant infection (approx. 0.5cm lesion) when compared with BXO1 (figure 2). Hence, this clearly reveals that PPL1, PPL2, PPL3 are non-pathogenic to the host.

169 In-house whole genome sequencing and assembly

Whole genome sequencing of PPL1, PPL2 and PPL3 strains were carried out using in-house
Illumina MiSeq platform. The genome size of all strains was approx. 5Mb with genome coverage
ranging from 78x to 109x. Assembly statistics for all the strains are given in table 2.

173 Phylogenomic analysis:

Core genome tree was constructed using PhyML. For analysis, total 36 species were used 174 (including 32 type and representative strains of different Xanthomonas species, along with three 175 strains PPL1, PPL2 and PPL3 isolated in this study). Stenotrophomonas maltophilia ATCC13637 176 was used as an outgroup. Out of 36 species, 27 were clubbed in one group as previously reported 177 [7] including X. pisi, X. vesicatoria, X. citri, X. codiaei, X. fragariae, X. bromi, X. campestris, X. 178 dyei, X. phaseoli, X. hortorum, X. arboricola, X. cynarae, X. cucurbitae, X. vasicola, X. 179 180 floridensis, X. perforans, X. euvesicatoria, X. maliensis, X. gardneri, X. axonopodis, X. cassavae, 181 X. nasturtii, X. alfalfae, X. prunicola, X. oryzae, X. melonis, and X. populi. Whereas 8 strains fallen in second group including PPL1, PPL2, PPL3, X. sacchari, X. theicola, X. translucens, X. 182 hyacinthi, X. albilineans. PPL1^T, PPL2 and PPL3 formed a monophyletic clade distinguishing 183

184 them from other strains. However, *X. sacchari* is the closest neighbour of these strains. Amongst

185 PPL strains, PPL1 and PPL2 are distinct from PPL3 with 100 bootstrap value (figure 3).

186 Genome based taxonogenomic status:

187 The orthoANI (figure 4) and dDDH values (table 3) of PPL1, PPL2 and PPL3 with type and representative strains of genus Xanthomonas species were below the cut-off for species 188 delineation. These strains have X. sacchari as 189 their closest relative with ANI values of ~ 94%, establishing novel species status of these strains. All the PPL1, PPL2 and PPL3 strains 190 showed dDDH values of around 55% with X. sacchari and less than 35% with other species of the 191 192 genus Xanthomonas. Interestingly, at the whole genome level, PPL1 and PPL2 strains were found to be clonal (ANI- 99.95%, dDDH- 99.5%) whereas, PPL3 was not clonal (ANI- 97.6%, dDDH-193 194 \sim 78%) however, it belonged to the same species.

195 **Description of** *Xanthomonas sontii* sp. nov.

Xanthomonas sontii (N.L. masc. gen. n. *sontii* named in honour of Ramesh V. Sonti, a renowned
Indian bacterial and plant molecular geneticist).

198 Cells are Gram-negative, aerobic, rod shape, motile and form yellow, round, smooth, convex and 199 circular colonies after 24hrs. Cells can grow on nutrient agar (NA), peptone sucrose agar (PSA) 200 and glucose yeast extract calcium carbonate agar (GYCA) media. Optimum temperature for growth is 28 °C. They are able to utilize of D-maltose, D-trehalose, D-cellobiose, gentiobiose, 201 202 sucrose, D-turanose, α -D-lactose, D-melibiose, β -methyl-D-glucoside, D-salicin, N-acetyl-D-203 glucosamine, α -D-glucose, D-mannose, D-fructose, D-galactose, L-fucose, glycerol, gelatin, Lalanine, L-aspartic acid, L-glutamic acid, pectin, quinic acid, methyl pyruvate, L-lactic acid, citric 204 acid, L-malic acid, , tween 40, propionic acid, acetic acid. Strains were able to grow at pH 6.0 and 205 resistant to 1% NaCl, 1% sodium lactate, and antibiotics like rifamycin SV, lincomycin, 206 207 vancomycin, tetrazolium violet, tetrazolium blue, lithium chloride. Taxonogenomic and 208 phylogenomic analysis revealed distinctness of these species with orthoANI and dDDH values 209 below established cutoff values i.e. 96% for ANI and 70% for dDDH. Core genome tree analysis 210 showed separate grouping PPL1, PPL2 and PPL3 from other Xanthomonas strains. Further, 211 amongst PPL strains PPL3 differ at clone level forming distinct clade than PPL1 and PPL2. 212 Therefore, we propose PPL1, PPL2 and PPL3 as novel species X. sontii of the genus Xanthomonas with PPL1 as type strain PPL1^T (CFBP8688^T = ICMP23426^T = MTCC12491^T). 213

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276	Figur	e legends
277	Figur	e 1: Transmission electron microscopy image of PPL1 ^T strain with monopolar flagella.
278	Figure	e 2: In planta infected leaves of rice (Pusa Basmati 1121) (a) leaves showing symptoms of

disease after 14 dpi (b) lesion length measured in cm for positive (BXO1), negative (PBS) controland PPL strains. Error bar indicates standard deviation of readings from 10 inoculated leaves and

- from two independent experiments.
- **Figure 3**: Whole genome based phylogeny considering all type strains and representative strains
- of genus *Xanthomonas*. The scale bar shows the number of nucleotide substitution per site. $PPL1^{T}$,

284 PPL2 and PPL3 strains (highlighted in coloured box) formed a distinct cluster. S. maltophilia

- ATCC1637 was used as an outgroup.
- **Figure 4**: Heat map showing ANI strains values of PPL1^T, PPL2 and PPL3 with type and
- 287 representative strains of the genus *Xanthomonas*.
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296 Tables

- **Table 1**: Comparison of biochemical characteristics of PPL1^T, PPL2, PPL3, and their closest
- 298 neighbor X. sacchari NCPPB 4341^T. Symbols represents; + : positive, : negative, +/- :
- borderline, s: X. albilineans LMG 494^T strain characteristics already reported in [23], NA : data
- 300 not available in literature.

	PPL1 ^T	PPL2	PPL3	X. sacchari NCPPB 4341 ^T	X. albilineans [§] LMG 494 ^T
Dextrin	+	+	+	+	-
D-Maltose	+	+	+	+	-
D-Trehalose	+	+	+	+	-
D-Cellobiose	+	+	+	+	+
Gentiobiose	+	+	+	+	-
Sucrose	+	+	+	+	+
D-Turanose	+	+	+	+	-
рН 6	+	+	+	+	NA
D-Raffinose	-	-	-	-	-
a-D-Lactose	+	+	+	+	-
D-Melibiose	+	+	+	+	-
β-Methyl-D-Glucoside	+	+	+	+	-
D-Salicin	+	+	+	+	NA
N-Acetyl-D-Glucosamine	+	+	+	+	+
N-Acetyl-β-D-Mannosamine	-	-	-	+/-	*
N-Acetyl-D-Galactosamine	+	+	+/-	+/-	-
N-Acetyl Neuraminic Acid	-	-	-	-	NA
1% NaCl	+	+	+	+	NA
4% NaCl	+	+	+	+	NA
8% NaCl	-	-	-	-	NA
a-D-Glucose	+	+	+	+	+
D-Mannose	+	+	+	+	+
D-Fructose	+	+	+	+	+
D-Galactose	+	+	+	+	-
L-Fucose	+	+	+	+	+
L-Rhamnose	-	-	-	+/-	-
Inosine	-	-	-	-	-
1% Sodium Lactate	+	+	-	+	NA
Fusidic Acid	-	-	-	-	NA
D-Sorbitol	-	-	-	-	-
D-Mannitol	-	-	-	-	-
D-Arabitol	-	-	-	-	-
Glycerol	+	+	+/-	+	-
D-Aspartic Acid	-	-	-	-	NA

Strain			PPL1 T	PPL2	PPL3
Rifamycin SV	+	+	+	+	NA
Minocycline	-	-	-	-	NA
Gelatin	+	+	+	+	NA
L-Alanine	+	+	+	+	-
L-Arginine	-	-	-	+/-	NA
L-Aspartic Acid	+	+	+	+	-
L-Glutamic Acid	+	+	+	+	+/-
L-Pyroglutamic acid	-	-	-	-	-
Lincomycin	+	+	+	+	NA
Pectin	+	+	+	+	NA
D-Gluconic Acid	-	-	-	-	-
Quinic Acid	+	+	+	+	-
D-Saccharic Acid	-	-	-	-	-
Vancomycin	+	+	+	+	NA
Tetrazolium Violet	+	+	+	+	NA
Tetrazolium blue	+	+	+	+	NA
p-Hydroxy Phenylacetic acid	-	-	-	+	-
Methyl Pyruvate	+	+	+/-	+	+
D-Lactic Acid Methyl Ester	-	-	-	-	NA
L-Lactic Acid	+	+	+	+	+
Citric Acid	+	+	+	+	-
L-Malic Acid	+	+	+	+	NA
Bromo-Succinic acid	+	+	+	+	-
Lithium Chloride	+	+	+	+	NA
Tween 40	+	+	+	+	-
Propionic Acid	+	+	+	+	-
Acetic Acid	+	+	+	+	-
Aztreonam	+/-	+	+	+	NA

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Table 2: Genome assembly statistics of PPL1^T, PPL2 and PPL3 strains.

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S.	Strain	Completeness/	Fold (Y)	N50	Contigs	Genome	GC	CDS	rRN	Isolation source	Accessio	
19.	Ivanie	Contamination	(A)	(KD)		Size	(70)		tRNA	source fi No.		
1	PPL1 ^T	96.31/0.22	78	32.9	332	4.8	69	4149	3+51	Rice seeds	NQYO	
2	PPL2	95.79/0.07	109	48.1	200	4.9	68.8	4105	2+50	Rice seeds	NQYP	
3	PPL3	98.56/0.55	80	40.3	231	4.8	68.8	4106	3+52	Rice seeds	NMPO	

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Table 3: The dDDH values of strains PPL1^T, PPL2 and PPL3 with other *Xanthomonas* strains.

32.4	32.4	32.5
33	33	33
33.6	33.6	33.4
28.5	28.4	28.4
55.1	55.3	55.2
78.2	78.7	100
100	99.5	78.2
99.5	100	78.7
23.1	23.1	23.1
23	23	23.1
23.2	23.2	23
23.4	23.4	23.3
23.4	23.4	23.4
23.6	23.6	23.7
23.4	23.4	23.4
22.9	22.8	22.8
23.1	23	23.3
22.9	22.9	23
22.5	22.4	22.4
23.6	23.5	23.4
22.7	22.6	22.4
22.9	22.8	22.8
23	23	22.9
23.1	23	22.9
23.1	23.1	23.1
23	23	23
22.9	22.9	22.9
23.1	23.1	23
23.3	23.3	23.3
23	23.1	23
23	23	23
23.6	23.6	23.6
23.4	23.4	23.3
22.8	22.8	22.9
22.9	22.8	23
22.9	22.9	22.9
	32.4 33 33.6 28.5 55.1 78.2 100 99.5 23.1 23 23.4 23.1 23 23.4 23.1 23.4 23.4 23.4 23.4 23.4 23.4 23.4 22.9 23.1 22.9 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.2 23.4 22.8 22.9 23.4 22.8 22.9 23.4 22.9 <tr tr=""></tr>	32.4 32.4 33 33 33.6 33.6 28.5 28.4 55.1 55.3 78.2 78.7 100 99.5 99.5 100 23.1 23.1 23.2 23.2 23.4 23.4 23.1 23.1 23.2 23.2 23.4 23.4 23.4 23.4 23.6 23.4 23.1 23 23.4 23.4 23.6 23.4 23.1 23 22.9 22.8 23.1 23 22.9 22.8 23.1 23 23.1 23 23.1 23 23.1 23 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.1 23.3 23.1 23.4 23.4 23.5 22.9 23.1 23.1

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(A)

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S.maltophilia ATCC13637 (T)	PPL1 (T)	PPL2	P L S		X.sacchari CFBP4641 (T)	X.hyacinthi CFBP1156	X.translucens DSM18974 (T)	X.theicola CFBP4691	X.albilineans CFBP2523 (T)	X.campestris ATCC33913 (T)	X.vesicatoria LMG 911 (T)	X.pisi DSM18256	X.dyei CFBP7245 (T)	X.codiaei CFBP4690	X.floridensis WHRI8848 (T)	X.cassavae CFBP4642 (T)	X.cucurbitae CFBP2542 (T)	X.melonis CFBP4644	X.fragariae PD885	X.arboricola CFBP2528 (T)	X.hortonum MO81	X.cynarae CFBP4188 (T)	X.gardneri ICMP7383	X.populi CFBP1817 (T)	X.citri LMG 9322 (T)	X.phaseoli CFBP412	X.euvesicatoria LMG 27970	X.perforans 91-118	X.alfalfae LMG 495 (T)	X.axonopodis DSM3585 (T)	X.bromi CFBP1976 (T)	X.vasicola NCPPB2717 (T)	X.oryzae ATCC35933 (T)	X.prunicola CFBP8353 (T)	X.nasturtii WHRI8853 (T)	X.maliensis LMG 27592 (T)	
100	79.17	7 79.	.21 79.	.04 7	79.05	79.07	79.22	78.89	76.68	77.76	77.06	77.6	77.19	78.1	77.33	77.69	77.59	77.65	76.69	77.63	77.05	77.24	77.53	77.14	77.16	77.25	77.37	77.32	77.34	77.28	77.19	76.71	77.11	77.25	77.63	77.16	S.maltophilia ATCC13637 (T)
79.17	100		.95 97	7.6 9	94.14	87.68	87.03	86.99	84.33	79.42	79.1	79.14	79.21	79.94	79.63	79.79	79.88	79.76	78.67	79.98	79.16	79.32	79.27	78.97	79.16	79.11	79.27	79.26	79.32	78.88	79.05	78.6	78.9	78.98	79.22	79.75	PPL1 (T)
79.21	99.9	5 10	97.	.66 9	94.23	87.55	87.02	86.8	84.49	79.48	79.15	79.41	79.17	80.11	79.76	79.67	79.56	79.79	78.63	79.97	79.27	79.39	79.33	79.02	79.13	79.3	79.28	79.34	79.21	79.14	79.14	78.65	78.9	79.02	79.4	79.65	PPL2
79.04	97.6	õ 97.	.66 10	900	94.14	87.6	87.14	87.05	84.43	79.35	79	79.47	79.22	80.15	79.65	79.76	79.76	79.7	78.55	79.85	79.32	79.39	79.28	79.03	79.1	79.15	79.33	79.43	79.35	79.06	79.15	78.56	78.89	78.87	79.32	79.56	PPL3
79.05	94.14	4 94.	.23 94	.14	100	87.65	87.21	86.84	84.16	79.34	78.96	79.31	79.18	79.84	79.58	79.69	79.56	79.91	78.64	79.95	79.17	79.13	79.21	78.77	79.06	79.26	79.19	79.27	79.42	79	78.92	78.72	78.69	78.89	79.41	79.61	X.sacchari CFBP4641 (T)
79.07	87.68	8 87.	.55 87	7.6 8	87.65	100	92.53	90.68	83.69	79.84	79.52	79.83	79.6	80.38	80.08	80.05	79.86	79.88	78.78	80.38	79.75	79.88	79.71	79.12	79.42	79.49	79.31	79.41	79.67	79.33	79.42	79.05	79.13	79.18	79.72	79.65	X.hyacinthi CFBP1156
79.22	87.03	3 87.	.02 87.	.14 8	87.21	92.53	100	90.13	83.61	79.82	79.69	79.75	79.72	80.41	79.98	80.05	79.79	80.19	78.81	80.38	79.74	79.8	79.74	79.45	79.4	79.56	79.7	79.69	79.63	79.56	79.59	79.24	79.42	79.44	79.77	79.71	X.translucens DSM18974 (T)
78.89	86.9	9 86	6.8 87.	.05 8	86.84	90.68	90.13	100	83.31	79.55	79.27	79.58	79.52	80.11	79.62	79.74	79.6	79.96	78.48	79.99	79.21	79.44	79.28	78.84	79.26	79.33	79.28	79.4	79.59	79.27	79.22	78.6	79.08	79.09	79.4	79.48	X.theicola CFBP4691
76.68	84.3	3 84.	.49 84	.43 8	84.16	83.69	83.61	83.31	100	77.41	77.22	77.63	77.51	77.71	77.48	77.53	76.98	77.79	76.78	77.7	77.05	77.43	77.28	76.73	77.23	77.22	77.09	77.26	77.5	77.04	77	76.94	77.29	76.9	77.18	77.48	X.albilineans CFBP2523 (T)
77.76	79.42	2 79.	.48 79.	.35 7	79.34	79.84	79.82	79.55	77.41	100	84.99	85.5	85.42	85.51	85.49	85.59	85.16	84.97	84.92	86.11	85.53	85.81	85.84	84.97	85.09	85.07	85.19	85.2	85.2	84.92	85.06	84.89	84.74	84.89	85.53	82.66	X.campestris ATCC 33913 (T)
77.06	79.1	79.	.15 79	97	78.96	79.52	79.69	79.27	77.22	84.99	100	90.29	90.61	87.18	87.9	87.56	86.56	86.02	85.64	86.73	86.51	86.18	86.25	85.62	86.22	85.94	86.18	86.07	86.17	85.74	86.78	85.91	85.79	86.18	87.36	82.37	X.vesicatoria LMG 911 (T)
77.6	79.14	4 79.	.41 79.	.47 7	79.31	79.83	79.75	79.58	77.63	85.5	90.29	100	91.34	87.39	88.96	87.61	86.94	86.47	86.01	87.78	87.09	86.78	86.97	86.19	86.47	86.35	86.46	86.5	86.54	86.17	86.85	86.11	86.13	87.58	87.38	82.56	X.pisi DSM18256
77.19	79.2 ⁻	1 79.	.17 79.	.22 7	79.18	79.6	79.72	79.52	77.51	85.42	90.61	91.34	100	87.13	88.05	87.36	86.5	86.17	85.96	86.92	86.68	86.6	86.74	85.98	86.27	86.21	86.22	86.18	86.25	85.85	86.81	86	85.93	86.49	87.41	82.4	X.dyei CFBP7245 (T)
78.1	79.94	4 80.	0.11 80	0.15 7	79.84	80.38	80.41	80.11	77.71	85.51	87.18	87.39	87.13	100	91.55	91.68	89.55	88.15	85.56	86.71	86.08	86.15	86.18	85.61	86.3	86.47	86.38	86.36	86.46	86.06	86.43	85.85	86.18	86.05	86.88	82.99	X.codiaei CFBP4690
77.33	79.63	3 79.	.76 79.	.65 7	79.58	80.08	79.98	79.62	77.48	85.49	87.9	88.96	88.05	91.55	100	93.2	89.54	87.9	85.59	87.14	86.45	86.29	86.4	85.76	86.49	86.53	86.57	86.45	86.52	86.15	86.84	85.97	86.05	86.39	87.16	83.02	X.floridensis WHR18848 (T)
77.69	79.79	9 79.	.67 79.	.76 7	79.69	80.05	80.05	79.74	77.53	85.59	87.56	87.61	87.36	91.68	93.2	100	89.5	87.76	85.68	87.54	86.46	86.49	86.56	85.78	86.62	86.72	86.84	86.89	87.03	86.41	86.81	86.18	86.24	86.39	87.21	82.85	X.cassavae CFBP4642 (T)
77.59	79.88	8 79.	.56 79.	.76 7	79.56	79.86	79.79	79.6	76.98	85.16	86.56	86.94	86.5	89.55	89.54	89.5	100	87.82	84.87	86.3	85.62	85.51	85.65	84.94	85.8	85.9	85.89	85.87	86.04	85.49	85.87	85.39	85.4	85.55	86.24	82.74	X.cucurbitae CFBP2542 (T)
77.65	79.76	6 79.	.79 79	9.7 7	79.91	79.88	80.19	79.96	77.79	84.97	86.02	86.47	86.17	88.15	87.9	87.76	87.82	100	84.88	86.23	85.5	85.48	85.54	84.89	85.76	85.76	85.93	85.81	85.89	85.46	85.7	85.39	85.45	85.52	86	82.67	X.melonis CFBP4644
76.69	78.6	7 78.	.63 78.	.55 7	78.64	78.78	78.81	78.48	76.78	84.92	85.64	86.01	85.96	85.56	85.59	85.68	84.87	84.88	100	87.4	87.37	87.37	87.47	87.12	85.92	85.87	85.88	85.97	86.05	85.72	86.09	85.95	85.93	85.89	86.67	82	X.fragariae PD885
77.63	79.98	8 79.	.97 79.	.85 7	79.95	80.38	80.38	79.99	77.7	86.11	86.73	87.78	86.92	86.71	87.14	87.54	86.3	86.23	87.4	100	89.49	89.53	89.5	88.52	87.06	86.94	87.02	87.03	86.98	86.58	86.9	86.76	86.64	87.1	87.68	83.23	X.arboricola CFBP2528 (T)
77.05	79.16	6 79.	.27 79.	.32 7	79.17	79.75	79.74	79.21	77.05	85.53	86.51	87.09	86.68	86.08	86.45	86.46	85.62	85.5	87.37	89.49	100	95.87	95.75	90.71	86.54	86.37	86.45	86.47	86.67	86.17	86.58	86.42	86.47	86.44	87.56	82.41	X.hortorum MO81
77.24	79.3	2 79.	.39 79.	.39 7	79.13	79.88	79.8	79.44	77.43	85.81	86.18	86.78	86.6	86.15	86.29	86.49	85.51	85.48	87.37	89.53	95.87	100	98.31	90.71	86.58	86.37	86.59	86.38	86.62	86.13	86.55	86.43	86.42	86.39	87.45	82.52	X.cynarae CFBP4188 (T)
77.53	79.2	7 79.	.33 79.	.28 7	79.21	79.71	79.74	79.28	77.28	85.84	86.25	86.97	86.74	86.18	86.4	86.56	85.65	85.54	87.47	89.5	95.75	98.31	100	90.78	86.59	86.4	86.63	86.54	86.62	86.19	86.57	86.36	86.33	86.58	87.45	82.49	X.gardneri ICMP7383
77.14 ^{bic}	DRxiv78.9	d oi: http s//do i certified by	pi org/10.110 1/768 y peer review) is th	8047; this version he author/fun	sion p osto d Augu dep All rights res	u s t 6 6, 2049. The served. No reuse	e comright holde allowed without	er f or this preprin It per nfiesion.	nt (which was not	84.97	85.62	86.19	85.98	85.61	85.76	85.78	84.94	84.89	87.12	88.52	90.71	90.71	90.78	100	85.9	85.83	85.85	85.92	85.91	85.69	85.85	85.87	85.73	85.82	86.57	82.06	X.populi CFBP1817 (T)
77.16	79.16	6 79.	.13 79	9.1 7	79.06	79.42	79.4	79.26	77.23	85.09	86.22	86.47	86.27	86.3	86.49	86.62	85.8	85.76	85.92	87.06	86.54	86.58	86.59	85.9	100	93.67	94.04	94.28	94.14	93.07	89.32	89.91	89.93	89.81	88.83	82.61	X.citri LMG 9322 (T)
77.25	79.11	1 79	9.3 79.	.15 7	79.26	79.49	79.56	79.33	77.22	85.07	85.94	86.35	86.21	86.47	86.53	86.72	85.9	85.76	85.87	86.94	86.37	86.37	86.4	85.83	93.67	100	94.14	94.06	94.11	93.16	89.15	89.52	89.65	89.58	88.63	82.78	X.phaseoli CFBP412
77.37	79.2	7 79.	.28 79.	.33 7	79.19	79.31	79.7	79.28	77.09	85.19	86.18	86.46	86.22	86.38	86.57	86.84	85.89	85.93	85.88	87.02	86.45	86.59	86.63	85.85	94.04	94.14	100	98.58	98.54	92.96	89.26	90.15	90.07	89.77	88.73	82.89	X.euvesicatoria LMG 27970
77.32	79.20	6 79.	.34 79.	.43 7	79.27	79.41	79.69	79.4	77.26	85.2	86.07	86.5	86.18	86.36	86.45	86.89	85.87	85.81	85.97	87.03	86.47	86.38	86.54	85.92	94.28	94.06	98.58	100	98.78	93.07	89.15	90.06	90.08	89.76	88.82	82.89	X.perforans 91-118
77.34	79.32	2 79.	.21 79.	.35 7	79.42	79.67	79.63	79.59	77.5	85.2	86.17	86.54	86.25	86.46	86.52	87.03	86.04	85.89	86.05	86.98	86.67	86.62	86.62	85.91	94.14	94.11	98.54	98.78	100	93.17	89.33	90.06	90.09	89.85	88.99	82.96	X.alfalfae LMG 495 (T)
77.28	78.8	8 79.	.14 79.	.06	79	79.33	79.56	79.27	77.04	84.92	85.74	86.17	85.85	86.06	86.15	86.41	85.49	85.46	85.72	86.58	86.17	86.13	86.19	85.69	93.07	93.16	92.96	93.07	93.17	100	89.31	89.64	89.64	89.43	88.58	82.77	X.axonopodis DSM3585 (T)
77.19	79.0	5 79.	.14 79.	.15 7	78.92	79.42	79.59	79.22	77	85.06	86.78	86.85	86.81	86.43	86.84	86.81	85.87	85.7	86.09	86.9	86.58	86.55	86.57	85.85	89.32	89.15	89.26	89.15	89.33	89.31	100	89.52	89.43	89.61	88.93	82.36	X.bromi CFBP1976 (T)
76.71	78.6	õ 78.	.65 78.	.56 7	78.72	79.05	79.24	78.6	76.94	84.89	85.91	86.11	86	85.85	85.97	86.18	85.39	85.39	85.95	86.76	86.42	86.43	86.36	85.87	89.91	89.52	90.15	90.06	90.06	89.64	89.52	100	91.3	91.11	88.78	82.34	X.vasicola NCPPB2717 (T)
77.11	78.9) 78	3.9 78.	.89 7	78.69	79.13	79.42	79.08	77.29	84.74	85.79	86.13	85.93	86.18	86.05	86.24	85.4	85.45	85.93	86.64	86.47	86.42	86.33	85.73	89.93	89.65	90.07	90.08	90.09	89.64	89.43	91.3	100	91.23	88.78	82.5	X.oryzae ATCC 35933 (T)
77.25	78.98	8 79.	.02 78.	.87 7	78.89	79.18	79.44	79.09	76.9	84.89	86.18	87.58	86.49	86.05	86.39	86.39	85.55	85.52	85.89	87.1	86.44	86.39	86.58	85.82	89.81	89.58	89.77	89.76	89.85	89.43	89.61	91.11	91.23	100	88.73	82.28	X.prunicola CFBP8353 (T)
77.63	79.22	2 79	9.4 79.	.32 7	79.41	79.72	79.77	79.4	77.18	85.53	87.36	87.38	87.41	86.88	87.16	87.21	86.24	86	86.67	87.68	87.56	87.45	87.45	86.57	88.83	88.63	88.73	88.82	88.99	88.58	88.93	88.78	88.78	88.73	100	82.73	X.nasturtii WHR18853 (T)
77.16	79.7	5 79.	.65 79.	.56 7	79.61	79.65	79.71	79.48	77.48	82.66	82.37	82.56	82.4	82.99	83.02	82.85	82.74	82.67	82	83.23	82.41	82.52	82.49	82.06	82.61	82.78	82.89	82.89	82.96	82.77	82.36	82.34	82.5	82.28	82.73	100	X.maliensis LMG 27592 (T)

global 76.68 96 100