1 Urbanization impacts apex predator gene flow 2 but not genetic diversity across an urban-rural divide 3 4 Trumbo DR¹, Salerno PE¹, Logan KA², Alldredge M³, Gagne RB⁴, Kozakiewicz CP⁵, Kraberger S⁴, 5 Fountain-Jones N⁶, Craft ME⁶, Carver S⁵, Ernest HB⁷, Crooks K⁸, VandeWoude S⁴, Funk WC^{1,9} 6 7 ¹Department of Biology, Colorado State University, Fort Collins, CO 80523 USA 8 ²Colorado Parks and Wildlife, Montrose, CO 81401 USA 9 ³Colorado Parks and Wildlife, Fort Collins, CO 80526 USA 10 ⁴Department of Microbiology, Immunology, and Pathology, Colorado State University, Fort Collins, CO 11 80523 USA 12 ⁵Department of Biological Sciences, University of Tasmania, Hobart, TAS 7005 Australia 13 ⁶Department of Veterinary Population Medicine, University of Minnesota, Saint Paul, MN 55108 USA 14 ⁷Department of Veterinary Sciences, University of Wyoming, Laramie, WY 82070 USA 15 ⁸Department of Fish, Wildlife, and Conservation Biology, Colorado State University, Fort Collins, CO 16 80523 USA 17 ⁹Graduate Degree Program in Ecology, Colorado State University, Fort Collins, CO 80523 USA 18

Abstract

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

Apex predators are important indicators of intact natural ecosystems. They are also sensitive to urbanization because they require broad home ranges and extensive contiguous habitat to support their prey base. Pumas (*Puma concolor*) can persist near human developed areas, but urbanization may be detrimental to their movement ecology, population structure, and genetic diversity. To investigate potential effects of urbanization in population connectivity of pumas, we performed a landscape genomics study of 134 pumas on the rural Western Slope and more urbanized Front Range of Colorado, USA. Over 12,000 single nucleotide polymorphisms were genotyped using double-digest, restriction site-associated DNA sequencing (ddRADseq). We investigated patterns of gene flow and genetic diversity, and tested for correlations between key landscape variables and genetic distance to assess the effects of urbanization and other landscape factors on gene flow. Levels of genetic diversity were similar for the Western Slope and Front Range, but effective population sizes were smaller, genetic distances were higher, and there was more overall population substructure in the more urbanized Front Range. Forest cover was strongly positively associated with puma gene flow on the Western Slope, while impervious surfaces restricted gene flow and more open, natural habitats enhanced gene flow on the Front Range. Landscape genomic analyses revealed differences in puma movement and gene flow patterns in rural versus urban settings. Our results highlight the utility of dense, genome-scale markers to document subtle impacts of urbanization on a wide-ranging carnivore living near a large urban center. **Keywords:** landscape genomics, gene flow, genetic diversity, effective population size, urbanization,

Puma concolor

Urbanization is a major threat to biodiversity, and in particular to apex predators with broad home

Introduction

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

ranges (Cohen 2003; Theobald 2005; Crooks et al. 2017). Habitat fragmentation due to urbanization can have important impacts on predator movement, disease, and survival (Markovchick-Nicholls et al 2008; Carver et al. 2016; Fountain-Jones et al. 2017). This reduced connectivity can lead to smaller, more isolated populations, where less gene flow and genetic diversity, as well as smaller effective population sizes (Riley et al. 2006; Vandergast et al. 2007; Ernest et al. 2014) ultimately cause local and regional extirpations through environmental and demographic stochasticity and inbreeding depression (Allendorf et al. 2013). Moreover, increased human recreational activities in wildlife habitats associated with nearby urbanization can change wildlife movement patterns and habitat usage, exacerbating the impacts of fragmentation (McKinney 2002; Lewis et al. 2015). As human populations continue to expand worldwide, urban areas are becoming larger and more extensive on the landscape. However, we do not fully understand how urbanization affects natural ecosystems near wildland-urban interfaces (Radeloff et al. 2005; Magle et al. 2012). Large carnivores are important indicators of intact natural ecosystems, as they require an abundant and sustainable prey base, as well as high habitat connectivity to support their broad home ranges (Sergio et al. 2006, 2008). However, understanding the effects of urbanization on large carnivores is difficult due to their low population densities and secretive nature (Logan and Sweanor 2001; Riley et al. 2006; Hornocker and Negri 2009). Camera traps, radio-telemetry, and GPS collars provide valuable information on animal home ranges and population sizes (e.g., Lewis et al. 2015; Blecha et al. 2018), but these studies are expensive, time consuming, and can only monitor a small fraction of the total population for limited time periods. Population and landscape genetics can provide additional, complementary techniques for a more detailed understanding of wildlife populations (Epps et al. 2007; Lowe and Allendorf 2010; Balkenhol et al. 2016). Genetic studies provide an indicator of functional landscape connectivity through measures of gene flow, effective population sizes of breeding individuals, and cost-efficient monitoring of

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

genetic diversity across broad geographic areas (McRae et al. 2005; Solberg et al. 2006). Moreover, recent high-throughput sequencing technologies enable the genotyping of many more thousands of loci than previously possible, providing higher power to detect the often subtle population genetic structure of wideranging species such as large carnivores (Luikart et al. 2003; Holderegger et al. 2006). Pumas (*Puma concolor*; other common names include mountain lions, cougars, panthers, catamounts) are a large, apex predator with one of the broadest latitudinal ranges of any terrestrial carnivore, spanning western North America, Central America, and South America (Hornocker and Negri 2009). Pumas are sensitive to urbanization, requiring broad-scale landscape connectivity to persist, and are thus useful indicators for monitoring the effects of urban fragmentation (Beier 1995; Crooks 2002; Maletzke et al. 2017). Given sufficient habitat area and landscape connectivity, however, pumas can still persist within and adjacent to urban systems (Wilmers et al. 2013; Riley et al. 2014; Lewis et al. 2015; Zeller et al. 2017; Blecha et al. 2018). Furthermore, the substantial area requirements of large carnivores such as pumas can enhance their role as "umbrella" species, whose protection also benefits co-occurring species through broad-scale habitat preservation (Thorne et al. 2006). The southern Rocky Mountains in western Colorado, USA support natural habitats with high puma densities, as well as many rural and urban human developments (Hornocker and Negri 2009). The Western Slope of the Rocky Mountains primarily consists of large areas of contiguous public wildlands with an abundant prey base for pumas, interspersed with small rural and exurban developments, including the Uncompangre Plateau region near the town of Montrose (Western Slope Study Area; Figure 1). In contrast, the Front Range is a rapidly urbanizing, major metropolitan area on the Eastern Slope of the Continental Divide, where urbanization is spreading from lower elevation areas in and around the Denver Metropolitan Area into adjacent wildland habitats in the foothills of the Rocky Mountains. Pumas continue to persist near this wildland-urban interface, including adjacent to the city of Boulder on the western edge of the Denver Metropolitan Area (Front Range Study Area; Figure 1; Lewis et al. 2015; Moss et al. 2016a). From 2010 – 2017, Colorado was the 8th fastest growing U.S. state by population (577,829

residents added) and the 6th fastest by percentage (11.5% population growth; U.S. Census Bureau 2017), with most of this growth occurring along the eastern edge of the Front Range. Thus, comparative studies of puma movement and gene flow in one of the most populous states in the mid-continental USA, which also supports a robust puma population, can provide insight into the effects of urbanization on this important apex predator.

Here, we tested how different landscape factors, including urbanization, enhance or restrict gene flow and genetic diversity in a large apex predator across an urban-rural divide in Colorado, USA. A large sample (n = 134) of pumas were utilized from (a) the rural Western Slope and (b) the more urbanized Front Range (Figure 1). We used double digest restriction site associated DNA sequencing (ddRADseq) to genotype pumas at 12,444 single nucleotide polymorphism (SNP) loci to evaluate the potential differences in gene flow, effective population sizes, genetic diversity, and population structure in these two different landscapes. We tested landscape genomic hypotheses by correlating key landscape factors with puma genetic distance measures. We hypothesized that pumas in the more urbanized Front Range would have (a) smaller effective population sizes, (b) lower levels of genetic diversity, and (c) more landscape factors related to urbanization that restrict gene flow, relative to the rural Western Slope landscape.

Materials and Methods

Samples and sequences

Puma blood and tissue samples were collected as part of ongoing monitoring efforts by Colorado Parks and Wildlife in both the Western Slope and Front Range regions of the southern Rocky Mountains of Colorado, USA (Figure 1; Lewis *et al.* 2015; Carver *et al.* 2016). Samples were collected from 2005-2014 on the Western Slope and 2007-2013 on the Front Range. Western Slope samples consisted of 36 males and 42 females, and Front Range samples consisted of 24 males, 31 females, and 1 puma of unknown sex. Our sampling represents a large proportion of the resident pumas present in both regions during the sampling period, as Lewis *et al.* (2015) estimated 14.4 (S.E. 1.6) and 14.7 (S.E. 1.3) resident

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

pumas occupying the Western Slope and Front Range study areas at a single time point, respectively, from motion camera and telemetry data collected in 2009 and 2010. Genomic DNA was extracted from tissue or blood using QIAGEN DNeasy Blood & Tissue kits (QIAGEN Inc., Valencia, CA). We genotyped a total of 78 individuals from the Western Slope and 56 individuals from the Front Range using the ddRADseq protocol described in Peterson et al. (2012) and sequenced on Illumina HiSeq 2500 and 4000 machines (Illumina, San Diego, California) using 100bp single-end sequencing at the University of Oregon Genomics Facility (gc3f.uoregon.edu). We tested 9 different combinations of restriction enzymes on puma samples for digestion efficiency and evaluated the size ranges of fragment distributions using an Agilent Tapestation 2200 (Agilent Genomics, Santa Clara, California). We chose the digest enzymes EcoRI-HF (6bp recognition) and NlaIII (4bp recognition) and a target fragment size range of 300–400 bp (excluding adapters). We used a Blue Pippin with a 2%, internal standard, 100-600 bp gel cartridge (Sage Science, Beverly, Massachusetts) for size selection and a biotinylated P2 adapter with DynaBeads[®] (Peterson et al. 2012) to purify the polymerase chain reaction (PCR) template for the final enrichment. PCR was performed for 12 cycles and five reactions were tested for each pool of individuals. We initially genotyped 16 individuals multiplexed into an Illumina 2500 HiSeq lane to estimate maximum multiplexing based on a target of >12X coverage per locus. After assessment of locus coverage, we proceeded to multiplex 48 and 70 individually-barcoded samples on Illumina 2500 and 4000 HiSeq lanes, respectively, using the Peterson et al. (2012) flex adaptors. Bioinformatics pipeline and filters We evaluated read quality for each sequencing lane using FastQC (bioinformatics.babraham.ac.uk) and assembled our SNP dataset de novo using Stacks v 1.41 (Catchen et al. 2013). Details on Stacks code and parameter settings used are on the GitHub repository; github.com/pesalerno/PUMAgenomics. We demultiplexed and filtered sequencing reads using the program process_radtags in Stacks. Due to sensitivity of downstream genotyping with different Stacks

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

parameter settings (Mastretta-Yanes et al. 2015; Paris et al. 2017), we incorporated individual sample replicates in library preparations. In each library, we included 3 within and 3 between library replicates, which were used for estimating genotyping error rates for different combinations of parameters used to construct loci with the *denovo map.pl* Stacks pipeline. We ran 11 different *de novo* assemblies varying 4 different Stacks parameters that affect locus, allele, and SNP error rates and the number of loci genotyped, consisting of (1) minimum number of identical, raw reads required to create a stack (-m), (2) number of mismatches allowed between loci when processing a single individual (-M), (3) number of mismatches allowed between loci when building the catalog (-n), and (4) maximum number of stacks at a single de novo locus (-max locus stacks) (Table S1; Mastretta-Yanes et al. 2015). Locus error rate was calculated as the number of loci present in only one of the samples of a replicate pair divided by the total number of loci, allele error rate was the number of allele mismatches between replicate pairs divided by the number of loci, and SNP error rate was the proportion of SNP mismatches between replicate pairs. After identifying the most supported parameter settings that minimized locus, allele, and SNP error rates, while maximizing the number of SNPs (-m = 3, -M = 4, -n = 4, max_locus_stacks = 3; Table S1), we exported the SNP matrix with the *populations* program in Stacks (Catchen *et al.* 2013), retaining SNPs that were present in at least 20% of individuals by population, and retaining a single random SNP per locus. This matrix was further filtered for missing data in Plink v. 1.07, first by locus, then by individual, and then by minor allele frequency (MAF) using multiple combinations of thresholds for reducing missing data in the matrix (see github.com/pesalerno/PUMAgenomics). After evaluating missing data from SNP matrices, we retained the matrix with a more stringent locus filter (excluding loci missing >25% individuals) and a less stringent filter on minor allele frequency (excluding loci with MAF < 0.01). We additionally filtered loci that were found at position 95 (the last position of our reads) due to a higher number of SNPs present in this position, suggesting increased error rates due to low sequence quality towards the end of the sequencing read. In order to compare landscape resistances with putatively neutral loci, we used a Principal Components Analysis (PCA) to identify loci showing strong signatures of

selection relative to neutral background genomic variation with the program PCAdapt (Luu *et al.* 2016). We found twelve, putatively adaptive, outlier loci using a false discovery rate of 10%, so we filtered these outliers out for downstream landscape genomic analyses to avoid confounding neutral demographic patterns with patterns generated by loci under selection.

Population genomics and structure

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

Population genomic statistics were calculated for the two sampling regions, the Western Slope and Front Range (Figure 1). Observed and expected heterozygosity (H_{obs} and H_{exp}), nucleotide diversity (π), inbreeding coefficient (F_{IS}), and population genetic differentiation (F_{ST}) were calculated using the populations program in Stacks with SNP loci that passed previous filters, excluding a single individual (sample_1382) that did not pass the 75% missing data threshold. We estimated allelic richness (A_r) using HP-RARE 1.0 (Kalinowski 2005), which corrects for variance in sample sizes using rarefaction. Two complementary, individual-based genetic distances were calculated: proportion of shared alleles distance (D_{DS}; Bowcok et al. 1994) using the adegenet R v. 3.3.3 package and relatedness distance (r; Smouse and Peakall 1999) using the PopGenReport R package. We then calculated mean genetic distance among individuals for each region, corrected for geographic distance (i.e., genetic distance per km), since individuals that are farther apart are expected to have higher genetic distances due to neutral isolation by distance population processes (Wright 1942; Balkenhol et al. 2016). Effective population sizes (N_e) were estimated using the linkage disequilibrium method in NeEstimator v. 2.01 (Do et al. 2014), using the correction for chromosome number (Waples et al. 2016), which has been shown to be a robust method for inferring N_e using SNP datasets and large sample sizes (Waples 2016; Waples et al. 2016). We evaluated overall genetic structure as well as genetic differentiation among the two sampling sites (Western Slope and Front Range) using PCA and Discriminant Analysis of Principal Components (DAPC) in the R package adegenet (Jombart 2008) and Admixture ancestry analysis (Alexander et al. 2009). We used the function assignplot to identify individuals that were putative migrants or admixed based on the individual

DAPC assignment probabilities. We used the find clusters command in adegenet and minimized cross validation error in Admixture to estimate the number of populations (i.e., K).

Landscape genomics

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

Geographic Information Systems (GIS) data were collected for different landscape factors that we hypothesized would affect puma dispersal and gene flow in Colorado. Table 1 provides details on GIS data sources, spatial resolution, and ecological justification for each landscape factor. Study area extents were calculated and landscape variables were compared across regions by buffering individual data points by a typical female puma dispersal distance of 34.6 km (Logan and Sweanor 2001), dissolving overlapping buffers, and calculating zonal statistics within each region (Western Slope and Front Range) using ArcGIS v. 10.1 (ESRI, Redlands, California). Landscape data were converted into resistance surfaces using the Reclassify and Raster Calculator tools in ArcGIS. The following hypothesized relationships of landscape factors with puma gene flow were modeled: percent impervious surface cover (negative effect on gene flow), land cover (forested, open-natural, and developed: positive, neutral, and negative effects on gene flow, respectively), percent tree canopy cover (positive effect), vegetation density (positive effect), river and stream riparian corridors (positive effect), roads (negative effect), minimum temperature of the coldest month (negative effect), annual precipitation (positive effect), topographic roughness (positive effect), and elevation (negative effect). Additionally, we included an isolation by geographic distance model, which would be supported if none of the landscape variables had an effect on gene flow except for straight line, Euclidean distance between individuals (Wright 1942; Balkenhol et al. 2016). Table S2 describes methods and justification for converting raw landscape variables to resistance surfaces. Two genetic distance measures were used as response variables in landscape genomic analyses: proportion of shared alleles distance (D_{ps}; Bowcok et al. 1994) and relatedness distance (r; Smouse and Peakall 1999). Environmental resistances among individuals were calculated using Circuitscape (McRae

217

218

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

2006) for each landscape resistance surface (McRae 2006; Row et al. 2017). Circuitscape resistances are a useful tool in landscape genetics because they summarize all potential movement pathways simultaneously, as opposed to least cost paths that evaluate only a single idealized pathway, and thus assume the study organism has complete knowledge of the landscape and always chooses the ideal pathway (McRae 2006; Balkenhol et al. 2015). Landscape variables were tested for multicollinearity, both prior to and after calculating environmental resistances in Circuitscape, to ensure Pearson's r correlations < 0.7 and variance inflation factor (VIF) scores < 5 in final landscape genomics models, as collinearity can cause instability in parameter estimation in regression models (Tables S3 and S4; Warren et al. 2010; Dormann et al. 2012; Rowe et al. 2017). Two complementary methods were used to estimate the effects of environmental resistances on genetic distances: multiple regression on distance matrices (MRDM; Legendre et al. 1994) using PERMUTE v.3.4 and maximum likelihood of population effects (MLPE; Clark et al. 2002; van Strien et al. 2012; Row et al. 2017) using the lme4 R package. MRDM is a permutational, distance matrix-based approach that has been traditionally used in landscape genetic analyses, whereas MLPE is a newer linear mixed effects modeling technique that models pairwise comparisons as a random effect and environmental resistances as fixed effects (Balkenhol et al. 2016). Recent evaluations of landscape genetic approaches found linear mixed effects modeling using MLPE to be more accurate, although both approaches performed well (Shirk et al. 2017). Therefore, we included the traditional MRDM approach as well as MLPE in order to utilize multiple, complementary techniques for inferring associations between landscape features and gene flow. For MRDM and MLPE, genetic distances were the response variable and environmental resistances were explanatory variables. Additionally for MLPE, a random effect matrix of individual comparisons was included to control for the non-independent, pairwise structure of the data, and landscape resistances were standardized to units of standard deviation centered on the mean (van Strien et al. 2012; Row et al. 2017). Models were ranked using the Bayesian information criterion (BIC), and top models within 5 BIC units are reported (Richards 2015).

241 242 Results 243 Genotyping and filtering SNP matrices 244 Initial Stacks processing retained a single random SNP per 95 bp read and SNPs present in at least 245 20% of individuals by population, resulting in a matrix of 98,813 SNPs. These SNPs were further filtered 246 in Plink by removing loci that were present in less than 75% of individuals, which resulted in a matrix of 247 20,355 SNPs. Only a single individual was excluded based on our >75% missing loci per individual 248 threshold. After excluding SNPs present in the 95th sequencing base position and with minor allele 249 frequency <0.01, we retained 12,456 SNPs. PCAdapt detected twelve outlier loci, putatively under 250 selection, while accounting for population structure (K=2). After removing these putatively adaptive loci, 251 the final neutral dataset contained 12,444 SNPs (Table S1; github.com/pesalerno/PUMAgenomics). 252 253 Population genomics and structure 254 The two study areas encompass similar geographic extents: 11,889 km² for the Western Slope and 255 11,958 km² for the Front Range (Table 2). Measures of genetic diversity (H_{obs} , H_{exp} , π , A_r ,) and inbreeding 256 (F_{IS}) were similar for the Western Slope and Front Range (Table 2). However, the effective population 257 size (N_e) was smaller, mean genetic distances among individuals $(D_{PS}/km \text{ and } r/km)$ were higher, and there 258 was more overall population substructure in the more urbanized Front Range (Table 2, Figure 2). We also 259 calculated N_e using subsets of individuals (i.e., pre and post-2010 individuals in the Front Range, pre and 260 post-2011 individuals in the Western Slope), since multiple overlapping generations may bias effective 261 population size estimates low or high (Waples 2016; Waples et al. 2016). N_e remained consistently higher 262 in the Western Slope, although it differed between the earlier and later sampling periods there, and 263 indicated the population may be expanding (Table S5). We found a detectable signature of population 264 differentiation between the Western Slope and Front Range regions based on a PCA and DAPC, and 265 Admixture ancestry analysis indicated K=2 was the best supported value of K by minimizing cross

validation error (Figure 2; Alexander *et al.* 2009). The proportion of correct individual assignment to populations based on DAPC (Figure 2b), which attempts to minimize within population distances and maximize between population distances (Jombart 2008), was high for most individuals in both the Western Slope (0.98) and the Front Range (0.96). However, the DAPC assignplot also identified admixed individuals and putative migrants between regions, including a female and a male in the Front Range that assigned mostly to the Western Slope, and an admixed male in the Western Slope that assigned mostly to the Front Range (Figure 2b). We also analyzed both regions separately for population substructure (Figure S1), and there was no signature of population differentiation within the Western Slope or Front Range, further supporting two populations.

Landscape Genomics

The Front Range has more urban development than the Western Slope, with more impervious surface cover and a higher density of roads (Figure 1, Table 3, Table S2). The Front Range also has more tree canopy cover, higher vegetation density, and higher annual precipitation than the Western Slope (Table 3), likely due to the high desert habitats (i.e., the Colorado Plateau ecoregion) in the Western Slope being drier than the grassland and shrub habitats found at lower elevations of the Front Range (i.e., the Great Plains ecoregion; McMahon *et al.* 2001).

Prior to running Circuitscape, landscape raster surfaces were largely uncorrelated (i.e., Pearson's r < 0.7), with the exception of elevation, which was positively correlated with annual precipitation and negatively correlated with minimum temperature of the coldest month in both regions, and vegetation density, which was negatively correlated with annual precipitation in the Front Range (Table S3). After Circuitscape analyses, environmental resistance variables showed more collinear relationships than raw raster surfaces (Table S4), likely due to Circuitscape resistances being higher for individuals separated by larger geographic distances (McRae 2006). Therefore, we removed landscape variables from both regions that were strongly correlated with many other variables, until all VIF scores were less than 10 (Row *et al.*

2017). Variables retained were geographic distance, river and stream riparian corridors, roads, impervious surface cover, tree canopy cover, vegetation density, and minimum temperature of the coldest month. However, vegetation density was still correlated with geographic distance in both regions, and impervious surface was correlated with geographic distance and tree canopy cover in the Western Slope (Table S4). We removed these variables as well, resulting in Pearson's r correlations less than 0.7 and VIF scores less than or equal to 4.1 and 3.5 in the Western Slope and Front Range, respectively, for all explanatory variables. Thus final MRDM and MLPE models for the Western Slope included geographic distance, tree canopy cover, stream and river riparian corridors, roads, and minimum temperature of the coldest month; and for the Front Range included the same landscape variables plus impervious surface cover.

Landscape genomic patterns of pumas were different in the rural Western Slope compared to the more urbanized Front Range, with the exception of geographic distance being supported in both regions (Tables 4 and 5). In the Western Slope, tree canopy cover was consistently positively correlated with gene flow in MRDM and MLPE models, and low minimum temperatures of the coldest month (i.e., those found in high elevation, alpine tundra habitats) were negatively correlated gene flow in one MLPE model (Tables 4 and 5). In contrast, in the Front Range, tree canopy cover and percent impervious surface cover were negatively associated with gene flow in the top MLPE models (Table 5). Since the relationship between tree cover and gene flow was the opposite of what we hypothesized in the Front Range, we also inverted the tree cover resistance surface (i.e., making higher tree cover = higher resistance), reran Circuitscape and MLPE analyses, and higher tree cover still showed significant negative correlations with gene flow in this region.

Discussion

The apex predator puma (*Puma concolor*) persists in many urbanized regions throughout its range, yet the localized effects of recent urban sprawl remain unclear. Here, we compared patterns of genomic landscape connectivity and diversity of pumas across two regions that span an urban-rural divide in

Colorado, USA. Landscape genomic connectivity patterns differed between regions, such that genetic distances were higher and urbanization (i.e., percent impervious surface cover) restricted gene flow in the more urbanized Front Range, whereas forest cover was most important for enhancing gene flow on the rural Western Slope. Despite finding reductions in gene flow associated with urbanization on the Front Range, population-level genetic diversity and inbreeding measures were similar to those on the rural Western Slope. This suggests that recent urban sprawl in the Colorado Front Range has not yet had a substantial impact on the genetic diversity of pumas. This is in contrast to more isolated puma populations in other highly urbanized landscapes such as southern California and Florida, which exhibit reduced genetic diversity and strong evidence of inbreeding compared to Colorado pumas (Ernest *et al.* 2003, 2014; Johnson *et al.* 2010). However, a smaller effective population size, higher among-individual genetic distances, and higher population substructure in the recently urbanized Front Range suggest habitat fragmentation has already impacted this population and could cause further reductions of genetic diversity as urbanization continues to expand in Colorado (Theobald 2005; U.S. Census Bureau 2017). If puma populations decline, this could have important cascading effects into lower trophic levels, such as overgrazing of vegetation by ungulate herbivores (Markovchik-Nicholls *et al.* 2008).

Population genomics and structure

The Western Slope and Front Range were resolved as two genetically distinct groups (i.e., K=2; Figures 1 and 2). Minimum temperature of the coldest month was also negatively associated with gene flow in one of the top landscape genomic models on the Western Slope (Table 5), suggesting there may be restricted gene flow through high elevation, alpine tundra habitats (McMahon *et al.* 2001). However, potential immigrants and admixed individuals were identified moving in both directions (Figure 2) and overall genetic differentiation between the two populations was low (pairwise $F_{ST} = 0.02$; Table 2). Since our sample archive consisted of opportunistically collected samples, our analyses were restricted to populations in two distinct regions, whereas pumas occur throughout the southern Rocky Mountains in

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

Colorado. Therefore, potential immigrants and admixed individuals are not necessarily moving between our specific Western Slope and Front Range study areas, but may originate from other unsampled populations that share genetic ancestry with our two study regions. Nevertheless, results from our study suggest pumas may be somewhat limited in dispersing across the high elevation peaks of the Continental Divide, and future studies should attempt to sample more intensively across the entire region to further investigate this trend. We identified similar levels of genetic diversity and inbreeding between the rural Western Slope and more urbanized Front Range (Table 2), suggesting urbanization is not yet having a large impact on the genetic diversity of pumas in Colorado. One potential explanation is that urbanization in the Front Range is primarily occurring on the eastern edge of the region, possibly creating a relatively impermeable urban boundary on the eastern border, but not isolating pumas in fragments or limiting their connectivity to wildland habitat to the west (Figure 1; Lewis et al. 2015; Blecha et al. 2018). Another possibility is that many of the SNPs we sampled may not have high enough mutation rates to show a strong genomic signature of the relatively recent effects of rapid urbanization occurring in the Front Range (Haasl and Payseur 2011; Allendorf et al. 2013). As the human population continues to expand, future urbanization could result in more fragmented populations and reductions in genetic diversity, as has been detected in other more urbanized landscapes like southern California and Florida (Ernest et al. 2003, 2014; Johnson et al. 2010). Despite similar geographic extents and levels of genetic diversity in the Western Slope and Front Range, mean genetic distances among individuals were higher in the urban Front Range (Table 2), suggesting that fragmentation due to urbanization may be limiting puma dispersal and gene flow. In addition, a larger effective population size (N_e) of pumas was detected on the rural Western Slope $(N_e=69.3)$ compared to the urban Front Range $(N_e=40.2;$ Table 2), with the caveat that some assumptions of this estimator are violated in both regions (e.g., closed populations with no immigration, nonoverlapping generations). The effect of non-overlapping generations on N_e is difficult to predict (Waples et al. 2016), and this assumption is expected to be violated similarly in both the Western Slope and Front Range populations. Immigration, however, is expected to downwardly bias N_e by creating linkage disequilibrium through a multi-locus Wahlund effect (Wahlund 1928; Waples and England 2011). Thus, it is possible that the Front Range may be showing a lower N_e due to having more immigrants from outside populations than the Western Slope. This is possible, and perhaps likely, given the higher overall population substructure in the Front Range (Figure 2), which could indicate more potential immigrants into this region. On the other hand, if immigration rates are similar for both regions, the relatively smaller Front Range N_e may be due to (1) urbanization and fragmentation impacting and limiting population size, and/or (2) species range limit theory (Abundant Center Hypothesis) predicting that smaller population sizes are likely to occur at the edge of the geographic range relative to core areas (Brown 1984; Sagarin and Gaines 2002). These potential underlying factors are not mutually exclusive and may both be acting together. However, the lack of difference in most genetic diversity measures, in addition to slightly lower allelic richness in the Front Range, which is the most sensitive metric to recent bottlenecks (Allendorf *et al.* 2013), suggests lower effective population size on the Front Range may be more consistent with recent urbanization impacts than historical range boundary effects.

Landscape genomics

With regard to general landscape genomics methodology, we found MRDM to be a much more conservative approach that adds fewer explanatory variables to the models than MLPE (Tables 4 and 5). Conversely, MLPE results in more complex models with more explanatory variables and higher r^2 values (genetic variation explained) than MRDM (Tables 4 and 5). The different genetic distance measures we used (D_{PS} and r) showed largely consistent relationships with landscape variables, but still provided a few different insights, particularly using MLPE (Tables 4 and 5). Overall r^2 values were somewhat low (r^2 = 0.04 - 0.08 for MRDM, r^2 = 0.11 - 0.17 for MLPE), but this is expected for a large carnivore with extreme

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

414

long distance dispersal abilities (e.g., Short Bull et al. 2011, Balkenhol et al. 2016). Isolation by distance was important across models for both regions (Tables 4 and 5). On the rural Western Slope, tree canopy cover was most important for enhancing gene flow, suggesting pumas prefer to disperse through forests rather than more open shrub and grassland habitats in this landscape (Table 5). Forests provide more cover for concealment and ambush predation (Logan and Sweanor 2001; Hornocker and Negri 2009; Warren et al. 2016). Use of open areas may also increase susceptibility to mortality by hunters and ranchers (Newby et al. 2013), which are both more prevalent in the rural Western Slope than the more urbanized Front Range. In addition, non-forested areas on the Western Slope are dry, high elevation desert habitats (i.e., the Colorado Plateau ecoregion; McMahon et al. 2001), which may provide less prey and water resources, and thus be poorer habitats for hunting and dispersal (Sweanor et al. 2000; McRae et al. 2005; Dickson et al. 2013). In the more urbanized Front Range, impervious surface cover restricted gene flow (Table 5). This suggests urbanization is limiting gene flow, despite high levels of genetic diversity (Table 2). Similarly, Lewis et al. (2015) found pumas were less likely to be detected in habitats with residential development, even low-density exurban developments, which are increasingly encroaching into the foothills of the Front Range region. Genetic studies on pumas from more urbanized and fragmented populations in southern California and Florida have detected strong inbreeding and isolation associated with urbanization (Ernest et al. 2003, 2014; Johnson et al. 2010; Riley et al. 2014). Our study detected more subtle impacts of urbanization in a less fragmented landscape, within mountainous wildland habitats adjacent to a major metropolitan center, which experiences high levels of human outdoor recreation activities such as hiking and skiing (Figure 1). In addition, in contrast with the rural Western Slope and contrary to our initial hypotheses, forest cover was negatively associated with gene flow on the Front Range (Table 5). This pattern suggests pumas are more willing to disperse through open shrub and grassland habitats in this region. The reasons for this are unclear, but pumas living in the more developed Front Range may be more

acclimated to human activities and thus more willing to travel outside of forested habitats, demonstrating

that pumas have a range of adaptable behaviors and will use and move through different types of habitat (Dickson *et al.* 2005; Blecha *et al.* 2018). Pumas may also be hunting more urban mesopredators, domestic, and agricultural animals in these open habitats on the more developed Front Range, which was shown in a prior study using stable isotope analysis of Front Range puma diets (Moss *et al.* 2016b). There is also less hunting of pumas in the Front Range compared to the rural Western Slope, so pumas may be less wary of open areas, although this effect would be expected to be counteracted in part by higher traffic mortality in the more urbanized region (Beier 1995; Crooks 2002).

Conclusions

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

430

431

432

433

434

435

436

437

438

439

Our findings are consistent with prior comparative landscape genetic studies that have revealed varying effects of landscape factors on movement and gene flow across different portions of a species' geographic range (e.g., Vandergast et al. 2007; Short Bull et al. 2011; Trumbo et al. 2013). We found that in the rural Western Slope with high hunting pressure, forests with high tree canopy cover are most important for conserving puma genetic connectivity. In contrast, in the more urbanized Front Range, nonforested habitats such as shrublands and grasslands habitats are utilized for dispersal and gene flow, effective population sizes are smaller, genetic distances among individuals are higher, and gene flow is being restricted by urbanization (Tables 2, 4, and 5). Next generation sequencing techniques can provide dense, genome-scale SNP datasets of thousands of putatively neutral markers, which gives researchers increased power to detect the often subtle effects of landscape factors, such as urbanization, on gene flow (Luikart et al. 2003; Lowe and Allendorf 2010; Allendorf et al. 2013). This is particularly important for wide-ranging species with broad geographic distributions, since landscape effects on gene flow occur at broader geographic scales and may be weaker and more difficult to detect compared to more dispersallimited species with smaller home ranges (Holderegger et al. 2006; Epps et al. 2007; Balkenhol et al. 2016). Indeed prior work on pumas using 16 microsatellites found no population structure across the southern Rocky Mountains of Colorado and northern New Mexico (McRae et al. 2005). Our results

441

442

443

444

445

446

447

448

449

450

451

452

453

454 455

456

457

458

459

460

461

462

463

464

465

466

467

468

469

470

471

472

473

demonstrate that large SNP datasets can allow researchers to identify impacts of urbanization on gene flow, effective population sizes, and patterns of population genetic structure of wide-ranging species, even before fragmentation is extensive enough to greatly reduce genetic diversity. Maintaining genetic connectivity in these "umbrella" species can have outsized benefits towards conserving biodiversity, since preserving broad swaths of contiguous habitats that are necessary for their persistence also benefits many other species with smaller home ranges and narrower habitat requirements (Sergio et al. 2006, 2008; Thorne et al. 2006). Acknowledgements Funding was provided by the National Science Foundation, Ecology of Infectious Disease Program (NSF-EID 1413925 and 723676). Samples were collected by Colorado Parks and Wildlife. We also thank Michael Antolin, Kelly Pierce, and Jill Gerberich at Colorado State University for assistance in the lab. References Alexander DH, Novembre J, Lange K (2009) Fast model-based estimation of ancestry in unrelated individuals. Genome Research, 19, 1655–1664. Allendorf FW, Luikart G, Aitken SN (2013) Conservation and the genetics of populations. Wiley-Blackwell, Chichester, West Sussex, United Kingdom. Balkenhol N, Cushman SA, Storfer AT, Waits LP (2016) Landscape genetics: concepts, methods, applications. Wiley-Blackwell, Chichester, West Sussex, United Kingdom. Beier P (1995) Dispersal of juvenile cougars in fragmented habitat. Journal of Wildlife Management, 59, 228-237. Blecha KA, Boone RB, Alldredge MW (2018) Hunger mediates apex predator's risk avoidance response in wildland–urban interface. *Journal of Animal Ecology*, **87**, 609-622. Bowcock AM, Ruiz-Linares A, Tomfohrde J, et al. (1994) High resolution of human evolutionary trees with polymorphic microsatellites. *Nature*, **368**, 455–457. Brown JH (1984) On the relationship between abundance and distribution of species. American Naturalist, **124**, 255–279. Carver S, Bevins SN, Lappin MR, et al. (2016) Pathogen exposure varies widely among sympatric populations of wild and domestic felids across the United States. Ecological Applications, 26, 367-381. Catchen JM, Hohenlohe PA, Bassham S, et al. (2013) Stacks: an analysis tool set for population genomics.

Molecular Ecology, **22**, 3124–3140.

- Clarke RT, Rothery P, Raybould AF (2002) Confidence limits for regression relationships between distance matrices: estimating gene flow with distance. *Journal of Agricultural Biological and Environmental Statistics*, **7**, 361–372.
 - Cohen JE (2003) Human population: the next half century. Science, 302, 1172–1175.

- Crooks KR (2002) Relative sensitivities of mammalian carnivores to habitat fragmentation. *Conservation Biology*, **16**, 488-502.
- Crooks KR, Burdett CL, Theobald DM, *et al.* (2017) Quantification of habitat fragmentation reveals extinction risk in terrestrial mammals. *Proceedings of the National Academy of Sciences*, **114**, 7635-7640.
- Cushman SA, Gutzweiler K, Evans JS, McGarigal K (2010) The gradient paradigm: a conceptual and analytical framework for landscape ecology. In Cushman SA, Huettman F (editors) Spatial complexity, informatics, and wildlife conservation. Springer, New York, USA
- Dickson BG, Jenness JS, Beier P (2005) Influence of vegetation, topography, and roads on cougar movement in southern California. *Journal of Wildlife Management*, **69**, 264-276.
- Dickson BG, Roemer GW, McRae BH, Rundall JM (2013) Models of regional habitat quality and connectivity for pumas (*Puma concolor*) in the southwestern United States. *Plos One*, **8**, 1-11.
- Do C, Wapels RS, Peel D *et al.* (2014) NeEstimator v2; re-implementation of software for the estimation of contemporary effective population size (Ne) from genetic data. *Molecular Ecology Resources*, **14**, 209–214.
- Dormann CF, Elith J, Bacher S, *et al.* (2012) Collinearity: a review of methods to deal with it and a simulation study evaluating their performance. *Ecography*, **36**, 27-46.
- Epps CW, Wehausen JD, Bleich VC, et al. (2007) Optimizing dispersal and corridor models using landscape genetics. *Journal of Applied Ecology*, **44**, 714-724.
- Ernest HB, Boyce WM, Bleich VC, et al. (2003) Genetic structure of mountain lion (*Puma concolor*) populations in California. *Genetics*, **4**, 353-366.
- Ernest HB, Vickers TW, Morrison SA, *et al.* (2014) Fractured genetic connectivity threatens a southern California puma (*Puma concolor*) population. *Plos One*, **9**, 1-12.
- Fountain-Jones NM, Craft ME, Funk WC, *et al.* (2017) Urban landscapes can change virus gene flow and evolution in a fragmentation-sensitive carnivore. *Molecular Ecology*, **26**, 6487-6498.
- Haasl RF, Payseur BJ (2011) Multi-locus inference of population structure: a comparison between single nucleotide polymorphisms and microsatellites. *Heredity*, **106**, 158-171.
- Hijmans RJ, Cameron SE, Parra JL, *et al.* (2005) Very high resolution interpolated climate surfaces for global land areas. *International Journal of Climatology*, **25**, 1965–1978
- Hilty JA, Merenlender AM (2004) Use of riparian corridors and vineyards by mammalian predators in northern California. *Conservation Biology*, **18**, 126-135.
- Holderegger R, Kamm U, Gugerli F (2006) Adaptive vs. neutral genetic diversity: implications for landscape genetics. *Landscape Ecology*, **21**, 797-807.
- Homer C, Huang C, Yang L, et al. (2004) Development of a 2001 national land cover database for the United States. *Photogrammetric Engineering and Remote Sensing*, **70**, 829–840.
- Hornocker M, Negri S (2009) Cougar: ecology and conservation. University of Chicago Press, Chicago, Illinois, USA.
- Johnson WE, Onorato DP, Roelke ME, *et al.* (2010) Genetic restoration of the Florida panther. *Science*, **329**, 1641-1645.
- Jombart T (2008) Adegenet: a R package for the multivariate analysis of genetic markers. *Bioinformatics*, **24**, 1403-1405.
- Kalinowski ST (2005) Hp-rare 1.0: a computer program for performing rarefaction on measures of allelic richness. *Molecular Ecology Notes*, **5**, 187–189.
- Legendre P, Lapointe FJ, Casgrain P (1994) Modeling brain evolution from behavior: a permutational regression approach. *Evolution*, **48**, 1487–1499.

- Lewis JS, Logan KA, Alldredge MW, *et al.* (2015) The effects of urbanization on population density, occupancy, and detection probability of wild felids. *Ecological Applications*, **25**, 1880-1895.
- Logan KA, Sweanor LL (2001) Desert puma: evolutionary ecology and conservation of an enduring carnivore. Island Press, Washingington, DC, USA.
 - Lowe WH, Allendorf FW (2010) What can genetics tell us about population connectivity? *Molecular Ecology*, **19**, 3038-3051.
 - Luikart G, England PR, Tallmon D, *et al.* (2003) The power and promise of population genomics: from genotyping to genome typing. *Nature Reviews Genetics*, **4**, 981-994.
 - Luu K, Bazin E, Blum MGB (2016) PCAdapt: an R package to perform genome scans for selection based on principal component analysis. *Molecular Ecology Resources*, **17**, 67-77.
 - Magle SB, Hunt VM, Vernon M, Crooks KR (2012) Urban wildlife research: past, present, and future. *Biological Conservation*, **155**, 23-32.
 - Maletzke B, Kertson B, Swanson M, *et al.* (2017) Cougar response to a gradient of human development. *Ecosphere*, **8**, 1-14.
 - Markovchick-Nicholls L, Regan HM, Deutschman DH, *et al.* (2008) Relationships between human disturbance and wildlife land use in urban habitat fragments. *Conservation Biology*, **22**, 99-109.
 - Mastretta-Yanes A, Arrigo N, Alvaraz N, *et al.* (2015) Restriction site-associated DNA sequencing, genotyping error estimation and de novo assembly optimization for population genetic inference. *Molecular Ecology*, **15**, 28-41.
 - McKinney ML (2002) Urbanization, biodiversity, and conservation. *BioScience*, **52**, 883–890.
 - McMahon G, Gregonis SM, Waltman SW, *et al.* (2001) Developing a spatial framework of common ecological regions for the conterminous United States. *Environmental Management*, **28**, 293-316.
 - McRae BH, Beier P, Dewald LE, *et al.* (2005) Habitat barriers limit gene flow and illuminate historical events in a wide-ranging carnivore, the American puma. *Molecular Ecology*, **14**, 1965-1977.
 - McRae BH (2006) Isolation by resistance. Evolution, 60, 1551–1561.

- Moss WE, Alldredge MW, Logan KA, Pauli JN (2016a) Human expansion precipitates niche expansion for an opportunistic apex predator (*Puma concolor*). *Scientific Reports*, **6**, 1-5.
- Moss WE, Alldredge MW, Pauli JN (2016b) Quantifying risk and resource use for a large carnivore in an expanding urban–wildland interface. *Journal of Applied Ecology*, **53**, 371-378.
- Naiman RJ, Decamps H, Polluck M (1993) The role of riparian corridors in maintaining regional biodiversity. *Ecological Applications*, **3**, 209-212.
- Newby JR, Mills LS, Ruth TK, *et al.* (2013) Human-caused mortality influences spatial population dynamics: pumas in landscapes with varying mortality risks. *Biological Conservation*, **159**, 230-239.
- Paris JR, Stevens JR, Catchen JM (2017) Lost in parameter space: a road map for Stacks. *Methods in Ecology and Evolution*, **8**, 1360-1373.
- Peterson BK, Weber JN, Kay EH, *et al.* (2012) Double digest RADseq: an inexpensive method for de novo SNP discovery and genotyping in model and nonmodel species. *PLoS One*, **7**, 1–11.
- Radeloff VC, Hamer RB, Stewart SI, *et al.* (2005) The wildland-urban interface in the United States. *Ecological Applications*, **15**, 799-805.
- Richards SA (2015) Likelihood and model selection. Ecological Statistics: Contemporary Theory and Application. Fox GA, Negrete-Yankelevich S, Sosa VJ. Oxford University Press, 58-80.
- Riley SP, Pollinger JP, Sauvajot RM, *et al.* (2006) A southern California freeway is a physical and social barrier to gene flow in carnivores. *Molecular Ecology*, **15**, 1733-1741.
- Riley SP, Serieys LEK, Pollinger JP, *et al.* (2014) Individual behavior dominates the dynamics of a mountain lion population in a highly fragmented urban landscape. *Current Biology*, **24**, 1989-1994.

Row JR, Knick ST, Oyler-McCance SJ, *et al.* (2017) Developing approaches for linear mixed modeling in landscape genetics through landscape-directed dispersal simulations. *Ecology and Evolution*, **7**, 3751–3761.

- Sagarin RD, Gaines SD (2002) The abundant centre distribution: to what extent is it a biogeographical rule? *Ecology Letters*, **5**, 137–147.
- Sergio F, Newton I, Marchesi L, Pedrini P (2006) Ecologically justified charisma: preservation of top predators delivers biodiversity conservation. *Journal of Applied Ecology*, **43**, 1049-1055.
- Sergio F, Caro T, Brown D, et al. (2008) Top predators as conservation tools: ecological rationale, assumptions, and efficacy. Annual Review of Ecology, Evolution, and Systematics, 39, 1-19.
- Shirk AJ, Landguth EL, Cushman SA (2017) A comparison of regression methods for model selection in individual-based landscape genetic analysis. *Molecular Ecology Resources*, **18**, 55-67.
- Short Bull RA, Cushman SA, Mace R, *et al.* (2011) Why replication is important in landscape genetics: American black bear in the Rocky Mountains. *Molecular Ecology*, **6**, 1092-1107.
- Smouse PE, Peakall R (1999) Spatial autocorrelation analysis of individual multiallele and multilocus genetic structure. *Heredity*, **82**, 561-573.
- Solberg KH, Bellemain E, Drageset OM, *et al.* (2006) An evaluation of field and non-invasive genetic methods to estimate brown bear (*Ursus arctos*) population size. *Biological Conservation*, **128**, 158-168.
- Sweanor LL, Logan KA, Hornocker MG (2000) Cougar dispersal patterns, metapopulation dynamics, and conservation. *Conservation Biology*, **14**, 798-808.
- Theobald DM (2005) Landscape patterns of exurban growth in the USA from 1980 to 2020. *Ecological Society*, **10**, 1–34.
- Thorne JH, Cameron D, Quinn JF (2006). A conservation design for the central coast of California and the evaluation of mountain lion as an umbrella species. *Natural Areas Journal*, **26**, 137-148.
- Trumbo DR, Spear SF, Baumsteiger J, Storfer A (2013) Rangewide landscape genetics of an endemic Pacific northwestern salamander. *Molecular Ecology*, **22**, 1250-1266.
- U.S. Census Bureau (2017) Cumulative estimates of resident population change for the United States, regions, states, and Puerto Rico and region and state rankings: April 1, 2010 to July 1, 2017, Table 2 (NST-EST2017-02). Source: U.S. Census Bureau, Population Division; Release date: December 2017; www.census.gov/programs-surveys/popest.html.
- Vandergast AG, Bohanak AJ, Weissman DB, Fisher RN (2007) Understanding the genetic effects of recent habitat fragmentation in the context of evolutionary history: phylogeography and landscape genetics of a southern California endemic Jerusalem cricket (Orthoptera: Stenopelmatidae: *Stenopelmatus*). *Molecular Ecology*, **16**, 977-992.
- van Strien MJ, Keller D, Holderegger R (2012) A new analytical approach to landscape genetic modelling: least-cost transect analysis and linear mixed models. *Molecular Ecology*, **21**, 4010-4023.
- Wahlund S (1928) Zuzammensetzung von populationen und korrelation-serscheiunungen von standpunkt der vererbungslehre aus betrachtet. *Hereditas*, **11**, 65–106
- Waples RS, England PR (2011) Estimating contemporary effective population size on the basis of linkage disequilibrium in the face of migration. *Genetics*, **189**, 633-644.
- Waples RS (2016) Making sense of genetic estimates of effective population size. *Molecular Ecology*, **25**, 4689-4691.
 - Waples RK, Larson WA, Waples RS (2016) Estimating contemporary effective population size in non-model species using linkage disequilibrium across thousands of loci. *Heredity*, **117**, 233-240.
- Warren DL, Glor RE, Turelli M (2010) ENMTools: a toolbox for comparative studies of environmental niche models. *Ecography*, **33**, 607–611.
- Warren MJ, Wallin DO, Beausoleil RA, Warheit KI (2016) Forest cover mediates genetic connectivity of northwestern cougars. *Conservation Genetics*, **17**, 1011-1024.

- Wilmers CC, Wang Y, Nickel B, *et al.* (2013) Scale dependent behavioral responses to human development by a large predator, the puma. *PLoS One*, **8**, 1-11.
- 620 Wright S (1942) Isolation by distance. *Genetics*, **28**, 114–138.
 - Zeller KA, Vickers TW, Ernest HB, Boyce WM (2017) Multi-level, multi-scale resource selection functions and resistance surfaces for conservation planning: pumas as a case study. *PloS One*, **12**, 1-20.

Author Contributions

D.R.T. performed laboratory work, analyzed landscape and population genomic data, and wrote the manuscript; P.S., R.B.G., C.P.K, S.K., and N.F.J. performed laboratory work and analyzed landscape and population genomic data; K.L. and M.A. directed fieldwork and collected field data; M.E.C., S.C., H.B.E., K.C., S.V., and W.C.F. conceived of study questions and directed research; and all authors contributed input to draft and final versions of the manuscript.

Data Accessibility

ddRADseq data used in genomic analyses will be uploaded to Dryad (datadryad.org) upon acceptance of the manuscript for publication.

Supporting Information

- **Table S1:** Library replicate analysis of error rates from different Stacks parameter settings for minimum number of identical raw reads required to create a stack (-m), number of mismatches allowed between loci when processing a single individual (-M), number of mismatches allowed between loci when building the catalog (-n), and maximum number of stacks at a single de novo locus (-max_locus_stacks). Locus error rate was the number of loci present in only one of the samples of a replicate pair divided by the total number of loci, allele error rate was the number of allele mismatches between replicate pairs divided by the number of loci, and SNP error rate was the proportion of SNP mismatches between replicate pairs.
- **Table S2**: Landscape resistance transformations for the Western Slope and Front Range.
- **Table S3**: Correlations (Pearson's r) between environmental raster surfaces used in landscape genomic analyses for the Western Slope and Front Range regions of Colorado. Pearson's correlations > 0.7 are in bold.
- **Table S4**: Correlations (Pearson's r) between Circuitscape environmental resistances used in landscape genomic analyses for the Western Slope and Front Range regions of Colorado. Pearson's correlations > 0.7 are in bold.
- **Figure S1**: Principle Components Analyses (PCAs) and Admixture plots of (a) 78 Western Slope pumas and (b) 56 Front Range pumas, analyzed separately within each region.

Tables

Table 1: Environmental variables used for landscape genomic analyses, data sources, spatial resolution, and ecological justification.

Category	Landscape Variable	Code	Description	Data Source, Spatial Resolution	Calculation	Ecological Justification
Distance	Isolation by geographic distance	Geo. dist.	Euclidean, straight- line distance between individuals	No environmental data; model assumes only distance affects gene flow, 30 meter	ArcGIS Reclassify tool, Circuitscape	Model of isolation by straight-line distance (Wright 1942).
Land cover	Land cover: forested, open- natural, and developed	Land cover	Multiple land cover categories collapsed into 3 costs of movement: forested (lowest), open natural areas (medium), and developed (highest)	National Land Cover Database (mrlc.gov/nlcd2011.php; Homer <i>et al.</i> 2011), 30 meter	ArcGIS Spatial Analyst	Forested habitats provide the most cover for hunting and dispersal, open natura areas are intermediate, are developed areas are the least suitable habitat for dispersal (Crooks 2002; Lewis et al. 2015).
	Percent impervious surface cover	Imperv.	Percentage of impervious surface	National Land Cover Database (mrlc.gov/nlcd2011.php; Homer <i>et al.</i> 2011), 30 meter	ArcGIS Spatial Analyst	Human development resu in increased noise, lights, and hunter access, limiting dispersal (Riley et al. 2006 Ernest et al. 2014; Maletzl et al. 2017).
	Road corridors	Roads	Roads, with 50 meter buffers on each side	Colorado Department of Transportation (dtdapps.coloradodot.info /otis), 30 meter	ArcGIS Analysis Tools, Spatial Analyst	Roads increase mortality, noise, lights, and hunter access, limiting dispersal (Riley et al. 2006; Newby al. 2013; Maletzke et al. 2017).
	River and stream riparian corridors	Riparian	River and stream riparian corridors, with 50 meter buffers on each side	National Hydrography Dataset (nhd.usgs.gov), 30 meter	ArcGIS Analysis Tools, Spatial Analyst	River and stream riparian corridors provide vegetati and topographical cover f dispersal, as well as water sources attracting prey species (Naiman et al. 199 Hilty and Merenlender 20 Dickson et al. 2005).
Vegetation	Percent tree canopy cover	Tree cover	Percentage of tree canopy cover	National Land Cover Database (mrlc.gov/nlcd2011.php; Homer et al. 2011), 30 meter	ArcGIS Spatial Analyst	Low tree canopy limits corfor ambush predation and concealment, and restrict dispersal (Sweanor et al. 2000; Logan and Sweanor 2001; Warren et al. 2016; Blecha et al. 2018).
	Enhanced vegetation index	Veg. density	Density of vegetation calculated from chlorophyll reflectance in visual and near-infrared spectra	Moderate Resolution Imaging Spectroradiometer (modis.gsfc.nasa.gov), 250 meter	ArcGIS Spatial Analyst	Low vegetation density limits cover for ambush predation and concealmer and restricts dispersal (Sweanor et al. 2000; Hilty and Merenlender 2004; Warren et al. 2016; Bleche et al. 2018).
Climate	Minimum temperature of the coldest month	Min. temp.	Mean annual minimum temperature of the coldest month (°C) calculated from	Global Climate Data (worldclim.org/bioclim; Hijmans <i>et al</i> . 2005), 1 kilometer	ArcGIS Spatial Analyst	Low minimum temperatur and high snowfall, found a high elevation mountain ridgelines (e.g., alpine tundra habitats) restrict

			1970-2000 weather station data, interpolated between stations			hunting, breeding, and dispersal (Hornocker and Negri 2009).
	Mean annual precipitation	Ann. precip.	Mean annual precipitation accumulation (mm) calculated from 1970-2000 weather station data, interpolated between stations	Global Climate Data (worldclim.org/bioclim; Hijmans <i>et al.</i> 2005), 1 kilometer	ArcGIS Spatial Analyst	Dry habitats with low precipitation accumulation limit prey species for hunting and vegetative cover, restricting dispersal (Logan and Sweanor 2001; McRae et al. 2005).
Topography	Topographic roughness	Topo. rough.	Topographic complexity based on variance in elevation within a moving window	National Elevation Dataset (Ita.cr.usgs.gov/ned) National Map Tool (viewer.nationalmap.gov), 30 meter	Geomorphometric and Gradient Metric Toolbox (Cushman et al. 2010), ArcGIS Spatial Analyst	Steep, topographically- complex canyons and mountain slopes provide cover for hunting and dispersal (Dickson et al. 2005; Hornocker and Negri 2009).
	Elevation	Elev.	Elevation calculated from digital elevation models.	National Elevation Dataset (Ita.cr.usgs.gov/ned) National Map Tool (viewer.nationalmap.gov), 30 meter	ArcGIS Spatial Analyst	Low minimum temperatures and high snowfall, found at high elevation mountain ridgelines (e.g., alpine tundra habitats) restrict hunting, breeding, and dispersal (Hornocker and
666 667						Negri 2009).

Table 2: Study areas (km²), number of individuals genotyped (N_{gen}), and population genomic parameter estimates from the Western Slope and Front Range of Colorado. Population genomic measures are observed heterozygosity (H_{obs}), expected heterozygosity (H_{exp}), nucleotide diversity (π), allelic richness (A_r), inbreeding coefficient (F_{IS}), genetic differentiation among populations (pairwise F_{ST}), mean genetic distance among individuals corrected for geographic distance (D_{PS} and r per km) with standard errors (S.E.), and effective population size (N_e) with 95% confidence intervals (C.I.) based on parametric bootstrapping.

Region	Area (km²)	N_{gen}	H_{obs}	H_{exp}	π	A_{r}	F_{IS}	F_{ST}	D_{PS}/km (S.E.)	r/km (S.E.)	N _e (95% C.I.)
Western Slope	11889	78 indiv.	0.240	0.272	0.0029	1.93	0.117	0.024	0.28 (0.05)	0.15 (0.03)	69.3 (66.2-72.4)
Front Range	11958	56 indiv.	0.242	0.263	0.0028	1.89	0.084	0.024	0.46 (0.16)	0.24 (0.09)	40.2 (38.7-41.7)

Table 3: Habitat differences between the Western Slope and Front Range of Colorado. Units are percent cover for impervious surface and tree canopy cover; resistance values for land cover, river and stream riparian corridors, and roads; degrees Celsius for temperature; millimeters for precipitation; meters for elevation; and unitless measurements based on chlorophyll reflectance and variance in elevation, respectively, for enhanced vegetation index and topographic roughness.

Landscape Data	Western Slope Front Range						•			
	Min	Max	Median	Mean	Std Dev	Min	Max	Median	Mean	Std Dev
Elevation (m)	1453.5	4362.9	2354	2418.0	552.5	1474.5	4347.1	2365	2374.9	629.3
Tree canopy cover (%)	0	100	20	29.9	31.4	0	100	32	35.0	33.6
Impervious surface (%)	0	100	0	0.5	4.1	0	100	0	4.0	13.5
Minimum temp. coldest month (°C)	-20.2	-9.5	-13.3	-13.9	2.8	-19.9	-8.3	-12.7	-12.6	2.8
Annual precipitation (mm)	208	1137	458	483.3	171.5	359	1006	452	496.4	121.9
Enhanced vegetation index	-1806	8955	4634	4434.9	1858.3	-1969	9132	5416	4957.8	1800.9
Topographic roughness	0	27924.6	11	53.1	129.6	0	20067.0	25	56.2	100.8
Landcover	1	10	1	3.2	3.0	1	10	1	4.2	3.7
Roads	1	10	1	1.7	2.5	1	10	1	2.7	3.6
Riparian	1	10	10	9.4	2.3	1	10	10	9.4	2.3

Table 4: Multiple regression on distance matrices (MRDM) landscape genomic results from the Western Slope and Front Range of Colorado. Response variables were individual-based genetic distances, i.e., proportion of shared alleles (Dps) and relatedness (r). Explanatory variables, after removing correlated variables, were the geographic (Euclidean) distance model (geo. dist.), percent impervious surface cover, percent tree canopy cover, river and stream riparian corridors, roads, and minimum temperature of the coldest month. Forward selection followed by backward elimination was performed, with 1,000 random permutations of the dependent distance matrix per step, using Bonferroni-corrected p-to-enter and p-to-remove alpha values of 0.05. Standardized beta coefficients were used to assess the direction of effect of each landscape variable on gene flow. Only univariate models were supported.

Region	Genetic distance	Landscape factors	Direction of effect	r ²	р
Western Slope	Dps	tree cover	+	0.08	0.001
	r	geo. dist.	-	0.04	0.001
Front Range	Dps	geo. dist.	-	0.05	0.001
	r	geo. dist.	_	0.04	0.001

Table 5: Maximum likelihood of population effects (MLPE) landscape genomic results from the Western Slope and Front Range of Colorado. Response variables were individual-based genetic distances, i.e., proportion of shared alleles (Dps) and relatedness (r). Pairwise comparisons of individuals were controlled as a random effect. Fixed effects, after removing correlated variables, were the geographic (Euclidean) distance model (geo. dist.), percent impervious surface cover, percent tree canopy cover, vegetation density, river and stream riparian corridors, roads, and minimum temperature of the coldest month. Standardized beta coefficients were used to assess the direction of effect of each landscape variable on gene flow. Models reported are within the top 5 BIC units. Landscape factors are in order of standardized beta coefficients (largest to smallest).

Region	Genetic distance	Landscape factors	Direction of effect	r^2	ΔΒΙϹ
Western Slope	Dps	tree cover	+	0.15	0
		min. temperature	=		
		tree cover	+	0.15	0.6
		geo. dist.	-		
		tree cover	+	0.14	3.0
	r	geo. dist.	-	0.17	0
		tree cover	+		
		geo. dist.	-	0.17	3.8
Front Range	Dps	geo. dist.	-	0.12	0
		tree cover	-		
		impervious surface	-		
		geo. dist.	-	0.11	2.9
		tree cover	-		
	r	geo. dist.	-	0.13	0
		tree cover	-		
		impervious surface	-		
		geo. dist.	-	0.13	0.5
		tree cover	-		

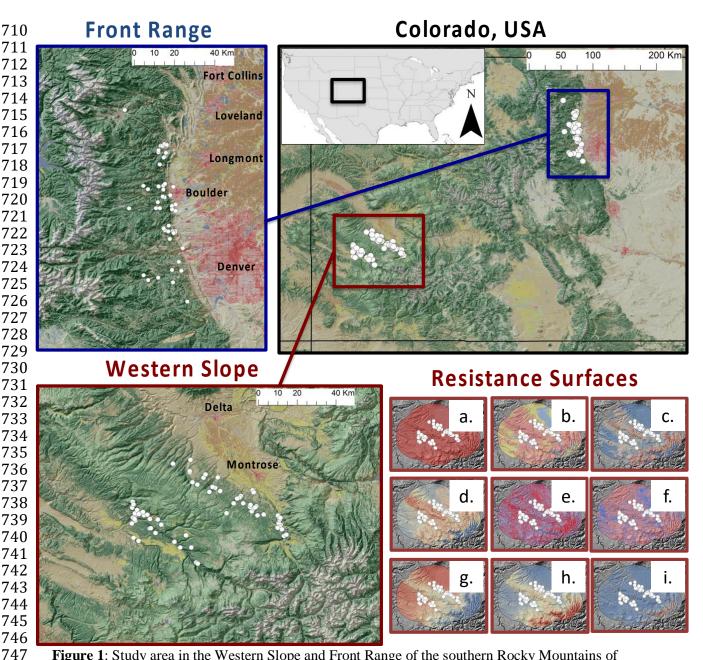


Figure 1: Study area in the Western Slope and Front Range of the southern Rocky Mountains of Colorado, USA. Landscape genomic analyses included 78 pumas from the Western Slope and 56 pumas from the Front Range (white circles). Resistance surfaces, shown for the Western Slope, represent alternative hypotheses of the effects of landscape variables on puma dispersal and gene flow (red=high gene flow, blue=low gene flow) for: (a) percent impervious surface cover (negative effect on gene flow), (b) land cover (forested, open-natural, and developed: positive, neutral, and negative effects on gene flow), (c) percent tree canopy cover (positive effect), (d) vegetation density (positive effect), (e) river and stream riparian corridors (positive effect), (f) roads (negative effect), (g) minimum temperature of the coldest month (negative effect), (h) annual precipitation (positive effect), and (i) topographic roughness (positive effect). We also tested isolation by geographic Euclidean distance. Land cover base maps show forests (green), shrub and grasslands (tan), urban areas (red), agriculture and ranchlands (brown and yellow), and alpine tundra (grey).

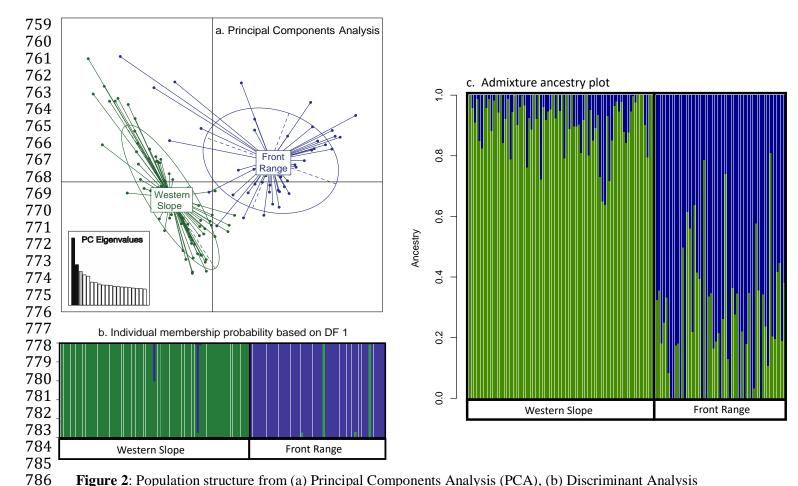


Figure 2: Population structure from (a) Principal Components Analysis (PCA), (b) Discriminant Analysis of Principal Components (DAPC), and (c) Admixture analysis. Individuals assigned to the Western Slope and Front Range are green and blue, respectively. K=2 was most supported in Admixture analysis using cross validation error.