

1 **Title:** Placental DNA methylation signatures of maternal smoking during pregnancy
2 and potential impacts on fetal growth

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47

48 **Competing interests**

49 The authors declare no competing interests.

50

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109 **Abstract**

110 Maternal smoking during pregnancy (MSDP) contributes to poor birth
111 outcomes, in part through disrupted placental functions, which may be reflected in the
112 placental epigenome. We meta-analyzed the associations between MSDP and placental
113 DNA methylation (DNAm) and between DNAm and birth outcomes within the
114 Pregnancy And Childhood Epigenetics (PACE) consortium (7 studies, N=1700, 344
115 with any MSDP). We identified 1224 CpGs that were associated with MSDP, of which
116 341 associated with birth outcomes and 141 associated with gene expression. Only 6 of
117 these CpGs were consistent with the findings from a prior meta-analysis of cord blood
118 DNAm, demonstrating substantial tissue-specific responses to MSDP. The placental
119 MSDP associated CpGs were enriched for growth-factor signaling, hormone activity,
120 inflammation, and vascularization, which play important roles in placental function. We
121 demonstrate links between placental DNAm, MSDP and poor birth outcomes, which
122 may better inform the mechanisms through which MSDP impacts placental function and
123 fetal growth.

124

125 **Keywords:** DNA methylation, epigenetics, EWAS, pregnancy, smoking, birth weight,
126 gestational age

127

128 Almost 1 in 10 pregnancies are impacted by the effects of maternal smoking during
129 pregnancy (MSDP), with state-specific prevalence ranging from as low as 1.8% to as
130 high as 27.1% in the USA¹, while in Europe, the prevalence of MSDP is estimated to
131 range between 4.2% and 18.9%². Consequently, the numerous health effects of MSDP
132 remain a significant public health concern. The impact of this exposure on fetal
133 development has been the source of significant investigation, resulting in MSDP being
134 recognized as a cause of multiple negative pregnancy and birth outcomes³.

135 The mechanisms that underlie this reproductive and developmental toxicity are
136 partially understood and include molecular and anatomical changes of the placenta^{4,5}.
137 Additionally, experimental mouse models have recently highlighted the critical roles of
138 proper placental function in ensuring successful pregnancy outcomes⁶. Epigenetic
139 responses to prenatal exposures have emerged as potential intermediate links between
140 early life exposures and developmental health outcomes, and epidemiologic studies of
141 DNAm, particularly multi-cohort collaborative efforts, are powerful approaches to
142 investigate these types of research questions⁷. So far, most studies of MSDP and
143 epigenetics have focused on DNA methylation (DNAm) variations in cord blood,
144 though some studies of placenta, peripheral blood and lung tissues have also been
145 performed⁸. The Pregnancy and Childhood Epigenetics (PACE) consortium⁹ published
146 a large meta-analysis identifying thousands of MSDP-associated variations in DNAm
147 within cord blood and child peripheral blood¹⁰. However, the placental epigenome has
148 not been as thoroughly studied, though the placenta is likely a critical target organ of
149 MSDP-associated toxicity. A handful of prior studies have examined the relationships
150 between MSDP and (DNAm) in human placenta¹¹⁻¹⁴, identifying MSDP-associated loci
151 some of which have been suggested partially mediate the effects of MSDP on lower
152 birth weight¹⁵.

153 These studies have begun to characterize the impact that MSDP has on the
154 human placental epigenome but have been limited by small sample sizes and have not
155 adjusted for placental tissue heterogeneity. We aimed to address these gaps by
156 performing a fixed effects meta-analysis examining the relationships between MSDP
157 and variations in the placental methylome across seven independent studies that are
158 members of the PACE consortium. We also aimed to gain insights into the potential
159 biological and health-related impacts of these associations by performing additional
160 analyses with nearby mRNA expression, as well as functional, regulatory, and
161 phenotypic enrichment, and a secondary meta-analysis of the associations between
162 placental DNAm and birth outcomes. This study represents the largest and most
163 comprehensive examination of MSDP associations with placental DNAm in human
164 populations and provides novel insights into the placental molecular responses to MSDP
165 and how these relate to fetal development.
166

167 **Results**

168 *Study population*

169 Seven American, Australian, and European studies (N=1,700) contributed to the
170 epigenome-wide association study (EWAS) linking MSDP to placental DNAm:
171 including Asking Questions about Alcohol in pregnancy (AQUA)¹⁶, Study on the pre-
172 and early postnatal determinants of child health and development (EDEN)¹⁷, Genetics of
173 Glucose regulation in Gestation and Growth (Gen3G)¹⁸, Genetics, Early Life
174 Environmental Exposures and Infant Development in Andalusia (GENEIDA),
175 Environment and Childhood Project (INMA)¹⁹, New Hampshire Birth Cohort Study
176 (NHBCS)²⁰ and Rhode Island Child Health Study (RICHS)²¹. For this meta-analysis,
177 344 (20.2%) mothers reported any MSDP, defined as any cigarette smoking during any
178 trimester of pregnancy. Any MSDP tended to be less prevalent in the cohorts from
179 Canada and the USA compared to those from Australia and Europe (Table 1). Three
180 cohorts (N=795, EDEN, GENEIDA and INMA) contributed to the EWAS of sustained
181 MSDP, defined as maternal smoking throughout pregnancy, among which 163 (20.5%)
182 mothers reported sustained MSDP. Distributions of covariates by cohort are provided in
183 the supplementary materials (Excel Table S1).

184 *Genome-wide DNAm meta-analyses*

185 We produced four statistical models for each CpG site, modeling the
186 associations between DNAm with both any and sustained MSDP, with and without
187 adjustment for putative cellular heterogeneity which was estimated with a reference free
188 deconvolution algorithm²². Models were adjusted for maternal age, parity, and maternal
189 education. Genomic inflation factors from the meta-analyses ranged from $\lambda=2.0$ to 2.8
190 (Excel Table S2, Supplemental Figure S1). Any MSDP was associated with DNAm at
191 532 CpG sites after Bonferroni adjustment (Excel Table S3), while sustained MSDP

192 was associated with DNAm at 568 CpG sites (Excel Table S4). After adjusting for
193 putative cellular heterogeneity, any MSDP was associated with 878 CpGs (Excel Table
194 S5), while sustained MSDP was associated with 894 CpGs (Excel Table S6). Among
195 these, 10.7% of models for any MSDP and 3.9% of models for sustained MSDP, not
196 adjusted for cell type, yielded heterogeneity p-values < 0.01 , while only 5.5% of the
197 models for any MSDP and 3.5% for sustained MSDP, adjusted for putative cellular
198 heterogeneity, produced heterogeneity p-values < 0.01 . Thus, heterogeneity in the
199 associations across cohorts was lowest in the analyses that were adjusted for putative
200 cellular heterogeneity and the CpGs from these models that yielded Bonferroni-
201 significant associations (1224 unique CpGs) were carried forward for secondary
202 analyses.

203 Overall, the Bonferroni significant CpGs were distributed throughout the
204 genome, and the majority (68%) of CpGs exhibited lower DNAm in association with
205 any and sustained MSDP (Figure 1). Of the 548 CpGs that yielded Bonferroni
206 significant associations with both any and sustained MSDP, the absolute values of the
207 parameter estimates were greater for the models of sustained MSDP compared to the
208 estimates for any MSDP (Excel Table S7). In a secondary analysis that restricted the
209 any MSDP models to the same three cohorts that contributed to the sustained MSDP
210 models (EDEN, GENEIDA, and INMA), we again found that almost all CpGs (544 out
211 of 548) exhibited increased magnitudes of association with increased duration of
212 exposure (Excel Table S7, Supplemental Figure S2).

213 The most notable association was observed at cg27402634, located upstream of
214 the *LEKRI* gene and the non-coding RNA *LINC00886*, which showed the largest
215 differential DNAm and smallest p-values in all meta-analyses. Placentas that were
216 exposed to any MSDP had 22.95% lower DNAm (95% CI: 21.01-24.99% lower

217 DNAm; p-value = 5.35E-119) and those exposed to sustained MSDP had 25.78% lower
218 DNAm (95% CI: 24.02-27.54% lower DNAm; p-value = 7.22E-181) when compared to
219 mothers that did not smoke at all during pregnancy. Though all cohorts observed
220 substantial hypomethylation with MSDP at this CpG, the actual estimates of the
221 associations were highly variable between cohorts for models of any MSDP
222 (heterogeneity p-value = 2.66E-15), but relatively consistent for models of sustained
223 MSDP (heterogeneity p-value = 1.55E-01) (Figure 2A). Overall, we consistency in the
224 associations across cohorts for the vast majority of the 1224 CpGs that yielded
225 Bonferroni-significant associations via meta-analysis: 92% and 97% of these CpGs
226 yielded heterogeneity p-values > 0.01, for their associations with any and sustained
227 MSDP respectively. In addition to cg27402634, we highlight those relationships that
228 yielded that largest magnitudes of association: ($|\beta_{\text{Any MSDP}}|$) > 0.05 for cg26843110
229 (*EDC3*), cg20340720 (*WBP1L*), and cg17823829 (*KDM5B*) (Figure 2B-2D). We
230 identified numerous other noteworthy relationships but due to the large number of
231 Bonferroni-significant associations from our meta-analyses, we highlight the
232 relationship among the 20 most statistically significant CpGs from the primary meta-
233 analysis of any MSDP going forward (Table 2).

234 ***Expression Quantitative trait methylation (eQTM) analyses***

235 We tested whether the DNAm levels at MSDP-associated CpGs were associated
236 with the expression of nearby (\pm 250kb of the CpG) mRNA from 194 placental samples
237 in the RICHS cohort, and corrected for multiple testing with the less conservative false
238 discovery rate (FDR) due to the smaller sample size. We identified 170 associations
239 between DNAm and gene expression within a 5% FDR (Excel Table S8), including 141
240 unique CpGs and 140 unique mRNAs; 72.4% of the eQTMs exhibited inverse
241 associations, while inverse associations were most common and statistical significance

242 was strongest for CpGs that were closer to the transcription start site (Supplemental
243 Figure S3). Among the top 20 CpGs from our meta-analysis, eight were identified as
244 eQTM (5% FDR) (Table 3). Higher placental DNAm at cg27402634, was associated
245 with lower expression ($\beta = -2.32$, and FDR < 5%) of *LEKRI*. Additionally, DNAm at
246 cg17823829 (annotated to *KDM5B*) was most strongly associated lower expression of
247 *PPFIA4* (FDR < 5%), and DNAm at cg26843110 (annotated to *EDC3*) was nominally
248 associated with higher expression of *CSK*, while cg20340720 (annotated to *WBPIL*)
249 was not associated with any nearby mRNA expression.

250 ***Functional and regulatory enrichment analyses***

251 Enrichment analyses were performed to gain insights into which biological
252 processes may be impacted by these MSDP-associated CpGs. We performed gene-set
253 enrichment analyses, which included 565 genes annotated to CpGs associated with any
254 MSDP and 581 genes for sustained MSDP. 143 and 25 pathways were significantly (q-
255 value < 0.05) enriched among CpGs associated with any MSDP (Excel Table S9) and
256 sustained MSDP (Excel Table S10), respectively. In both gene-sets, “signaling by nerve
257 growth factor (NGF)” was the most significant pathway (q-value = 2E-04 for any
258 MSDP and q-value = 9.6E-03 for sustained MSDP). Other significantly enriched
259 pathways were related to growth factors (VEGF, EGF, PDGF, IGF1R), hormones
260 (aldosterone, insulin, TSH, GnRH), interleukins (IL2, IL4, IL7), myometrial
261 contraction, vascular smooth muscle contraction, thrombin and platelet activation,
262 signaling and aggregation. We also tested whether the genes annotated to MSDP-
263 associated CpGs were enriched for regulatory regions of specific transcription factors
264 (TFs). Most notably, our MSDP-associated CpGs were enriched for genes regulated by
265 GATA1 and GATA2, as well as by RUNX1, TP63, SMAD4, AR, TP53 or PPARG
266 (Excel Tables S11-S12).

267 We then examined whether the MSDP-associated CpG sites were enriched for
268 CpG island locations, allele-specific germline differentially methylated regions
269 (gDMR)²³, regulatory features from the placenta specific 15-chromatin state annotation
270 from ROADMAP²⁴, or placenta specific partially methylated domains (PMD)²⁵, which
271 contain placenta-specific repressed genes (annotated to the results files in
272 Supplementary Tables S3-S6). The MSDP-associated CpGs were depleted in PMDs
273 (Supplemental Figure S4), highly enriched in placenta enhancers and depleted in
274 transcription start sites and inactive states (Supplemental Figure S5). Additionally,
275 MSDP-associated CpGs were depleted in CpG islands and shores, while enriched in
276 CpG island shelves and open sea positions (Supplemental Figure S6). We identified 3
277 CpGs (cg05211790 and cg16360861 at *RAI14*, and cg05575921 at *AHRR* gene) that
278 were within two candidate maternal gDMRs, but our overall set of MSDP-associated
279 CpGs were neither depleted nor enriched for confirmed gDMRs (Supplemental Figure
280 S7).

281 ***Phenotype enrichment analyses***

282 The genes annotated to our MSDP-associated CpGs were tested for phenotype
283 enrichment using data from the database of Genotypes and Phenotypes (dbGAP). Our
284 MSDP-associated genes were enriched for numerous phenotypes in dbGAP, including
285 several adiposity phenotypes (body mass index (BMI), waist-hip ratio, and obesity),
286 blood pressure, cardiovascular diseases, type 2 diabetes, asthma and respiratory function
287 (Excel Tables S13-S14).

288 ***Proximity to genetic variants for birth outcomes***

289 We aimed to understand whether the MSDP-associated CpGs that we identified
290 co-localize within the same genomic regions as genetic variants that have been
291 associated with birth outcomes via genome-wide association studies (GWAS). Thus we

292 investigated whether MSDP-associated CpGs were within ± 0.5 Mb (1 Mb window) of
293 single nucleotide polymorphisms (SNPs) that have been associated with birth weight
294 (BW), birth length (BL), head circumference (HC) and gestational age (GA)²⁶⁻³¹, which
295 have been added as annotations to the meta-analysis results files (Excel Tables S3-S6).
296 Of the 324 birth outcome SNPs in autosomal chromosomes, 94 SNPs (83 loci) and 108
297 (97 loci) were within 0.5 Mb of CpGs that were associated with any or sustained
298 MSDP, respectively (Excel Tables S15). Overall ~16% of the 1224 MSDP-associated
299 CpG sites were within 0.5 Mb of birth outcome SNPs, including cg27402634 (*LEKRI*),
300 cg26843110 (*EDC3*) and cg20340720 (*WBPIL*). We also explored whether our MSDP-
301 associated CpGs may be biased by methylation quantitative trait loci (mQTLs), in
302 which SNPs influence the methylation levels at nearby CpGs. Two studies have
303 examined this question in human placenta, identifying 866³² and 4,342³³ placental
304 mQTLs. Our findings did not appear to be biased by genetic variation as only 9 of the
305 1,224 MSDP-associated CpGs are previously characterized placental mQTLs.

306 ***Association of DNAM at smoking associated loci and smoking related birth outcomes***

307 We also performed a second meta-analysis to examine the relationships between
308 DNAM with gestational age at birth, preterm birth, BW, BL, and HC z-scores, outcomes
309 that are known to be related to maternal smoking. Of the 1224 CpGs tested, 341
310 (27.9%) were related to at least one birth outcome after Bonferroni-adjustment
311 (0.05/1224). The majority of birth outcome associations were related to gestational age
312 at birth (298 CpGs) (Excel Table S16). Preterm delivery, for which only two cohorts
313 could contribute data (EDEN and NHBCS), produced similar associations found for
314 GA, though fewer CpGs were statistically significant (Excel Table S17). We also found
315 that numerous loci were associated birth size z-scores, with the majority of these being
316 associated with BW (43 CpGs) (Excel Table S18), followed by BL (20 CpGs) (Excel

317 Table S19) and HC (4 CpGs) (Excel Table S20). Some of the CpGs associated with GA
318 were also associated with birth size measurements, even though they were standardized
319 for GA, suggesting independent associations with both gestational duration and fetal
320 growth (Supplemental Figure S8A), including 6 CpGs (annotated to *SYNJ2*, *PXN*,
321 *PTPRE*, *IGF2BP2*, 4q21.1) shared between GA and BW, and 2 (annotated to *POLR3E*
322 and *LOC441869*) shared between GA and BL. Among the CpGs that yielded at least
323 one Bonferroni-significant association with birth outcomes, CpGs that tended to have
324 positive associations with birth outcomes clustered together and were typically
325 hypomethylated with MSDP, while CpGs that exhibited inverse associations with birth
326 outcomes tended to be hypermethylated with exposure to MSDP (Supplemental Figure
327 S8B).

328 Among our top 20 CpGs that were associated with any MSDP, 5 were associated
329 with GA at birth and 4 were associated with BW z-scores after Bonferroni adjustment
330 (Table 3). DNAm at cg27402634 (*LEKRI*) and cg20340720 (*WBPIL*), both located
331 close to BW-SNPs and for which MSDP associated with lower DNAm, were associated
332 with larger BW (p-value = 6.71E-07, and p-value = 2.42E-07). On the other hand,
333 DNAm at cg26843110 (*EDC3*; hypomethylated in response to MDSP and also close to
334 BW-SNPs) and at cg17823829 (*KDM5B*; hypermethylated) were associated with longer
335 and shorter gestational ages at birth, respectively (p-value = 5.09E-12, and p-value =
336 9.12E-06). Forest plots of BW z-scores and GA for these four CpGs are shown in
337 Figure 3.

338 We summarize the results for all secondary analyses with a circos plot, for those
339 548 CpGs that yielded Bonferroni significant associations with both any and sustained
340 MSDP (Figure 4). Among these, 21 CpGs were associated both with mRNA expression

341 (FDR < 5%) and at least one birth outcome (Bonferroni-significant), which have been
342 annotated with the gene symbols from their respective eQTM models.

343 ***Comparison with smoking-sensitive CpGs in cord blood***

344 We assessed whether the DNAm signatures of MSDP in the placenta were
345 consistent with MSDP associations in cord blood previously reported by the PACE
346 consortium¹⁰. Only nine CpGs annotated to seven unique genes (*AHRR*, *CYP1A1*,
347 *GNG12*, *PXN*, *RNF122*, *SLC23A2*, and *ZBTB4*) yielded Bonferroni-significant
348 associations in both tissues, out of 1224 CpGs from our study and 568 CpGs from the
349 cord blood study (Table 4). Of note, the CpGs within *CYP1A1* and *RNF122* showed
350 opposite directions of association with MSDP in cord blood and placenta. We also
351 compared the parameter estimates from our study that yielded associations with MSDP
352 within 5% FDR to the parameter estimates of those CpGs described in cord blood also
353 within a 5% FDR¹⁰. There was no overall correlation ($r^2 < 0.1$) of the regression
354 coefficients across these two tissues (Supplemental Figure S9).

355

356 **Discussion**

357 We identified 1224 CpG sites with placental methylation levels that were
358 associated with any or sustained MSDP. Differential DNAm was greater with increased
359 duration of exposure at all loci that were associated with both any and sustained MSDP,
360 and a large proportion of the MSDP-associated CpGs were related to birth outcomes.
361 Those CpGs that were observed to have higher DNAm associated with MSDP, tended
362 to be inversely associated with gestational age and birth size, while CpGs exhibiting
363 lower DNAm with MSDP tended to be positively associated with gestational age and
364 birth size.

365 The MSDP-associated loci with the most statistically significant association
366 (cg27402634), also identified in prior smoking EWAS of placental tissues¹⁵, yielded
367 dramatically lower DNAm levels in association with MSDP exposure. This effect-size
368 is much larger in magnitude (~25% difference for sustained MSDP) compared to what
369 has generally been observed in most exposure-focused EWAS, though within the same
370 range as a CpG site in *AHRR* (cg05575921; 18% difference between exposed and
371 unexposed) from a prior EWAS of current smoking and blood DNAm³⁴. Additionally,
372 decreased placental DNAm at cg27402634 correlates with increased expression of
373 *LEKRI*, and associates with smaller BW and BL. Thus MSDP-associated
374 hypomethylation at this CpG would be consistent with the well-known effect of
375 maternal smoking resulting in shorter gestation and smaller birth size.

376 The functional activities of cg27402634, or corresponding *LEKRI* gene, in
377 human placental tissues are not known. However, GWAS findings provide evidence
378 that genetic variants within this region might be involved in fetal growth and possibly
379 metabolic programming. For instance, the SNP rs1482852 or its proxies (rs900400;
380 rs13322435) have been associated with different parameters of fetal growth³⁵, adiposity

381 in newborns^{36,37}, maternal adiponectin levels, cord blood leptin³⁷, and insulin release
382 after an oral glucose challenge³⁸. These findings from genetic studies in combination
383 with our current study, implicate that this locus on chromosome 3 (3q25.31) contains
384 very active determinants of growth regulation and metabolic activity, and that placental
385 DNAm at cg27402634 is highly responsive to maternal smoking. Future mechanistic
386 work is necessary to investigate whether the placental epigenetic regulation at this locus
387 specifically influences placental functions and/or overall growth and metabolic
388 functions of the developing fetus.

389 We identified numerous other notable MSDP-associated loci in addition to those
390 of cg27402634 (*LEKRI*), and highlight those CpGs yielding the strongest magnitudes of
391 effect (cg20340720, cg26843110, and cg17823829). MSDP was associated with lower
392 DNAm at cg20340720, located within *WBPIL* (also annotated as *C10orf26*), while
393 lower DNAm at this CpG correlated with lower with BW and BL z-scores. Genetic
394 variants nearby to this CpG have been related to BW³¹ and blood pressure³⁹. We also
395 observed lower DNAm with MSDP at cg26843110, which is within the body of the
396 *EDC3* gene, and is nearby to SNPs associated with BW (rs3784789³¹). Lower DNAm at
397 cg26843110 associated with shorter GA at birth, and decreased expression of *CSK*,
398 which is involved in trophoblast differentiation⁴⁰ as well as blood pressure and
399 aldosterone regulation⁴¹. Finally, cg17823829 (annotated to *KDM5B*) was
400 hypermethylated with MSDP. Higher DNAm at this CpG correlated with shorter GA at
401 birth and with lower expression of *PPFIA4* gene, which can be induced in response to
402 hypoxia⁴².

403 Our enrichment analyses identified numerous pathways that are critical to
404 placental growth and development, such as vascularization, hormone signaling and
405 inflammatory cytokines. Multiple pathways involving vascular endothelial growth

406 factors (VEGF) and nerve growth factors (NGF) populated the top of our enrichment
407 lists. The VEGFs and their receptors are required for all steps of placental
408 vascularization⁴³, while nerve growth factor (NGF) modulates immune activity,
409 inflammation and angiogenesis in the placenta⁴⁴. Thus, our findings may be related to
410 perturbed placental vascularization or angiogenic signaling. Altered placental
411 vasculature is the most common placental pathology identified in pregnancy
412 complications⁴³, and MSDP can result in placental vascular remodeling⁵. Our findings
413 may represent, in part, an epigenetic footprint of the placental vascular and angiogenic
414 response to MSDP. Our CpGs were also enriched for genes regulated by specific
415 transcription factors, most notably GATA1 and GATA2. Together with PPARG and
416 TP63, GATA factors are part of the core transcriptional regulatory circuit that guides
417 and maintains proper trophoblast differentiation^{45,46}. Placentas lacking *PPARG* have
418 lethal defects in placental vascularisation⁴³, and angiogenic activity is reduced in
419 placentas lacking *GATA2*⁴⁷. Additionally, our MSDP-associated CpGs were enriched in
420 placental enhancers, while depleted in transcription start sites, inactive states, PMDs²⁵
421 and gDMRs²³, overall suggesting that the CpG sites that we identified are located in
422 active regulatory regions.

423 Genes that are annotated to the CpGs that we studied have been linked to human
424 health and disease traits via dbGAP, including a number of conditions that are part of
425 the metabolic syndrome (BMI, obesity, cholesterol, blood pressure), for which prior
426 links to MSDP have been reported^{48,49}. This may indicate that the placental genes whose
427 regulation is impacted by MSDP, are involved in energy uptake and expenditure, lipid
428 and glucose metabolism, blood pressure regulation, and inflammation, which are some
429 of the key physiological processes that are disrupted in the pathogenesis of metabolic
430 syndrome⁵⁰. The MSDP-associated CpGs were also enriched for genes linked to asthma

431 and impaired respiratory function, which are known to be caused by MSDP⁵¹, but it is
432 unclear if altered regulation of these genes in the placenta has consequences on the
433 development of the respiratory system. Furthermore, 128 of the 324 SNPs that have
434 previously been associated with birth size or gestational age at birth²⁶⁻³¹ were in similar
435 genomic proximity (within 0.5 Mb) to our CpGs, suggesting the MSDP-associated
436 differential methylation in the placenta occurs in regions of the genome that are heavily
437 involved in growth and development. Many of these SNPs have been shown to be
438 related to birth outcomes, glycemic traits, blood pressure, and height, while the largest
439 GWAS of BW to date concludes that the link between lower BW and later cardio-
440 metabolic traits are largely driven by shared genetic effects³¹. Future research is needed
441 to characterize this convergence of genetic effects on growth and placental epigenetic
442 responsiveness to MSDP within similar genomic proximities. It is possible that MSDP,
443 genetic variation, and placental DNAm in these regions yield additive or interactive
444 effects on birth outcomes; prior studies have addressed this question in blood^{52,53}.
445 Additionally, while our MSDP-associated CpGs are in similar genomic regions (1 Mb
446 windows) with birth outcome SNPs, very few of our CpGs (9 of 1224) and very few of
447 the birth outcome SNPs (3 of 324) are known placental mQTLs^{32,33}.

448 We compared our findings to those of a previous PACE meta-analysis of MSDP
449 and cord blood DNAm¹⁰. Two of the CpGs that we identified within the *AHRR* gene
450 (cg05575921 an eQTM for *AHRR* in placenta, and cg21161138), have been consistently
451 observed to be sensors of MSDP exposure in cord blood. Only four other CpG sites,
452 (annotated to *GNG12*, *PXN*, *ZBTB4*, and *SLC23A2*) were differentially methylated in
453 both placenta and cord blood with the same direction of association in both tissues.
454 Additionally, three CpG sites within *CYP1A1* and *RNF122* were identified in both meta-
455 analyses but with different directions of association in cord blood versus placenta.

456 Interestingly, we observed *CYP11A1* to be hypomethylated in placenta with exposure to
457 MSDP, which is consistent with studies of adipose, skin, and lung tissues⁵⁴, but this
458 CpG was hypermethylated in the cord blood meta-analysis¹⁰. Additionally, the most
459 statistically significant association with MSDP in placenta (cg27402634, *LEKRI*) was
460 not associated with MSDP in the cord blood meta-analysis. These observations, and the
461 lack of overall correlation in regression coefficients when comparing placental and cord
462 blood responses to MSDP, suggest that there are unique tissue-specific molecular
463 responses to this exposure.

464 The above findings should be interpreted within the context this study's
465 limitations. MSDP was self-reported and subject to misclassification, though differential
466 misclassification likely would have biased our findings towards the null. We modeled
467 two different definitions for MSDP and found that the models with sustained MSDP
468 produced larger magnitudes of association, as has been previously found in studies of
469 blood DNAm⁵⁵. Although these findings suggest that increased duration of MSDP is
470 associated with greater differential DNAm, we did not assess dose-response patterns (ie.
471 number of cigarettes or cotinine concentrations), which should be the focus of future
472 investigations. Our study predominantly consisted of samples from mother-infant pairs
473 of European ancestry, and thus additional studies involving diverse racial and ethnic
474 backgrounds are needed in order to improve the generalizability of these findings.
475 While it is unlikely that placental DNAm would influence maternal smoking, the
476 observed associations between DNAm levels and reproductive outcomes could be due
477 to reverse causation. Placenta is a heterogeneous tissue with multiple different cell
478 types⁵⁶ that serve different functions and thus have different epigenetic states⁵⁷. To
479 correct for this, we estimated and adjusted for variability in placental DNAm that may
480 be due to tissue heterogeneity. We utilized a data driven approach that was not based on

481 a methylome reference, as no references for placental cell-type methylomes are
482 currently available. Adjustments for these estimates of putative cellular admixtures did
483 reduce heterogeneity in the meta-analyses, thus improving the consistency of observed
484 associations between MSDP and DNAm across these independent cohorts. However, it
485 is possible that residual confounding may have influenced some of our results.

486 Despite these limitations, our study had numerous strengths, including a large
487 overall sample-size and seven independent studies to identify these relationships. We
488 used harmonized definitions of exposure variables and covariates, standardized
489 protocols for quality control and pre-processing of DNAm data, and standardized
490 methods for estimating/adjusting for tissue heterogeneity and for statistical analyses.
491 We also performed secondary analyses involving mRNA expression, functional and
492 phenotype enrichment, overlap with GWAS hits for reproductive outcomes, and meta-
493 analyses of DNAm variation with birth outcomes to provide biological and health-
494 related interpretations of our findings.

495 We identified a DNAm signature of MSDP in the placenta that shows substantial
496 differences from that observed in cord blood, most notably the CpG in close proximity
497 to *LEKRI*. Many of the identified MSDP-associated loci are involved in biological
498 process that are known to play critical roles in placental development, including
499 vascularization, angiogenesis, and inflammation. Additionally, a large proportion of the
500 MSDP-associated CpGs were also associated with GA at birth, or birth size z-scores,
501 suggesting these placental epigenetic variations may be intermediate molecular markers
502 between MSDP and these outcomes. Further study is required to determine whether
503 these epigenetic variations are causal mediators within these relationships or reflecting
504 other processes.

505

507 **Material and methods**

508 *Participating cohorts*

509 Cohorts that are members of the PACE consortium were identified for
510 participation in the current study if they had existing DNAm data quantified from
511 placental tissue via the Illumina Infinium HumanMethylation450 BeadChip and if they
512 had obtained information on self-reported smoking during pregnancy. The seven
513 cohorts that contributed to the meta-analysis of any MSDP included AQUA¹⁶, EDEN¹⁷,
514 Gen3G¹⁸, GENEIDA, INMA¹⁹, NHBCS²⁰ and RICHS²¹. EDEN, GENEIDA and INMA
515 also contributed to the sustained MSDP stratified analyses. RICHS contributed RNAseq
516 data for analyses with mRNA expression. All cohorts acquired ethics approval and
517 informed consent from participants prior to data collection through local ethics
518 committees. Exclusion criteria for this study were: non-singleton births, pre-eclampsia
519 and DNAm not assessed in the fetal side of the placenta. All participants in the study
520 were of European ancestry, except 1.85% of EDEN mothers. Detailed methods for each
521 cohort are provided in the Supplementary Material (Supplemental Methods File).

522 *Tobacco smoking definitions*

523 Any MSDP was defined as “yes” if mothers reported smoking cigarettes at any
524 time during pregnancy. Sustained MSDP was defined as “yes” when mothers reported
525 smoking cigarettes at least in the 1st and 3rd trimester of pregnancy. For both exposure
526 variables, the comparison group was defined as the mothers that reported no smoking
527 during any of the pregnancy.

528 *Placental genome-wide DNAm data acquisition, quality control and normalization*

529 Placental DNAm from the fetal side was assessed with the Infinium Human-
530 Methylation450 array (Illumina, San Diego, CA USA). See Supplementary Methods file
531 for extra details on placenta collection, DNA extraction and DNAm acquisition in each

532 cohort. Quality control of DNAm was standardized across all cohorts. Low quality
533 samples were filtered out and probes with detection p-values > 0.01 were excluded.
534 Beta-values were normalized via functional normalization⁵⁸ and beta-mixture quantile
535 normalization (BMIQ)⁵⁹ was applied to correct for the probe type bias. Cohorts applied
536 ComBat to remove batch effects when applicable. Probes that hybridize to the X/Y
537 chromosomes, cross-hybridizing probes and probes with SNPs at the CpG site,
538 extension site, or within 10 bp of the extension site with an average minor allele
539 frequency > 0.01 were filtered out⁶⁰. Overall, 418,639 probes and 415,396 were
540 available for modelling any MSDP and sustained MSDP, respectively. Finally, DNAm
541 extreme outliers ($<25^{\text{th}}$ percentile $- 3*\text{IQR}$ or $>75^{\text{th}}$ percentile $+ 3*\text{IQR}$ across all the
542 samples) were trimmed.

543 *Estimates of putative cellular heterogeneity*

544 Placental putative cellular heterogeneity was estimated from DNAm data using a
545 reference-free cell-mixture decomposition method⁶¹. The number of surrogate variables
546 ranged from 2 to 5 depending on the cohort. Models for differential DNAm were
547 corrected for the number of surrogate variables minus one to reduce multi-collinearity.

548 *Genome-wide differential DNAM analyses*

549 Within each cohort, robust linear regression from the “MASS” package⁶² in R
550 were used to account for potential heteroskedasticity while testing the associations
551 between normalized DNAm beta values at each CpG with any MSDP and sustained
552 MSDP. Models were adjusted for maternal age, parity, maternal education and cohort-
553 specific variables first unadjusted for putative cellular heterogeneity then adjusted for
554 cellular heterogeneity. We performed inverse variance-weighted fixed-effects meta-
555 analyses using METAL⁶³. The meta-analysis was performed independently by two
556 groups to ensure consistent results. CpGs not retained in at least 2 cohorts were filtered

557 out. We used the Bonferroni adjustment to control for multiple testing. To examine
558 whether increased duration of exposure (sustained smoking versus any smoking)
559 yielded increased magnitudes of association, we calculated the percent change in the
560 coefficients between the two models ($|\beta_{\text{Sustained}}| - |\beta_{\text{Any}}|/|\beta_{\text{Any}}| * 100$). Secondary analyses
561 were only performed on CpGs that yielded Bonferroni significant associations with any
562 or sustained MSDP in models that were adjusted for putative cellular heterogeneity.

563 *Expression quantitative trait methylation (eQTM) loci*

564 We performed expression quantitative trait methylation (eQTM)⁶⁴ analyses in
565 the RICHS cohort. Transcription was measured via RNA-seq on 194 placentas. The
566 details of sample collection, assay, and QC for the RNA-seq data are presented in detail
567 elsewhere⁶⁵, and summarized in the Supplementary Material (Supplementary Methods
568 File). In this dataset, we identified 6523 unique transcripts annotated to an Ensembl ID
569 (GrCh37/hg19) and with a transcriptional start site (TSS) within 250 kb upstream or
570 downstream of 1184 out of the 1224 candidate CpGs. The association between DNAm
571 and expression levels was assessed via 10295 linear regression models using the MEAL
572 package⁶⁶ in R. We report the results for all models yielding nominally significant
573 associations (raw p-values <0.05), statistically significant eQTMs were determined at
574 5% FDR.

575 *CpG site annotation*

576 We annotated CpGs to genes and CpG islands with notations from the Illumina
577 HumanMethylation 450K annotation file, and with several regulatory features using
578 publicly available data: placental 15-chromatin states⁶⁷ released from the ROADMAP
579 Epigenomics Mapping Consortium²⁴ (ChromHMM v1.10), placental partially
580 methylated domains (PMDs)²⁵ and placental germline differentially methylated regions
581 (gDRMs)²³.

582 ***Enrichment analyses***

583 Functional enrichment analyses were performed at the gene level via
584 ConsensusPathDB⁶⁸ using KEGG, Reactome, Wikipathways, Biocarta as reference
585 gene-sets. ConsensusPathDB performs a hypergeometric test and corrects multiple-
586 testing with FDR. Enrichment for transcription factors and for phenotypes were
587 assessed at the gene level with EnrichR using ENCODE and ChEA consensus TFs from
588 ChIP-X database, and dbGaP database, respectively. EnrichR results were ranked using
589 the combined score (P-value computed using Fisher exact test combined with the z-
590 score of the deviation from the expected rank).⁶⁹ Enrichment for regulatory features was
591 assessed with the hypergeometric test, and P-values were Bonferroni-corrected for 15
592 (placental chromatin 15-states) and 6 (relation to CpG island) tests, respectively.

593 ***Overlap of MSDP-sensitive CpG sites and birth outcome SNPs***

594 Co-localization between MSDP-associated CpGs in placenta with previously
595 identified BW, BL, HC and GA SNPs from the largest genome-wide association studies
596 (GWAS) to date²⁶⁻³¹ was assessed using the GenomicRanges package in R⁷⁰. We
597 identified which CpGs were located within 1 Mb windows (± 0.5 Mb) surrounding each
598 of the 324 autosomal SNPs, which correspond to 280 potential unique loci. Unique loci
599 were defined based on the criteria in Warrington et al. 2019³¹, and linkage
600 disequilibrium in Europeans ($r^2 > 0.1$ in < 2 Mb).

601 ***Association between DNAm and birth outcomes***

602 Within each cohort, robust linear regression models were utilized to test the
603 association between normalized DNAm beta values at each CpG as the independent
604 variable and gestational age at birth (inverse normal transformation of sex residuals),
605 BW z-scores, BL z-scores, and HC z-scores as the dependent variables. Logistic
606 regression was used to examine the relationships between DNAm and pre-term birth

607 (defined as <37 weeks of gestation). Birth size z-scores were calculated using
608 international references from the INTERGROWTH-21st Project⁷¹ and standardized by
609 both gestational age and newborn sex. Models were adjusted for maternal age, parity,
610 maternal education, cohort-specific variables (see Supplemental Methods) and putative
611 cellular heterogeneity. Inverse variance-weighted fixed-effects meta-analyses⁶³ were
612 again used to estimate pooled associations. Multiple testing was controlled with the
613 Bonferroni adjustment ($\alpha = 0.05/1224$).

614 *Comparison of MSDP-sensitive CpGs in placenta with cord blood*

615 We examined the consistency between MSDP-sensitive CpGs in placenta and in
616 cord blood¹⁰. First we compared the coefficients from the models for sustained MSDP
617 in cord blood, unadjusted for cellular heterogeneity, to results for both any and
618 sustained MSDP in placenta, adjusted for cellular heterogeneity, using Pearson
619 correlation coefficients.

620 All DNAm data processing and analyses were conducted in R, with the
621 exception of the meta-analyses which were performed with METAL.

622

623

624 **References**

- 625 1. Curtin, S. C. & Mathews, T. J. Smoking prevalence and cessation before and
626 during pregnancy: data from the birth certificate, 2014. *Natl. Vital Stat. Reports*
627 **65**, (2016).
- 628 2. Smedberg, J., Lupattelli, A., Mårdby, A.-C. & Nordeng, H. Characteristics of
629 women who continue smoking during pregnancy: a cross-sectional study of
630 pregnant women and new mothers in 15 European countries. *BMC Pregnancy*
631 *Childbirth* **14**, (2014).
- 632 3. United States Department of Health and Human Services. The Health
633 Consequences of Smoking—50 Years of Progress A Report of the Surgeon
634 General. *A Rep. Surg. Gen.* 1081 (2014). doi:NBK179276
- 635 4. Jauniaux, E. & Burton, G. J. Morphological and biological effects of maternal
636 exposure to tobacco smoke on the feto-placental unit. *Early Hum. Dev.* **83**, 699–
637 706 (2007).
- 638 5. Zdravkovic, T., Genbacev, O., McMaster, M. T. & Fisher, S. J. The Adverse
639 Effects of Maternal Smoking on the Human Placenta: A Review. *Placenta* **26**,
640 S81–S86 (2005).
- 641 6. Perez-Garcia, V. *et al.* Placentation defects are highly prevalent in embryonic
642 lethal mouse mutants. *Nature* **555**, 463–468 (2018).
- 643 7. Felix, J. F. & Cecil, C. A. M. Population DNA methylation studies in the
644 Developmental Origins of Health and Disease (DOHaD) framework. *J. Dev.*
645 *Orig. Health Dis.* (2018). doi:10.1017/S2040174418000442
- 646 8. Richmond, R. C. & Joubert, B. R. Contrasting the effects of intra-uterine
647 smoking and one-carbon micronutrient exposures on offspring DNA methylation.
648 *Epigenomics* **9**, 351–367 (2017).

- 649 9. Felix, J. F. *et al.* Cohort profile: Pregnancy and childhood epigenetics (PACE)
650 consortium. *Int. J. Epidemiol.* **47**, 22-23u (2018).
- 651 10. Joubert, B. R. *et al.* DNA Methylation in Newborns and Maternal Smoking in
652 Pregnancy: Genome-wide Consortium Meta-analysis. *Am. J. Hum. Genet.* **98**,
653 680–696 (2016).
- 654 11. Wilhelm-Benartzi, C. S. *et al.* In Utero Exposures, Infant Growth, and DNA
655 Methylation of Repetitive Elements and Developmentally Related Genes in
656 Human Placenta. *Environ. Health Perspect.* **120**, 296–302 (2012).
- 657 12. Chhabra, D. *et al.* Fetal lung and placental methylation is associated with in utero
658 nicotine exposure. *Epigenetics* **9**, 1473–1484 (2014).
- 659 13. Maccani, J. Z. J., Koestler, D. C., Houseman, E. A., Marsit, C. J. & Kelsey, K. T.
660 Placental DNA methylation alterations associated with maternal tobacco smoking
661 at the RUNX3 gene are also associated with gestational age. *Epigenomics* **5**,
662 619–630 (2013).
- 663 14. Suter, M. *et al.* Maternal tobacco use modestly alters correlated epigenome-wide
664 placental DNA methylation and gene expression. *Epigenetics* **6**, 1284–1294
665 (2011).
- 666 15. Morales, E. *et al.* Genome-wide DNA methylation study in human placenta
667 identifies novel loci associated with maternal smoking during pregnancy. *Int. J.*
668 *Epidemiol.* **45**, 1644–1655 (2016).
- 669 16. Muggli, E. *et al.* Study protocol: Asking QUestions about Alcohol in pregnancy
670 (AQUA): A longitudinal cohort study of fetal effects of low to moderate alcohol
671 exposure. *BMC Pregnancy Childbirth* **14**, (2014).
- 672 17. Heude, B. *et al.* Cohort Profile: The EDEN mother-child cohort on the prenatal
673 and early postnatal determinants of child health and development. *Int. J.*

- 674 *Epidemiol.* **45**, 353–363 (2016).
- 675 18. Guillemette, L. *et al.* Genetics of Glucose regulation in Gestation and Growth
676 (Gen3G): a prospective prebirth cohort of mother–child pairs in Sherbrooke,
677 Canada. *BMJ Open* **6**, e010031 (2016).
- 678 19. Guxens, M. *et al.* Cohort Profile: The INMA—Infancia y Medio Ambiente—
679 (Environment and Childhood) Project. *Int. J. Epidemiol.* **41**, 930–940 (2012).
- 680 20. Gilbert-diamond, D., Emond, J. A., Baker, E. R., Korrick, S. A. & Karagas, M.
681 R. Relation between in Utero Arsenic Exposure and Birth Outcomes in a Cohort
682 of Mothers and Their Newborns from New Hampshire. *Environ. Health*
683 *Perspect.* **124**, 1299–1307 (2016).
- 684 21. Appleton, A. A. *et al.* Prenatal Programming of Infant Neurobehaviour in a
685 Healthy Population. *Paediatr. Perinat. Epidemiol.* **30**, 367–375 (2016).
- 686 22. Houseman, E. A. *et al.* Reference-free deconvolution of DNA methylation data
687 and mediation by cell composition effects. *BMC Bioinformatics* **17**, 259 (2016).
- 688 23. Hamada, H. *et al.* Allele-Specific Methylome and Transcriptome Analysis
689 Reveals Widespread Imprinting in the Human Placenta. *Am. J. Hum. Genet.* **99**,
690 1045–1058 (2016).
- 691 24. Roadmap Epigenomics Consortium *et al.* Integrative analysis of 111 reference
692 human epigenomes. *Nature* **518**, 317–329 (2015).
- 693 25. Schroeder, D. I. *et al.* The human placenta methylome. *Proc. Natl. Acad. Sci.*
694 **110**, 6037–6042 (2013).
- 695 26. Horikoshi, M. *et al.* Genome-wide associations for birth weight and correlations
696 with adult disease. *Nature* **538**, 248–252 (2016).
- 697 27. Beaumont, R. N. *et al.* Genome-wide association study of offspring birth weight
698 in 86 577 women identifies five novel loci and highlights maternal genetic

- 699 effects that are independent of fetal genetics. *Hum. Mol. Genet.* **27**, 742–756
700 (2018).
- 701 28. van der Valk, R. J. P. *et al.* A novel common variant in DCST2 is associated with
702 length in early life and height in adulthood. *Hum. Mol. Genet.* **24**, 1155–1168
703 (2015).
- 704 29. Taal, H. *et al.* Common variants at 12q15 and 12q24 are associated with infant
705 head circumference. *Nat. Genet.* **44**, 532–538 (2012).
- 706 30. Zhang, G. *et al.* Genetic Associations with Gestational Duration and Spontaneous
707 Preterm Birth. *N. Engl. J. Med.* **377**, 1156–1167 (2017).
- 708 31. Warrington, N. M. *et al.* Maternal and fetal genetic effects on birth weight and
709 their relevance to cardio-metabolic risk factors. *Nat. Genet.* **51**, 804–814 (2019).
- 710 32. Do, C. *et al.* Mechanisms and Disease Associations of Haplotype-Dependent
711 Allele-Specific DNA Methylation. *Am. J. Hum. Genet.* **98**, 934–955 (2016).
- 712 33. Delahaye, F. *et al.* Genetic variants influence on the placenta regulatory
713 landscape. *PLOS Comput. Biol.* **14**, e1007785 (2018).
- 714 34. Joehanes, R. *et al.* Epigenetic Signatures of Cigarette Smoking. *Circ Cardiovasc*
715 *Genet* **9**, 436–447 (2016).
- 716 35. Mook-Kanamori, D. O. *et al.* Variants near CCNL1/LEKR1 and in ADCY5 and
717 fetal growth characteristics in different trimesters. *J. Clin. Endocrinol. Metab.* **96**,
718 E810-5 (2011).
- 719 36. Urbanek, M. *et al.* The chromosome 3q25 genomic region is associated with
720 measures of adiposity in newborns in a multi-ethnic genome-wide association
721 study. *Hum. Mol. Genet.* **22**, 3583–96 (2013).
- 722 37. Hivert, M.-F. *et al.* Genetic determinants of adiponectin regulation revealed by
723 pregnancy. *Obesity* **25**, 935–944 (2017).

- 724 38. Andersson, E. A. *et al.* The Birth Weight Lowering C-Allele of rs900400 Near
725 LEKR1 and CCNL1 Associates with Elevated Insulin Release following an Oral
726 Glucose Challenge. *PLoS One* **6**, e27096 (2011).
- 727 39. Li, C. *et al.* Genome-Wide Association Study Meta-Analysis of Long-Term
728 Average Blood Pressure in East Asians. *Circ. Cardiovasc. Genet.* **10**, e001527
729 (2017).
- 730 40. Daoud, G., Le bellego, F. & Lafond, J. PP2 regulates human trophoblast cells
731 differentiation by activating p38 and ERK1/2 and inhibiting FAK activation.
732 *Placenta* **29**, 862–70 (2008).
- 733 41. Kim, S.-M., Kang, J.-O., Lim, J. E., Hwang, S.-Y. & Oh, B. Csk Regulates Blood
734 Pressure by Controlling the Synthetic Pathways of Aldosterone. *Circ. J.* **82**, 168–
735 175 (2018).
- 736 42. Chan, M. C. *et al.* Tuning the transcriptional response to hypoxia by inhibiting
737 Hypoxia-inducible Factor (HIF) prolyl and asparaginyl hydroxylases. *J. Biol.*
738 *Chem.* **291**, 20661–20673 (2016).
- 739 43. Chen, D.-B. & Zheng, J. Regulation of Placental Angiogenesis. *Microcirculation*
740 **21**, 15–25 (2014).
- 741 44. Sahay, A. S., Sundrani, D. P. & Joshi, S. R. Neurotrophins: Role in Placental
742 Growth and Development. *Vitam. Horm.* **104**, 243–261 (2017).
- 743 45. Paul, S., Home, P., Bhattacharya, B. & Ray, S. GATA factors: Master regulators
744 of gene expression in trophoblast progenitors. *Placenta* **60**, S61–S66 (2017).
- 745 46. Bai, Q. *et al.* Dissecting the First Transcriptional Divergence During Human
746 Embryonic Development. *Stem Cell Rev.* **8**, 150–162 (2012).
- 747 47. Ma, G. T. *et al.* GATA-2 and GATA-3 regulate trophoblast-specific gene
748 expression in vivo. *Development* **124**, 907–14 (1997).

- 749 48. Dior, U. P. *et al.* Parental smoking during pregnancy and offspring cardio-
750 metabolic risk factors at ages 17 and 32. *Atherosclerosis* **235**, 430–7 (2014).
- 751 49. Behl, M. *et al.* Evaluation of the Association between Maternal Smoking,
752 Childhood Obesity, and Metabolic Disorders: A National Toxicology Program
753 Workshop Review. *Environ. Health Perspect.* **121**, 170–180 (2012).
- 754 50. Stančáková, A. & Laakso, M. Genetics of metabolic syndrome. *Rev. Endocr.*
755 *Metab. Disord.* **15**, 243–252 (2014).
- 756 51. Zacharasiewicz, A. Maternal smoking in pregnancy and its influence on
757 childhood asthma. *ERJ Open Res.* **2**, 00042–02016 (2016).
- 758 52. Dogan, M. V., Beach, S. R. H. & Philibert, R. A. Genetically contextual effects
759 of smoking on genome wide DNA methylation. *Am. J. Med. Genet. Part B*
760 *Neuropsychiatr. Genet.* (2017). doi:10.1002/ajmg.b.32565
- 761 53. Gao, X., Thomsen, H., Zhang, Y., Breitling, L. P. & Brenner, H. The impact of
762 methylation quantitative trait loci (mQTLs) on active smoking-related DNA
763 methylation changes. *Clin. Epigenetics* **9**, 1–13 (2017).
- 764 54. Tsai, P., Glastonbury, C. A., Eliot, M. N. & Bollepalli, S. Smoking induces
765 coordinated DNA methylation and gene expression changes in adipose tissue
766 with consequences for metabolic health. *bioRxiv Genomics* 1–21 (2018).
767 doi:10.1101/353581
- 768 55. Joubert, B. R. *et al.* Maternal smoking and DNA methylation in newborns: In
769 utero effect or epigenetic inheritance? *Cancer Epidemiol. Biomarkers Prev.* **23**,
770 1007–1017 (2014).
- 771 56. Wang, Y. & Zhao, S. Cell Types of the Placenta. (2010).
- 772 57. Fogarty, N. M. E., Burton, G. J. & Ferguson-Smith, A. C. Different epigenetic
773 states define syncytiotrophoblast and cytotrophoblast nuclei in the trophoblast of

- 774 the human placenta. *Placenta* **36**, 796–802 (2015).
- 775 58. Fortin, J.-P. *et al.* Functional normalization of 450k methylation array data
776 improves replication in large cancer studies. *Genome Biol.* **15**, 503 (2014).
- 777 59. Teschendorff, A. E. *et al.* A beta-mixture quantile normalization method for
778 correcting probe design bias in Illumina Infinium 450 k DNA methylation data.
779 *Bioinformatics* **29**, 189–196 (2013).
- 780 60. Chen, J. *et al.* CpGFilter: model-based CpG probe filtering with replicates for
781 epigenome-wide association studies. *Bioinformatics* **32**, 469–471 (2016).
- 782 61. Houseman, E. A., Molitor, J. & Marsit, C. J. Reference-free cell mixture
783 adjustments in analysis of DNA methylation data. *Bioinformatics* **30**, 1431–1439
784 (2014).
- 785 62. Venables, W. N. & Ripley, B. D. *Modern Applied Statistics with S.* (Springer,
786 2002).
- 787 63. Willer, C. J., Li, Y. & Abecasis, G. R. METAL: fast and efficient meta-analysis
788 of genomewide association scans. *Bioinformatics* **26**, 2190–2191 (2010).
- 789 64. Teschendorff, A. E. & Relton, C. L. Statistical and integrative system-level
790 analysis of DNA methylation data. *Nat. Rev. Genet.* **Nov 13**, (2017).
- 791 65. Deysenroth, M. A. *et al.* Whole-transcriptome analysis delineates the human
792 placenta gene network and its associations with fetal growth. *BMC Genomics* **18**,
793 (2017).
- 794 66. Ruiz-Arenas, C. & Gonzalez, J. MEAL: Perform methylation analysis. *R Packag.*
795 *version 1.10.1* (2018).
- 796 67. Ernst, J. & Kellis, M. ChromHMM: automating chromatin-state discovery and
797 characterization. *Nat. Methods* **9**, 215–6 (2012).
- 798 68. Kamburov, A. *et al.* ConsensusPathDB: Toward a more complete picture of cell

- 799 biology. *Nucleic Acids Res.* **39**, 712–717 (2011).
- 800 69. Chen, E. Y. *et al.* Enrichr: interactive and collaborative HTML5 gene list
801 enrichment analysis tool. *BMC Bioinformatics* **14**, (2013).
- 802 70. Lawrence, M. *et al.* Software for Computing and Annotating Genomic Ranges.
803 *PLOS Comput. Biol.* **9**, e1003118 (2013).
- 804 71. Villar, J. *et al.* International standards for newborn weight, length, and head
805 circumference by gestational age and sex: The Newborn Cross-Sectional Study
806 of the INTERGROWTH-21st Project. *Lancet* **384**, 857–868 (2014)

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811

812 **Author contributions**

813 TME, MVU, CJM, JL, MFH and MB conceived of and designed the study. Study-
814 specific analyses were completed by JMC, (AQUA), ES (EDEN), AC (Gen3G), PCS
815 and JMM (GENEIDA), MVU (INMA) and TME (NHBCS and RICHS). MV meta-
816 analysed these results. TME, MV and MB performed the follow-up analyses. TME,
817 MVU, JS, CJM, MFH, and MB interpreted the results. TME, MVU, and MB wrote the
818 first draft of the manuscript. All authors (TME, MVU, ES, AC, ML, JMC, CL, ERB,
819 NFJ, BH, PP, BGA, JH, MAD, MRK, CI, LB, PCS, YJL, KH, TB, MAC, JMM, EM,
820 JC, MFF, JT, AGM, SJL, JS, CJM, JL, MFH, MB) read and critically revised drafts of
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26 Table 1: Distribution of any and sustained MSDP within participating cohorts.

Cohort	Country	Any MSDP			Sustained MSDP		
		Total N	N (%) Non-smokers	N (%) Smokers	Total N	N (%) Non-smokers	N (%) Smokers
AQUA	Australia	99	75 (75.76%)	24 (24.24%)	-	-	-
EDEN	France	647	446 (68.93%)	201 (31.07%)	570	446 (68.93%)	124 (21.75%)
Gen3G	Canada	151	138 (91.39%)	13 (8.61%)	-	-	-
GENEID A	Spain	87	67 (77.01%)	20 (22.99%)	82	67 (77.01%)	15 (18.29%)
INMA	Spain	166	119 (71.69%)	47 (28.31%)	143	119 (71.69%)	24 (16.78%)
NHBCS	USA	310	290 (93.55%)	20 (6.45%)	-	-	-
RICHS	USA	240	221 (92.08%)	19 (7.92%)	-	-	-
TOTAL		1700	1356 (79.76%)	344 (20.24%)	795	632 (79.50%)	163 (20.50%)

27 MSDP = Maternal smoking during pregnancy.

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30 Table 2. Meta-analysis results from models of any and sustained MSDP, for the 20 CpGs with the smallest
 31 p-values for the model of any MSDP, and the percent increase in effect size between sustained and any
 32 MSDP (% Change); all models adjusted for maternal age, parity, education and putative cellular
 33 heterogeneity; CpGs that were not annotated with a gene name in the Illumina 450K annotation file have
 34 been annotated with their genomic region (ie. 4q12).

CpG Annotations			Any MSDP			Sustained MSDP			% Change
CpG ID	Location	Annotated Gene (Region)	β_1	S.E.	P-value	β_1	S.E.	P-value	
cg17823829	chr1:202765754	<i>KDM5B</i> (Body)	0.06	0.006	1.79E-22	0.086	0.008	3.80E-27	42.98
cg26045080	chr1:36807363	<i>STK40</i> (Body)	0.024	0.003	6.66E-19	0.032	0.003	4.86E-21	31.12
cg00534380	chr2:101766586	<i>TBC1D8</i> (Body)	-0.044	0.004	2.53E-25	-0.061	0.006	1.03E-28	38.46
cg19246018	chr2:240031588	<i>HDAC4</i> (Body)	0.015	0.002	1.39E-20	0.016	0.002	6.52E-20	8.28
cg23752985	chr2:85803571	<i>VAMP8</i> (TSS1500)	-0.019	0.002	1.31E-19	-0.025	0.003	3.77E-16	32.98
cg00666842	chr2:88366145	<i>SMYD1</i> (TSS1500)	0.033	0.003	1.17E-22	0.049	0.004	2.23E-34	48.17
cg27402634	chr3:156536860	3q25.31	-0.23	0.01	5.40E-119	-0.258	0.009	7.20E-181	12.33
cg09491670	chr4:53529646	4q12	0.016	0.002	1.60E-19	0.023	0.002	3.96E-29	43.31
cg25585967	chr5:14452105	<i>TRIO</i> (Body)	0.04	0.005	7.53E-19	0.061	0.006	2.16E-23	51.74
cg12291408	chr7:100037572	7q22.1	-0.036	0.004	1.43E-18	-0.052	0.005	2.34E-22	45.15
cg14214914	chr9:131870304	<i>CRAT</i> (Body)	0.036	0.004	1.57E-20	0.05	0.005	1.20E-26	41.29
cg20340720	chr10:104512523	<i>WBPI1</i> (Body)	-0.054	0.004	5.38E-42	-0.076	0.005	2.19E-58	40.52
cg26648103	chr11:66791718	<i>SYT12</i> (5'UTR)	-0.033	0.004	8.98E-21	-0.043	0.005	4.30E-20	30.51
cg26115089	chr11:93846406	<i>HEPHL1</i> (3'UTR)	0.025	0.002	1.33E-28	0.032	0.003	2.00E-27	24.8
cg26843110	chr15:74935742	<i>EDC3</i> (Body)	-0.056	0.005	1.74E-29	-0.082	0.007	1.74E-36	47.84
cg26433445	chr16:81764289	16q23.3	0.024	0.003	5.85E-20	0.034	0.003	1.34E-23	40.25
cg24177452	chr17:27494295	<i>MYO18A</i> (5'UTR)	0.025	0.003	3.40E-18	0.031	0.004	2.84E-18	27.35
cg06716730	chr17:35851459	<i>DUSP14</i> (5'UTR)	-0.022	0.003	1.40E-18	-0.032	0.003	4.47E-22	46.79
cg03313447	chr19:41829042	<i>CCDC97</i> (3'UTR)	-0.02	0.002	1.26E-22	-0.029	0.002	1.93E-34	45.73
cg02341503	chr20:45947123	<i>ZMYND8</i> (Body)	-0.02	0.002	1.81E-18	-0.025	0.003	4.83E-18	25.74

35 β_1 = Coefficient of the association between DNAm and MSDP; MSDP = Maternal smoking during
 36 pregnancy; S.E. = Standard Error.
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39 Table 3. Results from eQTM models, DNAm versus gestational age at birth, and DNAm versus BW Z-score
 40 models that yielded at least nominally significant associations, among the 20 CpGs that yielded the most
 41 statistically significant associations with any MSDP in the primary meta-analysis (sorted in the same order
 42 as Table 2).

CpG ID	Annotate d Gene	eQTM				Gestational Duration			BW Z-score		
		eQTM Gene	β_1	S.E.	P-value	β_1	S.E.	P-value	β_1	S.E.	P-value
cg17823829	<i>KDM5B</i>	<i>PPFIA4</i>	-2.53	0.7	4.05E-04	-	0.2	9.12E-06	-	0.2	9.72E-05
cg26045080	<i>STK40</i>	<i>SH3D21</i>	-4.26	0.7	8.34E-08	-	0.4	4.19E-03	-	-	-
cg00534380	<i>TBC1D8</i>	<i>TBC1D8</i>	-2.29	0.5	1.13E-05	3.53	0.3	9.26E-09	1.48	0.3	1.62E-04
cg19246018	<i>HDAC4</i>	-	-	-	-	-	0.5	1.74E-02	-	-	-
cg23752985	<i>VAMP8</i>	<i>CAPG</i>	6.33	1.7	3.96E-05	5.47	0.6	1.40E-09	-	-	-
cg00666842	<i>SMYD1</i>	-	-	-	-	-	-	-	-	-	-
cg27402634	3q25.31	<i>LEKR1</i>	-2.32	0.6	7.14E-04	-	-	-	0.92	0.1	6.71E-07
cg09491670	4q12	<i>USP46</i>	-3.59	-	4.91E-04	-	-	-	-	-	-
cg25585967	<i>TRIO</i>	-	-	-	-	-	0.3	8.64E-03	-	0.3	2.06E-04
cg12291408	7q22.1	<i>ACTL6B</i>	-4.89	-	1.12E-02	-	-	-	1.31	0.3	6.82E-04
cg14214914	<i>CRAT</i>	<i>CRAT</i>	-4.76	0.5	6.03E-09	0.96	0.3	1.35E-02	-	-	-
cg20340720	<i>WBP1L</i>	-	-	-	-	-	-	-	1.86	0.3	2.42E-07
cg26648103	<i>SYT12</i>	-	-	-	-	1.06	0.4	1.29E-02	1.66	0.4	7.56E-05
cg26115089	<i>HEPHL1</i>	-	-	-	-	-1.1	0.5	4.43E-02	1.48	0.5	6.71E-03
cg26843110	<i>EDC3</i>	<i>CSK</i>	0.57	0.2	2.18E-05	2.3	0.3	5.09E-03	1.28	0.3	1.19E-04
cg26433445	16q23.3	-	-	-	-	-	-	-	2.99	0.6	9.06E-07
cg24177452	<i>MYO18A</i>	<i>MYO18</i>	-1.37	0.4	1.79E-03	-	-	-	-	-	-
cg06716730	<i>DUSP14</i>	<i>DUSP14</i>	-3.07	0.9	1.59E-03	3.72	0.5	7.87E-04	-	-	-
cg03313447	<i>CCDC97</i>	<i>TGFB1</i>	-	1.4	7.27E-13	-	0.7	7.79E-04	2.29	0.7	1.58E-03
cg02341503	<i>ZMYND8</i>	<i>ZMYND8</i>	-1.72	0.6	7.37E-03	1.44	0.6	2.29E-02	3.19	0.6	2.70E-07

13 β_1 = Coefficient of the association between DNAm and the outcome variable (mRNA expression,
 14 Gestational duration, or BW Z-score); BW = Birth weight; S.E. = Standard Error.
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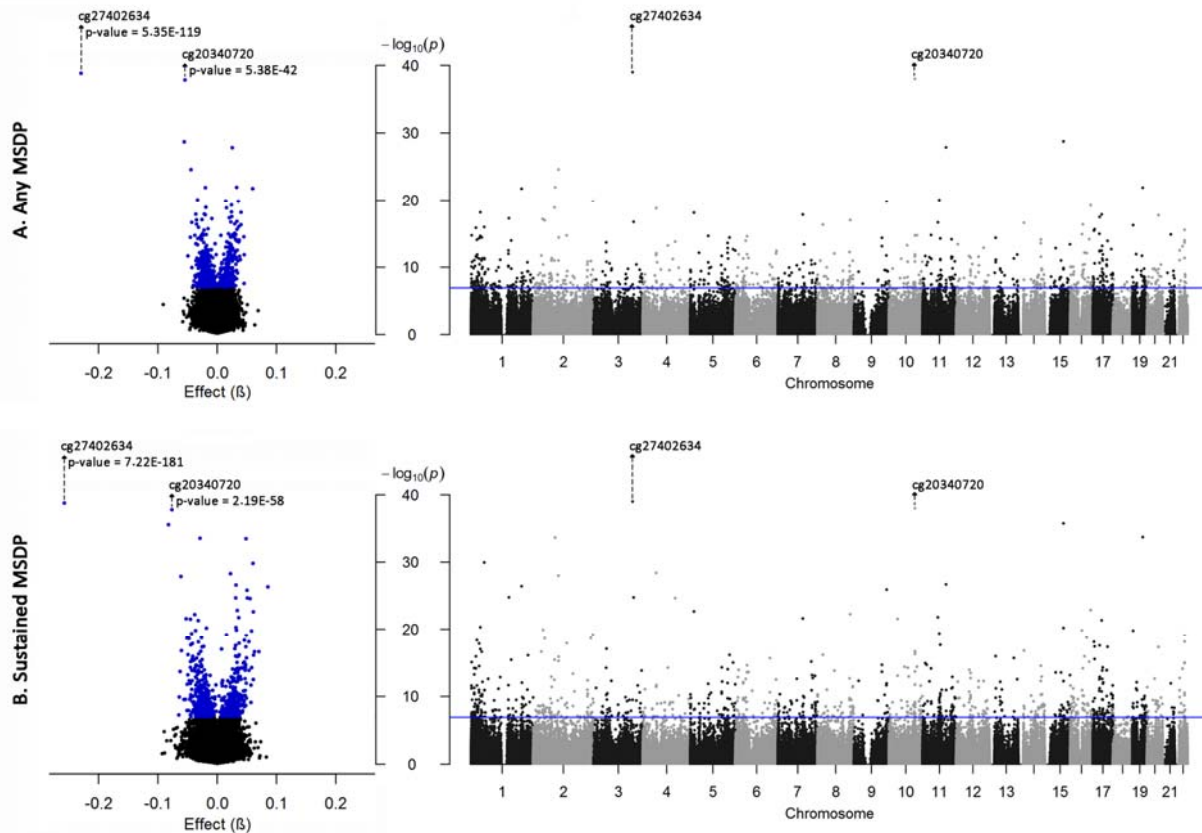
18 Table 4: Differentially methylated CpGs with MSDP in placenta (from our meta-analysis) and cord blood
 19 (from a published PACE meta-analysis¹⁰) that yielded Bonferroni-significant associations in both tissues.

CpG Annotation		Cord blood* (Sustained MSDP)			Placenta (Sustained MSDP)			Placenta (Any MSDP)		
CpG ID	Gene	β_1	S.E.	P-value	β_1	S.E.	P-value	β_1	S.E.	P-value
cg25189904	<i>GNGL2</i> (TSS1500)	-0.024	0.002	1.38E-32	-0.020	0.003	2.54E-13	-0.014	0.002	1.99E-10
cg05575921	<i>AHRR</i> (Body)	-0.064	0.002	1.64E-193	-0.030	0.005	5.01E-09	-0.015	0.004	1.83E-04
cg21161138	<i>AHRR</i> (Body)	-0.022	0.001	1.78E-54	-0.026	0.004	5.25E-11	-0.018	0.003	2.55E-10
cg08327744	<i>RNF122</i> (Body)	-0.010	0.002	1.37E-08	0.020	0.003	3.12E-09	0.016	0.003	2.21E-09
cg15893360	<i>PXN</i> (Body)	-0.011	0.002	1.66E-08	-0.024	0.004	3.61E-08	-0.015	0.003	3.43E-06
cg12101586	<i>CYP1A1</i> (TSS1500)	0.045	0.003	3.67E-50	-0.045	0.008	7.15E-08	-0.026	0.006	9.12E-06
cg23680900	<i>CYP1A1</i> (TSS200)	0.003	0.001	8.73E-08	-0.018	0.002	6.26E-21	-0.006	0.001	7.35E-06
cg07565956	<i>ZBTB4</i> (5'UTR)	-0.006	0.001	2.50E-10	-0.023	0.003	4.02E-18	-0.016	0.002	2.26E-16
cg16547579	<i>SLC23A2</i> (5'UTR)	-0.013	0.002	2.00E-10	-0.048	0.009	1.47E-08	-0.031	0.007	1.42E-06

50 β_1 = Coefficient of the association between DNAm and MSDP; MSDP = Maternal smoking during
 51 pregnancy; S.E. = Standard Error

52 * Main model in Joubert et al: sustained MSDP adjusted for cell type heterogeneity

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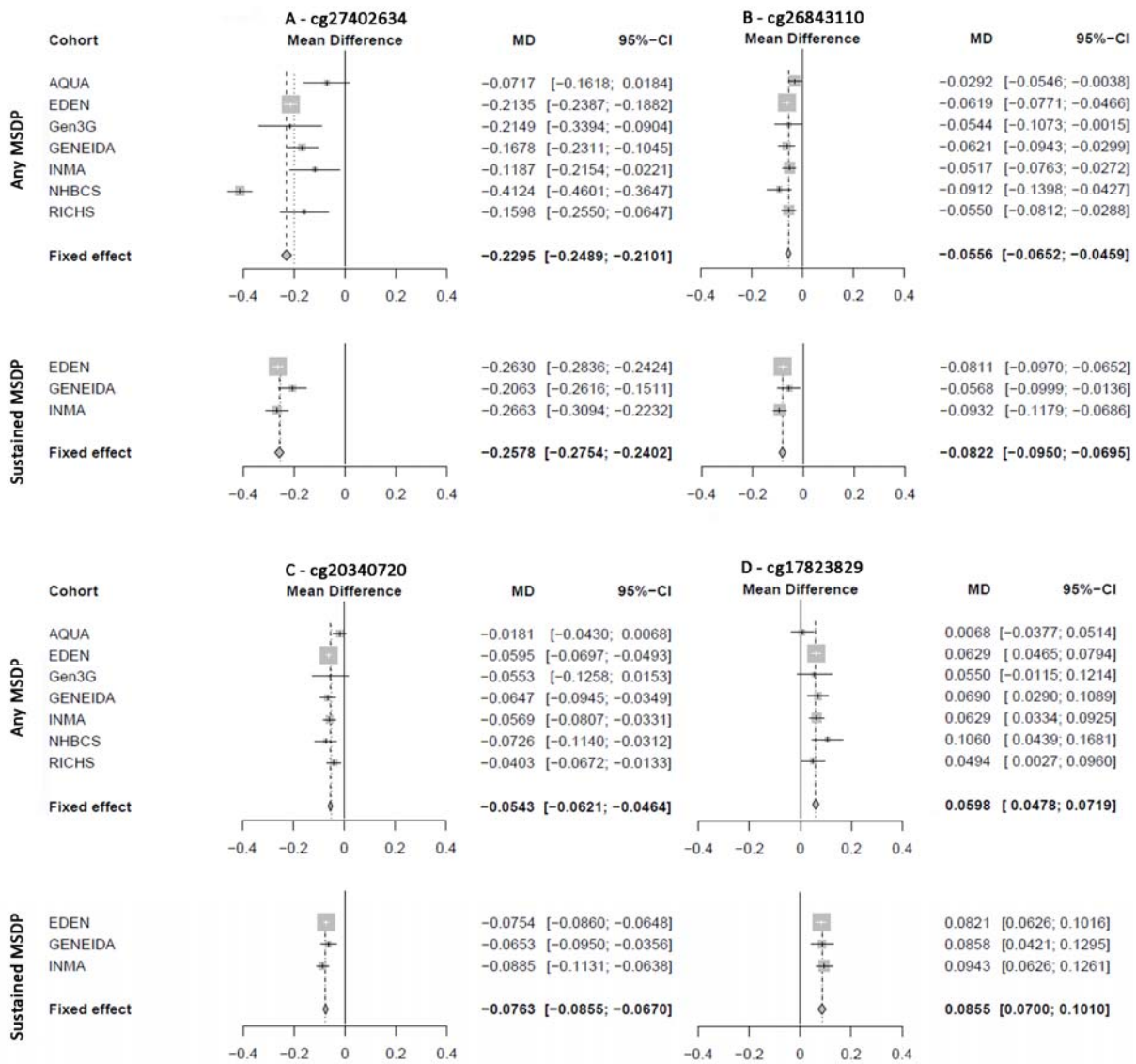
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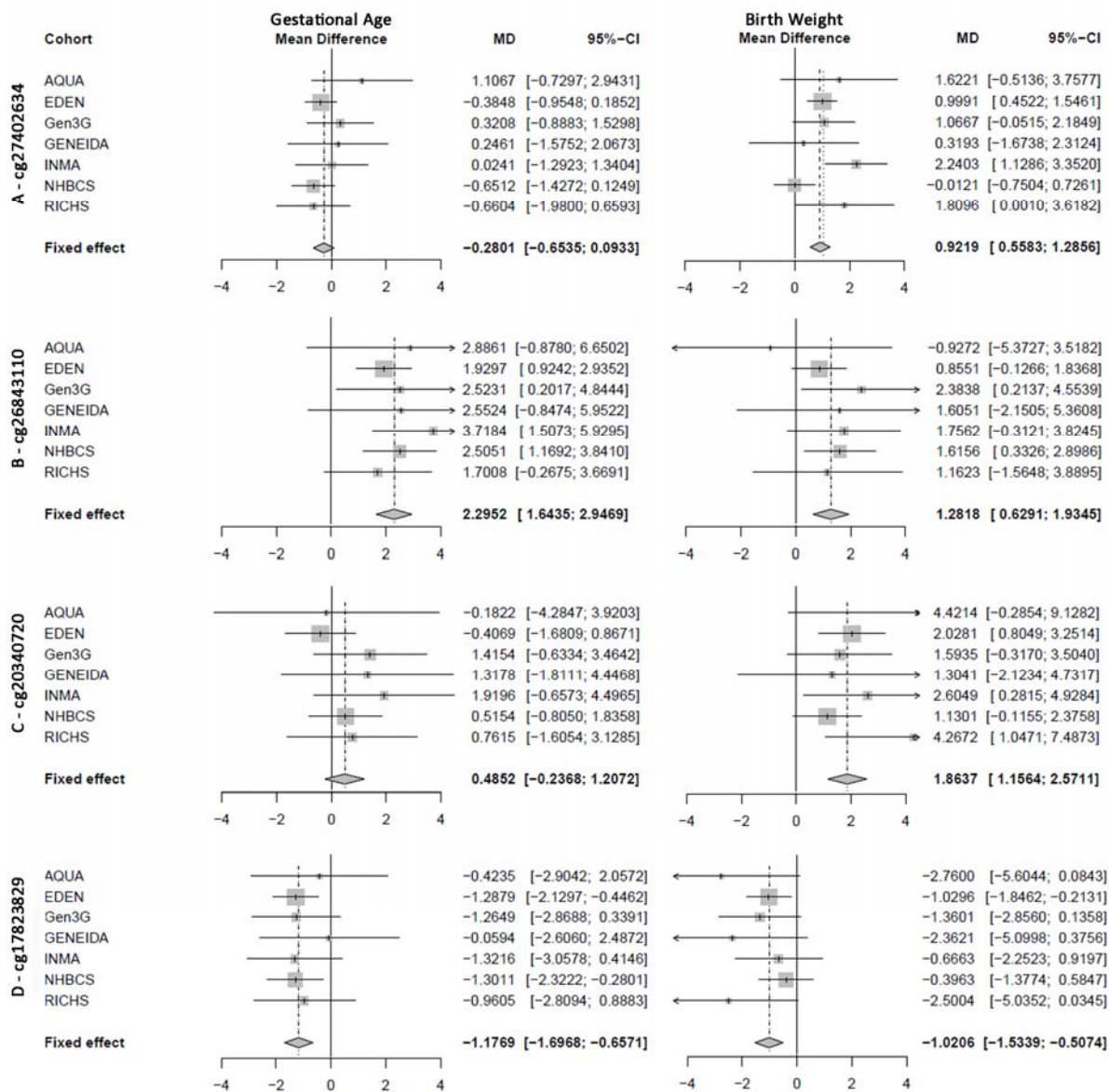
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Figure 1. Volcano and Manhattan plots of the association between any MSDP (total N=1700, exposed = 344) (A) and sustained MSDP (total N=795, exposed = 163) (B) with placental DNAm adjusted for maternal age, parity, maternal education and putative cellular heterogeneity. In the volcano plots, the x-axis shows the difference in DNAm (effect) with a possible range between 0 and 1, while the x-axis in the Manhattan plot represents genomic location; both plots share the same y-axis with $-\log_{10}(P)$. Bonferroni thresholds for statistical significance are shown as blue dots and a blue horizontal line, for volcano and Manhattan plots, respectively. The y-axes were truncated to a minimum p-value of 1×10^{-40} (or maximum $-\log_{10}(P)$ of 40), to allow for better visualization of the majority of our results. The CpGs that were impacted by y-axis truncation are indicated with arrows and annotated with their actual p-values.



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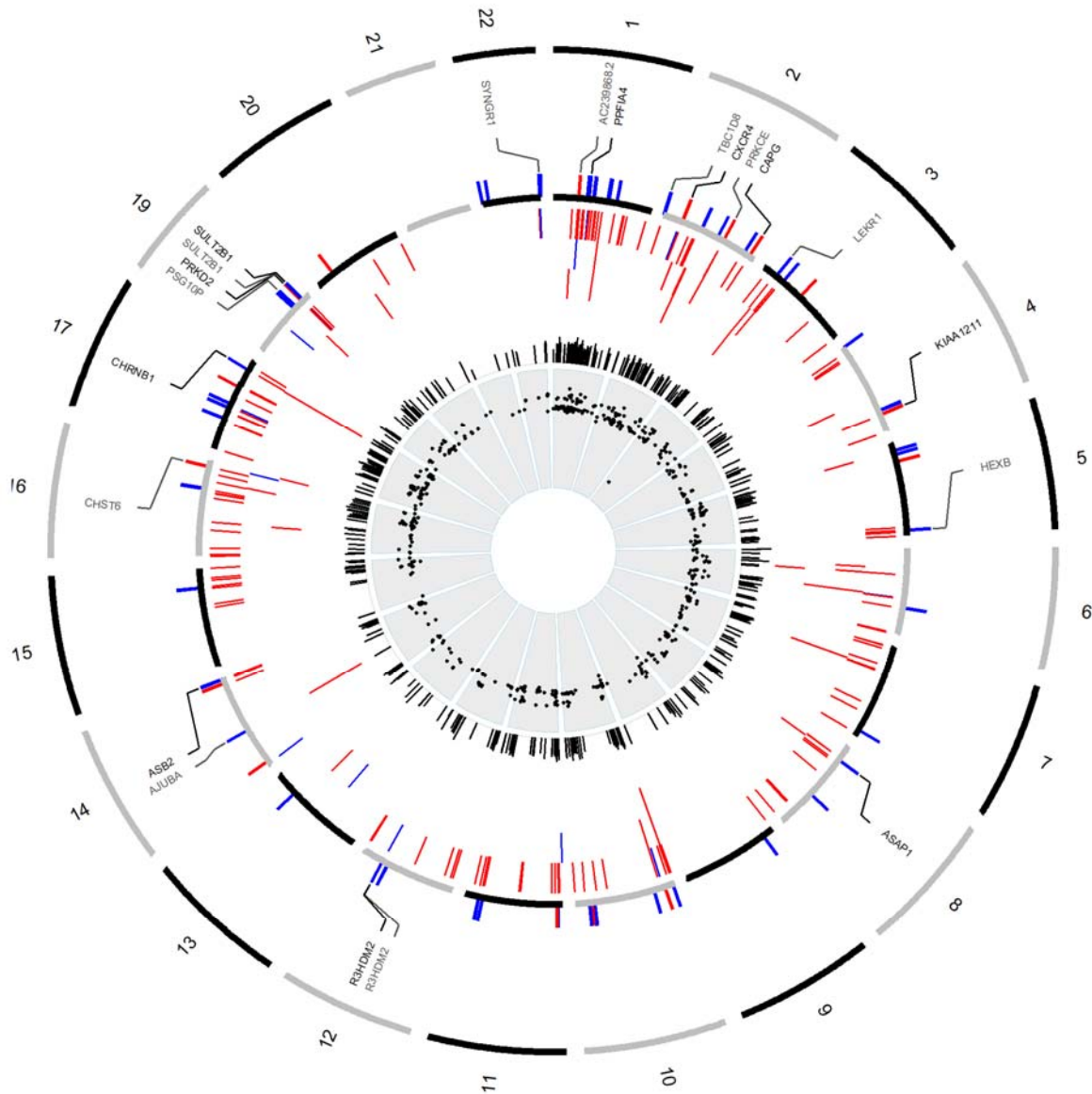
Figure 2. Forest plots of the cohort specific estimates of association and fixed-effect meta-analysis estimates of association between placental DNAm levels at (A) cg27402634, (B) cg26843110, (C) cg20340720, and (D) cg17823829 with any MSDP and sustained MSDP; models adjusted for maternal age, parity, maternal education, and putative cellular heterogeneity.



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Figure 3. Forest plots of the cohort specific estimates of association and fixed-effect meta-analysis estimates of association between higher levels of placental DNAm at (A) cg27402634, (B) cg26843110, (C) cg20340720, and (D) cg17823829 with gestational age and birth weight z-scores; models adjusted for maternal age, parity, maternal education, and putative cellular heterogeneity.

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38 **Figure 4.** Circos plot summarizing all analyses for the 548 CpG sites that yielded Bonferroni-significant
39 associations with both any and sustained MSDP. From inner to outer-band: Black points = differential
40 DNAm between any- and no MSDP; Black Bars = percent increase in magnitude of association when
41 comparing results from any MSDP to sustained MSDP; Color Bars = Positive (red) or Inverse (blue)
42 associations between DNAm and HC, BL, BW, and GA, that yielded Bonferroni-significant associations;
43 Color Bars = Positive (red) or Inverse (blue) associations between DNAm and nearby gene expression, that
44 yielded an association at a 5% FDR; Names of eQTM genes were annotated to the outer band for CpGs that
45 yielded an association between DNAm and expression with a 5% FDR and yielded an association between
46 DNAm and at least one birth outcome after Bonferroni-adjustment.

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