

1                   **Phylogenetic relationships in the genus *Avena* based on**  
2   **the nuclear *Pgk1* gene**

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## 22 **Abstract**

23           The phylogenetic relationships among 76 *Avena* taxa, representing 14 diploids,  
24 eight tetraploids, and four hexaploids were investigated by using the nuclear plastid  
25 3-phosphoglycerate kinase gene (*Pgk1*). A significant deletion (131 bp) was detected  
26 in all the C genome homoeologues which reconfirmed a major structural divergence  
27 between the A and C genomes. Phylogenetic analysis indicated the C<sub>p</sub> genome is  
28 more closely related to the polyploid species than is the C<sub>v</sub> genome. Two haplotypes  
29 of *Pgk1* gene were obtained from most of the AB genome tetraploids. Both types of  
30 the *barbata* group showed a close relationship with the A<sub>s</sub> genome diploid species,  
31 supporting the hypothesis that both the A and B genomes are derived from an A<sub>s</sub>  
32 genome. Two haplotypes were also detected in *A. agadiriana*, which showed close  
33 relationships with the A<sub>s</sub> genome diploid and the A<sub>c</sub> genome diploid, respectively,  
34 emphasizing the important role of the A<sub>c</sub> genome in the evolution of *A. agadiriana*.  
35 Three homoeologues of the *Pgk1* gene were detected in five hexaploid accessions.  
36 The homoeologues that might represent the D genome were tightly clustered with  
37 the tetraploids *A. marrocana* and *A. murphyi*, but did not show a close relationship  
38 with any extant diploid species.

## 39 **Introduction**

40           The genus *Avena* L. belongs to the tribe Aveneae of the grass family  
41 (Poaceae). It contains approximately 30 species [1-4] reflecting a wide range of  
42 morphological and ecological diversity over the temperate and subtropical regions  
43 [5]. The evolutionary history of *Avena* species has been discussed for decades, and

44 remains a matter of debate despite considerable research effort in this field.  
45 Cytologically, three ploidy levels are recognized in the genus *Avena*: diploid,  
46 tetraploid, and hexaploid, with a base number of seven chromosomes [6, 7]. The  
47 diploids are divided clearly into two distinct lineages with the A and C genomes. All  
48 hexaploid species share the same genomic constitution of ACD, corroborated by  
49 fertile interspecific crosses among each other, as well as by their similar genome  
50 sizes [8]. With less certainty, the tetraploids have been designated as AB or AA, AC or  
51 DC, and CC genomes [9]. It is noteworthy that the B and D genomes within the  
52 polyploid species have not been identified in any extant diploid species. There are  
53 three C genome diploid species, which have been grouped into two genome types  
54 ( $C_p$  and  $C_v$ ) according to their karyotypes [10]. Both types show a high degree of  
55 chromosome affinity to the polyploid C genome [9-14], but none have been  
56 undisputedly identified as the C genome progenitor of the polyploids.

57         The A genome origin of polyploid oats has also been under intense scrutiny.  
58 However, there is no conclusive evidence regarding which the A genome diploid  
59 contributed to the polyploid oats. There are up to 12 species designated as A  
60 genome diploids. These species have been further subdivided into five sub-types of  
61  $A_c$ ,  $A_d$ ,  $A_l$ ,  $A_p$  and  $A_s$  genomes, according to their karyotypes [6, 7]. Most research  
62 based on karyotype comparisons [6, 15], in situ hybridization [11, 16-18], as well as  
63 the alignments of nuclear genes [13, 14] suggest that one of the  $A_s$  genome species  
64 may be the A genome donor of polyploid oats. Alternatively, some studies have

65 proposed the  $A_c$  genome diploid *A. canariensis* [19], or the  $A_l$  genome diploid *A.*

66 *longiglumis* [9, 12] as the most likely A genome donor.

67         The absence of diploids with the B and D genomes complicates the B and D  
68 genome donor identification. It is generally accepted that both B and D genomes are  
69 derived from A genomes, due to the high homology between the B and A genomes  
70 [11, 20], as well as between the D and A genomes [16, 19, 21]. Our recent study  
71 based on high-density genotyping-by-sequencing (GBS) markers [9] provided strong  
72 evidence that the three tetraploid species formerly designated as AC genomes are  
73 much closer to the C and D genomes of the hexaploids than they are to the  
74 hexaploid A genome. These findings suggest that the hexaploid D genome exists in  
75 the extant tetraploids. However, no extant diploid species, even the  $A_c$  genome  
76 diploid *A. canariensis*, which was considered as the most likely D genome progenitor  
77 based on direct evidence from morphological features [22] and indirect evidence  
78 from fluorescent in situ hybridization (FISH) [18], showed enough similarity to the D  
79 genome of tetraploid and hexaploid oats to warrant consideration as a direct D  
80 genome progenitor.

81         In the case of the B genome, an initial study of chromosome pairing of  
82 hybrids between the AB genome tetraploids and the  $A_s$  genome diploids suggested  
83 that the B genome arose from the  $A_s$  genome through autopoloidization [23].  
84 Recently, another GBS study [19] showed that the AB genome tetraploid species fell  
85 into a tight cluster with  $A_s$  genome diploids, also supporting the hypothesis that the  
86 B genome arose through minor divergence following autopoloidization. However,

87 other evidence from C-banding [24], FISH [17], RAPD markers [25], and DNA  
88 sequence alignment [14] has indicated a clear distinction between A and B genomes,  
89 suggesting an allotetraploid origin of the AB genome tetraploid species. The most  
90 probable A genome progenitor of the AB genome tetraploids is assumed to be an  $A_s$   
91 genome diploid species, while the B genome of these species remains controversial.

92 Single or low copy nuclear genes are widely used in phylogenetic analyses  
93 due to their bi-parental inheritance and to the informativeness of mutations. Such  
94 studies have successfully revealed multiple polyploid origins, and clarified  
95 hybridization events in a variety of plant families [26, 27]. In a previous study [14],  
96 we investigated the relationships among *Avena* species by sequencing the  
97 single-copy nuclear acetyl-coA carboxylase gene (*Acc1*). The results provided some  
98 useful clues to the relationships of *Avena* species.

99 The *Pgk1* gene, which encodes the plastid 3-phosphoglyceratekinase, is  
100 another nuclear gene that has been widely used to reveal the evolutionary history of  
101 the *Triticum/Aegilops* complex due to its single copy status per diploid chromosome  
102 in grass [26, 28, 29]. The *Pgk1* gene is now considered to be superior to the *Acc1*  
103 gene in phylogenetic analysis, since it has more parsimony informative sites than the  
104 *Acc1* gene [26, 29]. In the present study, we sequenced cloned *Pgk1* gene copies  
105 from 76 accessions representing the majority of *Avena* species, in an attempt to  
106 further clarify evolutionary events in this important genus.

## 107 **Materials and Methods**

## 108 Plant materials

109 A total of 76 accessions from 26 *Avena* species were investigated to represent  
 110 the geographic range of six sections in *Avena*, together with one accession from  
 111 *Trisetopsis turgidula* as a functional outgroup (Table 1). All seeds were provided by  
 112 Plant Gene Resources of Canada (PGRC) or the National Small Grains Collection,  
 113 Agriculture Research Service, United States Department of Agriculture (USDA, ARS)  
 114 with the exception of the three accessions of *A. insularis*, which were kindly provided  
 115 by Dr. Rick Jellen, Brigham Young University, Provo, UT, USA. The species *A.*  
 116 *atherantha*, *A. hybrida*, *A. matritensis* and *A. trichophylla* described in Baum's [1]  
 117 monograph and *A. prostrata* described by Ladizinsky [30] were not included due to a  
 118 lack of viable material.

119 **Table 1. List of materials used in the present study including species, haplomes,**  
 120 **accession number, origin, abbreviation displayed in MJ network, and the sequence**  
 121 **number in Genbank (<https://www.ncbi.nlm.nih.gov>).**

Taxa	Haplomes	Accession Number	Origin *	Abbreviation	Genbank Accession
Section <i>Ventricosa</i>					
<i>A. clauda</i> Dur.	C <sub>p</sub>	CN 19242	Turkey	CLA1_1	KU888786
		CN 21378	Greece	CLA2_1	KU888787
		CN 21388	Algeria	CLA3_1	KU888804
		CN 24695	Turkey	CLA4_1	KU888784
<i>A. eriantha</i> Dur. (syn <i>A. pilosa</i> Bieb.)	C <sub>p</sub>	Clav 9050	United Kingdom	ERI1_1	KU888785
		PI 367381	Madrid, Spain	ERI2_1	KU888805
<i>A. ventricosa</i> Balansa ex Coss.	C <sub>v</sub>	CN 21405	Algeria	VEN1_1	KU888806
		CN 39706	Azerbaijan	VEN2_1	KU888807
Section <i>Agraria</i>					
<i>A. brevis</i> Roth	A <sub>s</sub>	Clav 1783	German	BRE1_1	KU888707

		Clav 9113	Europe	BRE2_1	KU888718
		PI 258545	Portugal	BRE3_1	KU888710
<i>A. hispanica</i> Ard.	A <sub>s</sub>	CN 25676	Portugal	HIS1_1	KU888714
		CN 25727	Portugal	HIS2_1	KU888711
		CN 25766	Portugal	HIS3_1	KU888709
		CN 25778	Portugal	HIS4_1	KU888712
<i>A. nuda</i> L.	A <sub>s</sub>	PI 401795	Netherlands	NUD1_1	KU888734
<i>A. strigose</i> Schreb.	A <sub>s</sub>	PI 83722	Australia	STR1_1	KU888719
		PI 158246	Lugo, Spain	STR2_1	KU888713
		Clav 9066	Ontario, Canada	STR3_1	KU888708
Section <i>Tenuicarpa</i>					
<i>A. agadiriana</i> Baum & Fedak	AB	CN 25837	Africa: Morocco	AGA1_1	KU888753
				AGA1_2	KU888774
		CN 25854	Africa: Morocco	AGA2_1	KU888777
				AGA2_2	KU888754
		CN 25856	Africa: Morocco	AGA3_1	KU888776
				AGA3_2	KU888751
		CN 25863	Africa: Morocco	AGA4_1	KU888775
		CN 25869	Africa: Morocco	AGA5_1	KU888752
				AGA5_2	KU888778
<i>A. atlantica</i> Baum & Fedak	A <sub>s</sub>	CN 25849	Africa: Morocco	ATL1_1	KU888757
		CN 25859	Africa: Morocco	ATL2_1	KU888756
		CN 25864	Africa: Morocco	ATL3_1	KU888739
		CN 25887	Africa: Morocco	ATL4_1	KU888737
		CN 25897	Africa: Morocco	ATL5_1	KU888736
<i>A. barbata</i> Pott ex Link	AB	PI 296229	Northern, Israel	BAR1_1	KU888723
		PI 337802	Izmir, Turkey	BAR2_1	KU888722
				BAR2_2	KU888732
		PI 337826	Greece	BAR3_1	KU888720
		PI 282723	Northern, Israel	BAR4_1	KU888729
		PI 337731	Macedonia, Greece	BAR5_1	KU888731
		PI 367322	Beja, Portugal	BAR6_1	KU888730
<i>A. canariensis</i> Baum et al	A <sub>c</sub>	CN 23017	Canary Islands	CAN1_1	KU888779
		CN 23029	Canary Islands	CAN2_1	KU888782
		CN 25442	Canary Islands	CAN3_1	KU888780
		CN 26172	Canary Islands	CAN4_1	KU888783
		CN 26195	Canary Islands	CAN5_1	KU888781
<i>A. damascena</i> Rajah & Baum	A <sub>d</sub>	CN 19457	Syria	DAM1_1	KU888744
		CN 19458	Syria	DAM2_1	KU888745

		CN 19459	Syria	DAM3_1	KU888747
<i>A. hirtula</i> Lag.	A <sub>s</sub>	CN 19530	Antalya, Turkey	HIR1_1	KU888738
		CN 19739	Algeria	HIR2_1	KU888762
<i>A. longiglumis</i> Dur.	A <sub>l</sub>	CN 21703	Morocco	HIR3_1	KU888717
		Clav 9087	Oran, Algeria	LON1_1	KU888741
		Clav 9089	Libya	LON2_1	KU888749
<i>A. lusitanica</i> Baum	A <sub>s</sub>	PI 367389	Setubal, Portugal	LON3_1	KU888750
		CN 25885	Morocco	LUS1_1	KU888746
		CN 25899	Morocco	LUS2_1	KU888748
		CN 26265	Portugal	LUS3_1	KU888742
<i>A. wiestii</i> Steud.	A <sub>s</sub>	CN 26441	Spain	LUS4_1	KU888763
		PI 53626	Giza, Egypt	WIE1_1	KU888715
		Clav 9053	Ontario, Canada	WIE2_1	KU888716
<i>Section Ethiopica</i>					
<i>A. abyssinica</i> Hochst.	AB	PI 411163	Seraye, Eritrea	ABY1_1	KU888724
		PI 411173	Tigre, Ethiopia	ABY2_1	KU888740
				ABY2_2	KU888725
<i>A. vaviloviana</i> Mordv.	AB	PI 412761	Eritrea	VAV1_1	KU888743
				VAV1_2	KU888728
		PI 412766	Shewa, Ethiopia	VAV2_1	KU888726
				VAV2_2	KU888735
<i>Section Pachycarpa</i>					
<i>A. insularis</i> Ladiz.	AC(DC)	sn	Sicily, Italy	INS1_1	KU888794
				INS1_2	KU888705
		6-B-22	Sicily, Gela, Italy	INS2_1	KU888706
				INS2_2	KU888796
		INS-4	Sicily, Gela, Italy	INS3_1	KU888790
				INS3_2	KU888704
<i>A. maroccana</i> Grand. (syn. <i>A. magna</i> Murphy et Terrell)	AC(DC)	Clav 8330	Morocco	MAR1_1	KU888773
				MAR1_2	KU888799
		Clav 8331	Khemisset, Morocco	MAR2_1	KU888721
				MAR2_2	KU888800
<i>A. murphyi</i> Ladiz.	AC(DC)	CN 21989	Spain	MUR1_1	KU888767
				MUR1_2	KU888802
		CN 25974	Morocco	MUR2_1	KU888769
				MUR2_2	KU888788
<i>Section Avena</i>					
<i>A. fatua</i> L.	ACD	PI 447299	Gansu, China	FAT1_1	KU888768
				FAT1_2	KU888795
		PI 544659	United States	FAT2_1	KU888764



				FAT2_2	KU888760
				FAT2_3	KU888798
<i>A. occidentalis</i> Dur.	ACD	CN 4547	Canary Islands, Spain	OCC1_1	KU888791
		CN 23036	Canary Islands, Spain	OCC2_1	KU888755
				OCC2_2	KU888803
				OCC2_3	KU888771
		CN 25942	Morocco	OCC3_1	KU888733
				OCC3_2	KU888789
				OCC3_3	KU888758
		CN 25956	Morocco	OCC4_1	KU888801
				OCC4_2	KU888772
<i>A. sativa</i> L.	ACD	PI 194896	Gonder, Ethiopia	SAT1_1	KU888727
				SAT1_2	KU888759
				SAT1_3	KU888793
		PI 258655	Russian Federation	SAT2_1	KU888797
				SAT2_2	KU888766
				SAT2_3	KU888761
<i>A. sterilis</i> L.	ACD	PI 411503	Alger, Algeria	STE1_1	KU888765
		PI 411656	Tigre, Ethiopia	STE2_1	KU888792
				STE2_2	KU888770
Outgroup					
<i>Trisetopsis turgidula</i>					
Röser & A. Wölk		PI 364343	Maseru, Lesotho		KU888808

122 \* Origin represents the collection site of wild material where this information is  
 123 available, otherwise it represents the earliest source for which information is  
 124 available.

## 125 DNA isolation, cloning and sequencing

126 Genomic DNA was isolated from fresh leaves of single plants following a  
 127 standard CTAB protocol [31]. *Pgk1* gene sequences were amplified by using a pair of  
 128 *Pgk1*-specific primers, PGKF1 (5'-TCGTCCTAAGGGTGTTACTCCTAA-3') and PGKR1  
 129 (5'-ACCACCAGTTGAGATGTGGCTCAT-3') described by Huang et al. [28]. Polymerase

130 chain reactions (PCR) were carried out under cycling conditions reported previously  
131 [26]. After estimating the size by 1.0% agarose gel, PCR products were purified using  
132 the QIAquick gel extraction kit (QIAGEN Inc., USA). The purified products were  
133 cloned into the pMD19-T vector (Takara) following the manufacturer's instructions.  
134 Initially, 6-8 positive clones from each of four accessions from 4 diploid  
135 species, including *A. canariensis* ( $A_c$ ), *A. longiglumis* ( $A_l$ ), *A. strigosa* ( $A_s$ ), and *A. clauda*  
136 ( $C_p$ ), were sequenced to confirm that the *Pgk1* gene was present in *Avena* diploid  
137 species as a single copy. After confirming its single copy status in diploid species, 2-3  
138 positive clones were selected and sequenced from each accession of the remaining  
139 diploid species. In order to isolate all possible homoeologous sequences in polyploid  
140 species, 4-6 positive clones from each accession of the tetraploid species and 5-10  
141 positive clones from each accession of the hexaploid species were selected and  
142 sequenced. All the cloned PCR products were sequenced on both strands by a  
143 commercial company (Sangon Biotech Co., Ltd., Shanghai, China) based on Sanger  
144 sequencing technology.

## 145 **Sequence alignment and phylogenetic analysis**

146 The homology of sequences was verified using the BLAST program in NCBI. In  
147 order to reduce the matrix size of the dataset, redundant sequences were removed,  
148 keeping one representative sequence if several identical sequences were derived  
149 from the same accession. Sequences were aligned using ClustalW software with  
150 default parameters [32] followed by manual correction. Substitution saturation of  
151 *Pgk1* sequences was examined using DAMBE version 5 [33] by calculating and plotting

152 pairwise rates of transitions and transversions against sequence divergence under  
153 the TN93 model. Phylogenetic trees were created by using Maximum parsimony  
154 (MP), and Bayesian inference (BI). MP analysis was performed on PAUP\* 4.0b10 [34]  
155 using the heuristic search with 100 random addition sequence replicates and Tree  
156 Bisection-Reconnection (TBR) branch swapping algorithms. Bootstrapping with 1000  
157 replicates was estimated to determine the robustness of formed branches [35]. Gaps  
158 in the sequence alignment were disregarded using the option 'gapmode=missing',  
159 which is consistent with an assumption that insertion/deletion events are an  
160 independent stochastic process from SNP substitutions. BI analysis was carried out  
161 by using MrBayes v3.2 [36]. The best-fit substitution model for BI analysis was  
162 GTR+ $\Gamma$ +I, which was determined by using MrModelTest v2.3 under Akaike  
163 information criteria (AIC) (<http://www.ebc.uu.se/systzoo/staff/nylander.html>). Four  
164 Markov chain Monte Carlo (MCMC) chains with default priors settings were run  
165 simultaneously. To ensure the two runs converged onto the stationary distribution,  
166 6,000,000 generations were run to make the standard deviation of split frequencies  
167 fall below 0.01. Samples were taken every 100 generations. The first 25% samples  
168 from each run were discarded as the "burn-in". The 50% majority-rule consensus  
169 tree was constructed from the remaining trees. Posterior probability (PP) values  
170 were used to evaluate the statistical confidence of each node.

## 171 **Network analysis**

172 The median-joining (MJ) network [37] method has been demonstrated to be  
173 an effective method for assessing the relationship in closely related lineages [38],

174 and thus was applied in this study. As MJ algorithms are designed for  
175 non-recombining molecules [37], DNA recombination was test by using a pragmatic  
176 approach-Genetic Algorithm Recombination Detection (GARD), described by Pond et  
177 al. [39]. The test was carried out on a web-based interface for GARD at  
178 <http://www.datamonkey.org/GARD/>. Building upon this test, the intron data was  
179 used for MJ reconstruction due to the absence of recombination signal, while  
180 potential recombination signals were detected in the exon regions. The MJ network  
181 analyses was performed using the Network 4.6.1.4 program (Fluxus Technology Ltd,  
182 Clare, Suffolk, UK).

## 183 **Results**

### 184 **Sequence analysis**

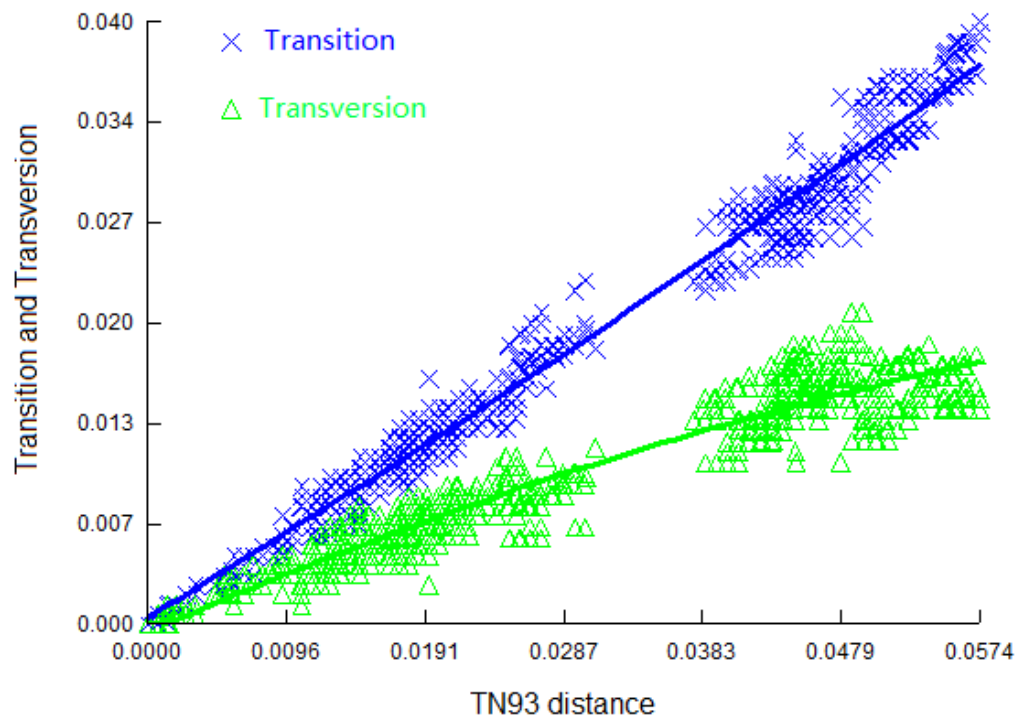
185 A total of 237 clones were sequenced from 76 accessions of 26 *Avena* species.  
186 Following removal of the redundant sequences within each accession, 104  
187 sequences were identified, including one from each of the 44 diploid accessions, 37  
188 unique sequences from 22 tetraploids, and 23 from 10 hexaploids. Theoretically, 44  
189 homoeologues should be isolated from 22 tetraploid accessions, and 30 single-copy  
190 homoeologues were expected from 10 hexaploid accessions. However, the full  
191 number of expected homoeologues were not isolated from every polyploid species  
192 for various potential reasons. In particular, within the AB genome tetraploid species  
193 *A. barbata*, only one copy was detected in five of its six accessions, whereas two very  
194 similar (only one site varied in exon 2) copies were detected in the sixth accession.  
195 This also happened in the hexaploid species *A. sterilis*, for which two accessions

196 provided only two homoeologues each. For these taxa, the missing genome type  
197 might be detected by screening a larger number of positive clones, but it is also  
198 possible that these accessions contain genomes of high similarity or autopolyploid  
199 origin. Another possibility that cannot be ruled out within the polyploids is the loss  
200 of one gene copy through homoeologous recombination or deletion.

201 All of the *Pgk1* gene sequences isolated in this study contain 5 exons and 4  
202 introns, covering a total length from 1391 bp to 1527 bp, which is consistent with  
203 previous studies of this gene in wheat [28] and *Kengyilia* [26]. The alignment of *Pgk1*  
204 sequences including both exons and introns resulted in a matrix of 1539 nucleotide  
205 positions, of which 11.6% (179/1539) were variable, and 10.1% (155/1539) were  
206 parsimony informative. The nucleotide frequencies were 0.264 (A), 0.304 (T), 0.199  
207 (C), and 0.232 (G). A significant (131-bp) insertion/deletion feature (Fig 1A) occurred  
208 at position 968, whereby all non-C genome type sequences contained the inserted  
209 (or non-deleted) region. Further analysis indicated that this region is likely an  
210 inserted inverted repeat, which belongs to the MITE stowaway element. Its  
211 secondary structure is shown in Fig 1B. This insertion/deletion event could be used  
212 as a genetic marker for rapid diagnosis of *Avena* species containing the C genome.



217 The substitution plot for *Pgk1* (Fig 2) indicated that the *Pgk1* gene was not  
218 saturated and that it could be used for phylogenetic analysis. Phylogenetic trees of 76  
219 *Avena* accessions with the oat-like species *Trisetopsis turgidula* as outgroup were  
220 generated through maximum parsimony and Bayesian inference approaches on the  
221 non-redundant dataset. The parsimony analysis resulted in 80 equally parsimonious  
222 trees (consistency index (CI) =0.632, retention index (RI) =0.954). BI analysis inferred  
223 an almost identical tree topology as the MP analysis, so the MP results were selected  
224 to describe this study (Fig 3).



225 **Fig 2. Saturation plot for transition and transversion of *Pgk1* gene sequences.** The  
226 crosses are the number of transition events; the triangles are the number of  
227 transversion events. The x axis shows the genetic distance based on the TN93 model;  
228 the y axis is the proportion of transitions or transversions, which was calculated by

229 using the number of transitions or transversions divided by the sequence length. The  
 230 curves show the trends of the variance of transitions and transversions with the  
 231 genetic distance increasing.



232 **Fig 3. Maximum parsimony tree derived from *Pgc1* sequence data.** The tree was  
 233 constructed using a heuristic search with TBR branch swapping. Numbers above and  
 234 below the branches are bootstrap support (BS) values  $\geq 50\%$  and Bayesian posterior



235 probability (PP) values  $\geq 90\%$ . Accession number, species name and haplome are  
236 indicated for each taxon.

237 Fig 3 shows that the *Pgk1* gene sequences from 76 *Avena* accessions were  
238 split into two distinct clades with high BS (100% and 92%) and PP (100% and 100%)  
239 support. One clade contained all C-genome type sequences, hence referred to as the  
240 C genome clade. The other clade contained all sequences from the species carrying  
241 the A genome, henceforth, referred to as the A genome clade. The C genome clade  
242 was composed of two major subclades. All  $C_v$  genome diploids and two  $C_p$  genome  
243 diploid accessions formed the subclade C1 with 63% BS and 91% PP support, while  
244 subclade C2 included four  $C_p$  diploids accessions, seven AC(DC) genome tetraploid  
245 accessions and nine hexaploid accessions with 71% BS and 97% PP support. The *Pgk1*  
246 gene sequences in the A genome clade were further split into six major subclades.  
247 The AC(DC) genome tetraploid species *A. insularis* was distinct from the other  
248 species, consequently forming a monophyletic clade (A1) with high BS (87%) and PP  
249 (100%) support. All five accessions of the  $A_c$  genome diploid species *A. canariensis*  
250 and one genome homoeologue of the AB genome tetraploid species *A. agadiriana*  
251 clustered together into subclade A2. Subclade A3 was composed of four accessions  
252 of the AB genome tetraploids *A. agadiriana*, five hexaploid accessions (*A.*  
253 *occidentalis* CN 23036 and CN 25942, *A. sativa* PI 194896 and PI 258655, *A. fatua* PI  
254 544659) and four  $A_s$  genome diploid accessions (*A. atlantica* CN25849 and CN 25859,  
255 *A. lusitanica* CN 26441, and *A. hirtula* CN 19739). One genome sequence of the  
256 AC(DC) genome tetraploids (without *A. insularis*) and the hexaploids formed a

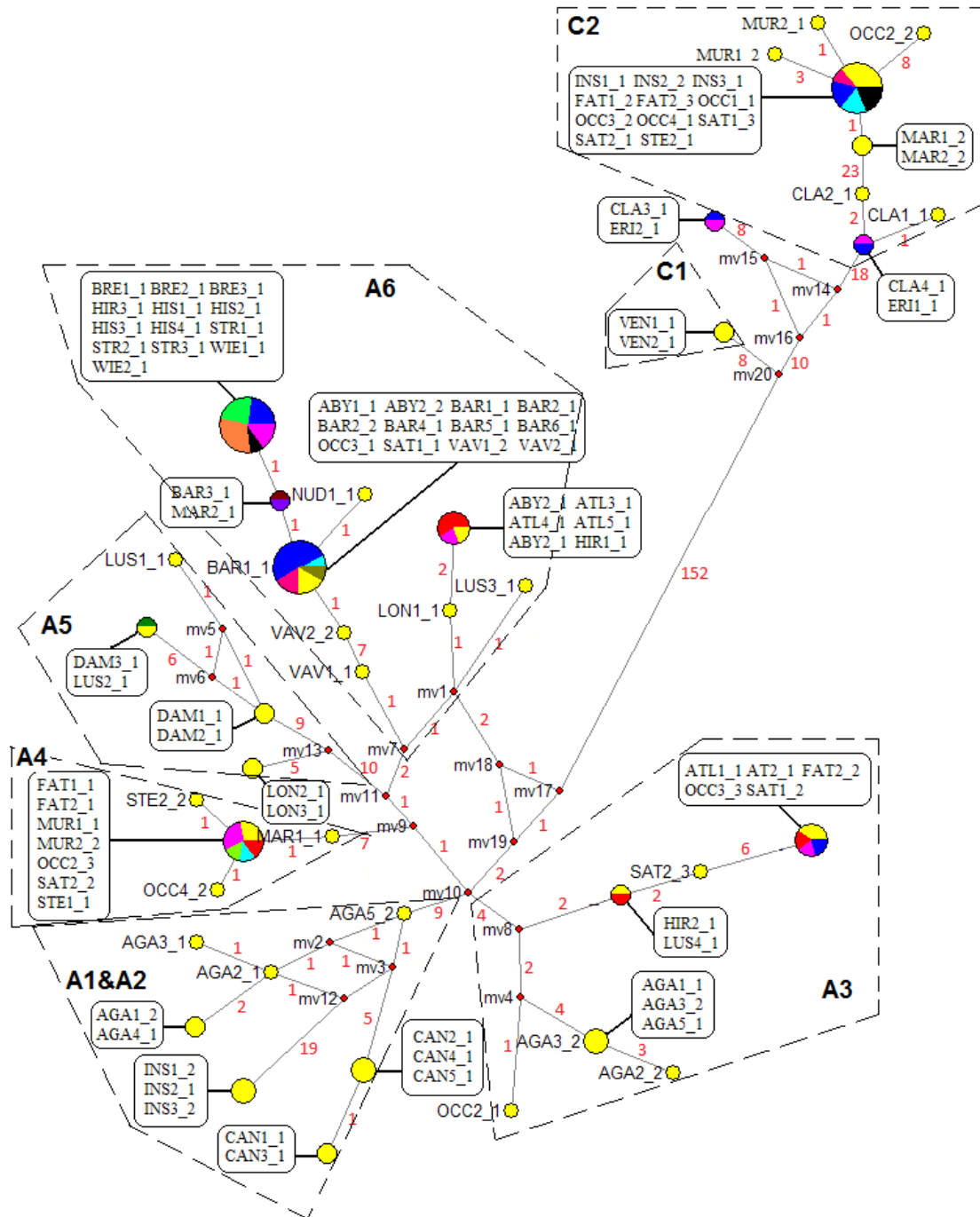
257 homogeneous clade (A4) that was separated from other species with high BS (100%)  
258 and PP (100%) support. The subclade A5 consisted of the A<sub>d</sub> genome diploid *A.*  
259 *damascena*, the A<sub>i</sub> genome diploid *A. longiglumis*, and the A<sub>s</sub> genome diploid *A.*  
260 *lusitanica*. The remaining sequences from the A genome diploids and the AB genome  
261 tetraploids (without *A. agadiriana*) formed a relatively broader cluster A6, together  
262 with two hexaploid accessions (*A. sativa* PI 194896 and *A. occidentalis* CN 25942)  
263 and one AC (DC) genome tetraploid accession (*A. maroccana* Clav 8831).

264 Three groups of haplotypes of *Pgk1* sequences were identified in five  
265 hexaploid accessions (*A. fatua* PI 544659, *A. occidentalis* CN 25942, CN 23036, and *A.*  
266 *sativa* PI 194896, PI 258655). These sequences fell into four subclades. One group  
267 clustered with the C genome diploids in subclade C2, and one group clustered with  
268 AC(DC) genome tetraploids in subclade A4. We hypothesize that these two types  
269 represent homoeologues from the C and D genomes, respectively. A third and fourth  
270 group fell into subclades A3 and A6. Since these two groups are highly separated, it  
271 is possible that they represent different A-genome events leading to different  
272 hexaploid lineages.

## 273 **Network analysis**

274 To gain better insight into relationships within closely related lineages, MJ  
275 network reconstruction based on the haplotypes of *Pgk1* sequences was employed.  
276 Due to the potential presence of recombination in the exon regions, the intron data  
277 was used for MJ network reconstruction. A total of 40 haplotypes were derived from  
278 104 *Pgk1* gene sequences (Fig 4). This low level of haplotype diversity demonstrates

279 the high conservation of this gene within genus *Avena*. The MJ network recovered a  
280 nearly identical phylogenetic reconstruction to that based on the MP and BI trees,  
281 therefore we identified the clades from the MP results (Fig 3) within the MJ network  
282 (Fig 4). Based on the topology and frequency of haplotypes, the MJ network was split  
283 into two main groups. The two major groups representing two distinct types of  
284 haplotypes (A and C genomes) were distinguished due to the 131 bp  
285 insertion/deletion. Ten C genome haplotypes were observed, which were much less  
286 diverse than the 30 A genome haplotypes. The two main groups were further  
287 subdivided into clusters corresponding to the eight MP-based subclades discussed  
288 earlier. The only divergence was that the AC(DC) genome tetraploids *A. insularis*,  
289 which formed a separate clade (A1) in MP and BI trees, fell into together with the AB  
290 genome tetraploid *A. agadiriana* and the A<sub>c</sub> genome diploid *A. canariensis* to form a  
291 relatively broad cluster in the MJ network (A1&A2).



292 **Fig 4. Median-joining networks based on 40 *Pgl1* gene haplotypes of intron regions**  
 293 **derived from 26 *Avena* species.** Each circular node represents a single haplotype,  
 294 with relative size being proportional to the frequency of that haplotype. Distinct  
 295 colors in the same haplotype node represent different species sharing the same  
 296 haplotype (colors are arbitrary). Median vectors (mv) represent the putative missing

297 intermediates. Numbers along network branches indicate the number of bases  
298 involved in mutations between two nodes. Clusters (surrounded by dashed lines) are  
299 named based on clade names shown in the MP tree (Fig 3). Three-letter  
300 abbreviations of species names are listed in Table 1. The numbers immediately after  
301 each species abbreviation represent different accessions of the same species, and  
302 the number following the underscore identifies different haplotypes from the same  
303 accession.

## 304 **Discussion**

### 305 **Two distinct diploid lineages exist in genus *Avena*.**

306 A significant 131 bp insert/deletion separated all *Avena* diploid species into  
307 two distinct groups representing the A and C genomes, respectively (Figs 1 and 4).  
308 These groups were also separated based on the MP or BI analysis that ignored gaps  
309 (Fig 3), indicating that the separation of A and C genomes is the most ancient major  
310 articulation in the genus *Avena*, a result that is consistent with most other literature  
311 [13, 14, 40]. MJ network analysis revealed that the C genome diploids have much  
312 lower levels of haplotype diversity than the A genome diploids. Within the C genome  
313 diploids, the C<sub>p</sub> genome haplotypes were relatively more diverse than those of the C<sub>v</sub>  
314 genome. These results might be explained by the geographic distribution of these  
315 species. The A genome diploids are distributed in a large region between latitude 20  
316 and 40° N, while the C genome diploid species are restricted to a narrow territory  
317 along the Mediterranean shoreline [1]. The geographic distributions of the C genome

318 diploid species are overlapping, but the range of the C<sub>p</sub> genome diploid species is  
319 much broader than that of the C<sub>v</sub> genome diploid species [41].

320 The A genome diploid species are the most diverse set of species in genus  
321 *Avena*, and chromosome rearrangements have occurred during the divergence of  
322 A-genomes from a common progenitor [41], resulting in the subdivision of the A  
323 genome into five types, of which we have investigated four. Our results showed that  
324 species with genome types A<sub>c</sub>, A<sub>l</sub>, and A<sub>d</sub> formed groups that correspond well with  
325 previously reported structural differences. However, the A<sub>s</sub> genome diploids appear  
326 to be much more diverse than previously reported, and are scattered into different  
327 subclades (Fig 3). Baum [1] divided all A<sub>s</sub> genome diploids into two sections, section  
328 *Agraria* and section *Tenuicarpa*. All species of section *Agraria* have florets with a  
329 domesticated (non-shattering) base, whereas the other A<sub>s</sub> species share relatively  
330 narrow spikelets. However, classification based on simple morphological traits is  
331 increasingly controversial. In this study, the A<sub>s</sub> genome diploid species of section  
332 *Agraria* showed high degree of genetic homogeneity, consistently forming their own  
333 subclade A6, but other A<sub>s</sub> genome species in section *Tenuicarpa* did not have their  
334 own subclade. *A. wiestii* showed a close relationship with the species of section  
335 *Agraria*, suggesting that it may be better-classified within that section. This result is  
336 in agreement with previous studies based on RAPD (Perchuk et al. 2002) and  
337 karyotypic comparisons (Badaeva et al. 2005). Accessions of the other two A<sub>s</sub>  
338 genome species of section *Tenuicarpa* (*A. atlantica* and *A. hirtula*) were scattered  
339 into different subclades. These results were also observed in other studies (Peng et

340 al. 2010, Yan et al. 2014). *A. lusitanica*, another  $A_s$  species of section *Tenuicarpa*, was  
341 diverged from other  $A_s$  species, but showed a close relationship to those with the  $A_d$   
342 genome species *A. damascena*. This divergence has also been observed in many  
343 other studies [8, 9, 14, 40]. These, and other incongruences between morphological  
344 characters and genetic differences raise questions about appropriate taxonomical  
345 classifications among  $A_s$  genome species.

### 346 **The $A_s$ and $A_c$ genomes played roles in the AB tetraploid** 347 **formation.**

348 Four recognized species have been proposed to have an AB genome  
349 composition. Of these, *A. barbata*, *A. abyssinica* and *A. vaviloviana* are grouped into  
350 a biological species known as the *barbata* group, while *A. agadiriana* is distinct [25,  
351 42]. Our results confirmed the reported structural differences between these two  
352 groups (Fig 3). Two different *Pgk1* gene sequences were detected from most of the  
353 AB genome tetraploids, supporting their allotetraploid origins. However, the  
354 genomes of *A. barbata* showed the least divergence, with only one of six *A. barbata*  
355 accessions providing multiple sequences, both of which were very similar. It seems  
356 that little divergence has occurred within the genome of *A. barbata* compared with  
357 that of *A. abyssinica* and *A. vaviloviana*, suggesting that *A. barbata* is the ancestral  
358 version of the species within the *barbata* group. This is supported by two lines of  
359 evidence. First, both *A. abyssinica* and *A. vaviloviana* are semi-domesticated forms  
360 that occur almost exclusively in Ethiopia, whereas the wild *A. barbata* are more  
361 geographically distributed, but can still be found close to the *abyssinica* and

362 *vaviloviana* forms [43]. The second line of evidence was provided by FISH and  
363 Southern hybridization [17], which found some B chromosomes of *A. vaviloviana* are  
364 involved in inter-genomic translocations, while these rearrangements were not  
365 detected in *A. barbata*. There is little doubt that the A genome diploids have been  
366 involved in the formation of the *barbata* species. Some studies have suggested that  
367 both the A and B genomes of *barbata* species are diverged  $A_s$  genomes [16, 23, 44],  
368 while some others proposed that the B genome might have originated from other A  
369 genome diploid species [24, 25, 45]. In this study, both types of *Pgk1* sequences  
370 detected from the *barbata* group showed high degree of genetic homogeneity with  
371 the  $A_s$  genome diploids (Fig 3), thus it was impossible to determine which type  
372 represents the A or B genome.

373         The recently discovered tetraploid species *A. agadiriana* was also proposed  
374 to have an AB genome composition because of its high affinity with *A. barbata* [23].  
375 However, this designation has been questioned due to chromosomal divergences  
376 between *A. agadiriana* and the *barbata* species, as revealed by cytological studies  
377 [45, 46] and by molecular data [9, 13, 14]. In the current study, two distinct types of  
378 *Pgk1* sequences were obtained in *A. agadiriana*. One copy clustered with the  $A_c$   
379 genome species *A. canariensis*, whereas the other copy fell into cluster A3 with the  
380  $A_s$  species *A. atlantica*, *A. hirtula*, *A. lusitanica*, and the hexaploids *A. occidentalis*, *A.*  
381 *fatua* and *A. sativa* (Fig 3). These results were in agreement with our previous  
382 studies based on nuclear *Acc1* gene [14] and GBS markers [9], and they support the  
383 proposal that *A. agadiriana* contains a different combination of A and/or B genomes



384 from the *barbata* group, and that one of its two genomes originates from the A<sub>c</sub>  
385 genome species *A. canariensis*, whereas the other one is closely related to the A<sub>s</sub>  
386 species.

387 **The tetraploid species *A. maroccana* and *A. murphyi* are**  
388 **closely related to the hexaploids, while *A. insularis* is**  
389 **diverged.**

390 The other tetraploid group (*Avena* sect. *Pachycarpa*) contains three species, *A.*  
391 *maroccana*, *A. murphyi*, and the recently discovered *A. insularis*. Initial studies based  
392 on genomic in situ hybridization [47] supported an AC genome designation for these  
393 species. However, this designation has been challenged by FISH analysis, which has  
394 revealed that this set of tetraploid species, like the D chromosomes of the hexaploid  
395 oats, lacks a repetitive element that is diagnostic of the A genome [18]. This,  
396 together with other molecular evidence [14, 48] and our recent whole-genome  
397 analysis based on GBS markers [9], suggests that these tetraploid species contain the  
398 genome designated as D in hexaploid oats, and that they are more properly  
399 designated as DC genome species.

400 In the present study, two distinct *Pgk1* homoeologues were detected in each  
401 of the three AC(DC) species, with each pair falling consistently into two clusters  
402 within the C and the A genome clades, respectively (Fig 3). The C-copy sequences of  
403 these tetraploids clustered consistently with the C-type homoeologues of the  
404 hexaploids, while the A/D genome homoeologues, with the exception of these from  
405 *A. insularis* and one sequence from *A. maroccana* (Clav 8331) fell into subclade A4

406 along with a set of sequences from the hexaploid oats (Fig 3). Considering that the  
407 other *Pgk1* gene sequences from the hexaploid oats clustered with the C or A  
408 genome diploids, we deduced that the sequences falling in subclade A4 must  
409 represent the D genome homoeologues of the hexaploids and of the AC(DC) species  
410 *A. maroccana* and *A. murphyi*. This result is not fully consistent with our previous  
411 GBS study: although *A. maroccana* and *A. murphyi* were very similar to hexaploid oat  
412 and were designated as DC genomes, our GBS work suggested that *A. insularis* was  
413 also a DC genome that was even more similar to the hexaploids [9]. Examining the  
414 existing literature, all three of these tetraploid species have variously been  
415 considered as the tetraploid ancestor of the hexaploids [4, 9, 49]. In view of the  
416 genome structure of these tetraploids [24, 50] and the meiotic chromosome pairing  
417 of their interspecific hybrids [51], all of these tetraploids are proposed to have  
418 diverged from a common ancestral tetraploid after the occurrence of some large  
419 chromosome rearrangements [24, 50]. However, it cannot be ruled out that these  
420 tetraploids might have originated independently from different diploid ancestors,  
421 since they have shown close relationships with different diploid species [40].  
422 Interestingly, in network analysis (Fig 4), the A/D-type homoeologues of *A. insularis*  
423 fell into a group with the  $A_c$  genome species *A. canariensis* and the AB genome  
424 species *A. agadiriana*. In fact, previous studies have revealed that *A. canariensis* is  
425 closely related to the DC genome tetraploids [15]. These results suggest a possibility  
426 that *A. canariensis* was involved in contributing an early version of a D genome in all  
427 three AC(DC) genome tetraploids. Nevertheless, we do not have an explanation for

428 why the D genome copy of *Pgk1* in *A. insularis* could have diverged so far from the  
429 version found in the hexaploids, especially since the C genome copies remain  
430 identical.

### 431 **The genome origins of the hexaploid species.**

432 It is now generally accepted that two distinct steps were involved in the  
433 evolution of hexaploid oats. The first step would have been the formation of a DC  
434 genome hybrid from ancestral D and C genome diploids, followed by doubling of the  
435 chromosomes to form an allotetraploid. The second step would have involved  
436 hybridization of a DC tetraploid with a more recent A genome diploid, followed by  
437 doubling of the triploid hybrid [9, 13].

438 The diploid progenitor of the hexaploid C genome was probably restricted to  
439 the narrow geographic range where the three extant C genome diploids are  
440 distributed. However, numerous inter-genomic translocations among hexaploid  
441 chromosomes [9, 11, 52, 53] have decreased the homology between the C genome  
442 diploids and the hexaploid C genome, making the identification of the C genome  
443 donor of the hexaploids challenging. In this study, the  $C_p$  genome species shared the  
444 highest degree of genetic similarity with both the DC genome tetraploids, as well as  
445 with the hexaploids, leading us to conclude that a  $C_p$  genome species was the C  
446 genome donor of the polyploids. This conclusion is supported by other evidence  
447 from nuclear genes [13, 54]. This is important, since it was recently demonstrated  
448 that the maternal tetraploid and hexaploid genomes originated from an A genome  
449 species, not from a C genome species [55], rendering comparisons to the  $C_v$  vs  $C_p$

450 maternal genomes irrelevant in determining the origin of the nuclear C genome in  
451 the hexaploids.

452 The A genome origin of the hexaploids remains a matter of debate, and many  
453 A genome diploids have been suggested as putative diploid progenitors, as  
454 summarized by Peng et al [13]. FISH analysis showed that an  $A_5$ -specific DNA repeat  
455 was restricted to the  $A_5$  and  $A_1$  genomes, as well as the hexaploid A genome [18]. In  
456 this study, a close relationship between the  $A_5$  genome diploid *A. atlantica* was  
457 observed for some hexaploid haplotypes in the phylogenetic tree (Fig 3) and the MJ  
458 network (Fig 4). An *A. atlantica* genome origin is consistent with previous studies  
459 based on IGS-RFLP analysis [12] and the *ppcB1* gene [40]. However, there is evidence  
460 in our work that some hexaploids may have an alternate A genome origin, closer to  
461 the *Agraria* section of  $A_5$  diploids. The presence of multiple A genome origins could  
462 explain variable results that have been reported in studies of hexaploid phylogeny.

463 In this study, strong evidence is presented for a D genome origin in the  
464 tetraploids *A. maroccana* and *A. murphyi* (Figs 3-4). However, these D genome  
465 sequences did not show a close relationship with any diploid species investigated in  
466 this study. Other than the discrepancy with *A. insularis*, this result is consistent with  
467 our recent GBS study [9]. One factor that may hinder the discovery of a D genome  
468 progenitor is the presence of inter-genomic translations among all three genomes in  
469 the hexaploid [9, 53]. Two hexaploid accessions (*A. occidentalis* CN 25942 and *A.*  
470 *sativa* PI 194896) did not contribute haplotypes that clustered with the putative D  
471 genome sequences (Subclade A4 in Fig 3). Although this may be a result of

472 incomplete sampling, it may also result from inter-genomic translations that have  
473 duplicated or eliminated copies of *Pgk1*.

474 In conclusion, this is the most comprehensive study to date that investigates  
475 a phylogeny in genus *Avena* using a single informative nuclear gene. It confirms or  
476 clarifies most previous work, and presents strong evidence in support of a working  
477 hypothesis for the origin of hexaploid oat. However, many questions still remain, and  
478 these will be best addressed through further studies involving multiple nuclear genes  
479 or whole genomes. We are collaborating on work that will provide exome-based  
480 gene diversity studies, but this work will require complete hexaploid reference  
481 sequences before it can be properly analyzed. Such reference sequences are  
482 currently in progress, so the next few years may see a revolution in our  
483 understanding of *Avena* phylogeny. Nevertheless, as many researcher in this field are  
484 aware, the polyploid species in this genus have experienced extensive chromosome  
485 rearrangement, which will continue to complicate phylogenetic studies. It may even  
486 be necessary to generate a pan-genome hexaploid reference sequence before  
487 definitive statements can be made.

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## 494 Reference

- 495 1. Baum BR. Oats: wild and cultivated. A monograph of the genus *Avena* L.(Poaceae):  
496 Minister of Supply and Services; 1977.
- 497 2. Baum BR, Fedak G. *Avena atlantica*, a new diploid species of the oat genus from  
498 Morocco. Canadian Journal of Botany. 1985;63(6): 1057-1060.
- 499 3. Baum BR, Fedak G. A new tetraploid species of *Avena* discovered in Morocco. Canadian  
500 Journal of Botany. 1985;63(8): 1379-1385.
- 501 4. Ladizinsky G. A new species of *Avena* from Sicily, possibly the tetraploid progenitor of  
502 hexaploid oats. Genetic Resources and Crop Evolution. 1998;45(3): 263-269.
- 503 5. Lin L, Liu Q. Geographical distribution of *Avena* L.(Poaceae). Journal of Tropical &  
504 Subtropical Botany. 2015;2: 111-122.
- 505 6. Rajhathy T, Thomas H. Cytogenetics of oats (*Avena* L.): Genetics Society of Canada;  
506 1974.
- 507 7. Thomas H. Cytogenetics of *Avena*. In: Marshall HG, Sorrells ME, editors. Oat Science and  
508 Technology. Agronomy Monograph. Madison, WI: American Society of Agronomy, Crop  
509 Science Society of America; 1992. pp. 473-507.
- 510 8. Yan H, Martin SL, Bekele WA, Latta RG, Diederichsen A, Peng Y, et al. Genome size  
511 variation in the genus *Avena*. Genome. 2016;59(3): 209-220.

- 512 9. Yan H, Bekele WA, Wight CP, Peng Y, Langdon T, Latta RG, et al. High-density marker  
513 profiling confirms ancestral genomes of *Avena* species and identifies D-genome  
514 chromosomes of hexaploid oat. *Theoretical and Applied Genetics*. 2016;129(11):  
515 2133-2149.
- 516 10. Rajhathy T, Thomas H. Chromosomal differentiation and speciation in diploid *Avena*. III.  
517 Mediterranean wild populations. *Canadian Journal of Genetics and Cytology*. 1967;9(1):  
518 52-68.
- 519 11. Chen Q, Armstrong K. Genomic in situ hybridization in *Avena sativa*. *Genome*.  
520 1994;37(4): 607-612.
- 521 12. Nikoloudakis N, Katsiotis A. The origin of the C-genome and cytoplasm of *Avena*  
522 polyploids. *Theoretical and Applied Genetics*. 2008;117(2): 273-281.
- 523 13. Peng Y-Y, Wei Y-M, Baum BR, Yan Z-H, Lan X-J, Dai S-F, et al. Phylogenetic inferences in  
524 *Avena* based on analysis of *FL* intron2 sequences. *Theoretical and Applied Genetics*.  
525 2010;121(5): 985-1000.
- 526 14. Yan H-H, Baum BR, Zhou P-P, Zhao J, Wei Y-M, Ren C-Z, et al. Phylogenetic analysis of  
527 the genus *Avena* based on chloroplast intergenic spacer *psbA-trnH* and single-copy  
528 nuclear gene *Acc1*. *Genome*. 2014;57(5): 267-277.
- 529 15. Fominaya A, Vega C, Ferrer E. Giemsa C-banded karyotypes of *Avena* species. *Genome*.  
530 1988;30(5): 627-632.
- 531 16. Katsiotis A, Hagedimitriou M, Heslop-Harrison JS. The close relationship between the A  
532 and B genomes in *Avena L.* (Poaceae) determined by molecular cytogenetic analysis of

- 533 total genomic, tandemly and dispersed repetitive DNA sequences. *Annals of Botany*.  
534 1997;79(2): 103-109.
- 535 17. Irigoyen M, Loarce Y, Linares C, Ferrer E, Leggett M, Fominaya A. Discrimination of the  
536 closely related A and B genomes in AABB tetraploid species of *Avena*. *Theoretical and*  
537 *Applied Genetics*. 2001;103(8): 1160-1166.
- 538 18. Linares C, Ferrer E, Fominaya A. Discrimination of the closely related A and D genomes  
539 of the hexaploid oat *Avena sativa* L. *Proceedings of the National Academy of Sciences*.  
540 1998;95(21): 12450-12455.
- 541 19. Chew P, Meade K, Hayes A, Harjes C, Bao Y, Beattie AD, et al. A study on the genetic  
542 relationships of *Avena* taxa and the origins of hexaploid oat. *Theoretical and Applied*  
543 *Genetics*. 2016;129(7): 1405-1415.
- 544 20. Linares C, González J, Ferrer E, Fominaya A. The use of double fluorescence in situ  
545 hybridization to physically map the positions of 5S rDNA genes in relation to the  
546 chromosomal location of 18S-5.8S-26S rDNA and a C genome specific DNA sequence in  
547 the genus *Avena*. *Genome*. 1996;39(3): 535-542.
- 548 21. Leggett J, Markhand G, editors. The genomic structure of *Avena* revealed by GISH.  
549 *Proceedings of the Kew Chromosome Conference IV*; 1995.
- 550 22. Baum BR, Rajhathy T, Sampson DR. An important new diploid *Avena* species discovered  
551 on the Canary Islands. *Canadian Journal of Botany*. 1973;51(51): 759-762.
- 552 23. Leggett JM, Thomas H. Oat evolution and cytogenetics. In: Welch RW, editor. *The Oat*  
553 *Crop World Crop Series*: Springer, Dordrecht; 1995. pp. 120-149.



- 554 24. Fominaya A, Vega C, Ferrer E. C-banding and nucleolar activity of tetraploid *Avena*  
555 species. *Genome*. 1988;30(5): 633-638.
- 556 25. Badaeva E, Shelukhina OY, Goryunova S, Loskutov I, Pukhalskiy V. Phylogenetic  
557 relationships of tetraploid AB-genome *Avena* species evaluated by means of  
558 cytogenetic (C-banding and FISH) and RAPD analyses. *Journal of Botany*. 2010; 2010.
- 559 26. Fan X, Sha LN, Zeng J, Kang HY, Zhang HQ, Wang XL, et al. Evolutionary dynamics of the  
560 *Pgk1* gene in the polyploid genus *Kengyilia* (Triticeae: Poaceae) and its diploid relatives.  
561 *Plos One*. 2012;7(2): e31122.
- 562 27. Sha LN, Fan X, Wang XL, Dong ZZ, Zeng J, Zhang HQ, et al. Genome origin, historical  
563 hybridization and genetic differentiation in *Anthosachne australasica* (Triticeae;  
564 Poaceae), inferred from chloroplast *rbcl*, *trnH-psbA* and nuclear *Acc1* gene sequences.  
565 *Ann Bot*. 2017;119(1): 95-107.
- 566 28. Huang S, Sirikhachornkit A, Faris JD, Su X, Gill BS, Haselkorn R, et al. Phylogenetic  
567 analysis of the acetyl-CoA carboxylase and 3-phosphoglycerate kinase loci in wheat and  
568 other grasses. *Plant Molecular Biology*. 2002;48(5-6): 805-820.
- 569 29. Chen Q, Kang HY, Fan X, Wang Y, Sha LN, Zhang HQ, et al. Evolutionary history of  
570 *Triticum petropavlovskyi* Udacz. et Migusch. inferred from the sequences of the  
571 3-Phosphoglycerate kinase gene. *Plos One*. 2013;8(8): e71139.
- 572 30. Ladizinsky G. *Avena prostrata*: a new diploid species of oat. *Israel J Bot*. 1971: 297-301.
- 573 31. Doyle J. A rapid DNA isolation procedure for small quantities of fresh leaf tissue.  
574 *Phytochemical Bulletin*. 1987;19(1): 11-15.

- 575 32. Thompson JD, Higgins DG, Gibson TJ. CLUSTAL W: improving the sensitivity of  
576 progressive multiple sequence alignment through sequence weighting, position-specific  
577 gap penalties and weight matrix choice. *Nucleic Acids Research*. 1994;22(22):  
578 4673-4680.
- 579 33. Xia X. DAMBE5: A comprehensive software package for data analysis in molecular  
580 biology and evolution. *Molecular Biology and Evolution*. 2013;30(7): 1720-1728.
- 581 34. Swofford DL. PAUP: Phylogenetic analysis using parsimony (and other methods). Version  
582 4.0b.10. Sunderland: Sinauer Associates. 2003.
- 583 35. Felsenstein J. Confidence limits on phylogenies: an approach using the bootstrap.  
584 *Evolution*. 1985;39(4): 783-791.
- 585 36. Huelsenbeck JP, Ronquist F. MRBAYES: Bayesian inference of phylogenetic trees.  
586 *Bioinformatics*. 2001;17(8): 754-755.
- 587 37. Bandelt HJ, Forster P, Röhl A. Median-joining networks for inferring intraspecific  
588 phylogenies. *Molecular Biology and Evolution*. 1999;16(1): 37-48.
- 589 38. Kilian B, Özkan H, Deusch O, Effgen S, Brandolini A, Kohl J, et al. Independent wheat B  
590 and G genome origins in outcrossing *Aegilops* progenitor haplotypes. *Molecular Biology*  
591 *and Evolution*. 2007;24(1): 217-227.
- 592 39. Pond SLK, Frost SDW, Muse SV. HyPhy: hypothesis testing using phylogenies.  
593 *Bioinformatics*. 2005;21(5): 676-679.
- 594 40. Liu Q, Lin L, Zhou X, Peterson PM, Wen J. Unraveling the evolutionary dynamics of  
595 ancient and recent polyploidization events in *Avena* (Poaceae). *Scientific Reports*.  
596 2017;7: 41944.

- 597 41. Loskutov IG, Rines HW. *Avena*. In: Kole C, editor. Wild crop relatives: genomic and  
598 breeding resources: Springer; 2011. pp. 109-183.
- 599 42. Drossou A, Katsiotis A, Leggett JM, Loukas M, Tsakas S. Genome and species  
600 relationships in genus *Avena* based on RAPD and AFLP molecular markers. Theoretical  
601 and Applied Genetics. 2004;109(1): 48-54.
- 602 43. Ladizinsky G. Studies in Oat Evolution: Springer Berlin Heidelberg; 2012.
- 603 44. Holden JHW. Species relationships in the Avenue. Chromosoma. 1966;20: 75-124.
- 604 45. Shelukhina OY, Badaeva ED, Brezhneva TA, Loskutov IG, Pukhalsky VA. Comparative  
605 analysis of diploid species of *Avena* L. using cytogenetic and biochemical markers:  
606 *Avena canariensis* Baum et Fedak and *A. longiglumis* Dur. Russian Journal of Genetics.  
607 2008;44(6): 694-701.
- 608 46. Jellen EN, Gill BS. C-banding variation in the Moroccan oat species *Avena agadiriana*  
609 ( $2n=4x=28$ ). Theoretical and Applied Genetics. 1996;92(6): 726-732.
- 610 47. Jellen EN, Gill BS, Cox TS. Genomic in situ hybridization differentiates between A/D- and  
611 C-genome chromatin and detects intergenomic translocations in polyploid oat species  
612 (genus *Avena*). Genome. 1994;37(4): 613-618.
- 613 48. Oliver RE, Jellen EN, Ladizinsky G, Korol AB, Kilian A, Beard JL, et al. New Diversity Arrays  
614 Technology (DART) markers for tetraploid oat (*Avena magna* Murphy et Terrell) provide  
615 the first complete oat linkage map and markers linked to domestication genes from  
616 hexaploid *A. sativa* L. Theoretical & Applied Genetics. 2011;123(7): 1159-1171.
- 617 49. Ladizinsky G, Zohary D. Notes on species delimitation, species relationships and  
618 polyploidy in *Avena* L. Euphytica. 1971;20(3): 380-395.

- 619 50. Shelukhina OY, Badaeva ED, Loskutov IG, Pukhal'sky VA. A comparative cytogenetic  
620 study of the tetraploid oat species with the A and C genomes: *Avena insularis*, *A. magna*,  
621 and *A. murphyi*. Russian Journal of Genetics. 2007;43(6): 613-626.
- 622 51. Ladizinsky G. Cytogenetic relationships between *Avena insularis* (2n=28) and both *A.*  
623 *strigosa* (2n=14) and *A. murphyi* (2n=28). Genetic Resources and Crop Evolution.  
624 1999;46(5): 501-504.
- 625 52. Irigoyen ML, Linares C, Ferrer E, Fominaya A. Fluorescence in situ hybridization mapping  
626 of *Avena sativa* L. cv. SunII and its monosomic lines using cloned repetitive DNA  
627 sequences. Genome. 2002;45(6): 1230-1237.
- 628 53. Sanz MJ. A new chromosome nomenclature system for oat (*Avena sativa* L. and *A.*  
629 *byzantina* C. Koch) based on FISH analysis of monosomic lines. Theoretical and Applied  
630 Genetics. 2010;121(8): 1541-1552.
- 631 54. Cheng DW, Armstrong KC, Drouin G, Mcelroy A, Fedak G, Molnar SD. Isolation and  
632 identification of *Triticeae* chromosome 1 receptor-like kinase genes (*Lrk10*) from diploid,  
633 tetraploid, and hexaploid species of the genus *Avena*. Genome. 2003;46(1): 119-127.
- 634 55. Fu YB. Oat evolution revealed in the maternal lineages of 25 *Avena* species. Scientific  
635 Reports. 2018;8(1): 4252.

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