# Supergene formation is associated with a major shift in genome-wide patterns of diversity in a butterfly

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**Abstract:** Selection shapes genetic diversity around target mutations, yet little is known about how 27 selection on specific loci affects the genetic trajectories of populations, including their genome-28 wide patterns of diversity and demographic responses. Adaptive introgression provides a way to 29 assess how adaptive evolution at one locus impacts whole-genome biology. Here we study the 30 31 patterns of genetic variation and geographic structure in a neotropical butterfly, *Heliconius numata*, and its closely related allies in the so-called melpomene-silvaniform subclade. H. numata is known 32 to have evolved a supergene via the introgression of an adaptive inversion about 2.2 million years 33 ago, triggering a polymorphism maintained by balancing selection. This locus controls variation in 34 wing patterns involved in mimicry associations with distinct groups of co-mimics, and butterflies 35 show disassortative mate preferences and heterozygote advantage at this locus. We contrasted 36 patterns of genetic diversity and structure 1) among extant polymorphic and monomorphic 37 populations of *H. numata*, 2) between *H. numata* and its close relatives, and 3) between ancestral 38 lineages in a phylogenetic framework. We show that *H. numata* populations which carry the 39 40 introgressed inversions in a balanced polymorphism show markedly distinct patterns of diversity compared to all other taxa. They show the highest diversity and demographic estimates in the entire 41 clade, as well as a remarkably low level of geographic structure and isolation by distance across the 42 entire Amazon basin. By contrast, monomorphic populations of *H. numata* as well as its sister 43 species and their ancestral lineages all show the lowest effective population sizes and genetic 44 diversity in the clade, and higher levels of geographical structure across the continent. This suggests 45 46 that the large effective population size of polymorphic populations could be a property associated with harbouring the supergene. Our results are consistent with the hypothesis that the adaptive 47 introgression of the inversion triggered a shift from directional to balancing selection and a change 48 in gene flow due to disassortative mating, causing a general increase in genetic diversity and the 49 homogenisation of genomes at the continental scale. 50

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**Introduction:** Genetic diversity is shaped by selective processes such as stabilizing or disruptive 52 selection, and by demographic processes such as fluctuations in effective population size. Empirical 53 studies on genetic diversity within and among populations abound, fuelled by an increasing 54 availability of whole genome data, and spurred by our interest in understanding the underlying 55 causes of variation in diversity (e.g. Beichmann 2018, Muers 2009; Murray 2017; Nielsen et al. 56 57 2009). At the locus scale, strong directional or disruptive selection tends to reduce diversity within populations (Mitchell-Olds et al. 2007), while balancing selection tends to enhance diversity 58 (Charlesworth 2006). Genome-wide factors reducing diversity include low effective population 59 sizes, generating drift, while high genetic diversity is enhanced by large population sizes and gene 60 flow. Overall, it is well recognised that demographic changes should have a genome-wide effect on 61 diversity, while positive selection is expected to play a role on the sites within and around the genes 62 involved in trait variation (Glinka et al. 2003, Muers 2009, Nielsen et al. 2009). 63

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65 Variation in behaviour and life-history traits, for instance involving changes in offspring viability or

dispersal distance, may also affect species demography, and thus whole genome genetic diversity.

67 However, whether and how genetic variability in a population may be driven by phenotypic

evolution at certain traits is poorly understood, and confounding effects may affect patterns of

69 genomic diversity, such as variation in census population size or colonization history. Dissecting

how selection on a trait may affect genome-wide diversity can be tackled by comparing closely related populations differing at this trait coupled with knowledge of when the differences evolved.

72 Here, we took advantage of the dated introgressive origin of a chromosomal inversion associated

with major life-history variation to study the demographics and whole genome consequences ofchanges in the selection regime at a major-effect locus.

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Heliconius butterflies are aposematic, chemically-defended butterflies distributed over the 76 American tropics from Southern Brazil to Southern USA (Emsley 1965; Brown 1979) (Fig 1A). 77 Heliconius butterflies are well-known for visual resemblance among coexisting species, a 78 relationship called Müllerian mimicry which confers increased protection from bird predators 79 through the evolution of similar warning signals (Sheppard et al. 1985). Most species are locally 80 monomorphic, but their mimicry associations vary among regions, and most species display a 81 geographic mosaic of distinct mimetic "races" through their range. In contrast to most *Heliconius* 82 species, the tiger-patterned Heliconius numata is well-known for maintaining both mimicry 83 polymorphism within localities, with up to seven differentiated coexisting forms, and extensive 84 geographic variation in the distribution of wing phenotypes (Brown & Benson 1974; Joron et al. 85 86 1999). Forms of *H. numata* combine multiple wing characters conveying resemblance to distinct sympatric species in the genus *Melinaea* and other local Ithomiini species (Nymphalidae: 87 Danainae). Polymorphism in *H. numata* is controlled by a supergene, i.e. a group of multiple linked 88 functional loci segregating together as a single Mendelian locus, coordinating the variation of 89 distinct elements of phenotype (Brown & Benson 1974; Joron et al. 2006). Supergene alleles are 90 characterized by rearrangements of the ancestral chromosomal structure, forming three distinct 91 chromosomal forms with zero (ancestral type, Hn0), one (Hn1) or three chromosomal 92 rearrangements (Hn123) (Fig 1B). The ancestral arrangement, Hn0, devoid of inversions, is fixed in 93 94 most *Heliconius* species (although an inversion in the same region evolved independently in a distantly-related *Heliconius* lineage (Edelman et al. 2019)). Arrangement Hn1 contains a 400kb 95 inversion called  $P_1$  originating from an introgression event about 2.2 My ago from *H. pardalinus*, in 96 which  $P_1$  is fixed (Jav et al. 2018). This introgression is thought to be the founding event triggering 97 the formation of the supergene and the maintenance of polymorphism in *H. numata* (Jay et al. 98 2018). Arrangement Hn123 displays two additional inversions, P<sub>2</sub> and P<sub>3</sub>, in linkage with P<sub>1</sub>, and 99 therefore originated after the introgression of P<sub>1</sub> into the *H. numata* lineage (Jay et al. 2021). 100 101

*Heliconius numata* is widespread in the lowland and foothill tropical forests of the Amazon basin, 102 103 the Guianas, and the Brazilian Atlantic Forest (Mata Atlântica), but the frequencies of the three chromosome arrangements vary across the range. Ancestral type Hn0 is fixed in the Atlantic Forest 104 populations of Brazil (forms robiqus or ethra), but segregates at intermediate frequencies in all 105 other *H. numata* populations throughout the range (forms *silvana* and *laura*) (Fig 1C). Chromosome 106 type Hn1 is associated with the Andean mimetic form *bicoloratus* and is found in the Eastern 107 Andean foothills of Ecuador, Peru, and Bolivia. Chromosome type Hn123 is associated with a large 108 diversity of wing-pattern forms of intermediate allelic dominance, including tarapotensis, arcuella 109 and aurora, and is reported from Andean, lowland Amazonian and Guianese populations. Inversion 110 polymorphism is therefore structured across the range, with populations being fixed for the 111 112 ancestral chromosome (Atlantic Forest, see Text S1 & Table S1-2), or displaying a polymorphism with two (Amazon-Guiana) or three (Andes) chromosomal types in coexistence (Joron et al. 2011). 113 Monomorphic populations of the Atlantic forest, devoid of rearrangements at the supergene locus, 114 might represent the ancestral state displayed by *H. numata* populations before the evolution of the 115 supergene via introgression (Fig 1C). 116

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118 The wing patterns of *H. numata* are subject to selection on their resemblance to local co-mimics

(Chouteau et al. 2016), but the polymorphism is maintained by balancing selection on the

120 chromosome types. Balancing selection is indeed mediated by disassortative mating favouring

- mixed-form mating (Chouteau et al. 2017) and is likely to have evolved in the response to the deleterious mutational load carried by inversions, which causes heterozygous advantage in *H*.
- *numata* (Jay et al. 2021, Faria et al. 2019, Maisonneuve et al. 2019). The introgression of  $P_1$  and the
- formation of a supergene were associated with a major shift in the selection regime and in the

mating system and may therefore have profoundly affected the population biology of the recipient

126 species, *H. numata*. We investigate here whether the adaptive introgression of a balanced inversion

is associated with a signature in the genetic diversity and geographic structure. We analyse changes

in the demographic history of the clade containing *H. numata* and closely related taxa, as well as

129 their current patterns of diversity and demography, using three well separated populations of *H*.

*numata* representing different states of inversion polymorphism. Our results are consistent with the
 selection regime and mating system associated with supergene formation having enhanced gene

132 flow among populations and increased effective population size. Moreover, our findings highlight

that balancing selection and a shift in mating systems associated with chromosomal polymorphism

- 134 may reshape genomewide diversity, with crucial consequences on current patterns of genetic
- 135 structure and population ecology.
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### 137 Material and Methods

138 We used here whole genome resequencing from 137 specimens of *Heliconius*, including 68 *H*.

*numata*. Sampling included specimens from populations in the Andean foothills (3 chromosome

types), from the upper Amazon (2 chromosome types), from French Guiana (2 chromosome types)

and from the Brazilian Atlantic Forests (1 chromosome type) (Fig 1C; Table S3). Related taxa were

represented by the sister species *H. ismenius*, found west of the Andes (parapatric to *H. numata*), by

143 Amazonian representatives of the lineage *H. pardalinus* (donor of the inversion), *H. elevatus*, *H.* 

*ethilla*, *H. besckei* as well as *H. hecale*, and by *H. melpomene* and *H. cydno* as outgroups. Only
Andean, Amazonian and Guianese populations of *H. numata* display chromosomal polymorphism,

all other taxa being fixed for the standard gene arrangement (Hn0), or for the inverted arrangement

147 Hn1 (*H. pardalinus*) (Jay et al. 2018). Hereafter, *H. numata* populations from the Andes, Amazon

and French Guiana will be collectively referred to as "Amazonian", and populations from the

149 Atlantic Forest as "Atlantic". Butterfly bodies were preserved in NaCl saturated DMSO solution at

- 150 20°C and DNA was extracted using QIAGEN DNeasy blood and tissue kits according to the
- 151 manufacturer's instructions with RNase treatment. Illumina Truseq paired-end whole genome
- 152 libraries were prepared and 2x100 bp reads were sequenced on the Illumina HiSeq 2000 platform.

Reads were mapped to the *H. melpomene* Hmel2 reference genome (Davey et al., 2016) using
 Stampy (version 1.0.28; Lunter and Goodson, 2011) with default settings except for the substitution

rate which was set to 0.05 to allow for the expected divergence from the reference of individuals in

the so-called silvaniform clade (*H. numata*, *H. pardalinus*, *H. elevatus*, *H. hecale*, *H. ismenius*, *H.* 

- 157 *besckei* and *H. ethilla*). *H. melpomene* and *H. cydno* belonging to the so-called *melpomene* clade,
- their genomes were mapped with a substitution rate of 0.02. Alignment file manipulations were
- 159 performed using SAMtools v0.1.3 (Li et al. 2009). After mapping, duplicate reads were excluded

using the *MarkDuplicates* tool in Picard (v1.1125; http://broadinstitute.github.io/picard) and local

indel realignment using IndelRealigner was performed with GATK (v3.5; DePristo et al. 2011).

162 Invariant and polymorphic sites were called with GATK HaplotypeCaller, with options --

163 min\_base\_quality\_score 25 --min\_mapping\_quality\_score 25 -stand\_emit\_conf 20 --heterozygosity

164 0.015.

#### 165

 $F_{ST}$ ,  $d_{XY}$  and  $\pi$ , were calculated in overlapping windows of 25 kb based on linkage disequilibrium 166 decay (*Heliconius* Genome Consortium 2012) using custom scripts provided by Simon H. Martin 167 (https://github.com/simonhmartin), and the genome-wide average was calculated using our own 168 169 scripts (available from https://github.com/angelesdecara). Distance in km between sampling sites was measured along a straight line, not taking into account potential physical barriers. The slopes of 170 F<sub>ST</sub> versus distance was calculated using the R package *lsmeans* (Lenth 2016); the slope difference 171 among species or between populations within species was estimated with an ANOVA and its 172 significance evaluated with function pairs of this package (Text S1 and see example script on 173 github.com/angelesdecara/). 174

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Admixture (Alexander et al. 2009) analyses were run on a subset of the 68 *H. numata* genomes,

keeping only 15 individuals from Peru to have a more balanced representation of individuals across

the geographic distribution. Filters were applied to keep biallelic sites with minimum mean depth of

179 8, maximum mean depth of 200 and at most 50% genotypes missing. We only kept 1 SNP per

180 kilobase to remove linked variants, and we obtained the optimal number of clusters using cross-

validation for values of K from 1 to 10 (Alexander et al. 2009). Principal component analyses

182 (PCA) were performed with the same filters as for admixture, using the same *H. numata* genomes

as for the admixture analyses, using smartpca (Patterson et al. 2006).

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In order to estimate demographic parameters independently of the effect of selection on diversity,we performed stringent filtering on the dataset. We removed all predicted genes and their 10,000

187 base-pair flanking regions, before performing G-PhoCS (Gronau et al. 2011) analyses as detailed

188 below. Repetitive regions were masked using RepeatMasker and Tandem Repeat Finder (Benson

189 1999). GC islands detected with CpGcluster.pl with parameters 50 and 1E-5 (Hackenberg et al.,

190 2006) were also masked. Scaffolds carrying the supergene rearrangements (Hmel215006 to

191 Hmel215028) were excluded, as were scaffolds from the sex chromosome (Z), since those are

expected to show unusual patterns of diversity due to selection and different effective populationsizes.

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195 We analysed the demographic history of *H. cydno*, *H. numata*, *H. ismenius*, *H. pardalinus* and *H. elevatus* with G-PhoCS, which allows for the joint inference of divergence time, effective 196 population sizes and gene flow. In order to detect differences in demography correlating with the 197 presence of the supergene in *H. numata*, we conducted analyses separating the Atlantic population 198 of *H. numata* from Amazonian populations. G-PhoCS is an inference method based on a full 199 coalescent isolation-with-migration model. Inferences are conditioned on a given population 200 phylogeny with migration bands that describe allowed scenarios of post-divergence gene flow. The 201 model assumes distinct migration rate parameters associated with each pair of populations, and 202 allows for asymmetric gene flow. Given the computational burden of G-PhoCS, we selected two 203 204 individuals per taxon or population, retaining those with the highest sequencing depth (see Table S3). The input dataset consisted of 4092 genomic regions, each 1kb in length and spaced at 205 approximately 30kb intervals and with genotypes in at least one of the two samples of each taxon 206 We used as priors for coalescence times ( $\tau$ ) and genetic diversity ( $\theta$ ), Gamma functions with  $\alpha$ =1 207 and  $\beta$ =100, and for migration bands  $\alpha$ =0.002 and  $\beta$ =0.00001. These priors were chosen to allow 208 good convergence while also ensuring non informativity. In order to calculate the highest posterior 209 density interval, we used the library HDInterval in R, and to integrate such posterior densities we 210

used the library sfsmisc in R. We rescaled the results using a mutation rate of 1.9E-9 (Martin et al.

212 2016) and 4 generations per year (i.e., g=0.25). Migration bands were considered significant

following the criteria of Freedman et al. (2012): if the 95% HPD interval did not include 0 or if the

total migration was larger than 0.03 with posterior probability larger than 0.5.

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#### 218 Results

Using cross validation error as a measure of the optimal number of clusters with Admixture, we 219 found that K=2 was the optimal cluster number describing within-species genetic variation in *H*. 220 numata (Fig 2A). One cluster corresponds to the Atlantic population, forming a well-differentiated 221 genetic entity compared to all other *H. numata* populations. All Amazonian populations of *H*. 222 numata showed a remarkable uniformity, with the exception of a few individuals sharing some 223 224 variation with SE Brazil. This pattern is consistent with the population structure inferred using microsatellite markers (Fig S1). Population structure revealed by PCA is in line with the admixture 225 analysis (Fig 2B). Individuals from the Atlantic population of *H. numata* clustered together to one 226 side of the first PCA axis, whereas all other individuals from all other populations clustered to the 227 other side. The second axis of the PCA separates individuals from French Guiana from the other 228 samples of the Amazon. This clustering was not found with Admixture (i.e. with K=3), suggesting 229 that the divergence between Amazonian populations is very reduced. In accordance, pairwise 230 genome-wide estimates of differentiation (F<sub>ST</sub>) between *H. numata* populations showed elevated 231 232 values when comparing the Atlantic population to other populations, but very small values when comparing pairs of Amazonian populations, even at a large distance (Fig 2C, Table S4). For 233 instance, the population from La Merced in Peru shows an  $F_{ST}=0.032$  with the population from 234 French Guiana at a distance of 3019km, but an  $F_{ST}$ =0.311 (an order of magnitude higher) with the 235 Atlantic population at a similar distance. Isolation by distance among Amazonian populations of *H*. 236 237 *numata*, estimated using the proxy  $F_{ST}/km$ , shows a very different pattern to other species, with a highly significantly shallower increase in F<sub>ST</sub> with distance in *H. numata* compared to all other taxa 238 (Fig 2C, Table S4). By contrast, differentiation as a function of distance between Atlantic and 239 Amazonian populations of *H. numata* is close to what is observed in other species, and not 240 241 significantly different (see Supp. Text S1).

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Analyses of genetic diversity show that all populations of *H. numata*, except those from the Atlantic 243 Forest, have a similar high genetic diversity (Fig 3A). By comparison, closely related *Heliconius* 244 taxa show significantly lower genetic diversity (Fig 3A). These patterns are similar to those 245 obtained using G-PhoCS to analyse the demographic histories in a phylogenetic context, where 246 Amazonian populations of *H. numata* show higher population sizes compared to the Atlantic 247 population (Fig 3B, Table S5). G-PhoCS analyses also show a demographic history in which gene 248 flow plays a crucial role (Table S6). For instance, our analyses show strong significant gene flow 249 250 right at the beginning of the divergence between H. ismenius and the other silvaniforms, as well as in the divergence between *H. pardalinus* and *H. elevatus*. The effective population sizes inferred 251 from Atlantic genomes are one order of magnitude lower than that obtained using H. numata 252 populations from other localities (Fig 3A and Table S5). In our cladogram, the increase in *H*. 253 254 *numata* population size is restricted to the Amazonian branch, excluding Atlantic populations. 255

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#### 257 Discussion

Our results suggest that populations displaying inversion polymorphism in the *P* supergene in *H*. 258 *numata* also display distinctive population demography and gene flow. Differences in demographic 259 and differentiation regimes associated with structural variation at this locus are revealed when 260 comparing polymorphic populations of *H. numata* to closely-related monomorphic taxa, such as (1) 261 peripheral populations of *H. numata*, (2) sister taxa, and (3) inferred ancestral lineages. This 262 suggests that the existence of a mimicry supergene controlling polymorphism in *H. numata* is 263 associated, in time and in space, with major differences in population biology. We hypothesize this 264 to be due to a change in the balancing selection regime due to heterozygote advantage (Jay et al. 265 2021) and in the associated evolution of disassortative mating (Chouteau et al. 2017) following the 266 onset of inversion polymorphism, causing direct effects on ecological parameters such as gene flow, 267 immigration success and effective population size. 268

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270 Our analyses show large-scale variation in genetic diversity among closely related taxa in this clade of *Heliconius* butterflies. Within *H. numata*, the genetic diversity of polymorphic Amazonian 271 populations is one to two orders of magnitude higher than the diversity found in populations from 272 the Atlantic Forest. Generally, Amazonian populations of *H. numata* harbour the highest genetic 273 diversity in the entire *melpomene*/silvaniform clade, which contrasts with the low diversity found in 274 the most closely related taxa such as *H. ismenius* or *H. besckei*. Inferring historical demography 275 during the diversification of the *H. numata* lineage reveals that the large effective population size in 276 that species is only associated with the branch representing polymorphic, Amazonian H. numata 277 populations, while internal branches all show very low diversity estimates. This suggests that 278 ancestral monomorphic populations of *H. numata* were similar in their diversity parameters to 279 current sister species H. ismenius populations, or to current peripheral Atlantic H. numata 280 populations. Although low-diversity lineages could have lost diversity due to recent events such as 281 strong bottlenecks, the distribution of parameters across lineages rather suggests that the 282 Amazonian populations of *H. numata* underwent a dramatic increase in effective population size 283 posterior to their split with Central American (H. ismenius) and Atlantic populations. The 284 Amazonian branch of the *H. numata* radiation is characterized by the long-term maintenance of 285 inversion polymorphism, triggered by the introgression of a chromosomal inversion about 2.2 Ma 286 287 ago. Therefore, the major shift in demography between Amazonian and Atlantic populations indeed appears associated with the occurrence of inversion polymorphism, even though the lack of 288 replication of this event impedes firmly establishing causality here. 289 290

Another striking result is the low genetic structure displayed by *H. numata* across the Amazon, with 291 all Amazonian and Guianese populations forming a single genetic cluster. Only Atlantic 292 populations stand out and display high differentiation with other *H. numata* from the rest of the 293 range. French Guiana and Peruvian populations, separated by over 3000 km across the Amazon, 294 are remarkably genetically similar compared to pairs of populations at comparable distances in 295 296 other species, and show similar differentiation as pairs of *H. numata* populations taken at short distances. *H. numata* populations from the Amazon show significantly lower isolation by distance 297 than all other taxa, as measured by the change in  $F_{ST}$  across distance ( $F_{ST}$ /km) (Fig. 2C), with a very 298 distinctive, flat slope of isolation by distance. The only exception is found when comparing 299 300 Amazonian populations with Atlantic populations of Brazil, displaying a level of differentiation in 301 line with that of pairs of populations at similar distances within other taxa. 302

Effective population size is affected by census size, mating system, and the force and type of 303 selection acting on traits (Charlesworth 2009). Selection is often viewed as a force only affecting 304 the genetic variation around specific, functional loci in the genome, but it may also affect whole 305 genome diversity, for instance when its action is sufficient to modify local demography or mating 306 patterns. In *H. numata*, morphs and therefore inversion genotypes show disassortative mate 307 preferences, i.e., they preferentially mate with individuals carrying different chromosome types 308 (Chouteau et al. 2017). Disassortative mating enhances heterozygosity and the mating success of 309 individuals expressing rare alleles (negative frequency dependence) (Knoppien 1985; Hedrick et al. 310 2018). Consequently, immigrants expressing rare, recessive alleles have a mating advantage in *H*. 311 *numata*. Their recessive effect on wing pattern lets them escape negative selection caused by their 312 inadequate mimicry patterns. Disassortative mating associated with the supergene should therefore 313 bring an advantage to immigrant genomes in LD with recessive supergene alleles, enhancing 314 genome-wide gene flow. This effective migration regime is quite different to that observed in other 315 mimetic taxa such as *Heliconius melpomene* or *H. erato*, in which mimicry variation is controlled 316 by multiple loci with diverse dominance patterns. In those taxa, hybrid offspring display 317 recombinant patterns breaking down mimicry, even after multiple generations of backcrossing, and 318 pure forms mate assortatively with respect to wing pattern (McMillan et al. 1997, Mallet et al. 1998, 319 Jiggins et al. 2001); both processes select against mimetic variants migrating from adjacent areas 320 with distinct warning patterns. In *H. numata*, the evolution of a polymorphic mimicry supergene 321 and disassortative mate preferences could therefore explain the relative lack, compared to other 322 *Heliconius* taxa, of differentiation among polymorphic populations, even across large distances. 323 324 Furthermore, enhanced gene flow could also cause an increase in effective population size estimates (Slatkin 1987), putatively explaining why polymorphic populations of *H. numata* harbour the 325 highest genetic diversity, and display the highest *Ne* estimates in the entire *melpomene*-silvaniform 326 clade of Heliconius. 327

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Alternative processes may of course contribute to the observed patterns. Amazonian and Atlantic 329 populations may differ in other aspects that could also result in differences in genetic diversity. 330 Habitat availability and structure may be different, possibly entailing differences in the maintenance 331 of diversity. The Atlantic Forest is vast in area, but may represent a smaller biome compared to the 332 333 Amazon, and is isolated from the bulk of the range of *H. numata*, which could result in a population ecology displaying characteristics of peripheral populations with smaller effective population sizes 334 (Eckert et al. 2008). The other Heliconius species in the clade have much in common with H. 335 *numata* in terms of habitat and general ecology, yet their niche and life-history specificities and 336 their phylogenetic histories may result in consistent differences with the polymorphic *H. numata* 337 populations. All those specificities may contribute to the observed pattern in which polymorphic 338 Amazonian populations of *H. numata* display high effective population size and a lack of 339 geographic structure in genome-wide genetic variation. Yet this pattern of variation correlates 340 parsimoniously with the evolution of a supergene causing disassortative mating in certain *H*. 341 342 numata populations, which provides an elegant mechanism explaining their differences with extant and ancestral closely-related lineages. However, we cannot rule out a role for conjectural 343 differences in ecology and geography with all other taxa. 344 345

In conclusion, our results show a remarkable contrast in the demography and differentiation of

347 populations within the Amazonian range of *H. numata* compared to closely related taxa and

348 ancestral lineages, as well as with other taxa in the *melpomene*/silvaniform clade. Although those

- 349 populations may differ in many uncharacterized ways from all other taxa, one known and consistent
- difference is the maintenance of inversion polymorphism associated with a specific mating system
- and selection regime in Amazonian *H. numata*. This distinctiveness of the only widely polymorphic populations in the clade is consistent with the hypothesis that the evolution of a supergene
- 352 populations in the clade is consistent with the hypothesis that the evolution of a supergene
- maintained by balancing selection represents a major transition in this lineage, triggering changes in genome-wide patterns of diversity and population ecology over the last 2 million years since its
- genome-wide patterns of diversity and population ecology over the last 2 million years since its formation. If this hypothesis is correct, the evolution of a locus under balancing selection may
- therefore feed-back on population ecology and diversification, and consequently on speciation.
- 357 More work on the determinants of variation in effective population sizes in the *Heliconius* genus is
- 358 needed to determine the precise impact of the supergene on demography of *H. numata*. We believe
- 359 that our results emphasize a potential link between genomic architecture, selection and demography,
- and should inspire future theoretical and modelling studies. Finally, the eco-evolutionary feedbacks
- 361 between changes in genomic architecture and the ecological parameters of populations are well-
- 362 known when considering self-incompatibility loci in plants, but may be more common than
- 363 previously thought. Indeed, our result suggests that balancing selection maintaining structural
- 364 polymorphisms affecting life-history traits may have a profound influence on species ecology.
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## 366 **Contributions:**

- MARdC, PJ and MJ designed the study and wrote the manuscript. BH, AVLF, TTT, RRR, KLSB
   provided the Atlantic samples. CS provided the Colombian samples. MARdC and PJ performed
- 369 genomic analyses with input from AW. MARdC, PJ, MJ, FPP and MC collected the Peruvian and
- 370 Ecuatorian samples. MC performed microsatellite analyses and organized fieldworks and butterfly
- rearing. All authors contributed to editing the manuscript.

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# 387 Data availability:

The raw sequence data were deposited in NCBI SRA and accession numbers are indicated in Supplementary table 3.

## 391 References

 Alexander DH, Novembre J, Lange K (2009). Fast model-based estimation of ancestry in unrelated individuals. *Genome Research* 19:1655-1664.

2. Beichmann AC, Huerta-Sanchez E, Lohmueller KE (2018). Using Genomic Data to Infer 394 Historic Population Dynamics of Nonmodel Organisms. Annual Review of Ecology, 395 Evolution, and Systematics 49:433–56 396 3. Benson G (1999) Tandem repeats finder: a program to analyze DNA sequences. *Nucleic* 397 Acids Research 27:573-580. 398 4. Brown KS (1979). Ecologia Geográfica e Evolução nas Florestas Neotropicais. – Univ. 399 Estadual de Campinas, Campinas, Brazil. 400 5. Brown KS, Benson WW (1974). Adaptive polymorphism associated with multiple müllerian 401 mimicry in *Heliconius numata* (Lepid.: Nymph.). *Biotropica* **6**:205–228 402 Brown KS, Mielke OHH. 1972. The Heliconians of Brazil (Lepidoptera: Nymphalidae). Part 6. 403 II. Introduction and general comments, with a supplementary revision of the tribe. 404 Zoologica, New York, 57:1–40. 405 7. Charlesworth B (2009) Fundamental concepts in genetics: effective population size and 406 patterns of molecular evolution and variation. Nature Reviews Genetics 10:195-205. 407 408 8. Chouteau M, Arias M, Joron M (2016). Warning signals are under positive frequencydependent selection in nature. Proceedings of the National Academy of Sciences of the USA 409 410 113:2164-2169. 9. Chouteau M, Llaurens V, Piron-Prunier F, Joron M. (2017). Polymorphism at a mimicry 411 supergene maintained by opposing frequency-dependent selection pressures. *Proceedings of* 412 the National Academy of Sciences of the USA 114: 8325-8329. 413 414 10. Davey JW, Chouteau M, Barker SL, Maroja L, Baxter SW, Simpson F, et al. (2016). Major Improvements to the Heliconius melpomene Genome Assembly Used to Confirm 10 415 Chromosome Fusion Events in 6 Million Years of Butterfly Evolution. *G3* **6**:695–708. 416 doi:10.1534/g3.115.023655 417 11. DePristo MA, Banks E, Poplin R, Garimella KV, Maguire JR, Hartl C, Philippakis AA, del 418 Angel G, Rivas MA, Hanna M, McKenna A, Fennell TJ, Kernytsky AM, Sivachenko AY, 419 Cibulskis K, Gabriel SB, Altshuler D, Daly MJ (2011). A framework for variation discovery 420 and genotyping using next-generation DNA sequencing data. *Nature Genetics* 43:491–498. 421 12. Eckert CG, Samis KE, Lougheed SC (2008). Genetic variation across species' geographical 422 ranges: the central-marginal hypothesis and beyond. *Molecular Ecology* 17:1170–1188. 423 13. Edelman NB, Frandsen PB, Miyagi M, Clavijo B, Davey J, Dikow RB, García-Accinelli G, 424 Van Belleghem SM, Patterson N, Neafsey DE, Challis R, Kumar S, Moreira GRP, Salazar 425 C, Chouteau M, Counterman BA, Papa R, Blaxter M, Reed RD, Dasmahapatra KK, 426 Kronforst M, Joron M, Jiggins CD, McMillan WO, Di Palma F, Blumberg AJ, Wakeley J, 427 Jaffe D, Mallet J (2019). Genomic architecture and introgression shape a butterfly radiation. 428 Science 366:594-599. 429 14. Emsley MG 1965. Speciation in *Heliconius* (Lep., Nymphalidae): morphology and 430 431 geographic distribution. Zoologica, New York 50:191–254. 15. Faria R, Johannesson K, Butlin RK, Westram AM (2019). Evolving inversions. Trends in 432 *Ecology & Evolution* **34**:239-248. 433 16. Freedman AH, Gronau I, Schweizer RM, Ortega-Del Vecchyo D, Han E, et al. (2012) 434 Genome Sequencing Highlights the Dynamic Early History of Dogs. PLoS Genetics 435 10:e1004016. 436

437 438 439	17.	Glinka S, Ometto L, Mousset S, Stephan W, De Lorenzo D (2003) Demography and natural selection have shaped genetic variation in Drosophila melanogaster: a multi-locus approach. <i>Genetics</i> <b>165</b> :1269-1278.
440	18.	Gronau I, Hubisz MJ, Gulko B, Danko CG, Siepel A (2011). Bayesian inference of ancient
441		human demography from individual genome sequences. <i>Nature Genetics</i> <b>43</b> :1031-1034.
442	19.	Hackenberg M, Previti C, Luque-Escamilla PL, Carpena P, Martínez-Aroza J, Oliver JL.
443		Bioinformatics 7:446.
445	20.	Hedrick PW, Tuttle EM, Gonser RA (2018) Negative-Assortative Mating in the White-
446		Throated Sparrow. Journal of Heredity 109:223-231.
447	21.	<i>Heliconius</i> Genome Consortium (2012). Butterfly genome reveals promiscuous exchange of
448		mimicry adaptations among species. <i>Nature</i> <b>487</b> : 94–8.
449	22.	Jay P, Whibley A, Frézal L, Rodríguez de Cara MÁ, Nowell RW, Mallet J, Dasmahapatra
450		KK, Joron M. (2018). Supergene evolution triggered by the introgression of a chromosomal
451		inversion. <i>Current Biology</i> <b>28</b> :1839-1845.
452	23.	Jay P, Chouteau M, Whibley A, Bastide H, Parrinello H, Llaurens V, Joron M. (2021).
453		Mutation load at a mimicry supergene sheds new light on the evolution of inversion
454		polymorphisms. <i>Nature Genetics</i> 53:288-293.
455	24.	Jiggins C, Naisbit R, Coe R, Mallet J 2001. Reproductive isolation caused by colour pattern
456		mimicry. <i>Nature</i> <b>411</b> :302–305.
457	25.	Joron M, Wynne IR, Lamas G, Mallet J (1999) Variable selection and the coexistence of
458		multiple mimetic forms of the butterfly <i>Heliconius numata</i> . <i>Evol Ecol</i> <b>13</b> : 721–754.
459	26.	Joron M, Papa R, Beltran M, Chamberlain N, Mavarez J, et al. (2006) A conserved
460		supergene locus controls colour pattern diversity in <i>Heliconius</i> butterflies. <i>PLoS Biology</i>
461		<b>4</b> :e303
462	27.	Joron M, Frezal L, Jones RT, Chamberlain NL, Lee SF, Haag CR, Whibley A, Becuwe M,
463		Baxter SW, Ferguson L, Wilkinson PA, Salazar C, Davidson C, Clark R, Quail MA,
464		Beasley H, Glithero R, Lloyd C, Sims S, Jones MC, Rogers J, Jiggins CD, ffrench-Constant
465		RH (2011). Chromosomal rearrangements maintain a polymorphic supergene controlling
466		butterfly mimicry. <i>Nature</i> <b>477</b> :203–206.
467	28.	Knoppien P (1985) Rare male mating advantage: a review. <i>Biological Reviews</i> <b>60</b> :81-117.
468	29.	Lenormand T (2002) Gene flow and the limits to natural selection. <i>Trends in Ecology and</i>
469		Evolution <b>17</b> :183-189.
470	30.	Lenth RV (2016). Least-Squares Means: The R Package lsmeans. <i>Journal of Statistical</i>
471		Software <b>69</b> :1-33.
472	31.	Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin
473		R; 1000 Genome Project Data Processing Subgroup (2009). The Sequence Alignment/Map
474		format and SAMtools. <i>Bioinformatics</i> <b>25</b> :2078-2079.
475	32.	Lunter G, Goodson M (2011). Stampy: a statistical algorithm for sensitive and fast mapping
476		of Illumina sequence reads. <i>Genome Research</i> <b>21</b> :936-939.
477	33.	Maisonneuve L., Chouteau M, Joron M, Llaurens V (2021). Evolution and genetic
478		architecture of disassortative mating at a locus under heterozygote advantage. <i>Evolution</i>
479		<b>75</b> :149-165.

480 481	34. Mallet J, McMillan W, Jiggins C (1998). Estimating the mating behavior of a pair of hybridizing <i>Heliconius</i> species in the wild. <i>Evolution</i> <b>52</b> :503–510.
482	35. Martin SH, Möst M, Palmer WJ, Salazar C, McMillan WO, Jiggins FM, Jiggins CD (2016).
483	Natural Selection and Genetic Diversity in the Butterfly <i>Heliconius melpomene</i> . <i>Genetics</i>
484	<b>203</b> :525-541.
485	36. Mitchell-Olds T, Willis JH, Goldstein DB (2007). Which evolutionary processes influence
486	natural genetic variation for phenotypic traits? <i>Nature Reviews Genetics</i> <b>8</b> :845–856.
487 488	<ul><li>37. Muers, M (2009) Separating demography from selection, <i>Nature Reviews Genetics</i> 10:280–281.</li></ul>
489	38. Murray GGR, Soares AER, Novak BJ, Schaefer NK, Cahill JA, Baker AJ, Demboski JR,
490	Doll A, Da Fonseca RR, Fulton TL, Gilbert MTP, Heintzman PD, Letts B, McIntosh G,
491	O'Connell BL, Peck M, Pipes ML, Rice ES, Santos KM, Sohrweide AG, Vohr SH, Corbett-
492	Detig RB, Green RE, Shapiro B (2017). Natural selection shaped the rise and fall of
493	passenger pigeon genomic diversity. <i>Science</i> <b>358</b> :951-954.
494	39. McMillan W, Jiggins C, Mallet J (1997). What initiates speciation in passion-vine
495	butterflies? Proceedings of the National Academy of Sciences of the USA <b>94</b> :8628–8633.
496	40. Nadeau NJ, Pardo-Diaz C, Whibley A, Supple M A, Saenko SV, Wallbank RWR et al.
497	(2016). The gene <i>cortex</i> controls mimicry and crypsis in butterflies and moths. <i>Nature</i> ,
498	<b>534</b> :106–110.
499	41. Nielsen, R., Hubisz, M.J., Hellmann, I., Torgerson, D., Andres, A.M., Albrechtsen, A.,
500	Gutenkunst R, Adams MD, Cargill M, Boyko A, Indap A, Bustamante CD, and Clark AG
501	(2009). Darwinian and demographic forces affecting human protein coding genes. <i>Genome</i>
502	Research <b>19</b> :838–849.
503	42. Patterson N, Price AL, Reich D (2006) Population Structure and Eigenanalysis. PLoS
504	<i>Genetics</i> <b>2</b> : e190.
505	43. Rosser N, Phillimore AB, Huertas B, Willmott KR, Mallet J (2012) Testing historical
506	explanations for gradients in species richness in heliconiine butterflies of tropical America.
507	Biological Journal of the Linnean Society <b>105</b> :479–497.
508	44. Schiffels S, Durbin R (2014) Inferring human population size and separation history from
509	multiple genome sequences. <i>Nature Genetics</i> <b>46</b> :919-925.
510	45. Saenko SV, Chouteau M, Piron-Prunier F, Blugeon C, Joron M, Llaurens V (2019)
511	Unravelling the genes forming the wing pattern supergene in the polymorphic butterfly
512	Heliconius numata. EvoDevo <b>10</b> :1-12.
513	46. Sheppard PM, Turner JRG, Brown KS, Benson WW, Singer MC (1985) Genetics and the
514	evolution of Muellerian mimicry in Heliconius butterflies. Philosophical Transactions of
515	the Royal Society of London, B Biological Sciences 308: 433–610
516	47. Slatkin M (1987) Gene flow and the geographic structure of natural populations. Science
517	<b>236</b> :787-792
518	



- 520 **Figure 1** | **Genetic and population structure at the P supergene.**
- 521 **A.** Schematic phylogeny of the sampled species. It includes all members of the silvaniform clade
- and two outgroups, *H. melpomene* and *H. cydno*.
- 523 **B.** Schematic description of the genetic structure of the P supergene. Three chromosomal
- arrangements coexist in *H. numata* and are associated with different morphs.
- 525 **C.** Origin of *H. numata* specimens used for analyses and distribution of chromosome arrangements
- 526 across the neotropics. Numbers in brackets indicate sampled specimens in each locality (the
- 527 Tarapoto population lumps several neighbouring subsamples on the map)





529 **A.** Admixture plot for *H. numata*. The optimal cluster number for *H. numata* is two, and it splits *H*.

*numata* into two categories, whereas they come from Atlantic forest or the Amazon. BR=Brazil

531 (Atlantic), PR=Peru, VE=Venezuela, CO=Colombia, EC=Ecuador, FG=French Guiana. **B.** 

532 Principal component analysis computed on whole genome SNP. **C.** Relationship between genetic

533 differentiation (Fst) and geographical distance. Fst is measured between morphs/populations of the

same species. *H. numata* populations from the Amazon show low isolation by distance when

535 compared to related species.



#### 537 Figure 3 | Variation in present and past effective population size in *Heliconius* species

538 **A.** Variation in Pi in several *Heliconius* populations, showing higher genetic diversity in *H. numata* 

- 539 populations from the Amazon than other taxa. Population names indicates their origin as in Figure 2
- (e.g. PR=Peru), with the addition of PA=Panama. The *H. numata* population with a lowest diversity
  is the one from the Atlantic forest (Brazil). **B.** Schematic representation of Gphocs results
- 542 (presented in table S5-6). Gene flow was modelled but not represented graphically for clarity.
- 543 showing that Amazonian populations of *H. numata*, which have the P supergene, show a dramatic
- increase in population size posterior to their split with the Atlantic populations of Brazil, which lack
- 545 the supergene.

## 546 List of Supplementary Materials:

- 547 Table S1-6
- 548 Fig S1-2
- 549 Text S1
- 550