

1 Impact of maternal intermittent fasting during pregnancy on cardiovascular, metabolic and
2 renal function in adult rat offspring

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4 Alaa Alkhalefah^{1,2}, Heather J. Eyre³, Rezwana Hussain², Jocelyn D. Glazier⁴, Nick Ashton^{1*}

5

6 ¹Division of Cardiovascular Sciences, Faculty of Biology, Medicine and Health, University of
7 Manchester, Manchester Academic Health Science Centre, Manchester, UK

8

9 ²Maternal and Fetal Health Research Centre, Division of Developmental Biology and Medicine,
10 St. Mary's Hospital, Faculty of Biology, Medicine and Health, University of Manchester,
11 Manchester Academic Health Science Centre, Manchester, UK

12

13 ³Division of Pharmacy and Optometry, Faculty of Biology, Medicine and Health, University of
14 Manchester, Manchester Academic Health Science Centre, Manchester, UK

15

16 ⁴Division of Evolution and Genomic Sciences, Faculty of Biology, Medicine and Health,
17 University of Manchester, Manchester Academic Health Science Centre, Manchester, UK

18

19 * Corresponding author

20 Email: nick.ashton@manchester.ac.uk (NA)

21 **Abstract**

22

23 Pregnant Muslim women are exempt from fasting during Ramadan; however a majority are
24 reported to fast. The impact of this form of maternal intermittent fasting (IF) on fetal
25 development and offspring health is not well defined. Using a rat model, we have shown
26 previously that maternal IF results in fetal growth restriction accompanied by changes in
27 placental nutrient transport function. The aim of this study was to assess cardiovascular,
28 metabolic and renal function in adult offspring of IF-exposed dams. Food was withheld from
29 Wistar rats from 17:00 to 09:00 daily throughout pregnancy; controls had *ad libitum* access to
30 food. Birth weight was unaffected; however male IF pups grew more slowly up to 10 weeks of
31 age ($P < 0.01$) whereas IF females matched their control counterparts. Systolic blood pressure
32 (SBP), glucose tolerance and basal renal function at 14 weeks were not affected by IF
33 exposure. When offered saline solutions (0.9-2.1%) to drink, females showed a greater salt
34 preference than males ($P < 0.01$); however there were no differences between dietary groups.
35 A separate group of pups was weaned onto a 4% NaCl diet. SBP increased in IF pups sooner, at
36 7 weeks ($P < 0.01$), than controls which became hypertensive from 10 weeks. Renal function
37 did not appear to differ; however markers of renal injury were elevated in IF males ($P < 0.05$).
38 Maternal IF does not affect resting cardiovascular, metabolic and renal function; but when
39 challenged by dietary salt load male IF offspring are more prone to renal injury.

40 **Introduction**

41

42 During the month of Ramadan, healthy adult Muslims are required to fast, abstaining from
43 both food and drink between sunrise and sunset. Pregnant women are exempt from fasting
44 and can elect to make up for any missed days at a later date or to pay for someone else to be
45 fed (Fidyah) [1]. However, evidence suggests that up to 90% of women take part in the daily
46 fast for at least part of their pregnancy [1]. The impact of maternal fasting on fetal
47 development and the subsequent health of the child is not well defined. We have reported
48 recently in a systematic review of the literature that maternal fasting during Ramadan is
49 associated with a reduction in placental weight; however birth weight was not affected [2].
50 Other more serious but infrequent events such as stillbirth or neonatal death have not been
51 reported, reflecting the small sample sizes of many studies, and there is a lack of longer term
52 follow up studies looking at the health of children or adult offspring.

53

54 The availability of nutrients in the diet and the capacity of the placenta to transport resources
55 to the developing fetus are essential for normal growth [3]. Studies on human populations
56 subject to famine, such as that which occurred during the Dutch Hunger Winter in 1944-45,
57 have shown that depending on the timing of exposure, fetal growth can be reduced [4]. Later
58 in life, the adult offspring were more likely to have an increased body mass index and be at
59 greater risk of impaired glucose tolerance, hypertension and coronary heart disease [4]. These
60 observations have been recapitulated in a variety of animal models, including both under- and
61 over-nutrition [5], demonstrating that the intrauterine environment is critical in determining
62 long-term health and disease. When the supply of nutrients is inadequate, particularly during
63 the later stages of gestation, resources are diverted to protect growth of the brain at the
64 expense of visceral organs such as the liver and kidneys [6]. As a result, metabolism and

65 excretory capacity are altered predisposing the offspring to diabetes and cardiovascular
66 disease [6].

67

68 These phenomena are of interest as women are reported to participate in the Ramadan fast
69 across each trimester of pregnancy [7-9], and differences in outcomes may be related to the
70 gestational timing when fasting occurred [2, 8, 10]. Additionally, the timing of food
71 consumption is altered and accompanied commonly by a change in the quantity and quality of
72 foods consumed [7]. Hence fasting pregnant women may expose their developing babies *in*
73 *utero* to an altered nutrient and metabolic environment [11, 12], which may link to the
74 increased propensity to long-term health issues such as coronary heart disease and type 2
75 diabetes reported previously in the offspring of mothers who observed Ramadan fasting
76 during pregnancy [10]. However, despite the now well-established link between maternal diet
77 during pregnancy, the intrauterine environment and the risk of developing a range of diseases
78 in adulthood, the potential impact of maternal fasting during Ramadan on offspring health
79 remains poorly defined.

80

81 Hence in order to study the impact of maternal intermittent fasting (IF) on the development
82 and subsequent health of the offspring, we have developed a rat model to mimic aspects of
83 human IF during Ramadan [13]. Pregnant rats were subjected to IF overnight, during their
84 active phase, for the duration of pregnancy in order to maximise the impact on the developing
85 fetus. IF fetuses were growth restricted at gestational day (GD) 21 and placental transport
86 efficiency (as evidenced by the fetal:placental weight ratio) was reduced. Consistent with this,
87 placental function was affected, with changes in placental metabolites and a significantly
88 reduced transplacental flux by the sodium-dependent system A amino acid transporter.
89 Exposure to IF altered fetal plasma amino acid profiles, with reductions in the branched chain

90 amino acids in particular, as well as a reduction in fetal insulin concentration. We also
91 observed sex differences in the fetal response to maternal IF, with sex-dependent differences
92 in placental aromatic amino acids also apparent within the IF group [13].

93

94 On the basis of these observations and the widely reported effects of maternal dietary
95 manipulation on offspring cardiovascular [14], metabolic [15] and renal function [16] in other
96 models of dietary-induced developmental programming, we postulated that exposure to IF
97 during pregnancy would have an adverse effect on these physiological functions in the adult
98 offspring. Therefore the aim of this study was to characterise postnatal growth and to assess
99 the impact of exposure to IF on blood pressure, glucose metabolism and renal function in the
100 adult offspring. IF rats were also challenged with a dietary salt load in order to expose any
101 potential dysfunction in cardio-renal regulation of blood pressure. Furthermore, as we have
102 observed sex differences in the response of fetuses to maternal IF [13], we studied both male
103 and female offspring to determine whether sex has any effect on the outcome.

104 **Methods**

105

106 **Ethical Approval**

107

108 All experiments involving animals were conducted under the authority of a project licence (PPL
109 40/3646) issued in accordance with the UK Animals (Scientific Procedures) Act 1986. Local
110 ethical approval was granted by the University of Manchester Animal Welfare and Ethical
111 Review Body. All animal work was conducted at the University of Manchester.

112

113 **Animals**

114

115 Virgin female Wistar rats (250-275 g, Charles River UK Ltd, Margate, Kent, UK) were
116 acclimatised to the Biological Services Facility for one week, where they were held under a 12
117 h dark:light cycle (06:00-18:00) at 21-23°C and 65% humidity. All rats had free access to
118 standard rat chow (BK001 (E) SDS Rodent Breeder and Grower, LBS Biotec, Redhill, UK) and
119 water. Females were then paired with a Wistar male (275-300 g) until conception was
120 confirmed by the presence of a vaginal plug: this was designated as gestational day 1 (GD1).
121 Females were then randomised to either intermittent fasting (IF, N = 25 rats) or control (C, N =
122 21 rats) groups and housed singly. Food was removed from the IF dams at 17:00 and returned
123 at 09:00 daily, commencing at GD1 until GD22 (term is at GD23); water was available *ad*
124 *libitum* throughout. Control animals had access to both food and water *ad libitum* throughout.
125 At term following birth, litter sizes were reduced to 8 animals (4 male and 4 female where
126 possible). All dams had free access to both food and water during the suckling period; pups
127 were weaned at 4 weeks of age onto standard chow and the dams were then killed by cervical
128 dislocation under isoflurane anaesthesia (4% isoflurane in oxygen at 2 L /min).

129

130 **Blood Pressure**

131

132 Systolic blood pressure (SBP) was measured by tail cuff plethysmography (Model LE5001,
133 PanLab, Spain) in conscious male and female rats at 5, 7 and 10 weeks of age (control n = 80
134 pups from N = 11 litters; IF n = 104 pups from N = 13 litters). These rats went forward for use in
135 one of the experiments described below, after which they were killed by cervical dislocation
136 under anaesthesia.

137

138 **Glucose and Insulin Tolerance Tests**

139

140 Glucose (GTT) and insulin (ITT) tolerance tests were conducted in separate groups of 12 week
141 old rats (control n = 5-6 pups per sex from N = 5-6 litters; IF n = 6-7 pups per sex from N = 6-7
142 litters). Rats were fasted overnight for 16 h prior to the collection of a pin-prick blood sample
143 from the tail for the measurement of baseline glucose concentration using an Accu-Chek
144 Mobile blood glucose monitoring system (Roche Diagnostics, West Sussex, UK). Animals then
145 received either an i.p. injection of sterile glucose solution (10% glucose in 0.9% saline at 1 g/kg
146 body weight for the GTT) or human insulin (0.75 unit/kg body weight, I9278, Sigma Aldrich for
147 the ITT), following which pin-prick blood samples were collected up to 120 min post-injection.
148 At the end of the experiment animals were killed by cervical dislocation under isoflurane
149 anaesthesia (4% isoflurane in oxygen at 2 L /min).

150

151 **Nephron Number**

152

153 Nephron number was determined at postnatal day (PD)1 (control n = 4-5 pups per sex from N
154 = 5 litters; IF n = 3-4 pups from N = 4 litters) and PD12 (control n = 6 pups per sex from N = 6
155 litters; IF n = 5-6 pups from N = 6 litters), as described previously [17]. Pups were killed by
156 decapitation using a method appropriate to their age (following stunning at PD1 or under
157 isoflurane anaesthesia at PD12). Kidneys were decapsulated, minced and digested in 10 mL 1
158 M HCl at 37°C for 3 (PD1) or 15 min (PD12), after which 50 mL deionised water was added and
159 homogenates were stirred gently at 4°C for 8 h. Mature glomeruli were counted in 20 x 40 µL
160 aliquots per kidney; comma and S shaped bodies were not included.

161

162 **Renal Function**

163

164 Renal function was measured in 14 week old offspring (control n = 5 pups per sex from N = 5
165 litters; IF n = 7 pups per sex from N = 7 litters) under Inactin anaesthesia (sodium
166 thiobutabarbital 100 mg/kg body weight i.p., T133, Sigma Aldrich), as described previously
167 [17]. A surgical plane of anaesthesia was confirmed through the absence of a pedal reflex; this
168 was checked at regular intervals throughout the experiment. A priming dose of clearance
169 markers (0.148 MBq ³H inulin, PerkinElmer, Monza, Italy and 12 mg para-aminohippuric acid
170 (PAH), A3759, Sigma Aldrich in 0.2 mL 0.9% saline) was administered intravenously following
171 which animals were infused continuously with 0.9% saline containing ³H inulin (0.0333
172 MBq/mL) and PAH (1 mg/mL) at 50 µL/min for a 3 h equilibration period. Thereafter urine
173 samples were collected via a bladder catheter at 15 min intervals over 3 h; arterial blood
174 samples (400 µL) were collected once every hour over 3 h. Blood pressure was recorded
175 continuously via a carotid artery catheter (Powerlab 800/s, ADInstruments, Hastings, East
176 Sussex, UK). Haematocrit was measured at the end of the experiment for the subsequent
177 calculation of effective renal blood flow. Urine and plasma samples were analysed for ³H inulin

178 activity (2000CA Tri-Carb Liquid Scintillation Analyser, Canberra Industries, Meriden, CT, USA),
179 PAH (standard colorimetric assay), Na⁺ and K⁺ concentrations (flame photometer model 420,
180 Sherwood Scientific Ltd, Cambridge, UK) and osmolality (Vapour pressure osmometer model
181 5500, Wescor, Inc, Logan, UT, USA); plasma was also analysed for protein concentration
182 (absorbance at 280 nm, Nanodrop 2000c spectrophotometer, Thermo Fisher Scientific,
183 Waltham, MA, USA). Animals were killed by cervical dislocation at the end of the experiment.

184

185 **Salt Preference and Aversion Threshold**

186

187 A two-bottle choice protocol was used to determine salt preference and total fluid intake, as
188 described previously [18]. At 7 weeks of age rats (control n = 5 pups per sex from N = 5 litters;
189 IF n = 6 pups per sex from N = 6 litters) were housed individually and offered bottles containing
190 sterile water or 0.9% saline. Following 2 days acclimatisation, daily fluid intake was recorded
191 over 5 consecutive days. The concentration threshold for salt aversion was subsequently
192 determined by offering the rats a choice between sterile water and increasing concentrations
193 of saline (from 0.9% to 2.1 % in 0.3% increments every 3 days over 12 days) to determine the
194 concentration at which they switched their preference from saline to water. At the end of the
195 experiment animals were killed by cervical dislocation under isoflurane anaesthesia (4%
196 isoflurane in oxygen at 2 L /min).

197

198 **Extracellular Fluid Volume**

199

200 Extracellular fluid volume was determined in 12 week old offspring (control n = 5 pups per sex
201 from N = 5 litters; IF n = 6 pups per sex from N = 6 litters) under Inactin anaesthesia (100 mg/kg
202 body weight, i.p.) as described previously [18]. Following a laparotomy, both sets of renal

203 vessels were occluded using 3-0 mersilk and the abdomen was closed. 0.222 MBq ^3H inulin in
204 350 μL 0.9 % saline was injected intravenously, followed by a saline flush (total volume of
205 injectate was 500 μL). Following a 90 min equilibration period, blood samples (50 μL) were
206 collected every 10 min over 60 min to measure plasma ^3H inulin activity and to calculate the
207 dilution of the injected ^3H inulin. Animals were killed by cervical dislocation at the end of the
208 experiment and a sample of urine was taken from the bladder to confirm occlusion of the renal
209 vessels.

210

211 **High Salt Diet**

212

213 The impact of dietary salt loading on SBP and renal function was determined in a separate
214 group of rats. These animals were either exposed to maternal IF *in utero* or a control *ad libitum*
215 diet as described above. Following weaning at 4 weeks, rats were randomised to receive either
216 a high salt (4% NaCl) diet (BK001 (E) 4% NaCl SDS Rodent Breeder and Grower, LBS Biotec,
217 Redhill, UK) or a standard (1% NaCl) rat chow diet (BK001 (E) SDS Rodent Breeder and Grower,
218 LBS Biotec, Redhill, UK) until 14 weeks of age (control n = 5 pups per sex per diet from N = 5
219 litters; IF n = 6 pups per sex per diet from N = 6 litters). SBP was measured as described above
220 at 5, 7 and 10 weeks of age. At 12 weeks of age the rats were housed individually in
221 metabolism cages until they had voided sufficient urine for analysis (3 mL typically collected
222 over 3 h). Renal function was then determined in anaesthetised rats at 14 weeks of age, as
223 described above. Animals were killed by cervical dislocation at the end of the experiment.

224

225 **Urine Analysis**

226

227 Urinary creatinine concentration was determined using a colorimetric assay, according to the
228 manufacturer's instructions (DetectX urinary creatinine kit, Arbor Assays, MI, USA). Urinary
229 albumin concentration was determined using a rat albumin ELISA kit, according to the
230 manufacturer's instructions (Bethyl Laboratories, Inc, TX, USA). Urinary neutrophil gelatinase-
231 associated lipocalin (NGAL) concentration was determined using a rat Lcn2 ELISA kit, according
232 to the manufacturer's instructions (RAB0906, Sigma Aldrich, UK).

233

234 **Statistical Analysis**

235

236 Data are presented as box (with median) and whisker plots (whiskers represent 5th and 95th
237 centiles) or as mean \pm SEM. N represents the dam or litter and n represents the offspring from
238 a litter. Where measurements were recorded for the whole litter (e.g. body weight) the litter
239 average is presented. In all other experiments data are representative of individual offspring.
240 No more than 2 rats of each sex from any given litter were included in an experimental group.
241 Distribution of the data was evaluated using a Shapiro-Wilk test, after which two-way ANOVA
242 (with repeated measures where appropriate) and Tukey tests or Kruskal-Wallis and Dunn's
243 multiple comparison tests were applied, as appropriate, where more than 2 groups were
244 compared. For comparisons between 2 groups Student's unpaired t-tests were used. Data
245 were analysed using SPSS (version 22.0, IBM SPSS Statistics, IBM United Kingdom Ltd,
246 Hampshire, UK) and GraphPad Prism (version 7.0, GraphPad Software, Inc, La Jolla, CA, USA);
247 statistical significance was taken as $P < 0.05$.

248 **Results**

249

250 **Maternal intermittent fasting impairs maternal weight gain and slows growth of male**
251 **offspring**

252

253 Pregnant rats subjected to IF ate $25 \pm 1\%$ less food over the course of gestation than controls
254 with *ad libitum* access to food ($P < 0.001$, S1A Fig). Their *ad libitum* water intake tended to be
255 lower too, reaching statistical significance ($P < 0.05$) over several days in the second half of
256 gestation (S1B Fig). As a result, IF dams gained significantly less weight than controls from
257 GD18 onwards ($P < 0.001$, S1C Fig). Despite this reduction in weight gain by IF dams, their litter
258 sizes and the body weight of new born pups did not differ significantly from that of control
259 dams (Table 1), even though all IF dams delivered ~ 0.5 day earlier than controls. Although
260 body weight did not differ between IF and control pups at PD1, organ growth, particularly that
261 of the brain, was impaired. The brain:liver weight ratios of both male (control $N = 9$, $1.20 \pm$
262 0.05 vs IF $N = 11$, 1.02 ± 0.03 , $P < 0.05$) and female (control $N = 9$, 1.19 ± 0.04 vs IF $N = 11$, 1.05
263 ± 0.02 , $P < 0.05$) IF rats was significantly lower than that of the controls. This reflected a
264 reduction in brain weight rather than liver weight (data not shown). Kidney weight was also
265 reduced significantly in both male (control $N = 9$, 73.6 ± 2.5 vs IF $N = 11$, 58.4 ± 2.0 mg/g body
266 weight, $P < 0.01$) and female (control $N = 9$, 72.1 ± 2.5 vs IF $N = 11$, 53.3 ± 1.7 mg/g body
267 weight, $P < 0.001$) IF rats compared with controls.

268

269 **Table 1 Litter size and pup birth weight at PD1 in control and IF offspring**

270

Control (N = 11)	IF (N = 13)
------------------	-------------

Litter size	15 ± 1	13 ± 1
Pup body weight (g)	6.5 ± 0.2	6.3 ± 0.2

271

272 Data are shown as mean ± SEM. Pup body weight represents the average weight per litter; N is
273 the number of litters. Statistical comparisons were by unpaired t-test. No significant
274 differences were identified.

275

276 Up to 16 days of age, the growth curves of control and IF pups were similar (Fig 1A). However,
277 from PD18 onwards IF pups gained significantly less weight than controls ($P < 0.001$). Pups
278 were divided into males and females when weaned at 4 weeks old. At this point it became
279 apparent that the lower rate of weight gain was driven by the males, since female IF pups
280 gained weight at the same rate as their control counterparts from weeks 5 to 12 whereas IF
281 males were significantly lighter than control males ($P < 0.05$, Fig 1B).

282

283 **Fig 1 Body weight prior to (A) and after weaning (B) up to 12 weeks of age in the offspring of**
284 **control and IF pregnancies.** (A) Body weight is shown as the litter mean up to weaning at 4
285 weeks of age (control N = 11 closed square, IF N = 13 open square). (B) Pups were then split by
286 sex (male circle, female square) and weight was recorded from 5 to 12 weeks of age (control N
287 = 10 closed symbols, IF N = 13 open symbols). Data are presented as mean ± SEM, except when
288 SEM falls within the size of the symbol. Statistical comparisons were by two-way ANOVA with
289 repeated measures and Tukey's test. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ IF vs control.

290

291 **Maternal intermittent fasting does not affect offspring blood pressure or insulin resistance**

292

293 Neither SBP nor heart rate differed between control and IF offspring at 5, 7 and 10 weeks of
 294 age (Table 2). Blood pressure tended to increase with age, but there were no differences
 295 between the dietary groups or between sexes. Glucose (Fig 2A) and insulin (Fig 2B) tolerance
 296 tests did not reveal any differences between the dietary groups or between sexes either. The
 297 area under the blood glucose concentration curve of male and female IF offspring, following
 298 injection of either glucose or insulin, did not differ significantly from that of their control
 299 counterparts.

300

301 **Table 2 Systolic blood pressure (SBP) and heart rate in control and IF offspring at 5, 7 and 10**
 302 **weeks of age**

303

	Control (N = 5)		IF (N = 6)	
	Male	Female	Male	Female
SBP (mmHg)				
Week 5	104 ± 2	102 ± 2	101 ± 2	103 ± 2
Week 7	112 ± 1	109 ± 1	110 ± 1	109 ± 1
Week 10	109 ± 2	108 ± 1	109 ± 1	107 ± 1
Heart Rate				
(bpm)				
Week 5	431 ± 8	430 ± 6	435 ± 10	433 ± 6
Week 7	440 ± 3	434 ± 4	439 ± 2	434 ± 4
Week 10	432 ± 3	429 ± 6	438 ± 2	434 ± 1

304

305 Data are shown as mean \pm SEM. Values represent the average per litter; N is the number of
306 litters. Statistical comparisons were by two-way ANOVA (SBP) or Kruskal-Wallis test (heart
307 rate). No significant differences were identified.

308

309 **Fig 2 Blood glucose concentration over 2 h following i.p. injection of glucose (A) or insulin (B)**
310 **in control and IF offspring at 12 weeks of age.** Following a 16 h overnight fast male (circle) and
311 female (square) rats were injected with either glucose (1 g/kg body weight, control n = 6
312 closed symbols, IF n = 7 open symbols) or insulin (0.75 unit/kg body weight, control n = 5, IF n =
313 6). Blood samples were collected over 120 min. Data are presented as mean \pm SEM. Area
314 under the curve was calculated and compared by two-way ANOVA (glucose) or Kruskal-Wallis
315 test (insulin). No significant differences were identified.

316

317 **Nephron number and basal renal function are not altered in IF offspring**

318

319 Nephron number increased with age ($P < 0.001$); but it did not differ between control and IF
320 offspring either at birth (PD1), when the kidney is still undergoing rapid nephrogenesis, or
321 when nephrogenesis was complete at PD12 (Table 3) [19].

322

323 **Table 3 Nephron number in control and IF offspring at PD1 and PD12**

324

	Control (N = 5-6)		IF (N = 4-6)	
	Male	Female	Male	Female
PD1	2625 \pm 217	2955 \pm 219	2513 \pm 117	2450 \pm 125
PD12	22938 \pm 497	22488 \pm 499	22288 \pm 570	22200 \pm 584

325

326 Data are shown as mean \pm SEM. Values represent the average per litter; N is the number of
327 litters. Statistical comparisons were by three-way ANOVA. Nephron number increased
328 significantly with age ($P < 0.001$); however no significant differences were identified within
329 each age group.

330

331 Renal clearance was used to assess baseline renal function in anaesthetised male and female IF
332 offspring at 14 weeks of age. In agreement with measurements made in conscious animals at
333 5-10 weeks of age (Table 2), mean arterial blood pressure recorded directly under anaesthesia
334 did not differ between IF and control rats of either sex (Table 4). Plasma sodium and protein
335 concentrations, and plasma osmolality did not differ significantly between groups (Table 4).
336 However, the plasma potassium concentration of male IF offspring was significantly higher
337 than that of control males ($P < 0.05$, Table 4). Haematocrit was significantly lower ($P < 0.05$,
338 Table 4) in female rats compared with their male counterparts.

339

340 **Table 4 Mean arterial pressure (MAP), plasma electrolyte concentrations, osmolality, protein**
341 **concentration and haematocrit in anaesthetised control and IF offspring during renal**
342 **clearance measurements at 14 weeks of age**

343

	Control (N = 5)		IF (N = 7)	
	Male	Female	Male	Female
MAP (mmHg)	125 \pm 4	121 \pm 3	119 \pm 5	129 \pm 3
Na ⁺ (mmol/L)	141 \pm 3	134 \pm 4	144 \pm 3	141 \pm 3
K ⁺ (mmol/L)	3.0 \pm 0.2	3.1 \pm 0.2	3.7 \pm 0.1*	3.0 \pm 0.1#
Osmolality (mOsm/kg)	357 \pm 11	357 \pm 9	363 \pm 8	361 \pm 6

H₂O)

Protein (g/100 mL)	4.5 ± 0.1	4.1 ± 0.1	4.3 ± 0.2	3.9 ± 0.2
Haematocrit (%)	46.8 ± 0.7	41.9 ± 1.0 [#]	50.1 ± 1.0	43.1 ± 1.2 ^{###}

344

345 Data are shown as mean ± SEM. Values represent the average per litter; N is the number of
346 litters. Statistical comparisons were by two-way ANOVA and Tukey's test or Kruskal-Wallis and
347 Dunn's multiple comparison test. * $P < 0.05$ IF vs control; # $P < 0.05$, ### $P < 0.001$ female vs
348 male.

349

350 The measured renal variables were stable over the 3 h experimental period, therefore for
351 clarity data are shown as the average over time. Effective renal blood flow (ERBF, Fig 3A) was
352 significantly higher in females compared with males ($P < 0.01$), but did not differ between IF
353 and controls. In contrast there were no significant differences in glomerular filtration rate
354 (GFR, Fig 3B) between the sexes, or dietary groups. Both urine flow rate (UV, Fig 3C) and
355 urinary sodium excretion rate ($U_{Na}V$, Fig 3D) were significantly greater in females compared
356 with males ($P < 0.05$), as were the potassium and osmolar excretion rates ($P < 0.01$, S2A, C Fig),
357 but there were no differences between IF and control offspring. Fractional excretion of sodium
358 was significantly higher in control females compared with control males (control male $N = 5$,
359 2.1 ± 0.4 vs control female $N = 5$, $4.2 \pm 0.7\%$, $P < 0.05$), which was not reflected by the IF
360 animals (IF male $N = 7$, 1.7 ± 0.3 vs IF female $N = 7$, $2.8 \pm 0.3\%$, $P > 0.05$), but there were no
361 differences in fractional excretion of potassium or free water clearance between the sexes or
362 dietary groups (S2B, D Fig).

363

364 **Fig 3 Effective renal blood flow (A), glomerular filtration rate (B), urine flow rate (C) and**
365 **sodium excretion rate (D) in anaesthetised control and IF offspring at 14 weeks of age. Renal**

366 haemodynamics (A-B) and urinary excretion rates (C-D) were measured over 3 h during
367 continuous infusion of 0.9% saline at 50 μ L/min in male and female control (N = 5 open boxes)
368 and IF (N = 7 shaded boxes) offspring. Statistical comparisons were by two-way ANOVA and
369 Tukey's test or Kruskal-Wallis and Dunn's multiple comparison test (sodium excretion). * $P <$
370 0.05, ** $P <$ 0.01, *** $P <$ 0.001 male vs female.

371

372 **Females have a greater preference for salt**

373

374 Salt preference and the salt aversion threshold were assessed using two-bottle choice tests.
375 When given the opportunity to choose between 0.9% saline and water to drink, female rats
376 not only drank more fluid per 100 g body weight than males ($P <$ 0.05), they also showed a
377 stronger preference for saline over water ($P <$ 0.01, Fig 4). Females of both groups drank 78-
378 87% more fluid in total, relative to body weight, than their male counterparts. The amount of
379 saline that they drank, as a proportion of the total fluid intake, was 33-54% greater than that
380 ingested by males. Thus females of both groups showed a stronger preference for 0.9% saline
381 than males.

382

383 **Fig 4 Salt preference and fluid intake in control and IF offspring at 7 weeks of age.** Male (M)
384 and female (F) rats were offered a choice of water or 0.9% saline as drinking fluid (control N =
385 5, open bar = water, hatched bar = saline intake; IF N = 6, open bar = water, solid bar = saline
386 intake). Data are shown as the mean + SEM of 5 consecutive days. Statistical comparisons
387 were by two-way ANOVA and Tukey's test. * $P <$ 0.05, ** $P <$ 0.01 total fluid intake by male vs
388 female; ## $P <$ 0.01 saline vs water intake.

389

390 Interestingly, the greater preference for 0.9% saline shown by control females was not
391 reflected by a higher threshold for salt aversion. When saline of increasing % was offered to
392 rats, the threshold at which they switched their preference from saline to water was between
393 1.5% and 1.8% for control and IF males, as well as IF females (S3 Fig). However, in control
394 females, the aversion threshold was significantly lower ($P < 0.01$), falling between 1.2% and
395 1.5% (S3C Fig).

396

397 Despite showing a greater preference for salt, the extracellular fluid volumes of female control
398 ($N = 4, 21.5 \pm 1.0$) and female IF rats ($N = 6, 23.5 \pm 1.3$) did not differ from their respective
399 male counterparts (control male $N = 4, 24.6 \pm 1.4$; IF male $N = 7, 21.8 \pm 1.0$ mL/100 g body
400 weight), nor were there significant differences between the dietary groups.

401

402 **Salt loading has sex-dependent effects on body weight and blood pressure**

403

404 In order to challenge the cardio-renal systems of IF rats, and thus identify any underlying
405 impairment, rats were weaned at 4 weeks of age onto a high salt diet containing 4% NaCl or a
406 control standard diet containing 1% NaCl. There were some fluctuations in food intake, but
407 overall all rats receiving a high salt diet ate similar quantities of food to their counterparts on a
408 standard salt diet, when adjusted for body weight (S4A, B Fig). In contrast, rats eating a high
409 salt diet drank significantly more water, per 100 g body weight, than their counterparts on a
410 standard salt diet (S4C, D Fig). However, there were no differences between the sexes or IF
411 and control rats eating the high salt diet. Control male rats were heavier and gained more
412 weight over the 7 week experimental period than control females; however dietary salt intake
413 did not affect the body weight of either sex (Fig 5A). In contrast, dietary salt had a sex-
414 dependent effect on body weight in IF rats. Males were heavier than females throughout, as

415 expected; however while IF males on a high salt diet gained more weight than their
416 counterparts on a standard salt diet ($P < 0.01$), IF females on a high salt diet gained less weight
417 than their counterparts on a standard salt diet ($P < 0.01$, Fig 5B).

418

419 **Fig 5 Body weight of control (A) and IF (B) offspring fed either a 1% or 4% salt diet from 4**
420 **weeks to 12 weeks of age.** Male (circles) and female (squares) control (N = 5 per diet) and IF
421 (N = 6 per diet) rats were weaned at 4 weeks of age onto a diet containing either 1% NaCl (NS
422 – normal salt, solid symbols) or 4% NaCl (HS – high salt, open symbols). Data are presented as
423 mean \pm SEM, except when SEM falls within the size of the symbol. Statistical comparisons were
424 by two-way ANOVA with repeated measures and Tukey's test. * $P < 0.05$, ** $P < 0.01$, *** $P <$
425 0.001 HS diet vs NS diet.

426

427 After one week on the high salt diet (5 weeks of age), SBP did not differ between those rats fed
428 4% NaCl (Table 5) and those on the standard diet (Table 2). Subsequently, IF rats responded
429 more robustly to the high salt diet than controls rats, with both sexes of IF offspring exhibiting
430 significantly higher ($P < 0.01$) SBP than their counterparts on a standard salt diet at 7 weeks of
431 age (Table 2 vs Table 5). SBP did not differ between control males on a high salt versus
432 standard salt diet at 7 weeks of age, and control females fed a high salt diet only showed a
433 modest, albeit significant increase in SBP ($P < 0.05$, Table 2 vs Table 5). It was not until 10
434 weeks of age that all rats fed a high salt diet exhibited significantly higher SBP ($P < 0.01$) than
435 their counterparts on a standard salt diet (Table 2 vs Table 5). Despite the difference in
436 timescales for the onset of hypertension when rats were fed a high salt diet, overall there were
437 no statistically significant differences between IF and control rats of either sex. Heart rate was
438 similarly unaffected (Table 5).

439

440 **Table 5 Systolic blood pressure (SBP) and heart rate in control and IF offspring fed a 4% NaCl**
441 **diet at 5, 7 and 10 weeks of age**

442

	Control (N = 5)		IF (N = 6)	
	Male	Female	Male	Female
SBP (mmHg)				
Week 5	108 ± 2	107 ± 2	106 ± 3	107 ± 3
Week 7	115 ± 2	114 ± 2	117 ± 2	116 ± 2
Week 10	120 ± 3	118 ± 2	123 ± 2	118 ± 3
Heart Rate				
(bpm)				
Week 5	437 ± 4	435 ± 3	443 ± 6	433 ± 4
Week 7	437 ± 2	433 ± 3	441 ± 2	432 ± 6
Week 10	436 ± 1	432 ± 4	441 ± 2	436 ± 5

443

444 Data are shown as mean ± SEM. Values represent the average per litter; N is the number of
445 litters. Statistical comparisons were by two-way ANOVA. No significant differences were
446 identified.

447

448 **Salt loading causes renal injury in male IF offspring**

449

450 After 8 weeks of salt loading, at 12 weeks of age, spot urine samples were collected to look for
451 markers of renal damage. The urinary albumin:creatinine concentration ratio was significantly
452 higher ($P < 0.05$) in IF males compared to IF females (Fig 6A). Male IF rats also showed
453 significantly elevated ($P < 0.05$) concentrations of NGAL, an early marker of renal injury [20],

454 compared with control males (Fig 6B). IF females tended to have higher urinary NGAL
455 concentrations compared with control females; however this did not reach statistical
456 significance ($P = 0.066$).

457

458 **Fig 6 Urinary albumin:creatinine concentration ratio (A) and urinary NGAL concentration (B)**

459 **in control and IF offspring fed a 4% salt diet from weaning until 12 weeks of age.** Male and

460 female control (N = 5 open boxes) and IF (N = 6 shaded boxes) rats were held individually in

461 metabolism cages until they had voided sufficient urine for analysis. Data are presented as box

462 (with median) and whisker plots (5th and 95th centiles). Statistical comparisons were by two-

463 way ANOVA and Tukey's test. * $P < 0.05$ IF vs control; # $P < 0.05$ male vs female.

464

465 Despite these increases in biomarkers of renal injury, there were no overt changes in renal

466 function when clearance measurements were made at 14 weeks of age. Mean arterial

467 pressure, plasma composition and haematocrit did not differ between IF and control rats

468 maintained on a high salt diet, nor were there any differences between the sexes (Table 6).

469 ERBF and GFR were comparable between IF and controls, as well as across the sexes (Fig 7A,

470 B), and UV and $U_{Na}V$ remained elevated in females compared with males ($P < 0.01$, Fig 7C, D).

471 Similarly, urinary potassium excretion and osmolar excretion were increased significantly ($P <$

472 0.01) in females compared with males (S5A, C Fig), while fractional excretion of potassium and

473 free water clearance were unaltered (S5B, D Fig). The only notable difference in renal function

474 in rats maintained on a high salt diet that was not observed in animals fed a standard diet was

475 that the fractional excretion of sodium by IF males (N = 6, $3.3 \pm 0.7\%$) was significantly lower

476 than that of IF females (N = 6, $6.1 \pm 0.5\%$, $P < 0.05$).

477

478 **Table 6 Mean arterial pressure (MAP), plasma electrolyte concentrations, osmolality, protein**
 479 **concentration and haematocrit in anaesthetised control and IF offspring fed at 4% NaCl diet**
 480 **during renal clearance measurements at 14 weeks of age**
 481

	Control (N = 5)		IF (N = 7)	
	Male	Female	Male	Female
MAP (mmHg)	125 ± 4	121 ± 4	116 ± 4	118 ± 3
Na ⁺ (mmol/L)	146 ± 4	147 ± 3	142 ± 6	146 ± 2
K ⁺ (mmol/L)	3.4 ± 0.2	2.9 ± 0.2	3.5 ± 0.2	3.2 ± 0.1
Osmolality (mOsm/kg H ₂ O)	328 ± 16	335 ± 13	334 ± 15	327 ± 11
Protein (g/100 mL)	4.3 ± 0.2	4.4 ± 0.2	3.8 ± 0.2	3.9 ± 0.2
Haematocrit (%)	57.1 ± 1.2	50.4 ± 1.1	56.5 ± 2.6	51.4 ± 2.2

482
 483 Data are shown as mean ± SEM. Values represent the average per litter; N is the number of
 484 litters. Statistical comparisons were by two-way ANOVA or Kruskal-Wallis test. No significant
 485 differences were identified.

486
 487 **Fig 7 Effective renal blood flow (A), glomerular filtration rate (B), urine flow rate (C) and**
 488 **sodium excretion rate (D) in anaesthetised control and IF offspring fed a 4% salt diet from**
 489 **weaning until 14 weeks of age.** Renal haemodynamics (A-B) and urinary excretion rates (C-D)
 490 were measured over 3 h during continuous infusion of 0.9% saline at 50 µL/min in male and
 491 female control (N = 5 open boxes) and IF (N = 6 shaded boxes) offspring. Data are presented as
 492 box (with median) and whisker plots (5th and 95th centiles). Statistical comparisons were by
 493 two-way ANOVA and Tukey's test. ** $P < 0.01$, *** $P < 0.001$ male vs female.

494 **Discussion**

495

496 Having shown previously that exposure to IF during pregnancy resulted in a number of changes
497 in maternal, fetal and placental function [13], the primary aim of the current study was to
498 establish whether the intrauterine challenge posed by IF leads to altered cardiovascular,
499 metabolic or renal function in the offspring. The picture that has emerged is that, in contrast to
500 other models of dietary manipulation during pregnancy in the rat such as the maternal low
501 protein (LP) model [14, 17, 21], IF appeared to have minimal impact on the offspring under
502 basal conditions. It was not until IF offspring were subject to a second, postnatal dietary
503 challenge in the form of salt loading that a susceptibility to renal injury was revealed in males.

504

505 We have reported previously that the ~30% reduction in food intake by pregnant rats
506 subjected to the IF regimen was associated with a significant reduction in fetal weight at GD21
507 [13]. In the current study we allowed rats to deliver at term (23 days) and observed that the
508 birth weight of IF pups was not different from that of controls, even though we consistently
509 found that IF dams delivered ~0.5 day early. The rat fetus normally undergoes rapid growth
510 over the final 2 days of gestation [22, 23]; nonetheless the magnitude of the increase in body
511 weight by IF fetuses is striking, bearing in mind that both fetal sexes exhibit FGR at GD21 [13].
512 While control fetuses increased their body weight by $79 \pm 6\%$ between GD21 and term at 23
513 days, IF fetuses gained $97 \pm 5\%$ in order to catch up and achieve a comparable weight to
514 control pups at birth. This pattern of growth restriction followed by rapid catch up growth
515 contrasts with other models of developmental programming, such as the maternal LP model in
516 which fetal weight was greater at GD21 yet birth weight was reduced compared with controls
517 [24]. In humans, we reported recently in a meta-analysis of the impact of Ramadan fasting
518 during pregnancy that birth weight was unaffected by maternal fasting [2]. Therefore while the

519 pattern of fetal growth in our IF rat model differs from other developmental programming
520 models, it appears to reflect that seen in human Ramadan fasting as evidenced by the lack of
521 impact on birth weight.

522

523 In early postnatal life, growth of IF pups matched that of controls; however from PD18 until
524 weaning at 4 weeks IF pups grew more slowly. Interestingly, rat pups begin to move away from
525 a diet comprising milk alone to one which includes solid food from around PD16-18 [25],
526 implying that the retarded growth of IF pups prior to weaning was not due to a reduction in
527 the quality or quantity of milk provided by the dam. Indeed, the growth retardation continued
528 beyond weaning until week 12. However this was only apparent in male offspring; IF females
529 grew at the same rate as their control counterparts, indicating that there may be sex-
530 dependent differences in nutrient and energy utilisation which are affected by the *in utero*
531 nutrient environment. In male offspring the mechanism may involve changes in mitochondrial
532 function: a reduction in muscle mitochondrial DNA (mtDNA) has been observed in male rats
533 exposed to a LP diet *in utero* [26]. In contrast mitochondrial function was increased male
534 protein-restricted mice, leading to greater oxidative capacity of muscle, increased energy
535 expenditure and diminished weight gain [27]. A different picture has been reported in female
536 offspring exposed to maternal undernutrition, where postnatal weight gain was associated
537 with leptin resistance [28]. Hence maternal dietary stress appears to affect different parts of
538 the energy regulation pathway in males and females.

539

540 Despite the pups having comparable body weights to controls at birth, the growth of key
541 organs was not proportionate in new-born IF pups. The brain:liver weight ratio was reduced in
542 IF pups, suggesting that brain growth had not been spared relative to visceral organ growth.
543 This contrasts with other models of food restriction during pregnancy in which growth of the

544 brain is spared at the expense of visceral organs such as the liver [29]. There are no published
545 data on the impact of maternal fasting during pregnancy on brain growth in humans. However,
546 neonatal head circumference as a proxy of brain growth did not show any difference between
547 infants of fasted and non-fasted mothers [30, 31]. Yet, it is interesting to note that a large
548 census-based study of Muslim populations in Uganda (n = 80,000) and Iraq (n = 250,000) has
549 shown that exposure to Ramadan fasting during the first month of pregnancy increased the
550 offspring's risk of 'mental or learning disability' by 50% and of 'psychological disability' by 63%
551 [7]. These surveys were not designed to identify the nature of any cognitive deficit associated
552 with maternal IF. However, nutritional challenges during pregnancy leading to low birth weight
553 [32] and slow postnatal growth [33] have been linked to impaired cognitive function in humans
554 in later life. It is interesting to note, therefore, that Muslim children living in England who were
555 exposed to Ramadan fasting *in utero* during the first trimester in particular achieved
556 significantly lower scores in the Key Stage 1 maths, reading and writing tests (taken in primary
557 schools at the age of 7 years) than either Muslim children who were not exposed to Ramadan
558 fasting *in utero* or to Caribbean children matched for socioeconomic status [34].

559

560 Kidney weight was also reduced in IF pups at birth, suggesting that renal development was
561 compromised. Despite this, nephron number was not different from that in controls either at
562 birth or later at PD12. The kidney is particularly vulnerable to nutritional insults during
563 pregnancy. Renal mass and nephron number have been reported to be lower in rat offspring
564 exposed to calorific restriction [35] and LP diets [17] during pregnancy. Nephron deficit in
565 particular has been linked to the development of high blood pressure [36], although there is
566 evidence to suggest that low nephron number and hypertension may be independent features
567 of the LP rat model [37]. We acknowledge that we did not use the gold standard method to
568 determine nephron number in the form of non-biased stereology; nonetheless we did not see

569 a difference between control and IF offspring either at birth when nephrogenesis is still
570 underway or at PD12 when nephrogenesis is complete in the rat [19]. The reason for the
571 apparent mismatch between reduced kidney weight at birth and unaltered nephron number in
572 IF offspring is unclear. Li *et al.* [38] have reported that pre-term babies exhibit
573 disproportionate growth of the renal compartments: the cortex had undergone hypertrophy
574 whereas the medulla was under developed compared with term babies at 6 months of age.
575 This raises the possibility that despite the apparent lack of a nephron deficit in IF offspring,
576 growth rates across the renal compartments may have differed between IF and control
577 kidneys. Further studies are necessary to confirm this notion. It is also of interest to note that,
578 in contrast to the food-only model of IF described in the current study, a 30% reduction in
579 nephron number has observed in rat offspring when pregnant dams were subjected to
580 intermittent restriction of both food and water for the whole of pregnancy [39]; however
581 shorter periods of food and water restriction (3 days which is equivalent to 1 month of human
582 pregnancy) had no effect on nephron count [40].

583

584 Blood pressure, first measured at 5 weeks and finally at 14 weeks at the end of the study, did
585 not differ between IF and control offspring of either sex. This is in marked contrast to other
586 models of developmental programming, including calorific restriction [41], high fat [42] and LP
587 diets [14] as well as fetal exposure to glucocorticoids [43], in which hypertension is a common
588 feature. It is possible that IF rats may go on to develop high blood pressure later in life: Kahn *et*
589 *al.* [42] reported that the female offspring of dams fed a high fat diet were not hypertensive
590 until 180 days old, and that males still had not developed high blood pressure at 360 days of
591 age. Conversely, we have reported previously that offspring exposed to a LP diet have elevated
592 blood pressure from as early as 4 weeks of age [17]. As there is variability in the timescales
593 over which hypertension can develop in rat models of developmental programming, we would

594 need to assess blood pressure in IF rat offspring over a longer period before we could confirm
595 that the offspring do not develop hypertension later in life.

596

597 Renal function did not differ between adult (14 week old) IF and control offspring under basal
598 conditions, which is in accord with the absence of any change in either blood pressure or
599 extracellular fluid volume. We have reported previously that renal haemodynamics, assessed
600 in the same manner as the current study, are unaltered in 4 week old LP rats [17], while others
601 have recorded a reduction in creatinine clearance (a marker of glomerular filtration rate) at
602 this age [16]. Glomerular filtration rate did not differ in older LP rats (20 weeks); however
603 albuminuria was present [16] indicating that the glomerular filtration barrier was damaged.
604 Renal tubular function in the LP rat does, however, appear to be impaired from an early age.
605 We observed a natriuresis and diuresis in LP rats aged 4 weeks, which appeared to be driven
606 by a reduction in Na⁺K⁺ATPase activity in the renal medulla [44]. Despite the loss of sodium, LP
607 rats had increased extracellular fluid volume [18] and raised blood pressure [44]. Hence it
608 appears that renal function and extracellular fluid volume regulation in the LP rat differs from
609 that in the IF model.

610

611 In an attempt to uncover any underlying deficit in kidney function, we challenged a separate
612 group of IF rats by weaning them onto a high (4%) salt diet prior to assessing their renal
613 function at 14 weeks of age. Salt loading revealed subtle, but important, differences between
614 IF and control animals. For example, the high salt diet resulted in comparable increases in
615 blood pressure in both IF and control animals. However, blood pressure in the IF rats began to
616 increase at an earlier age than their control counterparts. This was particularly apparent in the
617 IF males which became hypertensive at 7 weeks whereas blood pressure did not begin to
618 increase in control males until 10 weeks of age. Renal haemodynamics did not differ between

619 IF and control rats maintained on a high salt diet and although the urinary sodium excretion
620 rate was higher than that in animals maintained on a standard (1%) salt diet, it did not differ
621 between IF and control animals fed a high salt diet. Yet there were indications that the high
622 salt diet had affected the kidneys of male IF rats. The urinary albumin:creatinine concentration
623 ratio, a marker of glomerular damage [45], and urinary NGAL concentration, an early marker of
624 renal injury [20], were both elevated in IF males. NGAL is produced by injured renal tubules
625 and as a result of macrophage infiltration following inflammation [46], which is commonly
626 associated with dietary salt loading [47]. The marked increase in urinary NGAL concentration in
627 IF males therefore suggests that they are more susceptible to renal inflammation following salt
628 loading compared with their control counterparts. This in turn raises the possibility that IF
629 males, in particular, may be more prone to a deterioration in renal function later in life.

630

631 The only other differences in renal function that we observed in rats fed either a 1% or 4% salt
632 diet was between the sexes. This reflects, in part, the experimental design as infusion rates
633 were not adjusted for body weight. Rather, in common with many other studies of renal
634 function [48, 49] as well as previous reports from our laboratory [17, 44], urinary outputs were
635 adjusted for body weight instead. We acknowledge that this is a limitation of our study;
636 nonetheless if there had been sex-related differences in the IF animals the sex * diet
637 interaction term in the ANOVA would have been able to highlight such effects. Therefore, it is
638 unlikely that the approach taken obscured any sex-related effect of exposure to IF.

639

640 In addition to assessing the impact of salt loading on blood pressure and renal function, we
641 also determined the salt preference and aversion threshold of IF rats. When given a choice
642 between water and low concentration solutions of saline to drink, rats express a preference
643 for saline [50]. This salt preference is increased in several models of developmental

644 programming, including the LP rat [18], Dahl salt-sensitive rats exposed to a low sodium diet *in*
645 *utero* [51] and rats whose mothers underwent a partial aortic ligation [52]. Similarly maternal
646 dehydration, which can be induced in the rat by subcutaneous injection of polyethylene glycol
647 in order to mimic human vomiting during pregnancy, also increased salt preference in the
648 offspring [53]. It is interesting to note in this context that the risk of hyperemesis gravidarum
649 (morning sickness) is increased among pregnant women fasting during Ramadan, particularly
650 in the first trimester [54], suggesting perhaps that their offspring may have a greater
651 preference for salt. However in the IF rat model, no such enhanced preference for salt was
652 observed. Both male and female IF rats drank similar quantities of saline, as a proportion of
653 their total fluid intake, compared with their control counterparts. There was, however, a
654 strong sex difference. Both IF and control females drank more fluid relative to their body
655 weight compared with males, and of that fluid, the females drank significantly more saline (72-
656 79% of total fluid intake) compared with their male counterparts (51-54%). Similar sex-
657 dependent differences in salt preference have been noted before and have been attributed to
658 oestrogen-mediated blunting of salt-sensitivity in female rats [55].

659

660 Although there was no within-sex difference in the salt preference of IF and control rats, there
661 was a difference in the salt aversion threshold of IF females compared with control females.
662 Control females began to show an aversion to saline when the concentration reached 1.5%
663 whereas the IF females tolerated concentrations up to 1.8%. The literature in this area is
664 unclear: in males the point of indifference (the concentration of saline at which rats express
665 equal preference for water and saline) is reported to be at 1.5% in both neonatal and adult
666 rats; saline solutions of 3% or higher are rejected completely [56]. Female rats are able to
667 detect lower concentrations of saline than males, while also being more tolerant of higher
668 concentrations [55]. However, the aversion threshold for saline does not appear to have been

669 reported in female rats. Oestrogen has been shown to regulate salt-sensitivity in female rats
670 [55]; however we have no data on sex steroids or their receptors in IF rats upon which to base
671 a hypothesis to explain the observed difference in the salt aversion threshold.

672

673 Both male and female IF offspring responded to glucose and insulin challenges in the same
674 manner as control rats, indicating that they were not insulin-resistant at 12 weeks of age.
675 Impaired glucose tolerance and insulin resistance have been reported in other models of
676 developmental programming, including calorific restriction [57] and LP diets [15], but again age
677 seems to be critical. Hales *et al.* [58] reported that LP offspring were actually more glucose
678 tolerant than controls at 3 months of age; it was not until they were 15 months old that they
679 became glucose intolerant and exhibited frank diabetes [15]. MicroRNA (miRNA) may act as an
680 early marker of the risk of developing diabetes in later life. Ferland-McCollough *et al.* [59]
681 reported that expression of miRNA-483-3p, which regulates growth/differentiation factor 3
682 (GDF3), is upregulated as early as 22 days of age in LP rats and remained increased at 3 months
683 of age. These authors proposed that, as a result of increased expression of miRNA-483-3p,
684 GDF3 is downregulated which in turn affects the ability of adipose cells to store lipids and
685 ultimately leads to insulin resistance. It would therefore be of interest to assess miRNA-483-
686 3p expression in IF rats, as a potential early marker of insulin resistance and diabetes in later
687 life.

688

689 A limitation of this study is that the pregnant dams were fasted for the whole of pregnancy.
690 Ramadan takes place over a lunar month, which is approximately equivalent to 3 days of a rat
691 pregnancy. Ramadan could fall at any stage of a woman's pregnancy, so in order to assess the
692 effects of exposure to IF at all stages of fetal development and to maximise the potential
693 impact upon the offspring we elected to fast animals for the whole of pregnancy. This

694 approach has the advantage that it captures the potential impact of IF on the development of
695 multiple organ systems in the same animal, thereby reducing the overall number of animals
696 necessary in accordance with the 3Rs. However we acknowledge that it does not fully replicate
697 the pattern of fasting as practiced by Muslims; therefore in future studies a more targeted
698 approach will be taken, focussing on critical periods of development e.g. early pregnancy for
699 development of the brain or GD13-15 for development of the metanephros.

700

701 In conclusion, this study has shown that exposure to IF *in utero* appears to have minimal
702 impact on the cardiovascular, metabolic and renal health of adult offspring, contrasting with
703 offspring outcomes from other developmental programming models in which maternal dietary
704 intake is altered during pregnancy. Despite experiencing FGR as a result of altered placental
705 nutrient transport function [13], IF pups have comparable birth weights with controls. As they
706 grow, blood pressure remains normal, glucose and insulin tolerance are unaltered and basal
707 renal function is unaffected up to the age of 14 weeks. When challenged by a dietary salt load,
708 blood pressure begins to increase sooner in IF rats compared with controls; however the
709 magnitude of the overall hypertensive response is no different. The kidneys of IF rats appear to
710 be able to accommodate the increase in sodium intake; however there are indications that
711 they have undergone injury as male IF rats in particular exhibited albuminuria and had raised
712 urinary concentrations of the kidney injury marker, NGAL. Therefore, IF rats may be more
713 susceptible to renal injury.

714

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716

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920 **Supporting Information**

921

922 **S1 Fig Maternal food intake (A), water intake (B) and weight gain (C) in pregnant rats**
923 **throughout gestation.** Food was removed from IF rats (N = 13 open squares) for 16 h per day
924 between 17:00 and 09:00 from GD1 until GD22; water was available *ad libitum*. Control rats (N
925 = 11 closed squares) had free access to food and water at all times. Data are presented as
926 mean \pm SEM, except when SEM falls within the size of the symbol. Statistical comparisons were
927 by two-way ANOVA with repeated measures and Tukey's test. * $P < 0.05$, ** $P < 0.01$, *** $P <$
928 0.001 IF vs control.

929

930 **S2 Fig Potassium excretion rate (A), fractional excretion of potassium (B), osmolar excretion**
931 **rate (C) and free water clearance (D) in anaesthetised control and IF offspring at 14 weeks of**
932 **age.** Urinary excretion was measured over 3 h during continuous infusion of 0.9% saline at 50
933 $\mu\text{L}/\text{min}$ in male and female control (N = 5 open boxes) and IF (N = 7 shaded boxes) offspring.
934 Data are presented as box (with median) and whisker plots (5th and 95th centiles). Statistical
935 comparisons were by two-way ANOVA and Tukey's test. ** $P < 0.01$, *** $P < 0.001$ male vs
936 female.

937

938 **S3 Fig Salt aversion threshold in control (A, C) and IF (B, D) offspring.** Male (A, B) and female
939 (C, D) control (N = 5) and IF (N = 6) rats were offered a choice of water or saline increasing in
940 concentration from 0.9% to 2.1% as drinking fluid. Saline intake is shown as a percentage of
941 total fluid intake over 3 consecutive days for each concentration. Data are presented as mean

942 + SEM. Statistical comparisons were by one-way ANOVA and Dunnett's test. * $P < 0.05$, ** $P <$
943 0.01 , *** $P < 0.001$ vs 0.9% saline.

944

945 **S4 Fig Food intake (A, B) and water intake (C, D) of control and IF offspring fed either a 1% or**
946 **4% salt diet from 4 weeks to 12 weeks of age.** Male (circles) and female (squares) control (N =
947 5 per diet A, C) and IF (N = 5 per diet B, D) rats were weaned at 4 weeks of age onto a diet
948 containing either 1% NaCl (NS – normal salt, closed symbols) or 4% NaCl (HS – high salt, open
949 symbols). Data are presented as mean \pm SEM, except when SEM falls within the size of the
950 symbol. Statistical comparisons were by two-way ANOVA with repeated measures and Tukey's
951 test. * $P < 0.05$, *** $P < 0.001$ HS diet vs NS diet.

952

953 **S5 Fig Potassium excretion rate (A), fractional excretion of potassium (B), osmolar excretion**
954 **rate (C) and free water clearance (D) in anaesthetised control and IF offspring fed a 4% salt**
955 **diet from weaning until 14 weeks of age.** Urinary excretion was measured over 3 h during
956 continuous infusion of 0.9% saline at 50 $\mu\text{L}/\text{min}$ in male and female control (N = 5 open boxes)
957 and IF (N = 6 shaded boxes) offspring. Data are presented as box (with median) and whisker
958 plots (5th and 95th centiles). Statistical comparisons were by two-way ANOVA and Tukey's test.
959 * $P < 0.05$ IF vs control; ## $P < 0.01$, ### $P < 0.001$ male vs female.

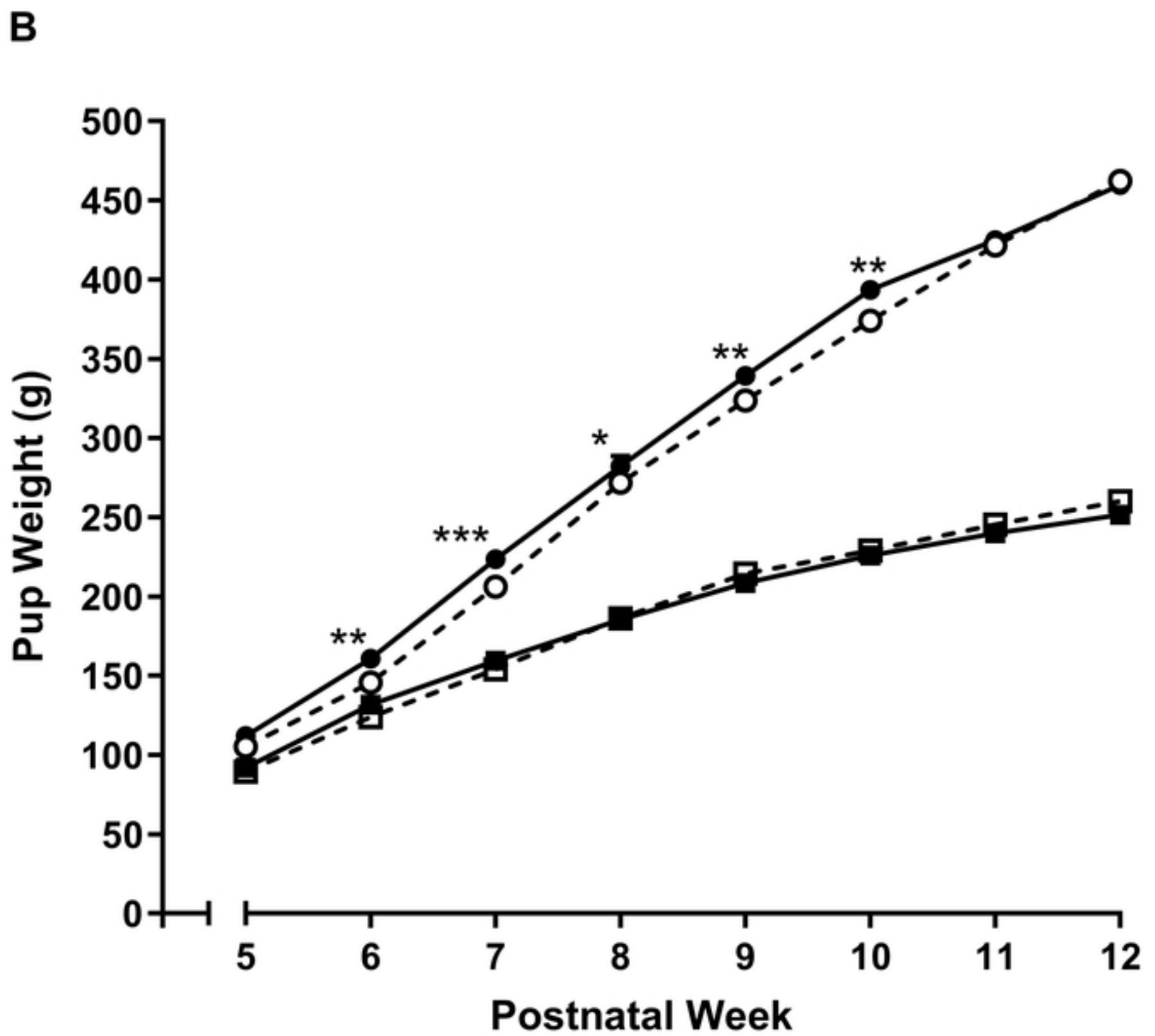
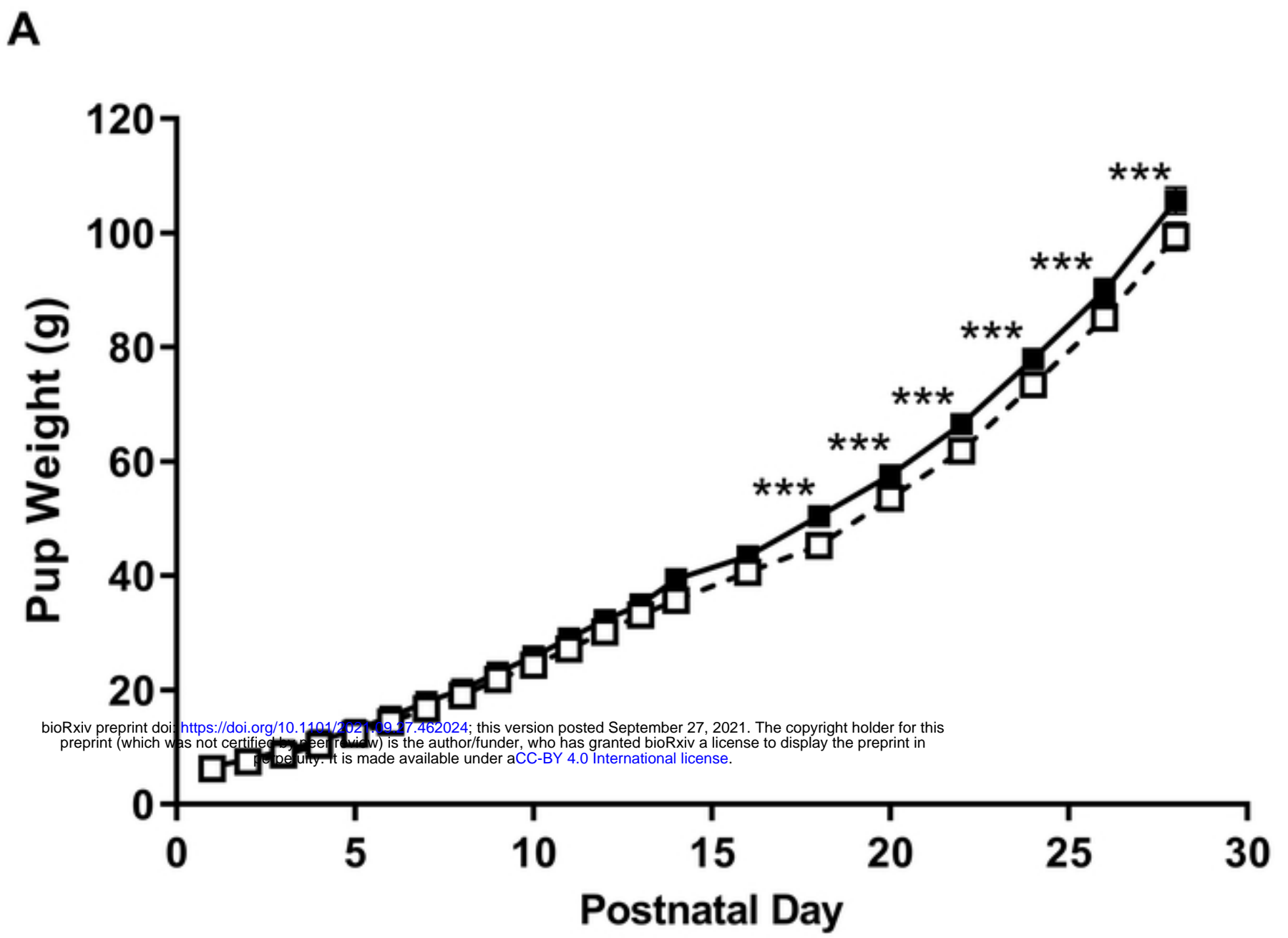


Fig 1

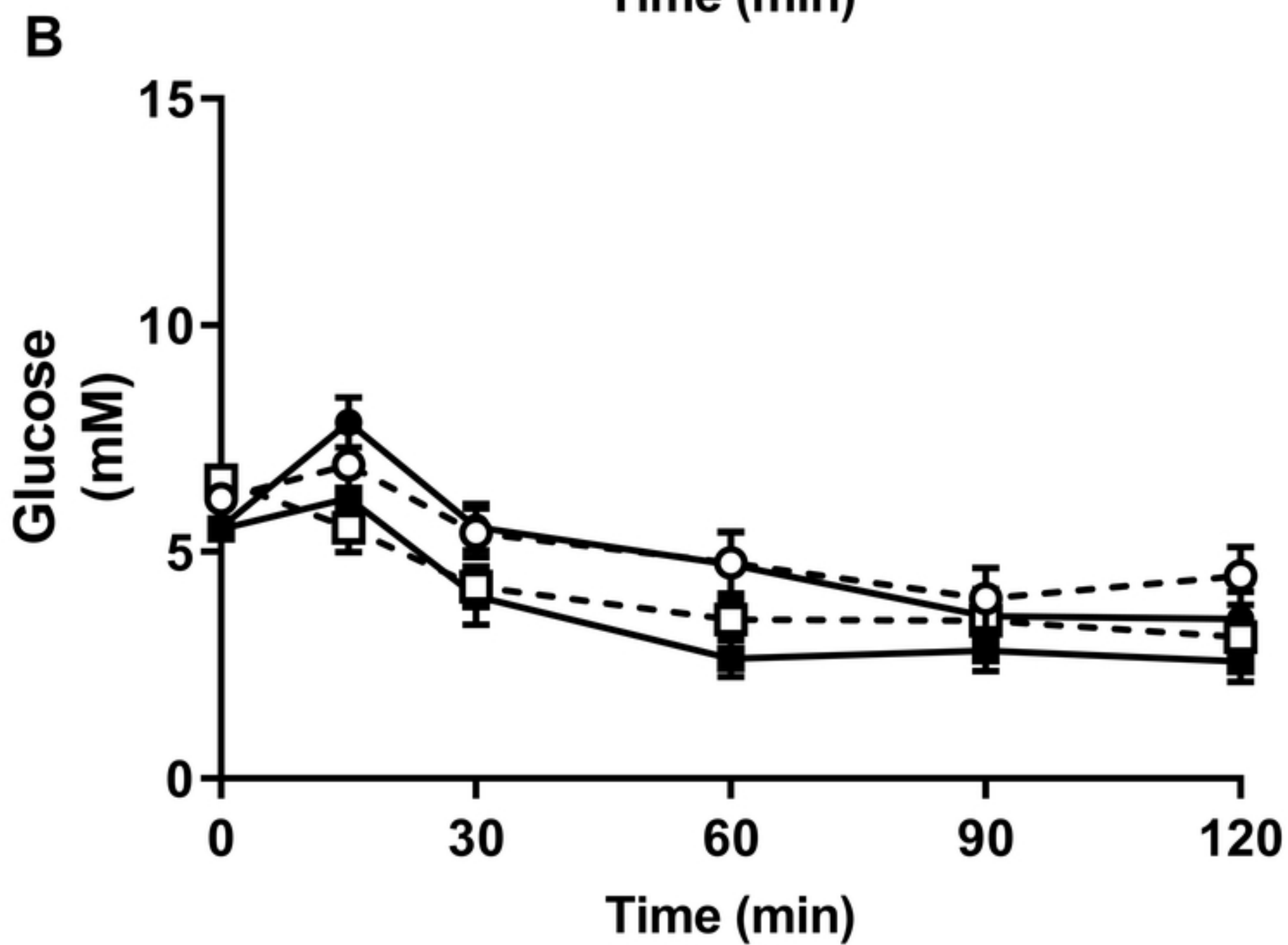
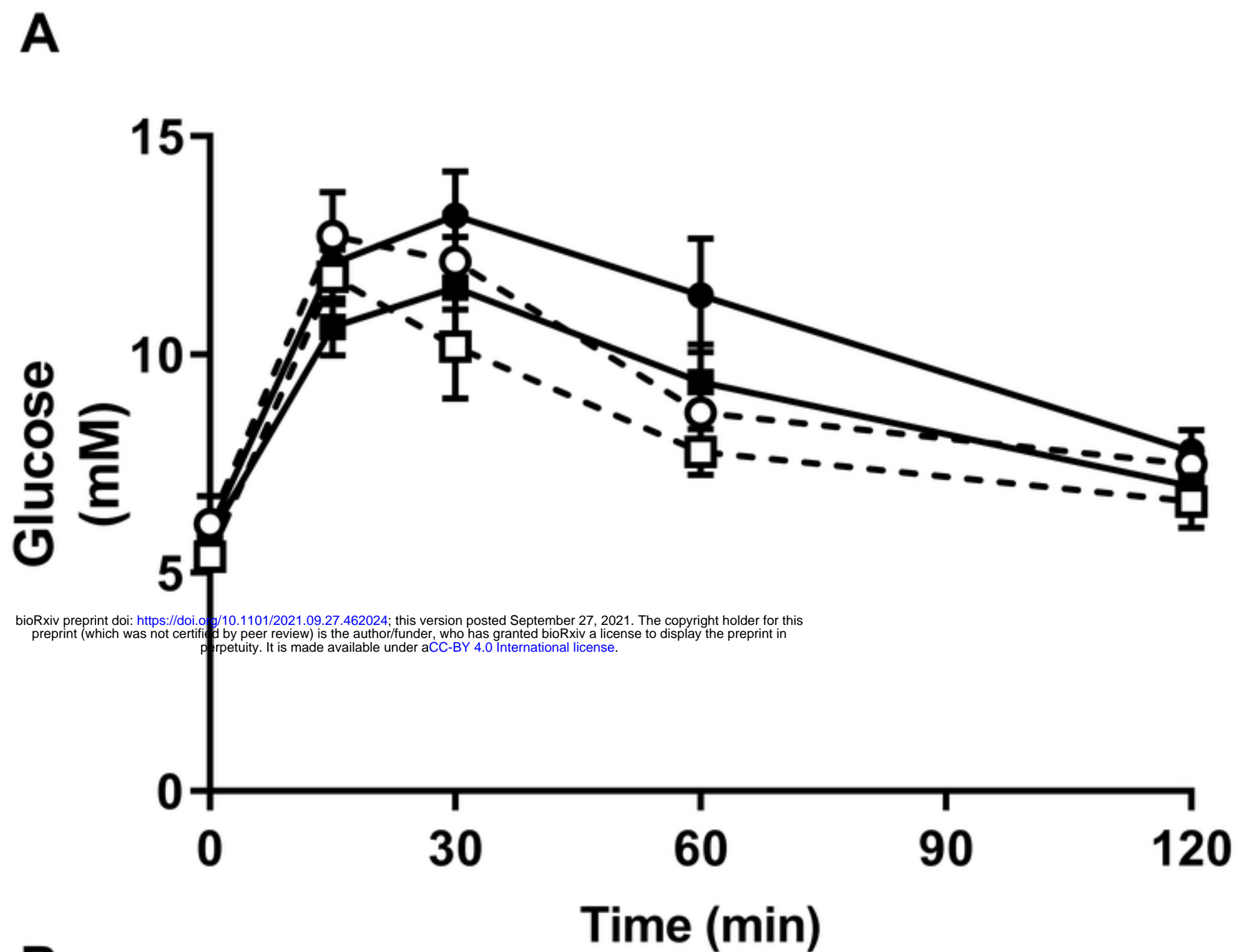


Fig 2

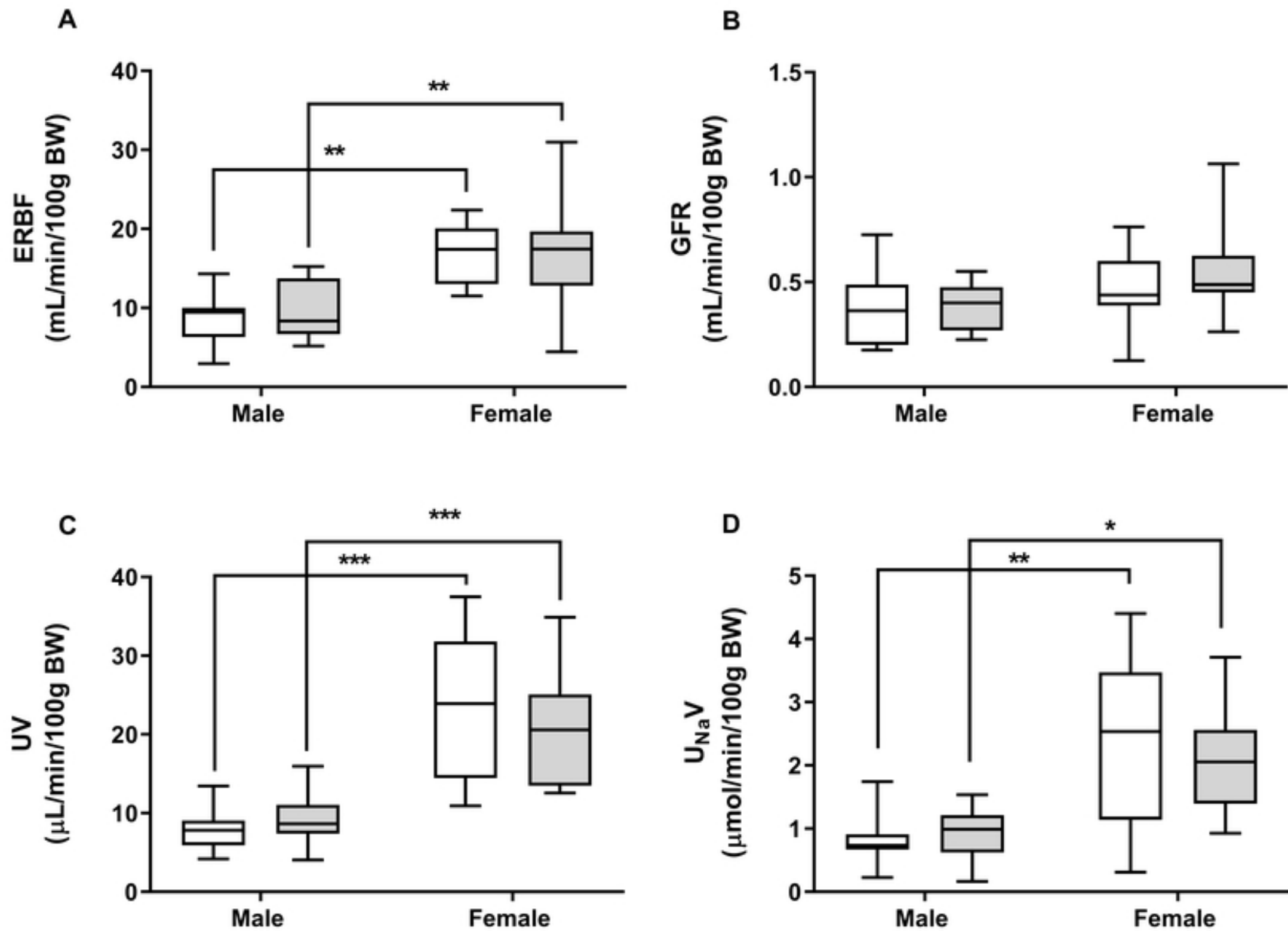


Fig 3

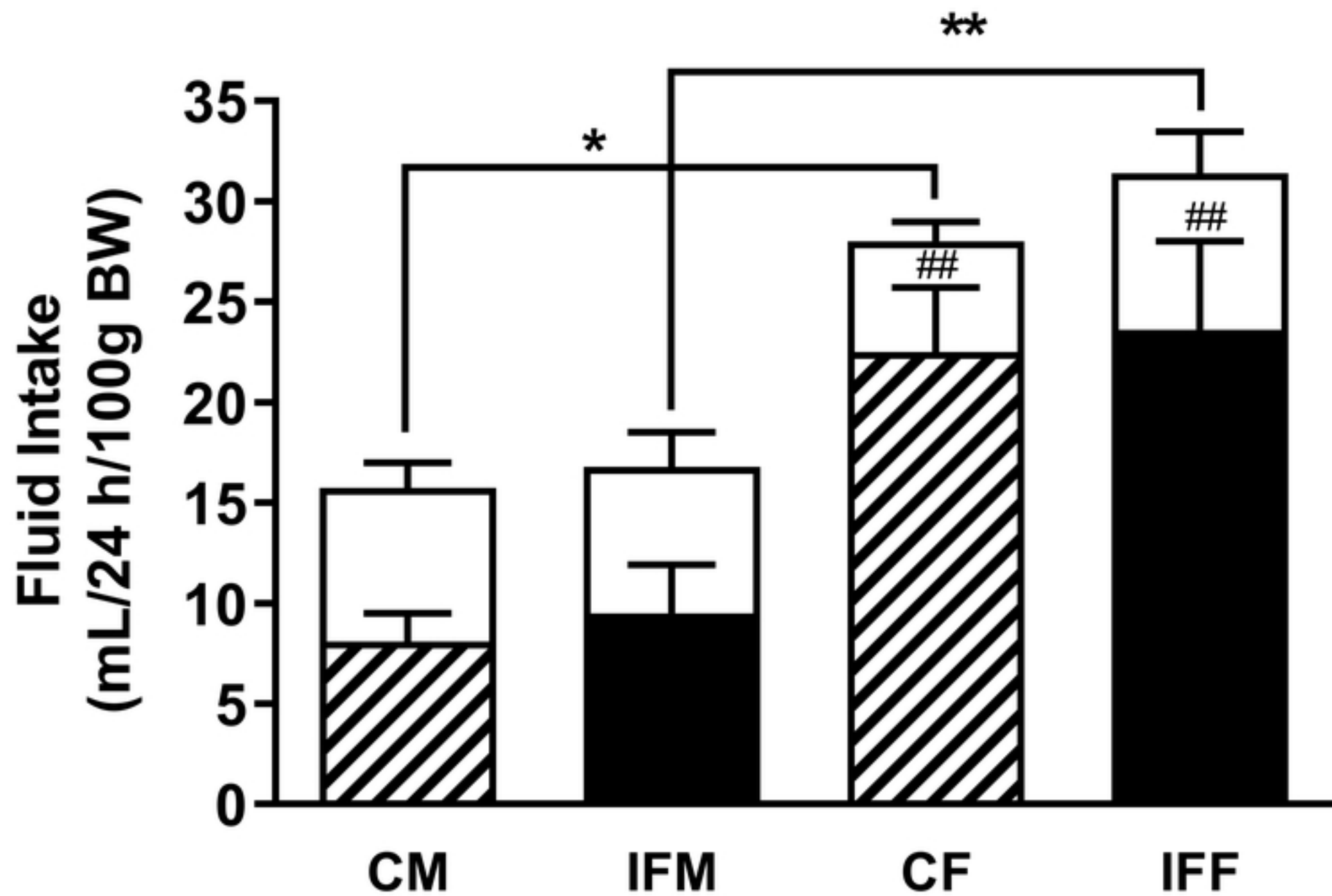


Fig 4

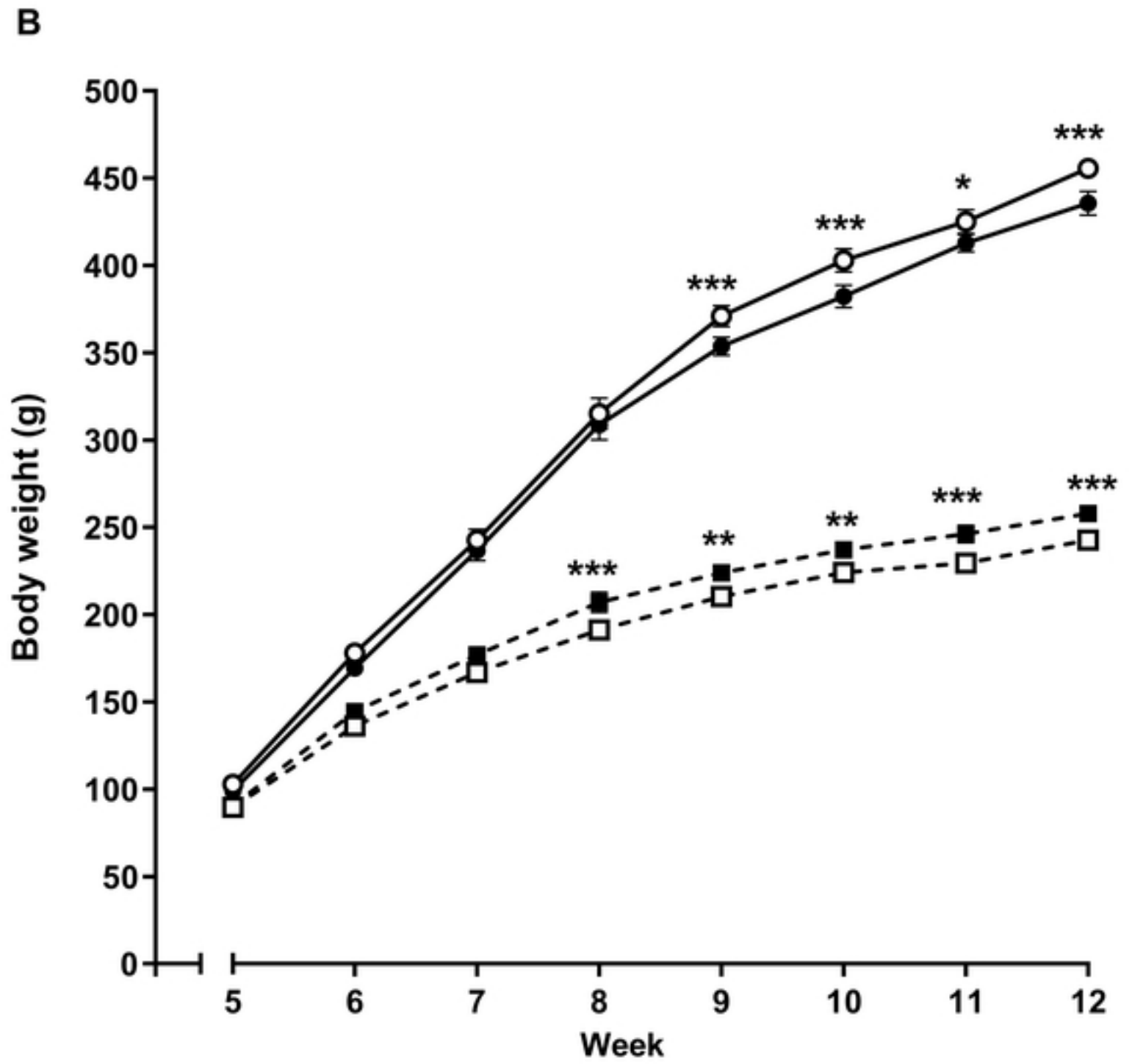
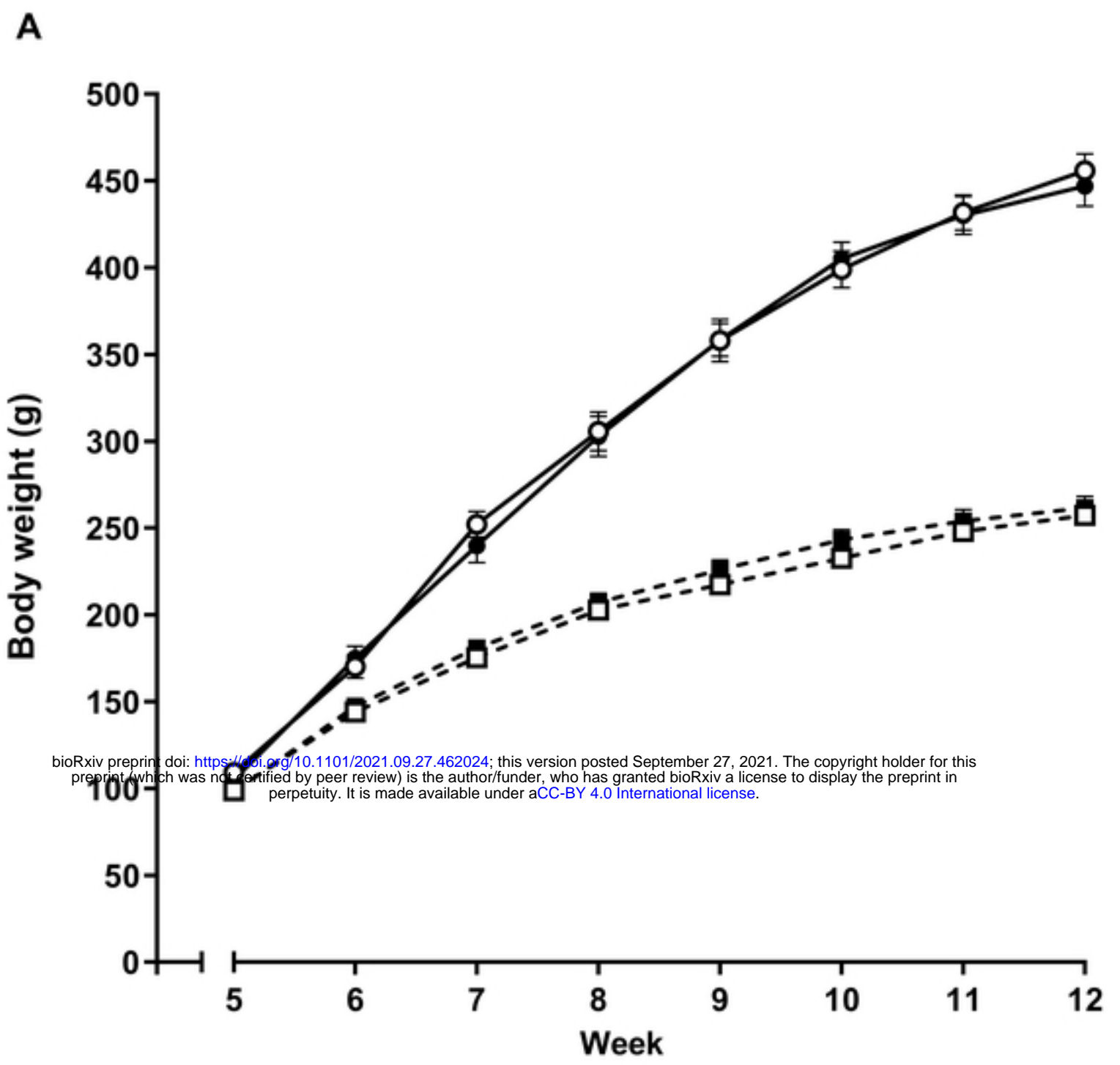


Fig 5

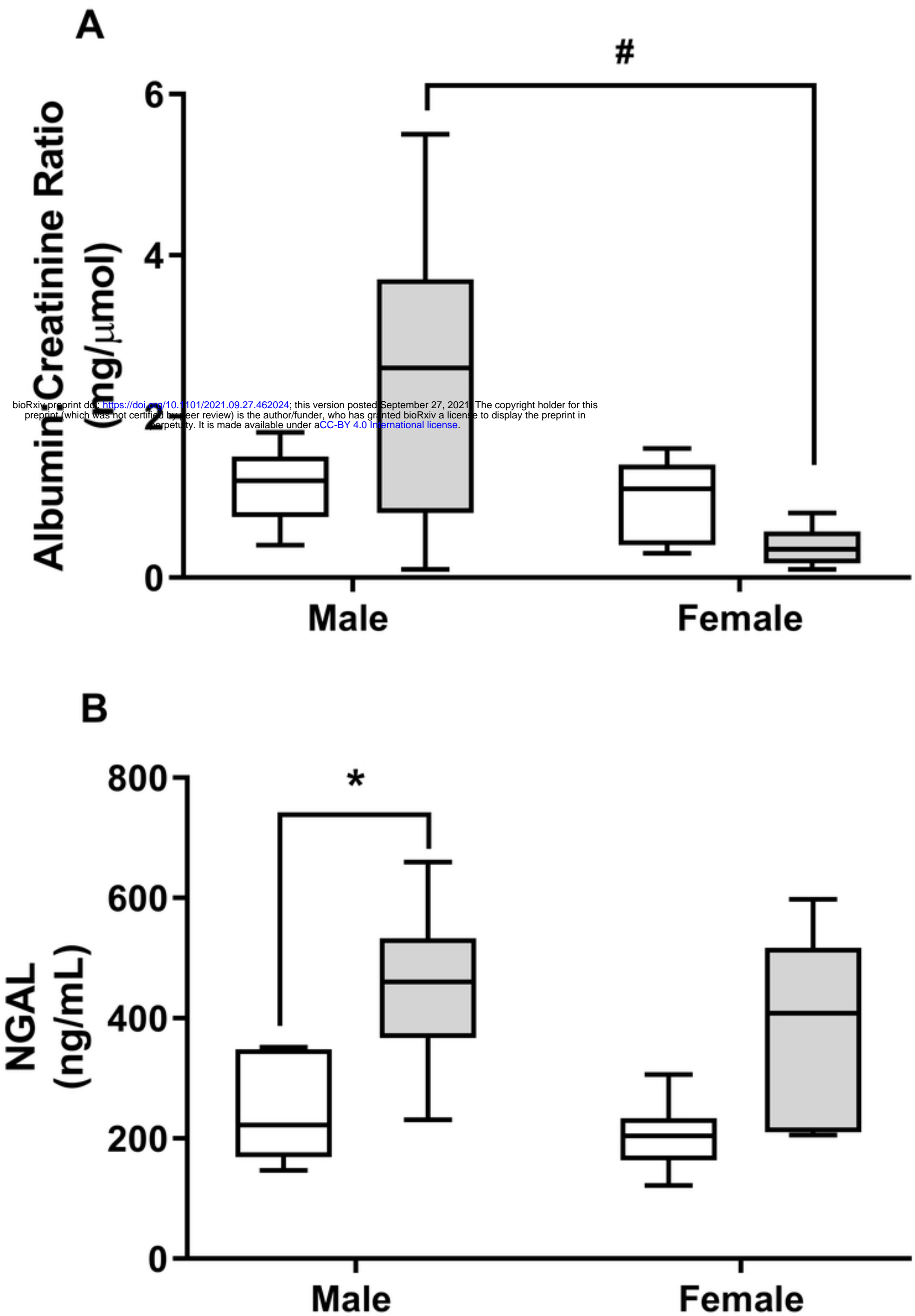


Fig 6

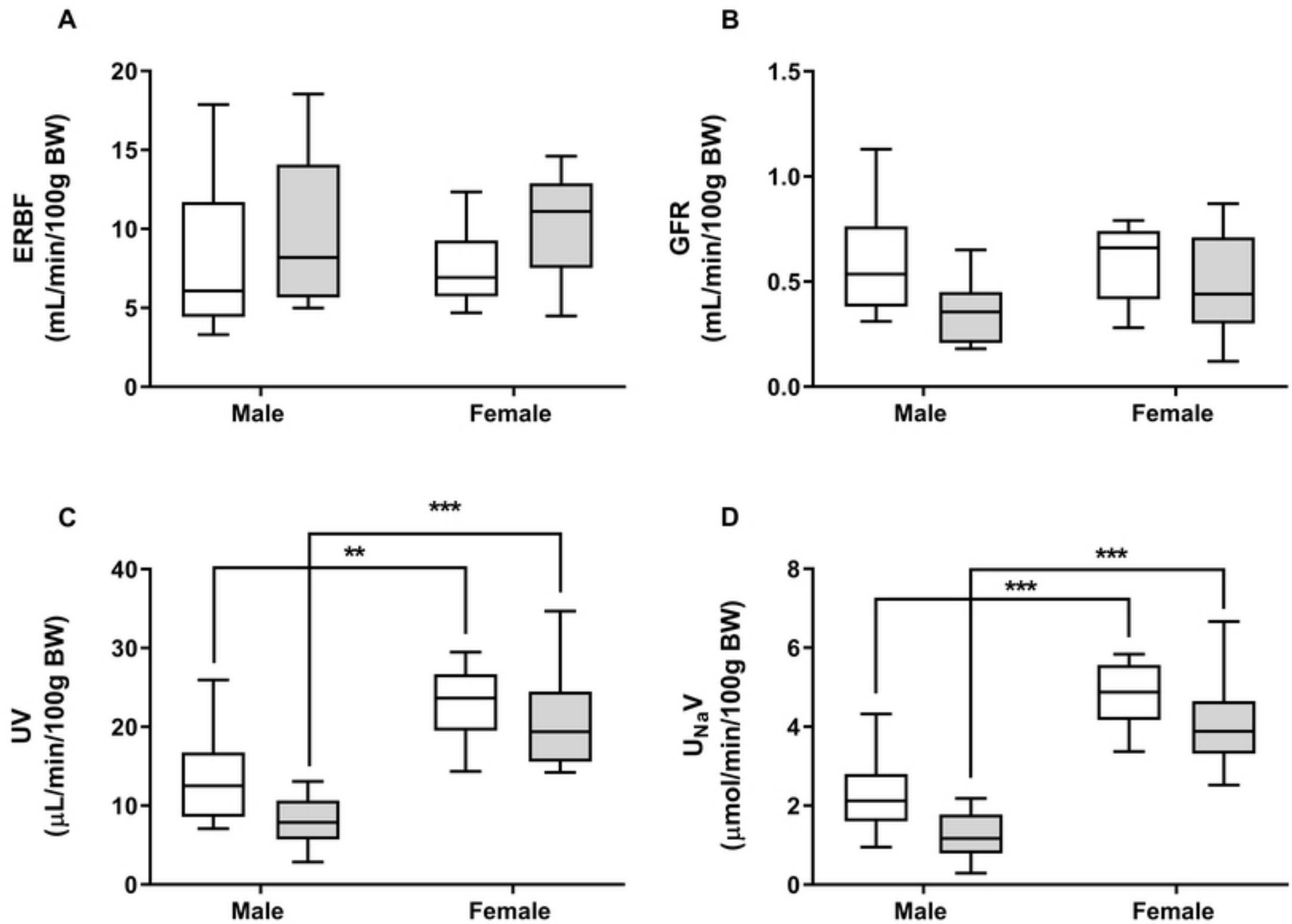


Fig 7