1	Vocal correlates of arousal in bottlenose dolphins (Tursiops spp.) in human care
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21 Abstract

Human-controlled regimes can entrain behavioural responses and may impact animal 22 23 welfare. Therefore, understanding the impact of schedules on animal behaviour can be a 24 valuable tool to improve welfare, however information on overnight behaviour and behaviour in the absence of husbandry staff remains rare. Bottlenose dolphins (Tursiops spp.) are highly 25 social marine mammals and the most common cetacean found in captivity. They 26 communicate using frequency modulated signature whistles which are individually 27 28 distinctive and used as a contact call. We investigated the vocal behaviour of ten dolphins housed in three social groups at uShaka Sea World dolphinarium to determine how acoustic 29 30 behavioural patterns link to dolphinarium routines. Investigation focused on overnight 31 behaviour, housing decisions, weekly patterns, and transitional periods between presence and absence of husbandry staff. Recordings were made from 17h00 – 07h00 over 24 nights, 32 spanning May to August 2018. Whistle production rate decreased soon after husbandry staff 33 left the facility, was low over night, and increased upon arrival. Results indicated elevate 34 arousal states associated with the morning arrival and presence of husbandry staff and 35 heightened excitement associated with feeding. Housing in pool configurations which limited 36 visual contact between certain groups were characterised by lower vocal production rates. 37 Production of signature whistles was greater over the weekends compared to weekdays 38 however total whistle production did not differ between weekends and weekdays. Heightened 39 arousal associated with staff arrival was reflected in the structural characteristics of signature 40 whistles, particularly maximum frequency, frequency range and number of whistle loops. 41 42 Overall, these results revealed a link between scheduled activity and associated behavioural responses, which can be used as a baseline for future welfare monitoring where changes in 43 normal behaviour may reflect shifts in welfare state. 44

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46 Introduction

Understanding and monitoring behaviour is a useful tool for welfare assessment [1] as 47 48 abnormal behaviour may be indicative of poorer welfare [2]. However, the behavioural cues 49 the animal gives must be correctly recognised and interpreted by the observer. Animals housed in captive facilities tend to have structured daily schedules of events such as food provision or 50 routine cleaning that are highly predictable through human-driven cues [3]. Patterns of animal 51 behaviour such as increased alertness [4] and increased vocalisation rates [3] may precede 52 regular events [5] Conversely, reduced activity of the animals has also been documented 53 preceding regular activities such as feeding [6] and public display [4]. 54

55 Arousal can be classified according to its level (high or low) and its valence (positive or negative) [7,8]. Vocal expression of arousal is a common feature of communication in both 56 57 humans and non-human animals [9-12] and can reflect responses to immediate experiences [13]. Understanding the vocal cues which act as markers of emotion in mammals [13-15] 58 may be used as a non-invasive tool to monitor welfare for animals held in human care [16-59 18]. In captive animals, both positive and negative associations with human care givers are 60 observed [19,20] as animals learn to associate humans with rewards and fear through both 61 62 classical and operant conditioning [21]. Human presence and handling can improve welfare of captive animals, for example in weanling pigs [22] and beef calves [19]. However, in some 63 livestock and poultry, human presence can induce fear and have a negative impact such as 64 decreased growth rates [20,23]. Responses to human presence associated with feeding are 65 often positive [13,24], and the combination of feeding and handling has been shown to play 66 an important role in the development of positive human-animal interactions [25]. Feeding 67

therefore seems to be a suitable candidate to study the behavioural response of a species to apositive interaction.

Animals in captivity are often cared for according to fixed daily schedules where these, as 70 well as other activities with positive outcomes, can elicit behavioural responses for example 71 in the form of anticipatory behaviours [26]. Anticipatory behaviour is a response evoked by 72 reward and is typically linked to elevated arousal states [24], however behaviour has been 73 74 characterised as either an increase [e.g., 27,28] or decrease [e.g., 29,30] in activity prior to the reward and has been best documented in domesticated mammals [28, 31, 32]. Responses such 75 as anticipatory behaviour can be used as an indicator of animal welfare through measuring 76 77 the frequency of certain behaviours [33].

Vocalisations often carry prosodic cues about the arousal state of the sender [10] which 78 79 makes the analysis of vocalisations a suitable tool to assess arousal states. Dolphins live in groups with many social interactions, with vocalisations being their primary method of 80 81 communication with and between groups. The vocal repertoire of bottlenose dolphins 82 consists of a range of pulsed and tonal sounds [34]. Tonal sounds include narrow-band, frequency modulated whistles to communicate during social interactions [35, 36]. Bottlenose 83 dolphins use 'signature whistles' to remain in contact [37-39] and address one another [40]. 84 Signature whistles are individually distinctive whistle types that encode identity information 85 within their frequency modulation pattern and are the most frequently emitted stereotyped 86 87 whistle type produced by an individual [41]. Although the whistle contour remains stable over time, shifts in whistle production rate, as well as frequency and duration characteristics, 88 89 may reflect underlying arousal of individual dolphins [38, 42, 43]. As might be expected for a cohesion call, individual production rates of signature whistles increase when motivation to 90 maintain contact or return to the group is strong, i.e., during separation [39] and isolation 91 92 events [42].

During the day, bottlenose dolphins housed in human care undergo multiple predictable 93 human-controlled events such as feeding, public presentations, training sessions and medical 94 examinations, making them good models for studying patterns in behaviour [44]. Predictable 95 events in captive facilities can evoke behavioural patterns in bottlenose dolphins [4] such as 96 an increase in spy-hopping frequency and surface-looking before any human-animal 97 interaction, including receiving of toys [45]. Vocal behaviour has been reported to be highest 98 when husbandry staff are present and during feeding/training sessions, and lowest at night 99 when numbers of caretakers are reduced [46] with peaks in activity overnight likely 100 101 associated with bouts of social activity [47]. The morning arrival of caretakers has also been associated with increased vocalisation rate in captive killer whales [48]. 102 As vocal behaviour indicates underlying emotional states, the link between behavioural 103 responses and daily events at a dolphinarium could provide insight into the animals' life in 104 human care. Additionally, this will provide insight into emotional responses and acoustic 105 106 cues of emotion in non-human animals. Therefore, the aim of this study was to investigate the overnight behaviour of bottlenose dolphins with a focus on patterns in detections and rates of 107 whistling in response to early morning regimes including husbandry staff presence and 108 109 feeding. We monitored the vocal response to scheduled events of the population as a whole, as well as at an individual level as animal personalities may vary. 110

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112 Methods

This study took place at uShaka Sea World which was established in 2004 and is located in
Durban, South Africa. The dolphinarium consists of a covered and open-air pool network
with seven interconnected pools of varying size with a combined volume of 11 000 m³. At

the time of data collection in 2018, the dolphinarium housed ten bottlenose dolphins: three 116 (two male and one female) common bottlenose dolphins (T. truncatus), one female Indo-117 Pacific bottlenose dolphin (T. aduncus) and six (four female and two male) hybrids of the two 118 species [49]. The individuals were held in three social groups (see Table 1 for more 119 information on the individuals and social groups) separated by gates which allowed partial 120 visual and full acoustic contact, but not free movement between social groups. All seven 121 122 pools were utilised by the dolphins during the course of the study, with the number and configuration of pools within which social groups were housed varying throughout the days 123 124 and overnight. We included pool configuration as a potential predictor variable since visual contact between the different social groups may affect arousal states. Each pool configuration 125 was defined based on which social group was housed in the outdoor presentation pool; 126 127 configuration 1 -all groups housed in the inside pools with one group having access to the outer interaction pool, and none having access to the outside presentation pool; configuration 128 2 - the female group housed in outside presentation pool while the mixed and male groups 129 housed in the inside pools with one group having access to the outer interaction pool; 130 configuration 3 – the male group housed in outside presentation pool while the mixed and 131 female groups housed in the inside pools with one group having access to the outer 132 interaction pool; and configuration 4 – the mixed group housed in outside presentation pool 133 while the male and female groups housed in the inside pools with one group having access to 134 135 the outer interaction pool. The outer interaction pool is directly linked to the outside presentation pool, allowing the dolphins in each of these pools to be in visual contact through 136 the gate. As potential arousal-eliciting activities, we noted morning activities at the 137 dolphinarium which include arrival of husbandry staff (hereafter referred to as 'staff') and 138 food preparation at 05h00 as well as feeding and vitamin administration at 06h00. The last 139

- public presentation occurs from 15h00 to 15h30 and the last trainer leaves between 17h00
- and 18h00, between which times all enrichment devices are removed from the pools.

Signature

whistle ID

F1

F2

F3

143 Table 1. Genetic, social grouping and individual data for the dolphins at uShaka Sea

Social Capture Name Species Sex **Parents** group date Female Affrika Τt F Captive born P1 + Ttgroup Zulu Ta-Tt Hybrid F Captive born P1 + P2F Khanya Ta-Tt F_2 Captive born M3 + hybrid

144 World in 2018 (adapted from Gridley et al., 2018)

	Tombi	<i>Ta-Tt</i> Hybrid	F	Captive born	P1 + P2	F4
	Khethiwe	Ta-Tt Hybrid	F	Captive born	P1 + P2	F5
Male	Ingelosi	<i>Ta-Tt</i> Hybrid	М	Captive born	P1 + P2	M1/M2
group	Khwezi	<i>Ta-Tt</i> Hybrid	М	Captive born	P1 + P2	M1/M2
	Kelpie	Tt	М	Captive born	Unknown <i>Tt</i> x <i>Tt</i>	M3
Mixed	Gambit	Tt	М	08/12/1976	Unknown <i>Tt</i> x <i>Tt</i>	P1
group	Frodo	Та	F	26/06/1979	Unknown <i>Ta</i> x <i>Ta</i>	P2

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146 Generating a signature whistle catalogue

The vocal behaviour of individuals was assessed through analysis of signature whistles. To determine the signature whistle for each dolphin, a catalogue was compiled using data collected during temporary isolation sessions in November 2016. Whistle contours are characterised by their time-frequency modulation patterns and in bottlenose dolphins, signature whistles can consist of a single contour or repeated contour (loop). A repeated contour, or multiloop whistle, is either connected where there are no breaks in the entire whistle contour or disconnected with a maximum inter-loop-interval of 0.25 seconds [43].

154	Acoustic data were collected using one to three High-Tec HTI-96 MIN (flat frequency
155	response of 2 Hz – 30 kHz \pm 1 dB) dipping hydrophone(s) attached to a Tascam digital
156	recorder (model DR-680) which sampled the data at 96 000 Hz. Simultaneous vocal notes of
157	observed behavioural data were recorded through a separate headset microphone. Time-
158	frequency spectrograms of acoustic recordings were analysed in Adobe Audition CC v 6.0
159	(FFT = 1024, frequency range = $0 - 60$ kHz, time series window = 10 seconds, Hann window,
160	50% overlap). To identify the signature whistle of each dolphin, they were temporarily
161	separated with one individual being kept in solitary in one pool between 10 and 20 minutes.
162	The signature whistle was defined as the most common whistle recorded during the
163	temporary isolations and matched to that individual by comparing the relative amplitude of
164	signals on the three hydrophones at different sites. Signature whistles were confirmed using
165	the SIGID bout analysis method [41] where stereotyped calls (at least 3 out of 4) occur in a
166	bout separated by $1 - 10$ seconds are considered highly likely to be signature whistles of an
167	individual.

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169

170 Investigating patterns of whistling behaviour

Acoustic data were collected over four periods in May, July and August 2018 using a single Sound Trap 300 HF hydrophone (Ocean Instruments, New Zealand, frequency response: 20 Hz – 150 kHz \pm 3 dB, sensitivity: 121 dB re. 1 µPa) sampling the data at 576 000 Hz. The hydrophone was placed in the 'link channel', an area central to the pool network to which the dolphins did not have access to but was within acoustic range of dolphins held in all pools. The hydrophone was attached to a 1 kg dive weight and suspended from a rope mid water (at 1.5 m depth, channel depth 2.5 m) held taut by attaching the rope to the roof with a carabiner clip. Ropes tied to the pool sides and roof were used to prevent movement which could
produce unnecessary noise on the hydrophone. Acoustic recording commenced in the late
afternoon after the final public presentation (between 15h30 and 17h00) and continued until
07h00 the following day. Recording was continuous, but files were restricted to standard 15minute durations which constrained file sizes.

The first 30 minutes of each overnight deployment was discarded to obtain data unbiased by 183 184 potential novelty effect as the dolphins may respond to the recorder's presence in the water. Thereafter, one 15-minute file was selected to represent each hour from 17h00 to 07h00, 185 using the recording which spanned the start of the hour. Each of the selected files were 186 187 analysed in Adobe Audition CC v 6.0 by visually locating whistle contours in the spectrogram display (FFT = 1024, frequency range = 0 - 60 kHz, time series window = 10 188 seconds, Hann window, 50% overlap). Each whistle was documented in a database and the 189 190 signal to noise ratio (SNR) was visually assessed using the following criteria: SNR - 1whistle is faint and barely visible, SNR 2 – whistle is clear and unambiguous, and SNR 3 – 191 192 whistle is prominent [50]. Whistles of good quality (those graded as SNR 2 or 3) were either matched to the established signature whistle catalogue and categorised as a signature whistle 193 from a specific individual or categorised as 'variable whistles' containing all non-signature 194 195 whistles from various individuals. The category 'unclassified whistles' was used for poor quality whistles (those graded as SNR 1) which did not match to the signature whistle 196 catalogue (Table 2). 197

198

200 Table 2. Whistle categories included in the analyses and recording quality rating based

201 on signal to noise assessment.

Whistle type	Description	SNR
Signature whistle	A whistle type unique to an	2 & 3, but can include
	individual that matches the	whistles with SNR 1 if we
	signature whistle catalogue	were confident that they are
		signatures
Signature whistle matching	Matching of an individual's	2 & 3
	signature whistle by another	
	individual identified through	
	overlapping of whistle	
	contours	
Variable whistle	Loud, unmasked whistles	2 & 3
	which do not match the	
	signature whistle catalogue	
Unclassified whistle	Masked or partial whistles	1
	that were unidentifiable and	
	faint	

202

In both the wild and captive facilities, bottlenose dolphins can copy the whistles of others. Such copies can be used in addressing or matching interactions, drawing attention from, or directing information to a particular individual [40, 51,52]. In some cases, the whistle match might not be an exact replication, but integrate features of the owner's whistle type or voice [52], however in other cases matches may be indistinguishable from the original. Whistle matching is difficult to identify unless they are overlapping in time. In general, where two 209 whistles with the same contour overlapped in time, we followed [53] in assigning the second whistle as the match and removed it from the signature whistle analyses. However, through 210 the analysis process we identified stereotyped matching behaviour distinct from all other 211 acoustic behaviour observed within the recordings whereby which led to the exclusion of 212 various whistles from the analysis. We noted that the contour of one individual (P2) was 213 emitted at various frequencies in prolonged series of matching interactions. For these 214 215 stereotyped acoustic interactions, we could not confidently assign whistle production to P2, therefore we termed these whistle sequences 'square copies'. These square copies, as well as 216 217 all whistles 30 seconds before and after each event, were removed from the statistical analyses to prevent over-representation of individual P2. All other whistle matching remained 218 in the dataset for the analyses including all whistle types. 219

220 To assess the production rate of whistles as a proxy of arousal, we counted the number of whistles in each category (Table 2) per 15-minute recording. We investigated how the 221 production rate of whistles (meaning all whistle types in Table 2 or only signature whistles) 222 was affected by several covariates using a generalised additive mixed model (GAMM) 223 approach using the package 'gamm4' version 0.2-6 [54] in RStudio version 4.0.3 [55]. This 224 225 approach allows fitting of non-linear relationships between variables as well as the inclusion of both fixed and random effects. Whistle production was investigated in terms of both 226 227 presence/absence and production rate (whistles per minute). A total of four models were built (two for all whistle types in Table 2, and two for signature whistles only) and for each, a 228 variation of the following covariates were tested: 'hour' (hour of the day/night), 'pool 229 configuration', 'time of week' (week vs weekend) and 'presence of signature whistles'. 230 Random effects for all four models included 'sampling day' to account for the variability of 231 whistles production between sampling days. The selection of covariates was based on testing 232 the assumption that patterns of activities and housing configuration in the dolphinarium lead 233

different levels of excitement which would reflect in dolphin vocal behaviour. Of most 234 interest were the activities of the dolphinarium which occur at scheduled times therefore 235 'hour' was of particular interest. Time of the week was of interest due to the increase of 236 visitors to uShaka Sea World over the weekend. Codes were assigned to working days, 237 'week', (1), which consisted of Monday to Friday, and weekend days, 'weekend', (2), which 238 consisted of Saturday and Sunday. The covariate 'pool configuration' also contained coded 239 240 subcategories which were based on which social group was housed in the outdoor presentation pool overnight. All models were run using data between the hours spanning 241 242 17h00 to 07h00, with one extreme outlier removed from the total whistle and signature whistle production rate models. 243

We did not incorporate signature whistle ID as a covariate in the models of the full data series 244 because the data were zero-inflated data due to a lack of whistles emitted in most hours 245 246 overnight. These models therefore investigate group, but not individual, calling behaviour for whistles. However, individual differences in vocal behaviour are of interest and relevant for 247 animal welfare. Therefore, a separate analysis was conducted to investigate individual 248 differences between 04h00 and 06h00 (04h00 - one hour before staff arrival and food 249 preparation; 05h00 - staff arrival and food preparation; 06h00 - feeding time) when a 250 251 sufficient number of calls for individual dolphins were available. Data were not normally distributed and had unequal variances (Shapiro-Wilks test and Levene's test, respectively) 252 even after transformations. Thus, Kruskal-Wallis tests were used to determine if certain 253 254 individuals significantly increased their signature whistle production rate in response to staff arrival and feeding time. Multiple Dunn *post hoc* tests followed a significant Kruskal-Wallis 255 result. We adjusted the alpha value using the Benjamini-Hochberg method to limit an 256 increase in type I error rate, which is caused by alpha-inflation due to multiple pairwise 257 comparisons. 258

In addition to changes in whistle production rate, the underlying arousal state may be 259 reflected in the structural parameters of whistles such as duration and frequency 260 characteristics [11, 42]. We investigated this for each individual by selecting five standard 261 whistle parameters from signature whistles namely minimum and maximum frequency. 262 frequency range, duration and number of loops, as well as production rate for each signature 263 whistle. Measurements were taken from time-frequency spectrograms (FFT = 1024, 264 265 frequency range = 0 - 40 kHz, time series window = 5 seconds, Hann window) of the fundamental frequency of 33 – 392 signature whistles from each animal in Raven Pro v1.6.1 266 267 [56]. For each signature whistle, production rate data were pooled into time periods representing staff absent (20h00 - 04h00) and staff present (05h00 - 07h00). Thereafter, 268 production rates per hour were recalculated for each time period. Times 17h00 – 19h00 were 269 270 omitted from this analysis due to the uncertainty of daily staff presence that occurred at these times. Data were not normally distributed and had unequal variances (Shapiro-Wilks test and 271 Levene's test, respectively) therefore paired-sample Wilcoxon tests were used to investigate 272 change in production rate between the pooled time. Additionally, all five signature whistle 273 parameters were individually compared between the two pooled time periods for six of the 274 ten individuals with sufficient data (at least five signature whistles for each individual for 275 each pooled time period). Again, data were not normally distributed and had unequal 276 277 variances (Shapiro-Wilks test and Levene's test, respectively), therefore Wilcoxon tests were 278 used to investigate change in the acoustic parameters of signature whistles between times of 'staff absent' and 'staff present'. 279

280

281 **Results**

A total of ten signature whistles were documented for the group of ten dolphins, of which eight were assigned to individuals (see Table 1, Fig 1A). The two remaining signature whistles could not be confidently differentiated between individuals M1 and M2, which are hybrid siblings which engage in a significant amount of whistly matching. All square whistle copying sequences were removed from the analyses (Fig 1B).

Fig 1. Whistle catalogues (A) Signature whistle catalogue of all ten dolphins housed at
uShaka Sea World in 2018 (F1, M3, P1 and P2 are single loop whistles; F2, F3 and F5 are
connected multiloop whistles; F4, M1 and M2 are disconnected multiloop whistles). (B)
Examples of (1) low, (2) middle and (3) high frequency square whistle copying, with a
similar contour shape to dolphin P2.

292

293 Nocturnal whistling behaviour was investigated from acoustic recordings made over 24 sampling nights (17 on weekdays and 7 on weekend days) which were conducted in four 294 hydrophone deployment periods ranging from five to seven nights in duration. From this, 88 295 hours of acoustic data were analysed with a total of 2640 documented whistles, 1647 of 296 which were signature whistles (62.4%), 168 variable whistles (6.3%), 569 unclassified 297 298 whistles (21.6%), 3 signature whistle matching (0.1%), 98 square whistles (3.7%) and 155 square whistle copying (5.9%) (Table 2). Square whistle copying behaviour was uncommon 299 and occurred sporadically with no particular temporal trend (S1 Fig). Furthermore, these 300 301 square copying events occurred predominantly during the night when overall signature 302 whistle production was at a minimum. In addition to P2 not confidently being assigned a whistle in these copying interactions, a closer inspection of these square whistle copying 303 304 events indicated that the vocal behaviour was more involved than a simple owner-copy interaction. In the 30 seconds before and after these whistle copying events that occurred 305

throughout the data set, whistles were produced most by three of the animals (S2 Fig). After
removing all whistles 30 seconds before and after each square whistle event, 2281 whistles,
1561 of which were signature whistles, were included in the analyses.

The GAMMs show a temporal trend in presence and production rate of whistles with two 309 clear peaks, one in the late afternoon and one in the morning (Fig 2; Table 3 – Models 1 to 4; 310 variable 'hour'; p < 0.001). These peaks correspond to staff presence, particularly to the time 311 312 of arrival of staff and preparation of food in the morning (05h00 - 06h00). Pool configuration also influenced overall whistle presence and production rate. Total whistle presence and 313 signature whistle presence significantly decreased when dolphins were housed in pool 314 315 configuration 2 (female group housed outside) compared to configuration 1 where all animals were housed in the indoor pools (Fig 3A; Table 3 – Model 1 and 3, variable 'pool 316 configuration'; p < 0.05). Total whistle production rate decreased when dolphins were housed 317 318 in pool configuration 4 (mixed group housed outside) and signature whistle production rate decreased in pool configuration 3 (male group housed outside) in comparison to 319 320 configuration 1 (Fig 3B; Table 3 – Model 2 and 4, variable 'pool configuration'; p < 0.05). The proportion of whistles produced during 'staff transition' (the afternoon period where staff 321 were leaving and/or had just left), 'staff absent' and 'staff present' indicates that this trend in 322 whistle production driven by pool configuration is prevalent during the overnight period 323 where staff were absent (Fig 3A, B). The likelihood of whistle or signature whistle 324 occurrence per hourly bin did not differ between weekdays and the weekend, however total 325 signature whistle production rate was greater over the weekend (Fig 4; Table 3; Model 4, 326 variable 'Time of Week'). Although not significant, the inclusion of this variable increased 327 328 model performance (R-squared from 0.178 to 0.233) which indicates that the time of the week plays a role in the production of signature whistles at the facility. Total whistle presence 329 and production are highly dependent on the presence of signature whistles (Table 3; Model 1 330

331	and 2, variable 'Signature whistle presence'). Although not significant in Model 1, this
332	variable significantly increased overall model performance (R-squared from 0.302 to 0.649).
333	The low-average adjusted R-squared values in all four models (Table 3; adjusted R-squared
334	range = $0.233 - 0.649$) indicate that there are other factors influencing whistle and signature
335	whistle production rates that cannot be explained by the covariates in these models.
336	Fig 2. GAMM summary of smoothing term 'hour' for all four models.
336 337	Fig 2. GAMM summary of smoothing term 'hour' for all four models. Fig 3. Total whistle and signature whistle production rate compared between pool
337	Fig 3. Total whistle and signature whistle production rate compared between pool

Fig 4. Signature whistle production rate compared between weekdays and weekend 340

341 days.

Model 1:		Hour	Pool configuration	Time of week		
Model 1:				Time of week	Signature whistle presence	Adjusted R-
Model 1:						squared
	347	< 0.001	0.042 (Intercept)	NA	-	0.649
Whistle presence			0.020 (config 2)			
Model 2:	346	< 0.001	0.044 (Intercept)	NA	< 0.001	0.521
Whistle count			0.025 (config 4)			
Model 3:	347	< 0.001	0.034 (Intercept)	NA	NA	0.279
Signature whistle			0.017 (config 2)			
presence						
Model 4:	346	< 0.001	0.001 (Intercept)	-	NA	0.233
Signature whistle			0.008 (config 3)			
count						

343 Table 3. GAMM results for all presence/absence and production rate models (only *p*-values of significant results presented)

There was a large difference in signature whistle production between individuals. Of the 345 1561 signature whistles identified $\sim 60\%$ were produced by three individuals (two female and 346 one male, Fig 5). The three least vocal animals contributing $\sim 7\%$ to the total signature 347 whistle production were all males (Fig 5). A comparison of individual signature whistle 348 production rate was investigated in the morning when total whistle production was at its 349 highest and there was sufficient data (04h00 to 06h00) to determine individual differences in 350 351 response to staff arrival (05h00) and feeding time (06h00). Kruskal-Wallis tests indicated that the signature whistle production rate of five of the dolphins increased significantly only in 352 353 response to feeding time (Fig 6A; no significant difference between 04h00 and 05h00, p < p0.05, n = 866). The remaining five individuals did not exhibit a significant increase and P1 354 produced no whistles during feeding time. Additionally, some of the animals were completely 355 silent overnight when no staff were present. Investigating individual differences in signature 356 whistle production between periods with staff absent (20h00 - 04h00) and with staff present 357 (05h00 – 07h00), F2 was highly vocal during both of these time periods (Fig 6B) and 358 signature whistle production from all dolphins increased with staff present (Fig 6B, p < 0.05359 for five of them, n = 1413). In the absence of staff, bouts of whistling occurred sporadically 360 between sample days, driven mostly by F2 as well as four other individuals. Although 361 signature whistle identification could not be included in the above-mentioned models as a 362 covariate, the results strongly suggest that individual differences in signature whistle 363 production are important. 364

Fig 5. Total count of signature whistles produced throughout the study period.

Fig 6. Individual vocal responses to feeding and staff presence. (A) Signature whistle production rate of each individual compared across three hours (04h00 – one hour before staff arrive, 05h00 – staff arrival and food preparation, 06h00 – feeding). Significance indicated by asterisks is consistent among individuals, with no significant shift between

370	04h00 and 05h00. (B) Whistle production rate per hour of all individuals when staff were
371	absent and when staff were present. Significance between time periods indicated by asterisks.

372

373	Matched pairs Wilcoxon tests showed significant differences in the acoustic parameters of
374	their signature whistles between periods of staff absence and presence (Fig 7). The three
375	animals which exhibited the most changes in whistle characteristics all showed a significant
376	decrease in maximum frequency and frequency range when staff were present. The two other
377	individuals did not exhibit any significant shifts in whistle characteristics. Overall, most of
378	the individuals showed significant decreases in maximum frequency (average: from 12.8 to
379	11.4 kHz), half of them significant decreases in frequency range (average: from 10 to 7.9
380	kHz) and number of loops (average: from 4.6 to 2.5 loops per whistle), while two individuals
381	exhibited significant decreases in whistle duration (average: from 1.35 to 1.05 s) and only one
382	individual a significant increase in minimum frequency (from 5.5 to 6.6 kHz).

Fig 7. Shifts in signature whistle parameters in response to staff presence. Asterisks indicate significance.

385

386 **Discussion**

The overall night-time vocal behaviour between the afternoon and morning peak of whistle production was fairly quiet, with some bouts of whistling behaviour during the night occurring randomly between recording nights. This is in accordance with [57] who found that peaks in nocturnal vocal activity of captive bottlenose dolphins may occur. Sleeping/resting occurs predominantly during the night and accounts for up to 87% of nocturnal behaviour of captive bottlenose dolphins [58]. This explains the decrease in vocal activity during this

night-time period, however from this study there has been no indication what might be 393 driving the bouts of social behaviour. It has also been shown that captive bottlenose dolphins 394 increase vocal and social interactions before their rest periods, possibly to promote 395 synchronous swimming, a behaviour observed in wild and captive dolphins [58]. Similarly, in 396 a study of captive bottlenose dolphins (T. truncatus) at facilities in Japan, vocal production 397 rates increased during the day when their human caretakers were present [46, 58] and vocal 398 399 behaviour, including whistle production rate, decreased at night when caretakers were absent [46]. These studies indicate a positive relationship between caretakers and animals in 400 401 captivity may be beneficial to the welfare of the animals [59].

Daily arousal patterns which coincided with presence of staff and caretakers at the facility. 402 specifically at the end of the day (last daily presentation) and early in the morning (food 403 preparation and feeding), suggest excitement associated with these activities. Excitement is a 404 positive arousal state [8, 60], and is less commonly studied than negative arousal. Vocal 405 406 responses indicating positive and negative emotional states are hard to differentiate as they may share vocal features [61]. In the current study, signature whistle production rates 407 increased in the morning during staff arrival and food preparation, significantly intensified 408 409 during feeding and then decreased one hour after feeding and was driven by half of the individuals in the facility. At feeding time, five dolphins significantly increased signature 410 whistle production. In wild bottlenose dolphins an increase in vocal activity is associated with 411 foraging which may be to maintain contact or to recruit individuals [62]. Food-anticipatory 412 413 activity describes increased arousal before feeding events on strict daily schedules [63] and 414 has been observed in captive animals during scheduled feeding times [46, 64]. The emotional value of anticipatory behaviour is thought to reflect the balance of the reward system 415 experienced by animals before and during feeding [27]. Increased vocalisations around 416 feeding time have also been documented in captive false killer whales (Pseudorca 417

crassidens), common bottlenose dolphins (*T. truncatus*) and common dolphins (*Delphinus delphis*), where vocalisation rates increased upon the arrival of their caretakers and was
maintained or intensified throughout feeding and decreased immediately after [48, 64]. In
wild animals, detection of food may lead to an increased arousal state followed by temporary
"elation" after capture of prey [8]. Other studies however observed a decrease in activity and
arousal before human-animal interactions including feeding, for dolphins [4] and
chimpanzees [6].

Of the four males in this analysis, the three younger males exhibited significant changes in 425 most of the whistle parameters, all including a decrease in frequency range from a period 426 with staff absent to a period with staff present. One male decreased whistle production rate 427 when staff were present and two males exhibited significant decreases in whistle duration, 428 one of which also decreased number of loops. The oldest male did not significantly shift any 429 whistle parameters between these two time periods. Of the two females included in this 430 431 analysis, F2 did not significantly shift any whistle parameters between these two time periods while the oldest female in the facility decreased maximum frequency and number of loops. 432 Some dolphins may emit whistle loops faster than normal when they are excited [65] 433 434 therefore duration is a function of number of loops [42]. Whistle duration has been shown to increase during isolation [42], and much like number of loops, in the context of excitement 435 and stress, whistle duration seems to be the opposite. Whistle frequency parameters in 436 bottlenose dolphins such as production rate and number of whistle loops are closely related to 437 438 the level of arousal of an individual [42, 43]. The number of loops and whistle production 439 rates vary by context and an increase in these parameters may indicate stress [43] or excitement [66] in bottlenose dolphins. In the current study, the whistle production rate 440 significantly increased in six of the ten individuals, however a significant decrease in the 441 442 number of loops was only exhibited by three of the six dolphins evaluated. An increase in the

number of loops in the context of stress [43] and a decrease in the number of loops in the
context of excitement (as in the current study) suggest that loop number may be a useful tool
in monitoring positive and negative arousal states in bottlenose dolphins. Although certain
shifts in signature whistle parameters appear to be indicative of individual arousal,
differences are not consistent across individuals [42]. The same applies to the dolphins at this
facility where individuals had different combinations of whistle parameter shifts, therefore
the dolphins should be individually considered and monitored.

In this study the dolphins were less vocal when the females were housed in the outdoor 450 451 presentation pool and produced whistles less frequently when the mixed group was housed outside and produced signature whistles less frequently when the male group was housed 452 outside and were most vocal in pool configuration 1, when they were all housed in the indoor 453 pools (Fig 3A). This is contradictory to what you might expect from a cohesion call [39, 42] 454 and indicates an emotive or social reaction in response to these different types of group 455 456 separation. Although pool configuration affects overall whistle production rates, a vocal response to staff presence or absence is much less clear (Fig 3B) and somewhat contrary. 457 When housed inside, human activity is in view of all dolphins, including the arrival of staff, 458 459 therefore the strongest behavioural response would be expected here. Whistle production rate in response to staff presence is higher in configuration 2 for total whistles, and similar in 460 configurations 1 and 2 for signature whistles, which is not what was expected. The social 461 dynamics are quite different among the three groups housed at uShaka Sea World and 462 although they are housed in separate social groups, they are always in acoustic (and mostly 463 464 visual) contact with one another. This may allow inter-group social bonds to be formed, which has also been documented in wild meerkats [67] and wild bottlenose dolphins [68]. 465 Bottlenose dolphins live in complex societies and depend greatly on close social bonds [69]. 466 Because of this, they are more likely to suffer from social-related stress in both the wild and 467

468 captivity [70] however there is great opportunity for these animals in captivity to achieve469 positive welfare states due to strong social bonds within their social groups [71].

470 Whistle production rate was higher over the weekend compared to weekdays. While the number of visitors were larger on weekends than during the week, the daily schedule was the 471 472 same regardless of the day of the week. Visitors to captive facilities play a role in the 473 behavioural responses of mammals, however not all captive mammals are affected by visitor presence and noise [72]. For example, negative responses to visitors in captivity has been 474 documented in nonhuman primates [73-76], whereas captive meerkats are behaviourally 475 476 unresponsive under different intensities of visitor behaviour [77]. Bottlenose dolphins undergo activities in the form of public presentations which has been documented to elicit 477 positive arousal rather than stress or negative arousal [4]. However, the relationship between 478 the magnitude of arousal proceeding a presentation and audience size has not yet been 479 investigated. Increased whistle production rate over the weekend could not be directly linked 480 481 to an increase in visitors as data were collected once visitors had left, however residual excitement from large crowds could have possibly been carried over into the overnight 482 recording periods. 483

Regarding the overall signature whistle production, all square copies were removed from the 484 analysis and in doing this, there was a possibility that signature whistles of P2 were 485 486 underrepresented. Square whistle copying interactions are important in terms of social interactions and bonds between certain animals and understanding this will give insight into 487 their social vocal behaviour as a whole. Of the ten dolphins in the study, the most vocal 488 489 during both overnight and morning periods was a female, the third youngest dolphin. The least vocal was a male, the oldest dolphin at the facility. Age has been documented to affect 490 signature whistle production rate in bottlenose dolphins where higher signature whistle rates 491

are present in younger dolphins and decreases with age, more quickly in males [42, 43].
Differences in whistle production in bottlenose dolphins between sexes has previously been
documented due to differences in social histories of males and females [78]. Because the
whistles of hybrids M1 and M2 could not be differentiated, we could not investigate the role
that age and sex play together in vocal activity of these dolphins in more detail.

497 The most vocal individuals in this study were predominantly hybrids, all of which are offspring of P1 and P2. Differences in behaviour between purebred and hybrid cetaceans has 498 been reported [79]. Hybridisation of captive bottlenose dolphins and other dolphin species 499 500 has been widely documented [58, 80-82], however hybridisation between bottlenose dolphin species in captivity has seldom been seen [see 49, 83]. Although hybridisation is naturally 501 occurring in 10% of animal species [84], in the wild there is little overlap of home ranges of 502 *Tusriops* spp., limiting inter-species mating opportunities of bottlenose dolphins [49]. 503 Hybridisation has been documented to influence behaviour in animals; for example, neonatal 504 505 hiding behaviour [85] and vocalisations [86] of hybrid deer species are strongly influenced by genetic history and display intermediate characteristics of the parent species. Similarly, 506 intermediate behaviour of free-ranging porpoise hybrids has been documented [79]. Although 507 508 not included in the analyses due to small sample sizes, species may play a role in signature whistle production of bottlenose dolphins. 509

510 Welfare assessments in the past were heavily focussed on monitoring behaviour indicative of 511 poorer welfare as good welfare was thought to result from lack of suffering [87].

512 Vocalisations in farm animals have been used as a measure of welfare on farms [88]. Animals

that express more arousal and anticipation for events or rewards may be in poorer welfare

514 however some level of arousal before a positive event, such as feeding, reflects positive

515 emotions [89]. One such emotion is excitement [8], which is the emotional state that the

dolphins in this facility appear to be reflecting when they increase the production rate of 516 social sounds. Additionally, some signature whistle characteristics reflect this excitement 517 associated with the presence of staff at the facility. Undesirable behaviour may only be 518 present in a certain group or sex, which emphasises the limitations of a unified approach to 519 monitoring welfare and the importance to explore individual welfare states [90]. This all 520 provides a useful index; where production rate and whistle characteristics would usually shift 521 522 within this population, an abnormal change at either a group or individual level could be indicative of a shift in welfare state. 523

524

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790 Supporting information

S1 Fig. Total number of square whistle copying events during each time bin throughout
the study period.

S2 Fig. Total whistle counts, as well as P2 as the initiator of the copying events (Kriesell
et al., 2014).













