

Interacting Phenotypes and the Coevolutionary Process

Running title: Trait interactions and coevolution

Stephen P. De Lisle^{1,2}, Daniel I. Bolnick¹, Edmund D. Brodie III³, Allen J. Moore⁴, and Joel W. McGlothlin⁵

Corresponding author: Stephen P. De Lisle Email: stephen.de_lisle@biol.lu.se

1. Department of Ecology & Evolutionary Biology
University of Connecticut
75 N. Eagleville Road
Storrs, Connecticut, USA 06269
2. Present address:
Evolutionary Ecology Unit, Department of Biology
Lund University
Solvegatan 37, Lund, Sweden
3. Department of Biology and Mountain Lake Biological Station
University of Virginia
485 McCormick Road
Charlottesville, VA 22904 USA
4. Department of Entomology
University of Georgia
Athens, GA 30602 USA
5. Department of Biological Sciences
Virginia Tech
2125 Derring Hall, 926 West Campus Drive
Blacksburg, Virginia, USA 24060

Author Contributions All authors contributed to all aspects of the manuscript.

Acknowledgements Funding was provided by grants from the Royal Swedish Academy of Sciences and Swedish Research Council to S. De Lisle (VR registration number 2019-03706), the University of Connecticut and the NIAID (1R01AI123659-01A1) to D. Bolnick, and the National Science Foundation (DEB 1457463) to J. McGlothlin.

Data Accessibility No data to be archived

1 **Interacting Phenotypes and the Coevolutionary Process**

2

3 **Abstract** Coevolution occurs when species interact to influence one another's fitness,
4 resulting in reciprocal evolutionary change. In many coevolving lineages, trait expression in
5 one species is modified by the genotypes and phenotypes of the other, forming feedback
6 loops reminiscent of models of intraspecific social evolution. Here, we adapt the theory of
7 within-species social evolution, characterized by indirect genetic effects and social selection
8 imposed by interacting individuals, to the case of interspecific interactions. In a trait-based
9 model, we derive general expressions for multivariate evolutionary change in two species and
10 the expected between-species covariance in evolutionary change across a selection mosaic.
11 We show that reciprocal interspecific indirect genetic effects can dominate the
12 coevolutionary process and drive patterns of correlated evolution beyond what is expected
13 from direct selection alone. In extreme cases, interspecific indirect genetic effects can lead to
14 coevolution when selection does not covary between species or even when one species lacks
15 genetic variance. Moreover, our model indicates that interspecific indirect genetic effects
16 may interact in complex ways with cross-species selection to determine the course of
17 coevolution. Importantly, our model makes empirically testable predictions for how different
18 forms of reciprocal interactions contribute to the coevolutionary process and influence the
19 geographic mosaic of coevolution.

20

21 **Key words:** Coevolution, cross-species selection, interspecific indirect genetic effects,
22 geographic mosaic, quantitative genetics, species interactions

23 **Introduction**

24 Coevolution occurs when interacting lineages evolve reciprocally in response to one another
25 (Janzen 1980, Thompson 1982). Although the concept of coevolution may be applied to
26 lineages that share genes, such as males and females of the same species (Arnqvist and Rowe
27 2002), it was originally invoked to explain patterns of correlated evolution between
28 interacting species (Ehrlich and Raven 1964). In some cases, coevolution can result in tightly
29 integrated mutualisms or spectacular arms races that drive the evolution of exceptional
30 phenotypes (Brodie et al. 2002, Pellmyr 2003, Johnson and Anderson 2010). Yet even
31 beyond these striking cases, coevolution is likely important for a wide range of interacting
32 lineages, including consumers and their resources, hosts and their pathogens, competitors,
33 and mutualists (Thompson 1982, 1994). Although coevolution has clearly played a major
34 role in the origins of diversity, much is still unknown about when and how species
35 interactions generate reciprocal evolutionary change.

36 Theoretical models of coevolution typically focus on the fitness effects of trait
37 interactions between coevolving species and how selection imposed by one species manifests
38 evolutionary change in an interacting species (Nuismer 2017). The interaction between
39 species in coevolving lineages is often intimate, with one species spending a greater part of
40 its life cycle in close contact with the other. Thus, coevolution bears a striking resemblance
41 to intraspecific social evolution (Stearns 2012). Like social evolution, coevolution often
42 includes interacting or extended phenotypes, which arise when traits can only be understood
43 within the context of interactions with others (Dawkins 1982, Moore et al. 1997).

44 In quantitative genetic models of intraspecific social evolution, social interactants
45 influence one another via two pathways, each of which has a counterpart in coevolutionary
46 theory. First, the phenotype of one individual may cause fitness effects in a social partner,
47 leading to a form of selection known as social selection (West-Eberhard 1979, West-

48 Eberhard 1983, West-Eberhard 1984, Wolf et al. 1999). At the heart of all coevolutionary
49 models is a form of reciprocal fitness interaction that resembles social selection, where the
50 fitness of individuals in one species is influenced by traits in an interacting species (Brodie
51 and Ridenhour 2003, Ridenhour 2005, Nuismer 2017). Second, models of social evolution
52 may also include indirect genetic effects, which occur when the phenotype of one individual
53 depends on the genotype of an interacting partner (Moore et al. 1997, Wolf et al. 1998). A
54 classic example of indirect genetic effects is maternal effects, in which offspring phenotype is
55 a function of both their own genes (a direct genetic effect) and maternal phenotypes such as
56 litter size and provisioning (an indirect genetic effect) (Kirkpatrick and Lande 1989,
57 Mousseau and Fox 1998, McAdam et al. 2002). When indirect genetic effects are reciprocal,
58 feedback effects may inflate the genetic variance available for response to selection,
59 drastically accelerating the rate of evolution (Moore et al. 1997). Although most models of
60 indirect genetic effects do not extend beyond species boundaries, indirect genetic effects may
61 also be common in species interactions between mutualists, competitors, parasites and hosts,
62 and predators and prey. Phenotypic plasticity in response to an interacting species is
63 common (Agrawal 2001), and when these influences on trait expression have a genetic basis
64 they may represent interspecific indirect genetic effects (IIGEs; Shuster et al. 2006).
65 Although IIGEs have received some attention in the context of community genetics (Shuster
66 et al. 2006, Witham et al. 2020), their potential role in driving trait coevolution has been
67 mostly unexplored.

68 To date, most explorations of IIGEs have been studies providing empirical support for
69 their likely existence and their contribution to trait variation. Examples of interspecific
70 phenotypic manipulation are common in nature (Table 1), and many of these cases can be
71 argued to be putative cases of IIGEs. One possible example occurs in arbuscular
72 mycorrhizae, where the genotypes of fungal mutualists can alter root traits in the plants they

Table 1. Coevolutionary interactions where IIGEs may be prevalent.

Interaction	IIGE synonym	Diffuse/individual interaction	Example References
host-parasite	host manipulation	both	(Thomas et al. 2012)
predator-prey	trait-mediated indirect interaction	both	(Peacor and Werner 2001)
plant-animal	inducible direct defense host-plant manipulations	both	(Weis and Abrahamson 1986, Chen 2008)
plant-microbe	joint trait microbially mediated trait	diffuse	(Friesen et al. 2011, O'Brien et al. 2021)
host-microbiome	host control host-microbiome interaction	diffuse	(Stappenbeck and Virgin 2016, Foster et al. 2017)
consumer-resource	toxin sequestration	both	(Züst et al. 2018)

73 inhabit (Gianinazzi-Pearson et al. 2007). In host-parasite systems, parasite manipulation of
74 host traits (such as behavior) and reciprocal host manipulation of parasite traits (such as
75 growth rate, via immune response) are key features of species interactions (Thomas et al.
76 2012). Importantly, in many host-parasite systems, both the host and the parasite experience
77 sustained interactions with a small number of individuals of the other species, often over key
78 periods of the life history. For example, helminth parasites excrete a variety of
79 immunomodulatory products that suppress or misdirect the immune system of their
80 individual host (Damian 1997, Schmid-Hempel 2008, Oladiran and Belosevic 2012). Thus,
81 host immune response to infection is controlled by the genotype of both the host and parasite
82 (e.g., Barribeau 2014). As a specific example, some populations of threespine stickleback
83 (*Gasterosteus aculeatus*) initiate a strong immune response to infection by the cestode
84 *Schistocephalus solidus* (Fig. 1A) involving granulocyte proliferation and fibrosis, which

85 effectively suppress cestode growth and viability (Weber et al. 2017). Other stickleback
86 populations do not exhibit this response and allow rapid cestode growth, perhaps representing
87 a tolerance strategy. Cestode growth is thus an indirect genetic effect of its host's genotype.
88 Conversely, the cestode has been shown to secrete compounds that suppress this host
89 response (Scharsack et al. 2004, Scharsack et al. 2007, Scharsack et al. 2013) and down-
90 regulate sticklebacks' pro-fibrotic gene expression (Fuess et al. 2020), suggesting reciprocal
91 indirect genetic effects.

92 In goldenrod (*Solidago*; Fig. 1B), size of galls produced by the gall fly *Eurosta* are
93 determined by genotypes of both fly and plant, and evolution of gall size is influenced in part
94 by cross-species selection imposed on *Eurosta* larvae by species at other trophic levels (Weis
95 and Abrahamson 1986, Weis et al. 1992, Abrahamson and Weis 1997). This type of
96 interaction is common across gall-forming insects and their plant hosts; in *Hormaphis* aphids,
97 variation the *bicycle* gene has been linked to variation in gall size (Korgaonkar et al. 2021).
98 In many herbivore-plant interactions, physical damage to leaves induces upregulation of
99 defensive compounds to deter further herbivory, which can be countered by matching
100 physiological changes in the herbivore (Ohgushi 2005). For example, *Littorina* snail
101 herbivory changes foliar chemistry of the brown seaweed *Ascophyllum nodosa* (increased
102 phlorotannin concentrations), which in turn reduces snail movement and consumption rates
103 (Borell et al. 2004).

104 Host-parasite, host-parasitoid, and some plant-herbivore interactions can entail
105 intimate long-term associations between individuals. In contrast, predator-prey interactions
106 tend to be more diffuse (Brodie and Brodie 1999). Prey sense predation risk through
107 chemical, auditory, or visual cues and change their morphology, physiology, or behavior in

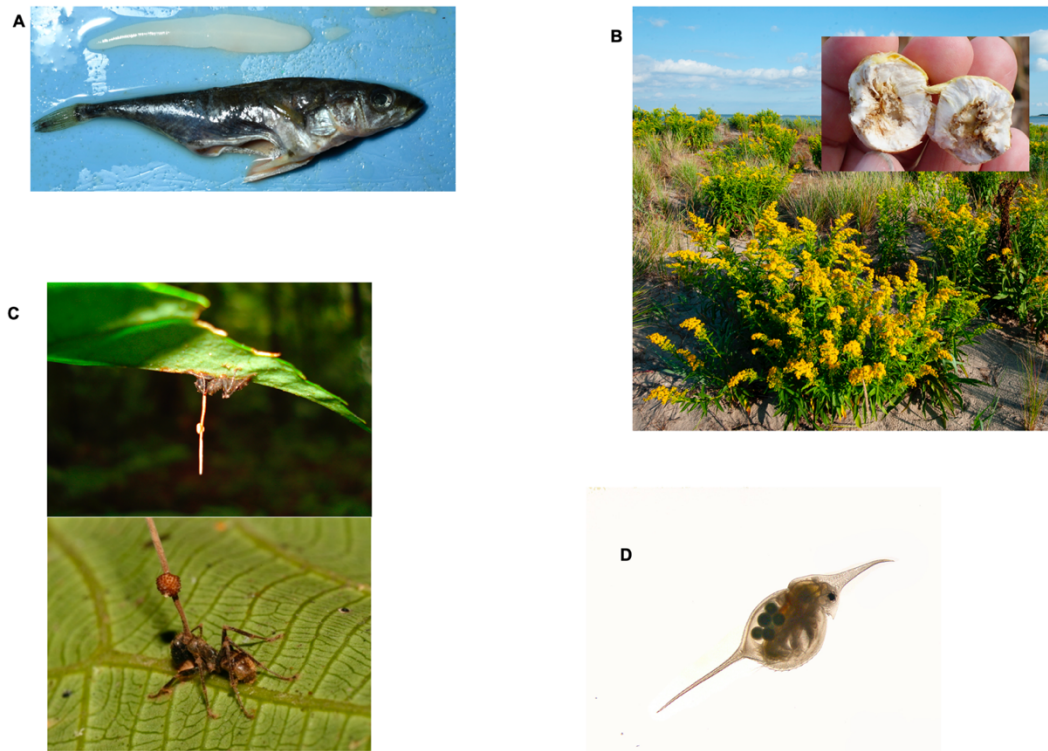


Figure 1. Examples of traits potentially mediated by IIGEs. IIGEs are a common feature of species interactions, particularly in host-parasite systems where prolonged contact between single individuals of both species determines fitness and trait expression for both. Here we highlight four examples where such IIGEs are likely contributing to trait expression and possibly patterns of among-population (co)variation. Panel A shows threespine stickleback fish (*Gasterosteus aculeatus*) and their cestode parasite *Schistocephalus*. Freshwater stickleback are the intermediate host for *Schistocephalus*, and each cestode acquires most of its lifetime resource pool while living inside a single host fish. *Schistocephalus* impose substantial reproductive and survival costs on hosts, and hosts have evolved an inducible (by the cestode) defense that suppresses cestode growth. Substantial among population-variation in this trait and in infection rates suggest these IIGEs may be mediating coevolution. Panel B shows goldenrod (*Solidago*) and a gall induced by the larvae (visible inside the gall) of the specialist gall-forming fly *Eurosta*. Gall size is induced by the genotype of the *Eurosta* larvae, and past work has shown that gall expression is a complex interaction between plant and fly genotype. Panel C shows the fungus *Ophiocordyceps unilateralis*, which manipulates behavior of its ant host prior to emergence of its fruiting body (Anderson et al. 2009). Panel D shows the water flea *Daphnia lumholtzi*, which induces growth of protective spines in response to chemical cues released by predatory fish (Agrawal 2001). Panel A and B main photos: S. De Lisle. Inset (B) photo: SriMesh / CC BY-SA (<https://creativecommons.org/licenses/by-sa/3.0>); C photo: "File:Ophiocordyceps unilateralis.png" by David P. Hughes, Maj-Britt Pontoppidan / CC BY 2.5; Panel D: "Water flea (*Daphnia lumholtzi*)" by Frupus / CC BY-NC 2.0.

108 ways that mitigate their risk of predator encounter (Werner and Peacor 2003, Preisser et al.
109 2005). Perhaps the most prominent example is the tendency of some *Daphnia* genotypes to
110 grow spines (Fig. 1D), or to migrate to other depths, when they detect scent cues
111 (kairomones) from predatory fish (Weber and Declerck 1997, Boersma et al. 1998). These
112 antipredator responses lead to systemic changes in gene expression and morphology (Tams et
113 al. 2019), which are controlled in part by fish traits (e.g., production of a scent cue). These
114 scents can themselves be variable and genetic, as illustrated by differences in how *Daphnia*
115 respond to cues from landlocked versus anadromous alewife (Walsh and Post 2011). The
116 *Daphnia* traits are thus IIGEs controlled by both *Daphnia* and fish genotypes. Unlike the
117 intimate host-parasite interactions, however, prey may be responding to diffuse cues from a
118 predator population as a whole.

119 Although IIGEs would appear to play a central role in trait interactions between many
120 coevolving species (broad categories summarized in Table 1), little is known about how these
121 effects influence the dynamics of coevolution (Scheiner et al. 2015). Past approaches, which
122 have included variance-partitioning models of community assembly (Shuster et al. 2006,
123 Whitham et al. 2020) and models of “joint traits” expressed together by interacting species
124 (Queller 2014, O’Brien et al. 2021), are suggestive of an important role for IIGEs in species
125 interactions. However, we lack a general understanding of how reciprocal IIGEs may affect
126 the coevolution of interacting phenotypes. The fact that indirect genetic effects within a
127 species can create feedback loops and other complex evolutionary dynamics, including in the
128 context of within-species coevolution (Drown and Wade 2014), suggests that IIGEs could
129 play a major role in mediating trait coevolution between species. Coopting concepts from
130 intraspecific social evolution theory, where trait-based IIGEs are well developed, thus
131 provides a natural way to understand the trait interactions that drive coevolution.
132 Importantly, this trait-based approach allows the contribution of indirect genetic effects to the

133 coevolutionary process to be fully explored. Our goal in this paper is to develop such models
134 to provide a comprehensive theoretical assessment of how IIGEs contribute to coevolution
135 between species.

136 Here we adapt the trait-based theory of intraspecific social evolution (Moore et al.
137 1997, Wolf et al. 1999, McGlothlin et al. 2010) to the case of two interacting species. Our
138 model applies to both pairwise interactions between individuals (e.g., stickleback and
139 cestodes) as well as diffuse interactions between species mean values (e.g., predatory fish and
140 *Daphnia*). Importantly, our model accommodates both IIGEs, where genes in individuals of
141 one species influence trait expression in individuals of another species, and cross-species
142 selection, where phenotypes of individuals in one species influence fitness of individuals of
143 another species (Figs. 1, 2; analogous to social selection in intraspecific social evolution
144 models). In addition to describing the contribution of these interspecific interactions to
145 evolutionary change, we develop expressions for the among-population evolutionary
146 covariance between traits of two interacting species; that is, the expected covariance in trait
147 means across populations between two coevolving species. In the process, we formalize an
148 interspecific analog of Zeng's (1988) quantitative genetic model of among-population trait
149 covariation, which we expand to incorporate interacting phenotypes. Our analysis shows that
150 IIGEs may have a central role in driving and mediating coevolution.

151

152 **Reciprocal evolutionary change in interacting species**

153 To model coevolution in two interacting species, we first decompose trait expression into
154 three components: direct genetic effects, environmental effects, and indirect effects mediated
155 by the phenotype of an interacting species. The phenotypic interface of coevolution involves
156 traits with interacting effects across individuals of two species x and y and can be written as

157

$$\mathbf{z}_x = \mathbf{a}_x + \mathbf{e}_x + \Phi_{xy}\mathbf{z}'_y \quad (1a)$$

$$\mathbf{z}_y = \mathbf{a}_y + \mathbf{e}_y + \Phi_{yx}\mathbf{z}'_x, \quad (1b)$$

158

159 where \mathbf{z}_i is a column vector of m traits expressed in an individual of species i (m is not
160 necessarily equal in each species), \mathbf{a}_i is the corresponding column vector of direct genetic
161 effects, and \mathbf{e}_i is an uncorrelated vector of residual environmental effects. The matrix Φ_{xy}
162 quantifies the effect of traits in an interacting individual of species y (\mathbf{z}'_y) on the expression of
163 traits in a focal individual of species x (Fig. 2, Table 2), while the matrix Φ_{yx} quantifies
164 such effects in the opposite direction. Thus, Φ_{ij} is an interspecific analog of the matrix of
165 conspecific indirect genetic effects, Ψ (Moore et al. 1997). The individual elements of Φ_{ij} ,
166 which we write as ϕ_{ij}^{kl} , represent partial regression coefficients of trait l in species j on trait k
167 in species i . When traits are standardized to the same scale, these coefficients will typically
168 be limited to a range of -1 to 1. Although we focus on interactions between pairs of
169 individuals for simplicity of notation, this model could be easily extended to describe
170 interactions with multiple individuals (cf. McGlothlin et al. 2010). For more diffuse
171 interactions (e.g., alewife and *Daphnia*), the elements of Φ_{ij} could represent a weighted
172 average of effects from integrating across the phenotype distribution of the interacting
173 population.

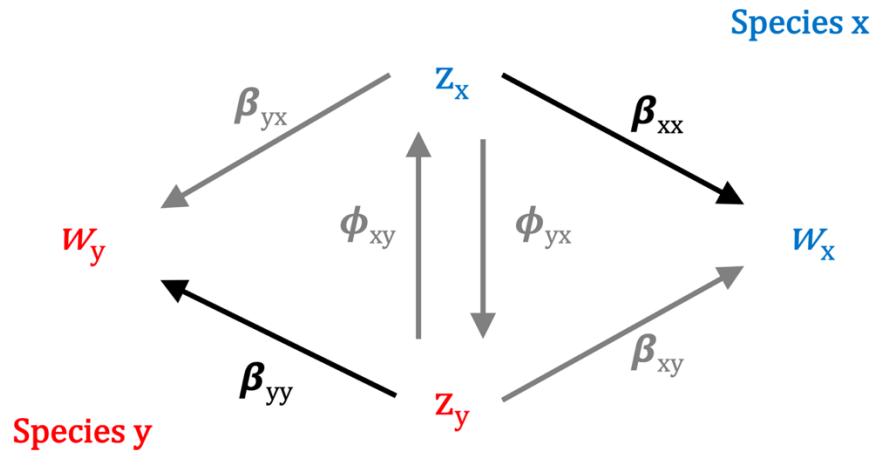


Figure 2. Path model of selection when traits and fitness are determined by interactions with heterospecifics. Individual trait values of two interacting species, x (blue) and y (red), are represented by z . Traits directly influence individual relative fitness (w) of the species that express them via natural selection (β_{xx} and β_{yy} , black arrows). Traits can also reciprocally influence expression of traits of heterospecific social partners, via interspecific indirect genetic effects Φ . Traits can also influence fitness of heterospecific social partners, via cross-species selection (β_{xy} and β_{yx} , grey arrows). Social effects are illustrated in grey arrows, direct effects in black.

Table 2. Definition of key parameters and expressions.

Expression / parameter	Biological definition
$\mathbf{z}_x, \mathbf{z}_y$	Vectors of individual trait values for species x and y , with a prime when present denoting partner traits.
$\mathbf{a}_x, \mathbf{a}_y$	Vectors of individual breeding values for species x and y .
Φ_{xy}, Φ_{yx}	Interspecific indirect effects (in matrix form). The effect of traits in individuals of species y on trait expression in species x , and vice-versa.
β_{xx}, β_{yy}	Vectors of within-species natural selection gradients for each species.
β_{xy}, β_{yx}	Vectors of cross-species selection gradients for each species. The effects of traits in species y on fitness of individual species x , and vice-versa.
$\mathbf{G}_{xx}, \mathbf{G}_{yy}$	Genetic covariance matrices of traits for species x and y .
$\mathbf{G}_{xy}, \mathbf{G}_{yx}$	Covariance matrices for breeding values of interacting individuals of species x and y within a population.
$\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y)$	Coevolutionary covariance, defined as the covariance in evolutionary change between interacting species, across populations. When standardized by the variances in evolutionary response for each species, becomes a scale-free correlation.
$\text{Cov}(\beta_{xx}, \beta_{yy}),$	Between-species, among-population covariance in selection gradients (shown here for within-species selection)

174 Assuming that the mean residual environmental effect is zero, the population mean
 175 phenotype vector for each species is

176

$$\bar{\mathbf{z}}_x = \bar{\mathbf{a}}_x + \mathbf{\Phi}_{xy} \bar{\mathbf{z}}_y \quad (2a)$$

$$\bar{\mathbf{z}}_y = \bar{\mathbf{a}}_y + \mathbf{\Phi}_{yx} \bar{\mathbf{z}}_x. \quad (2b)$$

177

178 Noting the change in the mean additive genetic value in each species is $\Delta \bar{\mathbf{a}} = \text{Cov}(\mathbf{a}, w)$

179 (Robertson 1966, Price 1970, 1972) where w is relative fitness, we can now define

180 evolutionary change in the multivariate mean phenotype as

181

$$\Delta \bar{\mathbf{z}}_x = \text{Cov}(\mathbf{a}_x, w_x) + \mathbf{\Phi}_{xy} \Delta \bar{\mathbf{z}}_y \quad (3a)$$

$$\Delta \bar{\mathbf{z}}_y = \text{Cov}(\mathbf{a}_y, w_y) + \mathbf{\Phi}_{yx} \Delta \bar{\mathbf{z}}_x. \quad (3b)$$

182

183 In equation (3), the first term on each right-hand side describes direct change due to natural

184 selection on the focal species, and the second term describes indirect change due to the

185 product of IIGEs and phenotypic change in the interacting species. It can be seen from

186 equation (3) that species will coevolve whenever there are IIGEs in both species, because the

187 change in phenotypic mean in species x depends upon the change of mean in species y and

188 vice versa whenever both $\mathbf{\Phi}_{ij} \neq \mathbf{0}$. Expanding equation (3) by substitution results in explicit

189 equations for evolutionary change in each species,

190

$$\Delta \bar{\mathbf{z}}_x = (\mathbf{I}_x - \mathbf{\Phi}_{xy} \mathbf{\Phi}_{yx})^{-1} [\text{Cov}(\mathbf{a}_x, w_x) + \mathbf{\Phi}_{xy} \text{Cov}(\mathbf{a}_y, w_y)] \quad (4a)$$

$$\Delta \bar{\mathbf{z}}_y = (\mathbf{I}_y - \mathbf{\Phi}_{yx} \mathbf{\Phi}_{xy})^{-1} [\text{Cov}(\mathbf{a}_y, w_y) + \mathbf{\Phi}_{yx} \text{Cov}(\mathbf{a}_x, w_x)]. \quad (4b)$$

191

192 where \mathbf{I}_x and \mathbf{I}_y are identity matrices with dimensionality equal to the number of traits in the
193 two species and the multiplier $(\mathbf{I} - \Phi_{ij}\Phi_{ji})^{-1}$ quantifies the feedback effect of reciprocal
194 IIGEs. Whenever IIGEs occur in both species such that $\Phi_{ij}\Phi_{ji} \neq \mathbf{0}$, this multiplier alters the
195 total amount of evolutionary change in both species. In order for such an effect to arise, there
196 must be a feedback loop in phenotypic expression. The simplest of these arises when there
197 are two traits with reciprocal IIGEs, such that trait k in species i affects trait l in species j and
198 trait l in turn influences the expression of trait k . In general, when ϕ_{ij}^{kl} and ϕ_{ji}^{lk} are of the
199 same sign, the magnitude of evolutionary change will be enhanced, and when they are of
200 opposite signs, evolutionary change will be diminished (Fig. 3).

201 To expand equation (4), we define the fitness of each interacting species using the
202 linear equations

203

$$w_x = \alpha_x + \mathbf{z}_x^T \boldsymbol{\beta}_{xx} + \mathbf{z}'_y^T \boldsymbol{\beta}_{xy} + \varepsilon_x \quad (5a)$$

$$w_y = \alpha_y + \mathbf{z}_y^T \boldsymbol{\beta}_{yy} + \mathbf{z}'_x^T \boldsymbol{\beta}_{yx} + \varepsilon_y \quad (5b)$$

204

205 where w is individual relative fitness, α is an intercept, ε is an error term, and the superscript
206 T denotes transposition. Directional selection in each species is partitioned into two selection
207 gradients. First, the within-species selection gradients ($\boldsymbol{\beta}_{xx}$ and $\boldsymbol{\beta}_{yy}$) describe the direct
208 effects of an individual's traits on its own fitness (Fig. 2; Lande and Arnold 1983). The
209 cross-species selection gradients ($\boldsymbol{\beta}_{xy}$ and $\boldsymbol{\beta}_{yx}$) relate the fitness of a focal individual of one
210 species (x or y) to the traits of individuals of the other coevolving species (y or x) (Fig. 2).
211 Both of these are linear components of selection; we consider nonlinear terms (interactions
212 for fitness between focal and partner traits) below (see Incorporating Specific Fitness
213 Models). The cross-species selection gradient is analogous to the directional social selection

214 gradient in within-species models (Wolf et al. 1999) and if pairs or groups of interacting
 215 individuals can be identified in a natural population, can be estimated in a similar way (cf.
 216 Ridenhour 2005).

217 We now substitute our definitions of fitness into equation (4) and expand, yielding
 218 predictive equations for evolutionary change in each species:

219

$$\Delta \bar{z}_x = (\mathbf{I}_x - \Phi_{xy} \Phi_{yx})^{-1} [\text{Cov}(\mathbf{a}_x, \mathbf{z}_x^T) \boldsymbol{\beta}_{xx} + \text{Cov}(\mathbf{a}_x, \mathbf{z}'_y^T) \boldsymbol{\beta}_{xy} + \Phi_{xy} \text{Cov}(\mathbf{a}_y, \mathbf{z}_y^T) \boldsymbol{\beta}_{yy} + \Phi_{xy} \text{Cov}(\mathbf{a}_y, \mathbf{z}'_x^T) \boldsymbol{\beta}_{yx}] \quad (6a)$$

220

$$\Delta \bar{z}_y = (\mathbf{I}_y - \Phi_{yx} \Phi_{xy})^{-1} [\text{Cov}(\mathbf{a}_y, \mathbf{z}_y^T) \boldsymbol{\beta}_{yy} + \text{Cov}(\mathbf{a}_x, \mathbf{z}_x^T) \boldsymbol{\beta}_{yx} + \Phi_{yx} \text{Cov}(\mathbf{a}_x, \mathbf{z}_x^T) \boldsymbol{\beta}_{xx} + \Phi_{yx} \text{Cov}(\mathbf{a}_x, \mathbf{z}'_y^T) \boldsymbol{\beta}_{xy}]. \quad (6b)$$

221

222 Each of these equations consists of four terms representing four components of the total
 223 response to selection. The first term ($\text{Cov}(\mathbf{a}_x, \mathbf{z}_x^T) \boldsymbol{\beta}_{xx}$) represents response to within-species
 224 selection ($\boldsymbol{\beta}_{xx}$ or $\boldsymbol{\beta}_{yy}$), the second ($\text{Cov}(\mathbf{a}_x, \mathbf{z}'_y^T) \boldsymbol{\beta}_{xy}$) represents response to cross-species
 225 linear selection ($\boldsymbol{\beta}_{xy}$ or $\boldsymbol{\beta}_{yx}$), and the last two terms ($\Phi_{xy} \text{Cov}(\mathbf{a}_y, \mathbf{z}_y^T) \boldsymbol{\beta}_{yy} +$
 226 $\Phi_{xy} \text{Cov}(\mathbf{a}_y, \mathbf{z}'_x^T) \boldsymbol{\beta}_{yx}$) represent the component of change caused by the change in mean of
 227 the interacting species. Note that the third and fourth terms of the first part of equation (6a)
 228 equal the first two terms of (6b) multiplied by the IIGE coefficient Φ_{xy} . Equation (6) also
 229 shows that the change in response to within-species selection depends on the covariance of
 230 additive genetic value with the phenotype of the same species, while change in response to
 231 cross-species selection depends on the covariance of additive genetic value with the
 232 phenotype of the opposite species.

233 To determine the components that give rise to these covariances, we expand our
 234 definition of the phenotypes in equation (2) and rearrange, yielding

235

$$\begin{aligned}
 236 \quad \Delta \bar{z}_x = & (\mathbf{I}_x - \Phi_{xy} \Phi_{yx})^{-1} [(\mathbf{G}_{xx} + \mathbf{G}_{xy} \Phi_{xy}^T)(\mathbf{I}_x - \Phi_{yx}^T \Phi_{xy}^T)^{-1} \boldsymbol{\beta}_{xx} + & (7a) \\
 237 & (\mathbf{G}_{xy} + \mathbf{G}_{xx} \Phi_{yx}^T)(\mathbf{I}_y - \Phi_{xy}^T \Phi_{yx}^T)^{-1} \boldsymbol{\beta}_{xy} + \\
 238 & \Phi_{xy}(\mathbf{G}_{yy} + \mathbf{G}_{yx} \Phi_{yx}^T)(\mathbf{I}_y - \Phi_{xy}^T \Phi_{yx}^T)^{-1} \boldsymbol{\beta}_{yy} + \\
 239 & \Phi_{xy}(\mathbf{G}_{yx} + \mathbf{G}_{yy} \Phi_{xy}^T)(\mathbf{I}_x - \Phi_{yx}^T \Phi_{xy}^T)^{-1} \boldsymbol{\beta}_{yx}]
 \end{aligned}$$

240

$$\begin{aligned}
 241 \quad \Delta \bar{z}_y = & (\mathbf{I}_y - \Phi_{yx} \Phi_{xy})^{-1} [(\mathbf{G}_{yy} + \mathbf{G}_{yx} \Phi_{yx}^T)(\mathbf{I}_y - \Phi_{xy}^T \Phi_{yx}^T)^{-1} \boldsymbol{\beta}_{yy} + & (7b) \\
 242 & (\mathbf{G}_{yx} + \mathbf{G}_{yy} \Phi_{xy}^T)(\mathbf{I}_x - \Phi_{yx}^T \Phi_{xy}^T)^{-1} \boldsymbol{\beta}_{yx} + \\
 243 & \Phi_{yx}(\mathbf{G}_{xx} + \mathbf{G}_{xy} \Phi_{xy}^T)(\mathbf{I}_x - \Phi_{yx}^T \Phi_{xy}^T)^{-1} \boldsymbol{\beta}_{xx} + \\
 244 & \Phi_{yx}(\mathbf{G}_{xy} + \mathbf{G}_{xx} \Phi_{yx}^T)(\mathbf{I}_y - \Phi_{xy}^T \Phi_{yx}^T)^{-1} \boldsymbol{\beta}_{xy}],
 \end{aligned}$$

245

246 where \mathbf{G}_{xx} and \mathbf{G}_{yy} represent within-species genetic (co)variance matrices and \mathbf{G}_{xy} and \mathbf{G}_{yx}
 247 represent cross-species genetic covariance. Note that in a model of pairwise interactions,
 248 $\mathbf{G}_{yx} = \mathbf{G}_{xy}^T$. The cross-species genetic covariance represents a covariance of breeding values
 249 between the species within the population of interest. This covariance may arise via any
 250 mechanism that leads to nonrandom genetic assortment between species x and y , including
 251 behavioral preference for certain traits in heterospecific partners, fine-scale population
 252 structure, and habitat preference. For example, a nonzero \mathbf{G}_{xy} could arise if a phytophagous
 253 insect showed genetic variance in host-plant choice based on genotypes of the plant. We
 254 elaborate on the contribution of \mathbf{G}_{xy} and Φ to phenotypic assortment between individuals of
 255 interacting species (C_{xy}) in equation (A1).

256 Response to within-species selection depends on the sum of within-species genetic
 257 variance, \mathbf{G}_{yy} or \mathbf{G}_{xx} , and $\mathbf{G}_{xy} \Phi_{xy}^T$ or $\mathbf{G}_{yx} \Phi_{yx}^T$, which represent an interaction between IIGEs
 258 and cross-species genetic covariance. Response to cross-species selection depends on the
 259 sum of cross-species genetic variance and $\mathbf{G}_{xx} \Phi_{yx}^T$ or $\mathbf{G}_{yy} \Phi_{xy}^T$, which describes the genetic
 260 variance created by IIGEs. Each term in equation (7) also contains an additional feedback
 261 multiplier, $(\mathbf{I}_y - \Phi_{xy}^T \Phi_{yx}^T)^{-1}$, which further enhances the response to selection when there
 262 are reciprocal IIGEs. Because of this additional multiplier, when IIGEs are reciprocal and of
 263 the same sign, their effects on can be massive, mirroring the effects observed for within-
 264 species IGEs (Moore et al. 1997, McGlothlin et al. 2010). As in previous equations, the last
 265 two terms in equation (7) represent a sort of evolutionary feedback that occurs across
 266 generations and is only present when there are IIGEs. These effects of Φ and cross-species
 267 selection on evolutionary response are illustrated in Fig. 3.

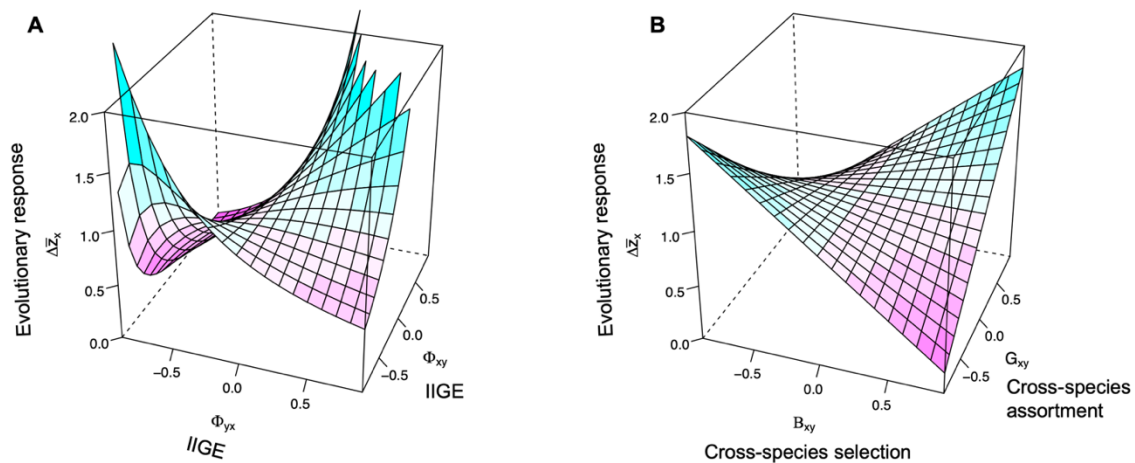


Figure 3. Reciprocal IIGEs and cross-species selection change evolutionary response in a single species. Panels show the separate effects on evolutionary response in species x of indirect genetic effects (Panel A) and cross-species selection with genetic assortment (Panel B). Panel A shows the effects of reciprocal IIGEs holding all other evolutionary parameters constant, and assuming no cross-species selection. Panel B shows the effects of cross-species selection imposed by species y on species x, in combination with the genetic assortment between interactants of the different species, and assuming IIGEs are absent. In both panels, $G_x = G_y = 1$, $\beta_{yy} = \beta_{xx} = 1$.

268 **Coevolution across a selection mosaic: the coevolutionary covariance**

269 When response to selection in one species depends on trait evolution in another species, the
270 species can coevolve. When selection varies through space or time due to varying ecological
271 conditions, so too does the position of each population of each species in trait space. For
272 coevolving lineages imposing reciprocal selection on each other, we expect correlated
273 evolution of interacting traits both through time and across replicate populations of both
274 species in space. In this case, local ecological drivers of natural selection may generate a
275 mosaic of varying selection across populations, mediating where each population falls on an
276 axis of correlated evolution and generating a “geographic mosaic” of coevolution (Thompson
277 1994, 2005). In a coevolutionary arms race, for example, this is expected to yield
278 populations where each interacting lineage has elevated trait values (“hot spots”), as well
279 other populations where the race appears deescalated (“cold spots”) (e.g., elevated toxin
280 levels and toxin resistance across prey and their predators; Brodie et al. 2002, Hanifin et al.
281 2008), and still other populations where one lineage appears to have the upper hand (Arnqvist
282 and Rowe 2002, Zangerl and Barenbaum 2003, Toju and Sota 2005, Perry and Rowe 2011,
283 Hague et al. 2020). Thus, a key feature of coevolving lineages is correlated evolution across
284 populations subject to varying ecological conditions, creating a geographic mosaic of
285 (co)varying mean phenotypes. Here, we seek to understand how the equations of selection
286 response can be used to understand how selection and IIGEs contribute to this shared among-
287 population divergence.

288 We can explore the contribution of interspecific social effects to correlated evolution
289 between interacting species by solving for the covariance in evolutionary change between
290 interacting species, $\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y)$, which we call the coevolutionary covariance. Such a
291 covariance represents the expected pattern of coevolution through time in a single pair of
292 populations of two species experiencing fluctuating selection, or perhaps more importantly,

293 across a set of populations in space under varying selection pressures. Although
294 mathematically equivalent, we focus on the latter scenario for its relevance to understanding
295 geographic variation among populations.

296 The coevolutionary covariance reflects the degree to which divergence among
297 population means in two species has occurred jointly. High absolute values of the
298 coevolutionary covariance indicate tightly-coupled coevolutionary change between the two
299 species, whereas values around zero indicate that evolutionary change occurs independently.
300 When scaled by the total amount of population divergence (variances in $\Delta\bar{z}_x$ and $\Delta\bar{z}_y$), this
301 becomes a scale-free correlation describing the proportion of total divergence shared between
302 the two species. Importantly, we expect the coevolutionary covariance to be related to the
303 covariance in population means, $\text{Cov}(\bar{z}_x, \bar{z}_y) \propto \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y)$. Thus, expanding
304 $\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y)$ allows the possibility to assess how selection and IIGEs contribute to patterns
305 of geographic variation in species mean phenotypes.

306 To simplify our analysis, we use models of a single trait in each species. Developing
307 a full equation including all sources of covariance quickly becomes cumbersome, so we focus
308 on three instructive special cases that illustrate the explicit impacts of considering cross-
309 species selection and interspecific interacting phenotypes. The simplest case occurs when
310 there are no IIGEs ($\phi_{xy} = \phi_{yx} = 0$) and heterospecific interactions occur at random within
311 populations ($G_{xy} = G_{yx} = 0$). If we make the simplifying assumption that genetic variance
312 does not differ among populations, the only source of covariance in selection response is
313 covariance in direct within-species selection, or

314

$$\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = G_{xx} G_{yy} \text{Cov}(\beta_{xx}, \beta_{yy}). \quad (8)$$

315

316 Equation (8) represents what is usually thought of as the source of a coevolutionary mosaic:
317 predictable variation in the type of selection occurring across space. Empirical data suggest
318 that selection varies substantially in magnitude across space (Siepielski et al. 2013) and that
319 spatially autocorrelated biotic selection plays a substantial role in driving divergence in trait
320 means (Urban et al. 2011). Importantly, not just any variance in selection will do. In order to
321 create covariance in evolutionary change, selection must covary between the interacting
322 species. Cross-species selection, however, does not play a role in equation (8) because it
323 does not contribute to an evolutionary response to selection in the absence of IIGEs and
324 cross-species genetic covariance. Equation (8) is analogous to the results of Zeng's (1988)
325 model of correlated trait evolution under directional selection, although for the case of traits
326 expressed in different species.

327 To see an effect of cross-species selection, we first add cross-species genetic
328 covariance but no IIGEs. For a one-trait (per species) model, $G_{xy} = G_{yx}$. Adding this effect
329 leads to three new sources of covariance among populations:

330

$$\begin{aligned} \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = & G_{xx}G_{yy}\text{Cov}(\beta_{xx}, \beta_{yy}) + G_{xy}^2\text{Cov}(\beta_{yx}, \beta_{xy}) + \\ & G_{xx}G_{xy}\text{Cov}(\beta_{xx}, \beta_{yx}) + G_{yy}G_{xy}\text{Cov}(\beta_{yy}, \beta_{xy}). \end{aligned} \quad (9)$$

331

332 The second term in equation (9) ($G_{xy}^2\text{Cov}(\beta_{yx}, \beta_{xy})$) shows that cross-species selection will
333 contribute to the coevolutionary covariance whenever there is cross-species genetic
334 covariance. Such covariances occur if, for example, parasite and host genotypes associate
335 nonrandomly and have traits that influence their partner's fitness. The third and fourth terms
336 ($G_{xx}G_{xy}\text{Cov}(\beta_{xx}, \beta_{yx}) + G_{yy}G_{xy}\text{Cov}(\beta_{yy}, \beta_{xy})$) represent a relationship between the effect
337 of species on itself and on its heterospecific partner. These will be nonzero if populations
338 with strong within-species selection also exhibit strong cross-species selection.

339 The most interesting effects on the coevolutionary covariance occur when we add
 340 IIGEs (ϕ_{xy} and ϕ_{yx}). With $G_{xy} = 0$, the covariance among populations in evolutionary
 341 response to selection is

$$\begin{aligned}
 \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = & \mathcal{U}[G_{xx}G_{yy}\text{Cov}(\beta_{xx}, \beta_{yy}) + \\
 & G_{xx}^2\phi_{yx}\text{Var}(\beta_{xx}) + G_{yy}^2\phi_{xy}\text{Var}(\beta_{yy}) + \\
 & G_{xx}G_{yy}\phi_{xy}\phi_{yx}\text{Cov}(\beta_{xx}, \beta_{yy}) + G_{xx}G_{yy}\phi_{xy}\phi_{yx}\text{Cov}(\beta_{xy}, \beta_{yx}) + \\
 & G_{xx}G_{yy}\phi_{xy}\text{Cov}(\beta_{xx}, \beta_{yx}) + G_{xx}G_{yy}\phi_{yx}\text{Cov}(\beta_{yy}, \beta_{xy}) + \\
 & 2G_{xx}^2\phi_{yx}^2\text{Cov}(\beta_{xx}, \beta_{xy}) + 2G_{yy}^2\phi_{xy}^2\text{Cov}(\beta_{yy}, \beta_{yx}) + \mathcal{V}],
 \end{aligned} \tag{10}$$

343
 344 where feedback effects of IIGEs are described by $\mathcal{U} = (1 - \phi_{xy}\phi_{yx})^{-4}$ and \mathcal{V} collects
 345 negligible third- and fourth-order ϕ terms:

$$\begin{aligned}
 \mathcal{V} = & G_{xx}^2\phi_{yx}^3\text{Var}(\beta_{xy}) + G_{yy}^2\phi_{xy}^3\text{Var}(\beta_{yx}) + \\
 & G_{xx}G_{yy}\phi_{xy}\phi_{yx}^2\text{Cov}(\beta_{yy}, \beta_{xy}) + G_{xx}G_{yy}\phi_{xy}^2\phi_{yx}\text{Cov}(\beta_{yx}, \beta_{xx}) + \\
 & G_{xx}G_{yy}\phi_{xy}^2\phi_{yx}^2\text{Cov}(\beta_{yx}, \beta_{xy}).
 \end{aligned} \tag{11}$$

347
 348 The first term in equation (10) is identical to equation (8) and represents covariance in direct
 349 within-species selection. The second ($G_{xx}^2\phi_{yx}\text{Var}(\beta_{xx})$) and third ($G_{yy}^2\phi_{xy}\text{Var}(\beta_{yy})$) terms
 350 show that in the presence of IIGEs, simple variance in within-species selection across
 351 populations can generate a coevolutionary covariance (or correlation, Fig. 4A; corresponding

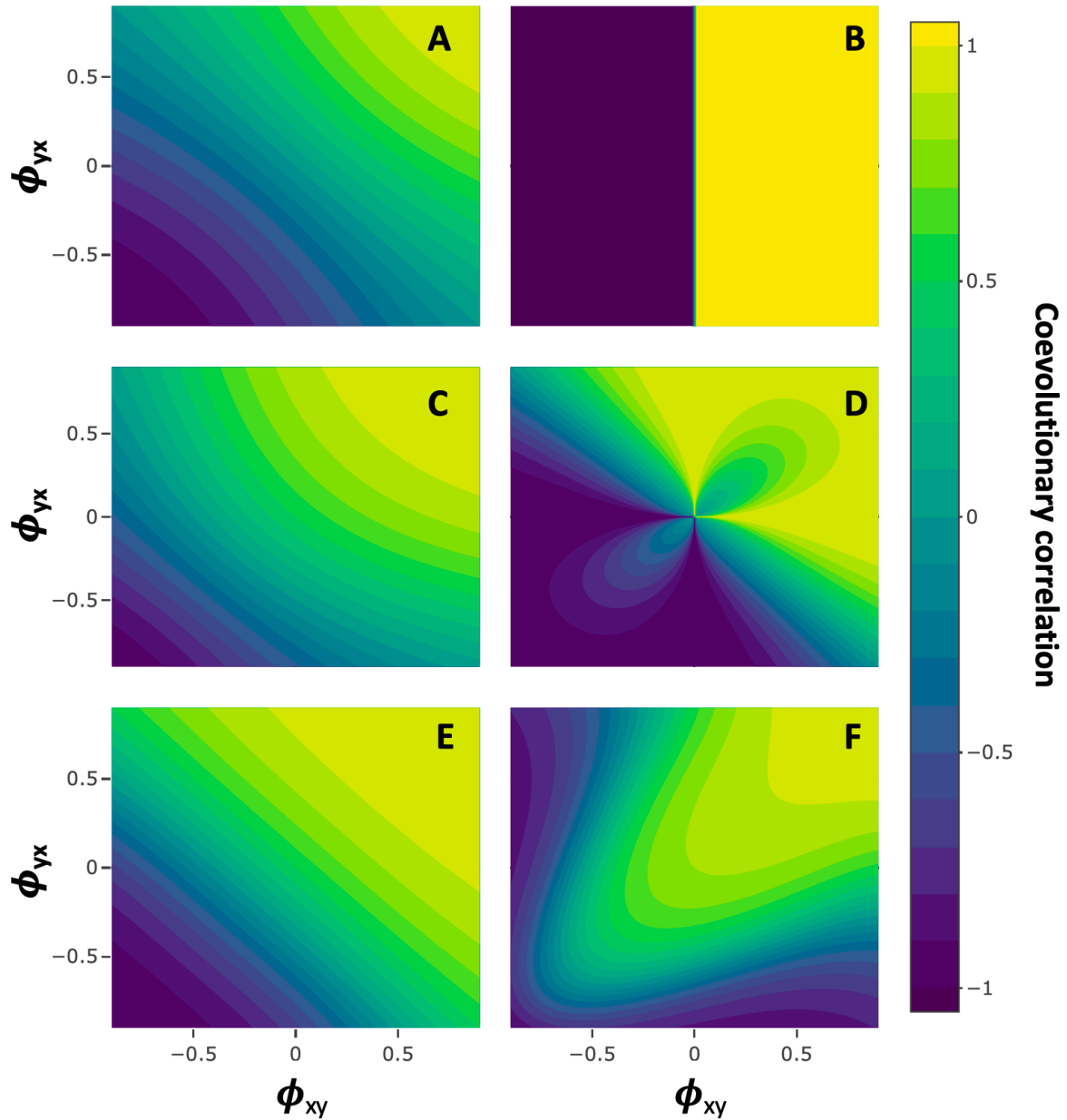


Figure 4. IIGEs drive and mediate coevolution between interacting species. Panels show the coevolutionary correlation between two interacting species as a function of the indirect genetic effect parameters ϕ , from equation (7) standardized by the evolutionary rates. In the absence of social interaction effects, correlated evolution (coevolution) between species is driven entirely by covariance in natural selection between the species (see text). Reciprocal IIGEs can generate coevolution even when there is not covariance in natural selection (Panel A), and can even drive coevolution even when one species lacks genetic variance (Panel B). IIGEs modify observed coevolutionary patterns when natural selection does covary ($\text{Cov}(\beta_{xx}, \beta_{yy}) = 0.5$; Panel C). When cross-species and IIGEs act together, coevolutionary patterns are a complex third order polynomial (Panel D; $\text{Cov}(\beta_{xy}, \beta_{yx}) = 0.5$, $\text{Var}(\beta_{xy}) = \text{Var}(\beta_{yx}) = 1$). When natural and cross-species selection both act ($\text{Var}(\beta_{xy}) = \text{Var}(\beta_{yx}) = \text{Var}(\beta_{yy}) = \text{Var}(\beta_{xx}) = 1$) and covary positively (0.5), (Panel E), effects of IIGEs become stronger in comparison to the case of (co)variance in natural selection alone. When covariance between natural and cross-species selection is negative (-0.8) a ridge is observed (Panel F).

353 evolutionary rates and covariances are plotted in Figs. S1 and S2, respectively). This
354 covariance in evolutionary response occurs as a necessary consequence of the dependence of
355 the trait mean of one species on that of the other. Thus, in the presence of IIGEs, a
356 coevolutionary covariance may occur even when selection is uncorrelated between the
357 species, and in extreme cases, when only one species varies in selection or when only species
358 has genetic variation (Fig. 4B).

359 The fourth and fifth terms ($G_{xx}G_{yy}\phi_{xy}\phi_{yx}\text{Cov}(\beta_{xx}, \beta_{yy}) +$
360 ($G_{xx}G_{yy}\phi_{xy}\phi_{yx}\text{Cov}(\beta_{xy}, \beta_{yx})$) show the effects of reciprocal IIGEs. When traits in the two
361 species exist in a feedback loop, the effect of covariance in within-species selection is
362 amplified if ϕ_{xy} and ϕ_{yx} are of the same sign and diminished if they are of opposite signs.
363 The combined effects of the second, third, and fourth terms lead to a complex relationship
364 between IIGEs and the coevolutionary covariance when within-species selection varies (Fig.
365 4C). In some cases, IIGEs can even reverse the sign of the correlation that would be
366 expected in their absence (Fig. 4C). In addition, reciprocal IIGEs may cause covariance in
367 cross-species selection to contribute to the coevolutionary covariance, causing an even more
368 complex relationship (Fig. 4D).

369 The last four terms in equation (10) show that in the presence of IIGEs, covariance
370 between within-species and cross-species selection may contribute to the coevolutionary
371 covariance (Fig. 4E-F). Because IIGEs inherently tie together the evolutionary responses of
372 the two species through their effects on the coevolutionary covariance, this can occur both
373 when there is covariance in gradients across species (terms 6 and 7) and when there is
374 covariance in gradients in the same species (terms 8 and 9). The total effect of IIGEs may be
375 quite complex when cross-species selection varies, with subtle changes in ϕ resulting in
376 dramatic changes in the expected among population correlation (Fig. 4D, E-F).

377 An equation incorporating both IIGEs and G_{xy} quickly becomes unwieldy, but we
378 present a compact form containing 10 (co)variance terms as equation (A2), which illustrates
379 that when present, non-random assortment and IIGEs together interact in complex ways to
380 influence coevolution.

381

382 **Incorporating specific fitness models**

383 Coevolutionary models often posit complex relationships between interacting phenotypes and
384 fitness (Nuismer 2017). Although the selection model we present here is linear, more
385 complex relationships can be incorporated by translating specific fitness functions to
386 selection gradients. The adaptive landscape represents the theoretical relationship between a
387 population's mean fitness and its phenotypic mean (Arnold et al. 2001). Selection gradients
388 represent the partial slope of the adaptive landscape with respect to a given phenotype, and
389 thus the multivariate selection gradient may be calculated using a vector of partial derivatives
390 if the adaptive landscape can be written as a differentiable function (Lande 1979, Lande and
391 Arnold 1983). In many cases, the multivariate selection gradient may be calculated using
392 partial derivatives of the individual fitness function as well (Lande and Arnold 1983, Abrams
393 et al. 1993, McGlothlin et al. 2021). Once selection gradients have been calculated for a
394 given model, they may be substituted into equations (6–7) to explore the effects of a given
395 fitness function on selection response and the coevolutionary covariance (see also Brodie and
396 Ridenhour 2003). This exercise also allows us to explore the effects of adding IIGEs to an
397 existing coevolutionary model.

398 First, we consider a “phenotypic difference,” or trait-matching, model of ecological
399 trait interaction (Nuismer et al. 2007, Nuismer 2017), where absolute fitness is a function of
400 the difference between interacting traits, $W_x \propto \exp(z_x - z_y)$ and $W_y \propto \exp(z_y - z_x)$.

401 Using the logarithm of the fitness function to calculate relative fitness (Lande and Arnold
402 1983),

403

$$\begin{aligned}w_x &= \alpha_x + b_x(z_x - z_y) + \varepsilon_x \\w_y &= \alpha_y + b_y(z_y - z_x) + \varepsilon_y.\end{aligned}\tag{12}$$

404

405 where b_x and b_y are constants that may vary across populations. Differentiating, the selection
406 gradients are then $\beta_{xx} = -\beta_{xy} = b_x$ and $\beta_{yy} = -\beta_{yx} = b_y$. Substituting these selection
407 gradients into equation (7) is trivial. However, is worth noting that under this fitness model,
408 the coevolutionary covariance simplifies to a function of just three (co)variance components,

409

$$\begin{aligned}\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) &= \mathcal{U}[G_{xx}G_{yy}(1 - \phi_{xy} - \phi_{yx} + 2\phi_{xy}\phi_{yx})\text{Cov}(b_x, b_y) + \\&G_{xx}^2\phi_{yx}(1 - 2\phi_{yx})\text{Var}(b_x) + G_{yy}^2\phi_{xy}(1 - 2\phi_{xy})\text{Var}(b_y) + \mathcal{V}],\end{aligned}\tag{13}$$

410

411 which corresponds to a relationship with ϕ as in Figs. 4F, S1F, and S2F. This analysis
412 illustrates that reciprocal IIGEs have greatest impact on mediating coevolutionary outcomes
413 in a trait-matching models when IIGEs are similar in both sign and magnitude. Biologically,
414 such a situation corresponds to a scenario where, for example, trait expression is reciprocally
415 escalated in response to heterospecific partners.

416

Another important fitness model to consider is the case of nonlinear fitness
417 interactions. Nonlinearity is fundamental to many coevolutionary models that invoke
418 epistatic interactions across species' genomes, such as the matching-allele or gene-for-gene

419 models of host-parasite coevolution (Dybdahl et al. 2014). To simplify, consider a fitness
420 model where relative fitness depends solely on the product of the two phenotypes:

421

$$\begin{aligned}w_x &= \alpha_x + b_{xy}z_xz_y + \varepsilon_x \\w_y &= \alpha_y + b_{yx}z_yz_x + \varepsilon_y.\end{aligned}\tag{14}$$

422

423 Under this fitness model, all selection gradients become dependent on the population means
424 of species x and y :

425

$$\begin{aligned}\beta_{xx} &= b_{xy}\bar{z}_y \\ \beta_{xy} &= b_{xy}\bar{z}_x \\ \beta_{yy} &= b_{yx}\bar{z}_x \\ \beta_{yx} &= b_{yx}\bar{z}_y\end{aligned}\tag{15}$$

426

427 This relationship may cause selection to (co)vary across populations even when b_{xy} and b_{yx}
428 are homogeneous. In the absence of IIGEs, this simplest case would lead to a coevolutionary
429 covariance defined by

430

$$\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = G_{xx}G_{yy}b_{xy}b_{yx}\text{Cov}(\bar{z}_x, \bar{z}_y).\tag{16}$$

431 Thus, any initial covariance in the population means leads to a covariance in the response to
 432 selection across species, lending a runaway aspect to the coevolutionary covariance. Adding
 433 IIGEs, this becomes

434

$$\begin{aligned}
 \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = & \mathcal{U}[G_{xx}G_{yy}b_{xy}b_{yx}\text{Cov}(\bar{z}_x, \bar{z}_y) \\
 & + G_{xx}^2\phi_{yx}b_{xy}^2\text{Var}(\bar{z}_y) + G_{yy}^2\phi_{xy}b_{yx}^2\text{Var}(\bar{z}_x) + \\
 & 2G_{xx}G_{yy}\phi_{xy}\phi_{yx}b_{xy}b_{yx}\text{Cov}(\bar{z}_x, \bar{z}_y) \tag{17} \\
 & + G_{xx}G_{yy}\phi_{xy}b_{xy}b_{yx}\text{Var}(\bar{z}_y) + G_{xx}G_{yy}\phi_{yx}b_{yx}b_{xy}\text{Var}(\bar{z}_x) + \\
 & 2G_{xx}^2\phi_{yx}^2b_{xy}^2\text{Cov}(\bar{z}_x, \bar{z}_y) + 2G_{yy}^2\phi_{xy}^2b_{yx}^2\text{Cov}(\bar{z}_x, \bar{z}_y) + \mathcal{V}].
 \end{aligned}$$

435

436 Although this equation is complex, it shows that when IIGEs are present any variance across
 437 populations in the trait mean of either species leads to a cross-species covariance in the
 438 response to selection (terms 2, 3, 5, and 6). Cross species covariance in selection response is
 439 also mediated further by covariance in trait means in this model (terms 1, 4, 7, and 8) as well
 440 as higher order products of IIGEs captured in \mathcal{V} (equation 11). These effects may lead trait
 441 means to become correlated across populations in future generations, further intensifying the
 442 geographic mosaic (Thompson 1994).

443

444 **Discussion**

445 Our model adapts the theory of trait-based intraspecific social evolution to the phenotypic
 446 interface between two coevolving species. We show that two forms of interspecific
 447 interaction, interspecific indirect genetic effects (IIGEs) and cross-species selection
 448 (analogous to within-species social selection), both contribute to correlated evolution
 449 between interacting species. Our analysis shows that reciprocal IIGEs modulate selection
 450 response, suggesting that IIGEs may play a major role in generating and mediating patterns

451 of correlated evolution between species. Further, we show that constant (across space) IIGEs
452 can generate a coevolutionary covariance (and thus a geographic mosaic) even in the absence
453 of covariance in selection, or even the absence of genetic variance in one species. When
454 selection does covary between species across populations, reciprocal IIGEs will promote
455 changes in the magnitude of coevolution and even reversals in the expected among-
456 population covariance. IIGEs also allow cross-species selection, which we model as the
457 effect of the traits of one species on the fitness of another, to influence evolutionary response.
458 Such a response may also be mediated by cross-species genetic assortment between
459 interacting individuals. When IIGEs and cross-species selection act together, effects on the
460 coevolutionary mosaic can be complex, with dramatic changes in the expected sign and
461 magnitude of correlated evolution occurring with subtle changes in these parameters of
462 interspecific social interaction. Our results indicate that whenever coevolving species
463 socially interact to modify expression of one another's phenotypes, these interspecific social
464 interactions are key to understanding coevolution.

465 IIGEs represent a scenario where phenotypes in individuals of one species influence
466 trait expression of individuals of another species. Thus, IIGEs are a specific type of
467 environmental effect (Moore et al. 1997, Drown and Wade 2014) where the environment is
468 the phenotypic value of the interspecific individual(s) with which an organism interacts. To
469 our knowledge, this type of environmental effect on between-species coevolution has been
470 considered in only two other theoretical studies (Scheiner et al. 2015, O'Brien et al. 2021; but
471 see Shuster et al. 2006, Witham et al. 2020 for a variance-partitioning approach). Scheiner et
472 al. (2015) consider a special case of our model, where IIGEs are present in only one of the
473 two interacting species. In this non-reciprocal model, they show a much more limited role
474 for IIGEs in coevolution. Our results are broadly consistent with this conclusion, in that
475 IIGEs in only a single species do not generate the reciprocal effects that lead to massive

476 inflation of evolutionary response. However, IIGEs in only a single species (e.g., $\Phi_{xy} = \mathbf{0}$,
477 $\Phi_{yx} \neq \mathbf{0}$) still play a role in mediating response to interspecific social selection whenever
478 interspecific social selection is a function of individual trait values (as opposed to the
479 population mean, as modeled by Scheiner et al. 2015). We also note that our fully
480 multivariate model accommodates the possibility that reciprocal IIGEs act across different
481 types or numbers of traits in the two interacting species. More recently, O'Brien et al. (2021;
482 see also Queller 2014) developed a model of coevolution between host plant and microbial
483 symbionts. Their parameterization differed from ours in that they consider evolution of a
484 single joint trait governed by genetic variation in host and symbiont, and so is most directly
485 applicable to plant-microbe systems or other intimate interactions. Nonetheless, their model
486 shows an important role for reciprocal fitness feedbacks, consistent with the conclusions of
487 our trait-based model.

488 Interspecific indirect genetic effects, or at least the potential for a prevalence of such
489 effects, appear to be commonplace in many biological systems. In Table 1, we provide in a
490 breakdown of types of biological interaction in which there is a large literature suggesting
491 importance of IIGE-like phenomena. These types of effects on trait expression across
492 species, widely appreciated in their own specific contexts (Weis and Abrahamson 1986,
493 Peacor and Werner 2001, Werner and Peacor 2003, Chen 2008, Thomas et al. 2012, O'Brien
494 et al. 2021), have taken on a variety of different forms. We suggest that these disparate
495 biological phenomena may nonetheless share a commonality—reciprocal effects on trait
496 expression across interspecific partners—that we have shown can affect the coevolutionary
497 process in dramatic, and in some cases predictable, ways.

498 Cross-species selection features prominently in verbal descriptions of the
499 coevolutionary process (Thompson 1982), and we show that such selection is especially
500 important in the presence of interspecific indirect genetic effects. When individual trait

501 values of one species affect individual fitness of another, focal species, this cross-species
502 selection can manifest evolutionary change in the focal species when there is phenotypic
503 assortment between interspecific interactants. This assortment, analogous to that required for
504 evolutionary response to social selection within species (Wolf et al. 1999, McGlothlin et al.
505 2010, Brodie et al. submitted), can be generated directly by a non-random genetic assortment,
506 or via IIGEs. Examples of processes that could generate direct genetic assortment between
507 interacting individuals of two different species include shared genetic structure. Such shared
508 genetic structure could arise through shared features of the environment that limit gene flow
509 and panmictic mating in both species, or alternatively, through variation in habitat preference
510 across individuals of both species. Direct genetic assortment could also arise through
511 behavioral preference for certain trait values in heterospecific partners. Such preferences
512 may be especially common in predator-prey interactions, where, for example, predator body
513 size may be expected to coevolve with behavioral preference for prey size (Troost et al.
514 2008).

515 Our results also indicate that nonlinear effects on cross-species selection can
516 contribute to coevolution even in the absence of genetic assortment or IIGEs. This form of
517 selection corresponds to an interaction between focal and interspecific-partner trait values for
518 focal individual fitness. The effect of this form of cross-species selection on evolutionary
519 response in a population depends on the mean genotype of the other species, and thus
520 represents a diffuse effect of population mean phenotype of the coevolving species. Such
521 interspecific interactions are potentially less intimate, for example diffuse predator chemical
522 cues in aquatic environments, than the individual level interactions (e.g., of host and parasite)
523 required to generate response from linear cross-species selection. Across populations,
524 nonlinear cross-species selection contributes to coevolution via covariance in mean genetic
525 values and/or linear selection between the species. This result is consistent with past models

526 of coevolution, verbal and mathematical, that indicate trait interactions for fitness are a key
527 feature of coevolution (Thompson 1982, 1994, 2005, Nuismer 2017), and in our model, such
528 interactions lead to a dependence between selection in one species and the mean trait value of
529 another. By defining these interaction terms in the framework of social evolution, our model
530 adds to past work by indicating that reciprocal IIGEs can substantially increase the degree to
531 which trait interactions for fitness contribute to reciprocal evolutionary change.

532 A unique feature of our model, compared to other models of interspecific coevolution,
533 is our development of a formal expression for the expected covariance in evolutionary
534 response, $\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y)$. This coevolutionary covariance is expected to be a key contributor
535 to generating among-population covariation in species mean trait values, a major focus in
536 many studies of coevolution (Thompson 1994, 1995, Zangerl and Berenbaum 2003, Toju and
537 Sota 2005, Hanifin et al. 2008, Hague et al. 2020). Importantly, similar to existing within-
538 species models of among-population quantitative genetic variation (Zeng 1988, Chenoweth et
539 al. 2010), defining this coevolutionary covariance illustrates how selection, IIGEs, and
540 genetics may contribute to patterns of trait variation across a geographic mosaic.

541 Our model subsumes mechanistic detail into broad statistical descriptions of species
542 interactions and thus provides a general description of how IIGEs and cross-species selection,
543 when present, contribute to reciprocal evolutionary change and correlated evolution across
544 populations. In contrast to our approach, some models of coevolution have focused instead
545 on specific ecological mechanisms that generate trait-fitness relationships between interacting
546 species (reviewed in Nuismer 2017). By highlighting the key parameters that contribute to
547 coevolution—covariance in natural selection, covariance in cross-species selection, and
548 IIGEs—our model indicates various pathways through which specific ecological mechanisms
549 may affect coevolution. Our framework can be tailored to specific scenarios by substituting
550 different fitness models into the general equations we present here.

551 Our model generates quantitative predictions for the shape of coevolution that are
552 directly testable with empirical data because it focuses on estimable statistical effects of
553 underlying ecological mechanisms rather than the mechanisms themselves, which are often
554 unknown (Wade and Kalisz 1990). For example, using an empirical estimate of Φ (which
555 could be measured using methods analogous to those used to measure within-species indirect
556 genetic effects; Bleakley and Brodie 2009, McGlothlin and Brodie 2009), one could use
557 matrix comparison of covariances among population means and the covariance terms
558 presented here to quantitatively test the contribution of IIGEs to among-population
559 covariance in selection response between two interacting species (e.g., see Chenoweth et al.
560 2010 for a within-species test of the predictions of Zeng's 1988 model). More generally,
561 Week and Nuismer (2019) have shown how datasets of among-population variation in trait
562 means can be used to test for conformation to expectations from coevolutionary models.
563 Concomitantly, our models show how environmental effects can be partitioned into terms
564 describing genotypes of other species in the ecological community, which could be useful in
565 understanding when and why evolutionary response fails to conform to predictions arising
566 from the standard breeder's equation.

567 Social interactions between individuals of the same species play a central role in the
568 evolutionary process. Within a single lineage, indirect genetic effects and social selection
569 fundamentally change selection response, the expression of genetic variance, and together
570 determine the course of social evolution (Moore et al. 1997, Wolf et al. 1998, Wolf et al.
571 1999, McGlothlin et al. 2010). We have shown that these effects of interactions among
572 individuals transcend species boundaries, and profoundly impact the dynamics of coevolution
573 between interacting lineages.

574 Literature Cited

- 575 Abrahamson, W. G., and A. E. Weis. 1997. Evolutionary Ecology Across Three Trophic
576 Levels: Goldenrods, Gallmakers, and Natural Enemies. Princeton University Press
577 Princeton.
- 578 Abrams, P. A., Y. Harada, and H. Matsuda. 1993. On the relationship between quantitative
579 genetic and ESS models. *Evolution* 47:982-985.
- 580 Agrawal, A. A. 2001. Phenotypic plasticity in the interactions and evolution of species.
581 *Science* 294:321-326.
- 582 Arnold, S. J., M. E. Pfrender, and A. G. Jones. 2001. The adaptive landscape as a conceptual
583 bridge between micro- and macroevolution. *Genetica* 112/113:9-32.
- 584 Arnqvist, G., and L. Rowe. 2002. Antagonistic coevolution between the sexes in a group of
585 insects. *Nature* 415:787-789.
- 586 Barribeau, S. M. 2014. Gene expression differences underlying genotype-by-genotype
587 specificity in a host-parasite system. *Proceedings of the National Academy of Sciences*
588 111:3496-3501.
- 589 Bijma, P., and M. J. Wade. 2008. The joint effects of kin, multilevel selection and indirect
590 genetic effects on response to genetic selection. *Journal of Evolutionary Biology*
591 21:1175-1188.
- 592 Bleakley, B. H., and E. D. Brodie III. 2009. Indirect genetic effects influence antipredator
593 behavior in guppies: Estimates of the coefficient of interaction ψ and the inheritance
594 of reciprocity. *Evolution* 63:1796-1806.
- 595 Boersma, M., P. Spaak, and L. De Meester. 1998. Predator-mediated plasticity in
596 morphology, life history, and behavior of *Daphnia*: the uncoupling of responses.
597 *American Naturalist* 152:237-248.
- 598 Borell, E. M., A. Foggo, and R. A. Coleman. 2004. Induced resistance in intertidal
599 macroalgae modifies feeding behavior of herbivorous snails. *Oecologia* 140:328-334.
- 600 Brodie, E. D., III, and E. D. Brodie, Jr. 1999. Predator-prey arms races. *BioScience* 49:557-
601 568.
- 602 Brodie, E. D. Jr., B. J. Ridenhour, and E. D. Brodie III. 2002. The evolutionary response of
603 predators to dangerous prey: hotspots and coldspots in the geographic mosaic of
604 coevolution between garters snakes and newts *Evolution* 56:2067-2082.
- 605 Brodie, E. D., III and B. J. Ridenhour 2003 Reciprocal selection at the phenotypic interface
606 of coevolution. *Integrative and Comparative Biology* 43:408-418
- 607 Brodie, E.D., III, P. A. Cook, R. A. Costello, V. A. Formica. Submitted. Phenotypic
608 assortment changes the landscape of selection. *Journal of Heredity*.
- 609 Chen, M.-S. 2008. Inducible direct plant defense against insect herbivores: A review. *Insect*
610 *Science* 15:101-114.
- 611 Chenoweth, S. F., H. D. Rundle, and M. W. Blows. 2010. The contribution of selection and
612 genetic constraints to phenotypic divergence. *American Naturalist* 175:186-196.
- 613 Damian, R. T. 1997. Parasite immune evasion and exploitation: reflections and projections.
614 *Parasitology* 115:S169-175.
- 615 Dawkins, R. 1982. *The Extended Phenotype*. Oxford Univ. Press, Oxford.
- 616 Drown, D. M., and M. J. Wade. 2014. Runaway coevolution: adaptation to heritable and
617 nonheritable environments. *Evolution* 68:3039-3046.
- 618 Dybdahl, M. F., C. E. Jenkins, and S. L. Nuismer. 2014. Identifying the molecular basis of
619 host-parasite coevolution: merging models and mechanisms. *American Naturalist*
620 184:1-13.
- 621 Ehrlich, P. R., and P. H. Raven. 1964. Butterflies and plants: a study in coevolution.
622 *Evolution* 18:586-608.

- 623 Foster, K. R., J. Schluter, K. Z. Coyte, and S. Rakoff-Nahoum. 2017. The evolution of the
624 host microbiome as an ecosystem on a leash. *Nature* 548:43-51.
- 625 Friesen, M. L., S. S. Porter, S. C. Stark, E. J. von Wettberg, J. L. Sachs, and E. Martinez-
626 Romero. 2011. Microbially mediated plant functional traits. *Annual Review of*
627 *Ecology and Systematics* 42:23-46.
- 628 Fuess, L., J. N. Weber, S. den Haan, N. C. Steinel, K. C. Shim, and D. I. Bolnick. 2020. A
629 test of the Baldwin effect: Differences in both constitutive expression and inducible
630 responses to parasites underlie variation in host response to a parasite. *bioRxiv*
631 <https://doi.org/10.1101/2020.07.29.216531>.
- 632 Gianinazzi-Pearson, V., N. Séjalon-Delmas, A. Genre, S. Jeandroz, and P. Bonfante. 2007.
633 Plants and arbuscular mycorrhizal fungi: cues and communication in the early steps of
634 symbiotic interactions. *Advances in Botanical Research* 46:181-219.
- 635 Hague, M. T. J., A. N. Stokes, C. R. Feldman, E. D. Brodie Jr., and E. D. Brodie III. 2020.
636 The geographic mosaic of arms race coevolution is closely matched to prey
637 population structure. *Evolution Letters* 164:1567–16.
- 638 Hanifin, C. T., E. D. Brodie Jr., and E. D. Brodie III. 2008. Phenotypic mismatches reveal
639 escape from arms-race coevolution. *PLoS Biology* 6:e60.
- 640 Janzen, D. H. 1980. When is it coevolution? *Evolution* 34:611-612.
- 641 Johnson, S. D., and B. Anderson. 2010. Coevolution between food-rewarding flowers and
642 their pollinators. *Evolution: Education and Outreach* 3:32-39.
- 643 Kirkpatrick, M. and R. Lande 1989. The evolution of maternal characters. *Evolution* 43:485-
644 503.
- 645 Korgaonkar, A., C. Han, A. L. Lemire, I. Siwanowicz, D. Bennouna, R. E. Kopec, P.
646 Andolfatto, S. Shigenobu, and D.L. Stern. 2021. A novel family of secreted insect
647 proteins linked to plant gall development. *Current Biology* 31:1-14
- 648 Lande, R. 1979. Quantitative genetic analysis of multivariate evolution, applied to brain:
649 body size allometry. *Evolution* 33:402-416.
- 650 Lande, R., and S. J. Arnold. 1983. The measurement of selection on correlated characters.
651 *Evolution* 37:1210-1226.
- 652 McAdam, A.G., S. Boutin, D. Réale, and D. Berteaux. 2002. Maternal effects and the
653 potential for evolution in a natural population of animals. *Evolution* 56: 846-851.
- 654 McGlothlin, J. W., and E. D. Brodie III. 2009. How to measure indirect genetic effects: the
655 congruence of trait-based and variance-partitioning approaches. *Evolution* 63:1785-
656 1795.
- 657 McGlothlin, J. W., E. Akçay, E. D. Brodie III, A. J. Moore, and J. Van Cleve. 2021. A
658 synthesis of game theory and quantitative genetic models of social evolution. *bioRxiv*
659 <https://doi.org/10.1101/2021.03.27.437341>
- 660 McGlothlin, J. W., A. J. Moore, J. B. Wolf, and E. D. Brodie III. 2010. Interacting
661 phenotypes and the evolutionary process. III. Social evolution. *Evolution* 64:2558-
662 2574.
- 663 Moore, A. J., E. D. Brodie III, and J. B. Wolf. 1997. Interacting phenotypes and the
664 evolutionary process: I. Direct and indirect genetic effects of social interactions.
665 *Evolution* 51:1352-1362.
- 666 Mousseau, T.A. and C.W. Fox. 1998. The adaptive significance of maternal effects. *Trends in*
667 *Ecology and Evolution* 13:403-407
- 668 Nuismer, S. L. 2017. *Introduction to Coevolutionary Theory*. W.H. Freeman and Company
669 New York
- 670 Nuismer, S. L., B. J. Ridenhour, and B. P. Oswald. 2007. Antagonistic coevolution mediated
671 by phenotypic differences between quantitative traits. *Evolution* 61:1823-1834.

- 672 Ohgushi, T. 2005. Indirect interaction webs: Herbivore-induced effects through trait change
673 in plants Annual Review of Ecology, Evolution, and Systematics 36:81-105.
- 674 Oladiran, A., and M. Belosevic. 2012. Immune evasion strategies of trypanosomes: a review.
675 Journal of Parasitology 98:284-292.
- 676 O'Brien, A. M., C. N. Jack, M. L. Friesen, and M. E. Frederickson. 2021. Whose trait is it
677 anyways? Coevolution of joint phenotypes and genetic architecture in mutualisms.
678 Proceedings of the Royal Society of London Series B. 288:20202483.
- 679 Peacor, S. D., and E. E. Werner. 2001. The contribution of trait-mediated indirect effects to
680 the net effects of a predator. Proceedings of the National Academy of Sciences
681 98:3904-3908.
- 682 Pellmyr, O. 2003. Yuccas, yucca moths, and coevolution: A review. Annals of the Missouri
683 Botanical Garden 90:35-55.
- 684 Perry, J., and L. Rowe. 2011. Sexual conflict and antagonistic coevolution across water
685 strider populations. Evolution 66:544-557.
- 686 Preisser, E. L., D. I. Bolnick, and M. F. Benard. 2005. Scared to death? The effects of
687 intimidation and consumption in predator-prey interactions. Ecology 86:501-509.
- 688 Price, G. R. 1970. Selection and covariance. Nature 227:520-521.
- 689 Price, G. R. 1972. Extension of covariance mathematics. Annals of Human Genetics 35:485-
690 490.
- 691 Queller, D. C. 2014. Joint phenotypes, evolutionary conflict and the fundamental theorem of
692 natural selection. Philosophical Transactions of the Royal Society B. 369:20130423.
- 693 Ridenhour, B. J. 2005. Identification of selective sources: Partitioning selection based on
694 interactions. The American Naturalist 166:12-25.
- 695 Robertson, A. 1966. A mathematical model of the culling process in dairy cattle. Animal
696 Production 8:95-108.
- 697 Scharsack, J. P., A. Gossens, F. Franke, and J. Kurtz. 2013. Excretory products of the
698 cestode, *Schistocephalus solidus*, modulate in vitro responses of leukocytes from its
699 specific host, the three-spined stickleback (*Gasterosteus aculeatus*). Fish and
700 Shellfish Immunology 35:1779-1787.
- 701 Scharsack, J. P., M. Kalbe, R. Derner, J. Kurtz, and M. Milinski. 2004. Modulation of
702 granulocyte responses in three-spined sticklebacks *Gasterosteus aculeatus* infected with
703 the tapeworm *Schistocephalus solidus*. Diseases of Aquatic Organisms 59:141-150.
- 704 Scharsack, J. P., K. Koch, and K. Hammerschmidt. 2007. Who is in control of the stickleback
705 immune system: interactions between *Schistocephalus solidus* and its specific
706 vertebrate host. Proceedings of the Royal Society of London Series B. 274:3151-
707 3158.
- 708 Scheiner, S. M., R. Gomulkiewicz, and R. D. Holt. 2015. The genetics of phenotypic
709 plasticity. XIV. Coevolution. American Naturalist 185:594-609.
- 710 Schmid-Hempel, P. 2008. Parasite immune evasion: a momentous molecular war. Trends in
711 Ecology and Evolution 23:318-326.
- 712 Shuster, S. M., E. V. Lonsdorf, G. M. Wimp, J. K. Bailey, and T. G. Whitham. 2006.
713 Community heritability measures the evolutionary consequences of indirect genetic
714 effects on community structure. Evolution 60:991-1003.
- 715 Siepielski, A. M., K. M. Gotanda, M. B. Morrissey, S. E. Diamond, J. D. DiBattista, and S.
716 M. Carlson. 2013. The spatial patterns of directional phenotypic selection. Ecology
717 Letters 16:1382-1392.
- 718 Stappenbeck, T. S., and H. W. Virgin. 2016. Accounting for reciprocal host-microbiome
719 interactions in experimental science. Nature 534:191-199.

- 720 Stearns, S. C. 2012. Evolutionary routes leading to host manipulation by parasites: Afterward
721 *in* F. Thomas, J. Brodeur, and D. P. Hughes, editors. Host manipulation by parasites.
722 Oxford University Press, Oxford.
- 723 Tams, V., J. H. Nickel, A. Ehring, and M. Cordellier. 2019. Insights into the genetic basis of
724 predator-induced response in *Daphnia* - a comparative transcriptomic approach.
725 bioRxiv. <https://doi.org/10.1101/503904>
- 726 Thomas, F., J. Brodeur, and D. P. Hughes. 2012. Host manipulation by parasites Oxford
727 University Press, Oxford.
- 728 Thompson, J. N. 1982. Interaction and Coevolution. University of Chicago Press, Chicago.
- 729 Thompson, J. N. 1994. The Coevolutionary Process. University of Chicago Press, Chicago.
- 730 Thompson, J. N. 2005. The Geographic Mosaic of Coevolution. University of Chicago Press,
731 Chicago.
- 732 Toju, H., and T. Sota. 2005. Imbalance of predator and prey armament: geographic clines in
733 phenotypic interface and natural selection. *American Naturalist* 167:105–117
- 734 Troost, T. A., B. W. Kooi, and U. Dieckmann. 2008. Joint evolution of predator body size
735 and prey size preference. *Evolutionary Ecology* 22:771-799.
- 736 Urban, M.C. 2011. The evolution of species interactions across natural landscapes. *Ecology*
737 *Letters* 14:723-732.
- 738 Wade, M. J., and Kalisz. 1990. The causes of natural selection. *Evolution* 44:1947-1955.
- 739 Walsh, M. R., and D. M. Post. 2011. The impact of intraspecific variation in a fish predator on
740 the evolution of phenotypic plasticity and investment in sex in *Daphnia ambigua*.
741 *Journal of Evolutionary Biology* 25:80-89.
- 742 Weber, A., and S. Declerck. 1997. Phenotypic plasticity of *Daphnia* life history traits in
743 response to predator kairomones: genetic variability and evolutionary potential
744 *Hydrobiologia* 360:89-99.
- 745 Weber, J. N., N. C. Steinel, K. C. Shim, and D. I. Bolnick. 2017. Recent evolution of extreme
746 cestode growth suppression by a vertebrate host. *Proceedings of the National*
747 *Academy of Sciences* 114:6575-6580.
- 748 Week, B. and S.L. Nuismer. 2019. The measurement of coevolution in the wild. *Ecology*
749 *Letters* 22:717-725
- 750 Weis, A. E., and W. G. Abrahamson. 1986. Evolution of host-plant manipulations by gall
751 makers: ecological and genetic factors in the Solidago-Eurosta system. *American*
752 *Naturalist* 127:681-695.
- 753 Weis, A.E., W.G. Abrahamson, and M.C. Anderson. 1992. Variable selection on Eurosta's
754 gall size, I: The extent and nature of variation in phenotypic selection. *Evolution*
755 46:1674-1697.
- 756 Werner, E. E., and S. D. Peacor. 2003. A review of trait-mediated indirect interactions in
757 ecological communities. *Ecology* 84:1083-1100.
- 758 West-Eberhard, M. J. 1979. Sexual selection, social competition, and evolution. *Proceedings*
759 *of the American Philosophical Society* 123:222-234.
- 760 West-Eberhard, M. J. 1983. Sexual selection, social competition, and speciation. *Quarterly*
761 *Review of Biology* 58:155-183.
- 762 West-Eberhard, M. J. 1984. Sexual selection, competitive communication and species-
763 specific signals in insects. Pages 283-342 *in* T. Lewis, editor. *Insect communication*.
764 Academic Press, New York.
- 765 Whitham, T. H., G. J. Allan, H. F. Cooper, and S. M. Shuster. 2020. Intraspecific genetic
766 variation and species interactions contribute to community evolution. *Annual Review*
767 *of Ecology and Systematics* 51:587-612.

- 768 Wolf, J. B., E. D. Brodie III, J. M. Cheverud, A. J. Moore, and M. J. Wade. 1998.
769 Evolutionary consequences of indirect genetic effects. *Trends in Ecology and*
770 *Evolution* 13:64-69.
- 771 Wolf, J. B., E. D. Brodie III, and A. J. Moore. 1999. Interacting phenotypes and the
772 evolutionary process. II. Selection resulting from social interactions. *American*
773 *Naturalist* 153:254-266.
- 774 Zangerl, A. R., and M. R. Berenbaum. 2003. Phenotype matching in wild parsnip and parsnip
775 webworms: causes and consequences. *Evolution* 57:806–815
- 776 Zeng, Z.-B. 1988. Long-term correlated response, interpopulation covariation, and
777 interspecific allometry. *Evolution* 42 363-374.
- 778 Züst, T., S. Mou, and A.A. Agrawal. 2018. What doesn't kill you makes you stronger: The
779 burdens and benefits of toxin sequestration in a milkweed aphid. *Functional Ecology*
780 8:1972-1981

Supplemental Figures

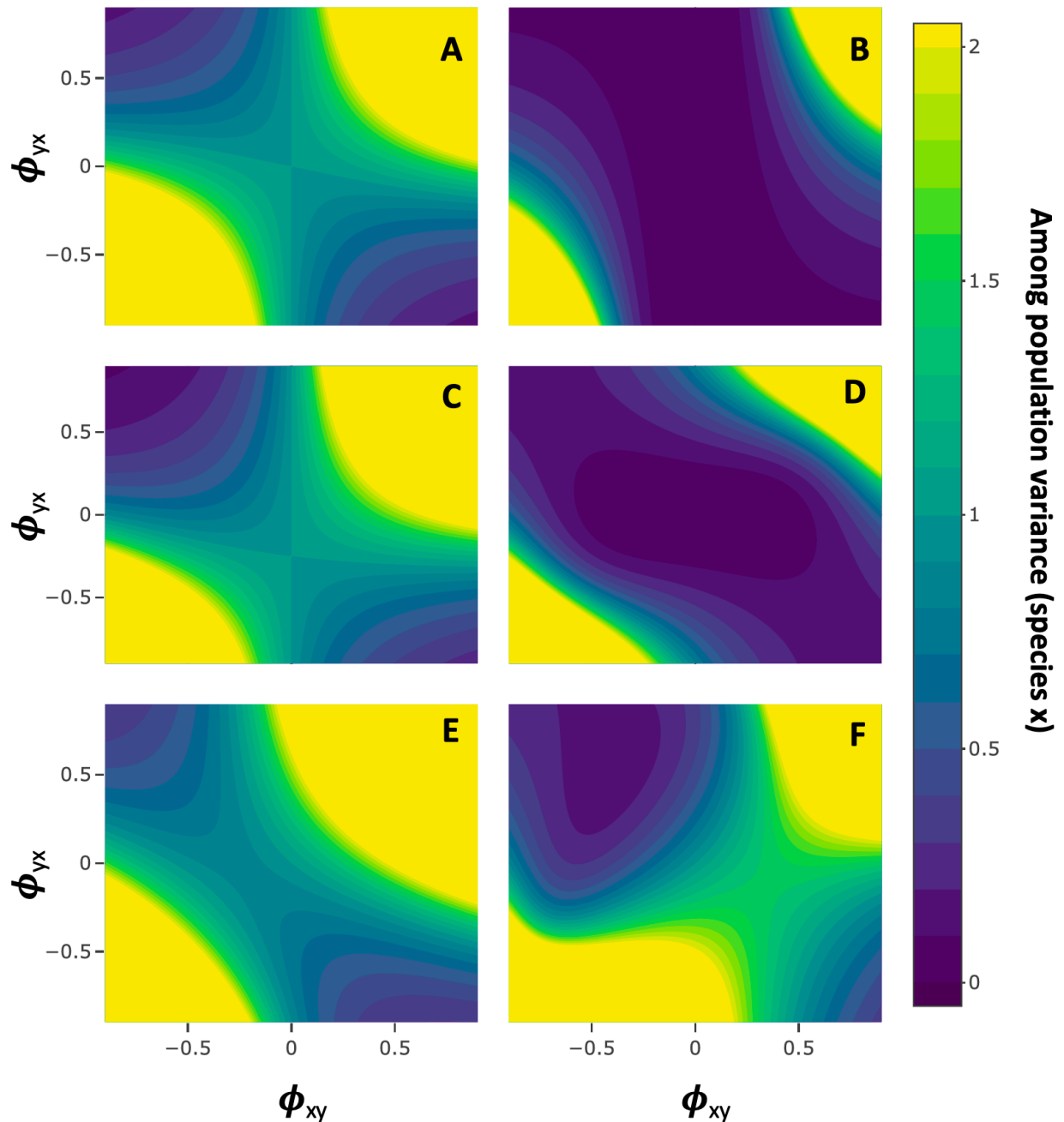


Figure S1. IIGEs accelerate evolutionary rate in a single species. Panels show the evolutionary rate, $\text{Var}(\Delta\bar{z}_x)$, of species x as a function of the indirect genetic effect parameters ϕ , under the same parameter values as in Figure 3. Reciprocal IIGEs between interacting species generally accelerate evolutionary rate. Note that in the absence of any other effects, the evolutionary rate is equal to the variance in natural selection, which is unity in Panels A, C, E, and F. In panel D, evolutionary rate is driven entirely by cross-species selection and IIGEs. In panel B, where $G_{xx}=0$, evolutionary rate in species x is driven entirely by reciprocal IIGEs and evolutionary change in species y .

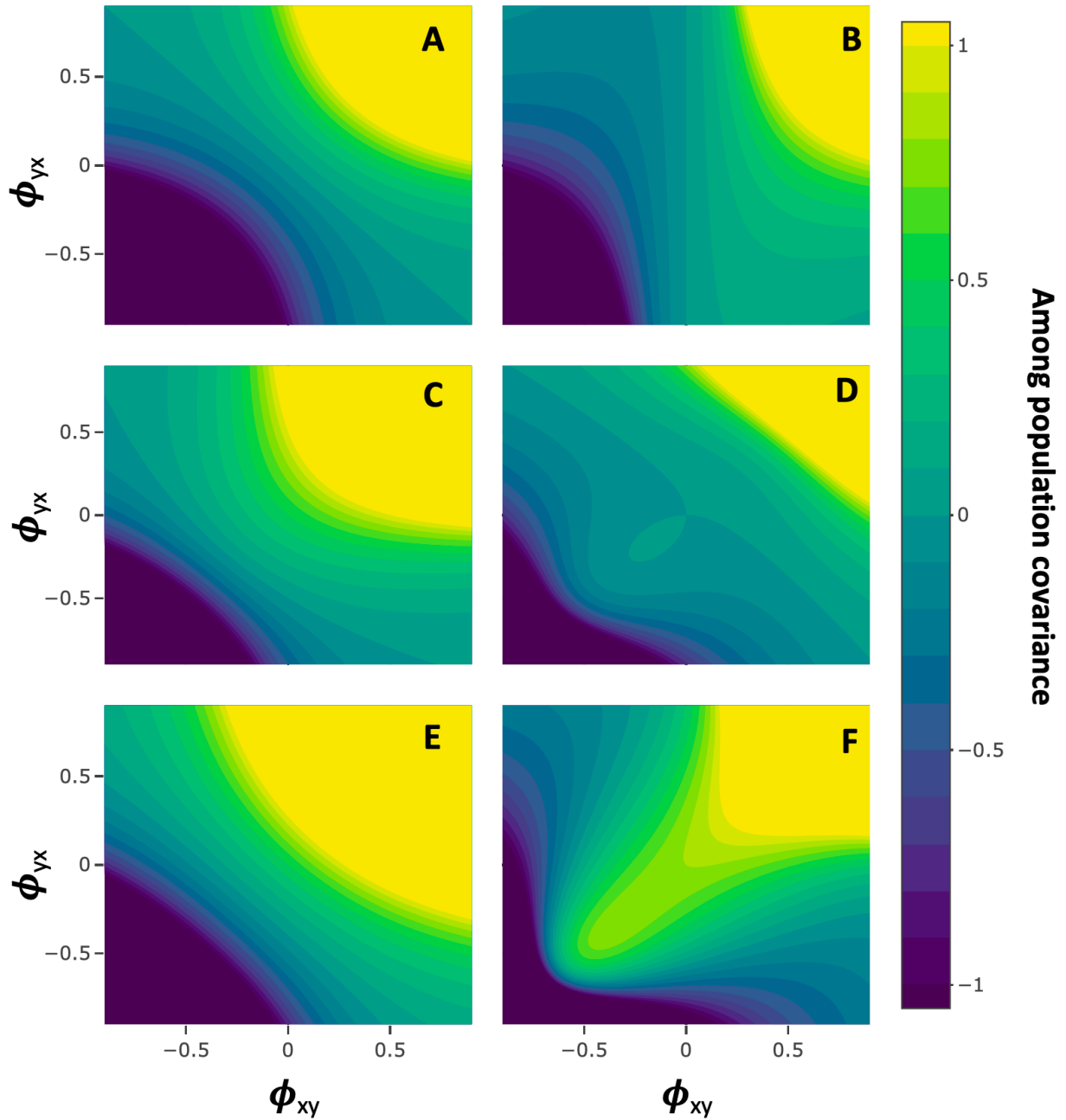


Figure S2. Among population covariances. Panels show the coevolutionary covariance between two interacting species as a function of the indirect genetic effect parameters ϕ , from equation (7) unstandardized. Parameter values are as in Figs. 4 and S1. For all panels, genetic variances were set to unity.

781 **Appendix**

782 **A1. Phenotypic covariance between interacting species**

783 For a single trait, we can solve for the covariance between z_y and z_x to partition the
 784 phenotypic covariance between individuals of two interacting species into terms describing
 785 the contribution of IIGEs and terms describing non-random genetic assortment. Assuming
 786 cross-species environmental covariance is zero,

787

$$C_{xy} = (1 - \phi_{yx}\phi_{xy})^{-2}[(1 + \phi_{yx}\phi_{xy})G_{xy} + \phi_{yx}(G_x + E_x) + \phi_{xy}(G_{yy} + E_x)]$$

788

789 where E_x and E_y represent within-species environmental variance and $G_{xy} = G_{yx}$. When
 790 IIGEs are absent, non-random genetic assortment G_{xy} is the sole contributor to the
 791 phenotypic association between individuals of coevolving species. When IIGEs are present,
 792 they can substantially change this phenotypic association.

793

794 **A2. Covariance in selection response with nonzero IIGEs and genetic assortment**

795 We expand the covariance in evolutionary response in two species when IIGEs are present
 796 and constant and when genetic assortment $G_{xy} = G_{yx}$ is present and constant,

797

$$\begin{aligned} \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = & \mathcal{U}[\mathcal{A}^2\phi_{yx}\text{Var}(\beta_{xx}) + \mathcal{B}^2\phi_{yx}\text{Var}(\beta_{xy}) + \mathcal{C}^2\phi_{xy}\text{Var}(\beta_{yy}) \\ & + \mathcal{D}^2\phi_{xy}\text{Var}(\beta_{yx}) + \mathcal{A}\mathcal{B}\mathcal{E}\text{Cov}(\beta_{xx}, \beta_{xy}) + \mathcal{A}\mathcal{C}\mathcal{E}\text{Cov}(\beta_{xx}, \beta_{yy}) \\ & + \mathcal{A}\mathcal{D}\mathcal{E}\text{Cov}(\beta_{xx}, \beta_{yx}) + \mathcal{B}\mathcal{C}\mathcal{E}\text{Cov}(\beta_{xy}, \beta_{yy}) + \mathcal{B}\mathcal{D}\mathcal{E}\text{Cov}(\beta_{xy}, \beta_{yx}) \\ & + \mathcal{C}\mathcal{D}\mathcal{E}\text{Cov}(\beta_{yy}, \beta_{yx})] \end{aligned}$$

802

803 where

804

805
$$\mathcal{U} = (1 - \phi_{xy}\phi_{yx})^{-4}$$

806
$$\mathcal{A} = G_{xx} + G_{xy}\phi_{xy}$$

807
$$\mathcal{B} = G_{xy} + G_{xx}\phi_{yx}$$

808
$$\mathcal{C} = G_{yy} + G_{xy}\phi_{yx}$$

809
$$\mathcal{D} = G_{xy} + G_{yy}\phi_{xy}$$

810
$$\mathcal{E} = 1 + \phi_{xy}\phi_{xy}$$