1 Unique protein features of SARS-CoV-2 relative to other Sarbecoviruses

3

4 Matthew Cotten^{1,2}, David L. Robertson², My V.T. Phan¹

5

6 Affiliations

- 7 1. MRC/UVRI & LSHTM Uganda Research Unit, Entebbe, Uganda
- 8 2. MRC-University of Glasgow Centre for Virus Research, Glasgow, UK
- 9 Correspondence: Matthew Cotten (Matthew.Cotten@lshtm.ac.uk)
- 10 Keywords: SARS-CoV-2, proteome changes, Sarbecovirus evolution, spike protein changes
- 11

12 Abstract

13 Defining the unique protein features of SARS-CoV-2, the viral agent causing Coronavirus 14 Disease 2019, may guide efforts to control this pathogen. We examined proteins encoded by the 15 Sarbecoviruses closest to SARS-CoV-2 using profile Hidden Markov Model similarities to identify 16 features unique to SARS-CoV-2. Consistent with previous reports, a small set of bat and pangolin-17 derived Sarbecoviruses show the greatest similarity to SARS-CoV-2. The analysis provided a measure 18 of total proteome similarity and showed that a small subset of bat Sarbecoviruses are closely related 19 but unlikely to be the direct source of SARS-CoV-2. Spike analysis reveals that the current SARS-CoV-20 2 variants of concern have sampled only 36% of the possible spikes changes which have occurred 21 historically in Sarbecovirus evolution. It is likely that new SARS-CoV-2 variants with changes in these 22 regions are compatible with virus replication and are to be expected in the coming months, unless global 23 viral replication is severely reduced.

24

25 Introduction

26 Since the first report of Coronavirus Disease 2019 (COVID-19) caused by SARS-CoV-2 in 27 December 2019 in Wuhan city, China(1)(2) and the World Health Organisation declaring COVID-19 a 28 global pandemic in March 2020, the disease has continued to affect every part of the world. The SARS-29 CoV-2 virus belongs to the Coronaviridae family of enveloped positive-sense single-stranded RNA 30 viruses, Betacoronavirus genus, Sarbecovirus subgenus. Other Sarbecoviruses include SARS-CoV 31 (the coronavirus causing the SARS outbreak in 2002-2004) and a large number of SARS-like bat 32 viruses that have been identified. The genomes of Sarbecoviruses are 30kb in length, encoding >14 33 open reading frames (ORFs). Among the structural proteins, the spike protein plays a crucial role in the 34 virus cell tropism, host range, cell entrance and infectivity and is considered the main protein target for 35 the host immune response. Other ORFs encode for accessory proteins, many of which modulate host 36 responses to infection. Investigation of the evolutionary history of SARS-CoV-2 show a clear link to 37 Sarbecoviruses from bats although no direct animal precursor for SARS-CoV-2 has been identified (3) 38 (4) (5) (6) (7). We sought to identify unique peptide regions of SARS-CoV-2 compared to all available 39 Sarbecoviruses to determine the features that might have allowed SARS-CoV-2 to replicate and

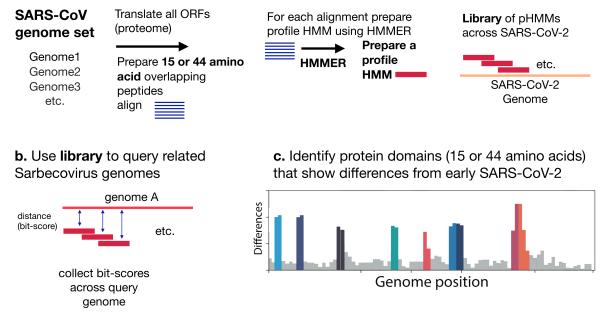
transmit efficiently in humans. Comparative analyses of viral proteins would aid in better understanding
of virus biology and pathology, providing insights into the origin of the virus and the conditions that led
to its zoonosis to humans, efficient spread without the need for adaptation, as well as providing leads
for drug and immune targets for effective treatments.

44

45 Results and Discussion

46 Protein domains and profile hidden Markov models. We have explored the genomes across 47 the Sarbecovirus subgenus using profile hidden Markov models (pHMMs) based on viral proteins 48 which provide a statistical description of the properties of amino acid sequences of viral proteins. The 49 differences in pHMMs between virus genomes can reveal differences among viral proteins (8) (9). 50 Efficient tools for preparing and comparing pHMMs are available with HMMER-3 (10) and the method 51 is useful for comparing large or difficult to align genomes and for identifying changes in functional 52 regions of the genomes. We have recently used these methods to identify and classify diverse 53 coronaviruses in the Coronaviridae family (11) and to explore large and unwieldy genomes such as 54 those from the African Swine Fever Virus (12). These methods were employed here to explore the 55 relationship between SARS-CoV-2 and the other known Sarbecoviruses to gain understanding of their 56 evolutionary history.

a. Prepare Agnostic Domains from a small set of early SARS-CoV-2 genomes



57 58 Figure 1. Analysis scheme. (A) Profile Hidden Markov Model (pHMM) domains were generated from 59 a set of 35 early lineage B SARS-CoV-2 genome sequences. All open reading frames were translated 60 and then sliced into either 44 amino acid peptides with a step size of 22 amino acids or 15 amino acid peptides with a step size of 8 amino acid. The peptides were clustered using Uclust (13), aligned with 61 62 MAFFT (14) and then each alignment was built into a pHMM using HMMER-3 (10). (B) The set of 63 pHMMs were used to query Sarbecovirus genome sequences, bit scores were collected as a 64 measure of similarity between each pHMM and the guery sequence. (C) Bit-scores were gathered an 65 analyzed to detect regions that differ between early SARS-CoV-2 genomes and guery genomes. 66

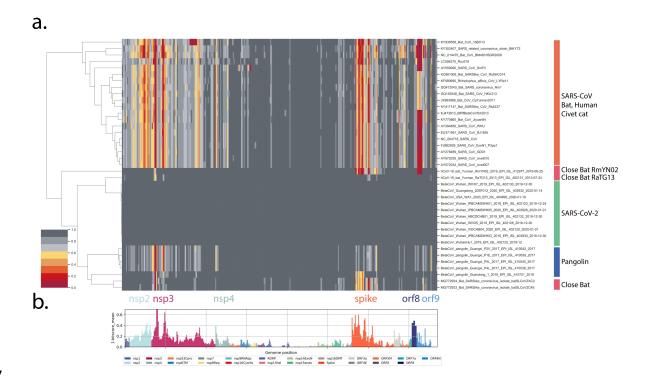
67 **Genome scans using custom pHMM domains.** We generated overlapping 44 or 15 amino 68 acid (aa) alignments of all SARS-CoV-2 encoded proteins derived from a set of 35 lineage B SARS-

69 CoV-2 genomes from early in the pandemic and then prepared pHMMs from each alignment (see Figure
70 1). The resulting libraries of pHMMs were then used to survey variations across all *Sarbecoviruses*71 relative to the initial SARS-CoV-2 found in human cases in late 2019/early 2020.

72 Using this method, ten early SARS-CoV-2 genomes were compared to a representative subset 73 of 29 Sarbecovirus genomes obtained from humans, bats, civet cats and pangolins (Figure 2) selected 74 after applying the same analysis to all available Betacoronavirus genomes to ensure that we were not 75 missing any unexpectedly close viral genome regions (results not shown). For each guery genome, the 76 bit-score from the corresponding pHMM from early B lineage SARS-CoV-2 were accumulated and 77 hierarchical clustering based on the normalized domain bits-scores was performed. The bit-score 78 describes the similarity between the 44aa (or 15aa) query sequence and the related sequence from 79 early SARS-CoV-2. The central region of the Sarbecovirus genome is conserved across the genome 80 set with all domains marked as dark or light grey in the Figure 2A clustermap indicating normalized bit-81 scores close to 1. This is not unexpected as this central genomic region spans nsp5 to nsp16 proteins 82 encoding the viral polymerase, other enzymes and non-surface exposed structural proteins of the virus, 83 which are presumably more constrained and less likely to change than other regions of the virus. In 84 contrast, the domains displayed in yellow, orange and red in Figure 2A indicate more increasingly 85 divergent regions between SARS-CoV-2 and the query Sarbecovirus genomes (much lower normalized 86 bit scores). For each domain across the proteome, a measure of the difference between the guery and 87 SARS-CoV-2 domain (1-mean bit-score) was plotted to identify regions that differed extensively across 88 the Sarbecoviruses genome set (Figure 2B). Further details on the relationship between amino acid 89 changes across a genome or gene set and the metric 1-mean bit-score are found in the Supplemental 90 Figure 1. The higher 1-mean bit-score values (≥ 0.3) were found in nsp2, nsp3, nsp4, spike and orf8 91 and orf9N (Figure 2B), indicating that these proteins differ in the set of SARS-CoV-2 and closely related 92 Sarbecoviruses. These regions may be dispensable, flexible and can tolerate change and may be 93 products of neutral evolution they may differ because selective pressure has resulted in new functional 94 roles or immune escape.

95

96





98 Figure 2. Proteome differences in SARS-CoV-2 vs close Bat, Human and Civet cat 99 Sarbecoviruses. All forward open reading frames from the 35 early lineage B SARS-CoV-2 genomes 100 were translated, and processed into 44 aa peptides (with 22 aa overlap), clustered at 0.65 identity using 101 Uclust (11), aligned with MAAFT (12) and converted into pHMMs using HMMER-3 (10). The presence 102 of these domains was sought in a set of Sarbecovirus genomes plus the SARS-CoV-2 genomes and 103 genomes were then clustered using hierarchical clustering based on the normalized domain bit-scores 104 (e.g. the similarity of the identified query domain to the reference lineage B SARS-CoV-2 domain). Each 105 row represents a genome, each column represents a domain. Domains are displayed in their order 106 across the SARS-CoV-2 genome, Red = low normalized domain bit-score (lower similarity to lineage B 107 SARS-CoV-2) = distant from SARS-CoV-2, Darkest grey = normalized domain bit-score = 1 = highly 108 similar to lineage B SARS-CoV-2. Groups of coronaviruses were indicated to the right of the figure. (A) 109 Domain differences across the Sarbecovirus subgenus. (B) For each domain the mean bit-score was 110 calculated across the entire set of Sarbecovirus genomes and the value 1-mean bit-score was plotted 111 for each domain. Domains are coloured by the proteins from which they were derived with the colour 112 code indicated below the figure.

113 114

115 The role of pangolins as an amplifying intermediate host of SARS-CoV-2 is important to 116 document securely, to guide efforts to prevent or prepare for future zoonotic events. A small number of 117 Sarbecoviruses have been identified in samples from trafficked pangolins in China (15) (7) (16), yet 118 there is no direct evidence that pangolins host the virus in their natural environment. It remains possible 119 that the positive pangolins identified in China were infected by viruses encountered after transport to 120 China. Five CoV sequences from pangolins were included in this analysis (Figure 2), including four 121 (EPI ISL 410538,-39,-40,-42) generated by Lam et al. (7) after sequencing the original samples described by Liu et al.(15); a 5th genome (Guandong 1 2019 EPI ISL 410721 2019) deposited by 122 123 Xiao et al. (16) was included despite unclear progeny for this sequence (17). Four additional CoV 124 sequences from pangolins from GISAID were excluded due to excessive gaps in the sequence which 125 precluded the domain analysis. Overall, the pangolin CoVs show many genomic regions of very close

domain homology although the overall similarity across the entire genome is not close enough to support of direct pangolin-human jump as the source of SARS-CoV-2 (see below).

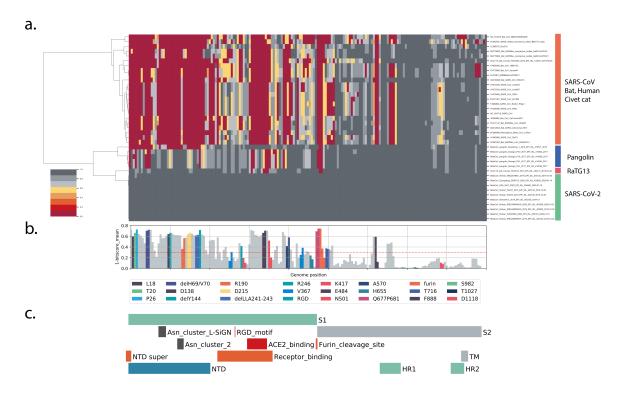
128 A number of closely related Sarbecoviruses have been reported in bats (18) (19). We focused 129 on the four bat Sarbecoviruses with closest similarity to SARS-CoV-2 (close bats: RaTG13, RmYn02, 130 MG772933,34). The clustermap and variance analysis (Figure 2A) showed higher similarity across most 131 of the genome (dark grey sectors) with three proteins (nsp3, spike and orf9) displaying high reduced 132 bit-scores from the SARS-CoV-2 sequences (Figure 2A, close bats, vellow/red domains). These limited 133 regions that differ between the 4 closely related bat Sarbecoviruses and SARS-CoV-2 indicate virus 134 elements that needed to adjust to human transmission. The spike differences have been explored in 135 detail (refs and see below) however it may be important to consider nsp3 and orf9 changes in the 136 complete analysis.

137 Spike changes with 15 amino acid domains. To provide more detailed resolution, 15 amino 138 acid pHMMs across the early lineage B spike protein were prepared and used to examine changes 139 across the set of close Sarbecoviruses and SARS-CoV-2 (Figure 3). The regions of change observed 140 in the spike protein show an expanded version of the total proteome pattern observed in Figure 2 plus 141 additional details. Adding the current knowledge of SARS-CoV-2 spike evolution reveals some 142 important patterns. The S1 domain of spike (the amino-terminal half of the protein) tolerates at large 143 amount of change with most of the low score domains (red) concentrated here (Figure 3A), consistent 144 with the surface exposure of the S1 region, and driven by pressure to avoid immune responses which 145 in turn require the virus to maintain a malleable structure in the exposed S1. Except for the 146 Guangdong 1 pangolin and RatG13 spikes, the central ACE2 receptor binding region is very different 147 between the close Sarbecoviruses and SARS-CoV-2. The furin cleavage site at the junction between 148 the S1 and S2 domains is also a region showing a lot of malleability in the Sarbecovirus spikes (Figure 149 3A) and is completely unique to SARS-CoV-2. This has been discussed in detail (20) and is also a site 150 of frequent change in the current SARS-CoV-2 Variant of Concern (VOC) spike sequence with Q677, 151 P681 and T717 flanking the furin site showing changes (Figure 3B).

Thirdly, the amino acid changes that have accumulated in the VOC spike protein are marked in color (Figure 3B) and cluster in regions with high variation (1-mean bit-score > 0.3) suggesting that *Sarbecoviruses* have made changes in these regions in previous evolutionary periods and are continuing to change in SARS-CoV-2 evolution. If all domains with 1-mean bit-score of 0.3 and above tolerate change in the current SARS-CoV-2 spike protein, this suggests that SARS-CoV-2 has many additional changes available for future immune evasion. Important regions that show high levels of historical change are RBD, the furin cleavage site and flanking regions and the NTD.

5

159

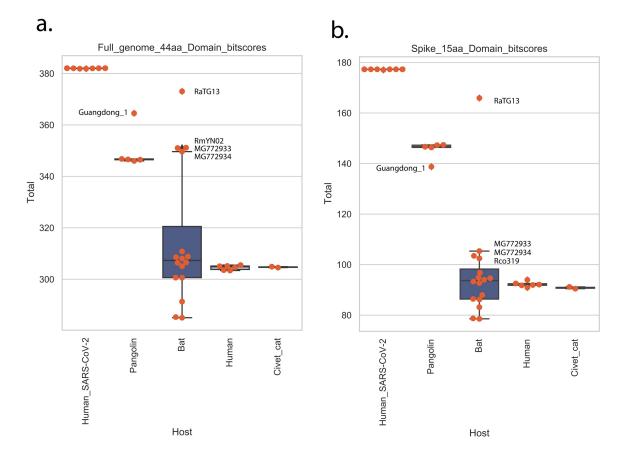


160

161 Figure 3. Spike differences in SARS-CoV-2 vs close Bat, Human and Civet cat Sarbecoviruses. 162 All forward spike open reading frames from the 35 early lineage B SARS-CoV-2 genomes were 163 translated, and processed into 15 aa peptides (with 8 aa overlap) and processed into an pHMM library 164 as described in Figure 2. (A) shows a hierarchical clustering of 15 amino acid domain bit-scores. (B) 165 shows the 1-mean of each domain bit-scores across the genome set, domain values, individual 166 domains that span known amino acid changes in the 5 VOC are colored (see key below panel B). (C) The locations of important spike protein features are indicated. NTD: N-terminal domain, RBD: receptor-167 168 binding domain, S1: spike 1, S1: Spike 2, TM: transmembrane domain, HR1: helical repeat 1, HR2: 169 helical repeat 2, NTD super: N-terminal domain supersite. 170

171 Global proteome similarities. A measure of the total protein distance between the SARS-172 CoV-2 and any query Sarbecovirus can be obtained by summing the normalized bit-scores (SNBS) 173 across the entire query proteome. The encoded proteomes from more distant viruses will show lower 174 SNBS values. We examined SNBSs grouped by virus host for both the 44 amino acid total genome 175 analysis as well as the 15 amino acid spike gene analysis. Overall, the patterns showed that the 176 similarity to SARS-CoV-2 follows the order Pangolin > Bat > human=civet cat (Figure 4) although the 177 small set of close bat Sarbecoviruses show some exceptions. Of special interest, the bat coronavirus 178 genome RaTG13 (GenBank MN996532.1) was identified as closely related to the SARS-CoV-2 lineage 179 {Citation} and supports a bat coronavirus being the zoonotic source of the epidemic, although the 180 genetic distance is too far for RaTG13 itself to be a direct source of the pandemic SARS-CoV-2 virus 181 (21)(3). Another close bat coronavirus RmYN02, shows some regions of close identity to SARS-CoV-2 (6), yet overall it is more distant from SARS-CoV-2 than the strain RaTG13 (Figure 4A) due its possible 182 183 recombinant nature. A single pangolin derived SARS-CoV-2 (Guangdong_1) showed an SNBS value 184 that was also elevated but not as high as the RaTG13 (Figure 4a), the 15 aa spike analysis showed

- similar patterns except that only the RaTG13 spike displayed the high similarity to SARS-CoV-2 (Figure
- 186 4b).
- 187



- 188
- 189

Figure 4. Total domain distances between virus groups. Normalized bit-score sums (NBSS) grouped into SARS-CoV-2 and *Sarbecoviruses* from pangolin, bat, human and civet cat NBSS for all domains for each genome were summed. The boxplot shows individual values marked in orange, median values indicated by horizontal black lines, 1st interquartile ranges marked with a box. The identities of several high scoring bat and pangolin genomes are indicated. (A) NBSS for 44 aa domains across the entire coronavirus genome. (B) NBSS for 15 aa domains across the spike protein.

197 Conclusions

198 What is special about SARS-CoV-2? Spike changes in SARS-CoV-2 compared to a large set 199 of known Sarbecovirus indicate that the immediate zoonotic source of SARS-CoV-2 is yet to be 200 identified and reinforces the unique nature of the SARS-CoV-2 genome. The more global analysis of 201 spike regions in SARS-CoV-2 genomes (Figure 3) revealed the changes that have occurred across the 202 Sarbecovirues. Combined with the current VOC spike changes, the patterns suggest that SARS-CoV-203 2 has a great deal of evolutionary possibilities to avoid immune pressure. This suggests that genomic 204 variant surveillance should continue and vaccine producers should be prepared to accommodate such 205 spike changes in the next generation of vaccine updates. In addition to the spike protein, additional 206 regions of high variance were observed in the nsp3 across all Sarbecoviruses (Figure 2) in close bat 207 and pangolins (Figure 3). The high variance regions flanked and partially overlapped the Macro domain, 208 which is frequently associated with ADP-deribosylase activity (22) (23). Variance observed in the ORF8

209 changes across the set was due to frequent deletion of this ORF, suggesting that the encoded protein 210 may be dispensable for human infection. Similar loss of ORF8 was observed with the original SARS-211 CoV (24) (25) and has been observed in several SARS-CoV-2 lineages as the virus adapted to humans 212 (26) (27) (28). The ORF9 (N protein) variance observed across Sarbecoviruses and the changes in this 213 protein in VOC strains suggest an additional region that may be adapting to human replication. The 214 regions of variance identified here may indicate either functional changes in SARS-CoV-2 proteins or 215 amino acid positions that can be changed without impairing the necessary functions of the protein. The 216 relatively high mutation rate of SARS-CoV-2 combined with the unprecedented number of SARS-CoV-217 2 infections in the world is resulting in massive viral adaptation. Additional experiments are required to 218 distinguish true functional changes from neutral evolution.

Finally, the detailed spike analysis of Figure 3 reveals 82 domains of 15-aa that have shown high variation (1-mean bit-score>= 0.3) in the *Sarbecoviruses* while 29 of these domains show changes in VOC relative to early lineage B SARS-CoV-2. In broad terms, the SARS-CoV-2 evolution observed in the current VOC has sampled only 36% (29/82) of the possible spikes changes which have occurred historically in *Sarbecovirus* evolution. It is highly likely that a large number of new SARS-CoV-2 variants with changes in these regions are possible, compatible with virus replication and expected in the coming months, unless global viral replication is severely reduced.

226

227 Acknowledgements

We thank all global SARS-CoV-2 sequencing groups for their open and rapid sharing of sequence data and GISAID for providing an effective platform for making these data available. The study is funded by the Wellcome, DFID - Wellcome Epidemic Preparedness – Coronavirus (AFRICO19, grant agreement number 220977/Z/20/Z) awarded to MC. DLR and MC receive funding from the MRC (MC_UU_1201412).

233

234 References

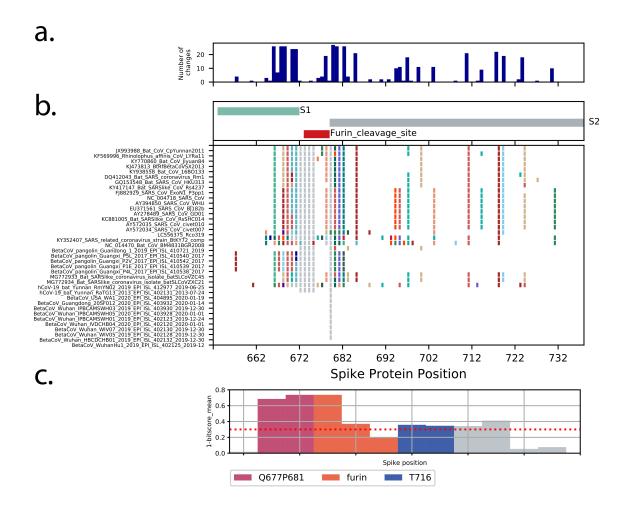
- Li,Q., Guan,X., Wu,P., Wang,X., Zhou,L., Tong,Y., Ren,R., Leung,K.S.M., Lau,E.H.Y., Wong,J.Y., *et al.* (2020)
 Early Transmission Dynamics in Wuhan, China, of Novel Coronavirus–Infected Pneumonia. *N. Engl. J. Med.*, 10.1056/NEJMoa2001316.
- 2. Yang,X., Yu,Y., Xu,J., Shu,H., Xia,J., Liu,H., Wu,Y., Zhang,L., Yu,Z., Fang,M., *et al.* (2020) Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study. *Lancet Respir. Med.*, 10.1016/S2213-2600(20)30079-5.
- 3. Andersen, K.G., Rambaut, A., Lipkin, W.I., Holmes, E.C. and Garry, R.F. (2020) The proximal origin of SARS-CoV 2. *Nat. Med.*, 10.1038/s41591-020-0820-9.
- 4. Boni,M.F., Lemey,P., Jiang,X., Lam,T.T.-Y., Perry,B.W., Castoe,T.A., Rambaut,A. and Robertson,D.L. (2020)
 Evolutionary origins of the SARS-CoV-2 sarbecovirus lineage responsible for the COVID-19 pandemic.
 Nat. Microbiol., 5, 1408–1417.
- 5. Zhang, T., Wu, Q. and Zhang, Z. (2020) Probable Pangolin Origin of SARS-CoV-2 Associated with the COVID-19
 Outbreak. *Curr. Biol.*, 10.1016/j.cub.2020.03.022.

- 6. Zhou,H., Chen,X., Hu,T., Li,J., Song,H., Liu,Y., Wang,P., Liu,D., Yang,J., Holmes,E.C., *et al.* (2020) A novel bat
 coronavirus reveals natural insertions at the S1/S2 cleavage site of the Spike protein and a possible
 recombinant origin of HCoV-19 Microbiology.
- 7. Lam,T.T.-Y., Jia,N., Zhang,Y.-W., Shum,M.H.-H., Jiang,J.-F., Zhu,H.-C., Tong,Y.-G., Shi,Y.-X., Ni,X.-B., Liao,Y.-S.,
 et al. (2020) Identifying SARS-CoV-2-related coronaviruses in Malayan pangolins. *Nature*, 583, 282–
 285.
- 254 8. Eddy, S.R. (1998) Profile hidden Markov models. *Bioinformatics*, **14**, 755–763.
- 255 9. Eddy, S.R. (1996) Hidden Markov models. *Curr. Opin. Struct. Biol.*, 6, 361–365.
- 256 10. Eddy, S.R. (2011) Accelerated Profile HMM Searches. *PLOS Comput. Biol.*, **7**, e1002195.
- 257 11. Phan,M.V.T., Ngo Tri,T., Hong Anh,P., Baker,S., Kellam,P. and Cotten,M. (2018) Identification and
 258 characterization of Coronaviridae genomes from Vietnamese bats and rats based on conserved
 259 protein domains. *Virus Evol.*, 4.
- 12. Masembe,C., Phan,M.V.T., Robertson,D.L. and Cotten,M. (2020) Increased resolution of African Swine
 Fever Virus genome patterns based on profile HMM protein domains Genomics.
- 262 13. Edgar, R.C. (2010) Search and clustering orders of magnitude faster than BLAST. *Bioinformatics*, 26, 2460–
 263 2461.
- 14. Katoh,K. and Standley,D.M. (2013) MAFFT Multiple Sequence Alignment Software Version 7:
 Improvements in Performance and Usability. *Mol. Biol. Evol.*, **30**, 772–780.
- Liu, P., Chen, W. and Chen, J.-P. (2019) Viral Metagenomics Revealed Sendai Virus and Coronavirus Infection
 of Malayan Pangolins (Manis javanica). *Viruses*, **11**, 979.
- 16. Xiao,K., Zhai,J., Feng,Y., Zhou,N., Zhang,X., Zou,J.-J., Li,N., Guo,Y., Li,X., Shen,X., *et al.* (2020) Isolation of
 SARS-CoV-2-related coronavirus from Malayan pangolins. *Nature*, 583, 286–289.
- 270 17. Chan,Y.A. and Zhan,S.H. (2020) Single source of pangolin CoVs with a near identical Spike RBD to SARS-CoV 271 2 Genomics.
- 18. Tang,X.C., Zhang,J.X., Zhang,S.Y., Wang,P., Fan,X.H., Li,L.F., Li,G., Dong,B.Q., Liu,W., Cheung,C.L., *et al.*(2006) Prevalence and Genetic Diversity of Coronaviruses in Bats from China. *J. Virol.*, **80**, 7481–7490.
- 19. Li,W. (2005) Bats Are Natural Reservoirs of SARS-Like Coronaviruses. *Science*, **310**, 676–679.
- 20. Hoffmann,M., Kleine-Weber,H. and Pöhlmann,S. (2020) A Multibasic Cleavage Site in the Spike Protein of
 SARS-CoV-2 Is Essential for Infection of Human Lung Cells. *Mol. Cell*, **78**, 779-784.e5.
- 277 21. Andrew Rambaut (2020) Divergence of nCoV-2019 to closest non-human relative. *Virological.org*.
- 278 22. Frick, D.N., Virdi, R.S., Vuksanovic, N., Dahal, N. and Silvaggi, N.R. (2020) Molecular Basis for ADP-Ribose
 279 Binding to the Mac1 Domain of SARS-CoV-2 nsp3. *Biochemistry*, 59, 2608–2615.
- 280 23. Lei, J., Kusov, Y. and Hilgenfeld, R. (2018) Nsp3 of coronaviruses: Structures and functions of a large multi 281 domain protein. *Antiviral Res.*, 149, 58–74.
- 24. Chiu,R.W.K., Chim,S.S.C., Tong,Y., Fung,K.S.C., Chan,P.K.S., Zhao,G. and Lo,Y.M.D. (2005) Tracing SARS Coronavirus Variant with Large Genomic Deletion. *Emerg. Infect. Dis.*, **11**, 168–170.
- 284 25. Tang, J.W., Cheung, J.L.K., Chu, I.M.T., Sung, J.J.Y., Peiris, M. and Chan, P.K.S. (2006) The Large 386-nt Deletion
 285 in SARS-Associated Coronavirus: Evidence for Quasispecies? J. Infect. Dis., 194, 808–813.

- 286 26. Su,Y.C.F., Anderson,D.E., Young,B.E., Linster,M., Zhu,F., Jayakumar,J., Zhuang,Y., Kalimuddin,S., Low,J.G.H.,
 287 Tan,C.W., *et al.* (2020) Discovery and Genomic Characterization of a 382-Nucleotide Deletion in
 288 ORF7b and ORF8 during the Early Evolution of SARS-CoV-2. *mBio*, **11**, e01610-20,
 289 /mbio/11/4/mBio.01610-20.atom.
- 27. Gong,Y.-N., Tsao,K.-C., Hsiao,M.-J., Huang,C.-G., Huang,P.-N., Huang,P.-W., Lee,K.-M., Liu,Y.-C., Yang,S.-L.,
 Kuo,R.-L., *et al.* (2020) SARS-CoV-2 genomic surveillance in Taiwan revealed novel ORF8-deletion
 mutant and clade possibly associated with infections in Middle East. *Emerg. Microbes Infect.*, 9, 1457–
 1466.
- 28. Young,B.E., Fong,S.-W., Chan,Y.-H., Mak,T.-M., Ang,L.W., Anderson,D.E., Lee,C.Y.-P., Amrun,S.N., Lee,B.,
 Goh,Y.S., *et al.* (2020) Effects of a major deletion in the SARS-CoV-2 genome on the severity of
 infection and the inflammatory response: an observational cohort study. *Lancet Lond. Engl.*, **396**,
 603–611.
- 298
- 299

300 Supplementary Figure 1 demonstrates the relationship between the 1-mean bit-score and amino acid 301 changes across a sequence alignment. A set of twelve, 15 aa pHMM domains across the central region 302 of the spike protein were used to demonstrate the relationship between the total amino acid changes 303 and the resulting pHMM 1-mean bit-scores. An alignment of 38 spike proteins from 10 early SARS-304 CoV-2 genomes, 4 close bat coronavirus Sarbecoviruses plus 24 additional close Sarbecoviruses was 305 prepared and trimmed to an 87 amino acid region spanning conserved and variable sequences across the furin cleavage site. The bit-scores(10) for the twelve, 15 aa pHMMs across each query sequence 306 307 were gathered, and the mean value for each domain across the set was calculated and converted to a 308 1-mean bit-score value for ease of visualization (higher value = greater difference from SARS-CoV-2).

309 The amino acid changes across the alignment are shown in Supplementary Figure 1 panel B. 310 with each colored bar indicating an amino acid change (or gap, indicated in grey) from the spike of reference genome NC-045512, The 3' end of the spike S1, the furin cleavage site and the 5' start of the 311 312 spike S2 region are indicated in Panel B. Peaks of variability are seen flanking and within the furin site 313 and in the S2 region (Supplementary Figure 1 panel A). The 1-mean bit-score values for the 12 domains 314 are plotted in Panel C. and show a related but more softened pattern of variation across the region. The 315 advantage the pHMM method and the 1-mean bit-score metrics is that the score is weighted to reflect 316 the type of amino acid change or insertion/deletions and can be determined guickly in large genome 317 sets using the HMMER tools(10). For our comparative analysis, the cutoff of 0.3 for the 1-mean bit-318 score was set to reflect about 25 amino changes in the total aa positions covered by each domain guery 319 (15 aa domain across 38 sequences = 570 total position differences possible).



320

321 Supplemental Figure 1. Demonstrating relationship between amino acid changes and 1-mean 322 bit score. (B) The spike proteins encoded by the set of Sarbecovirus genomes examined in Figure 2 323 and 3 were aligned and the amino acid changes from the early SARS-CoV-2 genomes (= NC-045512 324 sequence) were depicted as colored lines across each sequence. (A) The total number of proteins 325 showing a change from the early SARS-CoV-2 genomes (NC-045512) were plotted for each position 326 of the alignment. (C) The set of 12, 15 aa pHMMs spanning the central region of the spike (a subset of 327 the pHMMs used for Figure 3) were used to examine the Sarbecovirus spike sequences. For each 328 pHMM, the mean bit-score for the entire sequence sequencer set was calculate and the value 1-mean 329 bit-score is plotted for each of the 12 pHMMs. Domains containing known amino acid changes in VOC 330 (Q677, P681, T716) and the domains spanning the furin cleavage site are indicated by color (with the 331 legend found below panel C).

332