

1 Maximum likelihood inference of pathogen population size history from
2 a phylogeny

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10 ABSTRACT

11 Inference of effective population size from genomic data can provide unique information about
12 demographic history, and when applied to pathogen genetic data can also provide insights into
13 epidemiological dynamics. Non-parametric models for population dynamics combined with molecular
14 clock models which relate genetic data to time have enabled phylodynamic inference based on large sets
15 of time-stamped genetic sequence data. The theory for non-parametric inference of effective population
16 size is well-developed in the Bayesian setting, but here we develop a frequentist approach based on
17 non-parametric latent process models of population size dynamics. We appeal to statistical principles
18 based on out-of-sample prediction accuracy in order to optimize parameters that control shape and
19 smoothness of the population size over time. We demonstrate the flexibility and speed of this approach
20 in a series of simulation experiments and apply the models to genetic data from several pathogen data
21 sets.

22 INTRODUCTION

23 Past fluctuation in the size of a population are reflected in the genealogy of a sample of individuals
24 from that population. For example, under the coalescent model, two distinct lines of ancestry coalesce
25 (ie find a common ancestor) at a rate that is inversely proportional to the effective population size at
26 any given time (Kingman 1982; Griffiths and Tavaré 1994; Donnelly and Tavaré 1995). More coalescent
27 events are therefore likely when the population size is small compared to when the population size is
28 large. This causal effect of population size on genealogies can be reversed in an inferential framework
29 to recover past population size dynamics from a given pathogen genealogy. This approach to inference
30 of past demographic changes was first proposed 20 years ago (Pybus et al. 2000, 2001; Strimmer and
31 Pybus 2001) and has been fruitfully applied to many disease systems (Pybus and Rambaut 2009; Ho
32 and Shapiro 2011; Baele et al. 2016).

33 Population size analysis is often performed within the Bayesian BEAST framework (Suchard et al. 2018;
34 Bouckaert et al. 2019) which jointly infers a phylogeny and demographic history from genetic data. Here
35 we focus on an alternative approach in which the dated phylogeny is inferred first, for example using
36 *treedater* (Volz and Frost 2017), *TreeTime* (Sagulenko et al. 2018) or *BactDating* (Didelot et al. 2018),
37 and demography is investigated on the basis of the phylogeny. Although potentially less powerful, this
38 approach has the advantage of scalability to very large sequence data sets and allows more focus on
39 models and assumptions involved in the demographic inference itself as previously noted in studies
40 following the same post-processing strategy (Lan et al. 2015; Karcher et al. 2017; Volz and Didelot
41 2018). However, some of the methodology and results we describe here should be applicable in a joint
42 inferential setting too.

43 The reconstruction of past population size dynamics is usually based on a non-parametric model, since
44 the choice of any parametric function for the past population size would cause restrictions and be
45 hard to justify in many real-life applications (Drummond et al. 2005; Ho and Shapiro 2011). However,
46 even if a non-parametric approach offers a lot more flexibility than a parametric one, it does not fully
47 circumvent the question of which demographic model to use as the basis of inference. For example,
48 the *skygrid* model considers that the log population size is piecewise constant, with values following a
49 Gaussian Markov chain, in which each value is normally distributed around neighbouring values and
50 standard deviation determined by a precision hyperparameter (Gill et al. 2013). This model can be

51 justified as the discretisation of a continuous *skyride* model in which the log population size is ruled
52 by a Brownian motion (Minin et al. 2008). Alternatively, the *skygrowth* model is a similar Gaussian
53 Markov chain on the growth rate of the population size (Volz and Didelot 2018). Both models can be
54 conveniently extended to explore the association between population size dynamics and covariate data
55 (Gill et al. 2016; Volz and Didelot 2018).

56 The *skygrid*, *skygrowth* or other similar models can be assumed when performing the inference of
57 the demographic function, and the effect of this model choice has not been formally investigated.
58 Furthermore, these discretised non-parametric models require to select the number of pieces in the
59 demographic function, the location of boundaries between pieces, and the prior expectation for the
60 difference from one piece to another, all of which can have significant effect on the inference results.
61 Here we propose several statistical procedures to automatically select the best values for some of these
62 variables. In particular, the parameter controlling the smoothness of the population size function
63 is usually assumed to have an arbitrary non-informative prior distribution in a Bayesian inferential
64 setting (Minin et al. 2008; Gill et al. 2013), whereas we show here that it can be selected using a
65 frequentist statistical approach based on out-of-sample prediction accuracy. We tested the effect of
66 these procedures on simulated datasets, where the correct demographic function is known and can be
67 used to assess the relative value of inference under various conditions. We also reanalysed real datasets
68 from recent studies on viral and bacterial infectious diseases, and show that the new methods can lead
69 to improved epidemiological insights.

70 MATERIALS AND METHODS

71 Non-parametric Models

72 Let the demographic function $N_e(t)$ be piecewise linear with R pieces of equal lengths h . Let γ_i denote
73 the log of the effective population size in the i -th piece. In the *skygrid* model (Gill et al. 2013), the
74 values of γ_i follow a Gaussian Markov chain, with the conditional distribution of γ_{i+1} given γ_i equal
75 to:

$$\gamma_{i+1} \sim \mathcal{N}(\gamma_i, h/\tau) \quad (1)$$

76 By contrast, the *skygrowth* model (Volz and Didelot 2018) is defined using the effective population size
77 growth rates ρ_i which are assumed constant in each interval and are equal to:

$$\rho_i = \frac{\exp(\gamma_{i+1}) - \exp(\gamma_i)}{h \exp(\gamma_i)} \quad (2)$$

78 These growth rate values form a Gaussian Markov chain, with:

$$\rho_{i+1} \sim \mathcal{N}(\rho_i, h/\tau) \quad (3)$$

79 We also define a new model which we call *skysigma* based on the values σ_i of the second order differences
80 of the log of the effective population size:

$$\sigma_i = (\gamma_{i+1} - \gamma_i) - (\gamma_i - \gamma_{i-1}) = \gamma_{i+1} - 2\gamma_i + \gamma_{i-1} \quad (4)$$

81 Once again we consider a Gaussian Markov chain in which:

$$\sigma_{i+1} \sim \mathcal{N}(\sigma_i, h/\tau) \quad (5)$$

82 Each of the models above defines a demographic function $N_e(t)$ from which the likelihood of the
83 genealogy \mathcal{G} can be calculated. Let n denote the number of tips in \mathcal{G} , let $s_{1:n}$ denote the dates of

84 the leaves and $c_{1:(n-1)}$ denote the dates of the internal nodes. Let $A(t)$ denote the number of extant
85 lineages at time t in \mathcal{G} which is easily computed as the number of leaves dated after t minus the number
86 of internal nodes dated after t :

$$A(t) = \sum_{i=1}^n \mathbb{1}[s_i > t] - \sum_{i=1}^{n-1} \mathbb{1}[c_i > t] \quad (6)$$

87 This quantity is important because in the coalescent model, each pair of lineages coalesces at rate
88 $1/N_e(t)$, so that the total coalescent rate at time t is equal to:

$$\lambda(t) = \begin{cases} \frac{A(t)(A(t)-1)}{2N_e(t)}, & \text{if } A(t) \geq 2 \\ 0, & \text{otherwise.} \end{cases} \quad (7)$$

89 The full likelihood of the coalescent process is therefore computed as (Griffiths and Tavaré 1994;
90 Donnelly and Tavaré 1995):

$$L(\mathcal{G}|N_e(t)) = \exp\left(-\int_{-\infty}^{\infty} \mathbb{1}[A(t) \geq 2] \frac{A(t)(A(t)-1)}{2N_e(t)} dt\right) \prod_{i=1}^{n-1} \frac{1}{N_e(c_i)} \quad (8)$$

91 This computation is straightforward for the models considered here where the demographic function
92 $N_e(t)$ is piecewise constant.

93 Selection of the precision parameter

The precision parameter (also called the 'smoothing' parameter) τ controls how much subsequent values of the Markov N_e will vary when data is uninformative. The selection of this parameter is therefore shaped by competing aims of maximizing explanatory power while reducing overfitting. In frequentist statistics, a standard approach to selecting smoothing parameters is to minimize out-of-sample prediction error. Here, we pursue a k -fold cross-validation strategy where genealogical data is partitioned into k sets, $k-1$ of which are used for fitting, and the last one is used for prediction. This

procedure is equivalent to maximizing the following objective function:

$$f(\tau) = \prod_{j=1}^k P(\mathcal{G} \setminus X_j | \hat{N}_e(X_j, \tau)), \quad (9)$$

94 where $\hat{N}_e(X_j, \tau)$ is the maximum likelihood estimates of N_e on the partial data $X_j \subset \mathcal{G}$ and assuming
95 the precision parameter is τ . In this case $X_{j=1:k}$ represents a subset of the sample times and internal
96 node times of the genealogy \mathcal{G} .

97 This is a standard formulation of the cross-validation method, but the implementation depends on how
98 genealogical data is partitioned. We use the strategy of discretizing the coalescent likelihood (Equation
99 8) into intervals bordered by the time of nodes (tips s_i or internal nodes c_i of the tree) and/or the $R - 1$
100 times when the piecewise-constant N_e changes value. Given $R - 1$ change points, n tips, and $n - 1$
101 internal nodes of \mathcal{G} , there are $R + 2n - 3$ intervals $(\iota_1, \dots, \iota_{R+2n-3})$. Each cross-validation training set
102 is formed by taking a staggered sequence of intervals and collecting the genealogical data contained in
103 each, so that $X_k = \{\iota_j | \text{modulo}(j, k) \neq 0\}$.

104 Selection of the grid resolution

105 Before any of the non-parametric models described above can be fitted, the number R of pieces in the
106 piecewise demographic function needs to be specified. Setting R too low may lead to an oversimplified
107 output that does not capture all the information on past population changes suggested by the genealogy,
108 whereas setting R too high can lead to overfitting. We therefore propose to use well established
109 statistical methods to select the optimal value of R . First the model is fitted for multiple proposed
110 values of R , and then for each output we compute the Akaike information criterion (AIC), which is
111 equal to:

$$\text{AIC}_R = 2R - 2\log(L_R) \quad (10)$$

112 where L_R is the maximum value of the likelihood when using R pieces. The value of R giving the
113 smallest value of AIC_R is selected. We also implemented the Bayesian information criterion (BIC),
114 which are respectively equal to:

$$\text{BIC}_R = R\log(n - 1) - 2\log(L_R) \quad (11)$$

115 However, we found that the BIC was often overly conservative in the choice of the resolution as
116 previously noted (Kuha 2004; Weakliem 1999), and therefore we focus here in the use of AIC.

117 **Simulation of testing data**

118 In order to test the accuracy of our methodology, we implemented a new simulator of coalescent
119 genealogies given sampling dates and a past demographic function $N_e(t)$. When the demographic
120 function is constant, the simulation of coalescent genealogies is equivalent to simulating from a
121 homogeneous Poisson process, in which the waiting times from one event to the next are exponentially
122 distributed. To extend this to the situation where the demographic function is non-constant requires
123 to simulate from an equivalent non-homogeneous Poisson process. The approach we used to achieve
124 this is to consider a homogeneous Poisson process with a population size N_m which is lower than any
125 value of $N_e(t)$, ie $\forall t, N_e(t) \geq N_m$. We simulate this process using exponential waiting times, but filter
126 an event happening at time t according to the ratio $N_m/N_e(t)$. Specifically, we draw $u \sim \text{Unif}(0, 1)$ and
127 if $u < N_m/N_e(t)$ the event is accepted and otherwise rejected. The resulting filtered Poisson process
128 simulates from the non-homogeneous Poisson process as required (Ross 2014). The disadvantage of
129 this approach over other methods of simulations is that there may be many rejections if $N_e(t)$ takes
130 small values so that N_m needs to be small too. This is therefore not the most efficient method of
131 simulation. However, efficiency of simulation is not important for our purpose here, and this method
132 has the advantage to avoid the computation of integrals on the $N_e(t)$ function which other methods
133 would require.

134 **Implementation**

135 We implemented the simulation and inference methods described in this paper into a new R package
136 entitled *mlesky* which is available at <https://github.com/emvolz-phylogenetics/mlesky> . If
137 multiple CPU cores are available, these resources are exploited within the procedure of selection of
138 the smoothing parameter where the computation can be split between the different cross values in the
139 cross-validation. Multicore processing is also applied in the procedure of selection of the grid resolution
140 where computation can be split between different values of the number R .

141 RESULTS

142 Application to simulated dataset with constant population size

143 A dated phylogeny was simulated with 200 tips sampled at regular intervals between 2000 and 2020,
144 and a constant past population size function $N_e(t) = 20$ (Figure 1). To illustrate the importance of
145 the resolution R and precision τ parameters, we inferred the demographic function under the *skygrid*
146 model (cf Equation 1) for a grid of values with $R \in \{5, 20, 50\}$ and $\tau \in \{1, 10, 20\}$ (Figure 2). The
147 results look quite different depending on the parameters used, and in particular when R is large and τ
148 is small, fluctuations in the population size are incorrectly inferred. When applying the AIC procedure
149 to this dataset, the correct value of $R = 1$ was inferred for which the parameter τ becomes irrelevant.
150 In these conditions the effective population size was estimated to be 19.65 with confidence interval
151 ranging from 17.10 to 22.57 which includes the correct value of 20 used in the simulation. We repeated
152 the AIC procedure for 100 different phylogenies all which had been simulated under the same constant
153 population size conditions described above. For 65 of these phylogenies the AIC procedure selected
154 $R = 1$, with the third quartile falling on $R = 3$ and only one simulation giving $R > 10$.

155 Application to simulated dataset with sinusoidal population size

156 Next we simulated a dated phylogeny with the same number and dates of the tips as before, but using
157 a demographic function $N_e(t)$ that was sinusoidal with minimum 2 and maximum 22, with period 6.28
158 years. Figure 3 shows both the demographic function used and the resulting simulated phylogeny. We
159 attempted to reconstruct the demographic function based on the phylogeny under the three models
160 *skygrid*, *skygrowth* and *skysigma* described in Equations 1, 3 and 5, respectively. For each model
161 the precision parameter τ was optimised using our new cross-validation procedure and the number of
162 pieces was set to be $R = 20$ for ease of comparison. The results obtained in these conditions were
163 very similar under the three models (Figure 4). This suggests that when the precision parameter is
164 optimised using the cross-validation method, the choice between these three models becomes relatively
165 unimportant. The choice of using one model rather than another is therefore mostly guided by the
166 presence of covariate data and whether these are expected to correlate with the effective population

167 size directly or some other function of it such as the population growth rates (Gill et al. 2016; Volz
168 and Didelot 2018).

169 **Application to simulated dataset with bottleneck in population size**

170 We simulated another dated phylogeny with the same and dates of the tips as before, but using a
171 bottleneck function for $N_e(t)$ which was equal to 10 at all times except between 2005 and 2010 when it
172 was equal to 1. Figure 5 shows both this bottleneck function and the phylogeny simulated accordingly.
173 We reconstructed the demographic function using the *skygrid* model. The lowest value of the AIC was
174 obtained for $R = 14$, and the precision parameter was optimised using the cross-validation procedure
175 to $\tau = 0.87$. The inferred demographic function is shown in Figure 5, where the bottleneck between
176 2005 and 2010 has been accurately detected.

177 **Application to HIV dataset**

178 We analyzed 399 HIV-1 sequences from Senegal between 1990 and 2014 (Nascimento et al. 2020). All
179 sequences are subtype CRF02_AG. We used treedater (Volz and Frost 2017) to reconstruct a dated
180 phylogeny (Figure 6). This phylogeny has a common ancestor around 1972 and the number of lineages
181 through time having rapid change in the early 1980s when the HIV epidemic was expanding. We
182 applied the AIC procedure to determine the optimal number of pieces to be used for the demographic
183 function, which was found to be $R = 35$. The optimal value of the precision parameter was determined
184 using the cross-validation procedure, and found to be $\tau = 4.40$. The demographic function inferred
185 using these values of R and τ is shown in Figure 6. The whole analysis took less than 30 seconds on
186 a standard laptop computer.

187 **Application to cholera dataset**

188 We applied our methodology to a previously described collection of 260 genomes from the seventh
189 pandemic of *Vibrio cholerae* (Didelot et al. 2015). A genealogy was estimated in this previous study
190 using an early version of BactDating (Didelot et al. 2018), and it is reproduced in Figure 7. We applied

191 the AIC procedure to determine that the demographic function would be modelled using $R = 16$ pieces.
192 The precision parameter was optimised to a value of $\tau = 1.84$ using the cross-validation procedure.
193 The whole analysis took less than 20 seconds on a standard laptop computer. The result is shown in
194 Figure 7. A first peak was detected in the 1960s, followed by a second peak in the 1970s and finally a
195 third peak in the 1990s. This demographic function follows closely on the previously described three
196 “waves” of cholera spreading globally from the Bay of Bengal (Mutreja et al. 2011; Didelot et al. 2015;
197 Weill et al. 2017). However, these three waves had previously been described based on phylogeographic
198 reconstructions of the spread of the pandemic around the world. The fact that we found a similar
199 wave pattern in our analysis which did not include any information about the geographical origin of
200 the genomes provides further support for the validity of this phylodynamic reconstruction.

201 DISCUSSION

202 Non-parametric phylodynamic inference of population size dynamics is usually carried out in a Bayesian
203 framework (Minin et al. 2008; Gill et al. 2013; Volz and Didelot 2018). Here we presented methods
204 for performing such inference in a frequentist setting with a particular view towards model selection
205 and reducing over-fitting. Optimal smoothing can be obtained in a natural way using standard cross-
206 validation methods, and the optimal resolution of the discretised demographic function is achieved
207 using the well-established AIC criterion. This approach can be advantageous when prior distributions
208 are difficult to design or results are sensitive to arbitrarily chosen priors. Methods based on likelihood
209 maximization is also quite fast and scalable to data sets much larger than is conventionally studied
210 with Bayesian methods, and the selection of smoothing parameters does not require arbitrarily chosen
211 hyperparameters. Conventional AIC metrics also alleviate the difficulty of model selection. In most of
212 our simulations, we find relatively little difference in our estimates when parameterizing the model in
213 terms of $N_e(t)$ (Equation 1), or the growth rate of $N_e(t)$ (Equation 3) or its second order variation of
214 $N_e(t)$ (Equation 5).

215 Our methodology assumed that a dated phylogeny has been previously reconstructed from the genetic
216 data. It is therefore well suited for the post-processing analysis of the outputs from *treedater* (Volz
217 and Frost 2017) or *TreeTime* (Sagulenko et al. 2018). A key assumption of our method, as with
218 its Bayesian counterparts, is that all samples in the phylogeny come from a single population ruled
219 by a unique demographic function. To ensure that this is indeed the case, complementary methods
220 are emerging that can test for the presence or asymmetry or hidden population structure in dated
221 phylogenies (Dearlove and Frost 2015; Volz et al. 2020).

222 Past variations in the effective population size of a pathogen population can reveal key insights into past
223 epidemiological dynamics and help make predictions about the future. It is important to note that the
224 effective population size is not generally equal to or even proportional to the number of infections over
225 time (Volz et al. 2009; Dearlove and Wilson 2013). On the other hand, the growth rate of the effective
226 population size (Equation 2) can be used to estimate the basic reproduction number over time $R(t)$
227 which represents the average number of secondary infections caused by an infected individual (Volz
228 et al. 2013; Volz and Didelot 2018). Having good estimates of this quantity is especially important for
229 assessing the effect of infectious disease control measures (Fraser 2007), and phylodynamic approaches

230 provide a useful complementary approach to more traditional methods of estimation based on case
231 report data (Cori et al. 2013).

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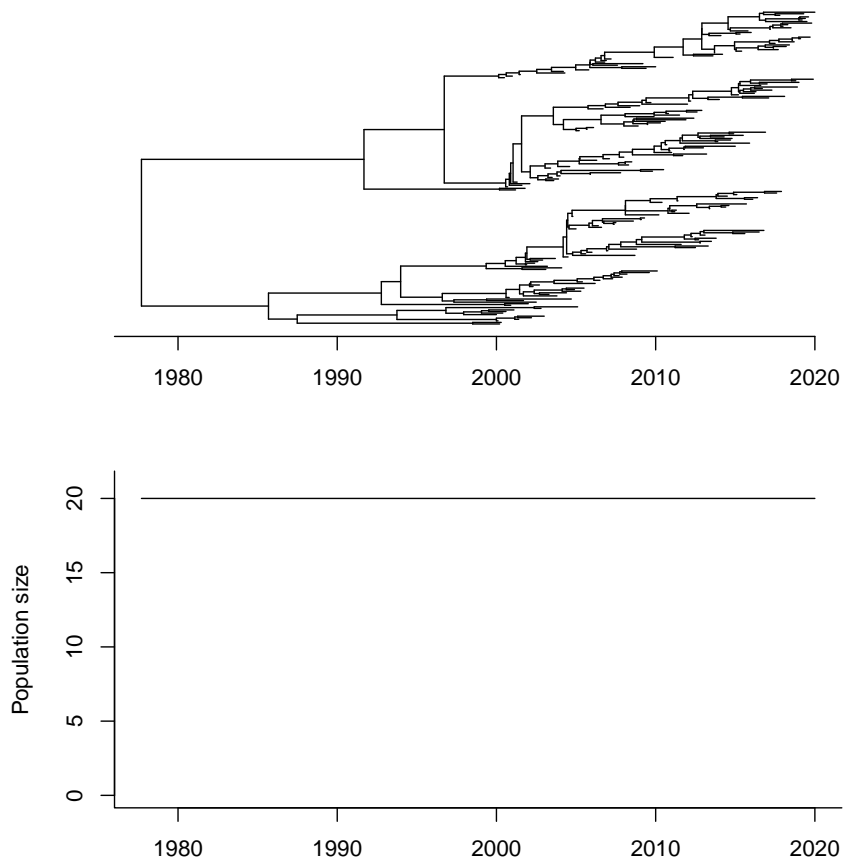


Figure 1: Simulated dataset using a constant past population size function.

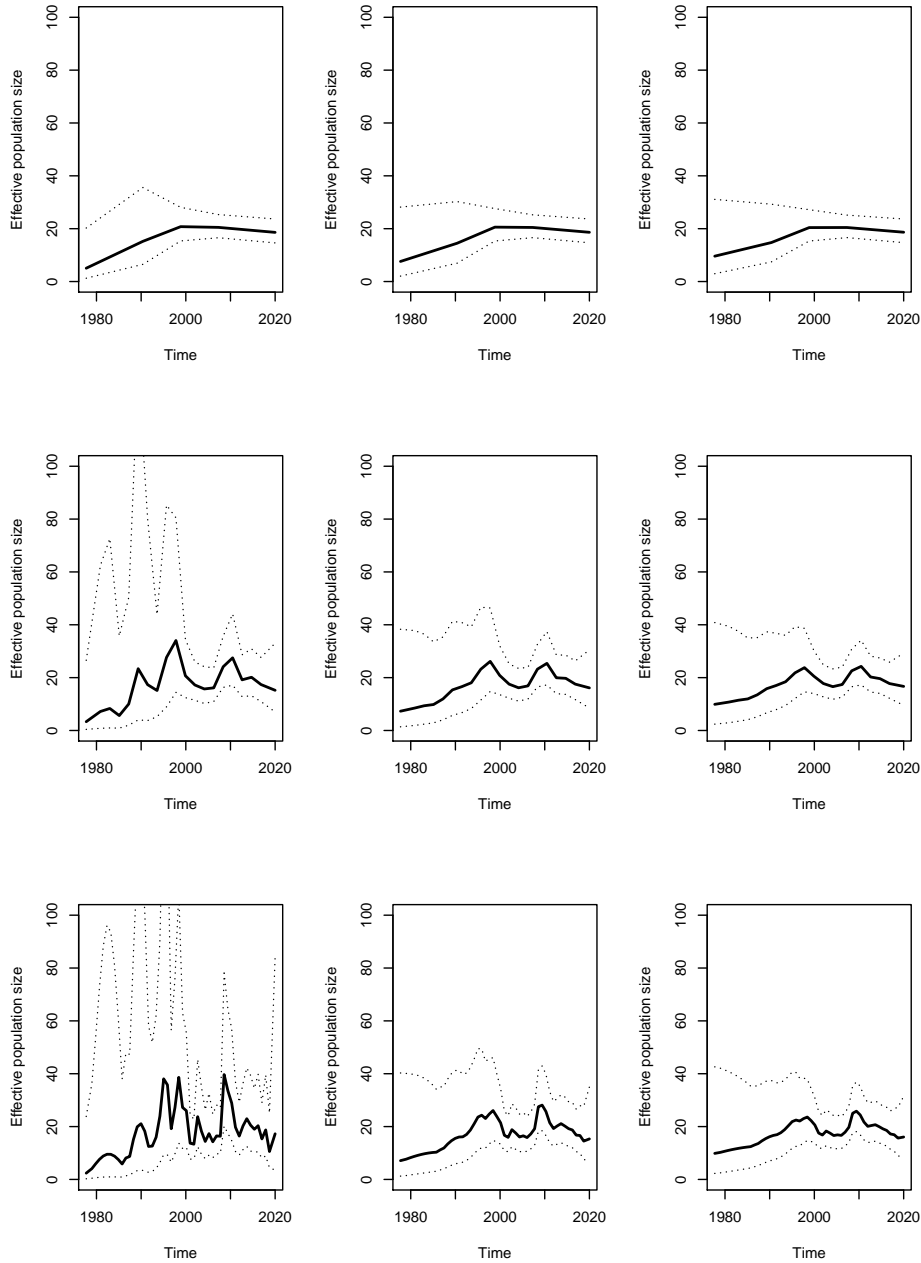


Figure 2: Result on simulated dataset shown in Figure 1 using the skyline model, from top to bottom $R = 5, 20, 50$ and from left to right $\tau = 1, 10, 20$.

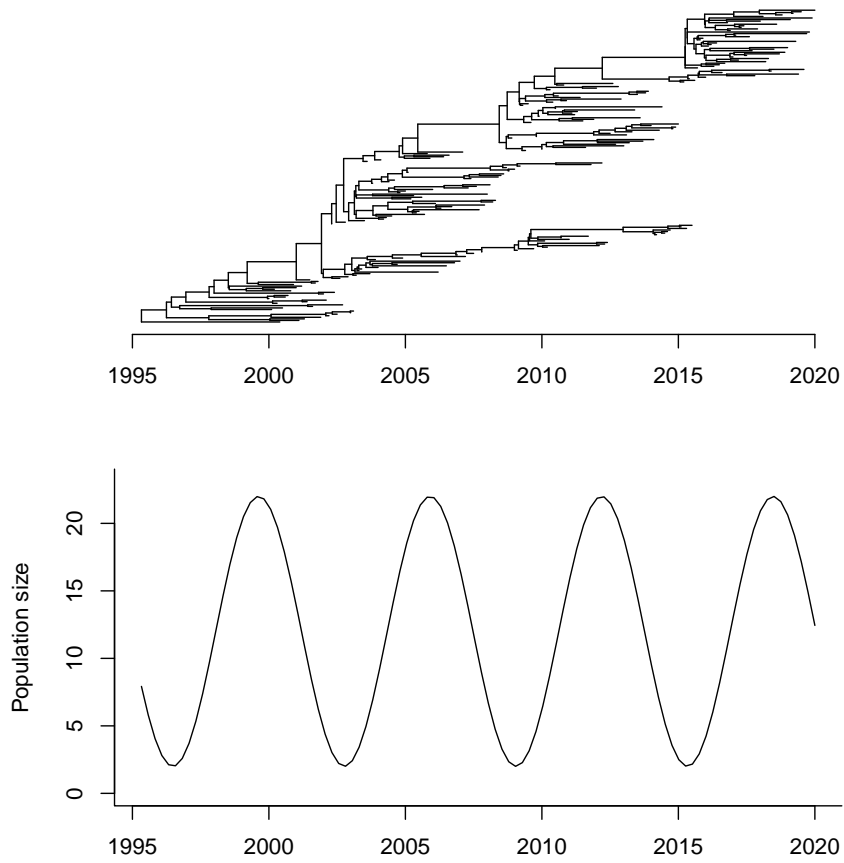


Figure 3: Simulated data using a sinusoidal past population size function.

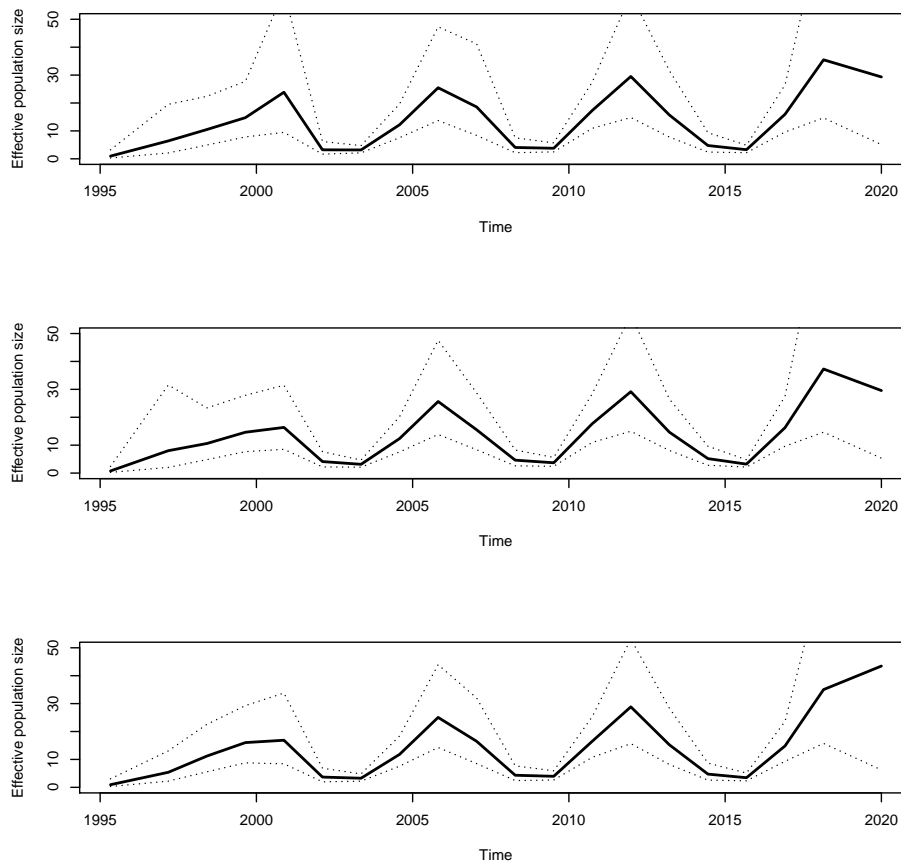


Figure 4: Result of applying the three different models to the phylogeny shown in Figure 3.

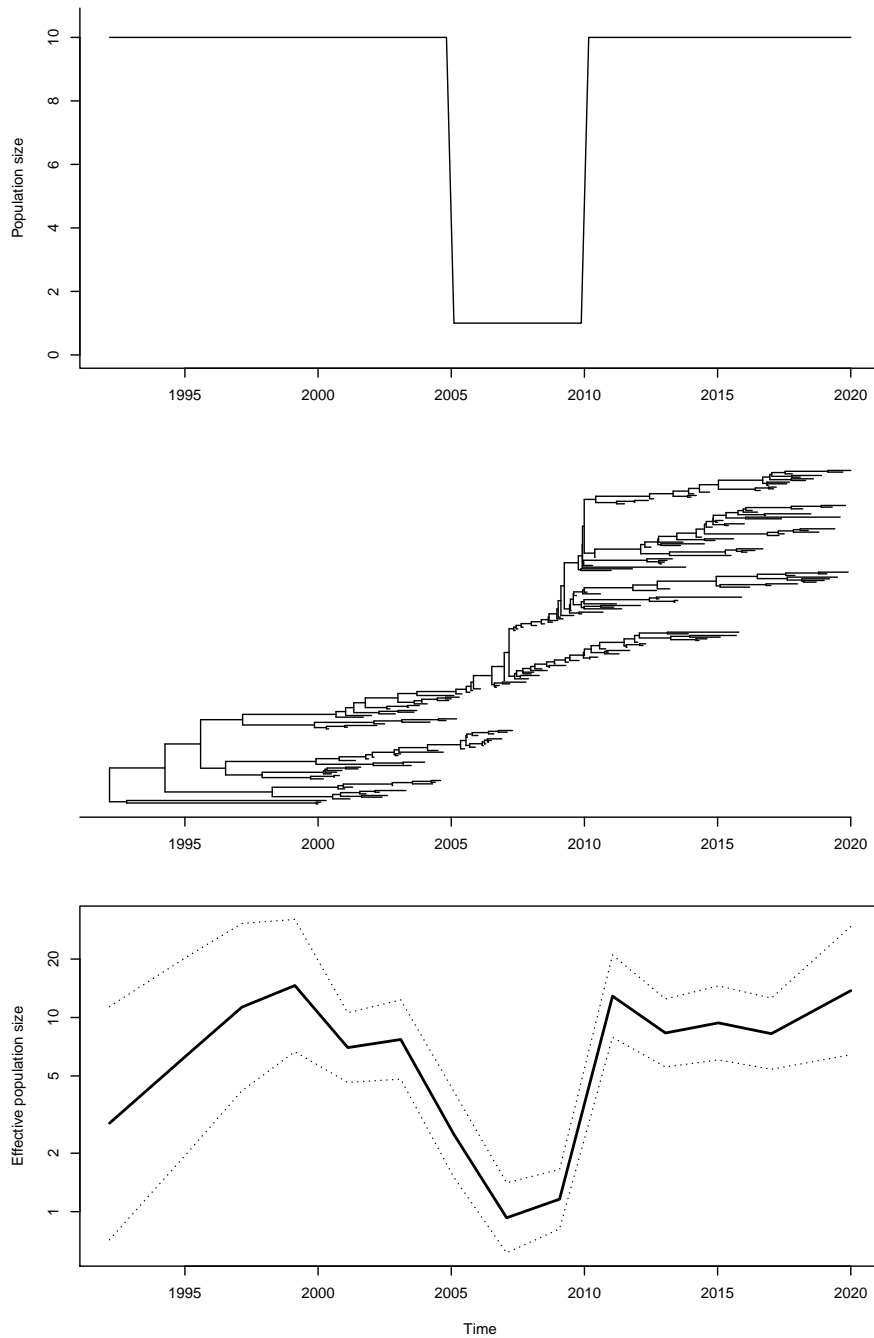


Figure 5: Demographic function (top), phylogeny (middle) and inferred demographic function (bottom) for a simulated dataset under a bottleneck model.

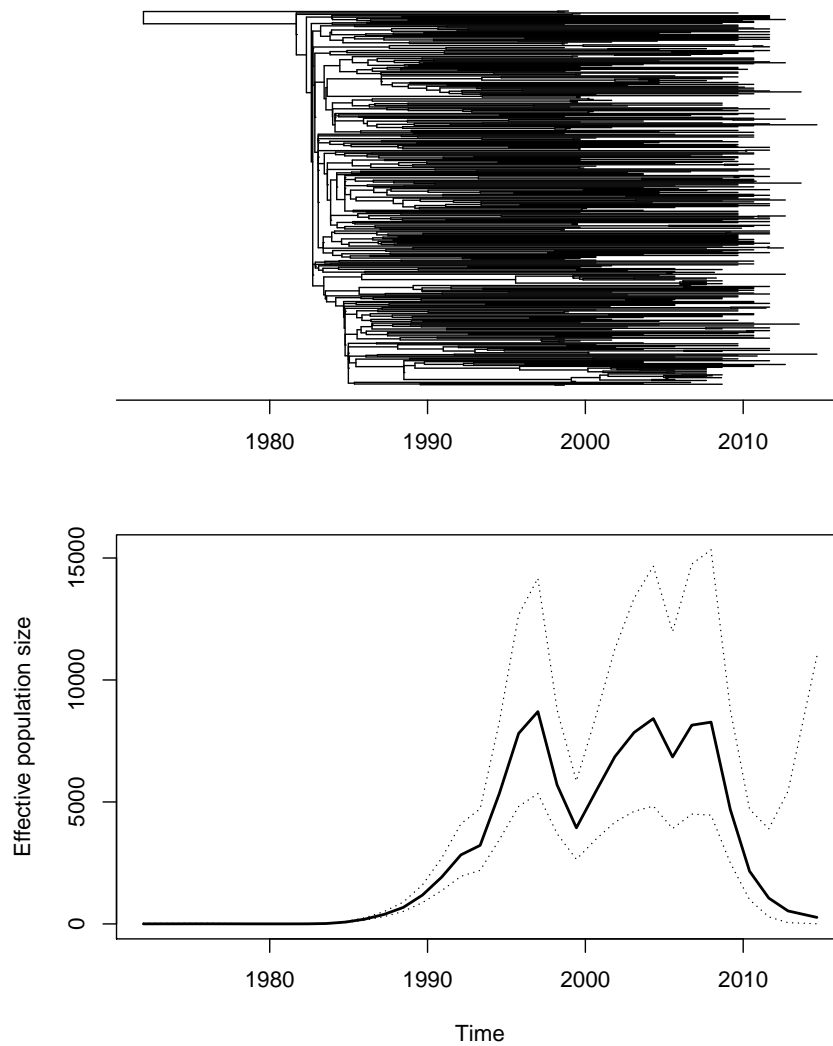


Figure 6: Analysis of the HIV dataset. Top: Dated phylogeny used as the starting point of past population size inference. Bottom: Demographic function reconstructed based on the phylogeny above.

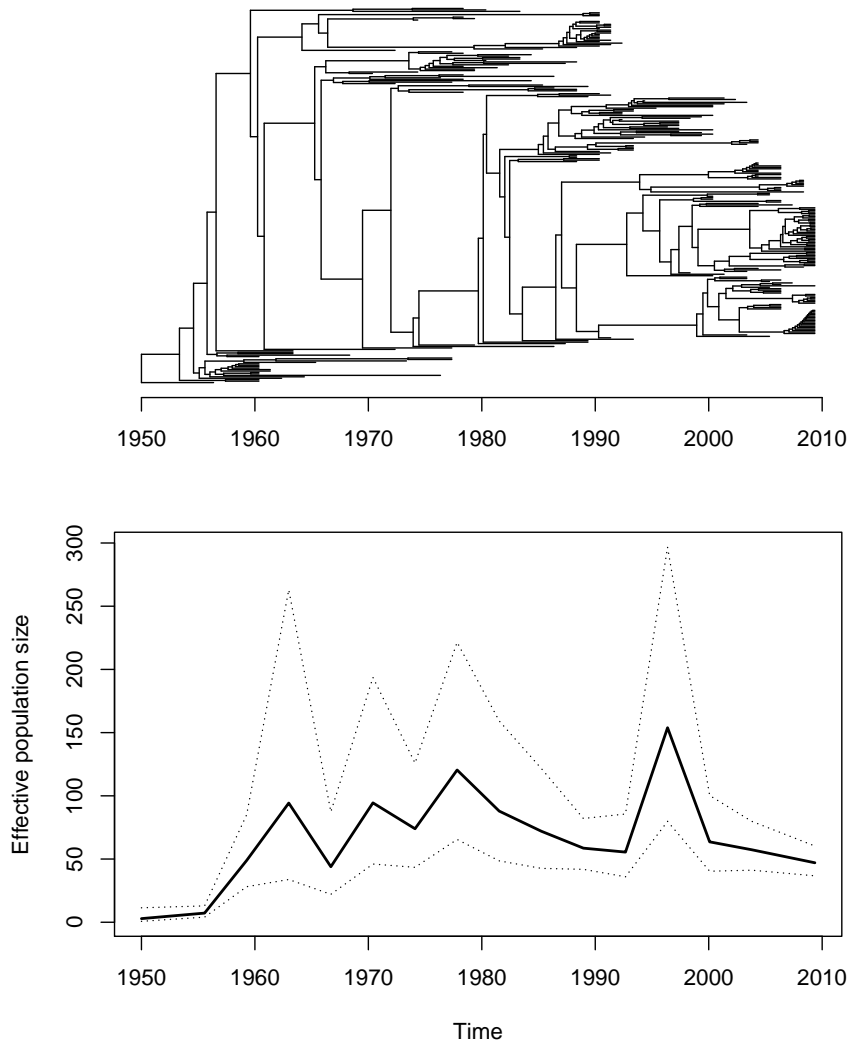


Figure 7: Analysis of the seventh pandemic of *Vibrio cholerae*. Top: Dated phylogeny used as the starting point of past population size inference. Bottom: Demographic function reconstructed based on the phylogeny above.