# Pulled Diversification Rates, Lineage-Through-Time Plots and Modern Macroevolutionary Modelling

ANDREW J. HELMSTETTER<sup>1,\*</sup>, SYLVAIN GLEMIN<sup>2</sup>, JOS KÄFER<sup>3</sup>, ROSANA ZENIL-FERGUSON<sup>4</sup>, HERVÉ SAUQUET<sup>5,6</sup>, HUGO DE BOER<sup>7</sup>, LÉO-PAUL M. J. DAGALLIER<sup>8</sup>, NATHAN MAZET<sup>9</sup>, ELIETTE L. REBOUD<sup>9</sup>, THOMAS L. P. COUVREUR<sup>8</sup>, AND FABIEN L. CONDAMINE<sup>9</sup>

<sup>1</sup> Fondation pour la Recherche sur la Biodiversité - Centre for the Synthesis and Analysis of Biodiversity, 34000 Montpellier, France

<sup>2</sup> CNRS, Ecosystèmes Biodiversité Evolution (Université de Rennes), 35000 Rennes, France

<sup>3</sup> Université de Lyon, Université Lyon 1, CNRS, Laboratoire de Biométrie et Biologie Evolutive UMR 5558, F-69622 Villeurbanne, France

<sup>4</sup> School of Life Sciences, University of Hawaii Manoa, Honolulu, HI, 96822, USA

<sup>5</sup> National Herbarium of New South Wales, Royal Botanic Gardens and Domain Trust, Sydney, New South Wales, 2000, Australia

<sup>6</sup> Evolution and Ecology Research Centre, School of Biological, Earth and

Environmental Sciences, University of New South Wales, Sydney, Australia <sup>7</sup> Natural History Museum, University of Oslo, 0318 Oslo, Norway

<sup>8</sup> IRD, DIADE, UNIV MONTPELLIER, MONTPELLIER, FRANCE

<sup>9</sup> CNRS, Institut des Sciences de l'Evolution de Montpellier (Université de Montpellier), 34095 Montpellier, France

\*ANDREW J. HELMSTETTER, FRB-CESAB, MONTPELLIER, 34000, FRANCE.

ANDREW.J.HELMSTETTER @GMAIL.COM

2

#### HELMSTETTER ET AL.

# Abstract

Estimating time-dependent rates of speciation and extinction from dated phylogenetic trees of extant species (timetrees), and determining how and why they vary is key to 2 understanding how ecological and evolutionary processes shape biodiversity. Due to an 3 increasing availability of phylogenies, a growing number of process-based methods relying 4 on the birth-death model have been developed in the last decade to address a variety of 5 questions in macroevolution. However, this methodological progress has regularly been 6 criticised such that one may wonder how reliable the estimations of speciation and 7 extinction rates are. In particular, using lineage-through-time (LTT) plots, a recent study (Louca and Pennell, 2020) has shown that there are an infinite number of equally likely 9 diversification scenarios that can generate any timetree. This has led to questioning 10 whether or not diversification rates should be estimated at all. Here we summarize, clarify, 11 and highlight technical considerations on recent findings regarding the capacity of models 12 and inferences to disentangle diversification histories. Using simulations we demonstrate 13 the characteristics of pulled diversification rates and their utility. We recognize the recent 14 findings are a step forward in understanding the behavior of macroevolutionary modelling, 15 but they in no way suggest we should abandon diversification modelling altogether. On the 16 contrary, the study of macroevolution using phylogenies has never been more exciting and 17 promising than today. We still face important limitations in regard to data availability and 18 methodological shortcomings, but by acknowledging them we can better target our joint 19 efforts as a scientific community. 20

<sup>21</sup> Key words: Birth-death models, extinction, speciation, phylogenetics

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

### INTRODUCTION

22

A major goal in evolutionary biology is to understand the large-scale processes that 23 have shaped biodiversity patterns through time. One important way to investigate this is 24 by modelling species diversification using speciation and extinction, which can vary over 25 time and among groups. It is commonplace to find areas, or clades, in phylogenetic trees 26 that accumulate lineages faster than others. Diversification models often aim to explain 27 this variation in diversification patterns by associating bursts of speciation or extinction 28 with factors such as time (Höhna et al., 2016b), lineages (Rabosky, 2014), character traits 29 (Maddison et al., 2007), or the environment (Morlon et al., 2016). 30

The growing number of large phylogenetic trees that capture a significant 31 proportion of living species provide increasing power and resolution for such studies (Jetz 32 et al., 2012; Smith and Brown, 2018; Upham et al., 2019). Furthermore, the availability of 33 a wide variety of methods and software (e.g. BAMM (Rabosky, 2014), -SSE models 34 (Maddison et al., 2007), RPANDA (Morlon et al., 2016), MEDUSA (Alfaro et al., 2009)) 35 have made diversification studies increasingly popular in the last decade. Approaches that 36 can link diversification to a particular process or trait are among the most appealing to 37 researchers in the field because they enable us to test long-standing hypotheses in 38 evolutionary biology and ecology including those related to the evolution of key 39 innovations (Silvestro et al., 2014), the colonisation of new areas (McGuire et al., 2014), 40 the effect of elevation (Lagomarsino et al., 2016; Quintero and Jetz, 2018) and the 41 latitudinal diversity gradient (Rolland et al., 2014; Pulido-Santacruz and Weir, 2016; 42 Rabosky et al., 2018; Igea and Tanentzap, 2020). 43

A recent study (Louca and Pennell, 2020) (abbreviated to LP) demonstrates how one approach, based on lineage-through-time (LTT) plots, cannot reliably estimate rates of speciation and extinction over time using extant timetrees. LP show how results of this approach can be misleading and provide potential solutions to the issues raised by proposing new summary statistics. This publication has provoked a response from the

4

### HELMSTETTER ET AL.

<sup>49</sup> community (Morlon et al., 2020) and stimulated considerable discussion, with some going <sup>50</sup> so far as to suggest that speciation and extinction cannot be estimated using phylogenetic <sup>51</sup> trees (Pagel, 2020). As a result, this study has called into question the meaning of <sup>52</sup> diversification rate estimates generated from any analytical framework. Here, we aim to <sup>53</sup> outline the major concepts discussed in LP in an accessible way, targeting a broad <sup>54</sup> audience. We then put the results and conclusions of LP into historical context and explore <sup>55</sup> how the implications of this study apply to macroevolutionary modelling today.

56

### MODELLING DIVERSIFICATION RATES

A typical workflow for diversification rate modelling using molecular phylogenetic 57 trees is as follows. DNA sequence data are obtained for species in a study group, which are 58 then used to estimate species relationships in the form of a phylogenetic tree. Typically, 59 this phylogenetic tree contains only extant species, and it is time-calibrated using ages 60 derived from different sources including fossils (Sauquet, 2013) (note, however, that fossils 61 are usually only used for calibration and tree shape estimation and not incorporated in 62 subsequent estimation of diversification rates). The output of this process is referred to as 63 an extant timetree. Once a tree has been generated, a birth-death model is fitted to 64 explain patterns of diversification in the tree. 65

The simplest birth-death models assume that each branch of a phylogenetic tree 66 shares the same rate of "birth" (speciation) events, as well as "death" (extinction) events 67 (Nee et al., 1994; Nee, 2006; Ricklefs, 2007; Morlon et al., 2011). The speciation rate (or 68  $\lambda(t)$  is the rate at which lineages arise at time t (or more precisely during a small time 60 interval, dt, between t and t + dt, formally called infinitesimal birth rate). Likewise, 70 extinction rate (or  $\mu(t)$ ) is the rate at which lineages disappear. Under this simple 71 framework  $\lambda$  and  $\mu$  are constant over time and the same across all clades. In addition, not 72 all extant taxa are included in the phylogenetic tree and the percentage of lineages present 73 is known as the sampling fraction (or  $\rho$ ). By making use of all of these parameters, a 74

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

birth-death model allows us to investigate whether the net diversification rate, defined as 75  $r(t) = \lambda(t) - \mu(t)$ , has varied over time or among clades (Morlon et al., 2011; Rabosky, 76 2014; Maliet et al., 2019; Barido-Sottani et al., 2020) and ultimately uncover the processes 77 that have given rise to extant biodiversity in the study group. 78

# A summary of the main concepts and findings in Louca and Pennell (2020) 79 The deterministic Lineage Through Time plot

80

The approach used by LP relies on the Lineage Through Time (LTT) plot (Nee 81 et al., 1992) (Fig. 1), which shows how extant lineages (i.e. only those existing in the 82 present-day) accumulated over time using a phylogenetic tree. Each point in an LTT 83 corresponds to a change in the number of lineages from the root of a phylogenetic tree at 84 t = 0 to the present day at t = T (Fig. 1a). This perspective is typically used when looking 85 at evolution from the coalescent standpoint. Alternatively, as in LP, time can be counted 86 backwards as an age (using the symbol  $\tau = T - t$ ), starting from the present day at  $\tau = 0$ 87 until  $\tau = T$  at the origin of the clade (Fig. 1b). This approach is often used when plotting 88 phylogenetic trees. For easier interpretation and consistency with LP, we will generally 89 consider timescale as age, or  $\tau$ , throughout this manuscript. 90

Simply put, when a clade diversifies faster, the slope of the LTT becomes steeper, 91 but when diversification slows, the slope of the LTT levels off. When only extant lineages 92 are considered, as in LP, LTT plots will never exhibit a drop in total lineage diversity over 93 time, that is, the slope will never be negative. However, this does not mean that extinction 94 does not have an effect on the shape of the LTT (Nee, 2006). By examining the shape of 95 the LTT plot we can begin to understand how diversification rates fluctuated over the 96 history of a clade (Ricklefs, 2007) and develop evolutionary hypotheses on why these 97 fluctuations occurred. 98

To study general properties of phylogenetic trees, a model of the branching process qq is used. Several models are available, but the birth-death model remains the most widely 100

6

### HELMSTETTER ET AL.

used, and is easily interpreted (Nee, 2006). The birth-death model is a continuous-time 101 Markov chain where at any given age  $(\tau)$  we can calculate the probability of speciation 102 (birth of a lineage) or extinction (death of a lineage) happening. The stochastic nature of 103 the model means that multiple simulations will result in different histories of 104 diversification. For stochastic models like the birth-death model we can calculate their 105 expected value, either by averaging over multiple realisations (simulations) or by 106 approximating it with a set of continuous equations, yielding a deterministic model. Such a 107 model directly yields the expected value one would get by averaging over infinitely many 108 realisations, thus it is deterministic because it is fully defined by the parameters, that is, 109 no uncertainty from stochasticity is involved. This latter approach is taken by LP who 110 model the birth-death process as a set of differential equations, which is advantageous 111 because these equations can be solved analytically. 112

LP refer to an LTT generated by such models as a deterministic LTT or dLTT, 113 which corresponds here to the expected LTT generated by trees with given speciation and 114 extinction rates. Empirical LTTs generated using extant timetrees can be compared to 115 model-generated dLTTs (where  $\lambda$  and  $\mu$  are known) to disentangle, on average, how 116 speciation and extinction have influenced patterns of diversity over time. To do this, the 117 probability of the data given the model, or the likelihood, is calculated and compared 118 across different models to select the one that fits best. Importantly, LP showed that, when 119  $\lambda$  and  $\mu$  are shared across the tree, the likelihood can be fully written as a function of the 120 observed LTT and the dLTT. Typically, as the parameters of the models are modified, the 121 likelihood will change, the model becoming more or less likely. The best-fitting model can 122 then be selected, representing our best hypothesis for how and to what extent speciation 123 and extinction rates varied over time. 124

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

125

### Model congruence and congruence classes

One might expect that every dLTT is represented by a single model - a single set of 126 histories or continuous functions for  $\lambda(t)$  and  $\mu(t)$  - but LP show that this is not the case. 127 Instead, different models can produce the same dLTT. These models may have very 128 different patterns of speciation and extinction over time, yet, if different models produce 129 the same dLTT then they will also share the same likelihood for any given LTT. This 130 makes it impossible to choose the model that is the best representative of the evolutionary 131 history of the clade in question. Models that generate the same dLTT fall into the same 132 "congruence class". These congruence classes contain an infinite number of models with 133 different parameter values that all produce the same dLTT. LP explain that when trying to 134 select the best model we often start with a relatively small set of allowed models that we 135 test. An example of such a difference is whether speciation rate is fixed, or allowed to vary 136 over time. LP suggest that instead of selecting the model closest to the true process, we are 137 instead selecting the model closest to the congruence class that includes the true process 138 (see Fig. 3 in LP). In extreme cases, the best fitting model could thus be further from the 139 true process than a model with a lower likelihood, just because the former is included in 140 the congruence class and the latter is not. However, LP concede that because we only 141 assess a limited set of models, it is unlikely that we encounter models belonging to the 142 same congruence class, but it is nevertheless possible. The consequence of multiple, equally 143 likely models with different speciation and extinction rates is that these rates cannot be 144 determined. This is a statistical phenomenon known as unidentifiability - the likelihood is 145 the same for multiple parameter values making it impossible to choose one over another. 146

147

# Unidentifiability

In macroevolutionary modelling we might be interested to know how both  $\lambda$  and  $\mu$ have changed over time (Alfaro et al., 2009). However, the unidentifiability issue outlined above means that we would not be able to ascertain the true parameter values of the

 $\overline{7}$ 

8

### HELMSTETTER ET AL.

<sup>151</sup> models that generate our dLTTs. Another well-known example of this in macroevolution is <sup>152</sup> the unidentifiability of  $\alpha$  and  $\theta$  from Ornstein-Uhlenbeck models of trait evolution (Ho and <sup>153</sup> Ané, 2014). This problem is not unique to macroevolutionary models, and, in fact, stems <sup>154</sup> from a basic mathematical issue Rannala (2002); Ponciano et al. (2012).

Consider a simple example of the concept in which we want to determine the 155 parameter values for x and y. For each value of x in equation 0.1 below, we can find a y 156 that satisfies this equation - and there are an infinite number of equally likely possibilities. 157 It is only when we add more information (in the form of equation 0.2) that we can 158 determine the unique pair of values for x and y. Put simply, a solution can be found only if 159 you possess at least the same number of equations as unknowns. In this case the 160 unidentifiability is caused by overparameterization - there is an excess of parameters such 161 that the model cannot estimate the values of any of them. 162

$$2x + y = 7 \tag{0.1}$$

163

174

$$3x + 2y = 12$$
 (0.2)

Though the LTT is generated through the use of many different observations and 164 elements (DNA, fossils for time-calibration, extant species sampling) it is represented by a 165 single curve made up of one observation at any given point in time that represents the 166 number of lineages in a clade (Fig. 1). Fitting a model to an LTT is like fitting two 167 parameters ( $\lambda$  and  $\mu$ ) for the slope ( $\lambda - \mu$ ), which gives you only one value. This problem 168 has been highlighted previously (Nee, 2006), where  $\lambda - \mu$  is the net diversification rate. If 169 we try to estimate  $\lambda$  and  $\mu$  separately we find it very difficult (Fig. 2a,b) but we are much 170 more precise when estimating  $\lambda - \mu$  (Fig. 2c). Estimates of  $\lambda$  and  $\mu$  are highly correlated 171 (see MCMC analyses, Fig. 2d) and we find a flat surface in the likelihood where different 172 pairs of values for  $\lambda$  and  $\mu$  are equally likely (Fig. 2e), signifying unidentifiability. 173

This issue of unidentifiability means that for any  $\lambda$  there exists a  $\mu$  that yields the

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

9

same dLTT. However, as LP show, if the rates vary over time, it is possible to choose 175 almost any function for  $\lambda(t)$ , and obtain a function  $\mu(t)$  that produces the same dLTT. So, 176 LP go beyond the classical unidentifiability issues for the parameters of a given model (as 177 in the example above) and extend it to the space of models itself. For example, using a 178 very large angiosperm phylogenetic tree (Smith and Brown, 2018), LP showed that the 179 observed LTT is congruent with two opposing scenarios (Fig. 2 in LP): either a continuous 180 increase or a continuous decline in both speciation and extinction rates through time 181 (though the resulting diversification rates of these two scenarios are very similar). 182 Therefore, if we observe a rapid increase in the number of lineages in our LTT (assuming 183 complete sampling) we cannot determine if it was caused by a decrease in extinction rate, 184 or an increase in speciation rate. If we want to use models to explain LTTs then our 185 variables are inadequate and we must look towards other possible solutions. 186

187

# Pulled rates and their interpretation

A consequence of using extant timetrees is that LTT plots will likely underestimate 188 the number of lineages at any given time because our trees are missing species (Silvestro 189 et al., 2018). Species can be missing for two reasons: (1) they went extinct or (2) they were 190 not sampled. However, these two factors will have differing effects on the LTT and our 191 estimates of diversification rates. The idea that extinction can affect estimates of diversity 192 when using data from only extant species may seem counter-intuitive, but a marked effect 193 has been shown (Nee, 2006). Extinction must occur in the past. Lineages that originated 194 recently have had less time to go extinct (Nee et al., 1994; Ricklefs, 2007), so the effect of 195 extinction on our estimates using only extant species is reduced towards the present. This 196 leads to an increase in the rate of lineage accumulation towards the present, as the effect of 197 extinction decreases, which occurs even when rates are constant, as seen in Figure 3f. 198 Conversely, incomplete sampling of a group occurs up to the present day and more 199 strongly affects the recent history (Phillimore and Price, 2008), as the deeper nodes in the 200

10

### HELMSTETTER ET AL.

<sup>201</sup> phylogeny can be reconstructed with only a few species. The relative importance of <sup>202</sup> extinction and sampling completeness will influence whether the PSR departs from  $\lambda$  more <sup>203</sup> in the past or in the present. In summary, the presence of extinction will cause us to <sup>204</sup> underestimate speciation rate further in the past, because the number of extinct species <sup>205</sup> increases as we consider more time, while incomplete sampling will lead to underestimates <sup>206</sup> of speciation rates that are more recent (Ricklefs, 2007).

<sup>207</sup> LP bring these two ideas together and define the 'pulled' speciation rate (PSR or <sup>208</sup>  $\lambda_p$ ) i.e. the speciation rate modified by extinction and sampling fraction. At a given time, <sup>209</sup> PSR is the estimated speciation rate multiplied by 1 minus the probability that a lineage is <sup>210</sup> missing from the tree due to extinction or incomplete sampling, *E*. We will not go into <sup>211</sup> details regarding the calculation of *E* here, but further information can be found in <sup>212</sup> supplementary materials of LP. PSR is shown in the following equation

$$\lambda_p(\tau) = \lambda(\tau)(1 - E(\tau)) \tag{0.3}$$

So, if all species are in the tree and there is no extinction (i.e. the probability of missing lineages, or E, is 0) then the PSR is equal to the (un-pulled) speciation rate. Any increase in extinction rate or the number of unsampled lineages (i.e. E > 0) will cause PSR to drop, or be 'pulled', below speciation rate (Figs. 2,4). The lower the extinction rate and the higher the sampling fraction, the closer the estimates of PSRs will be to speciation rate. The effect of a shift in speciation rate on PSR is demonstrated in Figure 4.

Similarly, LP also present pulled diversification rate (PDR or  $r_p$ ). This parameter is similar to the net diversification rate  $(\lambda - \mu)$  but, as above, is modified by another term. This new term is the relative  $(\frac{1}{\lambda})$  rate of change in speciation rate over time  $(\frac{d\lambda}{d\tau})$ . This causes the pulled diversification rate to lag behind the unpulled rate. The PDR can be represented by the following equation :

$$r_p = \lambda - \mu + \frac{1}{\lambda} \cdot \frac{d\lambda}{d\tau} \tag{0.4}$$

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

11

The perspective in which time is viewed is critical when interpreting PDR. When 224 reading time backwards (using  $\tau$ ) diversification decreases from present ( $\tau = 0$ ) to the past 225 and PDR does so faster than expected (plus sign in equation (0.4)), going "too far" before 226 stabilizing to the ancestral value. The first part of this equation is the net diversification 227 rate,  $\lambda - \mu$ , which is thus corrected by a 'pull' corresponding to the rate of change of the 228 speciation rate. The 'pull' of PDR is actually a delay in the response of this parameter 229 when compared to diversification rate. This is in contrast to the 'pull' of PSR, which refers 230 to a reduction in the estimated value of PSR relative to  $\lambda$ . This has some unexpected 231 consequences if speciation rate rapidly increases as the PDR will rapidly decrease before 232 starting to increase (Fig. 5a) as speciation rate stabilises. We note that LP also defined a 233 pulled extinction rate, (PER or  $\mu_p$ ), which is similar to PDR and we do not go into details 234 about its calculation here (see LP, Louca et al. (2018) for further details). 235

The difference between the true diversification rate and an estimated PDR can be 236 likened to a race between an amateur and a professional race car driver. The professional 237 driver, representing the true diversification rate in our analogy, hits the apex of each 238 corner, going smoothly around a racetrack until the finish line. The amateur, representing 230 PDR, will eventually arrive at the finish line, but may exceed track limits a few times 240 when doing so because of their poor reactions. However, if the track is simply a straight 241 line both will perform equally well. This is because the PDR is equal to the diversification 242 rate  $(r = \lambda - \mu)$  whenever  $\lambda$  is constant in time  $(\frac{d\lambda}{d\tau} = 0)$ , but differs from r when  $\lambda$  varies 243 with time (see Technical considerations below for more details). 244

<sup>245</sup> LP show mathematically that there is only one of each pulled rates (PSR, PDR, <sup>246</sup> PER) per congruence class. These pulled rates are useful because a single congruence class <sup>247</sup> can be represented by a single set of pulled rates, rather than the infinite number of <sup>248</sup> possibilities of speciation and extinction rates. This is because pulled rates are compound <sup>249</sup> parameters that are, as shown in Louca et al. (2018), constructed from the LTT, using its <sup>250</sup> slope and the rate of change of the slope (curvature). LP show that these are related to  $\lambda$ 

12

### HELMSTETTER ET AL.

and  $\mu$ : each pulled rate depends on both speciation and extincton. The shape of the dLTT plot is fully determined by any two of the rates, and the remaining rate can be calculated from the other two.

With these new variables we can begin to ask questions such as: has diversification 254 been constant over time? Pulled rates can be estimated using many commonly used models 255 of diversification (Louca and Pennell, 2020). The PSR is the speciation rate one would get 256 by constraining extinction to be 0 and assuming complete species sampling. For PDR this 257 involves making the speciation rate time-independent (i.e. speciation rate is constant over 258 time). In summary, PSR provides information about how speciation rate changes over time 259 while taking into account past extinction and the proportion of lineages sampled. PDR 260 provides a slightly delayed estimate of r with extreme responses to rapid changes in  $\lambda$ . 261 While the PSR can be very different from the underlying speciation and extinction rates, 262 the PDR is close to the net diversification rate as long as speciation rate does not change 263 too rapidly. A scenario with multiple, extremely rapid changes over a short time scale (e.g. 264 Fig. 5d) may be biologically possible, but from an empirical point of view one would not fit 265 models with many changes in small time intervals. 266

267

## TECHNICAL CONSIDERATIONS

268

# How continuously can speciation and extinction rates vary?

In their approach LP consider speciation and extinction to be a continuous property 269 of a species, which can have instantaneous values at any time. When speciation and 270 extinction are modelled as continuous processes, a change in the rates is immediately 271 visible in the dLTT plot. In real trees, however, we can only observe discrete events: either 272 a branch splits, goes extinct, or it continues. When working with a large phylogenetic tree 273 and many species, the LTT is smooth and the slope and curvature, which are necessary for 274 the estimation of the pulled rates, can be reliably estimated. However, many studies 275 attempt to estimate diversification rates with relatively small numbers of species (e.g. 276

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

13

<1000). When the tree used is small, the LTT plot resembles an irregular staircase, and 277 although one can calculate the slope by using sufficiently large time intervals, the 278 calculation of the curvature will be less reliable. So, one should not forget that allowing for 279 continuous rate variation is less applicable in those cases where the number of species 280 considered is small. The consequence of this is that rate variation can be difficult to detect 281 using birth-death models, particularly in small phylogenetic trees (Condamine et al., 2019). 282 In practice (with a limited number of species), what precision is reasonable to aim 283 for in the characterization of variation in diversification rate? Considering step-wise 284 changes, a straightforward limit is the total number of speciation events in a tree with n285 extant species, which is in n-1 speciation events. Above this limit some changes will not 286 be detectable because the number of parameters will exceed the number of observations. 287 Another limit concerns the speed of change, which is applicable even if the number of 288 parameters is limited. For example, consider sinusoidal variations with frequency f, where 289 only one parameter is to be inferred. Noting that the number of speciation events per unit 290 time is  $\lambda n$ , this sets an upper limit to f that can be detectable with the data. This also 291 shows that rapid changes are more difficult to infer as n decreases and highlights the 292 limitations of inferring pulled rates (or any rates inferred using continuous approximation) 293 with small phylogenetic trees. A reasonable approach is thus to consider relatively simple, 294 but biologically meaningful, scenarios without implausibly rapid rate variations. If so,  $r_{p}$ 295 will be (often very) close to r. 296

### 297

# The delay in PDR is a result of the lag time between extinction and speciation

<sup>298</sup> Consider a simple case with no extinction ( $\mu = 0$ ) so that changes in r that only <sup>299</sup> come from changes in  $\lambda$ . If so,  $r = \lambda$  but  $r_p$  is not exactly  $\lambda$  because of temporal variations <sup>300</sup> in  $\lambda$  (the term  $\frac{1}{\lambda} \cdot \frac{d\lambda}{d\tau}$  in equation (0.4)). LP suggest that "the pulled diversification rate can <sup>301</sup> be interpreted as the effective net diversification rate if  $\lambda$  was time-independent". In our <sup>302</sup> example, this means replacing a scenario where  $\mu$  is constant (at 0) and  $\lambda$  varies with a

14

### HELMSTETTER ET AL.

scenario where  $\lambda$  is constant and  $\mu$  varies. LP explain this from the point of view of holding speciation rate constant and using extinction rate to account for changes in diversification rate. To get the same changes in the LTT using changes in  $\mu$  instead of changes in  $\lambda$ (which is kept constant by construct), a delay is needed to wait for species to arise, before they can go extinct. The difficulty with this, as initially noted by Nee et al. (1992), is that there is a slight delay between the effect of speciation and the effect of extinction.

As mentioned previously, lineages that originated more recently have had less time 309 to go extinct. In a constant birth-death process, this is only visible in recent history: the 310 slope of the LTT is  $r = \lambda - \mu$  during most of the past but increases to  $\lambda$  for very recent 311 times where the stationary behaviour has not yet been reached. However, this phenomenon 312 is not unique to very recent times - it will also occur whenever there is a change in 313 speciation rate. Ultimately, this is the cause of the difference between PDR and r. For 314 example, a massive increase in the number of lineages caused by a burst of speciation 315 means that many new lineages become available to go extinct over a short time period. 316 However, the rate of extinction is still determined by how many lineages there were prior 317 to the burst. As time continues, these numerous new species will begin to go extinct, 318 meaning that extinction rate will increase to "catch up" to speciation rate and reach a new 319 stationary point. This effect is stronger when  $\lambda$  varies rapidly (i.e. high  $\frac{1}{\lambda} \cdot \frac{d\lambda}{d\tau}$ ). Conversely, 320 speciation cannot occur in a lineage after it has gone extinct, so there is no similar lag 321 caused by changes in extinction rate. This is also why variation in extinction rate would 322 not cause PDR to deviate from r (Fig. 5b). 323

324

### DISCUSSION

A recent study by Morlon et al. (2020) presents an alternative point of view that opposes the conclusions in LP. They focus on how a hypothesis-based framework allows us to overcome many of the issues that are raised in LP. Indeed, we are limiting our set of models to be tested to only those that represent our hypotheses about the factors shaping

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

diversification in a given group. We are not often interested in determining the precise values of speciation and extinction rate but rather how different diversification scenarios summarised by models containing  $\lambda$  and  $\mu$  explain patterns in a phylogenetic tree. The criticisms put forward by Morlon et al. (2020) will stimulate important discussion about key points that must be considered when using diversification models. We extend this discussion by highlighting several key points that must be considered in addition.

335

# Uses and limitations of LTTs

LTTs are a simplistic way to visualize and summarise a time-calibrated 336 phylogenetic tree, ignoring information related to branch lengths, tree topology and extinct 337 species (Morlon et al., 2011). However, under the assumption of  $\lambda$  and  $\mu$  being shared 338 across all species LP showed that the LTT contains the complete information about the 339 underlying branching process (See also Lambert and Stadler (2013)). This simplicity 340 provided the opportunity for LP to show mathematically how LTTs can lead to 341 misinterpretation. However, these issues are not new to macroevolutionary biology. A 342 review by Nee (2006) clearly demonstrated how an LTT may change when extinction is 343 present alongside speciation (birth-death), as opposed to speciation alone (pure-birth), 344 summarising theory from previous work (Nee et al., 1992, 1994; Harvey et al., 1994). If the 345 growth of an extant timetree is represented as an LTT on a semi-log scale (i.e. lineage 346 number is logarithmic, time is not, see Fig. 1) we would expect the trend to be linear 347 under a pure birth process (with constant speciation and no extinction). If extinction is 348 introduced then the LTT would deviate from this linearity. When both rates are constant 349 and greater than 0, the curve is expected to be linear over most of its history, but as time 350 reaches the present the rate of lineage accumulation will increase (i.e. the LTT slope will 351 become steeper), as shown in Figure 3a. With no prior knowledge of the parameters, this 352 could be because of increasing speciation rate towards the present (Fig. 3b), instead of 353 decreasing effect of extinction (Fig. 3). It is important to keep in mind that we are dealing 354

16

### HELMSTETTER ET AL.

with a phylogenetic tree made up of entirely extant species. The unobserved branches of 355 species that went extinct (and are therefore not in the extant timetree) do not contribute 356 to the LTT, making the estimated lineage accumulation rate lower in the past (or 'pulling' 357 it down). Nee et al. (1994) highlighted this issue 20 years ago in the context of models 358 where diversification rates were constant over time and now LP have provided an 359 important extension of this idea to models that allow for rates to vary through time. The 360 well-known limitations of LTTs for inferring speciation and extinction rates have continued 361 to be addressed in other studies (Ricklefs, 2007; Vamosi et al., 2018; Rabosky and Lovette, 362 2008; Crisp and Cook, 2009) since Nee et al. (1994), most recently in LP. This begs the 363 question: what other previously proposed ideas remain hidden in the literature that could 364 be useful to macroevolutionary modelling? 365

366

# Diversification rates vary among clades

The conclusions of LP imply that we can test hypotheses about whether 367 diversification rates deviate from constancy over time using pulled rates. We would be 368 unable to pin this on changes in speciation or extinction rate, but would get a sense of how 369 variable diversification has been. This would be useful for testing whether diversification in 370 particular clades has remained constant or been subject to large shifts in diversification 371 (e.g. mass extinctions) but not when diversification rate has shifted in a subclade (e.g. due 372 to the evolution of a key innovation). The first use of pulled rates was in Louca et al. 373 (2018), where they studied bacterial diversification, stating "Our findings suggest that, 374 during the past 1 billion years, global bacterial speciation and extinction rates were not 375 substantially affected during the mass extinction events seen in eukaryotic fossil records." 376 This might suggest that nothing particularly extraordinary happened in the 377 macroevolutionary dynamics of bacteria in the last billion years. However, it is important 378 to note that the models used in Louca et al. (2018) (and Louca and Pennell (2020)) do not 379 allow rates to vary among clades. The rates estimated using such clade-homogeneous 380

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

models will correspond to the average rates over time in the entire study group, therefore 381 missing out on any variation among clades - for example any difference in diversification 382 rates between those species that use terrestrial vs marine environments (Louca et al., 383 2018). Given the importance of subclades in driving inferred diversification patterns (see 384 (Morlon et al., 2011; Rabosky, 2020)), this may mean that we miss out on important and 385 interesting dynamics when using pulled rates. Louca et al. (2018) touch on this point 386 themselves: "It is possible that diversification within individual bacterial clades may have 387 been influenced by eukaryotic radiations and extinctions, and that these cases are 388 overshadowed when considering all bacteria together." Given the diversity of life on Earth, 389 it is unrealistic to assume that major events would have had the same effect on all lineages 390 of a large, cosmopolitan clade, with vast amounts of genetic, morphological and ecological 391 variation. The same criticism could be levelled at LP's use of a large phylogenetic tree of 392 angiosperms (Smith and Brown, 2018) that contains more than 65,000 of the roughly 393 300,000 known species, ranging from small ephemeral plants like Arabidopsis thaliana to 394 gigantic, long-lived trees such as *Eucalyptus regnans*. Furthermore, a large amount of 395 research has shown that diversification rates have varied significantly among 396 flowering-plant clades (e.g. (O'Meara et al., 2016; Igea et al., 2017; Vamosi et al., 2018; 397 Onstein, 2019; Soltis et al., 2019; Zenil-Ferguson et al., 2019; Magallón et al., 2019)). 398 Fortunately, the assumption of homogeneous rates among clades is not common in 390

modern approaches. For instance, Bayesian Analysis of Macroevolutionary Mixtures
(BAMM) (Rabosky, 2014) is one of several methods (Alfaro et al., 2009; Morlon et al.,
2016; Höhna et al., 2016a; Maliet et al., 2019; Barido-Sottani et al., 2020) that relaxes the
assumption that all lineages share the same evolutionary rates at a given point in time
(Rabosky, 2017). This is a key difference from the models used by LP because it allows
lineages to differ in their rates of speciation and extinction. With BAMM, the entire
phylogeny could be described using a model similar to what is used in Louca and Pennell
(2020), or alternatively, it could be described using multiple processes that explain rates of

18

### HELMSTETTER ET AL.

diversification on different parts of the tree. These non-homogeneous diversification rates
may help to alleviate the unidentifiability issues demonstrated by LP by taking into
account additional information included in tree topology and branch lengths.

Another model commonly used to estimate and compare diversification rates among 411 clades is the Binary-State Speciation and Extinction (BiSSE) model (Maddison et al., 412 2007), part of a family of models known as the state-dependent models of diversification 413 (-SSE models (Ng and Smith, 2014; O'Meara and Beaulieu, 2016; Beaulieu and O'Meara, 414 2016; Caetano et al., 2018)). These models are extensions of the birth-death model that 415 also include information about character states of extant species. They estimate ancestral 416 states at each node of the phylogenetic tree, as well as rates of transition between 417 character states. LP state that the likelihood functions of SSE models are too complex to 418 be addressed in their manuscript, but suggest that the same problems they uncover 419 probably still apply. The increased complexity of likelihood calculations LP refer to is 420 because BiSSE makes use of the full tree topology (Maddison et al., 2007), rather than just 421 the timing of branching events as in the LTT (Nee et al., 1994). LP further suggest that it 422 remains unclear how the dependence on character states (which, if removed, collapses 423 equations in BiSSE to those shown in Nee et al. (1994)) affects the unidentifiability issue 424 they raise. In the original BiSSE paper (Maddison et al., 2007), two important and 425 relevant assumptions were made: 426

 $_{427}$  1. Sampling fraction is assumed to be 100%

428

2. Speciation, extinction and transition rate are constant per character state.

These may allow the BiSSE model to overcome (or pre-empt) some of the problems raised by LP. LP show that  $\lambda$  equals PSR when sampling fraction is 100% and  $\mu = 0$ . The first of these was assumed in the original BiSSE model, though it has since been relaxed (FitzJohn et al., 2009). Extinction can easily be set to 0 in these models, which satisfies the second BiSSE assumption and allows estimation of  $\lambda$ . Similarly, PDR equals r when  $\lambda$ is constant, also an assumption in BiSSE. As a result, it appears that BiSSE models

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

estimate rates of per-character state diversification that are similar or even analogous to 435 pulled rates and may therefore be identifiable. Researchers using SSE models have noticed 436 that estimated speciation and extinction rates are often correlated within states and 437 therefore usually report net diversification rates. They also are often aware that the 438 diversification rate might vary over time, and that the whole diversification history cannot 439 be entirely captured in one value (or two values). LP's results suggest, however, that this is 440 a rather robust approach: as the time-dependent variation in speciation and extinction 441 rates cannot be uncovered reliably, one should consider the estimated diversification rate 442 as a "pulled" rate. Nevertheless, we stress that the likelihood of time-dependent 443 diversification models (as in LP) is not the same as the likelihood of state-dependent 444 diversification models (-SSE models) and what is unidentifiable in the former does not say 445 anything about identifiability in the latter. 446

It is unclear how lineage-dependent rates would affect model congruence, and how 447 the additional information included when using models such as BiSSE and BAMM would 448 affect the unidentifiability issues. However, what is clear is that the issues raised in LP 449 cannot be readily applied to commonly used macroevolutionary approaches without 450 further work to show that criticisms related to LTT-based approaches are applicable to 451 these more complex models. Alternatively, even if unidentifiability issues remain in such 452 models they may not be relevant in the questions the models were built to answer, for 453 example those models that test for variation in diversification rates in association with 454 particular clades or traits. In cases like these, it is not the precise values of rates that are 455 important but instead whether rates in one group of lineages are higher than another. 456

Perhaps most importantly, this means that we should not forego building models
that estimate diversification rates because one, simplistic approach has problems, but
instead continue to improve them and build upon the work done in LP. A case in point is
the issue of null model choice when using SSE models raised by Rabosky and Goldberg
(2015). This criticism spurred on innovation that led to the development of models with

20

### HELMSTETTER ET AL.

<sup>462</sup> hidden states (Beaulieu and O'Meara, 2016), which are now present in various new
<sup>463</sup> incarnations, e.g. (Caetano et al., 2018; Herrera-Alsina et al., 2019), of the SSE approach.

464

# Pulled rates are difficult to interpret

LP compared the usefulness of pulled rates to effective population size  $(N_e)$  in 465 population genetics. Like  $N_e$ , diversification rates and pulled diversification rates are not 466 real - they do not exist in nature. Instead their role is to help us interpret complex 467 processes like diversification, and if they do not do this, then they lack utility.  $N_e$  can be 468 broadly defined as the number of breeding individuals in an idealised population (e.g. 469 constant size, random mating) that would be able to explain the summary statistics in an 470 observed population.  $N_e$  is fairly intuitive and will react to biological phenomena in 471 expected ways (e.g. under population structure (Whitlock and Barton, 1997) or 472 non-random mating (Caballero and Hill, 1992)). 473

LP state that the variables they introduce are "easily interpretable". Their 474 terminology, however, is not completely consistent nor coherent with more traditional uses, 475 which can cause confusion. Given that  $r = \lambda - \mu$  one might intuitively think that 476  $r_p = \lambda_p - \mu_p$  but this is not the case - pulled rates are simply different ways of summarising 477 congruence classes and each one is calculated using both speciation and extinction rates. 478 PSR is reasonably intuitive, though given that extinction is also included it is more similar 479 to a diversification rate than a speciation rate. Indeed, the PSR is defined as the slope of 480 the LTT plot (Louca et al., 2018) (see Fig. 3f, 4f), which corresponds to the diversification 481 rate in times far enough before present, and, in the case all extant species are included, to 482 the speciation rate at present (Nee et al., 1992). 483

<sup>434</sup> PDR, however, is much more difficult to interpret, perhaps initially because the <sup>435</sup> 'pull' of PSR is not the same as the 'pull' of PDR. Whereas PSR decreases in value <sup>436</sup> relative to  $\lambda$ , PDR is delayed in time relative to r (Fig. 5) and could better be termed as <sup>437</sup> 'delayed' rather than 'pulled'. We simulated a variety of diversification scenarios from

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

simple to more complex (Fig. 5) and show that PDR and r are similar in each case. However, PDR is not as intuitive as r or  $N_e$  - for example, drastic increases in r can lead to sharp decreases in PDR (Fig. 5a). The inverted pattern PDR presents in this case would make it challenging to present in a clear and concise way. Given the added difficulty of its interpretation we question whether PDR provides us with a more useful estimate of the process of diversification than an estimate of r.

However, compared to other pulled rates, PDR could be especially useful, not as an 494 effective parameter, like  $N_e$ , but as a reasonable approximation of the true r. Indeed, we 495 noted above that when shifts in  $\lambda$  are not too strong nor too rapid, PDR is close to r (Fig. 49F 4). Under non-parametric scenarios, trying to biologically interpret fine-grain variations in 497 PDR would certainly lead to spurious conclusions. However, changes in PDR at a large 498 scale are good proxies for large scale variation in r. This is clearly illustrated in Figure 4a 499 where the main trend of the PDR is a recent increase in diversification, and in Figure 4d 500 where the main trend is the stability of diversification. 501

<sup>502</sup> Pulled rates can be estimated using only the shape of the LTT plot, without any <sup>503</sup> further information, i.e. they are non-parametric estimates that do not suffer from the <sup>504</sup> unidentifiability problems outlined previously. However, they cannot be directly <sup>505</sup> interpreted in biologically meaningful terms; to estimate rates that are meaningful (e.g.  $\lambda$ , <sup>506</sup>  $\mu \& r$ ), one needs to make further assumptions such as constant rates of speciation and <sup>507</sup> extinction over time.

508

# On the use of models

The discussion sparked by Louca and Pennell (2020) highlights an important issue: evolutionary biologists should be interested in the actual history of diversification of the clades they study, and the framework developed by Louca et al. (2018) shows how to do this using the shape of the LTT plot, without making strong assumptions about past speciation and extinction rates. This is classical knowledge, as we have argued; the slope of

22

### HELMSTETTER ET AL.

the LTT plot and variations therein contain information about the diversification history 514 of the clade. Much of the debate, however, focuses on the ability to recover a "true" history 515 of diversification. Indeed, the goal of a scientific study should be to find out what really 516 happened, but it becomes confusing if one considers a simulated birth-death process as the 517 "true" history. This birth-death process is determined by two parameters ( $\lambda$  and  $\mu$ ) that 518 can vary over time. These parameters are supposed to correspond to the rate that a lineage 519 splits into two lineages, or goes extinct. In reality, however, a species doesn't have a 520 speciation and an extinction rate in the same way it has a geographic distribution and a 521 population size. These rates only make sense when they are averaged over a number of 522 species and a certain amount of evolutionary time. That is, they are descriptive statistics 523 summarizing much more complex processes that are playing at the level of species, and 524 that would eventually lead to speciation or extinction. Louca and Pennell (2020) 525 convincingly show that one cannot estimate these statistics reliably from LTT plots, and 526 propose statistics that can be estimated more reliably. That these alternative statistics do 527 not exactly correspond to the parameters of the naive birth-death process is not a 528 problem; the birth-death process is only a model of diversification, and not the truth about 520 diversification itself. The framework built by Louca et al. (2018) and LP allows us to use 530 the LTT to test whether the diversification rate was constant or not. If a researcher wants 531 to know how speciation and extinction actually changed to give rise to this diversification 532 history, they will have to use other methods. 533

534

# CONCLUSION

Louca and Pennell (2020) have pointed out key issues with how we approach macroevolutionary modelling, namely the inability to distinguish historical diversification scenarios under certain circumstances. Their formalization of the unidentifiability issues in LTT-based models is an important step forward that provides us with the mathematical tools to study the associated issues further. LP highlights the avenues we must consider

### EXPLAINING AND EVALUATING PULLED DIVERSIFICATION RATES

and develop upon to ensure we do not make similar mistakes in the future. Whether 540 variations in diversification rate are due to changes in speciation or extinction is certainly 541 an interesting avenue of research, but LP have shown that exploring this would require 542 much more than just fitting a model with speciation and extinction rates to an LTT. 543 Indeed, more recent diversification models go beyond this by making use of additional 544 information that the LTT lacks. Awareness and consideration of potential unidentifiability 545 issues is important for macroevolutionary biologists going forward when they employ such 546 models of diversification. However, it is important to note that LP does not show that 547 speciation and extinction cannot be estimated with evolutionary trees (Pagel, 2020). 548 Instead, they show that when using extant timetrees with a single, LTT-based approach, 549 unidentifiability issues are encountered in the estimation of speciation and extinction rates, 550 and that these problems can be circumvented by making use of pulled rates. Further work 55 is needed to identify the extent to which the issues raised in LP apply to the more complex 552 models of diversification used today. In the meantime it is important that the field 553 continues to grow by using and building upon modern macroevolutionary methods, albeit 554 with a critical eye. 555

24

# HELMSTETTER ET AL.

### 556

# Acknowledgements

This article was initially conceived from discussions held with the Macroevolution
in Montpellier (MiM) group. We thank Stilianos Louca for comments on an earlier version
of this manuscript. We also thank Marcos Mendez, Sally Otto, Dan Schoen, Jürg
Schönenberger and the rest of the DiveRS group for comments and discussion throughout.
TLPC is supported by funding from the European Research Council (ERC) under the
European Union's Horizon 2020 research and innovation program (grant agreement No.
865787).

564

# SUPPLEMENTARY MATERIAL

565 Code associated with this manuscript is available from

http:/github.com/ajhelmstetter/pulled\_rates

### REFERENCES

### References

- Alfaro, M. E., F. Santini, C. Brock, H. Alamillo, A. Dornburg, D. L. Rabosky,
- G. Carnevale, and L. J. Harmon, 2009. Nine exceptional radiations plus high turnover
- explain species diversity in jawed vertebrates. Proceedings of the National Academy of
- Sciences 106:13410–13414. Publisher: National Academy of Sciences Section: Biological

572 Sciences.

- <sup>573</sup> Barido-Sottani, J., T. G. Vaughan, and T. Stadler, 2020. A Multitype Birth–Death Model
- <sup>574</sup> for Bayesian Inference of Lineage-Specific Birth and Death Rates. Systematic Biology
- <sup>575</sup> 69:973–986. Publisher: Oxford Academic.
- <sup>576</sup> Beaulieu, J. M. and M. J. Donoghue, 2013. Fruit Evolution and Diversification in
- <sup>577</sup> Campanulid Angiosperms. Evolution 67:3132–3144. \_eprint:
- https://onlinelibrary.wiley.com/doi/pdf/10.1111/evo.12180.
- <sup>579</sup> Beaulieu, J. M. and B. C. O'Meara, 2016. Detecting Hidden Diversification Shifts in
- <sup>550</sup> Models of Trait-Dependent Speciation and Extinction. Systematic Biology 65:583–601.
- <sup>581</sup> Caballero, A. and W. G. Hill, 1992. Effective size of nonrandom mating populations.
- Genetics 130:909–916. Publisher: Genetics Section: INVESTIGATIONS.
- Caetano, D. S., B. C. O'Meara, and J. M. Beaulieu, 2018. Hidden state models improve
  state-dependent diversification approaches, including biogeographical models: HMM
- AND THE ADEQUACY OF SSE MODELS. Evolution 72:2308–2324.
- <sup>586</sup> Condamine, F. L., N. S. Nagalingum, C. R. Marshall, and H. Morlon, 2015. Origin and
  <sup>587</sup> diversification of living cycads: a cautionary tale on the impact of the branching process
  <sup>588</sup> prior in Bayesian molecular dating. BMC Evolutionary Biology 15:65.
- <sup>589</sup> Condamine, F. L., J. Rolland, and H. Morlon, 2019. Assessing the causes of diversification
- <sup>590</sup> slowdowns: temperature-dependent and diversity-dependent models receive equivalent

0	0
•,	h
	U)

### REFERENCES

- <sup>591</sup> support. Ecology Letters 22:1900–1912. \_eprint:
- <sup>592</sup> https://onlinelibrary.wiley.com/doi/pdf/10.1111/ele.13382.
- <sup>593</sup> Crisp, M. D. and L. G. Cook, 2009. Explosive Radiation or Cryptic Mass Extinction?
- <sup>594</sup> Interpreting Signatures in Molecular Phylogenies. Evolution 63:2257–2265. \_eprint:

<sup>595</sup> https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.1558-5646.2009.00728.x.

- <sup>596</sup> FitzJohn, R. G., W. P. Maddison, and S. P. Otto, 2009. Estimating Trait-Dependent
- <sup>597</sup> Speciation and Extinction Rates from Incompletely Resolved Phylogenies. Systematic

<sup>598</sup> Biology 58:595–611.

- Harvey, P. H., R. M. May, and S. Nee, 1994. Phylogenies Without Fossils. Evolution
  48:523–529. \_eprint:
- https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.1558-5646.1994.tb01341.x.

Herrera-Alsina, L., P. van Els, and R. S. Etienne, 2019. Detecting the Dependence of
 Diversification on Multiple Traits from Phylogenetic Trees and Trait Data. Systematic
 Biology 68:317–328.

Ho, L. S. T. and C. Ané, 2014. Intrinsic inference difficulties for trait evolution with
 Ornstein-Uhlenbeck models. Methods in Ecology and Evolution 5:1133–1146. \_eprint:
 https://besjournals.onlinelibrary.wiley.com/doi/pdf/10.1111/2041-210X.12285.

Höhna, S., M. J. Landis, T. A. Heath, B. Boussau, N. Lartillot, B. R. Moore, J. P.

Huelsenbeck, and F. Ronquist, 2016a. RevBayes: Bayesian Phylogenetic Inference Using
Graphical Models and an Interactive Model-Specification Language. Systematic Biology
65:726–736.

- <sup>612</sup> Höhna, S., M. R. May, and B. R. Moore, 2016b. TESS: an R package for efficiently
- simulating phylogenetic trees and performing Bayesian inference of lineage
- diversification rates. Bioinformatics 32:789–791. Publisher: Oxford Academic.

### REFERENCES

- Igea, J., E. F. Miller, A. S. T. Papadopulos, and A. J. Tanentzap, 2017. Seed size and its
   rate of evolution correlate with species diversification across angiosperms. PLOS Biology
   15:e2002792.
- <sup>618</sup> Igea, J. and A. J. Tanentzap, 2020. Angiosperm speciation cools down in the tropics.
- Ecology Letters 23:692–700. \_eprint:
- https://onlinelibrary.wiley.com/doi/pdf/10.1111/ele.13476.
- Jetz, W., G. H. Thomas, J. B. Joy, K. Hartmann, and A. O. Mooers, 2012. The global diversity of birds in space and time. Nature 491:444–448.
- Lagomarsino, L. P., F. L. Condamine, A. Antonelli, A. Mulch, and C. C. Davis, 2016. The
- abiotic and biotic drivers of rapid diversification in Andean bellflowers (Campanulaceae).
   New Phytologist 210:1430–1442.
- Lambert, A. and T. Stadler, 2013. Birth–death models and coalescent point processes: The
  shape and probability of reconstructed phylogenies. Theoretical Population Biology
  90:113–128.
- Louca, S. and M. W. Pennell, 2020. Extant timetrees are consistent with a myriad of
   diversification histories. Nature 580:502–505.
- Louca, S., P. M. Shih, M. W. Pennell, W. W. Fischer, L. W. Parfrey, and M. Doebeli,
   2018. Bacterial diversification through geological time. Nature Ecology & Evolution
   2:1458–1467.
- Maddison, W. P., P. E. Midford, and S. P. Otto, 2007. Estimating a Binary Character's
   Effect on Speciation and Extinction. Systematic Biology 56:701–710.
- Magallón, S., L. L. Sánchez-Reyes, and S. L. Gómez-Acevedo, 2019. Thirty clues to the
   exceptional diversification of flowering plants. Annals of Botany 123:491–503. Publisher:
   Oxford Academic.

28

### REFERENCES

- Maliet, O., F. Hartig, and H. Morlon, 2019. A model with many small shifts for estimating
   species-specific diversification rates. Nature Ecology & Evolution 3:1086–1092.
- McGuire, J., C. Witt, J. V. Remsen, A. Corl, D. Rabosky, D. Altshuler, and R. Dudley,
- <sup>642</sup> 2014. Molecular Phylogenetics and the Diversification of Hummingbirds. Current
  <sup>643</sup> Biology 24:910–916.
- <sup>644</sup> Morlon, H., F. Hartig, and S. Robin, 2020. Prior hypotheses or regularization allow
- inference of diversification histories from extant timetrees. bioRxiv P. 2020.07.03.185074.
- <sup>646</sup> Morlon, H., E. Lewitus, F. L. Condamine, M. Manceau, J. Clavel, and J. Drury, 2016.

RPANDA: an R package for macroevolutionary analyses on phylogenetic trees. Methods
 in Ecology and Evolution 7:589–597.

- Morlon, H., T. L. Parsons, and J. B. Plotkin, 2011. Reconciling molecular phylogenies with
   the fossil record. Proceedings of the National Academy of Sciences 108:16327–16332.
- Nee, S., 2006. Birth-Death Models in Macroevolution. Annual Review of Ecology,
   Evolution, and Systematics 37:1–17.
- <sup>653</sup> Nee, S., R. M. May, and P. H. Harvey, 1994. The Reconstructed Evolutionary Process.
   <sup>654</sup> Philosophical Transactions: Biological Sciences 344:305–311.
- Nee, S., A. O. Mooers, and P. H. Harvey, 1992. Tempo and mode of evolution revealed from
   molecular phylogenies. Proceedings of the National Academy of Sciences 89:8322–8326.
- Ng, J. and S. D. Smith, 2014. How traits shape trees: new approaches for detecting
   character state-dependent lineage diversification. Journal of Evolutionary Biology
   27:2035–2045.
- O'Meara, B. C. and J. M. Beaulieu, 2016. Past, future, and present of state-dependent
   models of diversification. American Journal of Botany 103:792–795.

- <sup>662</sup> O'Meara, B. C., S. D. Smith, W. S. Armbruster, L. D. Harder, C. R. Hardy, L. C.
- Hileman, L. Hufford, A. Litt, S. Magallón, S. A. Smith, P. F. Stevens, C. B. Fenster, and
- P. K. Diggle, 2016. Non-equilibrium dynamics and floral trait interactions shape extant
- angiosperm diversity. Proceedings of the Royal Society B: Biological Sciences
- <sup>666</sup> 283:20152304.
- <sup>667</sup> Onstein, R. E., 2019. Darwin's second 'abominable mystery': trait flexibility as the <sup>668</sup> innovation leading to angiosperm diversity. New Phytologist P. nph.16294.
- Pagel, M., 2020. Evolutionary trees can't reveal speciation and extinction rates. Nature
   580:461-462.
- Phillimore, A. B. and T. D. Price, 2008. Density-Dependent Cladogenesis in Birds. PLOS
   Biology 6:e71. Publisher: Public Library of Science.
- Ponciano, J. M., J. G. Burleigh, E. L. Braun, and M. L. Taper, 2012. Assessing Parameter
  Identifiability in Phylogenetic Models Using Data Cloning. Systematic Biology
  61:955–972.
- Pulido-Santacruz, P. and J. T. Weir, 2016. Extinction as a driver of avian latitudinal
   diversity gradients: EXTINCTION AS A DRIVER OF DIVERSITY GRADIENTS.
   Evolution 70:860–872.
- <sup>679</sup> Quintero, I. and W. Jetz, 2018. Global elevational diversity and diversification of birds.
- Nature 555:246–250. Number: 7695 Publisher: Nature Publishing Group.
- Rabosky, D. L., 2014. Automatic Detection of Key Innovations, Rate Shifts, and
- <sup>682</sup> Diversity-Dependence on Phylogenetic Trees. PLoS ONE 9:e89543.
- 683 —, 2017. Phylogenetic tests for evolutionary innovation: the problematic link between
- key innovations and exceptional diversification. Philosophical Transactions of the Royal
- <sup>685</sup> Society B: Biological Sciences 372:20160417.

### REFERENCES

- Journal of Biogeography 47:1207–1217.
- Rabosky, D. L., J. Chang, P. O. Title, P. F. Cowman, L. Sallan, M. Friedman,
- K. Kaschner, C. Garilao, T. J. Near, M. Coll, and M. E. Alfaro, 2018. An inverse

latitudinal gradient in speciation rate for marine fishes. Nature 559:392–395. Number:

<sup>691</sup> 7714 Publisher: Nature Publishing Group.

Rabosky, D. L. and E. E. Goldberg, 2015. Model Inadequacy and Mistaken Inferences of
 Trait-Dependent Speciation. Systematic Biology 64:340–355.

Rabosky, D. L. and I. J. Lovette, 2008. Explosive evolutionary radiations: decreasing

speciation or increasing extinction through time? Evolution; International Journal of
 Organic Evolution 62:1866–1875.

- <sup>697</sup> Rannala, B., 2002. Identifiability of parameters in MCMC Bayesian inference of phylogeny.
   <sup>698</sup> Systematic Biology 51:754–760.
- <sup>699</sup> Ricklefs, R. E., 2007. Estimating diversification rates from phylogenetic information.
- Trends in Ecology & Evolution 22:601-610.
- Rolland, J., F. L. Condamine, F. Jiguet, and H. Morlon, 2014. Faster Speciation and
   Reduced Extinction in the Tropics Contribute to the Mammalian Latitudinal Diversity
   Gradient. PLOS Biology 12:e1001775. Publisher: Public Library of Science.
- Sauquet, H., 2013. A practical guide to molecular dating. Comptes Rendus Palevol
  12:355–367.
- <sup>706</sup> Silvestro, D., R. C. M. Warnock, A. Gavryushkina, and T. Stadler, 2018. Closing the gap
- <sup>707</sup> between palaeontological and neontological speciation and extinction rate estimates.
- <sup>708</sup> Nature Communications 9:5237. Number: 1 Publisher: Nature Publishing Group.
- <sup>709</sup> Silvestro, D., G. Zizka, and K. Schulte, 2014. Disentangling the Effects of Key Innovations

- on the Diversification of Bromelioideae (bromeliaceae). Evolution 68:163–175. \_eprint:
   https://onlinelibrary.wiley.com/doi/pdf/10.1111/evo.12236.
- <sup>712</sup> Slater, G. J., S. A. Price, F. Santini, and M. E. Alfaro, 2010. Diversity versus disparity
- and the radiation of modern cetaceans. Proceedings of the Royal Society B: Biological
- <sup>714</sup> Sciences 277:3097–3104. Publisher: Royal Society.
- <sup>715</sup> Smith, S. A. and J. W. Brown, 2018. Constructing a broadly inclusive seed plant
- <sup>716</sup> phylogeny. American Journal of Botany 105:302–314. \_eprint:
- <sup>717</sup> https://bsapubs.onlinelibrary.wiley.com/doi/pdf/10.1002/ajb2.1019.
- Soltis, P. S., R. A. Folk, and D. E. Soltis, 2019. Darwin review: angiosperm phylogeny and
   evolutionary radiations. Proceedings of the Royal Society B: Biological Sciences
   286:20190099.
- <sup>721</sup> Upham, N. S., J. A. Esselstyn, and W. Jetz, 2019. Inferring the mammal tree: Species-level
   <sup>722</sup> sets of phylogenies for questions in ecology, evolution, and conservation. PLOS Biology
   <sup>723</sup> 17:e3000494.
- <sup>724</sup> Vamosi, J. C., S. Magallón, I. Mayrose, S. P. Otto, and H. Sauquet, 2018.
- <sup>725</sup> Macroevolutionary Patterns of Flowering Plant Speciation and Extinction. Annual
- Review of Plant Biology 69:685–706.
- <sup>727</sup> Whitlock, M. C. and N. H. Barton, 1997. The Effective Size of a Subdivided Population.
- <sup>728</sup> Genetics 146:427–441. Publisher: Genetics Section: Investigations.
- <sup>729</sup> Zenil-Ferguson, R., J. G. Burleigh, W. A. Freyman, B. Igić, I. Mayrose, and E. E.
- <sup>730</sup> Goldberg, 2019. Interaction among ploidy, breeding system and lineage diversification.
- <sup>731</sup> New Phytologist 224:1252–1265.





Fig. 1. (a) The first example of a lineage through time plot (LTT), taken from Nee et al. (1992) and based on a phylogenetic tree of birds. On the y-axis is the number of lineages (log scale) and the x-axis is time since origin (present on the right hand side of the graph). "Each point corresponds to a change in the number of lineages. Line A, the pattern of origination of all 122 lineages; line B, same as A, but without the Passeri (line C) and the Ciconliformes (line D). Line B has been shifted downward to aid visual comparison. The diversification rate is quantified by the steepness of the slope." In this panel, time is displayed going forward, from past to present as time since origin (t). (b) Three LTTs from modern phylogenetic trees of Campanulids (Beaulieu and Donoghue, 2013), Cycadales (Condamine et al., 2015) and Cetacea (Slater et al., 2010). In this panel, time is read backwards, from present to past as an age  $(\tau)$ .



Fig. 2. An example of unidentifiability issues encountered when trying to estimate values of two parameters ( $\lambda \& \mu$ ) for the slope  $\lambda - \mu$ . We used a Bayesian Monte-Carlo Markov Chain approach to try to estimate the values of  $\lambda$  and  $\mu$  separately. We ran a chain for 5000 generations, sampling each generation. The traces for (a)  $\lambda$  and (b)  $\mu$  show a great deal of uncertainty in the parameter estimates compared to the estimates for (c)  $\lambda - \mu$ . True values are shown as black dashed lines in panels (a-c) and as orange circles in panels (d-e). We plotted  $\lambda$  against  $\mu$  and found that they two parameters were highly correlated (d). When then calculated the relative likelihood over a range of parameters values and found a flat ridge in the likelihood where different pairs of values for  $\lambda \& \mu$  are equally likely - or unidentifiability (e).



Fig. 3. A simple example of the relationship between constant diversification rates and corresponding pulled rates. Panel (a) shows values of speciation rate, extinction rate and diversification rate over time. An additional axis, at the top of panel (a) shows time going forward (t). Panel (b) shows how in the past, pulled speciation rate (PSR) is identical to the diversification rate (if sampling fraction = 1) while closer to the present PSR approaches speciation rate. The following two panels compare (c) r & pulled diversification rate (PDR) and (d) compares  $\mu \&$  pulled extinction rate (PER). In these two cases the pulled rates are identical to the un-unpulled rates. Panel (e) shows 50 LTT plots (grey lines) simulated with the parameters used in panels (a-d) and the mean LTT (black line). Panel (f) shows the slopes of the LTTs in panel (e) over time, matching PSR and depicting the expected increase towards the present caused by the lack of effect of extinction - species do not have enough time to go extinct towards the present. An interactive version of this plot, in which parameters can be modified, can be found at https://ajhelmstetter.shinyapps.io/pulled\_rates/.

### REFERENCES



Fig. 4. A slightly more complex example of the relationship between constant diversification rates and corresponding pulled rates where a single shift - an increase in speciation rate - has taken place. Panel (a) shows values of speciation rate, extinction rate and diversification rate over time. An additional axis, at the top of panel (a) shows time going forward (t). Panel (b) shows the gradual change in PSR during the shift in speciation rate. Panel (c) compares r and pulled diversification rate (PDR). The sudden increase in speciation rate causes PDR to decrease suddenly before recovering to the new diversification rate. Panel (d) compares extinction rate and pulled extinction rate (PER) and shows an inverse pattern to panel (c). Panel (e) shows 50 LTT plots (grey lines) simulated with the parameters used in panels (a-d) and the mean LTT (black line). Panel (f) shows the slopes of the LTTs in panel (e) over time, matching PSR and again depicting the expected increase towards the present caused by the lack of effect of extinction.



Fig. 5. Comparison of diversification rate and pulled diversification rate (PDR) under three simulated diversification scenarios that are commonly investigated (a-c) and a final, more complex scenario. Speciation rate, pulled speciation rate and extinction rate are also shown. Panel (a) shows a recent radiation where diversification rate and speciation rate sharply increase towards the present. An additional axis, at the top of panel (a) shows time going forward (t). Panel (b) shows a mass extinction event at 40 Ma in which extinction briefly but rapidly increases and then falls back to previous levels. Panel (c) shows a gradual increase in species turnover rate (both speciation and extinction increase slowly over time). Panel (d) shows a scenario where speciation and extinction rates are similar to each other but are in rapid fluctuation over time. This results in a relatively constant diversification rate and a rapidly fluctuating PDR that remains close to diversification rate (r).