

1 **Genetic composition and evolution of the prevalent *Mycobacterium tuberculosis* lineages**  
2 **2 and 4 in the Chinese and Zhejiang Province populations**

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30

31 **Keywords**

32 *Mycobacterium tuberculosis*; Whole-genome sequencing; Phylogenetic analysis; Bayesian

33 evolutionary analysis; Transmission

34 **Abstract**

35 The causative agent of tuberculosis (TB) comprises seven human-adapted lineages. Human  
36 movements and host genetics are crucial to TB dissemination. We analyzed whole-genome  
37 sequencing data for a countrywide collection of 1154 isolates and a provincial collection of  
38 1296 isolates, constructed the best-scoring maximum likelihood phylogenetic tree, conducted  
39 Bayesian evolutionary analysis to compute the most recent common ancestors of lineages 2  
40 and 4, and assessed the antigenic diversity in human T cell epitopes by calculating pairwise  
41 dN/dS ratios. Of the 1296 Zhejiang isolates, 964 (74.38%) belonged to lineage 2 and 332  
42 (25.62%) belonged to lineage 4. L2.2 is the most ancient sub-lineage in Zhejiang, first  
43 appearing approximately 6897 years ago (95% HDI: 6513-7298). L4.4 is the most modern  
44 sub-lineage, first appearing approximately 2217 years ago (95% HDI: 1864-2581). The dN/dS  
45 ratios revealed that the epitope and non-epitope regions of lineage 2 strains were significantly  
46 ( $P<0.001$ ) more conserved than those of lineage 4. An increase in the frequency of lineage 4  
47 may reflect its successful transmission over the last 20 years. The recent common ancestors  
48 and transmission routes of the sub-lineages are related to the entry of humans into China and  
49 Zhejiang Province.

## 50 **Introduction**

51 The causative agent of tuberculosis (TB), *Mycobacterium tuberculosis* (Mtb), is an obligate  
52 pathogen that comprises seven human-adapted lineages (Coscolla and Gagneux 2014). Mtb is  
53 one of the most successful human pathogens, having killed an estimated 1 billion people over  
54 the last 200 years (Gagneux 2012). In 2017, TB caused an estimated 1.6 million deaths,  
55 including 300,000 deaths in the HIV-positive population. Sustained reductions in disease  
56 incidence of up to 20% per year are required to meet the targets set out in the “WHO END TB”  
57 Strategy (Glaziou et al. 2013; Leung et al. 2018). However, current estimates suggest that the  
58 incidence is decreasing at a rate of only 1.5% per annum (WHO 2018).

59

60 It is well known that the social characteristics of human populations (Lonroth et al. 2009),  
61 host genetics (Gagneux 2012) and human interventions (e.g., the implementation of disease  
62 control programs) are crucial determinants of TB. Accumulating evidence indicates that  
63 human migrations and activities influence the population structure of Mtb (Nathanson et al.  
64 2010). As such, human-adapted Mtb lineages have shown a strong phylogeographic  
65 population structure in which different lineages are associated with distinct geographic  
66 regions (Filliol et al. 2006; Hershberg et al. 2008; Reed et al. 2009). A number of studies have  
67 found differences in virulence and immunogenicity among the seven lineages (Coscolla and  
68 Gagneux 2010; Parwati et al. 2010). Interestingly, the extent of their geographic distribution  
69 differs markedly, with some exhibiting a global distribution while others showing a strong  
70 geographic restriction. Widely distributed Mtb is more likely to spread. Therefore, identifying  
71 the predominant lineages in various regions can provide critical insight into the successful  
72 transmission and development of TB.

73

74 The human-adapted members of *Mycobacterium tuberculosis complex* (MTBC) can be  
75 classified into seven independent lineages (Coscolla and Gagneux 2014), all of which have  
76 humans as their only known host. Lineages 2 and 4 appear to be more virulent and  
77 transmissible on average than the other Mtb lineages (Coscolla and Gagneux 2014; Liu et al.  
78 2018). However, this is not always true, and there is a great deal of variation among the  
79 lineage 4 strains. Lineage 2, which is also known as the East-Asian lineage due to its

80 predominance in East Asia, includes the Beijing family of strains that have received particular  
81 attention because they are associated with drug resistance and virulence and are considered to  
82 be a ‘successful’ lineage (Nathanson et al. 2010). Molecular epidemiological studies have  
83 reported considerable variation in the transmission success of lineage 2 strains. For example,  
84 several studies using whole-genome sequencing (WGS) have demonstrated that lineage 4 can  
85 be further subdivided into several sub-lineages (Coll et al. 2014; Stucki et al. 2016). These  
86 sub-lineages partially reflected strain families that had been previously defined based on  
87 various genotyping techniques. The increase in human population density during the  
88 agricultural and industrial revolutions would then have selected for increased virulence in  
89 some Mtb lineages.

90

91 To understand the phenotypic consequence of between and within lineage diversity, one can  
92 look at its evolutionary conservation of protein residue (Shih et al. 2012), as between-lineage  
93 differences in the sharing of mutations may impact their phenotypes. Between-strain  
94 comparison of genomic regions encoding proteins that are recognized by human T cells has  
95 revealed that T cell epitopes are among the most conserved regions in the Mtb genomes; they  
96 exhibit lower frequencies of amino acid changes compared to essential genes and non-epitope  
97 antigen regions (Coscolla et al. 2015; Yrueala et al. 2016).

98

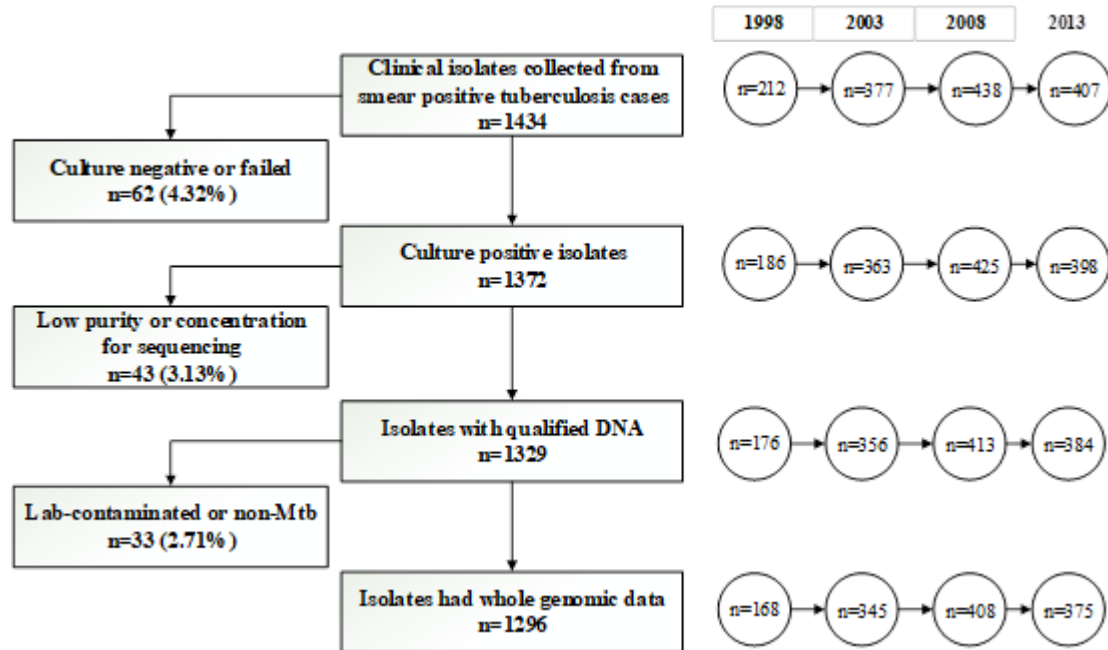
99 It remains unclear when epidemic forms of TB first arose in China, how the strains  
100 transmitted successfully within China, and what course these epidemics may have followed  
101 throughout Chinese history. In the present study, we reconstruct the phylogenomic history of  
102 epidemic TB in eastern China and use it to examine how the intersection of Mtb phylogeny,  
103 geography and demography has contribute to the widespread dispersal of TB in this country.  
104 We examine the SNPs (single nucleotide polymorphisms) shared by the predominant lineages  
105 in China as a means to explore the common genetic characteristics that have contributed to its  
106 wide transmission. Our analyses provide insights into the genomic polymorphism of the  
107 predominant TB lineages and the genetic basis for the widespread dissemination capacity and  
108 virulence of this important human disease.

109

## 110 Results

### 111 *Collection and genomic sequencing of 1296 Mtb isolates from Zhejiang Province*

112 From 1998 to 2013, a total of 1434 clinical isolates were collected; of them, 1372 (95.67%)  
113 were culture-positive and 1329 (96.87%) met our predefined criteria for the sequencing purity  
114 and concentration. Thirty-three isolates that were cross-contaminated or did not represent Mtb  
115 were excluded. In total, 1296 isolates were included for our analysis (Figure 1).



116 Figure 1. Clinical isolates collected in Zhejiang, 1998-2013.

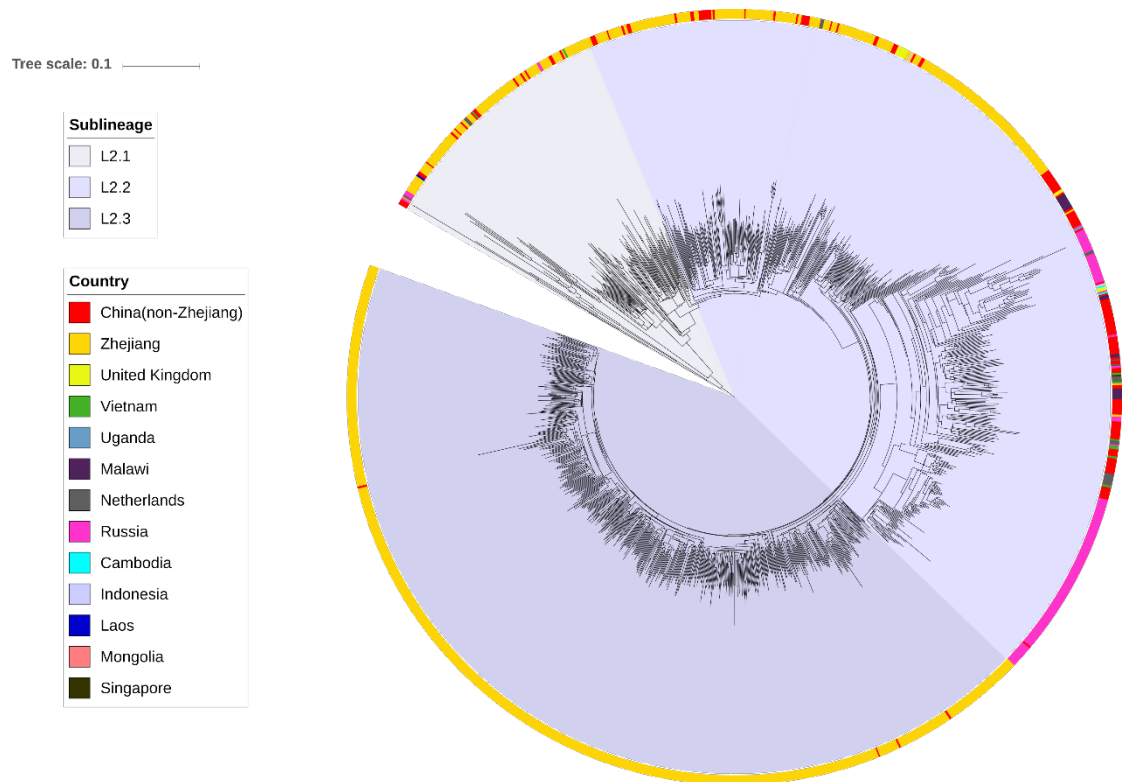
117

### 118 *Phylogenetic characteristics of the lineage 2 and lineage 4 strains*

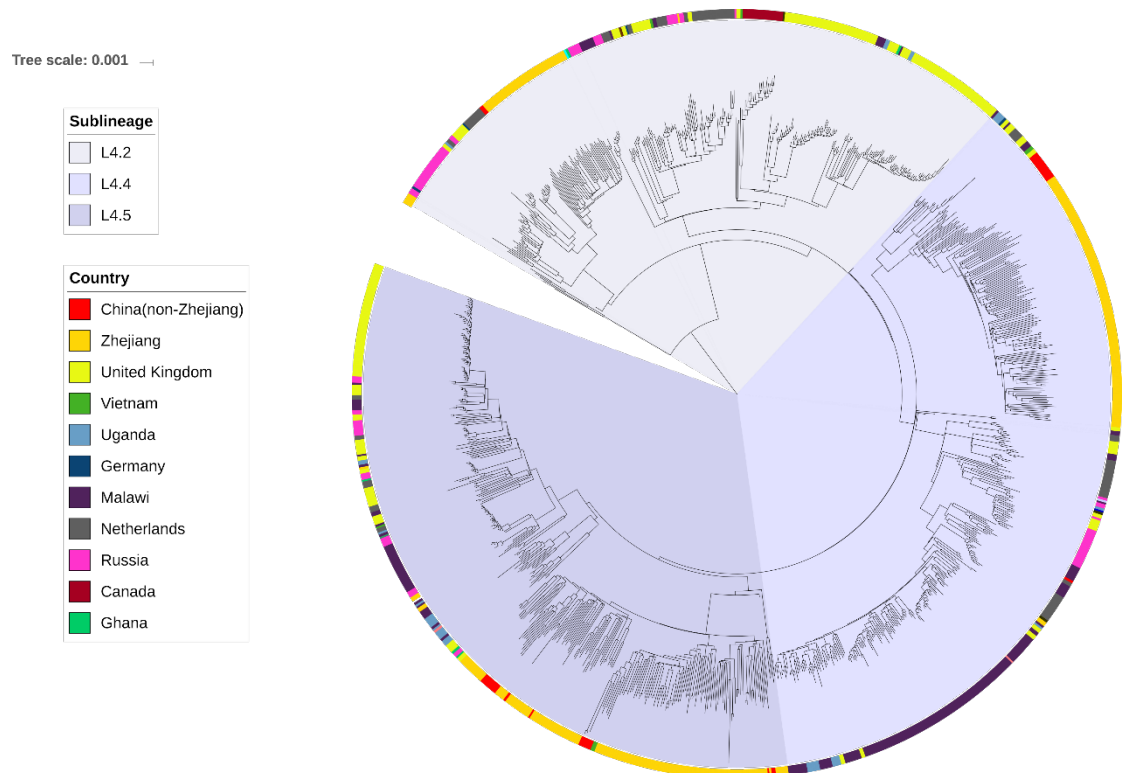
119 WGS data were obtained from the 1296 Mtb isolates from Zhejiang Province and downloaded  
120 for 1154 previously studied isolates that were obtained from around the world and represented  
121 the six main previously-defined phylogeographic lineages of Mtb. These data were used to  
122 construct phylogenetic trees (Figure 2). Of the 1296 Zhejiang isolates, 964 (74.38%) belonged  
123 to lineage 2 and 332 (25.62%) belonged to lineage 4. We next selected a subset of lineage 4  
124 clinical isolates (n=771) from 17 countries and a subset of lineage 2 clinical isolates (n=383)  
125 from 12 countries. To determine the placement of the Zhejiang strains along the evolutionary  
126 path of these lineages, we reconstructed maximum-likelihood phylogenies for lineages 2 and  
127 4. The phylogenetic trees showed that lineage 2 comprises three sub-lineages, L2.1 (10.17%),  
128 L2.2 (32.57%) and L2.3 (57.26%); among them, L2.3 (552 strains) was the predominant

129 sub-lineage in Zhejiang Province, accounting for 42.59% of the total strains. Lineage 4 was  
130 found to comprise three sub-lineages, L4.2 (18.07%), L4.4 (38.56%) and L4.5 (43.37%).

131 **a**



132 **b**



133 **Figure 2. Bayesian phylogeny of the Zhejiang *M. tuberculosis* isolates and 1154 globally**  
134 **distributed publicly available genomes for (a) lineage 2 and (b) lineage 4. Scale bar indicates the**

135 regions of origin. The *M. tuberculosis* sub-lineages, L2.1, L2.2, L2.3, L4.2, L4.4 and L4.5, are  
136 indicated respectively.

137

138 The distributions of sub-lineages varied between the administrative/geographic regions of  
139 Zhejiang Province (East, North, West, South and Middle). The lineage 4 types accounted for  
140 the largest proportion in Southern Zhejiang (40.10%), while Western Zhejiang had the lowest  
141 proportion (19.57%) of these lineages. Analysis of spatial-temporal trends in the distributions  
142 of lineage 2 and 4 isolates among the five districts indicated that the proportion of lineage 4  
143 isolates decreased in Northern and Southern Zhejiang over the 16-year study period, whereas  
144 it increased in Western Zhejiang (Supplementary\_Fig\_S1.).

145

#### 146 *Phylogeographic evolution of the major sub-lineages*

147 Published phylogeographic studies have indicated an African origin for Mtb, suggesting that it  
148 was introduced to other continents via human migration (Hershberg et al. 2008; Comas et al.  
149 2013). To further explore the evolutionary relationship of these strains and their geographical  
150 distribution, we used Bayesian evolutionary analysis (Table 1, Figure 3) to predict the  
151 divergence time of the most recent common ancestors of four sub-lineages  
152 (Supplementary\_Fig\_S2.).

153

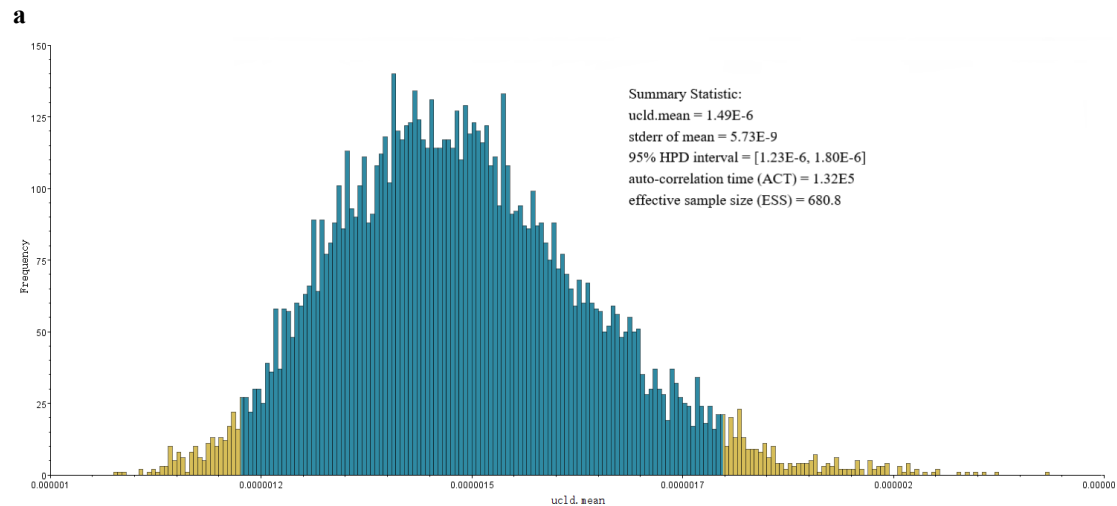
154 **Table 1. The most recent common ancestors of L2 and L4 sub-lineages in China**

Summary statistics	L2.2	L4.2	L4.4	L4.5
Mean (tMRCA)	10,763	8,530	7,800	7,446
SE of the mean	39.5	62.7	39.4	43.0
Median (tMRCA)	10,740	8,499	7,770	7,435
Geometric mean	10,711	8,456	7,747	7,406
95% HDI	[8,729-12,836]	[6,378-10,804]	[6,064-9,572]	[5,900-8,901]
ESS	711.5	323.7	531.5	319.1

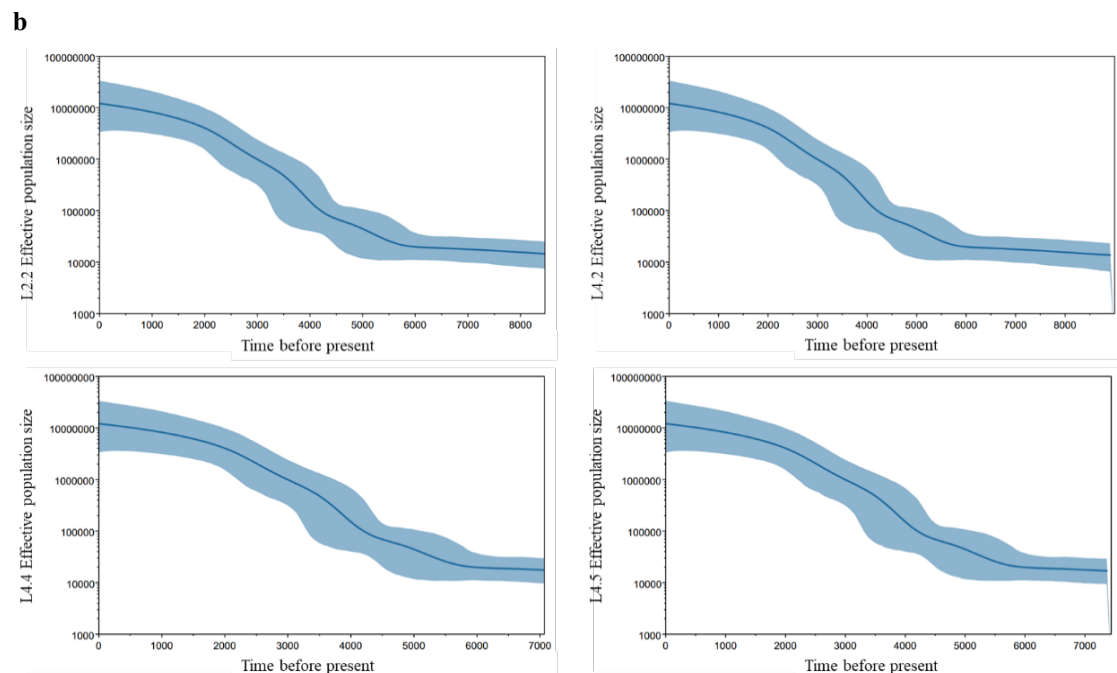
155 tMRCA: the most recent common ancestor; SE of the mean: standard error of the mean tMRCA; HDI:  
156 highest posterior density interval; ESS: effective sample size.



157



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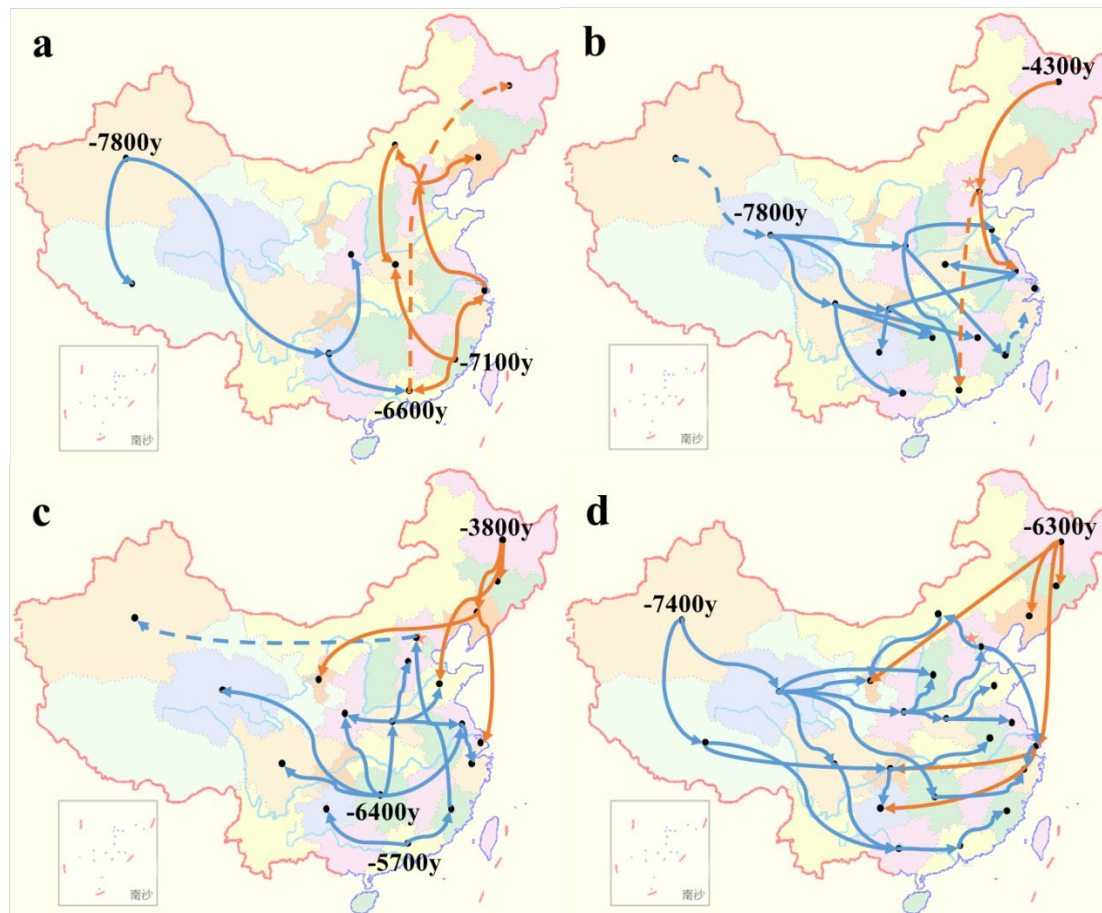
159 **Figure 3. Mutation rates and changes in sub-lineage diversity over time.** (a) The mutation rate was  
160 estimated using Beast. (b) Bayesian skyline plots indicating changes in the diversity of four  
161 sub-lineages over time. Shaded areas show the 95% HPD (high posterior density) intervals for the  
162 population-size estimations.

163

164 Our results revealed that L2.2 is the most ancient of the studied sub-lineages in China, with its  
165 tMRCA appearing around 10,763 years ago (95% HDI: 8729-12,836 years ago), whereas L4.5  
166 is the most modern of the studied sub-lineages in China, with its tMRCA appearing around  
167 7446 years ago (95% HDI: 5900-8901). As shown in Figure 3a, the substitution rate of Mtb  
168 was found to be a mean of  $4.35 \times 10^{-9}$  substitutions per genome per site per year [95% HPD  
169 interval:  $3.58 \times 10^{-9}$ - $5.26 \times 10^{-9}$ ; converted by the calculated annual mutation rate of each  
170 polymorphic locus (24,633 loci):  $uclid.mean=1.49 \times 10^{-6}$ ].

171

172 Given the times of origin for the four sub-lineages in China, the characteristics of the MCC  
173 tree (Supplementary\_Fig\_S2.), and historical information on the arrival and spread of modern  
174 humans in China (Comas et al. 2013), we propose two possible routes of propagation across  
175 China for each of the studied sub-lineages (Figure 4). For L2.2, one potential route of  
176 propagation originates in Xinjiang in Northwest China and spreads to the South and Southeast,  
177 while the other originates in Fujian and spreads to the north. For L4.2, one potential route of  
178 propagation originates in Qinghai Province in Western China and spreads to the East and  
179 Southeast, while the other originates in Heilongjiang Province in Northeast China and spreads  
180 to the South. For L4.4, one possible route of propagation originates in Guangdong and Hunan  
181 Provinces of Southern China and spreads to the North, while the other originates in  
182 Heilongjiang Province and spreads to the South. For L4.5, one possible route of propagation  
183 originates in Xinjiang Province and spreads to the East and Southeast, while the other  
184 originates in Heilongjiang Province and spreads to the South and Southwest. The origin times  
185 of some key propagation points are shown in Figure 4.



186 **Figure 4. Potential propagation routes of four sub-lineages in China.** Shown are routes for L2.2 (a),  
187 L4.2 (b), L4.4 (c) and L4.5 (d). The dotted line indicates that the distance is long and the evidence  
188 maybe weak (possibly due to a lack of strains). Blue lines indicate older transmission routes, while  
189 orange lines indicate more recent transmission routes.

190

191 We used a similar method to obtain the divergence times for the MRCAs of the six  
192 sub-lineages found in Zhejiang Province. As shown in Table 2, we found that L2.2 is the most  
193 ancient of the studied sub-lineages in Zhejiang, with its MRCA appearing around 6 897 years  
194 ago (95% HDI: 6513-7298 years), while L4.4 is the most modern of the studied sub-lineages  
195 in Zhejiang, with its MRCA appearing around 2217 years ago (95% HDI: 1864-2581 years).

196 **Table 2. The most recent common ancestor of L2 and L4 in Zhejiang**

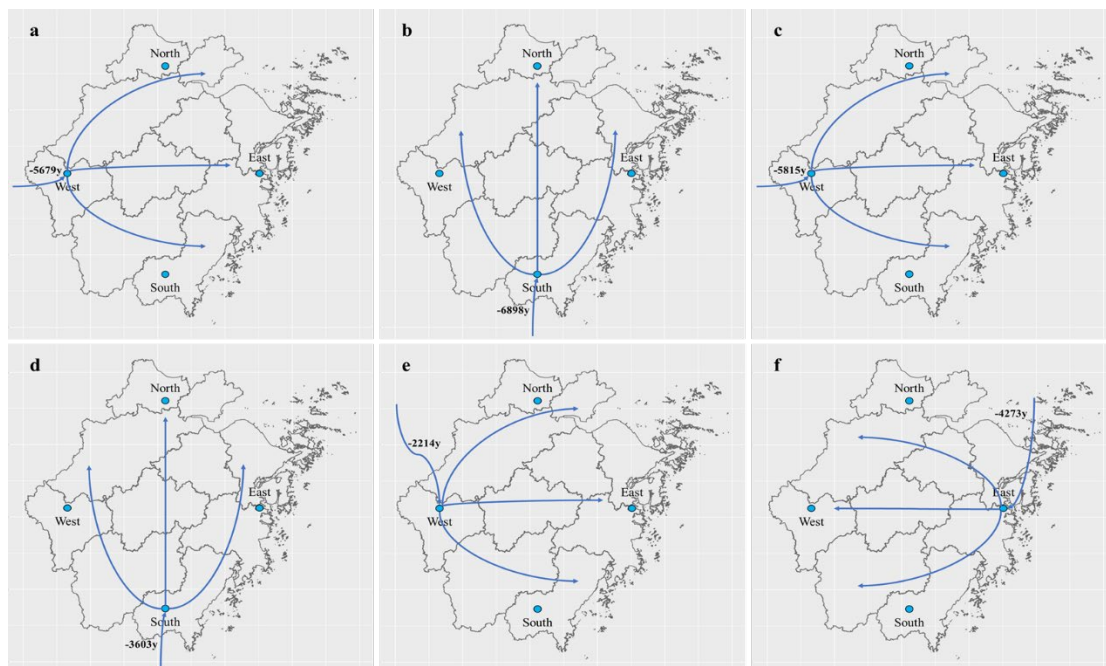
Summary statistics	L2.1	L2.2	L2.3	L4.2	L4.4	L4.5
Mean (tMRCA)	5,602	6,897	5,712	3,604	2,217	4,272
SE of the mean	14.6	4.6	13.4	13.2	10.7	6.9
Median (tMRCA)	5,679	6,898	5,815	3,603	2,214	4,273
Geometric mean	5,514	6,894	5,623	3,599	2,210	4,267
95% HDI	[5,077-6,123]	[6,513-7,298]	[5,202-6,229]	[3,220-4,012]	[1,864-2,581]	[3,841-4,670]
ESS	207.5	1894.6	229.8	238.9	291.6	958.1

197 tMRCA: the most recent common ancestor;

198 SE of the mean: standard error of the mean tMRCA;

199 HDI: highest posterior density interval; ESS: effective sample size.

200



201 **Figure 5. Potential propagation routes of six sub-lineages in Zhejiang Province.** Shown are L2.1  
 202 (a), L2.2 (b), L2.3 (c), L4.2 (d), L4.4 (e) and L4.5 (f). The curves without starting points indicate the  
 203 directions and years of the strains entering Zhejiang Province from other regions.

204

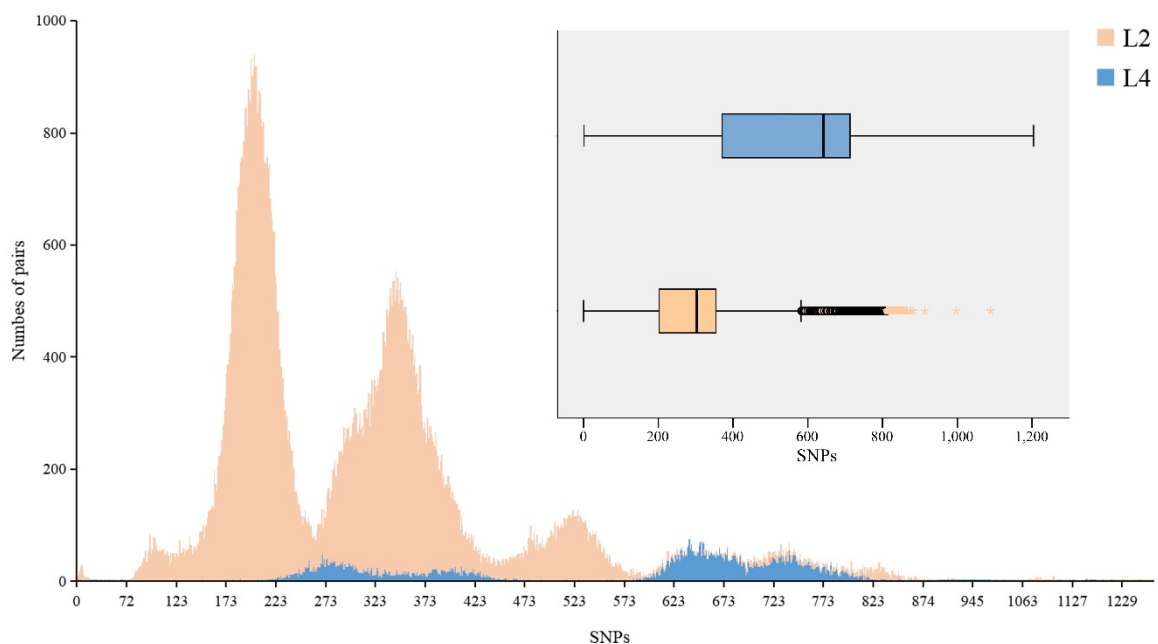
205 Given the origin times of the six sub-lineages in Zhejiang, the characteristics of the MCC tree  
 206 (Supplementary\_Fig\_S3.) and the above-described possible transmission routes of the four  
 207 sub-lineages in China, we inferred the potential propagation routes for the six sub-lineages in  
 208 Zhejiang, as shown in Figure 5. The directions and estimated years at which the strains  
 209 entered Zhejiang from other regions are basically consistent with the transmission routes of  
 210 the four sub-lineages (L2.2, L4.2, L4.4 and L4.5) in China. More specifically, L2.1 and L2.3,  
 211 which derived from 5,700 years ago, might be related to the origin and migration of Liangzhu  
 212 Culture (about 5,500 years ago), sharing similar original time and geographical distribution

213 (Yi 2019). L4.5, deriving from 3,600 years ago, might be related to the Battle of Mingtiao,  
214 which was the final battle of the Xia Dynasty (circa 1,600 BC). Shang Tang won the battle  
215 and Xia Jie retreated to Nanchao, adjacent to Zhejiang Province (Fan 2017). L4.4, deriving  
216 from 2,200 years ago, might be related to the war of Qin State destroying Chu State (circa 200  
217 BC). At that time, the territory of Chu included western and southeastern Henan, southern  
218 Shandong, Hubei, Hunan, Jiangxi, Anhui, Jiangsu, and Zhejiang. The marching route of Qin  
219 destroying Chu was consistent with the transmission route of L4.4 (Li 1981). Moreover, the  
220 spread of L2.2 might be related to the origin of Zhejiang's agricultural civilization and the  
221 transmission route of L4.5 began from sea, which may be related to the origin of the Maritime  
222 Silk Road (CCTV 2007).

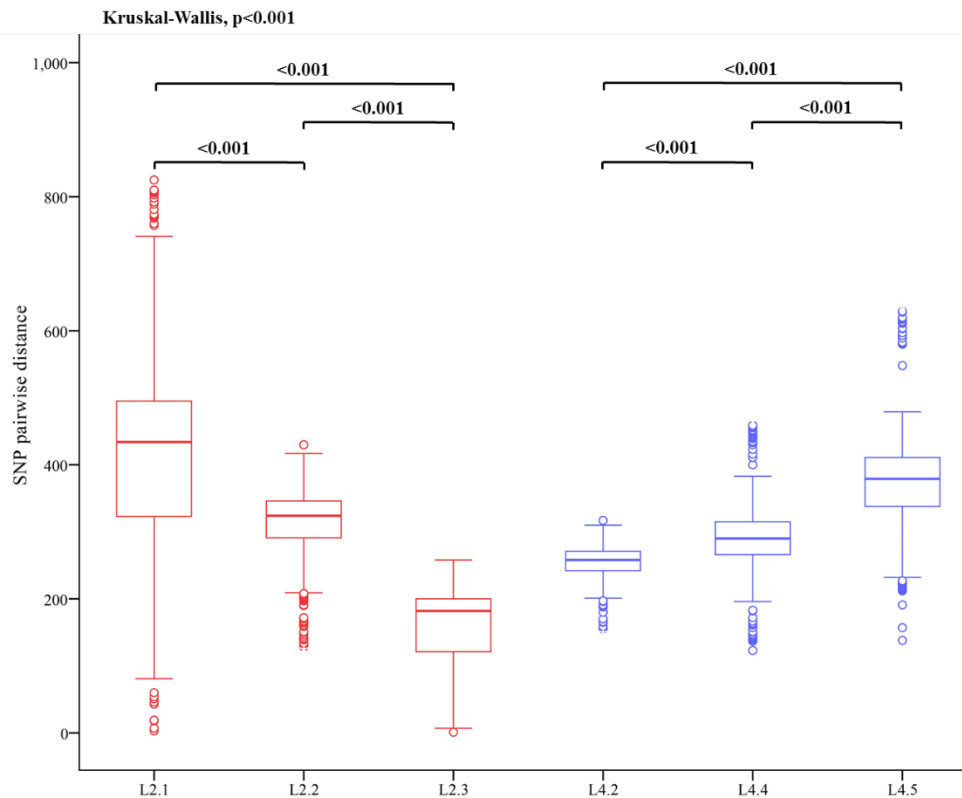
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#### 224 *Genomic features of lineages 2 and 4*

225 We compared the genetic diversity of the lineage 2 and 4 strains in Zhejiang Province to that  
226 of the global strains. As seen in the global strains, there was greater genetic diversity among  
227 the lineage 4 strains from Zhejiang Province than among the lineage 2 strains (Figure 6).  
228 Zhejiang lineage 4 strains harbored a mean diversity of 565 SNPs between any two strains,  
229 compared to 291 SNPs in lineage 2.



230 **Figure 6. Number of pairwise differences between Mtb strains for lineage 2 and lineage 4.** The  
231 alignment of 217 human-adapted Mtb clinical strains published previously (Comas et al., 2013) was  
232 used to calculate pairwise differences of global strains.



233 **Figure 7. Box plots of pairwise genetic distances (number of polymorphisms) for each**  
234 **sub-lineage.**

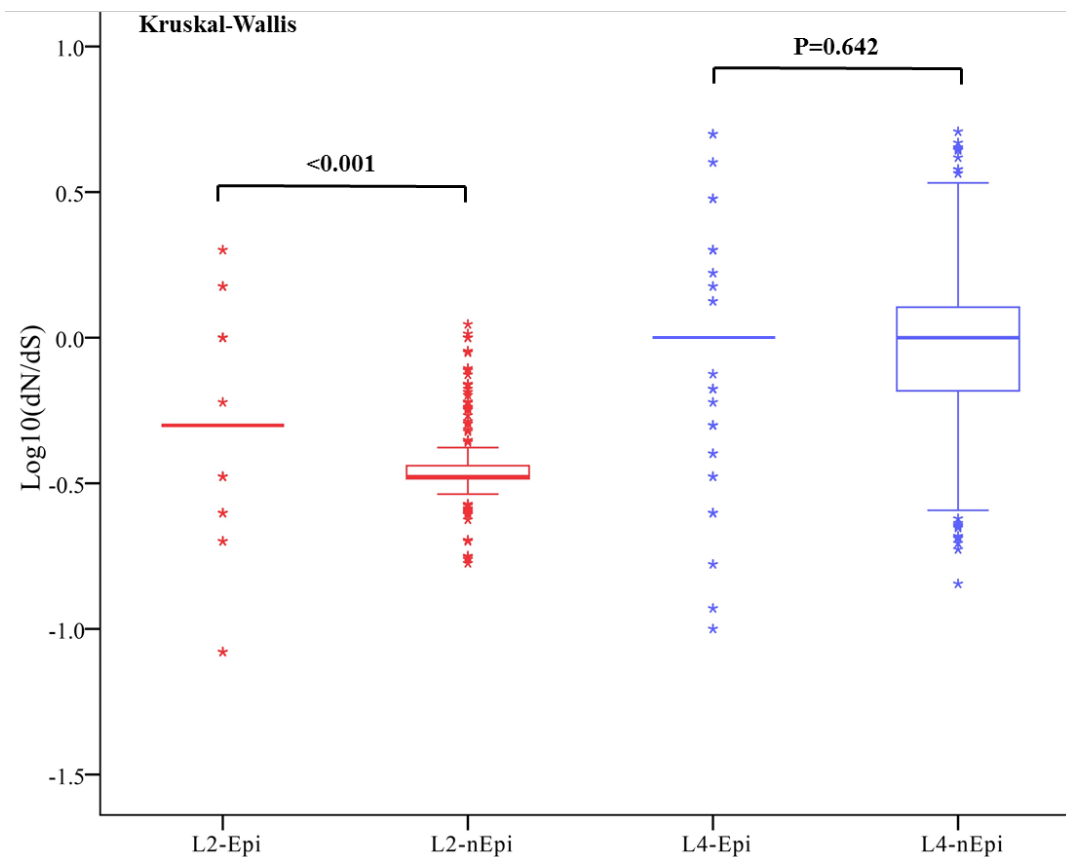
235

236 Our estimation of the genetic diversity among the sub-lineages of lineages 2 and 4 based on  
237 the SNP pairwise distances showed that L2.3, the predominant sub-lineage in lineage 2, was  
238 significantly more conserved than L2.1 (mean of 202 and 337 SNPs, respectively, shared  
239 between isolate pairs; Wilcoxon rank-sum test,  $P < 0.0001$ ). In lineage 4, we observed the  
240 opposite trend, as the predominant sub-lineage, L4.5, was more diverse than L4.2 (mean of  
241 385 and 253 SNPs, respectively; Wilcoxon rank-sum test,  $P < 0.0001$ ) (Figure 7).

242

243 To assess the genetic diversity of antigens in the lineage 2 and 4 strains, we calculated the  
244 non-synonymous to synonymous substitution (dN/dS) ratios for the epitope and non-epitope  
245 regions, along with the distribution of amino acid replacements in individual epitopes. We  
246 found that the dN/dS ratio of epitope and non-epitope regions exhibited significantly more  
247 conservation in lineage 2 strains than in lineage 4 strains. In lineage 2 strains, however, the T  
248 cell epitope regions showed significantly higher dN/dS ratios than the non-epitope regions  
249 (Figure 8). When we assessed the evolutionary conservation of human T cell epitopes in the

250 sub-lineages of lineage 2 and lineage 4 (Supplementary\_Fig\_S4.), we found that the dN/dS  
251 ratios for the sub-lineages of lineage 4 were similar to those of the overall lineage. For the  
252 sub-lineages of lineage 2, meanwhile, the dN/dS ratio of the lowest-prevalence sub-lineage,  
253 L2.1, differed from that of the overall lineage, whereas the ratios of the other sub-lineages  
254 were consistent with those of the overall lineage.



255 **Figure 8. Pairwise ratios for the rates of nonsynonymous to synonymous substitutions (dN/dS) in**  
256 **lineage 2 and 4 isolates, assessing epitope and non-epitope regions of T cell antigens.**

257

258 When we analyzed the distribution of amino acid replacements in individual epitopes, we  
259 found that a large majority (95%) of the 491 T cell epitopes showed no amino acid change  
260 (Supplementary\_Fig\_S5.). However, lineage 2 had more epitopes that harbored at least one  
261 amino acid change, compared to lineage 4. In lineage 2, four epitopes (*esxL*, *lpqH*, *fbpB* and  
262 *lppX*) harbored more than two variable positions.

263

## 264 Discussion

265 Whole-genome sequencing of 1296 Zhejiang Province strains and comparison with 1154



266 publically-available global MTBC genomes was used to elucidate the distribution of MTBC  
267 sub-lineages in the Chinese population. Genetic diversity and T cell epitopes were  
268 significantly different between sub-lineages.

269

270 We observed differences in the spatiotemporal characteristics of the lineage 2 and lineage 4  
271 strains. While the proportion of lineage 4 strains in Western Zhejiang was generally low, the  
272 proportion of cases arising from lineage 4 strains increased over time across the four survey  
273 periods. This increase may reflect the successful transmission of these strains over time. Other  
274 studies in various settings have reported that the higher fitness of lineage 2/ Beijing strains is  
275 reflected by increases in their frequency over time (Tuite et al. 2013). In contrast, the  
276 frequency of lineage 4 strains in Southern Zhejiang showed a downward trend, which is  
277 incompatible with the above hypothesis.

278

279 A previous study showed that migrants had an impact on the spread of Mtb in Russia  
280 (Mokrousov 2013). Lineage 4 was found at a high proportion in the Southern Zhejiang, which  
281 is typically the destination choice of migrant population from other provinces. Relatively low  
282 migration has been seen in the Western region of Zhejiang Province; however, due to  
283 developments in the economy and convenience of transportation, migration into this region  
284 increased significantly between 2000 and 2010. The similarity between the characteristics of  
285 migration and the trends in the proportion of lineage 4 suggest that there is likely to be a  
286 relationship between lineage 4 and migration. Future studies will be needed to assess whether  
287 migrants increase the risk of lineage 4 transmission in Zhejiang. Our Bayesian evolutionary  
288 analyses suggest that the identified sub-populations of Mtb emerged in China around 1000  
289 years ago, expanded in parallel from the 12<sup>th</sup> century onwards, and peaked (at a  
290 whole-population level) in the late 18<sup>th</sup> century. More recently, sub-lineage L2.3, which is  
291 indigenous to China and exhibits relatively high transmissibility and extensive global  
292 dissemination, came to dominate the population dynamics of Mtb in China.(Liu et al. 2018)

293

294 The tMRCAs that our Bayesian evolution model calculated for the four sub-lineages are  
295 related to the entry of modern humans into China, their migration routes, and the expansion of



296 the population in the Neolithic Age (about 10,000 years ago). We found that the population  
297 sizes all four sub-lineages increased significantly around 5000 years ago, which coincides  
298 with the origin of the Chinese civilization according to the historical record (Comas et al.  
299 2013). During that period, the population grew on a large scale and engaged in frequent social  
300 activities, presumably accelerating the evolution and spread of Mtb.

301

302 The Mtb strains differ genetically in their content of SNPs, and the more recently transmitted  
303 strains would be expected to have reduced levels of genetic diversity. Our findings show that  
304 number of pairwise differences between Mtb strains for lineage 2 in Zhejiang province was  
305 lower than that in global strains, whereas the opposite is true for lineage 4. The strains of  
306 lineage 2, which represent the predominant clades in Zhejiang, are separated by a smaller  
307 genetic distance, indicating more ongoing transmission. In contrast, the lineage 4 strains may  
308 be more likely to represent external inputs. The sub-lineages also differ in their genetic  
309 diversity, with sub-lineage L2.3 (the predominant within lineage 2) showing lower genetic  
310 distances compared to L2.1 and L2.2. Therefore, our results suggest this discrepancy supports  
311 the idea that there is an epidemiologic distinction between lineage 2 and lineage 4 in Zhejiang  
312 Province.

313

314 We detected three main potential routes for the spread of MTBC: the first originates in  
315 Xinjiang (about 8000 years ago) and may be traced back to human migration through the  
316 Eurasian continent from Europe to Central Asia, and then to East Asia (beginning around  
317 15,000-18,000 years ago) (Zhong et al. 2011); the second is consistent with the initial arrival  
318 of modern humans in South and Southeast Asia, followed by their entry into China by sea ~  
319 8000 years ago (Gray and Jordan 2000; Barton et al. 2009) and their subsequent spread to  
320 Southeastern China (Fujian, Guangdong and Hunan) about 6000 years ago; and the third and  
321 most modern route originates in Heilongjiang (3000-6000 years ago) and may trace back to  
322 Japan and Korea. These results are consistent with those of a previous study (Comas et al.  
323 2013). Our findings also support the idea that MTBC is a very old bacterium whose spread in  
324 China was achieved through the entry of modern humans into the country and their  
325 subsequent expansion and development of agricultural civilization (8000 years ago) (Wirth et

326 al. 2008).

327

328 The substitution rate per site per year obtained in our study was essentially the same as the  
329 genomic-level prediction ( $2.58 \times 10^{-9}$ , 95% HPD interval:  $1.66 \times 10^{-9}$  to  $2.89 \times 10^{-9}$ ) obtained by  
330 Comas et al. (Comas et al. 2013). However, this rate is much lower than recent estimates of  
331 short-term substitution rates for experimental models of TB and human outbreaks of the  
332 disease (Ford et al. 2011; Walker et al. 2013). Deleterious mutations tend to disappear during  
333 long-term evolution due to purifying selection, while the substitution rates tend to increase in  
334 experimental strains due to positive selection. This may explain why the substitution rate for  
335 long-term evolution is much lower than the short-term substitution rate.

336

337 We hypothesized that lineages that are predominant in a specific human population and  
338 undergoing ongoing transmission have a higher fitness and virulence (Rodrigo et al. 1997;  
339 Ernst 2012). In our study, as expected, essential genes were more conserved than nonessential  
340 genes, and a large majority of the currently known T cell antigens were completely conserved,  
341 in agreement with previous reports for the Mtb overall (Comas et al. 2010; Coscolla et al.  
342 2015). TB does not use antigenic variation as a main mechanism of immune evasion, and  
343 other studies found that reduced and/or delayed inflammatory responses were associated with  
344 increased Mtb virulence (Tsenova et al. 2005; Subbian et al. 2013). However, for both  
345 predominant lineage 2 and predominant sub-lineage L2.3, we obtained significantly higher  
346 dN/dS ratios for the T cell epitopes compared to the non-epitope regions. Other studies had  
347 found that although the majority of human T cell epitopes in Mtb were conserved (Comas et  
348 al. 2010) and relatively few of its antigens and epitopes exhibit evidence of diversifying  
349 selection and antigenic variation, the diverse regions exhibit nucleotide diversities and dN/dS  
350 ratios higher than the genome-wide average (Oleksyk et al. 2010). We identified four antigens  
351 that exhibited more than two nonsynonymous variations in the epitope regions of both  
352 lineages: *esxL*, *lpqH*, *fbpB* and *lppX*. Notably, these sites also exhibited diversity across the  
353 different successful sub-lineages. This natural sequence diversity suggests that variation in  
354 these particular antigens might benefit the pathogen, such as by allowing it to escape from  
355 human T cell recognition. Future studies will be needed to assess how the limited diversity in

356 Mtb T cell epitopes can impact immune escape. It is the conversation of most T cell epitopes  
357 in lineage 2 stains making them the delayed inflammatory immune response and increased  
358 virulence at a later stage, meanwhile, the diversity of some epitopes made them affect a wider  
359 population.

360

361 In conclusion, our study indicates that the spatiotemporal distribution characteristics of  
362 lineage 2 and 4 strains in Zhejiang Province are changing and the increase in the frequency of  
363 lineage 4 may reflect its successful transmission over the last 20 years. We reconstruct the  
364 phylogenomic history of TB transmission and analyze genomic features of lineages 2 and 4 in  
365 order to understand the intersection of phylogeny, geography, and demography to gain some  
366 insights about TB epidemics.

367

## 368 **Materials and Methods**

### 369 *Study population and samples*

370 The study population included patients with pulmonary disease and culture-positive TB  
371 sampled from 12 locations in Zhejiang Province of Eastern China during drug-resistance  
372 surveillances performed in 1998, 2003, 2008 and 2013. The same protocol was applied in all  
373 four surveillance periods. For each of the 12 locations, we randomly enrolled 30 new  
374 smear-positive patients and all previously treated smear-positive patients.

375

376 New cases were defined as those who had never received TB drugs or who had received  
377 treatment for less than 1 month. Previously treated cases were defined as those who had  
378 received previous TB treatment for 1 month or longer. All patients were active TB cases with  
379 bacteriological confirmation by sputum culture. Newly diagnosed patients provided three  
380 sputum specimens (spot, morning, and night) and previously treated patients provided two  
381 sputum specimens (spot and morning or night). Epidemiological data were collected by  
382 trained doctors at TB-designated hospitals, and patients were surveyed on site using a  
383 standard questionnaire. Demographic data for the study population are provided in  
384 Supplementary\_Table\_S1.

385

386 Samples were tested for Mtb by microscopy and culture in a manner consistent with national  
387 guidelines (China 2017). Isolates were cultured on Middlebrook medium for 4-6 weeks at  
388 37°C and DNA was extracted using magnetic beads (Tiangen Biotech Co., Ltd.). Rifampicin  
389 and isoniazid drug-susceptibility testing was performed using the proportion method in  
390 Löwenstein-Jensen medium (Aziz et al. 2008).

391

### 392 ***WGS of the 1296 Zhejiang Mtb strains***

393 Genomic DNA was sequenced using an Illumina HiSeq 2000 with an expected coverage of  
394 100X. Paired-end reads were mapped to the reference genome, H37Rv (GenBank AL123456),  
395 using the Bowtie 2 software. The SAMtools (version 1.6)/BCFtools suite was used to call  
396 fixed SNPs (frequency  $\geq 95\%$ ) (Li et al. 2009). We excluded all SNPs that were located in  
397 repetitive regions of the genome (e.g., PPE/PE/PGRS family genes, phage sequences,  
398 insertions and mobile genetic elements), as it is difficult to characterize such regions with  
399 short-read sequencing technologies (Yang et al. 2017). Small insertions or deletions, which  
400 were identified by VarScan (version 2.3.9) (Koboldt et al. 2012), were also excluded.

401

### 402 ***Collection of the relevant WGS data***

403 In order to construct phylogenetic trees including global strains and our samples, we curated a  
404 collection of MTBC representing geographic and genetic diversity. WGS data from global  
405 *Mycobacterium tuberculosis complex* (MTBC) lineage 2 and lineage 4 isolates was identified  
406 by searching PubMed for articles with WGS data. We downloaded the original sequencing  
407 reads from the European Nucleotide Archive (EMBL-EBI) and extracted the geographic  
408 origin and year of collection for each isolate from the relevant article. If the paper did not  
409 include this information, we sent an inquiry to the authors. Sequencing data were downloaded  
410 for 1154 MTBC isolates and geographic information was obtained for 1153 isolates  
411 (Supplementary\_Table\_S2).

412

### 413 ***Phylogenetic analysis and pairwise determination of SNP distances***

414 The fixed SNPs, excluding those in the proline-glutamic acid-proline-proline-glutamic acid  
415 sequence, the proline-glutamic acid-polymorphic GC-rich sequence and drug

416 resistance-associated genes, were combined into a concatenated alignment. The best-scoring  
417 maximum likelihood phylogenetic tree was computed using RAxML v7.4.2 (Alexandros 2014)  
418 based on the concatenated alignment of 98,672 sites spanning the whole genome. Given the  
419 considerable size of the dataset (1296 Zhejiang strains + 161 of 1154 global strains from  
420 China + 21 reference strains (Zhang et al. 2013; Liu et al. 2018); 98,672 SNP sites), the rapid  
421 bootstrapping algorithm ( $N=100$ ,  $x=12,345$ ) and maximum likelihood search were used to  
422 construct the phylogenetic tree. The resulting tree was rooted on *M. canettii* (GenBank  
423 accession number: NC\_019950.1). Lineage-defining nodes were based on 21 widely used  
424 isolates representing the six main phylogeographic lineages of MTBC. Bootstrap values were  
425 computed to assess the confidence of each clade, and to ensure that all lineage-defined nodes  
426 were highly supported (95-100%).

427

428 Filtered SNPs from isolates of lineages 2 and 4 were combined into a concatenated alignment  
429 as a fasta file. Pairwise SNP distances were calculated with the Bio:SeqIO package (Hackett  
430 et al. 2015). A pairwise SNP distance to all isolates of the same lineage was calculated for  
431 each isolate, and a distribution of the mean pairwise distance was plotted.

432

### 433 ***Bayesian-based coalescent analysis***

434 We randomly selected 197 Mtb strains from published studies (Zhang et al. 2013; Liu et al.  
435 2018) to represent the national diversity (31 out of the 34 provincial regions of China) of Mtb  
436 sub-lineages in China and 48 Mtb strains from Zhejiang to represent the provincial diversity  
437 (collected from four regions [eastern/northern/western/southern Zhejiang] in  
438 1998/2003/2008/2013, ignoring strains from middle Zhejiang to avoid confusion in  
439 constructing transmission routes) (Supplementary\_Table\_S3). The 197 and 48 strains were  
440 used for national and provincial phylogenetic reconstructions, respectively.

441

442 We applied Beast (Bayesian evolutionary analysis by sampling trees) (version 1.8.4)  
443 (Drummond et al. 2012), a genetic analysis software package based on the Monte Carlo  
444 Markov Chain algorithm (MCMC), to estimate the mutation rate, the divergence time of the  
445 Mtb strains and the times of the most recent common ancestors (tMRCAs) for lineages 2 and

446 4 and their sub-lineages. First, we imported the fasta file containing the genome sequencing  
447 information for the 197/48 strains into BEAUti software. To determine the Mtb genome  
448 substitution rate, we imposed a normal distribution for the substitution rate of Mtb with a  
449 mean of  $4.6 \times 10^{-8}$  substitutions per genome per site per year (95% highest posterior density  
450 [HPD] interval:  $3.0 \times 10^{-8}$  to  $6.2 \times 10^{-8}$ ), as described in a previous study (Bos et al. 2014).  
451 For the prior distribution of tMRCA, we imposed a normal distribution with a mean of 13,500  
452 and a SE of 3000, as previously applied by Lin et al. (N 2014). We used an uncorrelated  
453 log-normal distribution for the substitution rate, an optimal evolution model of GTR+Γ4  
454 (general time reversible + gamma-distributed rate variation with four rate categories), and the  
455 evolution model that was selected using Jmodeltest version 2.1.7.

456

457 To obtain reliable results, we ran a chain of  $1 \times 10^8$  generations, sampling every 10,000  
458 generations to ensure independent convergence of the chain. Convergence was assessed using  
459 Tracer (version 1.7.0) (Liu et al. 2018), ensure that all relevant parameters reached an  
460 effective sample size of >200. The first 10% of the chain was discarded as burn-in, and we  
461 used the remaining chain to construct a Maximum Clade Credibility Tree (MCC tree) using  
462 Tree Annotator (version 1.8.4). Phylogenetic trees were visualized using FigTree (version  
463 1.4.3). (Liu et al. 2018)

464

#### 465 *Calculation of dN/dS ratios*

466 To assess the antigenic diversity of human T cell epitopes among our Mtb samples, we chose  
467 a set of 491 epitopes corresponding to 130 non-overlapping regions in the antigen alignment  
468 (Comas et al. 2010). To assess how other regions of the genome are evolving, we also  
469 obtained alignments for essential and nonessential genes. Alignments of epitopes and  
470 non-epitope-containing regions for antigens, as well as essential and nonessential genes, were  
471 used to calculate pairwise dN/dS ratios for lineages 2 and 4. Pairwise dN and dS values within  
472 each lineage were calculated using the R package tool, seqinr, with the ka/ks function (Comas  
473 et al. 2010). To avoid having undetermined pairwise dN/dS values due to dN or dS being zero,  
474 we calculated a mean dN/dS value for each sequenced isolate by dividing its mean pairwise  
475 dN by its mean pairwise dS with respect to all other sequenced isolates within each lineage.

476

477 **Data Availability Statement**

478 The data underlying this article will be shared on reasonable request to the corresponding  
479 author.

480

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483

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489

490 **Author Contributions**

491 BW, YW, QW, XW and WW designed the study. BW, LZ, ZL, SC and XW collected and  
492 contributed the MTBC isolates analysed in this study. YW, QW, MB and WW analysed the  
493 sequencing reads and performed the genetic analysis. YW, LC and LB participated in the  
494 analysis of integrating tuberculosis history with Chinese human population history. QW and  
495 WZ performed the statistical analysis. BW, YW, QW, XW and WW drafted the manuscript.  
496 MB, BK revised the structure of this paper and polished the language. All authors critically  
497 reviewed and approved the final version of the manuscript.

498

499 **Declaration of Interests**

500 The authors declare that they have no competing interests.

## References

- Alexandros S. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**: 1312-1313
- Aziz, M.A, Wright, Muynck D, Laszlo. 2008. Anti-tuberculosis drug resistance in the world - third global report. *Geneva, Switzerland, World Health Organization [WHO], 2008* **12**: 257-261
- Barton L, Newsome SD, Chen FH, Wang H, Guilderson TP, Bettinger RL. 2009. Agricultural origins and the isotopic identity of domestication in northern China. *Proc Natl Acad Sci USA* **106**: 5523-5528.doi:10.1073/pnas.0809960106
- Bos KI, Harkins KM, Herbig A, Coscolla M, Weber N, Comas I, Forrest SA, Bryant JM, Harris SR, Schuenemann VJ, Campbell TJ, Majander K, Wilbur AK, Guichon RA, Wolfe Steadman DL, Cook DC, Niemann S, Behr MA, Zumarraga M, Bastida R, Huson D, Nieselt K, Young D, Parkhill J, Buikstra JE, Gagneux S, Stone AC, Krause J. 2014. Pre-Columbian mycobacterial genomes reveal seals as a source of New World human tuberculosis. *Nature* **514**: 494-497.doi:10.1038/nature13591
- CCTV. 2007. What is the "Silk Road" and "Maritime Silk Road"? ,
- China NHaFPCotPsRo. 2017. Diagnosis for pulmonary tuberculosis. *Health Industry Standards of the People's Republic of China*
- Coll F, McNerney R, Guerra-Assuncao JA, Glynn JR, Perdigao J, Viveiros M, Portugal I, Pain A, Martin N, Clark TG. 2014. A robust SNP barcode for typing Mycobacterium tuberculosis complex strains. *Nat Commun* **5**: 4812.doi:10.1038/ncomms5812
- Comas I, Chakravarti J, Small PM, Galagan J, Niemann S, Kremer K, Ernst JD, Gagneux S. 2010. Human T cell epitopes of Mycobacterium tuberculosis are evolutionarily hyperconserved. *Nat Genet* **42**: 498-503.doi:10.1038/ng.590
- Comas I, Coscolla M, Luo T, Borrell S, Holt KE, Kato-Maeda M, Parkhill J, Malla B, Berg S, Thwaites G, Yeboah-Manu D, Bothamley G, Mei J, Wei L, Bentley S, Harris SR, Niemann S, Diel R, Aseffa A, Gao Q, Young D, Gagneux S. 2013. Out-of-Africa migration and Neolithic coexpansion of Mycobacterium tuberculosis with modern humans. *Nat Genet* **45**: 1176-1182.doi:10.1038/ng.2744
- Coscolla M, Copin R, Sutherland J, Gehre F, de Jong B, Owolabi O, Mbayo G, Giardina F,



- Ernst JD, Gagneux S. 2015. M. tuberculosis T Cell Epitope Analysis Reveals Paucity of Antigenic Variation and Identifies Rare Variable TB Antigens. *Cell Host Microbe* **18**: 538-548.doi:10.1016/j.chom.2015.10.008
- Coscolla M, Gagneux S. 2010. Does M. tuberculosis genomic diversity explain disease diversity? *Drug Discov Today Dis Mech* **7**: e43-e59.doi:10.1016/j.ddmec.2010.09.004
- Coscolla M, Gagneux S. 2014. Consequences of genomic diversity in Mycobacterium tuberculosis. *Semin Immunol* **26**: 431-444.doi:10.1016/j.smim.2014.09.012
- Drummond AJ, Suchard MA, Xie D, Rambaut A. 2012. Bayesian phylogenetics with BEAUti and the BEAST 1.7. *Mol Biol Evol* **29**: 1969-1973.doi:10.1093/molbev/mss075
- Ernst JD. 2012. The immunological life cycle of tuberculosis. *Nat Rev Immunol* **12**: 581-591.doi:10.1038/nri3259
- Fan JJ. 2017. The pictures of the Battle of Mingtiao. *China surveying and mapping*: 61-63
- Filliol I, Motiwala AS, Cavatore M, Qi W, Hazbon MH, Bobadilla del Valle M, Fyfe J, Garcia-Garcia L, Rastogi N, Sola C, Zozio T, Guerrero MI, Leon CI, Crabtree J, Angiuoli S, Eisenach KD, Durmaz R, Joloba ML, Rendon A, Sifuentes-Osornio J, Ponce de Leon A, Cave MD, Fleischmann R, Whittam TS, Alland D. 2006. Global phylogeny of Mycobacterium tuberculosis based on single nucleotide polymorphism (SNP) analysis: insights into tuberculosis evolution, phylogenetic accuracy of other DNA fingerprinting systems, and recommendations for a minimal standard SNP set. *J Bacteriol* **188**: 759-772.doi:10.1128/JB.188.2.759-772.2006
- Ford CB, Lin PL, Chase MR, Shah RR, Iartchouk O, Galagan J, Mohaideen N, Ioerger TR, Sacchettini JC, Lipsitch M, Flynn JL, Fortune SM. 2011. Use of whole genome sequencing to estimate the mutation rate of Mycobacterium tuberculosis during latent infection. *Nat Genet* **43**: 482-486.doi:10.1038/ng.811
- Gagneux S. 2012. Host-pathogen coevolution in human tuberculosis. *Philos Trans R Soc Lond B Biol Sci* **367**: 850-859.doi:10.1098/rstb.2011.0316
- Glaziou P, Falzon D, Floyd K, Raviglione M. 2013. Global epidemiology of tuberculosis. *Semin Respir Crit Care Med* **34**: 3-16.doi:10.1055/s-0032-1333467
- Gray RD, Jordan FM. 2000. Language trees support the express-train sequence of Austronesian expansion. *Nature* **405**: 1052-1055.doi:10.1038/35016575

- Hackett R, Moulton OC, Raff JW. 2015. Biology Open: evaluating impact. *Biology Open* **4**: 1609
- Hershberg R, Lipatov M, Small PM, Sheffer H, Niemann S, Homolka S, Roach JC, Kremer K, Petrov DA, Feldman MW, Gagneux S. 2008. High functional diversity in *Mycobacterium tuberculosis* driven by genetic drift and human demography. *PLoS Biol* **6**: e311.doi:10.1371/journal.pbio.0060311
- Koboldt DC, Zhang Q, Larson DE, Shen D, McLellan MD, Lin L, Miller CA, Mardis ER, Ding L, Wilson RK. 2012. VarScan 2: somatic mutation and copy number alteration discovery in cancer by exome sequencing. *Genome Res* **22**: 568-576.doi:10.1101/gr.129684.111
- Leung CC, Chee CBE, Zhang Y. 2018. Tuberculosis updates 2018: Innovations and developments to end TB. *Respirology* **23**: 356-358.doi:10.1111/resp.13244
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, Genome Project Data Processing S. 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* **25**: 2078-2079.doi:10.1093/bioinformatics/btp352
- Li QHW, W. Z. 1981. The picture of Qin conquering the other six countries. *History Teaching*: 65
- Liu Q, Ma A, Wei L, Pang Y, Wu B, Luo T, Zhou Y, Zheng HX, Jiang Q, Gan M, Zuo T, Liu M, Yang C, Jin L, Comas I, Gagneux S, Zhao Y, Pepperell CS, Gao Q. 2018. China's tuberculosis epidemic stems from historical expansion of four strains of *Mycobacterium tuberculosis*. *Nat Ecol Evol* **2**: 1982-1992.doi:10.1038/s41559-018-0680-6
- Lonroth K, Jaramillo E, Williams BG, Dye C, Raviglione M. 2009. Drivers of tuberculosis epidemics: the role of risk factors and social determinants. *Soc Sci Med* **68**: 2240-2246.doi:10.1016/j.socscimed.2009.03.041
- Mokrousov I. 2013. Insights into the origin, emergence, and current spread of a successful Russian clone of *Mycobacterium tuberculosis*. *Clin Microbiol Rev* **26**: 342-360.doi:10.1128/CMR.00087-12
- N L. 2014. Genome-wide analysis of the populations of *Mycobacterium tuberculosis* from China.

- Nathanson E, Nunn P, Uplekar M, Floyd K, Jaramillo E, Lonnroth K, Weil D, Ravigliione M. 2010. MDR tuberculosis--critical steps for prevention and control. *N Engl J Med* **363**: 1050-1058.doi:10.1056/NEJMra0908076
- Oleksyk TK, Smith MW, O'Brien SJ. 2010. Genome-wide scans for footprints of natural selection. *Philos Trans R Soc Lond B Biol Sci* **365**: 185-205.doi:10.1098/rstb.2009.0219
- Parwati I, van Crevel R, van Soolingen D. 2010. Possible underlying mechanisms for successful emergence of the Mycobacterium tuberculosis Beijing genotype strains. *Lancet Infect Dis* **10**: 103-111.doi:10.1016/S1473-3099(09)70330-5
- Reed MB, Pichler VK, McIntosh F, Mattia A, Fallow A, Masala S, Domenech P, Zwerling A, Thibert L, Menzies D, Schwartzman K, Behr MA. 2009. Major Mycobacterium tuberculosis lineages associate with patient country of origin. *J Clin Microbiol* **47**: 1119-1128.doi:10.1128/JCM.02142-08
- Rodrigo T, Cayla JA, Garcia de Olalla P, Galdos-Tanguis H, Jansa JM, Miranda P, Brugal T. 1997. Characteristics of tuberculosis patients who generate secondary cases. *Int J Tuberc Lung Dis* **1**: 352-357
- Shih CH, Chang CM, Lin YS, Lo WC, Hwang JK. 2012. Evolutionary information hidden in a single protein structure. *Proteins* **80**: 1647-1657.doi:10.1002/prot.24058
- Stucki D, Brites D, Jeljeli L, Coscolla M, Liu Q, Trauner A, Fenner L, Rutaihwa L, Borrell S, Luo T, Gao Q, Kato-Maeda M, Ballif M, Egger M, Macedo R, Mardassi H, Moreno M, Tudo Vilanova G, Fyfe J, Globan M, Thomas J, Jamieson F, Guthrie JL, Asante-Poku A, Yeboah-Manu D, Wampande E, Ssenooba W, Joloba M, Henry Boom W, Basu I, Bower J, Saraiva M, Vaconcellos SEG, Suffys P, Koch A, Wilkinson R, Gail-Bekker L, Malla B, Ley SD, Beck HP, de Jong BC, Toit K, Sanchez-Padilla E, Bonnet M, Gil-Brusola A, Frank M, Penlap Beng VN, Eisenach K, Alani I, Wangui Ndung'u P, Revathi G, Gehre F, Akter S, Ntoumi F, Stewart-Isherwood L, Ntinginya NE, Rachow A, Hoelscher M, Cirillo DM, Skenders G, Hoffner S, Bakonyte D, Stakenas P, Diel R, Crudu V, Moldovan O, Al-Hajoj S, Otero L, Barletta F, Jane Carter E, Diero L, Supply P, Comas I, Niemann S, Gagneux S. 2016. Mycobacterium tuberculosis lineage 4 comprises globally distributed and geographically restricted

- sublineages. *Nat Genet* **48**: 1535-1543.doi:10.1038/ng.3704
- Subbian S, Bandyopadhyay N, Tsenova L, O'Brien P, Khetani V, Kushner NL, Peixoto B, Soteropoulos P, Bader JS, Karakousis PC, Fallows D, Kaplan G. 2013. Early innate immunity determines outcome of Mycobacterium tuberculosis pulmonary infection in rabbits. *Cell Commun Signal* **11**: 60.doi:10.1186/1478-811X-11-60
- Tsenova L, Ellison E, Harbacheuski R, Moreira AL, Kurepina N, Reed MB, Mathema B, Barry CE, 3rd, Kaplan G. 2005. Virulence of selected Mycobacterium tuberculosis clinical isolates in the rabbit model of meningitis is dependent on phenolic glycolipid produced by the bacilli. *J Infect Dis* **192**: 98-106.doi:10.1086/430614
- Tuite AR, Guthrie JL, Alexander DC, Whelan MS, Lee B, Lam K, Ma J, Fisman DN, Jamieson FB. 2013. Epidemiological evaluation of spatiotemporal and genotypic clustering of Mycobacterium tuberculosis in Ontario, Canada. *Int J Tuberc Lung Dis* **17**: 1322-1327.doi:10.5588/ijtld.13.0145
- Walker TM, Ip CL, Harrell RH, Evans JT, Kapatai G, Dedicoat MJ, Eyre DW, Wilson DJ, Hawkey PM, Crook DW, Parkhill J, Harris D, Walker AS, Bowden R, Monk P, Smith EG, Peto TE. 2013. Whole-genome sequencing to delineate Mycobacterium tuberculosis outbreaks: a retrospective observational study. *Lancet Infect Dis* **13**: 137-146.doi:10.1016/S1473-3099(12)70277-3
- WHO. 2018. WHO global tuberculosis control report 2018. .
- Wirth T, Hildebrand F, Allix-Beguec C, Wolbeling F, Kubica T, Kremer K, van Soolingen D, Rusch-Gerdes S, Locht C, Brisse S, Meyer A, Supply P, Niemann S. 2008. Origin, spread and demography of the Mycobacterium tuberculosis complex. *PLoS Pathog* **4**: e1000160.doi:10.1371/journal.ppat.1000160
- Yang C, Luo T, Shen X, Wu J, Gan M, Xu P, Wu Z, Lin S, Tian J, Liu Q. 2017. Transmission of multidrug-resistant Mycobacterium tuberculosis in Shanghai, China: a retrospective observational study using whole-genome sequencing and epidemiological investigation. *Lancet Infectious Diseases* **17**: 275-284
- Yi H. 2019. Liangzhu Culture and Huaxia Civilization. *The Central Plains Culture Research* **7**: 5-13.doi:10.16600/j.cnki.41-1426/c.2019.05.001
- Yruela I, Contreras-Moreira B, Magalhaes C, Osorio NS, Gonzalo-Asensio J. 2016.

Mycobacterium tuberculosis Complex Exhibits Lineage-Specific Variations Affecting Protein Ductility and Epitope Recognition. *Genome Biol Evol* **8**: 3751-3764.doi:10.1093/gbe/evw279

Zhang H, Li D, Zhao L, Fleming J, Lin N, Wang T, Liu Z, Li C, Galwey N, Deng J, Zhou Y, Zhu Y, Gao Y, Wang T, Wang S, Huang Y, Wang M, Zhong Q, Zhou L, Chen T, Zhou J, Yang R, Zhu G, Hang H, Zhang J, Li F, Wan K, Wang J, Zhang XE, Bi L. 2013. Genome sequencing of 161 Mycobacterium tuberculosis isolates from China identifies genes and intergenic regions associated with drug resistance. *Nat Genet* **45**: 1255-1260.doi:10.1038/ng.2735

Zhong H, Shi H, Qi XB, Duan ZY, Tan PP, Jin L, Su B, Ma RZ. 2011. Extended Y chromosome investigation suggests postglacial migrations of modern humans into East Asia via the northern route. *Mol Biol Evol* **28**: 717-727.doi:10.1093/molbev/msq247