

1 **FRONT MATTER**

2 **Title:** Disturbance Triggers Non-Linear Microbe-Environment Feedbacks

3
4 **Authors:** Aditi Sengupta^{1†}, Sarah J. Fansler^{2†}, Rosalie K. Chu³, Robert E. Danczak², Vanessa
5 A. Garayburu-Caruso², Lupita Renteria², Hyun-Seob Song⁴, Jason Toyoda³, Jacqueline Wells⁵,
6 and James C. Stegen^{2*}

7

8 **Affiliations:**

9 ¹California Lutheran University, Thousand Oaks, CA

10 ²Pacific Northwest National Laboratory, Ecosystem Science Team, Richland, WA

11 ³Environmental Molecular Sciences Laboratory, Richland, WA

12 ⁴University of Nebraska-Lincoln, Lincoln, NE

13 ⁵Oregon State University, Corvallis, OR

14

15 [†]Denotes equal contribution

16 *Correspondence: James.Stegen@pnnl.gov

17

18 **Abstract**

19 Conceptual frameworks linking microbial community membership, properties, and processes
20 with the environment and emergent function have been proposed but remain untested. Here we
21 refine and test a recent conceptual framework using hyporheic zone sediments exposed to
22 wetting/drying transitions. Throughout the system we found threshold-like responses to the
23 duration of desiccation. Membership of the putatively active community--but not the whole
24 community--responded due to enhanced deterministic selection (an emergent community
25 property). Concurrently, the thermodynamic properties of organic matter became less favorable
26 for oxidation (an environmental component) and respiration decreased (a microbial process).
27 While these responses were step functions of desiccation, we observed continuous monotonic
28 relationships among community assembly, respiration, and organic matter thermodynamics.
29 Placing the results in context of our conceptual framework points to previously unrecognized
30 internal feedbacks that are initiated by disturbance, mediated by thermodynamics, and that
31 cause the impacts of disturbance to be dependent on the history of disturbance.

32

33 **MAIN TEXT**

34

35 **Introduction**

36 Given the influence of microbes over ecosystem function, deeper knowledge of microbe-
37 environment relationships is needed to improve ecosystem models (1). In turn, there is strong
38 interest in quantifying and predicting microbe-environment relationships such as defining
39 microbial life history strategies as traits in ecosystem models (2), assessing microbial biomass
40 stoichiometry distributions in response to changing resource environments (3), and evaluating
41 the extent of microbial adaptation to changing environments and their role in biogeochemical
42 processes (4). To enhance and synthesize understanding of microbe-environment interactions,
43 it is useful to develop conceptual frameworks based on linkages among microbial characteristics

44 and ecosystem processes. Previous work has used such frameworks to improve mechanistic
45 representation and predictive capacity of microbe-environment interactions in ecosystem
46 models (5).

47

48 A recently developed framework by Hall et al. (6) poses a series of concepts that collectively
49 define the intersection between microbial and ecosystem ecology. Their framework draws
50 attention to causal relationships between microbial characteristics, environmental dynamics,
51 and cumulative ecosystems processes with the potential to incorporate relevant mechanistic
52 links into predictive ecosystem models. A powerful element of the Hall et al. (6) framework is
53 that it applies to diverse systems spanning natural (7), host-associated (8), and built (9)
54 environments as well as across spatiotemporal scales (10). While potentially very useful, the
55 Hall et al. (6) framework has seen little direct use in terms of explicitly defining and evaluating
56 the linkages within specific study systems (but see 3). To make full use of and continually
57 improve the framework, it is necessary to consider different realizations and interpretations of
58 the proposed linkages. In the following paragraphs we detail a modified interpretation of the
59 framework (**Fig. 1**) to enable its application to microbial communities and biogeochemistry
60 associated with hyporheic zone sediments experiencing hydrological disturbance. In turn, we
61 use data from a controlled laboratory experiment to evaluate key linkages within the modified
62 framework.

63

64 As in Hall et al. (6) we consider microbial membership to be directly influenced by environmental
65 conditions (arrow 4, Fig.1) and to underlie community-level properties (arrow 1, Fig. 1).
66 Determining microbial membership is relatively straightforward, and uses culture-independent
67 (11–14) and culture-dependent (15) techniques. Sequence-based assays using phylogenetic
68 markers are routine, with DNA-based (total community members) and RNA-based (putatively
69 active community members) (16–21) approaches providing the foundation to study community
70 properties.

71

72 While community membership is relatively straightforward, the identification of community
73 properties that are relevant to a given system and function is open to broader interpretation.
74 Here we propose using the relative influences of deterministic and stochastic community
75 assembly (22) as emergent properties of microbial communities that have implications for
76 biogeochemical function (23) including in ecosystems experiencing environmental disturbance.
77 Deterministic mechanisms are associated with systematic differences in reproductive success
78 imposed by the biotic and/or abiotic environment, while stochastic mechanisms are associated
79 with passive spatial movements of organisms and birth/death events that are not due to
80 systematic differences across taxa in reproductive success (24, 25). The relative contributions
81 of determinism and stochasticity can be inferred by coupling microbial community membership
82 and phylogeny to ecological null models (22, 25–27).

83

84 We propose the relative contributions of determinism and stochasticity to be an emergent
85 property that is greater than the sum of the individual components (i.e., taxa), and that is
86 complementary to the community properties proposed by Hall et al. (6), such as biomass and
87 gene expression. Note that here we conceptualize stochastic and deterministic events as

88 ecological community assembly processes and that the relative influences of these ecological
89 processes as an emergent community property. The ecological processes of community
90 assembly are distinct from ‘microbial processes’ associated with biogeochemical reactions. As
91 an emergent property, the relative influence of determinism and stochasticity is the result of
92 complex biotic and abiotic interactions (28) and also shapes cumulative microbial processes
93 that impact ecosystem biogeochemical functions (23) (arrow 2, Fig.1). For example, a stronger
94 influence of determinism over community assembly is hypothesized to cause higher respiration
95 rates (a microbial processes) due to a larger contribution of well-adapted taxa (23).

96
97 Analyses of microbial community assembly have been widely employed across environments
98 including soil (24, 29–32), sediment (26, 33–35), marine (36, 37), riverine (38), gut (39), and
99 engineered (40, 41) systems. Previous work has focused primarily on using DNA-derived
100 membership and phylogenetic data to study whole-community assembly. In contrast, recent
101 studies have also used an RNA-based approach to study the relative influence of stochasticity
102 over the assembly of the putatively active portion of microbial communities (31, 42). This RNA-
103 based approach is complementary to the DNA-based approach and may provide additional
104 insights into shorter-term dynamic linkages between emergent community properties and
105 microbial processes. Such linkages have not, however, been previously evaluated.

106
107 The Hall et al. (6) framework proposes that microbial processes (e.g., respiration rate) are
108 influenced by both microbial community properties and environmental factors. Here we propose
109 a revision of this structure that includes bidirectional links between the environment and
110 microbial processes (arrow 3, Fig. 1). Such bi-directional links between microbes and their
111 environment are common (43–46), and in hyporheic zone sediments may be particularly tied to
112 thermodynamic properties of organic matter (47) and influenced by hydrological disturbances
113 that are common in such environments. For example, preferential use OM by microbial
114 communities has potential to alter the thermodynamic properties of organic matter pools (33).
115 This microbe-driven shift in the environment could then feedback to impact microbial
116 metabolism due to the strong influence of OM thermodynamics on biogeochemical rates (34,
117 48–50). Bi-directional feedback between environmental factors and microbial processes is,
118 therefore, likely important to the link between microbial communities and ecosystem function.
119 Fundamental knowledge of these feedbacks and how they are modulated by hydrologic
120 disturbances in hyporheic zone sediments is largely unknown, however.

121
122 Within ecosystems, mechanistic associations between environmental factors, microbial
123 properties, and microbial processes underlie spatial and temporal distributions of
124 biogeochemical rates (Fig. 1 arrow 5). The resulting distributions (e.g., of respiration rates)
125 define cumulative system function and can be used to understand key phenomena such as
126 biogeochemical hot spots and hot moments (51). Developing concepts and models to predict
127 the influences of biogeochemical hot spots/moments is a major outstanding challenge. To
128 facilitate progress, Bernhardt et al. (52) proposed grouping hot spots/moments into the concept
129 of ecosystem control points that exert a disproportionate influence on ecosystem function.

130

131 While not called out explicitly in Bernhardt et al. (52), the control point concept is based on the
132 distribution of biogeochemical rates through space and/or time. Focusing on the shape of rate
133 distributions allows the notion of control points to be extended to the concept of control point
134 influence (CPI; Fig. 1 arrow 5). The CPI is a quantitative measurement of the contribution of
135 elevated biogeochemical rates in space and/or time to the net aggregated rate within a defined
136 system (53). While proposed conceptually and studied via simulation in Arora et al. (53),
137 empirical measurements of CPI are lacking. More generally, incorporating CPI into a modified
138 version of the Hall et al. (6) framework (Fig. 1), provides an integrated conceptualization for how
139 environmental factors, microbial properties, and microbial processes contribute to emergent
140 system function.

141
142 Some elements of our modified framework (Fig. 1) are generalizable across systems (e.g., CPI),
143 while others (e.g., OM thermodynamics) may have different levels of relevance across different
144 ecosystem types. Here we aim to generate fundamental knowledge of the linkages between
145 microbial community and ecosystem function, as well as reveal how hydrologic disturbance may
146 modify these linkages. We do so by studying the modified conceptual framework in context of
147 variably inundated hyporheic zone sediments exposed to different drying/wetting dynamics.
148 Hyporheic zones are biogeochemical active subsurface domains in river corridors through which
149 surface water flows and can mix with groundwater (51, 52, 54). These zones can have
150 disproportionate biogeochemical impacts on river corridors (54–58). Within variably inundated
151 streams (59, 60), hyporheic zones experience extreme changes in environmental conditions,
152 but the consequences of this variability for microbe-ecosystem linkages is poorly known.

153
154 To mimic natural disturbances we subjected sediments to wetting-drying transitions and focused
155 on a series of analyses tied to our modified framework. We specifically evaluated relationships
156 between: (i) the relative influence of stochastic assembly as a community property and
157 respiration rates as a microbial process (Fig. 1 arrow 2), and (ii) environmental features and
158 both microbial properties (Fig. 1 arrow 4) and processes (Fig. 1 arrow 3) that underlie aggregate
159 system function (Fig. 1 arrow 5). We hypothesized that: (i) Stronger influences of determinism
160 result in well-adapted microbes that will generate higher respiration rates, (ii) Longer duration in
161 an inundated state will result in greater influences of stochastic assembly--due to weaker
162 ecological selection--and lower respiration rates following re-inundation due to relatively
163 consistent abiotic conditions (61), and (iii) Microbial processes are facilitated by OM that is
164 thermodynamically more favorable for oxidation, leading to an association between respiration
165 rates and OM thermodynamics.

166

167 **Methods and Materials**

168 *Study site and sediment collection.* Hyporheic sediments were collected from the Columbia
169 River shoreline (approximately 46.372411°N, 119.271695°W) in eastern Washington State (Fig
170 S1) (34, 62–66) within the Hanford Site on January 14, 2019 at 9 am Pacific Standard Time.
171 Samples were aseptically collected to a depth of 10 cm at five sub-sampling locations within a
172 meter to form a composite sample that was sieved on site through a 2 mm sieve into a clean
173 glass beaker. Sieved sediment was stored on blue ice for 30 min while being transported back

174 to the laboratory. Once back at the laboratory sediment was stored at 4 °C until processing into
175 incubation vials (see below).

176

177 The sediments were subjected to increasing temporal environmental variance (as a function of
178 periodic wetting and drying transitions) and evaluated for associations between microbial
179 membership, microbial properties, microbial community assembly, OM chemistry, absolute
180 respiration rate (represented in this study as O₂ consumption rates), and cumulative respiration
181 rates represented as CPI. Aerobic respiration was chosen as the biogeochemical process since
182 it influences global-scale energy and material fluxes (67), and because the hyporheic zone
183 within the field system is predominantly aerobic (63, 68). Detailed experimental design and
184 methods are provided in the following paragraphs.

185

186 *Experimental design.* Sediments used in the batch reactors were sourced from one
187 homogenized sediment pool. In turn, 10 g of sediment from the homogenized pool was added to
188 each reactor vial. The sample set was then split into two groups, one inundated and the other
189 allowed to desiccate. These conditions were maintained for 23 days prior to the start of dynamic
190 moisture manipulation, from January 29th to February 21st, 2019. This initial ‘preconditioning’
191 period was used to avoid measuring the immediate impacts of sampling disturbance and to
192 allow time for desiccation. All replicate reactors were maintained in the dark, shaking at 100
193 rpm, at 21 °C and were covered with a gas permeable Breathe-Easy (Milli-Pore Sigma,
194 Burlington, MA) membrane that allowed for gas exchange and drying. After the preconditioning
195 period in which sediments were consistently inundated or allowed to continuously desiccate, the
196 transition regimes (**Fig. 2**) were applied to the reactors starting on February 22nd, 2019. We
197 refer to the time from Feb 22nd, 2019 onwards as the ‘transitions period’ for all treatments, even
198 though some did not experience transitions between being inundated and dry. Each treatment
199 had 6-7 replicates (detailed below). These regimes were designed around the number of
200 wet/dry transitions experienced by sediments within a given treatment. Treatment regimes also
201 caused variation in the cumulative number of days sediments were in a drying state. We
202 imposed six different experimental treatment regimes (**Fig. 2**) as follows:

203

- 204 • 0 Transitions and 0 days of desiccation: Sediments were maintained at field moisture
205 conditions for the preconditioning and transitions periods. This treatment had 6
206 replicates.
- 207 • 1 Transition and 34 days of desiccation: Sediments were dry during the preconditioning
208 and transitions periods, and transitioned once to the field moisture level prior to
209 respiration estimation. This treatment had 6 replicates.
- 210 • 2 Transitions and 4 days of desiccation: Sediments were held at field moisture levels for
211 the preconditioning period and then for the first 7 days of the transitions period, then
212 transitioned to a dried state for 4 days, and transitioned to field moisture conditions prior
213 to respiration estimation. This treatment had 7 replicates.
- 214 • 3 Transitions and 31 days of desiccation: Sediments were dry during the preconditioning
215 period and the first 4 days of the transitions period, then transitioned to field moisture
216 levels for three days, then transitioned to 4 days in a dried state, and transitioned again
217 to field moisture levels prior to respiration estimation. This treatment had 7 replicates.

- 218 ● 4 Transitions and 8 days of desiccation: Sediments were held at field moisture levels for
219 the preconditioning period and transitioned to a dried state for the first 4 days of the
220 transitions period, then transitioned to field moisture levels for 3 days, then transitioned
221 to 4 days in a dried state, and transitioned to field moisture levels prior to respiration
222 estimation. This treatment had 7 replicates.
223 ● 5 Transitions and 27 days of desiccation: Sediments were dry during the preconditioning
224 period and then transitioned to field moisture levels for the first 3 days of the transitions
225 period, transitioned to a dried state for 2 days, transitioned to field moisture levels for 2
226 days, transitioned to a dried state for 2 days, and transitioned to field moisture levels for
227 3 days prior to respiration estimation. This treatment had 7 replicates.
228

229 To avoid modifying electrical conductivity across experimental treatments, sterile deionized
230 water was added to reactors to achieve/maintain field moisture levels according to the defined
231 wet/dry regimes detailed above. For reactors with sediments that were below field moisture
232 levels, deionized water was added to achieve field moisture levels prior to respiration rate
233 estimation. Changes in the total mass of reactors and volumes of water added during the course
234 of the experiment are provided in Data file S1.

235
236 *Respiration rate measurements.* Laboratory incubations were performed in batch reactors to
237 quantify dissolved oxygen consumption rates. Borosilicate glass vials (20 ml) (I-Chem™ Clear
238 VOA Glass Vials, Thermo-Fisher, Waltham, MA) served as incubator reactors. Factory
239 calibrated oxygen sensor spots (Part# 200001875, diameter = 0.5 cm, detection limit 15 ppb, 0
240 – 100 % oxygen; PreSens GmbH, Regensburg, Germany) were adhered to the inner vials of the
241 reactor prior to sediment addition. Detailed description of sensor adhesion and non-destructive
242 measurements of DO consumption using these sensors is provided in Garayburo-Caruso et al.
243 (49). Vials were grouped into six treatment regimes (explained in the previous section)
244 representing inundation-drought transitions.
245

246 Sample processing and incubations were performed in a laboratory at 21 ± 1 °C. The reactors
247 were monitored for 2 hours, with measurement of dissolved oxygen (DO) concentration (μmol
248 L^{-1}) every 30 min. DO concentration in each bioreactor was measured with an oxygen optical
249 meter (Fibox 3; PreSens GmbH) connected to a 2 mm polymer optical fiber lined up to sense
250 the sensor dot every thirty minutes. Respiration rates ($\mu\text{mol L}^{-1} \text{ h}^{-1}$) were estimated as the slope
251 of the linear regression between DO concentration and incubation time for each sample. Some
252 non-linearity was observed in the relationship between DO concentration and time such that
253 only the first 4 data points--time zero to 2 hours--were used to fit a linear function. The slope of
254 the linear function was taken as an estimate of respiration rate.
255

256 *Microbial Analysis.* Post-incubation, the sediment slurry was transferred to centrifuge tubes
257 (Item#28-108 Genesee Scientific) and centrifuged for 5 min at 3200 rcf and 20°C. The
258 supernatant was removed and reserved for biogeochemistry analyses and sediment aliquots for
259 DNA and RNA extraction were flash-frozen in liquid N₂ and stored at -80 °C. The extraction,
260 purification, and sequencing of sediment microbial gDNA were performed according to
261 published protocol (29). The extraction of RNA was performed using the Qiagen PowerSoil RNA

262 extraction kit (Qiagen, Germantown, MD). RNA was treated with DNase and quantified with a
263 Qubit RNA kit (Thermo Fisher, Waltham, MA). An aliquot of the RNA extraction was used to
264 generate cDNA using SuperScript™ IV First-Strand Synthesis System (Thermo Fisher
265 Scientific, Waltham, MA). The 16S rRNA gene sequencing--for both gDNA and cDNA--followed
266 the established protocol by The Earth Microbiome Project (94). Sequence pre-processing,
267 operational taxonomic unit (OTU) assignment, and phylogenetic tree building were performed
268 using an in-house pipeline, HUNDO (69). Sequences were deposited at NCBI's Sequence Read
269 Archive PRJNA641165. The final sample count of gDNA and cDNA, respectively, for each
270 treatment regime, after dropping samples following quality filtering and rarefaction, was 5 and 5
271 (0 transition), 4 and 3 (1 transition), 5 and 4 (2 transitions), 7 and 6 (3 transitions), 7 and 6 (4
272 transitions), and 7 and 5 (5 transitions). Rarefaction levels are provided below in the Statistics
273 section.

274 *Biogeochemistry.* Reserved supernatant was filtered through a 0.22 µm polyethersulfone
275 membrane filter (Millipore Sterivex) and an aliquot was immediately removed for non-purgeable
276 organic carbon (NPOC) and the remainder was stored at -20C until further OM high resolution
277 analysis was conducted (see below). NPOC was determined by acidifying an aliquot of sample
278 with 15% by volume of 2N ultra-pure HCl (Optima grade, Fisher#A466-500). The acidified
279 sample was sparged with carrier gas (zero air, Oxarc# X32070) for 5 minutes to remove the
280 inorganic carbon component. The sparged sample was then injected into the TOC-L furnace of
281 the Shimadzu combustion carbon analyzer TOC-L CSH/CSN E100V with ASI-L auto sampler at
282 680°C using 150 uL injection volumes. The best 4 out of 5 injections replicates were averaged
283 to get the final result. The NPOC organic carbon standard was made from potassium hydrogen
284 phthalate solid (Nacalia Tesque, lot M7M4380). The calibration range was 0 to 70 ppm NPOC
285 as carbon.

286 Fourier Transform Ion Cyclotron Resonance Mass Spectrometry (FTICR-MS) of post-incubation
287 sediment slurry was conducted as per Danczak et al. (70). Sample processing, injection, and
288 data acquisition, processing and analysis was performed as per scripts provided in Danczak et
289 al. (70), with '*Start tolerance*' in Formularity changed to 8. Ten samples were dropped due to
290 poor calibration, resulting in 5 replicates for 0 transition, 4 replicates for 1 transition, 5 replicates
291 for 2 transitions, 6 replicates for 3 transitions, and 5 replicates each for 4 and 5 transition
292 regimes.

293 From the FTICR-MS data, as in previous work (32, 34, 47, 49), we followed LaRowe and Van
294 Cappellen (71) to calculate the Gibbs free energy for the half reaction of organic carbon
295 oxidation under standard conditions (ΔG^0_{Cox}). This calculation is based on elemental
296 stoichiometries associated with molecular formulae assigned to individual molecules observed
297 in the FTICR-MS data. The formulae assignments are part of the processing scripts described in
298 (70). As in previous work (32, 34, 47, 49), we interpret larger values of ΔG^0_{Cox} to indicate OM
299 that is thermodynamically less favorable for oxidation by microbes. That is, larger values of
300 ΔG^0_{Cox} indicate OM that provides less net energy to a microbial cell per oxidation event,
301 assuming all else is equal. Given the large numbers of assigned formulae within each sample,
302 this resulted in thousands of ΔG^0_{Cox} estimates within each sample, from which we estimated
303 mean ΔG^0_{Cox} for each sample.

305

306 **Statistics.**

307 *Estimating influences of community assembly processes.* The relative influences of community
308 assembly processes influencing microbial community membership are emergent properties that
309 cannot be calculated/inferred directly from knowledge of membership. To evaluate assembly
310 processes as a link between membership and microbial processes (reference framework
311 graphic) it is necessary to quantitatively estimate the relative influences of these processes. To
312 do so we use a well-established null modeling framework based on phylogenetic relationships
313 among microbial taxa (22, 24, 25, 27). We refer the reader to these previous studies for details.
314 In brief, randomizations were used to generate estimates of phylogenetic associations among
315 microbial taxa for scenarios in which microbial communities were stochastically assembled.
316 These stochastic (i.e., null) expectations were compared quantitatively to observed phylogenetic
317 associations to estimate the β -Nearest Taxon Index (β NTI) (22). We used cDNA sequences
318 rarefied to 27227 and gDNA sequences rarefied to 15106 sequences per sample to determine
319 putatively active community and whole community β NTI values, respectively. Samples falling
320 below these sequence counts were removed as indicated above in subsection *Microbial*
321 *Analysis.* A β NTI value of 0 indicates no deviation between the stochastic expectation and the
322 observed phylogenetic associations, thereby indicating the dominance of stochastic assembly
323 processes. As β NTI deviates further from 0, there is an increasing influence of deterministic
324 assembly processes that drive community membership away from the stochastic expectation.
325 β NTI values below -2 or above +2 indicate statistical significance, with negative and positive
326 values indicating less than or more than expected shifts in membership. β NTI is a pairwise
327 metric measured between any two communities/samples, such that shifts in membership are
328 related to changes between the pair of communities being evaluated. We used β NTI to study all
329 pairwise community comparisons within each experimental treatment. Each community from a
330 given reactor is therefore associated with multiple β NTI values due to being compared to
331 communities associated with other replicate reactors. In turn, the average β NTI was calculated
332 for each reactor. As in Stegen et al. (25), this provides a community-specific value for β NTI and
333 thus an estimate of the relative influences of stochastic and deterministic processes causing
334 deviations between a given community and all other communities within the same experimental
335 conditions (32). That is, the larger the absolute value of β NTI for a given community, the
336 stronger the influence of deterministic assembly processes acting on that community (25). In
337 turn, these community-specific estimates were related to reactor-specific measurements. For
338 example, respiration rates were regressed against β NTI to evaluate the link between emergent
339 properties (i.e., ecological assembly) and microbial processes (i.e., respiration rates).

340

341 *Evaluating relationships between microbial characteristics and environment.* Respiration rate
342 distributions, absolute β NTI values, and ΔG^0_{Cox} were summarized as box plots. Pairwise Mann-
343 Whitney test was performed to evaluate statistical differences between reactor-specific
344 measurements (e.g., respiration rates and thermodynamic properties) and treatment groups
345 (cumulative dry and inundated days). Continuous bivariate relationships were evaluated with
346 ordinary least squares regression. Prior to regression analyses, respiration rates were log-
347 transformed due to non-linearities resulting from respiration being constrained to be at or above

348 zero. Prior to log-transformation, half the smallest non-zero rate was added to each rate to
349 enable inclusion of rate estimates with a value of zero.
350

351 *Control point influence calculation.* To characterize respiration rate distributions we used the
352 control point influence (CPI) metric. CPI was recently developed (53) and is defined as the
353 fraction of cumulative function (R_{tot} ; e.g., total respiration rate) within a defined system that is
354 contributed by individual rates that are above the system's median rate (R_{med}). To define
355 cumulative function one must first define the system being evaluated. In our study, all replicate
356 batch reactors within a given experimental treatment were conceptualized as a representative
357 set of samples from a larger system experiencing the experimental conditions. R_{tot} for each
358 treatment was therefore estimated as the sum of respiration rates across a treatment's replicate
359 reactors. CPI was estimated as the sum of respiration rates that fell above the median rate for a
360 given treatment (R_{above}) divided by R_{tot} for that treatment. That is, $R_{\text{above}} = \sum_i^N R_i$, where R_i are
361 respiration rates from individual reactors that fell above R_{med} , and $CPI = R_{\text{above}} / R_{\text{tot}}$.
362

363 An important feature of CPI is that it makes no assumptions of distribution normality, and can be
364 estimated for rate distributions of any form (e.g., unimodal, multimodal, Gaussian, skewed, etc.).
365 In most cases, CPI is constrained to have a minimum value of 0.5 (for a perfectly normal
366 distribution with no outliers) and asymptotically approach 1 as a maximum value (e.g., for
367 heavily skewed distributions with a small number of very high rates). CPI therefore quantitatively
368 estimates the biogeochemical contribution of places in space or points in time that have
369 elevated biogeochemical rates.
370

371 **Results**

372

373 To link microbial membership to emergent microbial community properties (Fig. 1 arrow 1) we
374 used null modeling to estimate the contributions of stochastic and deterministic community
375 assembly. Results from the null models indicate a relatively balanced mixture of stochasticity
376 and determinism for both the whole community (gDNA-based) and putatively active community
377 (rRNA-based) (Fig. S2). More specifically, stochasticity and determinism each governed 50% of
378 turnover in microbial membership for the whole community and 33% and 67%, respectively, for
379 the putatively active community. The relative contributions of the two deterministic components
380 (homogeneous and variable selection) were strongly imbalanced. Homogeneous selection was
381 responsible for 94% and 91% of the deterministic component for the whole and putatively active
382 communities, respectively. The contributions of homogeneous and variable selection to the
383 deterministic component must sum to 1, such that the variable selection was responsible for 6%
384 and 9% of the deterministic component for the whole and putatively active communities,
385 respectively.
386

387 As shown in Figure 1 (arrow 2), we hypothesized a link between microbial community properties
388 and microbial processes realized as a relationship between the strength of determinism and
389 respiration rates. Such a relationship was not observed for the whole community (Fig. 3a), but
390 we did observe a non-linear decreasing relationship between respiration rates and the absolute

391 value of β NTI for the putatively active community (Fig. 3b). The direction of this relationship
392 (negative) was opposite of that expected.
393

394 In our conceptual framework there are multiple ways in which connections among the
395 environment, microbial properties, and microbial processes may be realized, in part due to the
396 environment having multiple components relevant to our study (Fig. 1 arrows 3,4). More
397 specifically, the environment was characterized here in terms of both disturbance (number of
398 dry days; imposed by the experimental manipulation) and OM thermodynamics (ΔG^0_{Cox} ; this is
399 an emergent aspect of the environment).

400 Disturbance influenced both microbial properties and processes. These influences appeared to
401 be non-linear with experimental treatments associated with the two largest number of dry days
402 (31 and 34) causing decreases in respiration rates (Fig. 4a) and stronger influences of
403 deterministic homogeneous selection for the putatively active community (Fig. 4b). Disturbance
404 had no clear influence on community assembly for the whole community (Figs. S3, S4a). Given
405 the apparent binary nature of these results, we evaluated statistical significance by combining
406 respiration rate data from treatments with 0-27 cumulative dry days and separately combining
407 data from treatments with 31 or 34 cumulative dry days (Fig. S5). Respiration rates were
408 significantly depressed in the treatments associated with 31 or 34 cumulative dry days ($W = 5$, p
409 < 0.001). The β NTI data are non-independent due to being based on all pairwise comparisons
410 within a treatment. Standard statistical tools are therefore not applicable for assigning statistical
411 significance when comparing β NTI distributions. However, as shown in Figures 4b and S4b,
412 there is an obvious shift to lower β NTI values for the putatively active community in the
413 treatments with 31 or 34 cumulative dry days.
414

415 The other aspect of the environment examined here (i.e., OM thermodynamics) also had
416 significant relationships with both microbial processes (Fig. 1, arrow 3) and properties (Fig. 1,
417 arrow 4). More specifically, respiration rates decreased significantly as a negative exponential
418 function of increasing ΔG^0_{Cox} ($R^2 = 0.26$; $p = 0.01$; Fig. 5a). This indicates a decrease in
419 respiration rate as OM thermodynamic properties shifted towards lower favorability for oxidation
420 (i.e., larger values of ΔG^0_{Cox}). Similarly, we found that the strength of deterministic assembly
421 associated with the putatively active community increased linearly with ΔG^0_{Cox} ($R^2 = 0.46$; $p <$
422 0.0001; Fig. 5b). The strength of deterministic assembly associated with the whole community
423 was unrelated to ΔG^0_{Cox} ($p = 0.64$) (Fig. S6).
424

425 The conceptual model described in Figure 1 focuses primarily on connections among
426 environmental and/or microbial attributes, but there are potentially important relationships within
427 attribute categories. In particular, within the environmental category there is the potential for an
428 influence of disturbance on OM thermodynamics. Such an effect was found for OM
429 thermodynamics as measured by ΔG^0_{Cox} (Fig. 5c). Using the same approach as for analyses
430 described above, we combined data for treatments with 0-27 cumulative dry days and
431 compared that distribution to data combined across treatments with 31 or 34 cumulative dry
432 days. A Mann-Whitney test comparing these distributions confirmed a significant change in the
433 ΔG^0_{Cox} distribution ($W = 189$, $p = < 0.001$) (Fig. S7).
434

435
436 The last component of the conceptual model considered here is the connection between
437 microbial processes occurring in a given location and cumulative system function that
438 aggregates across locations (Fig. 1, arrow 5). It is at the system level that the influence of
439 biogeochemical hot spots (or hot moments) can be evaluated. We conceptualized an aggregate
440 system as the collection of replicate batch reactors within a given experimental treatment.
441 Based on this definition, we estimated control point influence (CPI) as a measurement for the
442 influence of biogeochemical hot spots. We observed a large amount of variation in CPI across
443 experimental treatments, but there was no clear, direct influence of the treatments on CPI (Fig.
444 4). The largest value of CPI observed (> 0.9) was associated with the treatment that imposed 31
445 cumulative dry days. This treatment also had the lowest median respiration rates across all
446 treatments (Fig. 4).

447
448 **Discussion**
449 Mechanistic evaluation of microbe-environment interactions is fundamental to understanding
450 microbe-mediated ecosystem function. Inspired by a microbe-environment-ecosystem
451 framework proposed by Hall et al. (6) we proposed and evaluated a modified framework linking
452 microbial characteristics (membership, emergent properties, processes), the environment
453 (disturbance, OM thermodynamics), and cumulative ecosystem function (CPI) of hyporheic zone
454 sediments. Our results provide clear support for the overall conceptual framework and further
455 point to an iterative loop among OM thermodynamics, respiration rates, and microbial
456 community assembly that can be initiated by externally-imposed disturbance. Furthermore, our
457 results indicate that the iterative thermodynamics-assembly-respiration loop may be initiated
458 through threshold-like impacts of disturbance that were observed only after 31 or more
459 cumulative days of desiccation.

460
461 We first evaluated emergent community properties as a function of microbial membership by
462 studying the relative influences of stochasticity and determinism over community assembly.
463 Taking this approach, we found fully balanced stochastic-deterministic influences over the whole
464 community, in which each contributed to 50% of the variation in community composition. The
465 relative influences of stochasticity and determinism have been quantified for many microbial
466 systems and the estimates are highly variable (72, 73). In addition, within the deterministic
467 component of assembly, homogeneous selection had a far greater influence than variable
468 selection. Previous work has also observed a broad range of contributions from homogeneous
469 and variable selection (63, 74–77). As such, the assembly-associated outcomes observed here
470 for the whole community are not unexpected relative to previous work. Very few studies,
471 however, have examined the relative influences of different assembly components over
472 putatively active microbial communities.

473
474 For the putatively active communities we found that across all treatments both stochasticity and
475 determinism were important, though deterministic assembly had greater influence. This deviates
476 quantitatively from the whole community in which the influences of stochasticity and
477 determinism were more balanced. Consistent with the whole community results, however, was
478 the dominance of homogeneous selection within the deterministic component of assembly. The

479 strong influence of homogeneous selection is likely due to selection-based constraints imposed
480 by aspects of the experimental system that did not vary across treatments. For example,
481 mineralogy is known to strongly influence microbial communities (35, 78–82) and was
482 homogenized across the experimental batch reactors, thereby potentially imposing
483 homogeneous selection on both the whole and putatively active communities.
484

485 Our study uniquely evaluates null-model outcome of putatively-active community assemblies in
486 hyporheic zone sediments, where homogeneous selection was further enhanced by our
487 experimentally imposed hydrologic disturbances. Increased homogeneous selection in response
488 to disturbance is consistent with previous work in a soil system. In the soil system, disturbance
489 led to an immediate increase in homogeneous selection for the putatively active community
490 (31). The strong influence of homogeneous selection on the putatively active community is not
491 always observed, however, suggesting it may be tied to acute disturbance. That is, Jia et al.
492 (42) recently found that within a natural soil chronosequence, variable selection was stronger for
493 putatively-active communities while homogeneous selection influenced the whole community
494 assembly. Our results combined with these previous studies indicate that community assembly
495 of putatively-active members may be more closely linked to short-term environmental change
496 than assembly of the whole community.
497

498 In addition to being more sensitive to disturbance, we find that assembly of the putatively active
499 community was more strongly tied to microbial processes (i.e., respiration rate), than was the
500 whole community. A strong link between biogeochemical rates and the putatively active
501 community is consistent with previous studies (83, 84). More specifically, we observed a
502 negative relationship between respiration rate and absolute values of β NTI for the putatively
503 active community, but no relationship for the whole community. The direction of this relationship
504 is opposite to our hypothesis. While stronger selection should remove mal-adapted individuals,
505 leading to higher biogeochemical rates (23), increased selection in our experiment was imposed
506 by disturbance that appeared to directly suppress respiration rates due to desiccation (85, 86).
507 The simultaneous suppression of respiration and imposition of stronger selection led to the
508 negative relationship between respiration and the strength of selection. The lack of such
509 relationships when considering the whole community indicates that a greater focus on assembly
510 dynamics of putatively active communities could reveal new insights into the multi-component
511 linkages among microbes, the environment, and function.
512

513 Disturbance also impacted OM thermodynamics and respiration rates, potentially initiating an
514 iterative loop among microbial assembly, microbial processes, and the abiotic environment. In
515 this iterative loop the direction of causation between OM thermodynamics and microbial
516 processes (Fig. 1, arrow 3) is not clear due to feedbacks, though we interpret a direction of
517 causation from OM thermodynamics to microbial properties in terms of community assembly
518 (Fig. 1 arrow 4). As such, there may be a loop between OM thermodynamics and microbial
519 processes (i.e., respiration) embedded in a larger loop that also includes microbial properties
520 (i.e., community assembly). Such feedbacks are inherent in complex systems and often lead to
521 non-linear dynamics as observed here in terms of the threshold-like impact of desiccation on
522 multiple system components (87, 88).

523

524 As key elements of the inferred system of feedbacks, the links among OM thermodynamic
525 properties, respiration, and desiccation found here are consistent with recent work tied to the
526 same field system. That is, Garayburu-Caruso et al. (49) also showed decreasing aerobic
527 respiration with decreasing favorability for oxidation (i.e., larger values of ΔG^0_{Cox}) using
528 sediments sourced ~2 years previously from the same field system. In addition, the impacts of
529 desiccation found here are similar to Goldman et al. (62) after re-inundation. This impact of
530 desiccation on respiration contrasts with the Birch effect (89) in soils whereby desiccation
531 followed by re-wetting leads to enhanced respiration. The consistency across hyporheic zone
532 studies and deviation from classical soil phenomena points to potential consistency in governing
533 processes within the hyporheic zone that deviate from processes operating in soil systems.
534 Further evaluation is needed across additional hyporheic zone systems to rigorously evaluate
535 this inference, however.

536

537 In addition to linkages between the environment and microbial aspects of the system, our study
538 revealed connections within the environmental components of the conceptual framework. That
539 is, greater cumulative desiccation caused an increase in ΔG^0_{Cox} , indicating a significant change
540 in OM thermodynamics (Fig. 7). While our data cannot pinpoint governing mechanism(s), we
541 hypothesize that the ΔG^0_{Cox} response may have been tied to increased ion concentration
542 following desiccation. For example, OM chemistry may have been altered due to changes in
543 abiotic sorption, limitations of microbially accessible C due to water potential constraints, and/or
544 osmolyte production and formation of extracellular polymeric substances (90–92).

545

546 Irrespective of mechanisms, the shift in OM thermodynamics in response to desiccation was
547 associated with a decline in respiration. We infer a causal connection between OM
548 thermodynamics and respiration, potentially triggered by desiccation-driven shifts in OM
549 chemistry and/or microbial physiology. This causal connection is supported by recent work (49)
550 and the observation of a continuous function between ΔG^0_{Cox} and respiration rate that
551 transcended experimental treatments. Desiccation therefore likely influences and may even
552 initiate an iterative loop among OM thermodynamics, microbial assembly, and biogeochemistry
553 that underlies cumulative system function.

554

555 Cumulative system function can often be driven by ecosystem control points (52), but we
556 observed relatively little indication of such behavior. That is, estimates of control point influence
557 (CPI) were relatively low across most treatments. CPI is theoretically constrained to range from
558 0.5-1, with lower values indicating smaller influences of control points. In our study, all but one
559 treatment had CPI between ~0.5 and 0.7. The associated distributions of respiration rates did
560 not contain obvious outliers such that we interpret CPI values in the 0.5-0.7 range to be
561 relatively low and not strongly influenced by control points or biogeochemical hot
562 spots/moments (51). The treatment with 31 cumulative days of desiccation diverged from the
563 rest in having a CPI value of ~0.9. This large CPI was due to a single outlier (Fig. 5a) such that
564 most of the cumulative respiration across reactors was contributed by that single reactor. We
565 interpret that single reactor as a biogeochemical hot spot or control point within that

566 experimental treatment. It is unclear, however, what led to such behavior as disturbance did not
567 have any systematic influence on CPI.

568

569 A strength of CPI as a metric is that it allows for direct quantitative comparisons across studies,
570 systems, and scales. Ours is the first study to estimate CPI, however, such that we cannot yet
571 make comparisons to previous work. Through future comparisons it will be possible to evaluate
572 the strengths, weaknesses, and behavior of CPI. We expect that some patterns may emerge
573 such as CPI having a greater likelihood to reach very high values (near 1) in systems with
574 relatively low rates on average. In these conditions, even a modest quantitative increase in
575 biogeochemical rates can lead to a large proportional change such that most cumulative
576 function is from a single point in space and/or time, resulting in large CPI. We also expect that
577 some biogeochemical processes will show greater variation in CPI than others, potentially due
578 to variation in degree of functional redundancy (93). For example, processes such as respiration
579 can be performed by numerous microbial taxa (i.e., there is high functional redundancy), while
580 others are more constrained to a relatively small number of taxa (e.g., ammonia oxidation). We
581 hypothesize that CPI may be lower on average and less variable across systems and scales for
582 biogeochemical processes with greater functional redundancy. Additional work will be needed to
583 test this hypothesis.

584

585 In this study we coupled intrinsic characteristics of natural hyporheic zone sediments with
586 imposed constraints in the form of desiccation to evaluate an *a priori* conceptual framework
587 modified from Hall et al. (6). Our results demonstrated strong and often non-linear connections
588 among desiccation, OM thermodynamics, assembly of the putatively active microbial
589 community, and respiration rates. Collating our results points to further modification of the
590 framework into an *a posteriori* conceptual model containing nested feedback loops (**Fig. 6**). This
591 conceptual model is consistent with the recently proposed unification of microbial ecology
592 around the concepts of external forcing, internal dynamics, and historical contingencies (46).
593 That is, we hypothesize that external forcing imposed by desiccation initiates multiple internal
594 loops that drive biological and chemical dynamics that, in turn, underlie respiration responses to
595 re-wetting that are contingent on desiccation history. The development of conceptual models
596 such as this is key to incorporating additional mechanistic detail into predictive simulation
597 models (e.g., reactive transport codes). We encourage further evaluation and improvement of
598 both our *a priori* and *a posteriori* concepts across environmentally divergent conditions to
599 generate knowledge that is transferable across systems.

600

601 References

602

- 603 1. R. L. Bier, E. S. Bernhardt, C. M. Boot, E. B. Graham, E. K. Hall, J. T. Lennon, D. R.
604 Nemergut, B. B. Osborne, C. Ruiz-González, J. P. Schimel, M. P. Waldrop, M. D.
605 Wallenstein, Linking microbial community structure and microbial processes: an empirical
606 and conceptual overview. *FEMS Microbiol. Ecol.* **91** (2015), doi:10.1093/femsec/fiv113.
- 607 2. A. A. Malik, J. B. H. Martiny, E. L. Brodie, A. C. Martiny, K. K. Treseder, S. D. Allison,
608 Defining trait-based microbial strategies with consequences for soil carbon cycling under
609 climate change. *ISME J.* **14**, 1–9 (2020).

- 610 3. M. Manzella, R. Geiss, E. K. Hall, Evaluating the stoichiometric trait distributions of
611 cultured bacterial populations and uncultured microbial communities. *Environ. Microbiol.*
612 **21**, 3613–3626 (2019).
- 613 4. M. D. Wallenstein, E. K. Hall, A trait-based framework for predicting when and where
614 microbial adaptation to climate change will affect ecosystem functioning. *Biogeochemistry*.
615 **109**, 35–47 (2012).
- 616 5. W. R. Wieder, S. D. Allison, E. A. Davidson, K. Georgiou, O. Hararuk, Y. He, F. Hopkins,
617 Y. Luo, M. J. Smith, B. Sulman, K. Todd-Brown, Y.-P. Wang, J. Xia, X. Xu, Explicitly
618 representing soil microbial processes in Earth system models. *Glob. Biogeochem. Cycles*.
619 **29**, 1782–1800 (2015).
- 620 6. E. K. Hall, E. S. Bernhardt, R. L. Bier, M. A. Bradford, C. M. Boot, J. B. Cotner, P. A. del
621 Giorgio, S. E. Evans, E. B. Graham, S. E. Jones, J. T. Lennon, K. J. Locey, D. Nemergut,
622 B. B. Osborne, J. D. Rocca, J. P. Schimel, M. P. Waldrop, M. D. Wallenstein,
623 Understanding how microbiomes influence the systems they inhabit. *Nat. Microbiol.* **3**,
624 977–982 (2018).
- 625 7. J. A. Gilbert, J. K. Jansson, R. Knight, Earth Microbiome Project and Global Systems
626 Biology. *mSystems*. **3** (2018), doi:10.1128/mSystems.00217-17.
- 627 8. J. Lloyd-Price, C. Arze, A. N. Ananthakrishnan, M. Schirmer, J. Avila-Pacheco, T. W.
628 Poon, E. Andrews, N. J. Ajami, K. S. Bonham, C. J. Brislawn, D. Casero, H. Courtney, A.
629 Gonzalez, T. G. Graeber, A. B. Hall, K. Lake, C. J. Landers, H. Mallick, D. R. Plichta, M.
630 Prasad, G. Rahnavard, J. Sauk, D. Shungin, Y. Vázquez-Baeza, R. A. White, J. Braun, L.
631 A. Denson, J. K. Jansson, R. Knight, S. Kugathasan, D. P. B. McGovern, J. F. Petrosino,
632 T. S. Stappenbeck, H. S. Winter, C. B. Clish, E. A. Franzosa, H. Vlamakis, R. J. Xavier, C.
633 Huttenhower, Multi-omics of the gut microbial ecosystem in inflammatory bowel diseases.
634 *Nature*. **569**, 655–662 (2019).
- 635 9. X. Fu, Y. Li, Y. Meng, Q. Yuan, Z. Zhang, D. Norbäck, Y. Deng, X. Zhang, Y. Sun, *bioRxiv*,
636 in press, doi:10.1101/2020.01.05.893529.
- 637 10. S. König, A. Worrlich, T. Banitz, F. Centler, H. Harms, M. Kästner, A. Miltner, L. Y. Wick, M.
638 Thullner, K. Frank, Spatiotemporal disturbance characteristics determine functional stability
639 and collapse risk of simulated microbial ecosystems. *Sci. Rep.* **8**, 1–13 (2018).
- 640 11. S. Behrens, A. Kappler, M. Obst, Linking environmental processes to the in situ functioning
641 of microorganisms by high-resolution secondary ion mass spectrometry (NanoSIMS) and
642 scanning transmission X-ray microscopy (STXM). *Environ. Microbiol.* **14**, 2851–2869
643 (2012).
- 644 12. S. Norland, K. M. Fagerbakke, M. Heldal, Light element analysis of individual bacteria by x-
645 ray microanalysis. *Appl. Environ. Microbiol.* **61**, 1357–1362 (1995).
- 646 13. L. R. Thompson, J. G. Sanders, D. McDonald, A. Amir, J. Ladau, K. J. Locey, R. J. Prill, A.
647 Tripathi, S. M. Gibbons, G. Ackermann, J. A. Navas-Molina, S. Janssen, E. Kopylova, Y.
648 Vázquez-Baeza, A. González, J. T. Morton, S. Mirarab, Z. Zech Xu, L. Jiang, M. F.
649 Haroon, J. Kanbar, Q. Zhu, S. Jin Song, T. Kosciolka, N. A. Bokulich, J. Lefler, C. J.
650 Brislawn, G. Humphrey, S. M. Owens, J. Hampton-Marcell, D. Berg-Lyons, V. McKenzie,
651 N. Fierer, J. A. Fuhrman, A. Claes, R. L. Stevens, A. Shade, K. S. Pollard, K. D.
652 Goodwin, J. K. Jansson, J. A. Gilbert, R. Knight, A communal catalogue reveals Earth's
653 multiscale microbial diversity. *Nature*. **551**, 457–463 (2017).

- 654 14. M. Wagner, Single-cell ecophysiology of microbes as revealed by Raman
655 microspectroscopy or secondary ion mass spectrometry imaging. *Annu. Rev. Microbiol.* **63**,
656 411–429 (2009).
- 657 15. R. P. Bartelme, J. M. Custer, C. L. Dupont, J. L. Espinoza, M. Torralba, B. Khalili, P. Carini,
658 Influence of Substrate Concentration on the Culturability of Heterotrophic Soil Microbes
659 Isolated by High-Throughput Dilution-to-Extinction Cultivation. *mSphere*. **5** (2020),
660 doi:10.1128/mSphere.00024-20.
- 661 16. R. L. Barnard, C. A. Osborne, M. K. Firestone, Changing precipitation pattern alters soil
662 microbial community response to wet-up under a Mediterranean-type climate. *ISME J.* **9**,
663 946–957 (2015).
- 664 17. S. J. Blazewicz, R. L. Barnard, R. A. Daly, M. K. Firestone, Evaluating rRNA as an
665 indicator of microbial activity in environmental communities: limitations and uses. *ISME J.*
666 **7**, 2061–2068 (2013).
- 667 18. D. C. Cardoso, A. Sandionigi, M. S. Cretoiu, M. Casiraghi, L. Stal, H. Bolhuis, Comparison
668 of the active and resident community of a coastal microbial mat. *Sci. Rep.* **7**, 1–10 (2017).
- 669 19. P. J. Kearns, J. H. Angell, E. M. Howard, L. A. Deegan, R. H. R. Stanley, J. L. Bowen,
670 Nutrient enrichment induces dormancy and decreases diversity of active bacteria in salt
671 marsh sediments. *Nat. Commun.* **7**, 12881 (2016).
- 672 20. D. Shu, J. Guo, B. Zhang, Y. He, G. Wei, rDNA- and rRNA-derived communities present
673 divergent assemblage patterns and functional traits throughout full-scale landfill leachate
674 treatment process trains. *Sci. Total Environ.* **646**, 1069–1079 (2019).
- 675 21. N. I. Wisnoski, M. E. Muscarella, M. L. Larsen, A. L. Peralta, J. T. Lennon, Metabolic
676 insight into bacterial community assembly across ecosystem boundaries. *Ecology*. **101**,
677 e02968 (2020).
- 678 22. J. C. Stegen, X. Lin, A. E. Konopka, J. K. Fredrickson, Stochastic and deterministic
679 assembly processes in subsurface microbial communities. *ISME J.* **6**, 1653–1664 (2012).
- 680 23. E. B. Graham, J. C. Stegen, Dispersal-Based Microbial Community Assembly Decreases
681 Biogeochemical Function. *Processes*. **5**, 65 (2017).
- 682 24. F. Dini-Andreote, J. C. Stegen, J. D. van Elsas, J. F. Salles, Disentangling mechanisms
683 that mediate the balance between stochastic and deterministic processes in microbial
684 succession. *Proc. Natl. Acad. Sci.* **112**, E1326–E1332 (2015).
- 685 25. J. C. Stegen, X. Lin, J. K. Fredrickson, A. E. Konopka, Estimating and mapping ecological
686 processes influencing microbial community assembly. *Front. Microbiol.* **6** (2015),
687 doi:10.3389/fmicb.2015.00370.
- 688 26. J. C. Stegen, X. Lin, J. K. Fredrickson, X. Chen, D. W. Kennedy, C. J. Murray, M. L.
689 Rockhold, A. Konopka, Quantifying community assembly processes and identifying
690 features that impose them. *ISME J.* **7**, 2069–2079 (2013).
- 691 27. J. Zhou, D. Ning, Stochastic Community Assembly: Does It Matter in Microbial Ecology?
692 *Microbiol. Mol. Biol. Rev.* **81** (2017), doi:10.1128/MMBR.00002-17.
- 693 28. J. Grilli, G. Barabás, M. J. Michalska-Smith, S. Allesina, Higher-order interactions stabilize
694 dynamics in competitive network models. *Nature*. **548**, 210–213 (2017).
- 695 29. E. M. Bottos, D. W. Kennedy, E. B. Romero, S. J. Fansler, J. M. Brown, L. M. Bramer, R.
696 K. Chu, M. M. Tfaily, J. K. Jansson, J. C. Stegen, Dispersal limitation and thermodynamic
697 constraints govern spatial structure of permafrost microbial communities. *FEMS Microbiol.*

- 698 *Ecol.* **94** (2018), doi:10.1093/femsec/fiy110.
- 699 30. Y. Feng, R. Chen, J. C. Stegen, Z. Guo, J. Zhang, Z. Li, X. Lin, Two key features
700 influencing community assembly processes at regional scale: Initial state and degree of
701 change in environmental conditions. *Mol. Ecol.* **27**, 5238–5251 (2018).
- 702 31. S. D. Jurburg, I. Nunes, J. C. Stegen, X. Le Roux, A. Priemé, S. J. Sørensen, J. F. Salles,
703 Autogenic succession and deterministic recovery following disturbance in soil bacterial
704 communities. *Sci. Rep.* **7**, 1–11 (2017).
- 705 32. A. Sengupta, J. Indivero, C. Gunn, M. M. Tfaily, R. K. Chu, J. Toyoda, V. L. Bailey, N. D.
706 Ward, J. C. Stegen, Spatial gradients in the characteristics of soil-carbon fractions are
707 associated with abiotic features but not microbial communities. *Biogeosciences*. **16**, 3911–
708 3928 (2019).
- 709 33. E. B. Graham, M. M. Tfaily, A. R. Crump, A. E. Goldman, L. M. Bramer, E. Arntzen, E.
710 Romero, C. T. Resch, D. W. Kennedy, J. C. Stegen, Carbon Inputs From Riparian
711 Vegetation Limit Oxidation of Physically Bound Organic Carbon Via Biochemical and
712 Thermodynamic Processes. *J. Geophys. Res. Biogeosciences*. **122**, 3188–3205 (2017).
- 713 34. J. C. Stegen, T. Johnson, J. K. Fredrickson, M. J. Wilkins, A. E. Konopka, W. C. Nelson, E.
714 V. Arntzen, W. B. Chrisler, R. K. Chu, S. J. Fansler, E. B. Graham, D. W. Kennedy, C. T.
715 Resch, M. Tfaily, J. Zachara, Influences of organic carbon speciation on hyporheic corridor
716 biogeochemistry and microbial ecology. *Nat. Commun.* **9**, 1–11 (2018).
- 717 35. J. C. Stegen, A. Konopka, J. P. McKinley, C. Murray, X. Lin, M. D. Miller, D. W. Kennedy,
718 E. A. Miller, C. T. Resch, J. K. Fredrickson, Coupling among Microbial Communities,
719 Biogeochemistry and Mineralogy across Biogeochemical Facies. *Sci. Rep.* **6**, 1–14 (2016).
- 720 36. P. Starnawski, T. Bataillon, T. J. G. Ettema, L. M. Jochum, L. Schreiber, X. Chen, M. A.
721 Lever, M. F. Polz, B. B. Jørgensen, A. Schramm, K. U. Kjeldsen, Microbial community
722 assembly and evolution in subseafloor sediment. *Proc. Natl. Acad. Sci.* **114**, 2940–2945
723 (2017).
- 724 37. W. Wu, H.-P. Lu, A. Sastri, Y.-C. Yeh, G.-C. Gong, W.-C. Chou, C.-H. Hsieh, Contrasting
725 the relative importance of species sorting and dispersal limitation in shaping marine
726 bacterial versus protist communities. *ISME J.* **12**, 485–494 (2018).
- 727 38. W. Chen, K. Ren, A. Isabwe, H. Chen, M. Liu, J. Yang, Stochastic processes shape
728 microeukaryotic community assembly in a subtropical river across wet and dry seasons.
729 *Microbiome*. **7**, 138 (2019).
- 730 39. I. Martínez, J. C. Stegen, M. X. Maldonado-Gómez, A. M. Eren, P. M. Siba, A. R. Greenhill,
731 J. Walter, The Gut Microbiota of Rural Papua New Guineans: Composition, Diversity
732 Patterns, and Ecological Processes. *Cell Rep.* **11**, 527–538 (2015).
- 733 40. I. D. Ofitseru, M. Lunn, T. P. Curtis, G. F. Wells, C. S. Criddle, C. A. Francis, W. T. Sloan,
734 Combined niche and neutral effects in a microbial wastewater treatment community. *Proc.*
735 *Natl. Acad. Sci.* **107**, 15345–15350 (2010).
- 736 41. J. Zhou, W. Liu, Y. Deng, Y.-H. Jiang, K. Xue, Z. He, J. D. V. Nostrand, L. Wu, Y. Yang, A.
737 Wang, Stochastic Assembly Leads to Alternative Communities with Distinct Functions in a
738 Bioreactor Microbial Community. *mBio*. **4** (2013), doi:10.1128/mBio.00584-12.
- 739 42. X. Jia, F. Dini-Andreote, J. Falcão Salles, Comparing the Influence of Assembly Processes
740 Governing Bacterial Community Succession Based on DNA and RNA Data.
741 *Microorganisms*. **8**, 798 (2020).

- 742 43. R. A. Daly, M. A. Borton, M. J. Wilkins, D. W. Hoyt, D. J. Kountz, R. A. Wolfe, S. A. Welch,
743 D. N. Marcus, R. V. Trexler, J. D. MacRae, J. A. Krzycki, D. R. Cole, P. J. Mouser, K. C.
744 Wrighton, Microbial metabolisms in a 2.5-km-deep ecosystem created by hydraulic
745 fracturing in shales. *Nat. Microbiol.* **1**, 1–9 (2016).
- 746 44. G. E. Leventhal, M. Ackermann, K. T. Schiessl, Why microbes secrete molecules to modify
747 their environment: the case of iron-chelating siderophores. *J. R. Soc. Interface* **16**,
748 20180674 (2019).
- 749 45. C. Ratzke, J. Denk, J. Gore, Ecological suicide in microbes. *Nat. Ecol. Evol.* **2**, 867–872
750 (2018).
- 751 46. J. C. Stegen, E. M. Bottos, J. K. Jansson, A unified conceptual framework for prediction
752 and control of microbiomes. *Curr. Opin. Microbiol.* (2018), doi:10.1016/j.mib.2018.06.002.
- 753 47. E. B. Graham, A. R. Crump, D. W. Kennedy, E. Arntzen, S. Fansler, S. O. Purvine, C. D.
754 Nicora, W. Nelson, M. M. Tfaily, J. C. Stegen, Multi 'omics comparison reveals
755 metabolome biochemistry, not microbiome composition or gene expression, corresponds
756 to elevated biogeochemical function in the hyporheic zone. *Sci. Total Environ.* **642**, 742–
757 753 (2018).
- 758 48. K. Boye, V. Noël, M. M. Tfaily, S. E. Bone, K. H. Williams, J. R. Bargar, S. Fendorf,
759 Thermodynamically controlled preservation of organic carbon in floodplains. *Nat. Geosci.*
760 **10**, 415–419 (2017).
- 761 49. V. A. Garayburu-Caruso, J. C. Stegen, H.-S. Song, L. Renteria, J. Wells, W. Garcia, C. T.
762 Resch, A. E. Goldman, R. K. Chu, J. Toyoda, E. B. Graham, Carbon Limitation Leads to
763 Thermodynamic Regulation of Aerobic Metabolism. *Environ. Sci. Technol. Lett.* (2020),
764 doi:10.1021/acs.estlett.0c00258.
- 765 50. H.-S. Song, J. C. Stegen, E. B. Graham, J.-Y. Lee, V. A. Garayburu-Caruso, W. C. Nelson,
766 X. Chen, J. D. Moulton, T. D. Scheibe, *bioRxiv*, in press, doi:10.1101/2020.02.27.968669.
- 767 51. M. E. McClain, E. W. Boyer, C. L. Dent, S. E. Gergel, N. B. Grimm, P. M. Groffman, S. C.
768 Hart, J. W. Harvey, C. A. Johnston, E. Mayorga, W. H. McDowell, G. Pinay,
769 Biogeochemical Hot Spots and Hot Moments at the Interface of Terrestrial and Aquatic
770 Ecosystems. *Ecosystems* **6**, 301–312 (2003).
- 771 52. E. S. Bernhardt, J. R. Blaszcak, C. D. Ficken, M. L. Fork, K. E. Kaiser, E. C. Seybold,
772 Control Points in Ecosystems: Moving Beyond the Hot Spot Hot Moment Concept.
773 *Ecosystems* **20**, 665–682 (2017).
- 774 53. B. Arora, M. A. Briggs, J. Zarnetske, J. C. Stegen, J. Gomez-Velez, D. Dwivedi, C. I.
775 Steefel, in *Biogeochemistry of the Critical Zone*, A. Wymore, W. Yang, W. Silver, B.
776 McDowell, J. Chorover, Eds. (Springer-Nature, 2020).
- 777 54. F. Boano, J. W. Harvey, A. Marion, A. I. Packman, R. Revelli, L. Ridolfi, A. Wörman,
778 Hyporheic flow and transport processes: Mechanisms, models, and biogeochemical
779 implications. *Rev. Geophys.* **52**, 603–679 (2014).
- 780 55. R. M. Burrows, H. Rutledge, N. R. Bond, S. M. Eberhard, A. Auhl, M. S. Andersen, D. G.
781 Valdez, M. J. Kennard, High rates of organic carbon processing in the hyporheic zone of
782 intermittent streams. *Sci. Rep.* **7**, 1–11 (2017).
- 783 56. B. O. L. Demars, Hydrological pulses and burning of dissolved organic carbon by stream
784 respiration. *Limnol. Oceanogr.* **64**, 406–421 (2019).
- 785 57. H. Fischer, F. Kloep, S. Wilzcek, M. T. Pusch, A River's Liver – Microbial Processes within

- 786 the Hyporheic Zone of a Large Lowland River. *Biogeochemistry*. **76**, 349–371 (2005).
- 787 58. M. H. Kaufman, M. B. Cardenas, J. Buttles, A. J. Kessler, P. L. M. Cook, Hyporheic hot
788 moments: Dissolved oxygen dynamics in the hyporheic zone in response to surface flow
789 perturbations. *Water Resour. Res.* **53**, 6642–6662 (2017).
- 790 59. S. T. Larned, T. Datry, D. B. Arscott, K. Tockner, Emerging concepts in temporary-river
791 ecology. *Freshw. Biol.* **55**, 717–738 (2010).
- 792 60. A. M. Romaní, E. Vázquez, A. Butturini, Microbial Availability and Size Fractionation of
793 Dissolved Organic Carbon After Drought in an Intermittent Stream: Biogeochemical Link
794 Across the Stream–Riparian Interface. *Microb. Ecol.* **52**, 501–512 (2006).
- 795 61. H. F. Birch, Mineralisation of plant nitrogen following alternate wet and dry conditions.
796 *Plant Soil.* **20**, 43–49 (1964).
- 797 62. A. E. Goldman, E. B. Graham, A. R. Crump, D. W. Kennedy, E. B. Romero, C. G.
798 Anderson, K. L. Dana, C. T. Resch, J. K. Fredrickson, J. C. Stegen, Biogeochemical
799 cycling at the aquatic–terrestrial interface is linked to parafluvial hyporheic zone inundation
800 history. *Biogeosciences*. **14**, 4229–4241 (2017).
- 801 63. E. B. Graham, A. R. Crump, C. T. Resch, S. Fansler, E. Arntzen, D. W. Kennedy, J. K.
802 Fredrickson, J. C. Stegen, Coupling Spatiotemporal Community Assembly Processes to
803 Changes in Microbial Metabolism. *Front. Microbiol.* **7** (2016),
804 doi:10.3389/fmicb.2016.01949.
- 805 64. J. M. Zachara, P. E. Long, J. Bargar, J. A. Davis, P. Fox, J. K. Fredrickson, M. D. Freshley,
806 A. E. Konopka, C. Liu, J. P. McKinley, M. L. Rockhold, K. H. Williams, S. B. Yabusaki,
807 Persistence of uranium groundwater plumes: Contrasting mechanisms at two DOE sites in
808 the groundwater–river interaction zone. *J. Contam. Hydrol.* **147**, 45–72 (2013).
- 809 65. L. D. Slater, D. Ntarlagiannis, F. D. Day-Lewis, K. Mwakanyamale, R. J. Versteeg, A.
810 Ward, C. Strickland, C. D. Johnson, J. W. Lane, Use of electrical imaging and distributed
811 temperature sensing methods to characterize surface water–groundwater exchange
812 regulating uranium transport at the Hanford 300 Area, Washington. *Water Resour. Res.* **46**
813 (2010), doi:10.1029/2010WR009110.
- 814 66. E. Arntzen, Effects of fluctuating river flow on groundwater/surface water mixing in the
815 hyporheic zone of a regulated, large cobble bed river - Arntzen - 2006 - River Research
816 and Applications - Wiley Online Library (2006), (available at
817 <https://onlinelibrary.wiley.com/doi/abs/10.1002/rra.947>).
- 818 67. S. Faticchi, S. Manzoni, D. Or, A. Paschalis, A Mechanistic Model of Microbially Mediated
819 Soil Biogeochemical Processes: A Reality Check. *Glob. Biogeochem. Cycles.* **33**, 620–648
820 (2019).
- 821 68. E. B. Graham, A. R. Crump, C. T. Resch, S. Fansler, E. Arntzen, D. W. Kennedy, J. K.
822 Fredrickson, J. C. Stegen, Deterministic influences exceed dispersal effects on
823 hydrologically-connected microbiomes. *Environ. Microbiol.* **19**, 1552–1567 (2017).
- 824 69. J. Brown, N. Zavoshy, C. J. Brislaw, L. A. McCue, “Hundo: a Snakemake workflow for
825 microbial community sequence data” (e27272v1, PeerJ Inc., 2018), ,
826 doi:10.7287/peerj.preprints.27272v1.
- 827 70. R. E. Danczak, A. E. Goldman, R. K. Chu, J. G. Toyoda, V. A. Garayburu-Caruso, N. Tolić,
828 E. B. Graham, J. W. Morad, L. Renteria, J. R. Wells, S. P. Herzog, A. S. Ward, J. C.
829 Stegen, *bioRxiv*, in press, doi:10.1101/2020.02.12.946459.

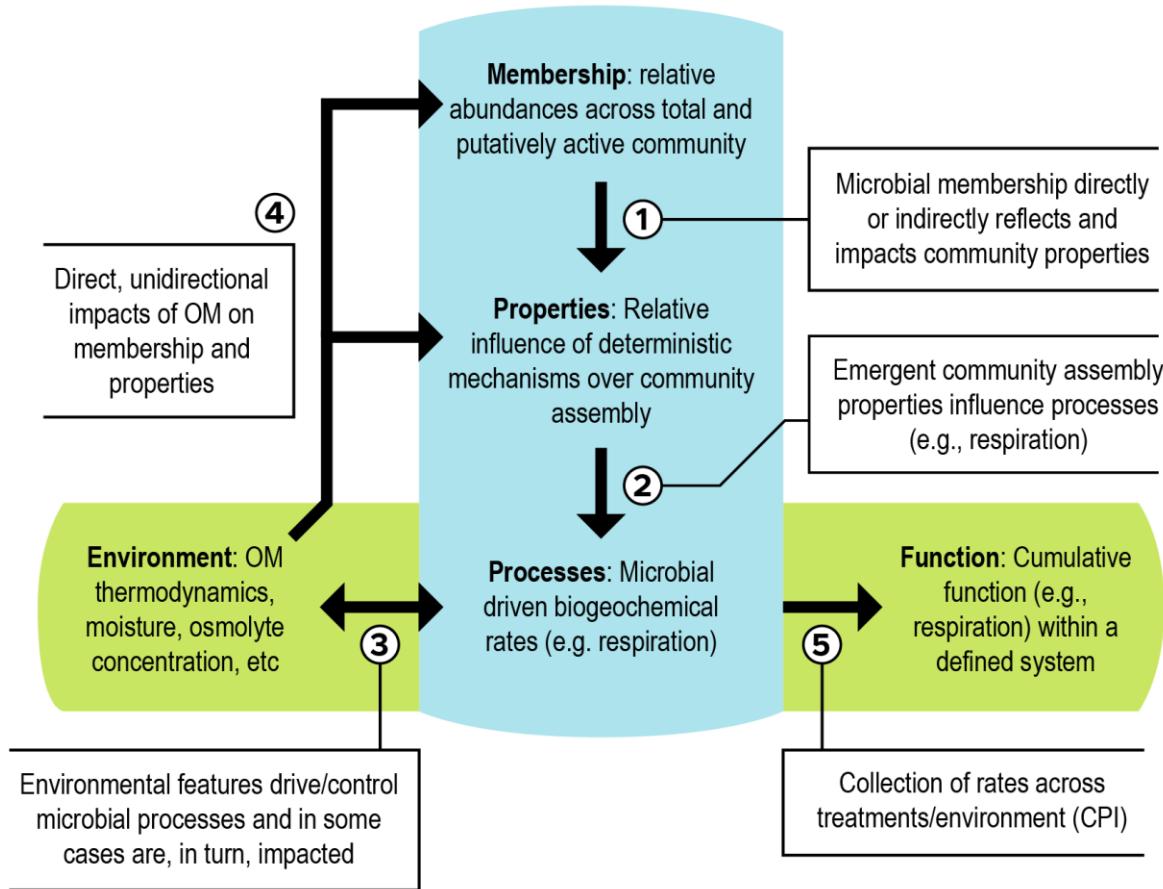
- 830 71. D. E. LaRowe, P. Van Cappellen, Degradation of natural organic matter: A thermodynamic
831 analysis. *Geochim. Cosmochim. Acta.* **75**, 2030–2042 (2011).
- 832 72. B. M. Tripathi, J. C. Stegen, M. Kim, K. Dong, J. M. Adams, Y. K. Lee, Soil pH mediates
833 the balance between stochastic and deterministic assembly of bacteria. *ISME J.* **12**, 1072–
834 1083 (2018).
- 835 73. J. Wang, J. Shen, Y. Wu, C. Tu, J. Soininen, J. C. Stegen, J. He, X. Liu, L. Zhang, E.
836 Zhang, Phylogenetic beta diversity in bacterial assemblages across ecosystems:
837 deterministic versus stochastic processes. *ISME J.* **7**, 1310–1321 (2013).
- 838 74. L. Fillinger, Y. Zhou, C. Kellermann, C. Griebler, Non-random processes determine the
839 colonization of groundwater sediments by microbial communities in a pristine porous
840 aquifer. *Environ. Microbiol.* **21**, 327–342 (2019).
- 841 75. Y. Li, Y. Gao, W. Zhang, C. Wang, P. Wang, L. Niu, H. Wu, Homogeneous selection
842 dominates the microbial community assembly in the sediment of the Three Gorges
843 Reservoir. *Sci. Total Environ.* **690**, 50–60 (2019).
- 844 76. A. Sengupta, J. C. Stegen, A. A. M. Neto, Y. Wang, J. W. Neilson, T. Tatarin, E. Hunt, K.
845 Dontsova, J. Chorover, P. A. Troch, R. M. Maier, Assessing Microbial Community Patterns
846 During Incipient Soil Formation From Basalt. *J. Geophys. Res. Biogeosciences.* **124**, 941–
847 958 (2019).
- 848 77. T. Whitman, R. Neurath, A. Perera, I. Chu-Jacoby, D. Ning, J. Zhou, P. Nico, J. Pett-Ridge,
849 M. Firestone, Microbial community assembly differs across minerals in a rhizosphere
850 microcosm. *Environ. Microbiol.* **20**, 4444–4460 (2018).
- 851 78. E. S. Boyd, D. E. Cummings, G. G. Geesey, Mineralogy Influences Structure and Diversity
852 of Bacterial Communities Associated with Geological Substrata in a Pristine Aquifer.
853 *Microb. Ecol.* **54**, 170–182 (2007).
- 854 79. J. K. Carson, L. Campbell, D. Rooney, N. Clipson, D. B. Gleeson, Minerals in soil select
855 distinct bacterial communities in their microhabitats. *FEMS Microbiol. Ecol.* **67**, 381–388
856 (2009).
- 857 80. S. Doetterl, A. A. Berhe, C. Arnold, S. Bodé, P. Fiener, P. Finke, L. Fuchsleger, M.
858 Griepentrog, J. W. Harden, E. Nadeu, J. Schnecker, J. Six, S. Trumbore, K. Van Oost, C.
859 Vogel, P. Boeckx, Links among warming, carbon and microbial dynamics mediated by soil
860 mineral weathering. *Nat. Geosci.* **11**, 589–593 (2018).
- 861 81. B. Fauvel, H.-M. Cauchie, C. Gantzer, L. Ogorzaly, Influence of physico-chemical
862 characteristics of sediment on the in situ spatial distribution of F-specific RNA phages in
863 the riverbed. *FEMS Microbiol. Ecol.* **95** (2019), doi:10.1093/femsec/fiy240.
- 864 82. B. S. Mauck, J. A. Roberts, Mineralogic Control on Abundance and Diversity of Surface-
865 Adherent Microbial Communities. *Geomicrobiol. J.* **24**, 167–177 (2007).
- 866 83. Z. B. Freedman, K. J. Romanowicz, R. A. Upchurch, D. R. Zak, Differential responses of
867 total and active soil microbial communities to long-term experimental N deposition. *Soil
868 Biol. Biochem.* **90**, 275–282 (2015).
- 869 84. D. J. Levy-Booth, I. J. W. Giesbrecht, C. T. E. Kellogg, T. J. Heger, D. V. D'Amore, P. J.
870 Keeling, S. J. Hallam, W. W. Mohn, Seasonal and ecohydrological regulation of active
871 microbial populations involved in DOC, CO₂, and CH₄ fluxes in temperate rainforest soil.
872 *ISME J.* **13**, 950–963 (2019).
- 873 85. D. S. Baldwin, A. M. Mitchell, The effects of drying and re-flooding on the sediment and soil

- 874 nutrient dynamics of lowland river–floodplain systems: a synthesis. *Regul. Rivers Res.*
875 *Manag.* **16**, 457–467 (2000).
- 876 86. S. Manzoni, J. P. Schimel, A. Porporato, Responses of soil microbial communities to water
877 stress: results from a meta-analysis. *Ecology*. **93**, 930–938 (2012).
- 878 87. S. Pérez Castro, E. E. Cleland, R. Wagner, R. A. Sawad, D. A. Lipson, Soil microbial
879 responses to drought and exotic plants shift carbon metabolism. *ISME J.* **13**, 1776–1787
880 (2019).
- 881 88. J. I. Prosser, J. B. H. Martiny, Conceptual challenges in microbial community ecology.
882 *Philos. Trans. R. Soc. B Biol. Sci.* **375**, 20190241 (2020).
- 883 89. H. F. Birch, M. T. Friend, Humus Decomposition in East African Soils. *Nature*. **178**, 500–
884 501 (1956).
- 885 90. N. Fierer, A. S. Allen, J. P. Schimel, P. A. Holden, Controls on microbial CO₂ production: a
886 comparison of surface and subsurface soil horizons. *Glob. Change Biol.* **9**, 1322–1332
887 (2003).
- 888 91. G. Gionchetta, F. Oliva, A. M. Romani, L. Baneras, Hydrological variations shape diversity
889 and functional responses of streambed microbes. *Sci. Total Environ.* **714**, 136838 (2020).
- 890 92. P. M. Homyak, J. C. Blankinship, E. W. Slessarev, S. M. Schaeffer, S. Manzoni, J. P.
891 Schimel, Effects of altered dry season length and plant inputs on soluble soil carbon.
892 *Ecology*. **99**, 2348–2362 (2018).
- 893 93. S. Louca, M. F. Polz, F. Mazel, M. B. N. Albright, J. A. Huber, M. I. O'Connor, M.
894 Ackermann, A. S. Hahn, D. S. Srivastava, S. A. Crowe, M. Doebeli, L. W. Parfrey, Function
895 and functional redundancy in microbial systems. *Nat. Ecol. Evol.* **2**, 936–943 (2018).
- 896 94. J. G. Caporaso, EMP 16S Illumina Amplicon Protocol (2018),
897 doi:10.17504/protocols.io.nuudeww.
- 898

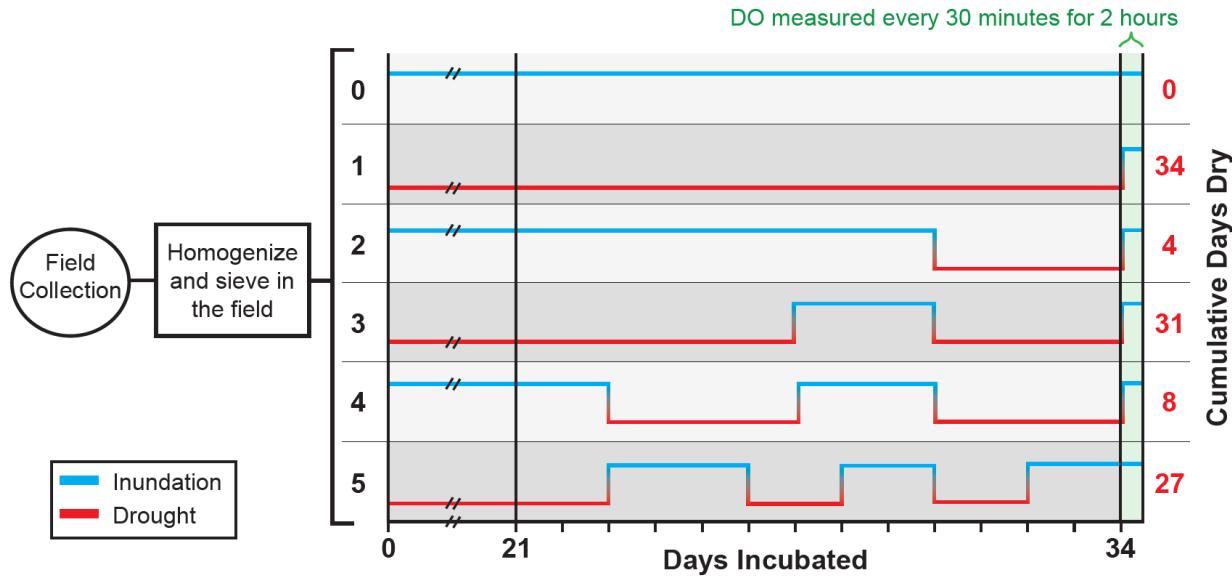
899 Acknowledgements

900 We thank Amy Goldman and Nathan Johnson for developing graphics. **Funding:** The initial
901 experimental stages of this work were supported by the PREMIS Initiative at the Pacific
902 Northwest National Laboratory (PNNL) with funding from the Laboratory Directed Research and
903 Development Program at PNNL, a multi-program national laboratory operated by Battelle for the
904 US Department of Energy under Contract DE-AC05-76RL01830. The later stages of this work
905 (e.g., data analysis, conceptual interpretation, manuscript development) were supported by the
906 U.S. Department of Energy-BER program, as part of an Early Career Award to JCS at PNNL. A
907 portion of the research was performed using EMSL, a DOE Office of Science User Facility
908 sponsored by the Office of Biological and Environmental Research. **Author contributions:** JCS
909 and SF conceptualized and designed the study; JT, JW, LR, RC, SF, and VAG-C performed the
910 experiment; AS, JCS, RED, and SF analyzed data; AS, JCS, and SF drafted the manuscript and
911 all authors contributed to further writing. **Competing Interests:** The authors declare no
912 competing interests. **Data and materials availability:** Data are available on the ESS-DIVE
913 archive at the following link (a data package will be published prior to the manuscript being
914 published, and the associated link will be provided). Sequence data is available on NCBI's
915 Sequence Read Archive PRJNA641165.

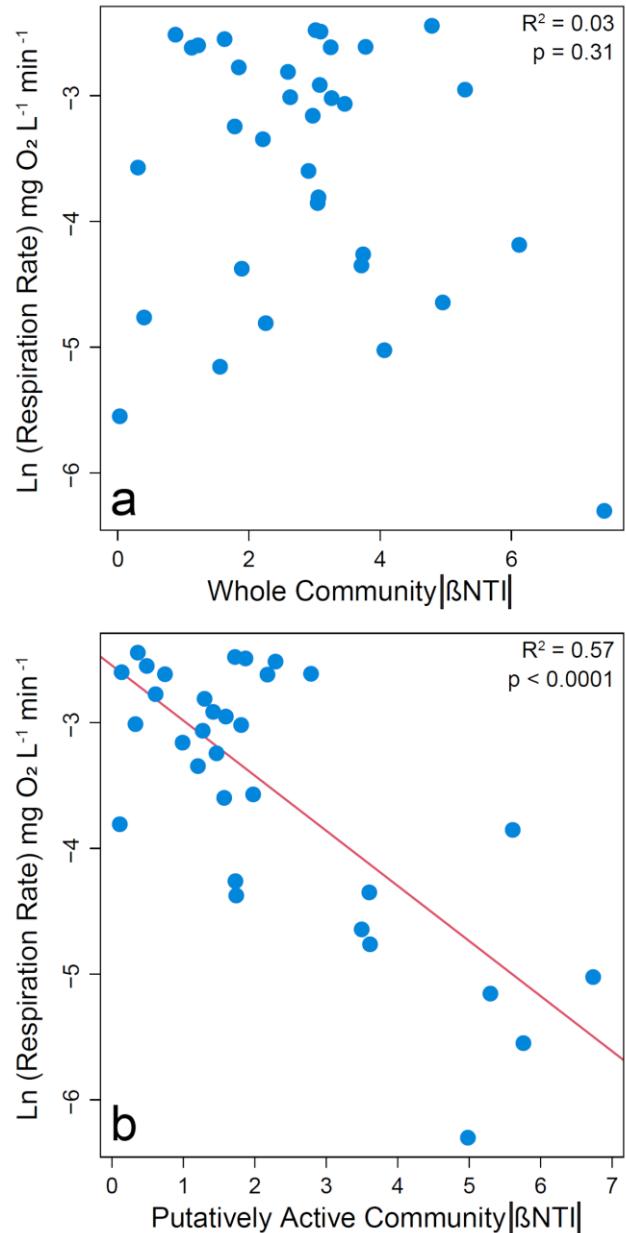
916 **Figures**
917



918
919
920 **Figure 1. Integrated conceptual framework.** The conceptual figure (modified from Hall et al.
921 (6)) details relationships (indicated by numbered arrows) between cumulative properties of the
922 microbial community (e.g., microbial membership, community assembly properties,
923 biogeochemical rates), environmental features (e.g., organic matter thermodynamics), and
924 emergent ecosystem function

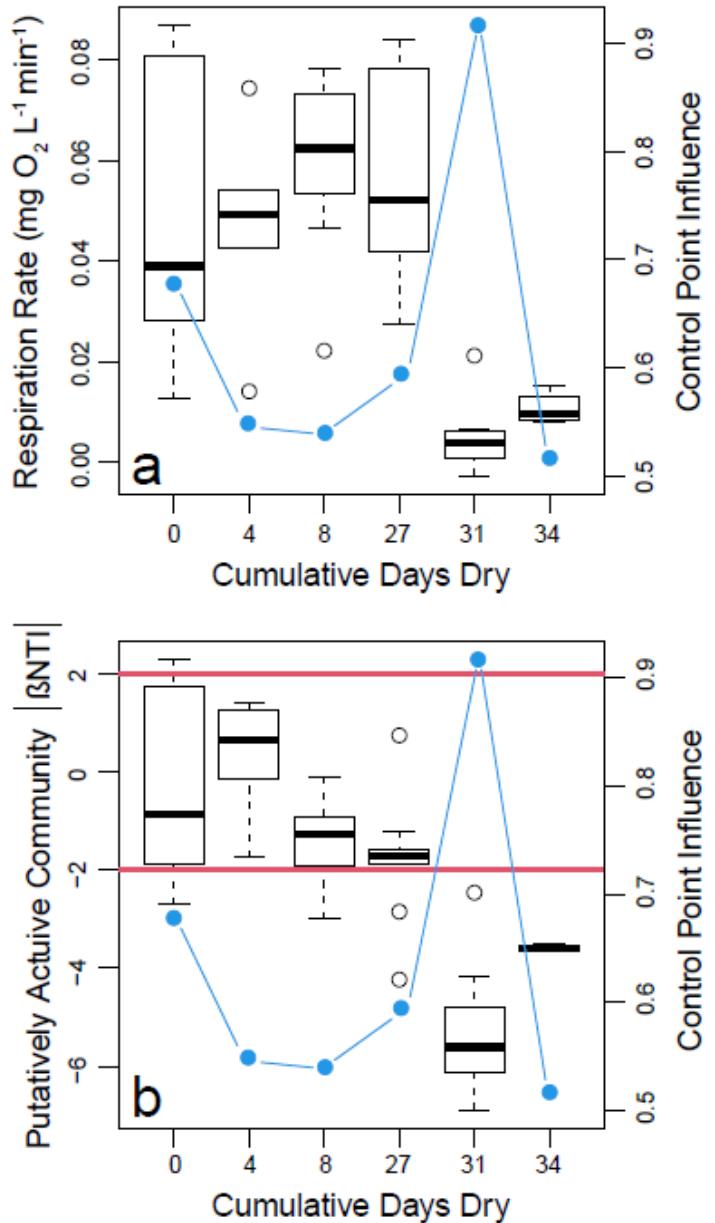


925
926
927
928 **Figure 2. Experimental design of batch reactor incubations subjected to six treatment**
929 **regimes of inundated (blue line) and dry (red line) conditions.** Black values on the left
930 indicate the number of inundated/dry transitions, including the final inundation that all treatments
931 experienced immediately prior to the measurement of respiration. Red values on the right
932 indicate the number of cumulative dry days (e.g., treatments with 1 or 3 transitions experienced
933 34 or 31 cumulative dry days, respectively. Transitions between inundated and dry conditions
934 started on day 24. All treatments were held at either an inundated or dry state prior to day 24.

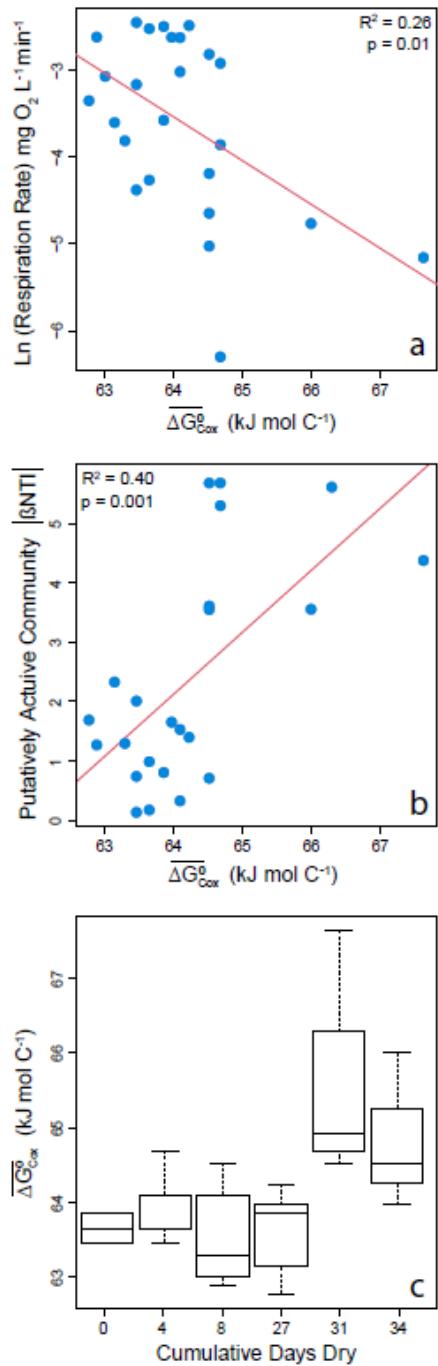


935
936

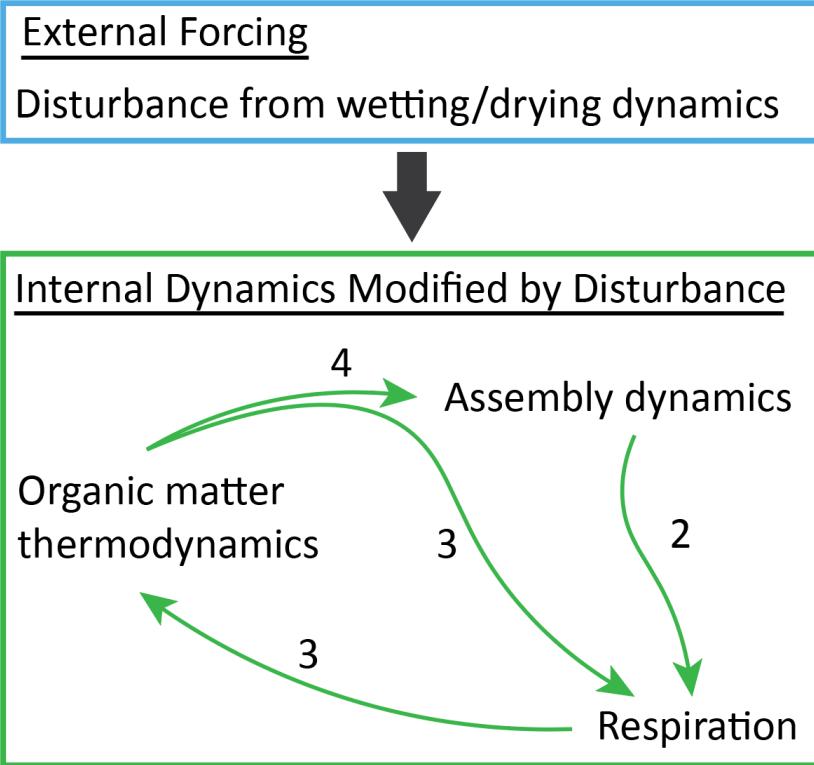
937 **Figure 3. Natural log transformed respiration rates (i.e., O₂ consumption) as a function of**
938 **the absolute value of βNTI for (a) whole communities or (b) putatively active**
939 **communities.** Larger absolute values of βNTI indicate stronger influences of deterministic
940 assembly. Nonlinearity was observed because the respiration rate has a lower limit of 0 such
941 that its relationship with βNTI was fit as a negative exponential function. The significant
942 regression model is shown as a red line, and statistics are provided on each panel.



943
944 **Figure 4. Boxplot representations of respiration rate (a) and putatively active community**
945 **βNTI (b) distributions as a function of the cumulative number of days reactors were in a**
946 **dried state.** Each value along the horizontal axis represents a different experimental treatment.
947 On both panels the right hand axis provides estimates of control point influence (blue circles and
948 lines) across the treatments. Horizontal red lines in (b) indicate significance thresholds; values
949 below -2 indicate deterministic homogenous selection, values above +2 indicate deterministic
950 variable selection, and values between -2 and +2 indicate stochastic assembly.



951
952 **Figure 5. Microbial processes and properties as a function of OM thermodynamics, and**
953 **impacts of disturbance on OM thermodynamics.** (a) Respiration rates (natural log
954 transformed) decreased with decreasing favorability for oxidation (larger values of ΔG_{Cox}⁰). (b)
955 The strength of deterministic selection measured as the absolute value of βNTI increased with
956 decreasing favorability for oxidation. Regression models are shown as red lines and statistics
957 are provided on each panel. (c) Boxplot representations of the distributions of OM
958 thermodynamics across experimental treatments. Significant increases were observed for
959 treatments with 31 or 34 cumulative dry days. See text and Figure S7 for a description of
960 statistics.

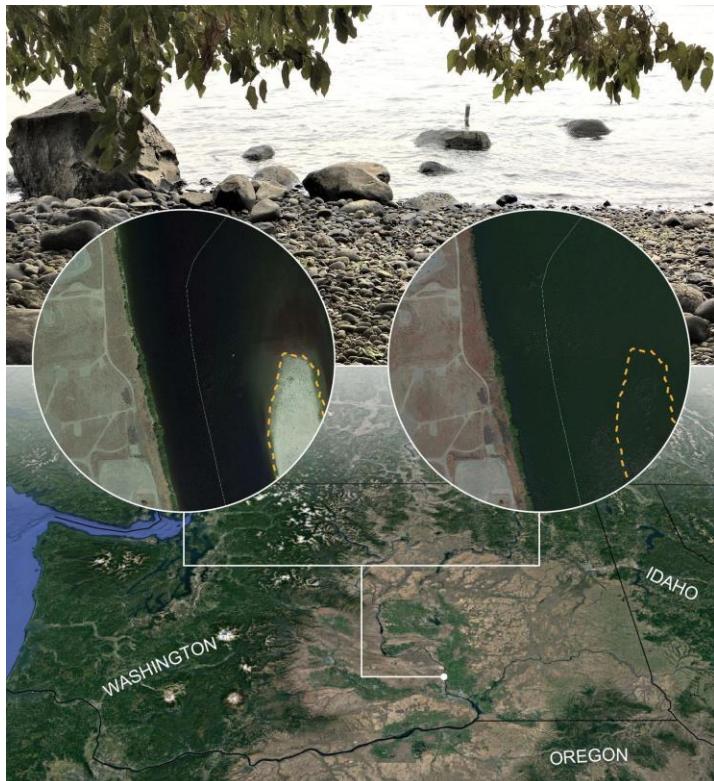


961
962

963 **Figure 6. Integrated conceptual interpretation of results from this study.** Collectively, our
964 results indicate that the external forcing imposed by disturbance leads to feedback between
965 assembly of the putatively active community and respiration rates that is modulated by coupled
966 dynamics in organic matter thermodynamics. Relative to Fig. 1, here external and internal
967 aspects of the environment are separated. The arrows within the internal dynamics component
968 are analogous to arrows 2,3, and 4 in Figure 1. The arrow from external to internal is not
969 considered in Figure 1, and represents the impact of external forcing on all aspects of the
970 internal system. These impacts are both direct effects of disturbance and indirect effects
971 mediated through the internal feedback that collectively lead to impacts of re-wetting that are
972 contingent on desiccation history.

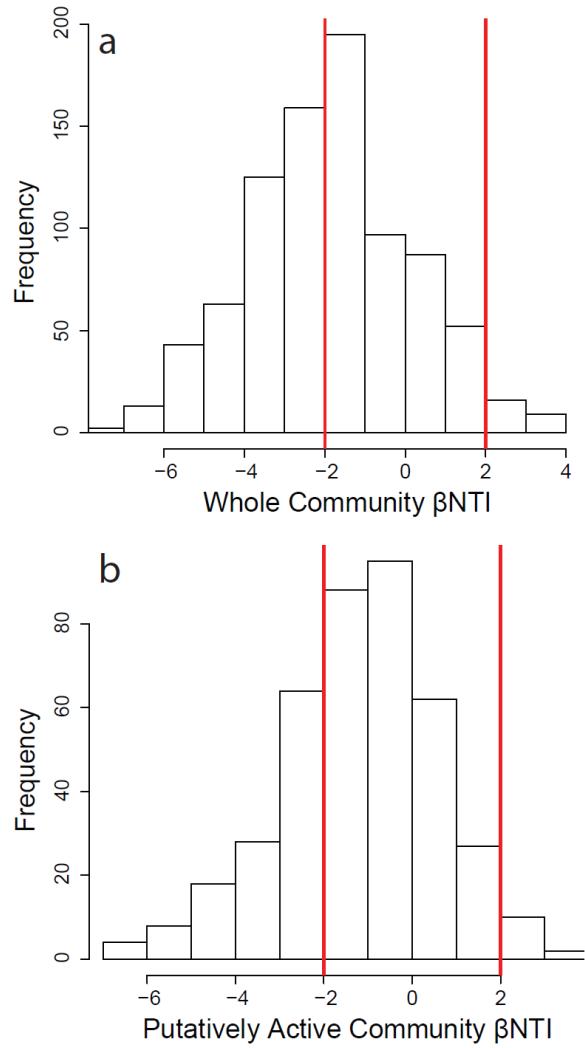
973 **Supplementary Materials**

974



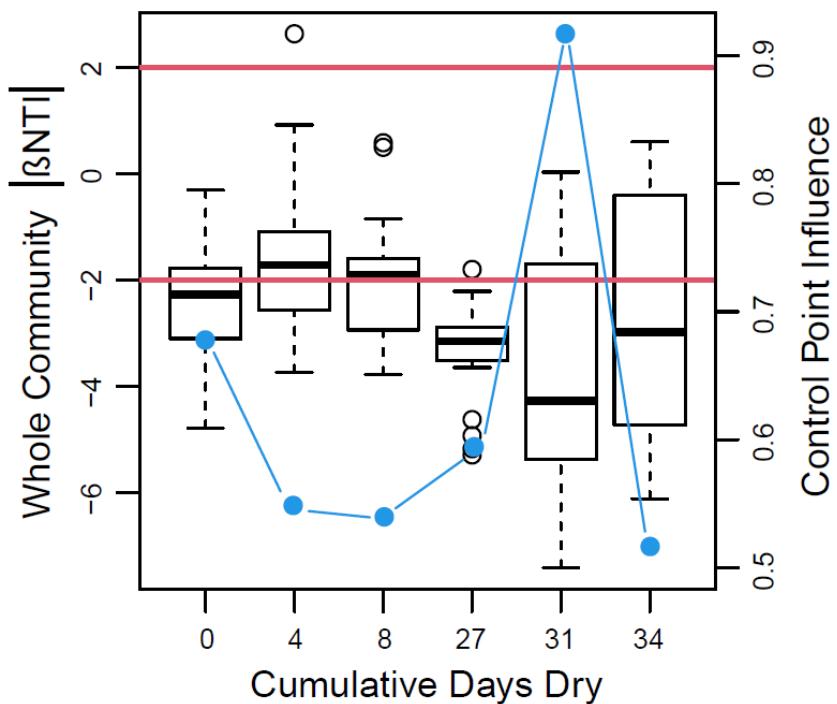
975
976

977 **Supplementary Figure S1. Field location from which sediments were collected.** The site is
978 along the Columbia River in southeast Washington State, as shown in the bottom portion of the
979 image. The system is characterized by variable inundation of the river bed sediments and
980 associated hyporheic zone. The middle images emphasize this variability in inundation. In those
981 images the dashed yellow line is in the same location and outlines the shoreline of an island that
982 is exposed during low water and inundated during high water. The photo at the top of the image
983 is from the location of sediment collection. As can be seen, the system contains large cobbles
984 and gravels, which are partially responsible for high hydrologic conductivity within the hyporheic
985 zone, resulting in aerobic conditions during both inundated and non-inundated conditions.

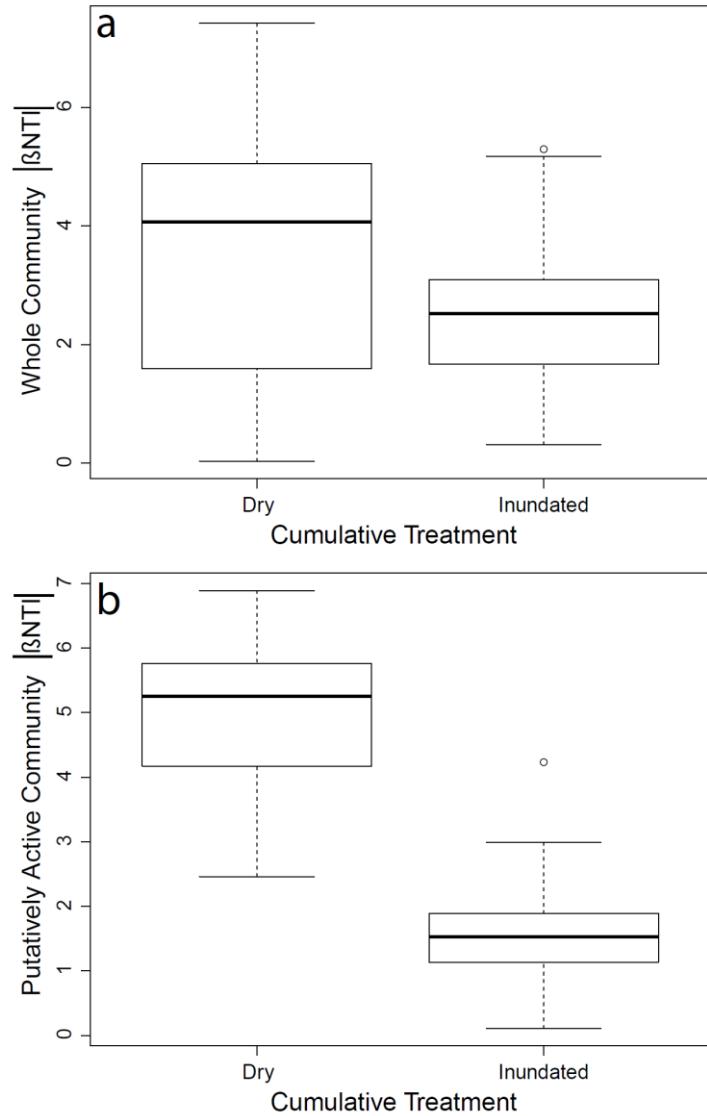


986
987

988 **Supplementary Figure S2. Histograms of β NTI associated with whole communities (a)**
989 **and putatively active communities (b).** Vertical red lines are the significant thresholds (-2 and
990 +2). Values below -2 indicate deterministic homogenous selection, values above +2 indicate
991 deterministic variable selection, and values between -2 and +2 indicate stochastic assembly.

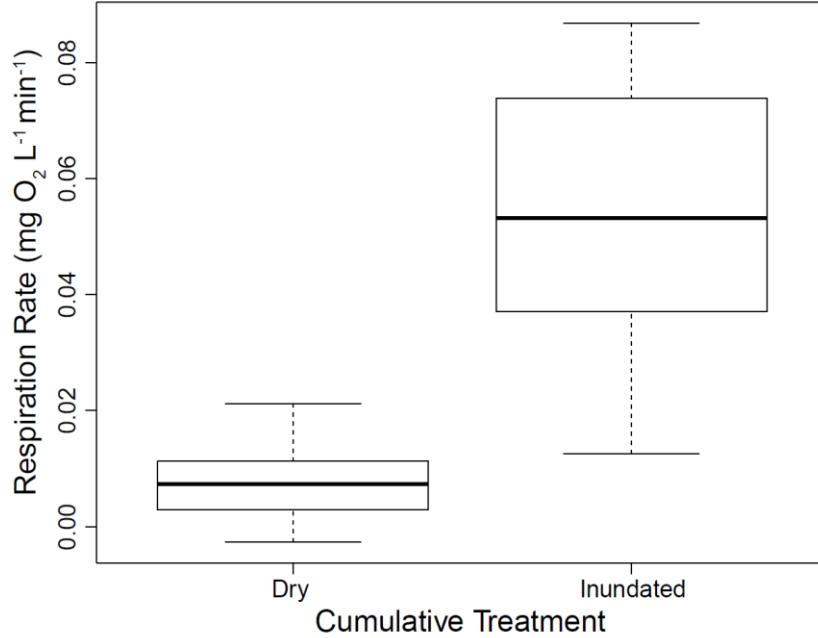


992
993 **Supplementary Figure S3. Boxplot representations of whole community βNTI**
994 **distributions as a function of the cumulative number of days reactors were in a dried**
995 **state.** Each value along the horizontal axis represents a different experimental treatment. The
996 right hand axis provides estimates of control point influence (blue circles and lines) across the
997 treatments. Horizontal red lines indicate significance thresholds for βNTI ; values below -2
998 indicate deterministic homogenous selection, values above +2 indicate deterministic variable
999 selection, and values between -2 and +2 indicate stochastic assembly.



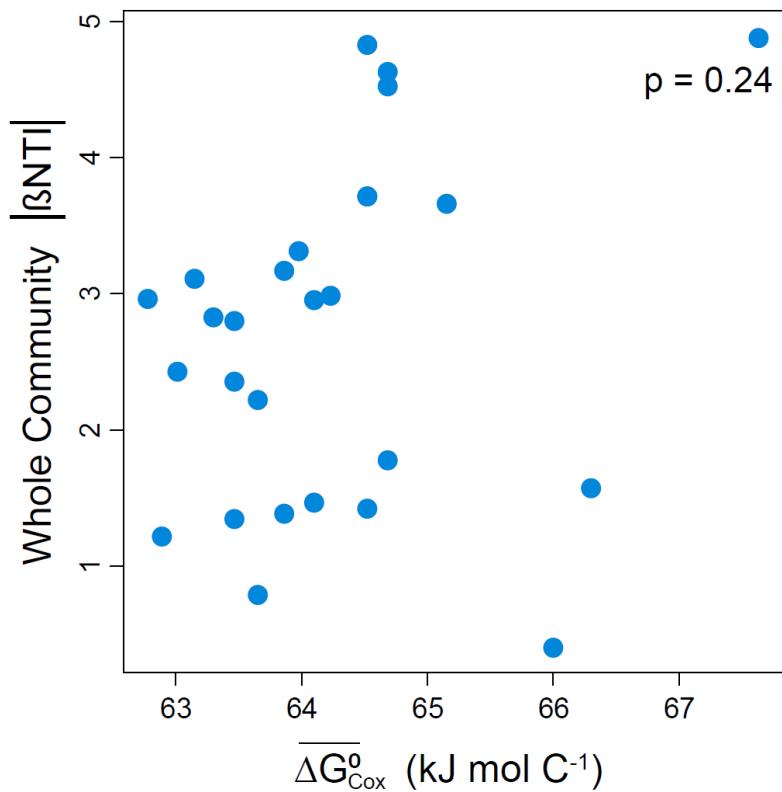
1000
1001
1002
1003
1004
1005
1006

Supplementary Figure S4. Boxplot representations of distributions of the absolute values of β_{NTI} for the whole community (a) and the putatively active community (b) across two different categories of disturbance. On the horizontal axis 'Dry' indicates data combined from across treatments with 31 or 34 cumulative dry days, while 'Inundated' indicates data combined from across treatments with 0-27 cumulative dry days.



1007
1008
1009
1010
1011
1012
1013

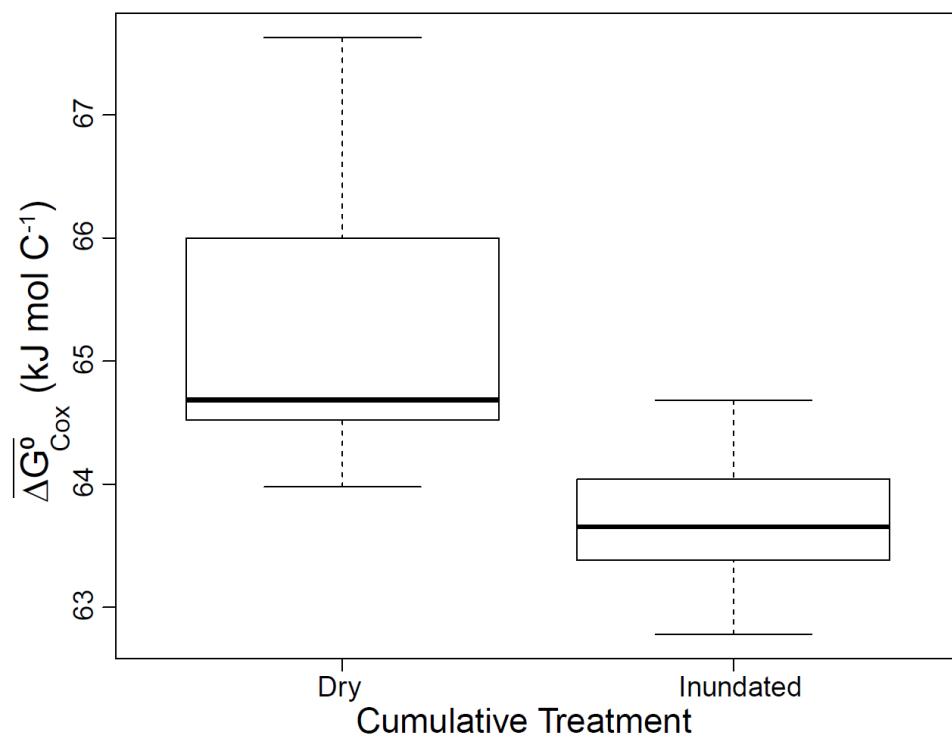
Supplementary Figure S5. Boxplot representations of respiration rate distributions across two different categories of disturbance. On the horizontal axis 'Dry' indicates data combined from across treatments with 31 or 34 cumulative dry days, while 'Inundated' indicates data combined from across treatments with 0-27 cumulative dry days. The two distributions were significantly different per a pairwise Mann-Whitney test ($W = 5, p < 0.001$).



1014
1015
1016
1017
1018

Supplementary Figure S6. Community assembly as a function of organic matter thermodynamics. The strength of deterministic assembly associated with the whole community as measured by βNTI was not related to organic matter thermodynamics.

1019



1020

1021 **Supplementary Figure S7. Boxplot representations of organic matter thermodynamics**
1022 **across two different categories of disturbance.** On the horizontal axis, 'Dry' indicates data
1023 combined from across treatments with 31 or 34 cumulative dry days, while 'Inundated' indicates
1024 data combined from across treatments with 0-27 cumulative dry days. The two distributions
1025 were significantly different per a pairwise Mann-Whitney test ($W = 189$, $p = < 0.001$).

1026

1027 **Data file S1. Time series of moisture content within vial batch reactors.** Columns indicate
1028 the date (month/day/year), sample name that maps to the metadata file found on ESS-DIVE, the
1029 number of inundated/dessicated transitions (referred to as cycles for compatibility with R
1030 Scripts provided in ESS-DIVE data package), the estimated dry sediment mass in grams, and
1031 the mass of water in grams per gram of dry sediment.