Supplementary Information

Table S1. Good's coverage estimation for concatenated MLSA sequences and individual loci. Values are calculated for unique sequences or for operational taxonomic units defined at a 0.01 nucleotide dissimilarity cutoff.

Phylogroup	concatenated		atpD		gyrB		recA		rpoB		trpB	
	unique	0.01	unique	0.01	unique	0.01	unique	0.01	unique	0.01	unique	0.01
MAN125	0.64	1	1	1	0.87	1	0.91	1	1	1	0.96	1
MAN196	0.62	1	0.98	1	0.91	1	0.98	1	1	1	0.96	1
WA1063	0.47	0.95	0.95	1	0.95	0.95	0.89	0.95	0.79	1	0.84	0.95
MS200	0.3	1	0.89	1	0.67	1	0.67	0.96	0.67	1	0.67	1
MS152	0.18	0.94	0.88	1	0.71	0.94	0.47	0.94	0.82	1	0.41	0.88
F34	0.3	1	0.83	0.97	0.78	1	0.97	1	0.89	1	0.78	0.97

Table S2. Reported values are *p*-values for the Shimodaira-Hasegawa test for phylogenetic incongruence. Rows represent maximum likelihood topologies and columns the nucleotide alignments used to evaluate incongruence. Corresponding phylogenic trees were constructed as defined in the methods and are depicted in Figure S4.

	atpD	gyrB	recA	rpoB	<i>trpB</i>	concatenated
atpD		0	0	0	0	0
gyrB	7.0E-04		1.8E-03	4.8E-02	2.3E-03	2.3E-03
recA	0	3.3E-03		7.5E-03	0	0
rpoB	0	0	0		0	0
<i>trpB</i>	4.0E-04	3.5E-03	1.7E-03	5.8E-02		3.3E-03
concatenated	1.6E-01	4.5E-01	3.8E-01	3.4E-01	4.7E-01	



Figure S1. The maximum likelihood tree was constructed with a GTRGAMMA evolution model using partial SSU rRNA gene sequences from three representatives for each phylogroup. Scale bar represents nucleotide substitutions per site. Branches with bootstrap support values greater than 75% are labeled, and the outgroup is *Mycobacterium smegmatis*. Strains are colored to reveal phylogroup membership as indicated in the color legend provided.



Figure S2. Structure analysis was performed on all sequences, irrespective of phylogroup membership. (This approach differs from that depicted in Figure 1 which shows the results of independent Structure analyses performed on each separate phylogroup.) The colored bars in outer rings indicate genetic contributions from different ancestral populations as indicated in the legend (a). Since WA1063 and MS200 share an ancestral population, we performed an additional Structure analysis that included only sequences from these phylogroups, and this analysis reveals clear subdivision between these phylogroups (b). Taxa names in the inner ring are colored to indicate the phylogroup designation of each strain as defined in the legend. The tree was constructed from concatenated MLSA loci nucleotide sequences using maximum likelihood with a GTRGAMMA evolution model. Scale bar represents nucleotide substitutions per site. The root was defined by *Mycobacterium smegmatis*. Nodes with bootstrap confidences > 80 are indicated with gray circles, and precise bootstrap values are found in Figure S4A.



Figure S3. The number of unique MLSA haplotypes is plotted against the percent of total isolates sampled for MLSA (Table 2). The rarefaction curves for phylogroups MS200 and F34 are identical and are depicted with dashed lines. Good's coverage estimations for each phylogroup can be found in Table S1.







----- 0.01







---- 0.01



E



trpB



Figure S4. Phylogenetic trees were constructed from nucleotide sequences of concatenated MLSA loci (A), *atpD* (B), *gryB* (C), *recA* (D), *rpoB* (E), *trpB* (F) by maximum likelihood with a GTRGAMMA evolution model. The scale bar represents nucleotide substitutions per site. Branches with bootstrap support values > 75 are labeled. For Figure S4A, representative isolates from PubMLST were included to orient phylogroup taxonomy to existing *Streptomyces* type strains. The root was defined by *Mycobacterium smegmatis* in each case. The outer color strip depicts phylogroup membership consistent with Figure 2 [green (MAN125), yellow (MAN196), teal (WA1063), blue (MS200), pink (MS152), purple (F34)]. The key for phylogroup isolate names can be found in Table 1.



Figure S5. Circles represent pairwise comparisons of Bray-Curtis dissimilarity and geographic distance between sites for all phylogroups. Bray-Curtis dissimilarity was calculated from unique allele composition across sites. The Mantel coefficient (r) and *p*-value under the null hypothesis of no correlation between the matrixes are reported.