Iroki: automatic customization for phylogenetic trees

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Abstract

Background

Phylogenetic trees are an important analytical tool for examining species and community diversity, and the evolutionary history of species. In the case of microorganisms, decreasing sequencing costs have enabled researchers to generate ever-larger sequence datasets, which in turn have begun to fill gaps in the evolutionary history of microbial groups. However, phylogenetic analyses of large sequence datasets present challenges to extracting meaningful trends from complex trees. Scientific inferences made by visual inspection of phylogenetic trees can be simplified and enhanced by customizing various parts of the tree, including label color, branch color, and other features. Yet, manual customization is time-consuming and error prone, and programs designed to assist in batch tree customization often require programming experience. To address these limitations, we developed Iroki, a program for fast, automatic customization of phylogenetic trees. Iroki allows the user to incorporate information on a broad range of metadata for each experimental unit represented in the tree.

Results

Iroki was applied to four existing microbial sequence datasets to demonstrate its utility in data exploration and presentation. Specifically, we used Iroki to highlight connections between viral phylogeny and host taxonomy, explore the abundance of microbial groups associated with Shiga toxin-producing *Escherichia coli* (STEC) in cattle, examine short-

term temporal dynamics of virioplankton communities, and to search for trends in the biogeography of Zetaproteobacteria.

Conclusions

Iroki is an easy-to-use application having both command line and web-browser implementations for fast, automatic customization of phylogenetic trees based on user-provided categorical or continuous metadata. Iroki enables hypothesis testing through improved visualization of phylogenetic trees, streamlining the process of biological sequence data exploration and presentation.

Availability

Iroki can be accessed through a web browser application or via installation through RubyGems, from source, or through the Iroki Docker image. All source code and documentation is available under the GPLv3 license at https://github.com/mooreryan/iroki. The Iroki web-app is accessible at www.iroki.net or through the VIROME portal (http://virome.dbi.udel.edu), and its source code is released under GPLv3 license at https://github.com/mooreryan/iroki_web. The Docker image can be found here: https://hub.docker.com/r/mooreryan/iroki.

Keywords

Phylogeny, visualization, sequence analysis, bioinformatics, metagenomics

62 **Iroki: automatic customization for phylogenetic trees**

63

64 **Background**

65 Community and population ecological studies often use phylogenetic trees as a means for
66 assessing the diversity and evolutionary history of organisms. In the case of
67 microorganisms, the declining cost of sequencing has enabled researchers to gather ever-
68 larger sequence datasets from unknown microbial populations within environmental
69 samples. While large sequence datasets have begun to fill in the gaps in the evolutionary
70 history of microbial groups [1–5]; they have also posed new analytical challenges as
71 extracting meaningful trends within such highly dimensional datasets can be cumbersome.
72 In particular, scientific inferences made by visual inspection of phylogenetic trees can be
73 simplified and enhanced by customizing various parts of the tree including label and branch
74 color, branch width, and other features. Though several tree visualization packages allow
75 for manual modifications [6–9], the process can be time consuming and error prone
76 especially when the tree contains many nodes. Moreover, these packages are typically not
77 capable of batch customization without prior computer programming experience [10–13].

78

79 Iroki, a program for fast, automatic customization of phylogenetic trees, was developed to
80 address these limitations and enable users to incorporate a broad array of metadata
81 information for each experimental unit represented in the tree. Iroki is available for use
82 through a web browser interface at www.iroki.net, through the VIROME portal
83 (http://virome.dbi.udel.edu), and through a UNIX command line tool. Results are saved in

the widely used Nexus format with color metadata tailored for use with FigTree [8] (a freely available and efficient tree viewer).

Implementation

Iroki enhances visualization of phylogenetic trees by coloring node labels and branches according to categorical metadata criteria or numerical data such as abundance information. Iroki can also rename nodes in a batch process according to user specifications so that node names are more descriptive. A tree file in Newick format containing a phylogenetic tree is always required. Additional required input files depend on the operation(s) desired. Coloring functions require a color map or a biom [14] file. Node renaming functions require a name map. The color map, name map, and biom files are created by the user and, along with the Newick file, form the inputs for Iroki.

Explicit tree coloring

Iroki's principle functionality involves coloring node labels and/or branches based on information provided by the user in the color map. The color map text file contains either two or three tab-delimited columns depending on how branches and labels are to be colored. Two columns, pattern and color, are used when labels and branches are to have the same color. Three columns, pattern, label color, and branch color, are used when branches and labels are to have different colors. Patterns are searched against node labels either as regular expressions or exact string matches.

Entries in the color column can be any of the 657 named colors in the R programming language [15] (e.g., skyblue, tomato, goldenrod2, lightgray, black) or any valid hexadecimal color code (e.g., #FF78F6). In addition, Iroki provides a 19 color palette with complementary colors based on Kelly's color scheme for maximum contrast [16]. Nodes on the tree that are not in the color map will remain black.

Depending on user-specified options, a pattern match to node label(s) will trigger coloring of the label and/or the branch directly connected to that label. Inner branches will be colored to match their descendent branches if all descendants are the same color, allowing quick identification of common ancestors and clades that share common metadata.

Tree coloring based on numerical data

Iroki provides the ability to generate color gradients based on numerical data, such as absolute or relative abundance, from a tab-delimited biom format file [14]. Single-color gradients use color saturation to illustrate numerical differences, with nodes at a higher level being more saturated than those at a lower level. For example, highly abundant nodes will be represented by more highly saturated colors. Two-color gradients show numerical differences through both color mixing and luminosity. Additionally, the biom file may specify numerical information for one group (e.g., abundance in a particular sample) or for two groups (e.g., abundance in the treatment group vs. abundance in the control group). For biom files with one group, single- or two-color gradients may be used. However, biom files specifying two-group metadata may only use the two-color gradient.

Renaming nodes

Some packages for generating phylogenetic trees restrict the use of special characters and spaces or require node names to be shorter than a specified length or (RAxML [17], PHYLIP [18], etc.). Name restrictions present challenges to scientific interpretation of phylogenetic trees. Iroki's renaming function uses a two-column, tab-delimited name map to associate current node names, exactly matching those in the tree file, with new names. The new name column has no restrictions on name length or character type. Iroki ensures name uniqueness by appending integers to the ends of names, if necessary.

Combining the color map, name map, and biom files

Iroki can be used to make complex combinations of customizations by combining the color map, name map, and biom files. For example, a biom file can be used to apply a color gradient based on numerical data to the labels of a tree, a color map can be used to separately color the branches based on user-specified conditions, and a name map can be used to rename nodes in a single command or web request. Iroki follows a specific order of precedence when applying multiple customizations. The color gradient inferred from the biom file is applied first. Next, the color map is applied to specified labels or branches, overriding the gradient applied in the previous step, if necessary. Finally, the name map is used to map current names to the new names (Fig. 1).

Output

Iroki outputs the modified tree in the Nexus format. When building the phylogenetic tree, FigTree uses the Nexus format file and interprets the color metadata output of Iroki.

Results & Discussion

Global diversity of bacteriophage

Viruses are the most abundant biological entity on Earth, providing an enormous reservoir of genetic diversity, driving evolution of their hosts, influencing composition of microbial communities, and affecting global biogeochemical cycles [19,20]. The viral taxonomic system developed by the International Committee on Taxonomy of Viruses (ICTV) is based on a suite of physical characteristics of the virion rather than on genome sequences. Noting this limitation, the phage proteomic tree was created to provide a genome-based taxonomic system for bacteriophage classification [21]. The phage proteomic tree was recently updated to include hundreds of new phage genomes from the Phage SEED reference database [22], as well as long assembled contigs from viral shotgun metagenomes (viromes) collected from the Chesapeake Bay (SERC sample) [23] and the Mediterranean Sea [24].

Taxonomy and host information metadata was collected for the viral genome sequences, a color map was created to assign colors based on viral family and host phyla, and Iroki was used to add color metadata to branches and labels of the phage proteomic tree. Since a large number of colors were required on the tree, Iroki's Kelly color palette was used to provide clear color contrasts. The tree was rendered with FigTree (Fig. 2).

Adding color to the phage proteomic tree with Iroki shows trends in the data that would be difficult to discern without color. Uncultured phage contigs from the Chesapeake Bay and Mediterranean viromes make up a large portion of all phage sequences shown on the tree, and are widely distributed among known phage. In general, viruses in the same family claded together, e.g., branch coloring highlights large groups of closely related Siphoviridae and Myoviridae. This label-coloring scheme also shows that viruses infecting hosts within same phylum are, in general, phylogenetically similar. For example, viruses within one of the multiple large groups of Siphoviridae across the tree infect almost exclusively host species within the same phylum, e.g., Siphoviridae infecting Actinobacteria clade away from Siphoviridae infecting Firmicutes or Proteobacteria.

Bacterial community diversity and prevalence of E. coli in beef cattle

Shiga toxin-producing *Escherichia coli* (STEC) are dangerous human pathogens that colonize the lower gastrointestinal tracts of cattle and other ruminants. STEC-contaminated beef and STEC shed in the feces of these animals are major sources of foodborne illness. To identify possible interactions between STEC populations and the commensal cattle microbiome, a recent study examined the diversity of the bacterial community associated with beef cattle hide [25]. Fecal and hide samples were collected over twelve weeks and SSU rRNA amplicon libraries were constructed and analyzed by Illumina sequencing [26]. The study indicated that the community structure of hide bacterial communities was altered when the hides were positive for STEC contamination.

194

195 Iroki was used to visualize changes in the relative abundance of each cattle hide bacterial
196 OTU according to the presence or absence of STEC. A Mann-Whitney U test comparing
197 OTU abundance between STEC positive and STEC negative samples was performed, and
198 those bacterial OTUs showing a significant change in relative abundance ($p < 0.5$) were
199 placed on a phylogenetic tree according to the 16S rRNA sequence. Branches of the tree
200 were colored based on whether there was a significant change in relative abundance with
201 STEC contamination (red: $p < 0.1$, blue: $p >= 0.1$). Node labels were colored along a
202 blue-green color gradient representing the abundance ratio of OTUs between samples with
203 STEC (blue) and without (green). Additionally, label luminosity was determined based on
204 overall abundance of each OTU (lighter: less abundant, darker: more abundant) (Fig. 3).
205 Iroki makes it clear that most OTUs on the tree showed a significant difference in
206 abundance (branch coloring) between STEC positive and STEC negative samples (node
207 coloring). Furthermore, we can see that most OTUs are at low abundance with only a few
208 highly abundant OTUs (label luminosity). The color gradient added by Iroki allows us to
209 see that the abundant OTUs were evolutionarily distant from one another and thus spread
210 out across many phylogenetic groups.

211

212 Iroki can be used to quickly test hypotheses without investing a large amount of time
213 annotating trees manually. A UPGMA tree was created based on unweighted UniFrac
214 distance [27] between 356 bacterial community profiles based on SSU rRNA amplicon
215 sequences from cattle hide and fecal samples (Fig. 4). Iroki was used to evaluate similarities

in sample bacterial communities according to the sampling location. Iroki colored branches based on whether the sample originated from feces (blue) or from hide (red). The coloring added by Iroki shows a clear partitioning of bacterial communities on the tree based on their sampling location (hide or feces). However, four fecal samples grouped with hide samples, and two hide samples grouped with fecal samples, highlighting the ability of Iroki to easily identify good candidates for more in-depth examination. Additionally, Iroki was used to illustrate a correlation between one of the most abundant bacterial families, Ruminococcaceae, and sampling location. Iroki colored node labels with a color gradient based on Ruminococcaceae family abundance, utilizing both a single color gradient (Fig. 4A) and a two color gradient (Fig. 4B). Custom trees were visualized using FigTree. Iroki's automatic color gradient and ability to label branches and nodes based on different criteria clearly show that Ruminococcaceae is more abundant in fecal samples than in hide samples.

Short-term dynamics of virioplankton

The gene encoding Ribonuclotide reductase (RNR) is common within viral genomes and thus can be used as a marker gene for studying viral diversity [23]. Moreover, RNR polymorphism is predictive of some of the biological and ecological features of viral populations [28]. A mesocosm experiment examined the short-term dynamics of phage populations using RNR amplicon sequences, specifically, sequences of class II RNRs of bacteriophages infecting cyanobacterial hosts. A phylogenetic tree was created from the Cyano II RNR amplicon sequences and Iroki was used to color nodes and branches based

238 on the time point (0 h, 6 h, 12 h) at which each amplicon sequence was observed. The
239 customized tree was then visualized using FigTree (Fig. 5). Iroki's coloring showed that no
240 phylogenetic clade was dominated by OTUs observed in any particular time point; rather,
241 time points were spread relatively evenly across clades. This analysis demonstrates Iroki's
242 utility for exploring sequence datasets, allowing the researcher to quickly and easily test
243 hypotheses.
244
245 *Phylogeny of Zetaproteobacteria within a biogeographic context*
246 Biogeographical studies assess the distribution of an organism's biodiversity across space
247 and time. The extent to which microorganisms exhibit geographic distribution patterns is an
248 open question in microbial ecology. The isolated nature of the microbial communities
249 associated with deep-ocean hydrothermal vents provides an ideal system for studying the
250 biogeography of microbes. In particular, iron-oxidizing bacteria have been shown to thrive
251 in vent fluids, sediments, and iron-rich microbial mats associated with the vents. Globally,
252 iron-oxidizing bacteria make significant contributions to the iron and carbon cycles. A
253 recent study analyzed multiple SSU rRNA clone libraries to investigate the biogeography of
254 Zetaproteobacteria, a phyla containing many iron-oxidizing bacterial species, between three
255 sampling regions of the Pacific Ocean (central Pacific—Loihi seamount, western Pacific—
256 Southern Mariana Trough, and southern Pacific (Vailulu'u Seamount/Tonga Arc/East Lau
257 Spreading Center/Kermadec Arc) [29]. Sequences were aligned and a phylogeny was
258 inferred as described in [29]. Iroki was used to examine the relationship between sampling
259 location and phylotype by adding branch and label color based on geographic location and

renaming original node labels with OTU and location metadata. The custom tree was visualized using FigTree (Fig. 6). In some cases, OTUs contained sequences from only one sampling location (e.g., OTUs 12, 15, and 16), whereas other OTUs are distributed among more than one sampling location (e.g., OTUs 1, 2, and 4). Often, sequences sampled from the same geographic location are in the same phylotype despite being members of different OTUs (e.g., OTUs 10 and 19).

Availability and requirements

A web browser version of Iroki can be accessed online at www.iroki.net or through the VIROME portal (http://virome.dbi.udel.edu/). For users who wish to run Iroki locally, a command line version of the program is installable via RubyGems, from GitHub (https://github.com/mooreryan/iroki). A Docker image is available for users who desire the flexibility of the command line tool, but do not want to install Iroki or manage its dependencies (https://hub.docker.com/r/mooreryan/iroki). Docker is a popular software container platform that allows bundling of an application with its dependencies in a portable, self-contained system [30,31]. The README file, accompanying the source code, provides detailed instructions for setting up and running Iroki. Further documentation and tutorials can be found at the Iroki Wiki (https://github.com/mooreryan/iroki/wiki).

License

Iroki and its associated programs are released under the GNU General Public License version 3 [32].

Conclusions

Iroki is a command line program and web browser application for fast, automatic customization of large phylogenetic trees based on user specified configuration files describing categorical or continuous metadata information. The output files include Nexus tree files with color metadata tailored specifically for use with FigTree. Various example datasets from microbial ecology studies were analyzed to demonstrate Iroki's utility. In each case, Iroki simplified the processes of data exploration, data presentation, and hypothesis testing. Though these examples focused specifically on applications in microbial ecology, Iroki is applicable to any problem space with hierarchical data that can be represented in the Newick tree format. Iroki provides a simple and convenient way to rapidly customize trees, especially in cases where the tree in question is too large to annotate manually or in studies with many trees to annotate.

List of Abbreviations

OTU: operational taxonomic

RNR: Ribonucleotide reductase

STEC: Shiga-toxengenic *Escherichia coli*

Ethics approval and consent to participate

Not applicable

Consent for publication

Not applicable

Availability of data and materials

Data and code used to generate figures are available on GitHub at https://github.com/mooreryan/iroki_manuscript_data

Funding

This project was supported by the USDA National Institute of Food and Agriculture award number 2012-68003-30155 and the National Science Foundation Advances in Bioinformatics program (award number DBI_1356374).

Competing Interests

The authors declare that they have no competing interests.

Authors' contributions

RMM and SMM conceived the project. RMM wrote the manuscript and implemented Iroki. AOH and RLM processed and analyzed Cyano II amplicons. All authors read, edited, and approved the final manuscript.

Acknowledgements

We would like to acknowledge Daniel J. Nasko and Jessica M. Chopyk for their work on the phage proteomic tree, and Barbra D. Ferrell for editing the manuscript.

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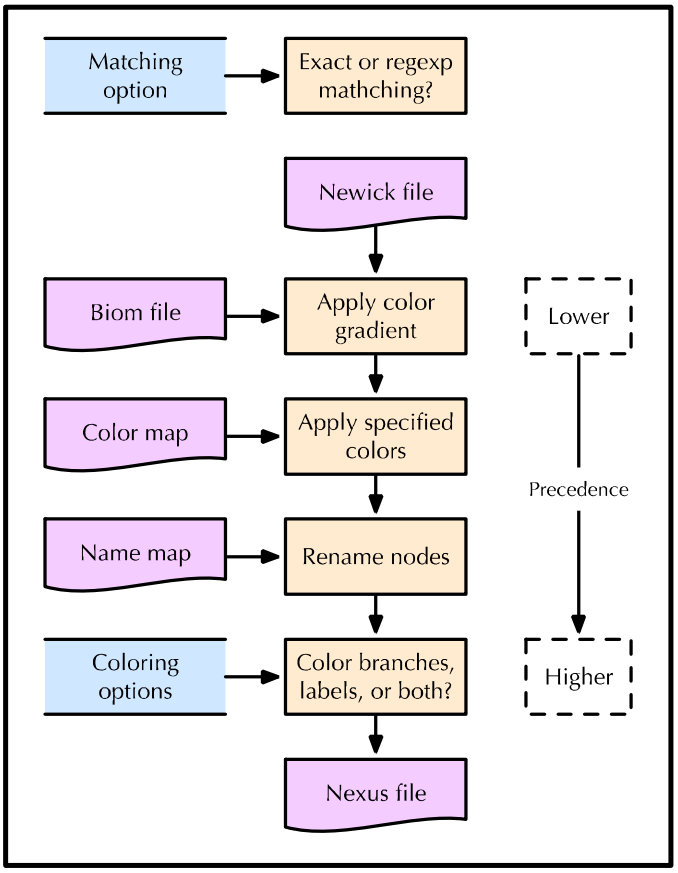


Fig. 1: Precedence of Iroki's customization pipeline

Flowchart illustrating the precedence of steps when performing multiple customizations with Iroki. Input/output files are purple, command line options are in blue, and processes are orange. The choice of exact or regular expression matching guides each subsequent step of the process. Iroki gives higher precedence to processes towards the bottom of the diagram. For example, given that a user selects the options for coloring both labels and branches, and provides both a biom file and color map with the color map specifying colors for the labels only, then the branches will be colored according to the color gradient inferred from the biom file, whereas the labels will be colored according to the rules specified in the color map.

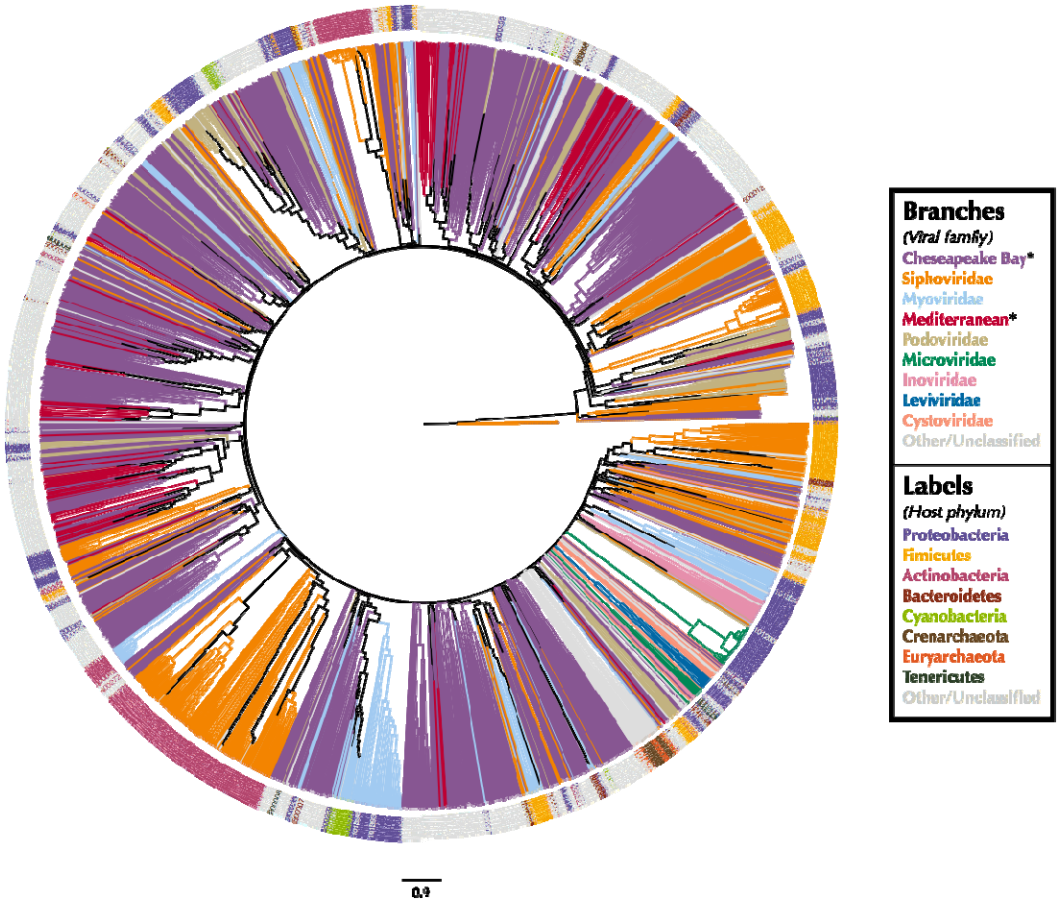


Fig. 2: Comparing phage and their host phyla

All phage genomes from Phage SEED with assembled virome contigs from the Chesapeake Bay and Mediterranean Sea. Iroki highlights phylogenetic trends after coloring branches according to viral family or sampling location in the case of virome contigs (marked with an asterisk in the legend), and coloring node labels according to host phylum of the phage.

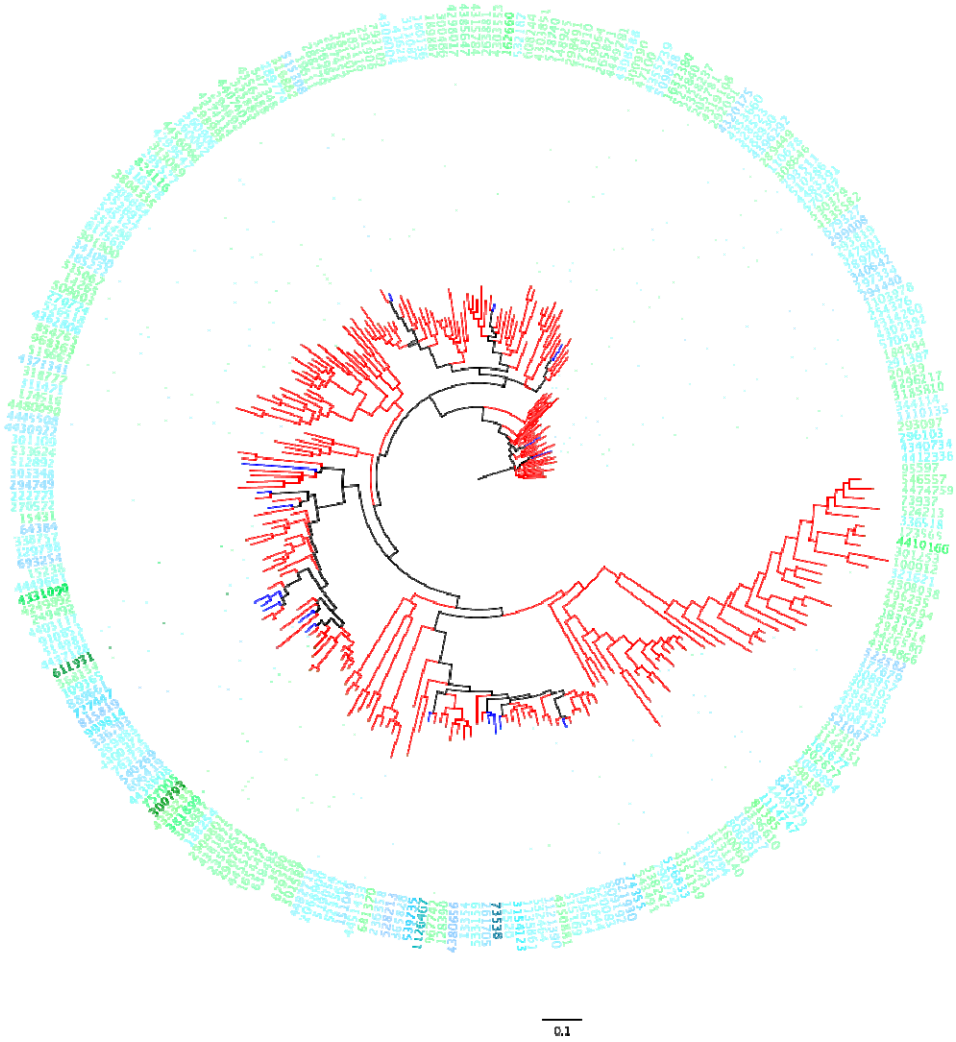
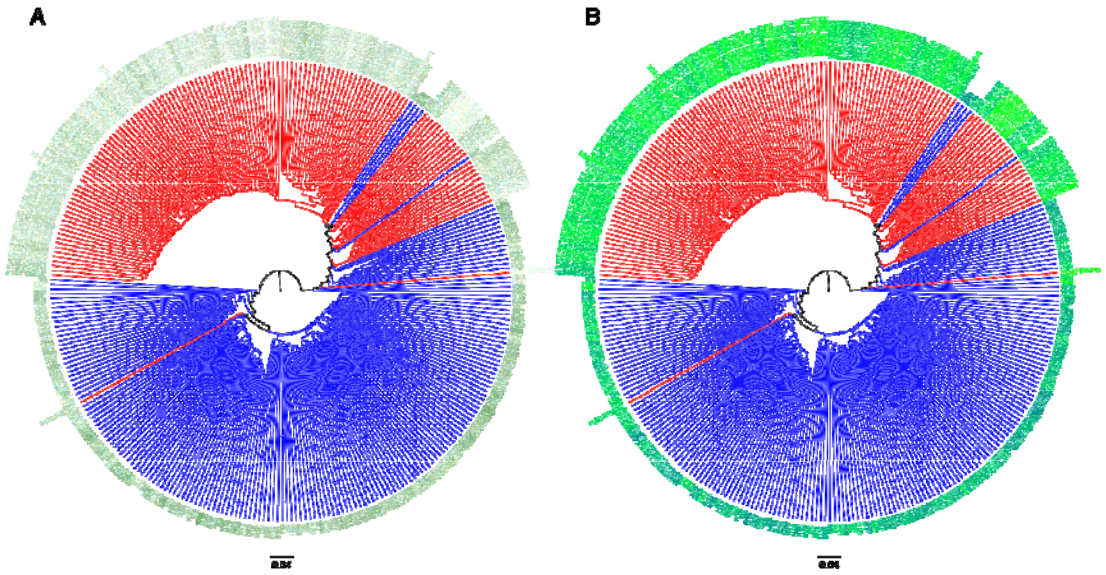


Fig. 3: Changes in OTU abundance in two sample groups

Approximate-maximum likelihood tree of OTUs that showed significant differences in relative abundance between STEC positive and STEC negative cattle hide samples. Branches show significance based on coloring by the p-value of a Mann-Whitney U test examining changes in abundance between samples positive for STEC ($p < 0.1$ – red) and samples negative for STEC, ($p >= 0.1$ – blue). Label color on a blue-green color gradient highlights OTU occurrence based on the abundance ratio between STEC positive samples (green) and STEC negative samples (blue). Labels that are darker green had a higher

abundance in STEC positive samples, and a lower abundance in STEC negative samples. For example, OTU 300793 (bottom left corner) is darker than most (indicating high overall abundance) and more green than blue (indicating higher abundance in STEC positive samples than in STEC negative samples). Node luminosity represents overall abundance with lighter nodes being less abundant than darker nodes.



436

437 **Fig. 4: Comparing cattle fecal and hide samples and the abundance of**

438 **Ruminococcaceae**

439 Phylogeny based on UPGMA tree of pairwise unweighted UniFrac distance between 356

440 bacterial community profiles based on SSU rRNA amplicon sequences from cattle hide and

441 feces. Branches are colored by feces (blue) and hide (red). Rapid testing of the hypothesis

442 that the abundance of one of the most abundant families, Ruminococcaceae, and sample

443 origin are correlated is enabled through node label coloring by (A) a green single-color

444 gradient (color saturation increases with increasing abundance of Ruminococcaceae OTUs)

445 and (B) a light green (low abundance of Ruminococcaceae OTUs) to dark blue (high

446 abundance of Ruminococcaceae OTUs) color gradient.

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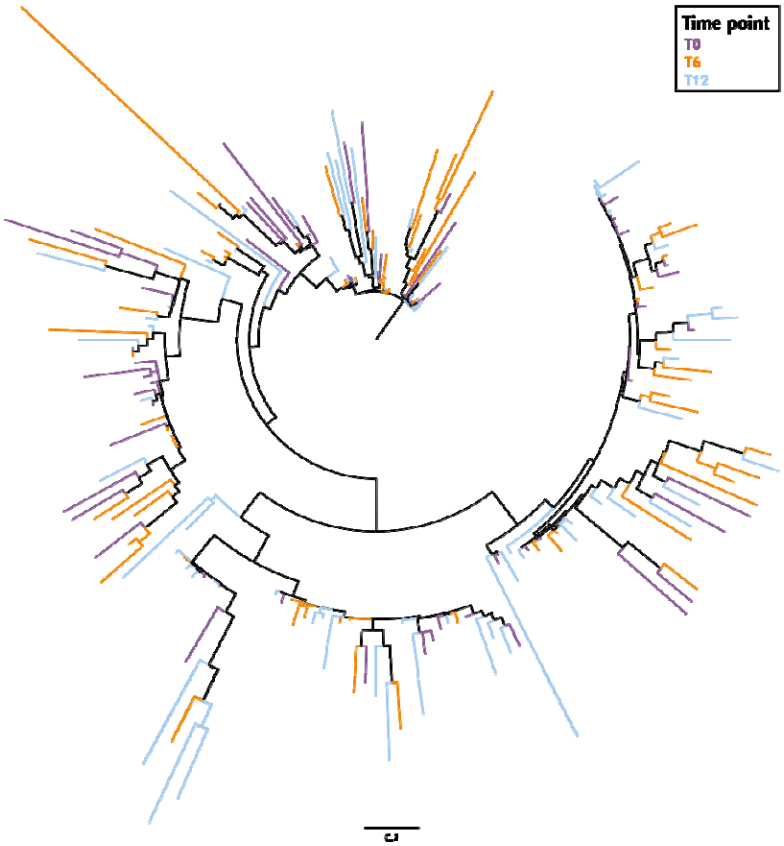

448

449 **Fig. 5: Temporal dynamics of virioplankton populations according to Cyano II**

450 **RNR amplicon phylogeny**

451 An approximately-maximum-likelihood phylogenetic tree of 200 randomly selected class II

452 Cyano RNR representative sequences from 98% percent clusters. Iroki was used to color

453 branches by time point: zero hours – purple, six hours – orange, and twelve hours – blue.

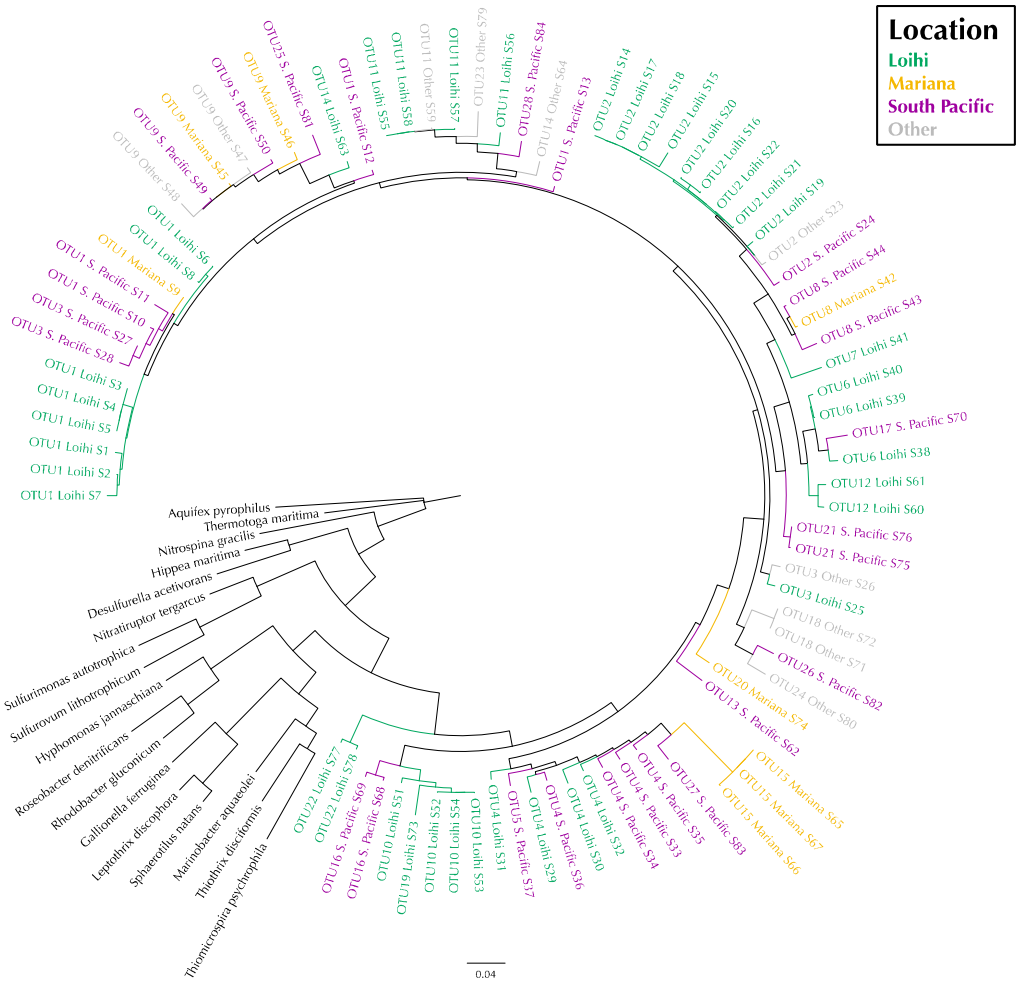

Fig. 6: Zetaproteobacteria show biogeographic partitioning

Phylogenetic tree showing placement of 84 full-length Zetaproteobacteria SSU rRNA sequences collected from three Pacific Ocean locations and 17 reference sequences. Iroki was used to color labels and branches by geographic location of the sampling site (Loihi – green, Mariana – gold, South Pacific – purple, and Other – gray), as well as to rename the nodes with OTU and sampling site metadata. Known reference Zetaproteobacterial species are shown in black.