The *Drosophila* pheromone Z4-11Al is encoded together with habitat olfactory cues and mediates species-specific communication

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Abstract

Mate recognition in animals evolves during niche adaptation and involves habitat and social olfactory signals. *Drosophila melanogaster* is attracted to fermenting fruit for feeding and egg-laying. We show that, in addition, female flies release a pheromone (*Z*)-4-undecenal (*Z*4-11Al), that elicits flight attraction in both sexes. The biosynthetic precursor of *Z*4-11Al is the cuticular hydrocarbon (*Z,Z*)-7,11-heptacosadiene (7,11-HD), which is known to afford reproductive isolation between *D. melanogaster* and *D. simulans*. A pair of alternatively spliced receptors, Or69aB and Or69aA, is tuned to *Z*4-11Al and to food olfactory cues, respectively. These receptors are co-expressed in the same olfactory sensory neurons, and feed into a neural circuit mediating species-specific, long-range communication: the close relative *D. simulans* does not respond. That Or69aA and Or69aB have adopted dual olfactory traits highlights the interplay of habitat and social signals in mate finding. These olfactory receptor genes afford a collaboration between natural and sexual selection, which has the potential to drive phylogenetic divergence.

Keywords

pheromone, kairomone, olfaction, reproductive isolation

Introduction

Volatile insect pheromones transmit species-specific messages over a distance. Premating communication with pheromones facilitates and accelerates mate-finding, and reduces predation risk and energy expenditure, which is particularly adapative in short-lived insects (1,2).

Sexual communication subserves mate-finding and reproduction. The sensory drive hypothesis predicts that mate recognition in animals evolves during niche adaptation and that premating sexual communication involves olfactory specialization to both social signals and habitat cues (3). Pheromones are released into an atmosphere that is filled with environmental, habitat-related olfactory cues, some of which signal mating sites and food sources. The response to pheromones and habitat odourants (kairomones) is under sexual and natural selection, respectively. Pheromones and kairomones are always perceived as an ensemble in a natural context and this leads to interaction of sexual and

natural selection during adaptive divergence of sexual signaling, which is thought to facilitate premating reproductive isolation (3-6).

Olfactory sexual communication is studied at cellular and molecular resolution in the fruit fly *Drosophila melanogaster*, but volatile pheromones encoding species-specific, long-range mate recognition have not yet been found. *Drosophila* is attracted to yeast and fruit odorants for feeding, mating and oviposition (7-9) and the interconnection between perception of pheromones and food semiochemicals is a current research theme (10, 11). For example, the male-produced sex pheromone cVA and food stimuli are integrated to coordinate feeding, courtship behavior and oviposition site selection (12-15). Perception of cVA is a current and outstanding paradigm for studying the molecular and neuronal logic of innate, olfactory-mediated reproductive behavior (14, 16, 17). cVA and other olfactory pheromones are active during courtship. Since they are shared with other *Drosophila* species, they cannot account for species-specific communication (18-20).

Interspecific matings of *D. melanogaster* with sibling species are inhibited by the female-produced cuticular hydrocarbon (*Z,Z*)-7,11-heptacosadiene (7,11-HD), which is not volatile and perceived through gustatory receptors only at close range (21-23). This raises the question whether *Drosophila* uses, in addition, volatile pheromone signals that mediate specific mate recognition at a distance.

We have identified the first long-range, species-specific pheromone in *D. melanogaster*. A pair of spliced olfactory receptors, feeding into the same neural circuit, has developed a dual affinity to this pheromone and to environmental semiochemicals, encoding adult and larval food. A blend of this pheromone and a food odourant specifically attracts *D. melanogaster*, but not the close relative *D. simulans*. This becomes an excellent paradigm to study the interaction of social signals and habitat olfactory cues in premating reproductive isolation and phylogenetic divergence.

Results

Drosophila melanogaster females produce a suite of volatile aldehydes.

We analysed volatile compounds released by *D. melanogaster* flies, by gas chromatography-mass spectrometry (GC-MS), and found 16 aliphatic aldehydes. Males and females shared saturated aldehydes with a carbon chain length of C7 to C18, but mono-unsaturated aldehydes were released by females only (Fig. 1a; Table 1). The most abundant compound was identified as (*Z*)-4-undecenal (*Z*4-11Al) and synthesized.

GC-MS analysis showed that *Z*4-11Al was present also in cuticular extracts of females, although in lower amounts (0.27 ± 0.12 ng/female, n = 5) than in headspace collections

(3.0 ± 0.81 ng/female, n = 10; P<0.01 Mann-Whitney test). Cuticular profiles of *Drosophila* flies have been investigated, but *Z*4-11Al or other aldehydes have not been reported (18, 24, 25).

The closely related *D. simulans* did not release *Z*4-11Al, or other monounsaturated aldehydes (Fig. 1a). Unlike *D. melanogaster*, *D. simulans* does not produce (*Z,Z*)-7,11-heptacosadiene (7,11-HD) (21). This led us to hypothesize that the production of mono-unsaturated aldehydes with a double bond in position 4 was linked to oxidation of di-unsaturated cuticular hydrocarbons. Oxidation of 7,11-HD is expected to generate two saturated aldehydes, heptanal and hexadecanal, and two unsaturated aldehydes, *Z*4-11Al and (*Z*)-4-eicosenal (Fig. 1b). This was experimentally verified by applying 100 ng synthetic 7,11-HD to a glass vial. After 60 min, 1.92 ± 0.42 ng *Z*4-11Al were retrieved (n = 3). Based on the cuticular hydrocarbon profile of *D. melanogaster* (24), 26 aldehydes are expected to be formed by oxidation, 16 of which were found in our headspace analysis (Table 1), others may have been below detection level.

Next, single sensillum electrophysiological recordings (SSR) from all basiconic, trichoid, coeloconic, and intermediate olfactory sensilla in *D. melanogaster* (Fig. 1c) and GC-coupled SSR recordings (GC-SSR) from ab9 sensilla (Fig. 1d) showed that *Z*4-11Al strongly activates ab9A olfactory sensory neurons (OSNs).

The olfactory receptor Or69aB responds to *Z*4-11Al.

ab9A OSNs express the olfactory receptor (Or) Or69a (26). We therefore screened ab9A OSNs with known ligands of Or69a (27) and *Z*4-11Al. In the *D. melanogaster* strains Canton-S and Zimbabwe, the monoterpene (*R*)-carvone elicited the strongest response from ab9A, although the response to *Z*4-11Al was not significantly different. In *D. simulans*, Z4-11Al elicited a significantly lower response than (*R*)-carvone (Fig. 3a).

The Or69a gene encodes two proteins, Or69aA and Or69aB, as a result of alternative splicing (Fig. 3d), prior to the split of the *D. obscura* and *D. melanogaster* groups (28, 29). Heterologous co-expression of both Or69a splice variants in ab3A (*Δhalo*) empty neurons (30) produced a response similar to native ab9A OSNs; whereas individual expression revealed distinct response profiles for Or69aA and Or69aB (Fig. 3a,b). Or69aB responds best to both isomers of carvone, followed by *Z*4-11Al. These compounds share a structural motif, a carbonyl functional group with an equidistant double bond in position 4 (Fig. 3b,c). Upon binding to the same Or, ligands adopt complementary bioactive conformations; the strain energy required to achieve the bioactive conformation should typically not exceed 5 kcal/mol (31). Conformational analysis showed that *Z*4-11Al aligns with (*R*)-carvone at a strain energy cost of only 1.5 kcal/mol. Or69aA, on the other hand, is tuned to terpenoid alcohols and responded significantly less to *Z*4-11Al. The most active ligands (*S*)-terpineol, (*S*)- and (*R*)-linalool, which again share the functional group and a double bond in position 4, align at 3.0 kcal/mol (Fig. 3b,c).

The Or69a splice event enabled adaptive changes in ligand tuning in the newly emerged twin receptor. Functional divergence has apparently been biased towards structurally related ligands (Fig. 3b,c; 27) and ecologically relevant odorant signals. Citrus peel, a preferred oviposition substrate (8), is a common source of all main ligands of Or69aA and Or69aB (32) and possibly a driver of Or69a ligand evolution. Diversification of the splice variants is, however, constrained to a behavioral theme, since both Ors feed into one OSN.

Z4-11Al elicits upwind flight attraction in *D. melanogaster*, but not in *D. simulans*.

*Z*4-11Al elicited upwind flight and landing at the source, in cosmopolitan Dalby and Canton-S strain *D. melanogaster* males and females. In contrast, males of the Zimbabwe strain and the sister species *D. simulans* were not attracted (Fig. 4a,b). This shows that *Z*4-11Al, in addition to its precursor 7,11-HD (Fig. 1) participates in sexual isolation between *D. melanogaster* and *D. simulans* (21), and between cosmopolitan and African *D. melanogaster* strains (33-35). Moreover, admixture of *Z*4-11Al eliminated *D. simulans* attraction to the yeast volatile (*R*)-linalool (31) (Fig. 4a). Finally, we used tetanus toxin mutants to verify that Or69a encodes *Z*4-11Al. Upwind flight attraction was significantly reduced when Or69a OSNs were disrupted (Fig. 4b).

The response rate to Z4-11Al released at a rate of 10 ng/min was similar to the upwind flight response to vinegar, when acetic acid, the main compound of vinegar headspace, was released at ca. 17 µg/min (Fig. 4a; 36). In summary, *Z*4-11Al is a powerful attractant that enables specific mate recognition at a distance and its interspecific role was enhanced through interaction with the food odour linalool.

Discussion

*Z*4-11Al is the first species-specific, long-range sex pheromone of *D. melanogaster*. It is produced by females and perceived by Or69aB in both sexes. We further show that Or69aB and its twin receptor Or69aA bind kairomonal terpenoids, such as linalool, found in fruit and yeast headspace (32, 37). Combined social and food odour tuning in the two Or69a splice variants underscores the tie between sexual and natural selection during the evolution of specific mate communication (3, 4). Tinting food odorants with sex pheromone foregrounds communication channels and facilitates mate finding. This is particularly adaptive in *Drosophila* when mating sites, fruit and berries, are abundant and widely spread.

Olfactory representations of other *Drosophila* Ors involved in food and pheromone perception project through separate channels to the LH, where they partially overlap and integrate (14). In stark contrast, Or69a is the first olfactory gene known to encode dual olfactory traits. Or69aA and Or69aB co-express in the same ab9A OSNs (26) and thus achieve a coordination of mating and food stimuli already in first order neurons, at the antennal periphery. This makes Or69a a target for selection during phylogenetic divergence. The tuning range of Ors evolves more rapidly than hardwired neural circuits in higher brain centres (38) and selection pressure is further relaxed following a splice event. Differential tuning of Or69a in closely related cosmopolitan and African strains of *D. melanogaster* corroborates this idea.

Tuning changes in the two splice forms of Or69a are constrained with respect to the behavioral and ecological role of their ligands, since they both feed into a neural circuit mediating sexual and habitat attraction, and courtship. The two splice forms provide, on the other hand, degrees of freedom during adaptive divergence, since they allow fly populations to adopt new kairomone and pheromone signals; alteration of either one produces a new communication channel. Reproductive isolation may arise as a byproduct and the Or69a gene therefore has the potential to drive speciation (39, 40).

Methods

Insects

Canton-S, Zimbabwe (S-29; Bloomington #60741) and Dalby-HL (Dalby, Sweden) (41) strains of *D. melanogaster* were used as wild type flies for behavioral experiments. Canton-S was used for comparison with knockouts of the same background. Further tests were done with the sister species *D. simulans*.

We used the Or69a-Gal4/UAS TeTx, tetanus toxin knockout line to verify the role of Or69a in flight attraction to *Z*4-11Al. Canton-S/UAS TeTx (Bloomington #28838 and 28997) and Canton-S/Or69a-Gal4 (Bloomington #10000) were used as parental controls.

Flies were reared on a standard sugar-yeast-cornmeal diet at room temperature (19 to 22°C) under a 16:8-h L:D photoperiod. Newly emerged flies were anesthetized under CO_2 and sexed under a dissecting microscope. Virgin flies were identified by the presence of meconium, and were kept together with flies of the same sex. Flies were kept in 30-ml Plexiglas vials with fresh food. Experiments were done with 3- to 5-d-old flies.

Chemicals

(*Z*)-4-undecenal (*Z*4-11Al) and (*E*)-4-undecenal (*E*4-11Al) were synthesized. A short description follows below, for a complete account of the chemical synthesis, see Supplemental Information.

(*Z*)-4-Undecenoic acid was synthesized via a modified version of Wube et al. (42) in 80% stereoisomeric purity. Esterification under acidic conditions with sulfuric acid in methanol resulted in 80% *Z*-isomer and a 93% yield over two steps. Stereoisomeric purity was controlled with NMR and GC-FID by comparing analysis for acid and ester, the appearance of a small quartet, in the NMR spectra, at 1.96 indicates the presence of *E*-isomer. Gas chromatographic separation on a polar Varian factorFOUR vf-23ms of *Z*- and *E*-ester proved that the stereochemistry was not affected by the acidic conditions during esterification. Methyl (*Z*)-4-undecenoate was purified on regular silica gel and on silver nitrate impregnated silica gel to obtain a stereoisomeric purity of 98.6%. Methyl (*Z*)-4-undecenoate was reduced to (*Z*)-4-undecenol with lithium aluminum hydride in diethylether and oxidized to *Z*4-11Al with Dess-Martin periodinane in dichloromethane.

A modified version of Virolleaud's (43) metathesis was used to produce (*E*)-4-undecenoic acid in a 56% yield (87.5% of the *E*-isomer). (*E*)-4-undecenoic acid was esterified under the same conditions as the (*Z*)-acid, without isomerisation of the double bond (according to GC-FID and ^{1}H-NMR). The methyl-(*E*)-4-undecenoate was reduced to the alcohol with lithium aluminum hydride in diethylether and purified on silver nitrate impregnated silica gel to obtain a purity of 99.8% of the (*E*)-isomer, which was oxidized with Dess-Martin periodinane in dichloromethane to obtain *E*4-11Al.

Commercially available compounds were: (*R*)-carvone (97% chemical purity, CAS #6485-40-1, Firmenich), (*S*)-carvone (98%, CAS #2244-16-8, Firmenich), (*S*)-terpineol (97%, CAS #10482-56-1, Aldrich), (*S*)-linalool (97%, CAS #126–91–0, Firmenich), (*R*)-linalool (97%, CAS #126-90-9, Firmenich), citronellol (99%, CAS #106-22-9, Aldrich), geraniol (98%, CAS #106-24-1, Aldrich), 3-octanol (99%, CAS #589-98-0, Aldrich), decanol (99%, CAS #112-30-1, Fluka), 11-Al (99%, CAS #112-44-7, Aldrich).

Odor collection and chemical analysis

Twenty *D. melanogaster* (Dalby), *D. melanogaster* (Canton) (*n* = 10) or 20 *D. simulans* (*n* = 10) virgin female or virgin male flies were placed in a glass aeration apparatus designed for collection of airborne pheromone (effluvia) (44). The flies were held in a glass bulb with a narrow open outlet (ø 1 mm), which prevented them from escaping. A charcoal-filtered air flow (100 mL/min) passed over the flies during 75 min. Fly effluvia were collected on the glass surface, breakthrough was monitored by attaching a 10-cm glass capillary (ø 1 mm). After 75 min, flies were removed, 100 ng of heptadecenyl

acetate (internal standard) was deposited in the glass bulb, which was then rinsed with 50 µl hexane, the solvent was concentrated to 10 µl in Francke vials.

Cuticular extracts (n = 5) were obtained by dropping 20 *D. melanogaster* females for 5 min in 400 µl hexane containing 100 ng heptadecenyl acetate. After 5 min, the extracts were transferred to Francke vials and concentrated to 10 µl before analysis. Fly extracts and volatile collections were stored at -20°C.

Oxidation of (*Z,Z*)-7,11-heptacosadiene (7,11-HD) was analysed by depositing 100 ng synthetic 7,11-HD in a 1.5-mL glass vial, at 19°C. Vials were rinsed with 10 µl of hexane, which contained 100 ng heptadecenyl acetate as an internal standard, after 15, 30, 45, 60 and 75 min (n = 3).

Samples were analysed by combined gas chromatography and mass spectrometry (GC-MS; 6890 GC and 5975 MS, Agilent technologies Inc., Santa Clara, CA, USA). Two µl were injected (injector temperature 225°C) splitless (30 s) into fused silica capillary columns (60 m x 0.25 mm), coated with HP-5MS UI (Agilent Technologies Inc., d_f = 0.25 µm) or DB-wax (J&W Scientific, Folsom, CA, USA, d_f = 0.25 µm), that were temperature-programmed from 30 to 225°C at 8°C/min. Helium was used as mobile phase at 35 cm/s. The MS operated in scanning mode over m/z 29-400. Compounds were tentatively identified based on their mass spectra and Kovats retention indices, using custom and NIST (Agilent) libraries, as well as authentic standards.

Behavioural assays

Upwind flight behavior was observed in a glass wind tunnel (30 x 30 x 100 cm) equipped with a piezo sprayer (36). The flight tunnel was lit diffusely from above, at 13 lux, temperature ranged from 22 to 24°C, relative humidity from 38% to 48% and charcoal filtered air, at a velocity of 0.25 m/s, was produced by a fan (Fischbach GmbH, Neunkirchen, Germany). Compounds were delivered from the centre of the upwind end of the wind tunnel via a piezo-electric micro-sprayer (45). Forty flies were flown individually to each treatment. "Attraction" was defined as upwind flight, directly from a release tube at the end of the tunnel over 80 cm towards the odor source, followed by landing. Unmated fed, 3-d-old Dalby wild-type males and females, *D. melanogaster* Zimbabwe strain males and *D. simulans* males were flown towards (*Z*)-4-undecenal (released at 10 ng/min), (*R*)-linalool (10 ng/min) and the blend of (*Z*)-4-undecenal and (*R*)-linalool (10 ng/min, each).

Heterologous expression of Or69aA and Or69aB

Or69aA and Or69aB receptors were cloned from antennae of *D. melanogaster*, Dalby line (46). Briefly, cDNA was generated from RNA extracts of antennae of 100 males and females using standard procedures. Or69a variants were PCR amplified with the following

primers: Or69aA_5': GTCATAGTTGAAACCAGGATGCAGTTGC, Or69aB_5': ATAATTCAGGACTAGATGCAGTTGGAGG, Or69aAB_3': TGCACTTTTGCCCTTTTATTTAAGGGAC.

The splice variants were amplified with unique 5' primers and a common 3' primer, reflective of genomic structure at this locus. These primers encompass the entire open reading frame of the receptor variants, and are located partially upstream and downstream of the start and stop codons. PCR amplicons were gel-purified and cloned into the pCR8/GW/Topo-TA Gateway entry vector (Thermo-Fisher Scientific, Waltham, MA, USA) according to standard procedure, with vector inserts sequenced to confirm fidelity of Or sequence. Or inserts were subsequently transferred to pUAS.g-HA.attB (47) with LR Clonase II enzyme (Thermo-Fisher Scientific), according to manufacturers protocol; vector inserts were sequenced to confirm fidelity of Or sequence.

Mini-prep purified pUAS.g-HA.attB plasmid with Or69aA or Or69aB insert were delivered to Best Gene Inc. (Chino Hills, CA USA) for generation of transgenic *D. melanogaster* flies. Using the PhiC31 targeted genomic-integration system (47) vectors with Or69aA or Or69aB were injected into the following fly strain, for integration on the 3rd chromosome: M{3xP3-RFP.attP}ZH-86Fb (with M{vas-int.Dm}ZH-2A) (Bloomington *Drosophila* Stock Number: 24749). For expression of single receptor variants in the empty neuron system, Or69a transgenes were crossed into the *Δhalo* background to give genotype: w; *Δhalo*/Cyo; UAS-DmelOr69a(A or B), and these flies were crossed to flies with genotype: w; *Δhalo*/Cyo; DmelOr22a-Gal4, as described previously (46). Experimental electrophysiology assays were performed on flies with genotype: w; *Δhalo*; UAS-DmelOr69a(A or B)/DmelOr22a-Gal4.

For co-expression of Or69aA and Or69aB in the same empty neurons, a second fly-line with Or69aB was generated with Or69aB present on the X-chromosome. The same UASg-HA.attB:Or69aB plasmid generated previously was injected into the following fly strain: y,w, P{CaryIP}su(Hw)attP8 (Bloomington *Drosophila* Stock Number: 32233). The Or69aB transgene was crossed into the DmelOr22a-Gal4 line in *Δhalo* background to give genotype: UAS-DmelOr69aB; *Δhalo*/Cyo; DmelOr22a-Gal4; these flies were crossed to flies with genotype: w; *Δhalo*/Cyo; UAS-DmelOr69aA. Experimental electrophysiology assays were performed on flies with genotype: UAS-DmelOr69aB/w; *Δhalo*; UAS-DmelOr69aA/DmelOr22a-Gal4.

Conformational analysis

MacroModel version 11.0 (Schrodinger LLC, New York, NY, USA) in the Maestro Version 10.4.017 were used to build, minimize and to perform conformational analysis of *Z*4-11Al, (*R*)-carvone, (*S*)-terpineol and (*R*)-linalool, using default settings (OPLS3 as force field, water as the solvent and mixed torsional/low-mode sampling method). The

assumed bioactive conformations of the conformationally more flexible compounds, *Z*4-11Al and (*R*)-linalool, were based on the positon of the shared functional groups in the conformationally more restricted compounds, (*R*)-carvone and (*S*)-terpineol. The carbonyl and the double bond atoms were kept fixed during minimization of the proposed bioactive conformation of Z4-11Al; the alcohol functional group and the double bond were kept fixed in (*R*)-linalool. The strain energies, the energy cost for adopting proposed bioactive conformations, were then calculated as the difference between the lowest energy conformations and the assumed bioactive conformation.

Electrophysiological recordings

Single sensillum recordings (SSR) were done as described earlier (18). Unmated males were restrained in 100-µl pipette tips, with half of the head protruding, the third antennal segment or palps were placed on a glass microscope slide and held by dental wax. For the initial screening, all basiconic, trichoid, coeloconic, and intermediate sensilla (26) were localized in *D. melanogaster* (Canton-S strain) males, under a binocular at 1000x magnification. Further recordings were made from small basiconic ab9 sensilla, in *D. melanogaster* (Canton-S and Zimbabwe strains) and in *D. simulans* males, and from large basiconic ab3 sensilla in mutant *D. melanogaster*, where Or69aA and Or69aB were heterologously expressed (see above).

Tungsten electrodes (diameter 0.12 mm, Harvard Apparatus Ltd, Edenbridge, United Kingdom) were electrolytically sharpened with a saturated KNO_3 solution. The recording electrode was introduced with a DC-3K micromanipulator equipped with a PM-10 piezo translator (Märzhäuser Wetzler GmbH, Germany) at the base of the sensilla. The reference electrode was inserted into the eye. The signal from olfactory sensory neurons (OSNs) was amplified with a probe (INR-02; Syntech), digitally converted by an IDAC-4-USB (Syntech) interface, and analyzed with Autospike software v. 3.4 (Syntech). Neuron activities were recorded during 10 s, starting 2 s before odor stimulation. Neuron responses were calculated from changes in spike frequency, during 500 ms before and after odor stimulation.

Odorants were diluted in redistilled hexane, 10 µg of test compounds in 10 µl hexane were applied to filter paper (1 cm^2), kept in Pasteur pipettes. The test panel contained the most active ligands known for Or69a (27) and several aldehydes. Diagnostic compounds for confirmation of sensillum identity were 2-phenyl ethanol (ab9) and 2-heptanone (ab3). Control pipettes contained solvent only. Puffs (2.5 ml, duration 0.5 s) from these pipettes, produced by a stimulus controller (Syntech GmbH, Kirchzarten, Germany), were injected into a charcoal-filtered and humidified airstream (0.65 m/s), which was delivered through a glass tube to the antenna.

For GC–SSR recordings, GC columns and the temperature programmes were the same as for the GC–MS analysis. At the GC effluent, 4 psi of nitrogen was added and split 1:1 in a

3D/2 low dead volume fourway-cross (Gerstel, Mühlheim, Germany) between the flame ionization detector and the antenna. Towards the antenna, the GC effluent capillary passed through a Gerstel ODP-2 transfer line, that tracked GC oven temperature, into a glass tube (30 cm x 8 mm ID), where it was mixed with charcoal-filtered, humidified air (20°C, 50 cm/s).

Statistical analysis

Generalized linear models (GLM) with a Bernoulli binomial distribution were used to analyse wind tunnel data. Landing at source and sex were used as the target effects. *Post-hoc* Wald pairwise comparison tests were used to identify differences between treatments. For all the electrophysiological tests, differences in spike activity derived from SSRs were analyzed with Kruskall Wallis H test followed by pairwise comparisons with Mann Whitney U *post hoc* test. All statistical analysis were carried out using R (R Core Team 2013) and SPSS Version 22 (IBM Corp.).

References

1. Greenfield M (1981) Moth sex pheromones: an evolutionary perspective. *Florida Entomol* 64, 4-17.
2. Wyatt TD (2010) Pheromones and signature mixtures: defining species-wide signals and variable cues for identity in both invertebrates and vertebrates. *J Comp Physiol A* 196, 685-700.
3. Endler JA (1992) Signals, signal conditions, and the direction of evolution. *Am Naturalist* 139:S125-S153.
4. Boughman JW (2002) How sensory drive can promote speciation. *Tr Ecol Evol* 17, 571-577.
5. Servedio MR, Van Doorn GS, Kopp M, Frame AM, Nosil P (2011) Magic traits in speciation: 'magic' but not rare? *Tr Ecol Evol* 26, 389-397.
6. Boughman JW, Svanbäck R (2016) Synergistic selection between ecological niche and mate preference primes diversification. *Evolution* (in press). doi:10.1111/evo.13089
7. Becher PG, Flick G, Rozpedowska E, Schmidt A, Hagman A, Lebreton S, Larsson MC, Hansson BS, Piskur J, Witzgall P, Bengtsson M (2012) Yeast, not fruit volatiles mediate attraction and development of the fruit fly *Drosophila melanogaster*. *Funct Ecol* 26, 822-828.
8. Dweck HK, Ebrahim SA, Kromann S, Bown D, Hillbur Y, Sachse S, Hansson BS, Stensmyr MC (2013) Olfactory preference for egg laying on citrus substrates in *Drosophila*. *Curr Biol* 23, 2472-2480.

9. Laturney M, Billeter JC (2014) Neurogenetics of female reproductive behaviors in *Drosophila melanogaster*. *Adv Genet* 85, 1-108.
10. Depetris-Chauvin A, Galagovsky D, Grosjean Y (2015) Chemicals and chemoreceptors: ecologically relevant signals driving behavior in *Drosophila*. Front Ecol Evol 3, 41. doi: 10.3389/fevo.2015.00041
11. Gorter JA, Jagadeesh S, Gahr C, Boonekamp JJ, Levine JD, Billeter JC (2016) The nutritional and hedonic value of food modulate sexual receptivity in *Drosophila melanogaster* females. *Sci Rep* 6, 19441.
12. Bartelt RJ, Schaner AM, Jackson LL (1985) cis-Vaccenyl acetate as an aggregation pheromone in *Drosophila melanogaster*. *J Chem Ecol* 11, 1747-1756.
13. Grosjean Y, Rytz R, Farine JP, Abuin L, Cortot J, Jefferis GSXE, Benton R (2011) An olfactory receptor for food-derived odours promotes male courtship in *Drosophila*. *Nature* 478, 236-U123.
14. Kohl J, Ostrovsky AD, Frechter S, Jefferis GSXE (2013) A bidirectional circuit switch reroutes pheromone signals in male and female brains. *Cell* 155, 1610-1623.
15. Lebreton S, Trona S, Borrero-Echeverry F, Bilz F, Grabe V, Becher PG, Carlsson MA, Nässel DR, Hansson BS, Sachse S, Witzgall P (2015) Feeding regulates sex pheromone attraction and courtship in *Drosophila* females. *Sci Rep* 5, 13132.
16. Pavlou HJ, Goodwin SF (2013) Courtship behavior in *Drosophila melanogaster*: towards a 'courtship connectome'. *Curr Op Neurobiol* 23, 76-83.
17. Clowney EJ, Iguchi S, Bussell JJ, Scheer E, Ruta V (2015) Multimodal chemosensory circuits controlling male courtship in *Drosophila*. *Neuron* 87, 1036-1049.
18. Dweck HK, Ebrahim SA, Thoma M, Mohamed AA, Keesey IW, Trona F, Lavista-Llanos S, Svatos A, Sachse S, Knaden M, Hansson BS (2015) Pheromones mediating copulation and attraction in *Drosophila*. *Proc Natl Acad Sc* 112, E2829-E2835.
19. Auer TO, Benton R (2016) Sexual circuitry in *Drosophila*. *Curr Opin Neurobiol* 38, 18-26.
20. El-Sayed AM (2016) The Pherobase: database of insect pheromones and semiochemicals. www.pherobase.com
21. Billeter JC, Atallah J, Krupp JJ, Millar JG, Levine JD (2009) Specialized cells tag sexual and species identity in *Drosophila melanogaster*. *Nature* 461, 987-U250.
22. Thistle R, Cameron P, Ghorayshi A, Dennison L, Scott K (2012) Contact chemoreceptors mediate male-male repulsion and male-female attraction during *Drosophila* courtship. *Cell* 149, 1140-1151.
23. Toda H, Zhao X, Dickson BJ (2012) The *Drosophila* female aphrodisiac pheromone activates ppk23+ sensory neurons to elicit male courtship behavior. *Cell Rep* 1, 599-607.
24. Everaerts C, Farine JP, Cobb M, Ferveur JF (2010) *Drosophila* cuticular hydrocarbons revisited: mating status alters cuticular profiles. *PLoS One* 5, e9607.
25. Dembeck LM, Böröczky K, Huang W, Schal C, Anholt RR, Mackay TF (2015) Genetic architecture of natural variation in cuticular hydrocarbon composition in *Drosophila melanogaster*. *eLife* 4, e09861.

26. Couto A, Alenius M, Dickson BJ (2005) Molecular, anatomical, and functional organization of the *Drosophila* olfactory system. *Curr Biol* 15, 1535-1547.

27. Münch D, Galizia CG (2016) DoOR 2.0 - comprehensive mapping of *Drosophila melanogaster* odorant responses. *Sci Rep* 6, 21841.

28. Robertson HM, Warr CG, Carlson JR (2003) Molecular evolution of the insect chemoreceptor gene superfamily in *Drosophila melanogaster*. *Proc Natl Acad Sci USA* 100, 14537-14542.

29. Conceicao IC, Aguade M (2008) High incidence of interchromosomal transpositions in the evolutionary history of a subset of Or genes in *Drosophila*. *J Molec Evol* 66, 325-332.

30. Dobritsa AA, van der Goes van Naters W, Warr CG, Steinbrecht RA, Carlson JR (2003) Integrating the molecular and cellular basis of odor coding in the *Drosophila* antenna. *Neuron* 37, 827-841.

31. Perola E, Charifson PS (2004) Conformational analysis of drug-like molecules bound to proteins: an extensive study of ligand reorganization upon binding. *J Medicin Chem* 47, 2499-2510.

32. Chisholm MG, Jell JA, Cass DM (2003) Characterization of the major odorants found in the peel oil of *Citrus reticulata* Blanco cv. Clementine using gas chromatography–olfactometry. *Flavour Fragrance J* 18, 275-281.

33. Ferveur JF, Cobb M, Boukella H, Jallon JM (1996) World-wide variation in *Drosophila melanogaster* sex pheromone: behavioural effects, genetic bases and potential evolutionary consequences. *Genetica* 97, 73-80.

34. Dallerac R, Labeur C, Jallon JM, Knipple DC, Roelofs WL, Wicker-Thomas C (2000) A Δ9 desaturase gene with a different substrate specificity is responsible for the cuticular diene hydrocarbon polymorphism in *Drosophila melanogaster*. *Proc Natl Acad Sci USA* 97, 9449-54.

35. Grillet M, Everaerts C, Houot B, Ritchie MG, Cobb M, Ferveur JF (2012) Incipient speciation in *Drosophila melanogaster* involves chemical signals. *Sci Rep* 2, 224.

36. Becher PG, Bengtsson M, Hansson BS, Witzgall P (2010) Flying the fly: long-range flight behavior of *Drosophila melanogaster* to attractive odors. *J Chem Ecol* 36, 599-607.

37. Carrau FM, Medina K, Boido E, Farina L, Gaggero C, Dellacassa E, Versini G, Henschke PA (2005) De novo synthesis of monoterpenes by *Saccharomyces cerevisiae* wine yeasts. *FEMS Microbiology Letters* 243(1):107-115.

38. Ramdya P, Benton R (2010) Evolving olfactory systems on the fly. *Tr Genet* 26, 307-316.

39. Orr HA, Masly JP, Presgraves DC (2004) Speciation genes. *Curr Op Genet Develop* 14(6):675-679.

40. Nosil P, Schluter D (2011) The genes underlying the process of speciation. *Tr Ecol Evol* 26(4):160-167.

41. Ruebenbauer A, Schlyter F, Hansson BS, Löfstedt C, Larsson MC (2008) Genetic variability and robustness of host odor preference in *Drosophila melanogaster*. *Curr Biol* 18, 1438-1443.

42. Wube AA, Hüfner A, Thomaschitz C, Blunder M, Kollroser M, Bauer R, Bucar F (2011) Design, synthesis and antimycobacterial activities of 1-methyl-2-alkenyl-4(1H)-quinolones. *Bioorg Medicinal Chem* 19, 567-579.

43. Virolleaud MA, Menant C, Fenet B, Piva O (2006) Total and formal enantioselective synthesis of lyngbic acid and hermitamides A and B. *Tetrahedron Letters* 47, 5127-5130.

44. Witzgall P, Frérot B (1989) Pheromone emission by individual females of carnation tortrix, *Cacoecimorpha pronubana*. *J Chem Ecol* 15, 707-717.

45. El-Sayed A, Godde J, Arn H (1999) Sprayer for quantitative application of odor stimuli. *Environm Entomol* 28(6):947-953.

46. Gonzalez F, Witzgall P, Walker WB (2016) Protocol for heterologous expression of insect odourant receptors in *Drosophila*. *Front Ecol Evol* 4, 24

47. Bischof J, Maeda RK, Hediger M, Karch F, Basler K (2007) An optimized transgenesis system for *Drosophila* using germ-line-specific phiC31 integrases. *Proc Natl Acad Sci USA* 104, 3312-3317.

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Author Contributions

S.L., P.G.B. and P.W. conceived the study. F.B.-E. and M.S. carried out behavioural studies, F.G. and W.B.W produced empty neuron flies, F.G. and H.D. did single sensillum recordings, supervised by B.S.H, E.W. and E.H. synthesized the compound, A.-L.G. calculated the confirmational analysis, M.B., G.B. and S.L. performed chemical analysis, P.W. wrote the paper with input from all co-authors.

Legends

Figure 1. Headspace analysis of *Drosophila* females and males by GC-MS and electrophysiological screening of the candidate pheromone compound *Z*4-11Al on male antennae.

(**a**) Chromatograms of headspace collections from *D. melanogaster* females (lilac traces; upper trace: amplified signal; lower trace: entire chromatogram), males (blue trace), and *D. simulans* females (green trace). The headspace of *D. melanogaster* females contains 16 yet undescribed compounds: heptanal (1), octanal (2), (*Z*)4-nonenal (3), nonanal (4), (*Z*)4-undecenal (*Z*4-11Al) (5), undecanal (6), dodecanal (7), (*Z*)4-tridecenal (8), tridecanal (9), tetradecanal (10), pentadecanal (11), (*Z*)4-hexadecenal (12), hexadecanal (13), (*Z*)4-octadecenal (14), octadecanal (15) and (*Z*)4-eicosenal (16) (see Table 1). Female-specific compounds are coloured, the most abundant cuticular hydrocarbon, 7-tricosene (17) is shown for reference, the internal standard (IS) was heptadecenyl acetate. Inset: mass spectrum of the most abundant female-specific compound *Z*4-11Al.

(**b**) Oxidation of the most abundant female cuticular hydrocarbon (*Z*,*Z*)-7,11-heptacosadiene (7,11-HD), affording two saturated and two unsaturated aldehydes, heptanal, hexadecanal, *Z*4-11Al and (*Z*)4-eicosenal.

(**c**) Single sensillum recordings (SSR) from all *D. melanogaster* olfactory sensory neurons (OSNs) with *Z*4-11Al (error bars show SEM; $n = 5$).

(**d**) SSR coupled to GC (GC-SSR), showing a response of ab9A to three different amounts of *Z*4-11Al.

Figure 2. SSR-Response of Or69a splice variants in native ab9A OSNs and in ab3A OSNs, following heterologous expression, to ten odourants.

(**a**) SSR from ab9A OSNs, in *D. melanogaster* males (Canton-S, Zimbabwe) and *D. simulans* males, which natively express both splice variants Or69aA and Or69aB.

(**b**) SSR from ab3A OSNs in *D. melanogaster,* heterologously expressing Or69aA and Or69aB, together and singly. Test panel includes the known most active ligands for Or69a (27) and three aldehydes. Cross-hatched bars indicate behaviorally active compounds (Fig. 3). Bars followed by different letters indicate statistically significant differences for each fly type ($P<0.05$; Mann-Whitney test, $n = 5$ for ab9A, $n = 10$ for ab3A).

(**c**) Key ligands for Or69aA, (*S*)-terpineol (*1*) and (*R*)-linalool (*4*), and for or Or69aB, (*R*)-carvone (*8*) and Z4-11Al (*10*).

(**d**) Alternative splicing of Or69a, where coloured boxes A and B show unique exons encoding the splice products; dark boxes show shared exons, generating co-expression of Or69aA and Or69aB in the same neurons in ab9 sensilla in *D*. *melanogaster*.

Figure 3. Z4-11Al mediates long range attraction in *D. melanogaster*.

(**a**) Upwind flights to 10 ng/min of *Z*4-11Al and (*R*)-linalool, followed by landing at the source, in *D*. *melanogaster* (Dalby) males and females, in *D*. *melanogaster* (Zimbabwe) males and *D*. *simulans* males. Lower case letters indicate statistical differences between test insect strains and species, for each treatment. Asterisks indicate significant differences between treatments (n = 40, P<0.001; binomial GLMs followed by post-hoc Wald pairwise comparison tests).

(**b**) Upwind flights to 10 ng/min of *Z*4-11Al in *D. melanogaster* (Canton-S) males expressing a tetanus toxin in OSNs expressing Or69a, and in the parental lines. Letters indicate statistical differences within treatments (n = 40, P<0.001; binomial GLM, followed by *Post-hoc* Wald pairwise comparison tests).

528 **Table 1. Saturated and unsaturated aldehydes found in headspace**
529 **collections of *D. melanogaster* (Dalby) females and males.**

Compound	Females (%±SD, *n*=10)	Males (%±SD, *n*=10)
heptanal	3.1 ± 0.8	9.9 ± 4.8
octanal	tr ***a***	0.2 ± 0.5
nonanal	4.1 ± 0.9	8.7 ± 7.9
(*Z*)-4-nonenal	tr	- ***b***
undecanal	tr	-
(*Z*)-4-undecenal	23.3 ± 1.8	-
dodecanal	7.9 ± 1.3	-
tridecanal	tr	-
(*Z*)-4-tridecenal	0.4 ± 0.9	-
tetradecanal	11.2 ± 0.8	4.9 ± 4.1
pentadecanal	3.2 ± 0.9	2.4 ± 2.1
hexadecanal	27.8 ± 3.3	69.1 ± 9.7
(*Z*)-4-hexadecenal	2.9 ± 0.3	-
octadecanal	4.7 ± 1.0	4.4 ± 3.9
(*Z*)-4-octadecenal	3.0 ± 0.6	-
(*Z*)-4-eicosenal	5.6 ± 1.7	-

530
531 ***a*** - traces
532 ***b*** - not found
533

Supplementary Information: Chemical Synthesis

Dry THF and dry Et_2O was obtained from a solvent purification system (Activated alumina columns, Pure Solv PS-MD-5, Innovative technology, Newburyport, USA) and used in the reactions when dry conditions were needed. All other chemicals were used without purification. Reactions were performed under Argon atmosphere unless otherwise stated. Flash chromatography was performed on straight-phase silica gel (Merck 60, 230–400 mesh, 0.040–0.063 mm, 10–50 g/g of product mixture) employing a gradient technique with an increasing concentration (0–100 %) of distilled ethyl acetate in distilled cyclohexane. In cases of very polar products chromatography was continued with ethanol in ethyl acetate (0-20 %). Thin-layer chromatography (TLC) was performed to monitor the progress of the reaction on silica gel plates (Merck 60, precoated aluminium foil), using ethyl acetate (40 %) in cyclohexane as an eluent, and plates were developed by means of spraying with vanillin in sulfuric acid and heating at 120°C. Purity of the product was checked with gas chromatography (GC) analysis on a Varian 3300 GC instrument equipped with a flame ionization detector (FID) using a capillary column Equity-5 (30 m x 0.25 mm i.d, d_f = 0.25 µm, with nitrogen (15 psi) as carrier gas and a split ratio of 1:20). The oven temperature was programmed at 50°C for 5 min followed by a gradual increase of 10°C min^{-1} to reach a final temperature of 300°C. An Agilent 7890 GC equipped with a polar capillary column FactorFOUR vf-23ms (30 m x 0.25 mm i.d., d_f = 0.25 µm) coupled to an Agilent 240 ion-trap MS detector for separation of some isomeric intermediates. The injector was operated in split mode (1:20) at 275°C, and a helium flow rate of 1 ml min^{-1} and a transfer line temperature of 280 °C. The analyses were performed in the external ionisation configuration. EI spectra were recorded with a mass range of m/z 50-300 at fast scan rate. Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker Avance 500 (500 MHz ^1H, 125.8 MHz ^{13}C) spectrometer using $CDCl_3$ as solvent and internal standard.

(*Z*)-4-Undecenoic acid. NaHMDS (6.78 mmol, 1 M in hexane) was added dropwise, during 30 min, to a suspension of (3-carboxypropyl)triphenylphosphonium bromide (1.45 g, 3.39 mmol) in THF (25 mL). The mixture was stirred for 2 h then cooled to 0°C on ice/water bath, and heptanal (0.387g, 3.39 mmol) in THF (2.5 mL) was added slowly during 15 min. The mixture was stirred for 5 h at 0°C then allowed to reach room temperature over night. The reaction was quenched with H_2O (20 mL) and the organic solvent was evaporated. The remaining water phase was extracted with Et_2O (3 x 20 mL), the obtained organic phases discarded and the basic aqueous phase was acidified with HCl (2M) until pH 1 and extracted with Et_2O (3 x 20 mL). The combined organic phases were dried over $MgSO_4$ (anhydr.) and the solvent evaporated off. The obtained crude product was dissolved in pentane, cooled at -18°C and filtered to remove the precipitated $OPPh_3$ followed by evaporation of the solvent to result in 0.547 g of a yellow oil (87.5% yield). ^1H-NMR: 5.52–5.30 (m, 2H), 2.35 (m, 4H), 2.04 (q, *J* = 6.5 Hz, 1.6H,

Z-isomer), 1.96 (q, *J*=6.5 Hz, 0.4H, *E*-isomer), 1.37–1.19 (m, 8H), 0.89 (t, *J*=7 Hz, 3H) ppm. The NMR data is in accordance with data previously reported (42, 48). The relationship by integration between protons at 2.04 and 1.95 indicates approximately a *Z*:*E*-ratio of 80:20 which is supported by GC-MS analysis on a Varian factorFOUR vf-23ms column. The obtained crude product was used in the next step without further purification.

Methyl (*Z*)-4-undecenoate. (*Z*)-4-Undecenoic acid (0.547 g, 2.97 mmol) from above was dissolved in methanol (15 mL) and 7 drops of concentrated H_2SO_4 were added followed by heating at 70°C over night. The mixture was allowed to reach room temperature and the methanol was evaporated and the remaining crude product was dissolved in Et_2O (15 mL). The organic phase was washed with H_2O (3 x 10 mL) and brine (2 x 10 mL), dried over Na_2SO_4 (anhydr.) and solvent evaporated resulting in 0.547 g of a yellow oil (92.8% yield). GC-MS (FactorFour vf-23ms) shows a *Z*:*E*-ratio of 80:20. ^{1}H-NMR($CDCl_3$): 5.4 (m, 2H), 3.67 (s, 3H), 2.3 (m, 4H), 2.03 (q, *J* = 6.5 Hz, 1.6H, *Z*-isomer), 1.96 (q, *J*=6.5 Hz, 0.4H, *E*-isomer), 1.33–1.21 (m, 8H), 0.89 (t, *J*=6.5 Hz, 3H) ppm (no data found in the literature). ^{13}C-NMR($CDCl_3$): 134.2, 119.9, 32.3, 31.9, 29.5, 29.3, 27.43, 22.7, 14.1 ppm. ^{13}C-NMR data similar to reported (49). Proton NMR shows a 80:20 *Z*:*E*-ratio between the diastereomers. Enrichment of the *Z*-isomer on $AgNO_3$ (10%) impregnated silica resulted in 63 mg of 98.6:1.4 *Z*:*E*-ratio according to GC-FID analysis on the vf-5 column as the diastereoisomeric purity was not possible to measure when using ^{1}H-NMR.

(*Z*)-4-Undecenol. Methyl (*Z*)-4-undecenoate (63 mg, 0.32 mmol) was dissolved in Et_2O (5 mL) and $LiAlH_4$ (2 spatel tips) was added followed by stirring at room temperature for 30 min. HCl (2 M, 2 mL) was added to quench the reaction and the mixture was extracted with Et_2O (2 x 3 mL), the combined organic layer was dried over $MgSO_4$ (anhydr.) and solvent was evaporated. Purification with flash chromatography on SiO_2 resulted in 37 mg. ^{1}H-NMR($CDCl_3$): 5.43-5.32 (m, 2H), 3.67 (m, 2H), 2.16–2.10 (m, 2H), 2.08–2.02 (m, 2H), 1.69–1.60 (m, 2H), 1.39–1.22 (m, 8H), 0.89 (t, *J*=6.5 Hz, 3H) ppm. NMR-data were similar to Kim and Hong (50) and Davis and Carlsson (51). Diastereomeric purity was checked with GC-FID before next step.

(*Z*)-4-Undecenal. (*Z*)-4-Undecenol (37 mg, 0.22 mmol) in DCM (3 mL) was added to Dess-Martin periodinane (0.140 g, 0.33 mmol) in DCM (0.5 mL). After 50 min, NaOH (2M, 10 mL) was added to quench the reaction. The two layers were separated and the aqueous phase was extracted with Et_2O (3 x 10 mL), the combined organic layers were washed with NaOH (2M, 10 mL), dried over $MgSO_4$ (anhydr.) and solvent was evaporated resulting in 30 mg of a yellow oil (81% yield). The crude product was purified with Kugelrohr distillation at bp 65–70 °C (1.6 mbar), resulted in 17 mg. ^{1}H-NMR($CDCl_3$): 9.77 (s, 1H), 5.48–5.22 (m, 2H), 2.47 (t, *J* = 7 Hz, 2H), 2.37 (q, *J* = 7 Hz, 2H), 2.04 (q, *J* = 7Hz, 2H), 1.37–1.23 (m, 8H), 0.88 (t, *J* = 7 Hz, 3H) ppm. ^{13}C-NMR($CDCl_3$): 202.1, 131.8,

127.0, 43.9, 31.8, 29.5, 29.0, 27.2, 22.6, 20.1, 14.1 ppm. Both ^{1}H- and ^{13}C-NMR data were in accordance with published results (52, 53). Analysis on GC-MS (FactorFour vf-23ms) resulted in a 98.6:1.4 *Z*:*E*-ratio, the *E*-isomer could not be detected by ^{1}H-NMR.

(*E*)-4-Undecenoic acid. 4-Pentenoic acid (0.5 g, 5 mmol) and 1-octene (2.8 g, 25 mmol) was dissolved in DCM (50 mL) and Grubbs II catalyst (85 mg, 0.1 mmol) was added and the reaction was refluxed. After 7 h was a second portion of Grubbs II catalyst (85 mg, 0.1 mmol) added and the reaction refluxed for 16 h. Reaction was allowed to reach room temperature and the solvent was evaporated. The obtained crude product was dissolved in Et_2O (50 mL) and filtered through a short pad of silica gel. The product was purified with flash chromatography by gradient elution (0–100% EtOAc in c-hexane followed by 0–10% EtOH in EtOAC) resulting in 0.52 g oil (56% yield). ^{1}H-NMR($CDCl_3$): 5.51–5.33 (m, 2H), 2.41 (q, *J*=7 Hz, 2H), 2.32 (q, *J*=7 Hz, 2H), 2.04 (q, *J*=6.5 Hz, 0.25 H, *Z*-isomer), 1.97 (q, *J*=6.5 Hz, 1.75H, *E*-isomer), 1.37–1.22 (m, 9H), 0.88 (t, *J*=7.5 Hz, 3H) ppm. The relation between proton at 2.04 and 1.97 reveals a 87.5:12.5 *E*:*Z*-ratio. The isolated product was used in the next step without further purification.

Methyl (*E*)-4-Undecenoate. (*E*)-4-Undecenoic acid (0.52 g, 2.82 mmol) was dissolved in methanol (25 mL) and a catalytic amount H_2SO_4 was added and the mixture was refluxed over night. After evaporation of the solvent the crude product was dissolved in Et_2O (10 mL) and washed with H_2O (20 mL). The aqueous phase was extracted with Et_2O (2 × 25 mL), the combined organic layer was washed with H_2O (20 mL) and brine (20 mL), dried over $MgSO_4$ (anhydr.) and evaporation of solvent resulted in 0.439 g (78% yield). ^{1}H-NMR($CDCl_3$): 5.51–5.33 (m, 2H), 3.67 (s,3H), 2.40–2.27 (m, 4H), 1.96 (q, *J*=6.5 Hz, 2H), 1.38–1.21 (m, 8H), 0.88 (t, *J*=6.5 Hz,3H) ppm. Purification with flash chromatography resulted in 0.401g (71.7% yield). GC-FID showed the same stereoisomeric ratio as for the acid above.

(*E*)-4-Undecen-1-ol. $LiAlH_4$ (0.055 g, 1.46 mmol) was added to methyl (*E*)-4-undecenoate (0.145 g, 0.73 mmol) dissolved in Et_2O (5 mL). After 30 minutes was HCl (2 M, 5 mL) added to quench the reaction. The acidic water phase was extracted with Et_2O (3 × 10 mL) and the combined organic layers were dried over $MgSO_4$ (anhydr.) and evaporation of solvent resulted in 0.104 g (99% yield). Enrichment of the *E*-isomer with medium pressure liquid chromatography (MPLC) on $AgNO_3$ (10% impregnated) silica resulted in 30 mg of a clear oil (>99.8 % *E*). ^{1}H-NMR($CDCl_3$): 5.43 (m, 2H), 3.65 (m, 2H), 2.08 (q, *J*=7 Hz, 2H), 1.97 (q, *J*=7 Hz, 2H), 1.63 (pent, 2H), 1.35–1.21 (m, 9H), 0.88 (t, *J*=6.5 Hz, 3H) ppm. ^{13}C-NMR($CDCl_3$): 134.4, 131.3, 129.4, 62.6, 32.6, 32.5, 31.7, 29.6, 29.5, 28.9, 28.8, 22.6, 14.1 ppm. All NMR-data were in accordance with previous published data (54).

(*E*)-4-Undecenal. Dess-Martin Periodinane (0.110 g, 0.26 mmol) was added to (*E*)-4-undecen-1-ol (0.030 g, 0.22 mmol) in DCM (4 mL). NaOH (2 M, 10 mL) was added after 1 h to quench reaction. The aqueous phase was extracted with Et_2O (3 × 10 mL) and the

combined organic layers were dried over $MgSO_4$ (anhydr.) and evaporation of the solvent resulted in 30 mg (98% yield). Purification of the crude with Kugelrohr distillation at 65°C (2 mbar) resulted in 10 mg of product (33% yield, 97% chemical purity, 3% undecenal). ^{1}H-NMR($CDCl_3$): 9.76 (t, *J*=1.5 Hz, 1H), 5.50–5.36 (m, 2H), 2.48 (d of t, *J*=7.5, 1.5 Hz, 2H), 2.33 (q, *J*=7 Hz, 2H), 1.97 (q, *J*=6.5 Hz, 2H), 1.32–1.19 (m, 8H), 0.87 (t, *J*=6.5 Hz, 3H). ^{13}C-NMR ($CDCl_3$): 202.5, 132.2, 127.6, 43.6, 32.5, 31.7, 29.4, 28.8, 25.2, 22.6, 14.1. The NMR-data were in accordance with published data (53, 54).

(*Z*)-4-Undecenoic acid was synthesized via a modified version of Wube and Hüfner (42) in 80% stereoisomeric purity. Esterification under acidic conditions with sulfuric acid in methanol resulted in 80% *Z*-isomer and a 93% yield over two steps. Stereoisomeric purity was controlled with NMR and GC-FID by comparing analysis for acid and ester, the appearance of a small quartet, in the NMR spectra, at 1.96 indicates the presence of *E*-isomer. Gas chromatographic separation on a polar Varian factorFOUR vf-23ms of *Z*- and *E*-ester proved that the stereochemistry was not affected by the acidic conditions during esterification. Methyl-(*Z*)-4-undecenoate was purified on regular silica gel and on silver nitrate impregnated silica gel to obtain a stereoisomeric purity of 98.6 %. Methyl-(*Z*)-4-undecenoate was reduced to (*Z*)-4-undecenol with lithium aluminum hydride in diethylether and oxidized to (*Z*)-4-undecenal with Dess-Martin periodinane in dichloromethane.

A modified version of Virolleaud´s (43) metathesis was used to produce the (*E*)-4-undecenoic acid and in a 56 % yield (87.5% of the *E*-isomer). (*E*)-4-undecenoic was esterified under the same conditions as the (*Z*)-acid and once again there was no isomerisation of the double bond (according to GC-FID and ^{1}H-NMR. The methyl-(*E*)-4-undecenoate was reduced to alcohol with $LiAlH_4$ in diethylether and purified on silver nitrate impregnated silica gel to obtain a purity of 99.8 % of the (*E*)-isomer, which was oxidized with Dess-Martin periodinane in dichloromethane to obtain the wanted (*E*)-4-undecenal.

48. Henderson BS, Larsen BS, Schwab JM (1994) Chemistry and photochemistry attending the inactivation of Escherichia coli beta-hydroxydecanoyl thiol ester dehydrase by an acetylenic diazoketone. *J Am Chem Soc* 116, 5025-5034.

49. Bus J, Sies I, Jie MSLK (1976) 13C-NMR of methyl, methylene and carbonyl carbon atoms of methyl alkenoates and alkynoates. *Chem Phys Lipids* 17, 501-518.

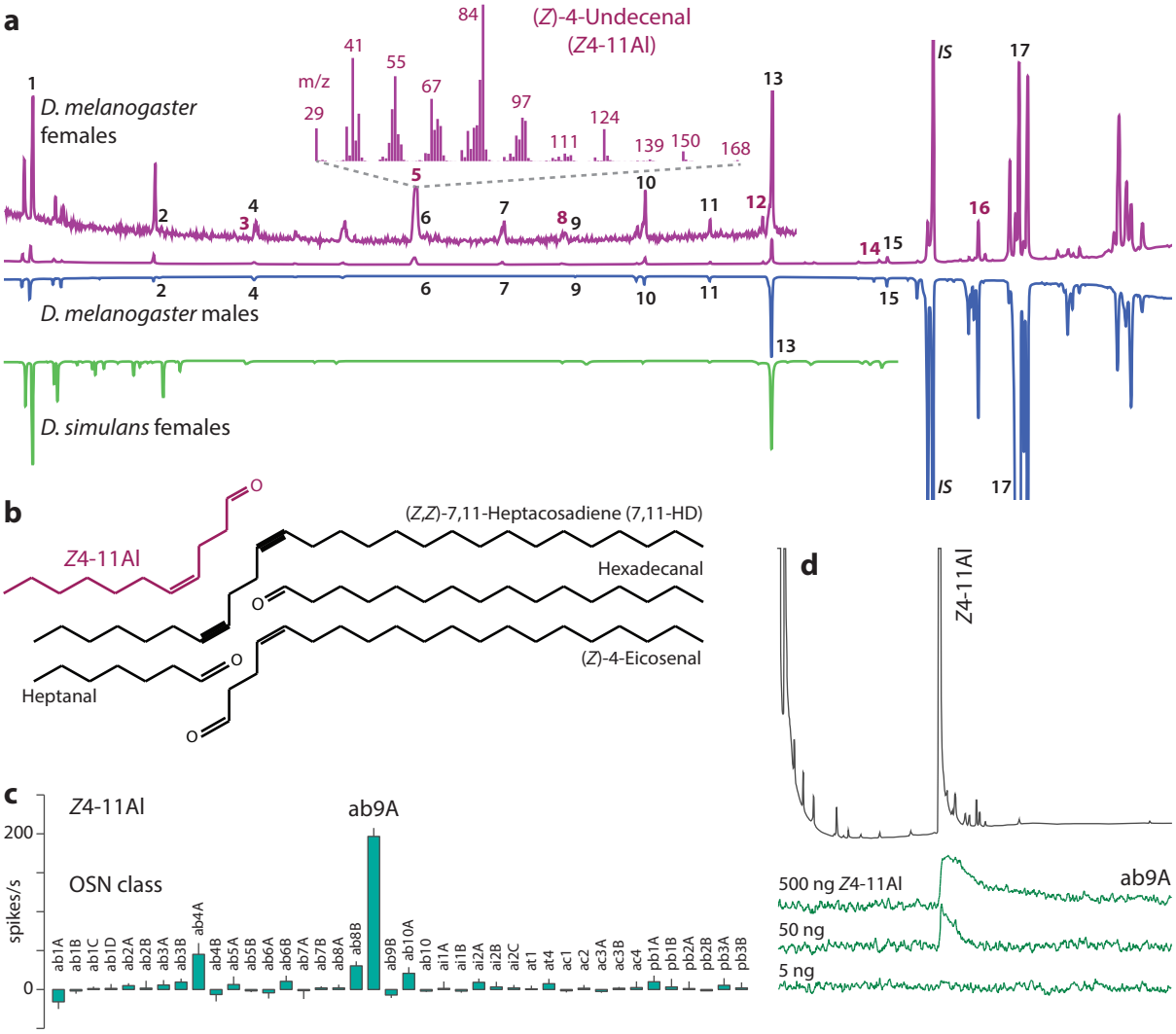
50. Kim SS, Hong YP (2011) The stereospecific synthesis of the rice leaffolder moth sex pheromone components from 1,5-cyclooctadiene. *Bull Korean Chem Soc* 32, 3120-3122.

51. Davis TL, Carlsson DA (1989) Synthesis of 7,11-dienes from enol ether and Grignard-reagents under Nickel catalysis: sex pheromones of *Drosophila melanogaster*. *Synthesis* 1989, 936-938.

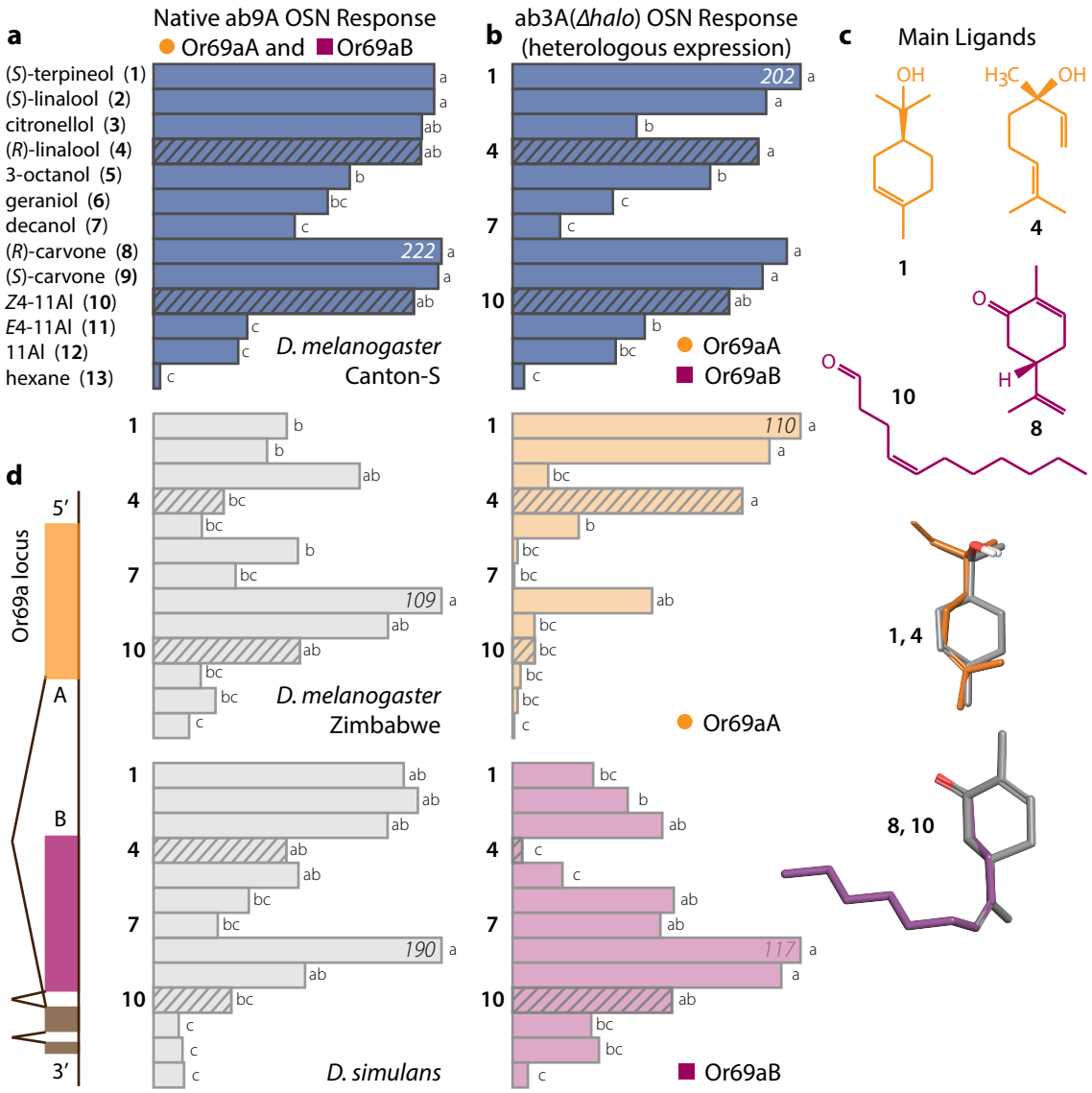
52. Wenkert E, Ferreira VF, Michelotti EL, Tingoli M (1985) Synthesis of acyclic, cis olefinic pheromones by way of nickel-catalyzed Grignard reactions. *J Org Chem* 50, 719-721.

53. Snowden RL, Brauchli R, Linder S (2011) A novel synthesis of γ,δ-unsaturated aldehydes from α-formyl-γ-lactones. *Helvetica Chim Acta* 94, 1216-1225.

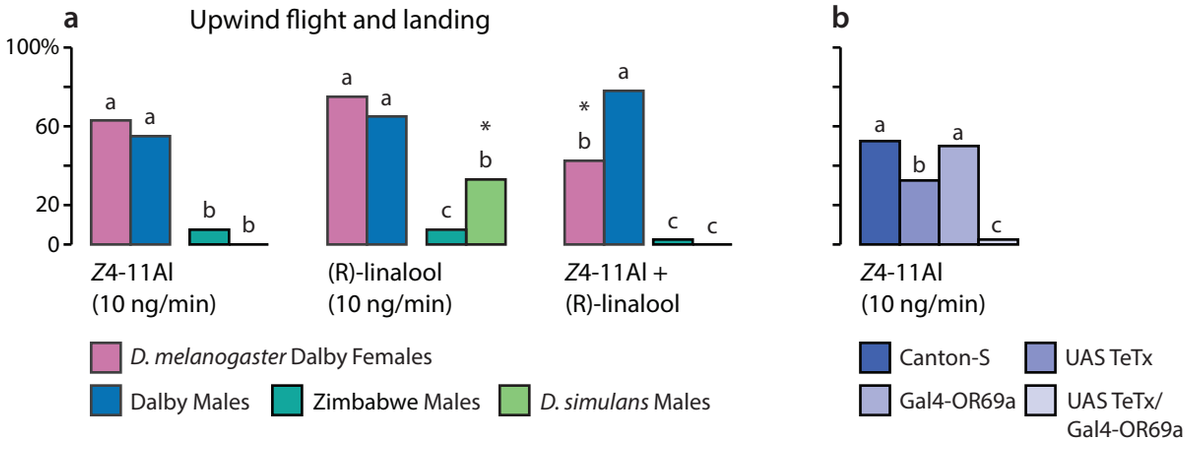
54. Pearson WH, Hutta DA, Fang WK (2000) Azidomercurations of alkenes: mercury-promoted Schmidt reactions. *J Org Chem* 65(24):8326-8332.



Lebreton Figure 1



Lebreton Figure 2



Lebreton Figure 3