

On the (un-)predictability of a large intragenic fitness landscape

Claudia Bank^{a,b,c,1}, Sebastian Matuszewski^{b,c,1}, Ryan T. Hietpas^{d,e}, Jeffrey D. Jensen^{b,c,1,*}

^a*Instituto Gulbenkian de Ciência, 2780-156 Oeiras, Portugal*

^b*School of Life Sciences, École Polytechnique Fédérale de Lausanne (EPFL), Lausanne, Switzerland*

^c*Swiss Institute of Bioinformatics (SIB), Lausanne, Switzerland*

^d*Eli Lilly and Company, Indianapolis, IN 46225*

^e*Department of Biochemistry & Molecular Pharmacology, University of Massachusetts Medical School, Worcester, MA 01605*

Abstract

The study of fitness landscapes, which aims at mapping genotypes to fitness, is receiving ever-increasing attention. Novel experimental approaches combined with NGS methods enable accurate and extensive studies of the fitness effects of mutations – allowing us to test theoretical predictions and improve our understanding of the shape of the true underlying fitness landscape, and its implications for the predictability and repeatability of evolution.

Here, we present a uniquely large multi-allelic fitness landscape comprised of 640 engineered mutants that represent all possible combinations of 13 amino-acid changing mutations at six sites in the heat-shock protein Hsp90 in *Saccharomyces cerevisiae* under elevated salinity. Despite a prevalent pattern of negative epistasis in the landscape, we find that the global fitness peak is reached via four positively epistatic mutations. Combining traditional and extending recently proposed theoretical and statistical approaches, we quantify features of the global multi-allelic fitness landscape. Using subsets of the data, we demonstrate that extrapolation beyond a known part of the landscape is difficult owing to both local ruggedness and amino-acid specific epistatic hotspots, and that inference is additionally confounded by the non-random choice of mutations for experimental fitness landscapes.

Key words: evolution, adaptation, epistasis, fitness landscapes

*To whom correspondence should be addressed. E-mail: jeffrey.jensen@epfl.ch

¹contributed equally

Author Summary

The study of fitness landscapes is fundamentally concerned with understanding the relative roles of stochastic and deterministic processes in adaptive evolution. Here, the authors present a uniquely large and complete multi-allelic intragenic fitness landscape of 640 systematically engineered mutations in yeast Hsp90. Using a combination of traditional and recently proposed theoretical approaches, they study the accessibility of the global fitness peak, and the potential for predictability of the fitness landscape topography. They report local ruggedness of the landscape and the existence of epistatic hotspot mutations, which together make extrapolation and hence predictability inherently difficult, if mutation-specific information is not considered.

Introduction

Since first proposed by Sewall Wright in 1932 [1], the idea of a fitness landscape relating genotype (or phenotype) to the reproductive success of an individual has inspired evolutionary biologists and mathematicians alike [2, 3, 4]. With the advancement of molecular and systems biology towards large and accurate data sets, it is a concept that receives increasing attention across other subfields of biology [5, 6, 7, 8, 9]. The shape of a fitness landscape carries information on the repeatability and predictability of evolution, the potential for adaptation, the importance of genetic drift, the likelihood of convergent and parallel evolution, and the degree of optimization that is (theoretically) achievable [4]. Unfortunately, the dimensionality of a complete fitness landscape of an organism – that is, a mapping of all possible combinations of mutations to their respective fitness effects – is much too high to be assessed experimentally. With the development of experimental approaches that allow for the assessment of full fitness landscapes of tens to hundreds of mutations, there is growing interest in statistics that capture the features of the landscape, and that relate an experimental landscape to theoretical landscapes of similar architecture, which have been studied extensively [10]. It is, however, unclear whether this categorization allows for an extrapolation to unknown parts of the landscape, which would be the first step towards quantifying predictability – an advance that would yield impacts far beyond the field of evolutionary biology, in particular for the clinical study of drug resistance evolution in pathogens and the development of effective vaccine and treatment strategies [8].

Existing research in this rapidly growing field comes from two sides. Firstly, different empirical landscapes have been assessed (reviewed in [4]), generally based on the combination of previously observed beneficial mutations or on the dissection of an observed adaptive walk (i.e., a combination of mutations that have been observed to be beneficial in concert). Secondly, theoretical research has proposed different landscape architectures (such as the House-of-cards, the Kauffman NK, and the Rough-Mount-Fuji model), studied their respective properties, and developed a number of statistics that characterize the landscape and quantify the expected degree of epistasis (i.e., interaction effects between mutations) [11, 12, 13, 10, 14].

The picture that emerges from these studies is mixed, reporting both smooth [15] and rugged [16, 17] landscapes with both positive epistasis (i.e., two mutations in concert are more advantageous than expected; [18]) and negative epistasis (i.e., two mutations in concert are more deleterious than expected; [19, 20]; but see [21, 22]). Current statistical approaches have been used to rank the existing landscapes by certain features [10, 14] and to assess whether they are compatible with Fisher’s Geometric Model [23]. A crucial remaining question is the extent to which the non-random choice of mutations for the experiment affects the topography of the landscape, and whether the local topography is indeed informative as to the rest of the landscape.

Here, we present an intragenic fitness landscape of 640 amino-acid changing mutations in the heat shock protein Hsp90 in *Saccharomyces cerevisiae* in a challenging environment imposed by high salinity. With all possible combinations of 13 mutations of various fitness effects at 6 positions, the presented landscape is not only uniquely large but also distinguishes itself

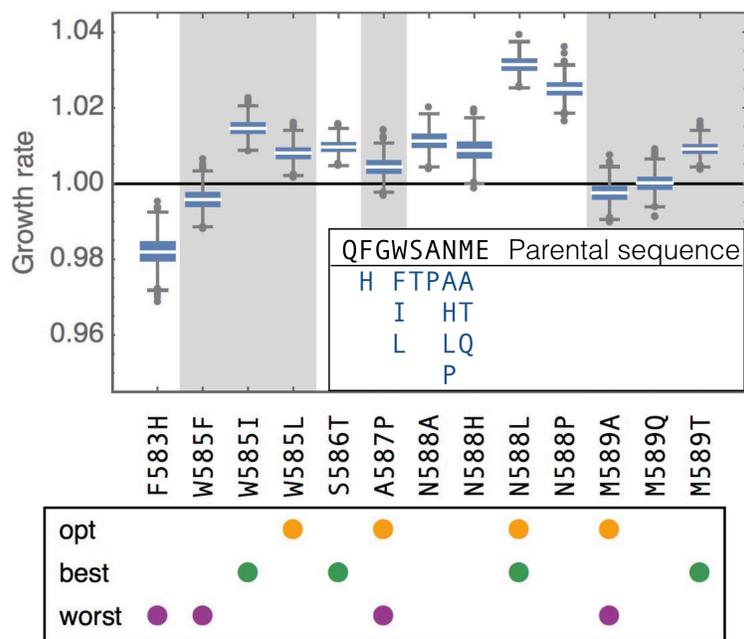


Figure 1 – Individual amino-acid substitutions and their effect on the parental background in elevated salinity, obtained from 1000 samples from the posterior distribution of growth rates. Boxes represent the interquartile range (i.e., the 50% C.I.), whiskers extend to the highest/lowest data point within the box ± 1.5 times the interquartile range, and circles represent outliers; gray and white background shading alternates by amino acid position. The box below indicates with colored dots which mutations are involved in the focal landscapes discussed throughout the main text: the four mutations leading to the global optimum (“opt”), the four individually most beneficial mutations (“best”) and the four mutations with the individually lowest growth rates (“worst”). Inset: parental sequence at positions 582-590 and assessed amino acids by position.

from previously published work regarding several other experimental features – namely, by its systematic and controlled experimental setup using engineered mutations of various selective effects, and by considering multiple alleles simultaneously. We begin by describing the landscape and identifying the global peak, which is reached through a highly positively epistatic combination of four mutations. Based on a variety of implemented statistical measures and models, we describe the accessibility of the peak, the pattern of epistasis, and the topography of the landscape. In order to accommodate our data, we extend several previously used models and statistics to the multi-allelic case. Using subsets of the landscape, we discuss the predictive potential of such modeling and the problem of selecting non-random mutations when attempting to quantify local landscapes in order to extrapolate global features.

Material and Methods

Here, we briefly outline the materials and methods used. A more detailed treatment of the theoretical work is presented in the Supporting Information.

Data generation

Codon substitution libraries consisting of 640 combinations (single, double, triple and quadruple mutants) of 13 previously isolated individual mutants within the 582-590 region of yeast Hsp90 were generated from optimized cassette ligation strategies as previously described in [24] and cloned into the p417GPD plasmid that constitutively expresses Hsp90.

Constitutively expressed libraries of Hsp90 mutation combinations were introduced into the *S. cerevisiae* shutoff strain DBY288 (*can1-100 ade2-1 his3-11,15 leu2-3,12 trp1-1 ura3-1 hsp82::leu2 hsc82::leu2 ho::pgals-hsp82-his3*) using the lithium acetate method [25]. Following transformation the library was amplified for 12 hours at 30deg C under nonselective conditions using galactose (Gal) medium with 100 $\mu\text{g}/\text{mL}$ ampicillin (1.7 g yeast nitrogen base without amino acids, 5 g ammonium sulfate, 0.1 g aspartic acid, 0.02 g arginine, 0.03 g valine, 0.1 g glutamic acid, 0.4 g serine, 0.2 g threonine, 0.03 g isoleucine, 0.05 g phenylalanine, 0.03 g tyrosine, 0.04 g adenine hemisulfate, 0.02 g methionine, 0.1 g leucine, 0.03 g lysine, 0.01 g uracil per liter with 1% raffinose and 1% galactose). After amplification the library culture was transferred to selective medium similar to Gal medium but raffinose and galactose are replaced with 2% dextrose. The culture was grown for 8 hours at 30deg C to allow shutoff of the wild-type copy of Hsp90 and then shifted to selective medium containing 0.5M NaCl for 12 generations. Samples were taken at specific time points and stored at -80degC.

Yeast lysis, DNA isolation and preparation for Illumina sequencing were performed as previously described [26]. Sequencing was performed by Elim Biopharmaceuticals, Inc and produced ≈ 30 million reads of 99% confidence at each read position based on PHRED scoring [27, 28]. Analysis of sequencing data was performed as previously described [29].

Estimation of growth rates

Individual growth rates were estimated according to the approach described by [20] using a Bayesian Monte Carlo Markov Chain (MCMC) approach proposed in [30]. Nucleotide sequences coding for the same amino acid sequence were interpreted as replicates with equal growth rates. The resulting MCMC output consisted of 10,000 posterior estimates for each amino acid mutation corresponding to an average effective samples size of 7,419 (minimum 725). Convergence was assessed using the Hellinger distance approach [31] combined with visual inspection of the resulting trace files.

Adaptive walks

In the strong selection weak mutation (SSWM) limit [32], adaptation can be modeled as a Markov process only consisting of subsequent fitness-increasing one-step substitutions that continue until an optimum is reached (so-called adaptive walks). This process is characterized

by an absorbing Markov chain with a total of n different states (i.e., mutants), consisting of k absorbing (i.e., optima) and $n - k$ transient states (i.e., non-optima). Defining $w(g)$ as the fitness of genotype g , and $g_{[i]}$ as the genotype g carrying a mutant allele at locus i , the selection coefficient is denoted by $s_j(g) = w(g_{[j]}) - w(g)$, such that the transition probabilities $p_{g,g_{[i]}}$ for going from any mutant g to any mutant $g_{[i]}$ are given by the selection coefficient normalized by the sum over all adaptive, one-mutant neighbors of the current genotype g . If g is a (local) optimum, $p_{g,g} = 1$. Putting the transition matrix \mathbf{P} in its canonical form and computing the fundamental matrix, then allows to determine the expectation and the variance in the number of steps before reaching *any* optimum, and to calculate the probability to reach optimum g when starting from genotype g' [33]. Robustness of the results and the influence of specific mutations were assessed by deleting the corresponding columns and rows in \mathbf{P} (i.e., by essentially treating the corresponding mutation as unobserved), and re-calculating and comparing all statistics to those obtained from the full data set.

Correlation of fitness effects of mutations

Strength and type of epistasis was assessed by calculating the correlation of fitness effects of mutations γ [14], which quantifies how the selective effect of a focal mutation is altered when put onto a different genetic background, averaged over all genotypes of the fitness landscape. Extending recent theory [14], we calculated the matrix of epistatic effects between different pairs of alleles (A_i, B_i) and (A_j, B_j) termed $\gamma_{(A_i, B_i) \rightarrow (A_j, B_j)}$ (eq. S1_8), the vector of epistatic effects between a specific pair of alleles (A_i, B_i) on all other pairs of alleles $\gamma_{(A_i, B_i) \rightarrow}$ (eq. S1_9), the vector of epistatic effects between all pairs of alleles on a specific allele pair (A_j, B_j) termed $\gamma_{\rightarrow (A_j, B_j)}$ (eq. S1_12), and the decay of correlation of fitness effects γ_d (eq. S1_15) with Hamming distance d averaged over all genotypes g of the fitness landscape.

Fraction of epistasis

Following [34] and [35], we quantified whether specific pairs of alleles between two loci interact epistatically, and if so whether these display magnitude epistasis (i.e., fitness effects are non-additive, but fitness increases with the number of mutations), sign epistasis (i.e., one of the two mutations considered has an opposite effect in both backgrounds) or reciprocal sign epistasis (i.e., if both mutations show sign epistasis). In particular, we calculated the type of epistatic interaction between mutations $g_{[i]}$ and $g_{[j]}$ (with $i \neq j$) with respect to a given reference genotype g over the entire fitness landscape. There was no epistatic interaction if $|s_i(g_{[j]}) - s_i(g)| < \varepsilon = 10^{-6}$, magnitude epistasis if $s_j(g)s_j(g_{[i]}) \geq 0$ and $s_i(g)s_i(g_{[j]}) \geq 0$, reciprocal sign epistasis if $s_j(g)s_j(g_{[i]}) < 0$ and $s_i(g)s_i(g_{[j]}) < 0$, and sign epistasis in all other cases [36].

Roughness-to-slope ratio

Following [11], we calculated the roughness-to-slope ratio ρ by fitting the fitness landscape to a multidimensional linear model using the least-squares method. The slope of the linear model corresponds to the average additive fitness effect [10, 23], whereas the roughness is

given by the variance of the residuals. Generally, the better the linear model fit, the smaller the variance in residuals such that the roughness-to-slope ratio approaches 0 in a perfectly additive model. Conversely, a very rugged fitness landscape would have a large residual variance and, thus, a very large roughness-to-slope ratio (as in the House-of-Cards model). In addition, we calculated a test statistic Z_ρ by randomly shuffling fitness values in the sequence space to evaluate the statistical significance of the obtained roughness-to-slope ratio from the data set ρ_{data} given by $Z_\rho = \frac{\rho_{\text{data}} - E[\rho_{\text{RSL}}]}{\sqrt{\text{Var}[\rho_{\text{RSL}}]}}$.

Results and Discussion

We used the EMPIRIC approach [24, 26] to assess the growth rate of 640 mutants in yeast Hsp90 (see Materials and Methods). Based on previous screenings of fitness effects in different environments [29] and on different genetic backgrounds [20], and on expectations of their biophysical role, 13 amino-acid changing point mutations at 6 sites were chosen for the fitness landscape presented here (Fig. 1). The fitness landscape was created by assessing the growth rate associated with each individual mutation on the parental background, and all possible mutational combinations. A previously described MCMC approach was used to assess fitness and credibility intervals ([30]; see Materials and Methods).

The fitness landscape and its global peak

Figure 2 presents the resulting fitness landscape, with each mutant represented based on its Hamming distance from the parental genotype and its median estimated growth rate. Lines connect single-step substitutions, with vertical lines occurring when there are multiple mutations at the same position (Fig. 1). With increasing Hamming distance from the parental type, many mutational combinations become strongly deleterious. This indicates strong negative epistasis between the substitutions that, as single steps on the background of the parental type, have small effects. This pattern is consistent with Fisher’s Geometric Model [37] when combinations of individually beneficial or small-effect mutations overshoot the optimum, and with classical arguments predicting negative epistasis based on mutational load [38]. It is also intuitively comprehensible on the protein level, where the accumulation of too many mutations is likely to destabilize the protein and render it dysfunctional [39].

The global peak of the fitness landscape is located 4 mutational steps away from the parental type (Fig. 2B), with 98% of posterior samples identifying the peak. The fitness advantage of the global peak reaches nearly 10% over the parental type, and is consistent between replicates (see Materials and Methods; Fig. S3_1). Though perhaps surprising given the degree of conservation of the studied genomic region ([24], their Fig. S5), it is important to note that these fitness effects are measured under highly artificial experimental conditions including high salinity, which are unlikely to represent a natural environment of yeast. The effects of the individual mutations comprising the peak in a previous experiment without added NaCl were -0.04135, -0.01876, -0.03816, -0.02115 for mutations W585L, A587P, N588L and M589A, respectively, emphasizing the potential cost of adaptation associated with the increased salinity environment (data from [20]; see also [29]).

Curiously, the global peak is not reached by combining the most beneficial single-step mutations, but via a highly synergistic combination of one beneficial and three ‘neutral’ mutations (i.e., mutations that are individually indistinguishable from the parental type in terms of growth rate). In fact, each one of the five beneficial local optima shows a similar signature of positive epistasis (Fig. S3_2). Figure 2C demonstrates that a combination of the four mutations involved in the global peak (termed “opt”) predicts only a 4% fitness advantage. Furthermore, even a combination of the four individually most beneficial single-step mutations in the data set (“best”; considering at most one mutation per position) only predicts a benefit of 6%. Notably, the actual combination of these four mutations is highly deleterious

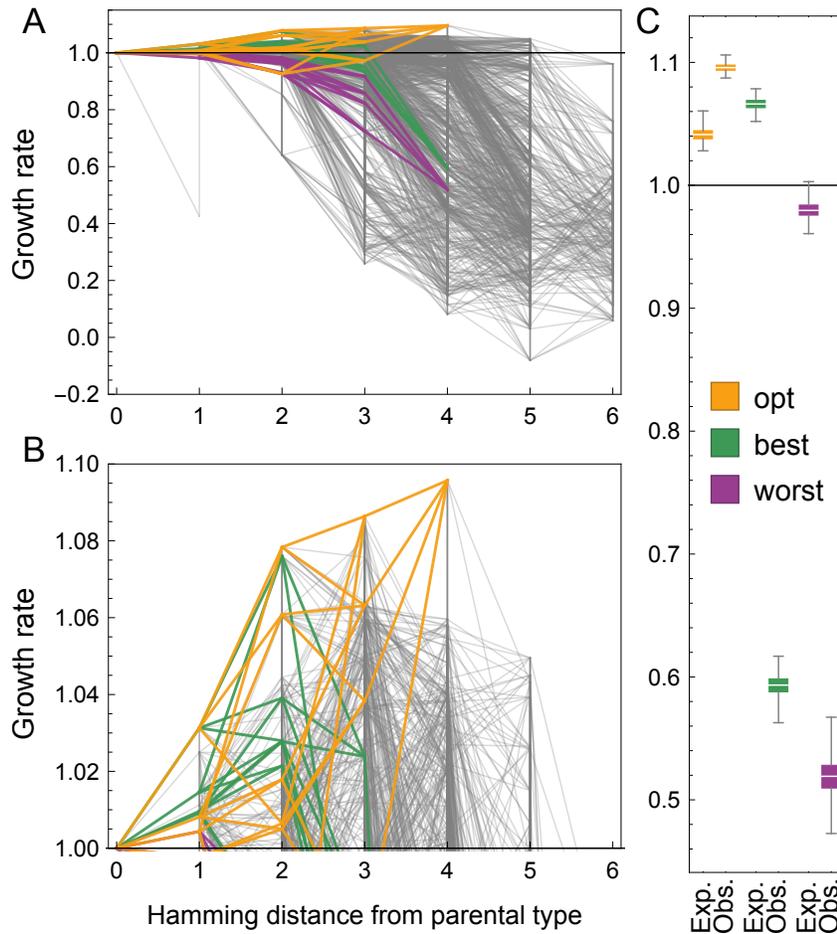


Figure 2 – (A) Empirical fitness landscape by mutational distance to parental type. Each line represents a single substitution. Vertical lines appear when multiple alleles have been screened at the same position. There is a global pattern of negative epistasis. The three focal landscapes are highlighted. (B) Close-up on the beneficial portion of the landscape. (C) Expected ("Exp.") versus observed ("Obs.") fitness for the focal landscapes, obtained from 1000 posterior samples. We observe strong positive epistasis in the landscape that contains the global optimum, whereas the other two are dominated by negative epistasis. In all panels, the y axis depicts growth rate as a proxy for fitness.

and thus exhibits strong negative epistasis. Although negative epistasis between beneficial mutations during adaptation has been reported more frequently, positive epistasis has also been observed occasionally [18, 40], particularly in the context of compensatory evolution. In fact, negative epistasis between beneficial mutations and positive epistasis between neutral mutations has been predicted by de Visser *et al.* [41]. Furthermore, our results support the pattern recently found in the gene underlying the antibiotic resistance enzyme TEM-1 beta-lactamase in *E. coli*, showing that large-effect mutations interact more strongly than small-effect mutations such that the fitness landscape of large-effect mutations tends to be more rugged than the landscape of small-effect mutations [13], and that mutations that were selected for their combined beneficial effect tend to interact synergistically, whereas mutations selected for their individual effects interact antagonistically [13, 4].

Adaptive walks on the fitness landscape

Next, we studied the empirical fitness landscape within a framework recently proposed by Draghi and Plotkin [42]. Given the empirical landscape, we simulated adaptive walks and studied the accessibility of the six observed local optima. In addition, we evaluated the length of adaptive walks starting from any mutant in the landscape, until an optimum is reached. In the strong selection weak mutation limit [43], we can express the resulting dynamics as an absorbing Markov chain, where local optima correspond to the absorbing states, and in which the transition probabilities correspond to the relative fitness increases attainable by the neighboring mutations (see Materials and Methods). This allowed us to derive analytical solutions for the mean and variance of the number of steps to reach a fitness optimum (see extended Materials and Methods), and the probability to reach a particular optimum starting from any given mutant in the landscape (Figs. 3, S3_3, S3_4).

Using this framework, we find that the global optimum can be reached with non-zero probability from almost 95% of starting points in the landscape, and is reached with high probability from a majority of starting points - indicating high accessibility of the global optimum (Fig. 3). The picture changes when restricting the analysis to adaptive walks initiating from the parental type (Figs. 3, S3_3, S3_4). Here, although 73% of all edges and 78% of all vertices are included in an adaptive walk to the global optimum, it is reached with only 26% probability. A local optimum two substitutions away from the parental type (Fig. 3C) is reached with a much higher probability of 47%. Hence, adaptation on the studied landscape is likely to stall at a sub-optimal fitness peak. This indicates that the local and global landscape pattern may be quite different, an observation that is confirmed and discussed in more detail below. In line with the existence of multiple local fitness peaks, we find that pairs of alleles at different loci show pervasive sign (30%) and reciprocal sign (8%) epistasis [34], whereas the remaining 62% are attributed to magnitude epistasis (i.e., there is no purely additive interaction between alleles; for a discussion of the contribution of experimental error see Fig. S3_5).

Epistasis measures and the topography of the fitness landscape

Next, we considered the global topography of the fitness landscape. Various measures of epistasis and ruggedness have been proposed, most of them correlated and hence capturing

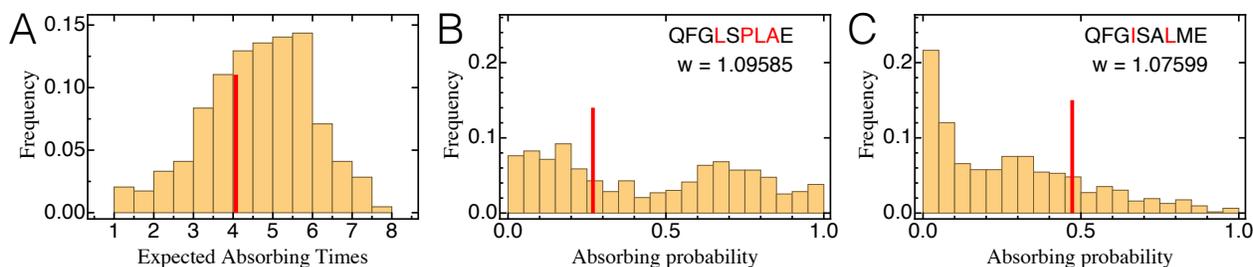


Figure 3 – (A) Distribution of average lengths of adaptive walks starting from any type in the full landscape (i.e., absorbing times of the Markov chain). The red line indicates mean path length for adaptive walks from the parental genotype. (B) & (C): Distribution of absorbing probabilities, that is, the probability to reach a specific optimum starting from a given genotype computed for all genotypes in the data set. The red line corresponds to the respective probability when starting from the parental sequence. The global optimum (panel B) is in general reached with a very high probability, but there are starting points from which it is poorly accessible.

similar features of the landscape [10]. However, drawing conclusions has proven difficult because the studied landscapes were created according to different criteria. Furthermore, the majority of published complete landscapes are too small to be divided into subsets [but see 10, 12], preventing tests for the consistency and hence the predictive potential of landscape statistics. The landscape studied here provides us with this opportunity. Moreover, because multiple alleles at the same site are contained within the landscape, we may study whether changes in the shape of the landscape are site- or amino-acid specific.

We computed various landscape statistics (roughness-to-slope ratio, fraction of epistasis, and the recently proposed gamma statistics; see SI 1) [11, 10, 14], and compared them to expectations from theoretical landscape models (NK, Rough-Mount-Fuji (RMF), House-of-Cards (HoC), Egg-Box landscapes; for brief definitions of these see SI 2). Whenever necessary, we provide an analytical extension of the used statistic to the case of multi-allelic landscapes (see Materials and Methods). To assess consistency and predictive potential, we computed the whole set of statistics for (1) all landscapes in which one amino acid was completely removed from the landscape (a cross-validation approach [44], subsequently referred to as the ‘drop-one’ approach), (2) all possible 360 di-allelic sub-landscapes, and (3) for all 1,570 di-allelic 4-step landscapes containing the parental genotype, highlighting as special examples the three focal landscapes discussed.

We find that the general topography of the fitness landscape resembles that of a RMF landscape with intermediate ruggedness, which is characterized by a mixture of a random HoC component and an additive component (Fig. 4A,B). Whereas the whole set of landscape statistics supports this topography and our conclusions, the gamma statistics measuring landscape-averaged correlations in fitness effects, recently proposed by [14], proved to be particularly illustrative. We will therefore focus on these in the main text; we refer to the Supplementary Material for additional results (e.g., measurement uncertainty and adaptive walks).

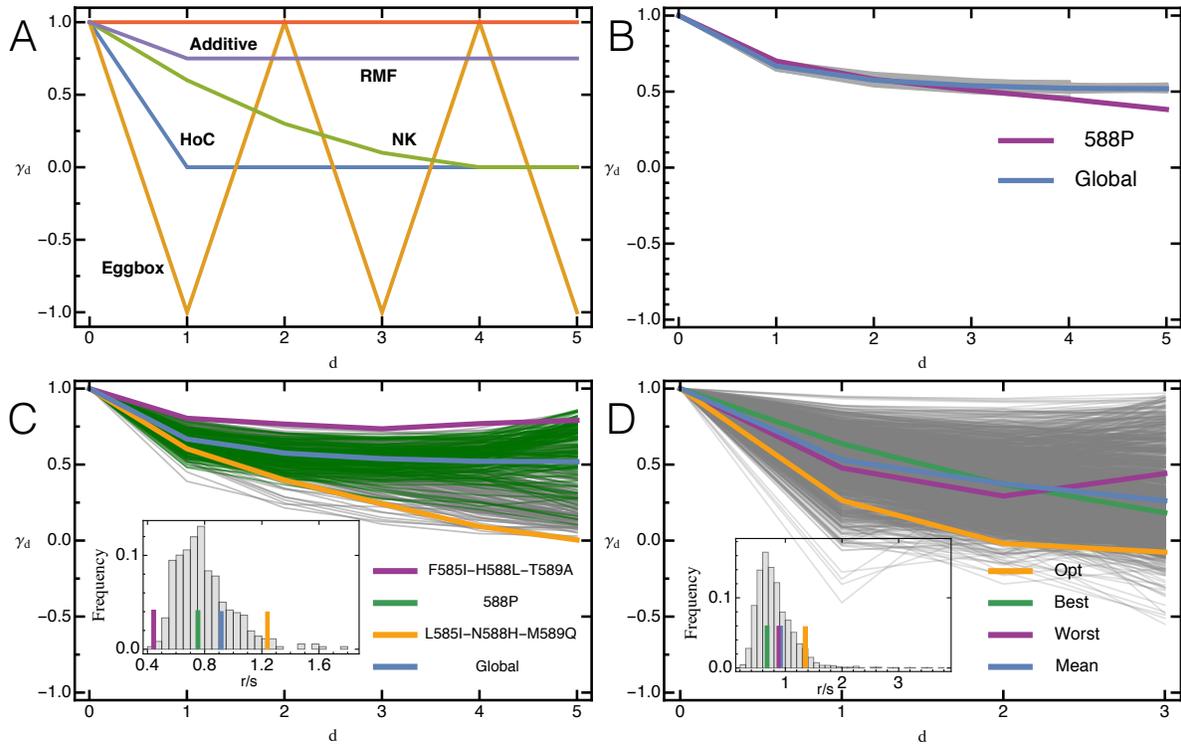


Figure 4 – (A) Expected pattern of landscape-wide epistasis measure γ_d (eq. S1_15) with mutational distance for theoretical fitness landscapes with 6 loci from [14]. (B) Observed decay of γ_d with mutational distance under the drop-one approach is quite homogenous, except when hotspot mutation 588P is removed. 95% C.I. are contained within the lines for the global landscape. (C) Observed decay of γ_d for all di-allelic 6-locus sub-landscapes. Depending on the underlying mutations, γ_d is vastly different suggesting qualitative differences in the topography of the underlying fitness landscape and in the extent of additivity in the landscape. Three focal landscapes representative of different types, and γ_d for the full landscape have been highlighted. (D) Observed decay of γ_d for all di-allelic 4-locus sub-landscapes containing the parental type, indicative of locally different landscape topographies. Highlighted are the three focal landscapes. Insets: Histograms of the roughness-to-slope ratio r/s for the respective subset, with horizontal lines indicating values for highlighted landscapes (r/s for global landscape (blue) is significantly different from HoC expectation, $p = 0.01$). Similar to γ_d , there is a huge variation in r/s for subparts of the fitness landscape, especially when considering the 4-locus subsets.

Predictive potential of landscape statistics

When computed based on the whole landscape and on a drop-one approach, the landscape appears quite homogeneous, and the gamma statistics show relatively little epistasis (Fig. 4B, 5B). On first sight, this contradicts our earlier statement of strong negative and positive epistasis but can be understood given the different definitions of the epistasis measures used: Above, we have measured epistasis based on the deviation from the multiplicative combination of the single-step fitness effects of mutations on the parental background. As these effects were small, epistasis was strong in comparison. Conversely, the gamma measure is independent of a reference genotype and captures the fitness decay with a growing number of substitutions as a dominant and quite additive component of the landscape.

Only mutation 588P has a pronounced effect on the global landscape statistics, and seems to act as an epistatic hotspot by making a majority of subsequent mutations (of individually small effect) on its background strongly deleterious (clearly visible in Fig. 4B, 5C). This can be explained by looking at the biophysical properties of this mutation. In wild-type Hsp90, amino acid 588N is oriented away from solvent and forms hydrogen bond interactions with neighboring amino acids [24]. Proline lacks an amide proton, which inhibits hydrogen bond interactions. As a result, substituting 588N with a proline could disrupt hydrogen bond interactions with residues that may be involved in main chain hydrogen bonding and destabilize the protein. In addition, the pyrrolidine ring of proline is extremely rigid and can constrain the main chain, which may restrict the conformation of the residue preceding it in the protein sequence [45].

The variation between inferred landscape topographies increases dramatically for the 360 di-allelic 6-locus sub-landscapes (Fig.4C). Whereas they are still largely compatible with an RMF landscape, the decay of landscape-wide epistasis with mutational distance (as measured by γ_d) shows a large variance, suggesting large differences in the degree of additivity. Interestingly, various sub-landscapes, typically carrying mutation *588P, show a relaxation of epistatic constraint with increasing mutational distance (i.e., increasing γ_d) that is not captured by any of the proposed theoretical fitness landscape models, suggestive of systematic compensatory interactions (but see the eggbox model for an explicit example featuring non-monotonicity in γ_d). The variation in the shape of the fitness (sub-)landscapes is also reflected in the corresponding roughness-to-slope ratio (inset of Fig. 4C-D), further emphasizing inhomogeneity of the fitness landscape with local epistatic hotspots.

Finally, the 1,570 di-allelic 4-locus landscapes containing the parental genotype, though highly correlated genetically, reflect a variety of possible landscape topographies (Fig.4D), ranging from almost additive to egg-box shapes, accompanied by an extensive range of roughness-to-slope ratios. The three focal landscapes discussed above are not strongly different compared with the overall variation; yet show diverse patterns of epistasis between substitutions (Fig. 5).

Thus, predicting fitness landscapes is difficult indeed. Extrapolation of the landscape, even across only a single mutation, may fail due to the existence of local epistatic hotspot mutations. While the integration of biophysical properties into landscape models is an important step forward [e.g. 46], we demonstrate that such models need to be mutation-specific. Con-

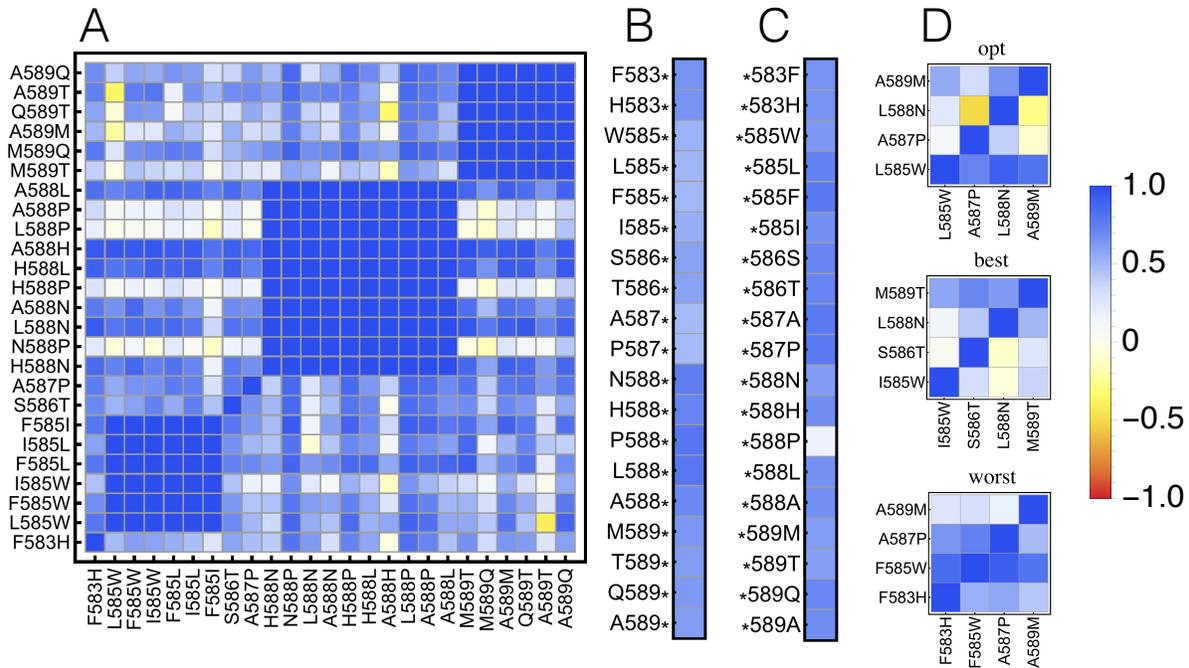


Figure 5 – (A) Epistasis measure $\gamma_{(A_i, B_i) \rightarrow (A_j, B_j)}$ (eq. S1_8) between any two substitutions, averaged across the entire landscape. The majority of interactions are small to moderate (blue). Parts of the fitness landscape show highly localized and mutation-specific epistasis, ranging from strong magnitude epistasis (white) to sign and reciprocal sign epistasis (yellow). (B) The average epistatic effect $\gamma_{A_i \rightarrow}$ (eq. S1_10) of a mutation occurring on any background is always small. (C) The average epistatic effect $\gamma_{\rightarrow A_j}$ (eq. S1_13) of a background on any new mutation is usually small, except for background *588P, which shows a strong magnitude effect. (D) Locus-specific gamma for the 4 mutations leading to the global optimum (top), the 4 largest single effect mutations (middle), and the single effect mutations with the lowest fitness (bottom). The “opt” landscape exhibits strong sign epistasis between locus 587 and 588, and 588 and 589. Also the “best” landscape exhibits pervasive epistasis with sign epistasis between locus 588 and locus 585 and 586, respectively. We observe almost no epistasis in the “worst” landscape.

sidering a site-specific model (e.g., BLOSUM matrix; [47]) is not sufficient. Newer models such as DeepAlign may provide the opportunity to allow integration of mutation-specific effects via aligning two protein structures based on spatial proximity of equivalent residues, evolutionary relationship and hydrogen bonding similarity [48].

Conclusion

Originally introduced as a metaphor to describe adaptive evolution, fitness landscapes promise to become a powerful tool in biology to address complex questions regarding the predictability of evolution and the prevalence of epistasis within and between genomic regions. Due to their high-dimensional nature, however, the ability to extrapolate will be paramount to progress in this area, and the optimal quantitative and qualitative approaches to achieve this goal are yet to be determined.

Here, we have taken an important step towards addressing this question via the creation and analysis of a landscape comprising 640 engineered mutants of the Hsp90 protein in yeast. The unprecedented size of the fitness landscape along with the multi-allelic nature allows us to test whether global features could be extrapolated from subsets of the data. Although the global pattern indicates a rather homogeneous landscape, smaller sub-landscapes are a poor predictor of the overall global pattern because of ‘epistatic hotspots’.

In combination, our results highlight the inherent difficulty imposed by the duality of epistasis for predicting evolution. In the absence of epistasis (i.e., in a purely additive landscape) evolution is globally highly predictable as the population will eventually reach the single fitness optimum, but the path taken is locally entirely unpredictable. Conversely, in the presence of (sign and reciprocal sign) epistasis evolution is globally unpredictable, as there are multiple optima and the probability to reach any one of them depends strongly on the starting genotype. At the same time, evolution may become locally predictable with the population following obligatory adaptive paths that are a direct result of the creation of fitness valleys owing to epistatic interactions.

The empirical fitness landscape studied here appears to be intermediate between these extremes. Although the global peak is within reach from almost any starting point, there is a local optimum that will be reached with appreciable probability, particular when starting from the parental genotype. From a practical standpoint, these results thus highlight the danger inherent to the common practice of constructing fitness landscapes from ascertained mutational combinations. However, this work also suggests that one promising way forward for increasing predictive power will be the utilization of multiple small landscapes used to gather information about the properties of individual mutations, combined with the integration of site-specific biophysical properties.

Acknowledgements

This project was funded by grants from the Swiss National Science Foundation (FNS) and a European Research Council (ERC) Starting Grant to JDJ. Computations were performed at the Vital-IT (<http://www.vital-it.ch>) Center for high-performance computing of the Swiss

Institute of Bioinformatics (SIB). We thank Dan Bolon, Brian Charlesworth, Pamela Cote, Inês Fragata, and Hermina Ghenu for helpful comments and discussion.

References

- [1] Wright S (1932) The roles of mutation, inbreeding, crossbreeding, and selection in evolution. *Proc. 6th Int. Congress Genet.* pp. 356–366.
- [2] Coyne JA, Barton NH, Turelli M (1997) Perspective: A Critique of Sewall Wright’s Shifting Balance Theory of Evolution. *Evolution* 51(3):643–671.
- [3] Gavrilets S (2004) *Fitness landscapes and the origin of species.* (Princeton University Press, Princeton, NJ).
- [4] de Visser JAGM, Krug J (2014) Empirical fitness landscapes and the predictability of evolution. *Nat Rev Genet* 15(7):480–490.
- [5] Costanzo M et al. (2010) The Genetic Landscape of a Cell. *Science* 327(5964):425–431.
- [6] Jiménez JI, Xulvi-Brunet R, Campbell GW, Turk-Macleod R, Chen IA (2013) Comprehensive experimental fitness landscape and evolutionary network for small RNA. *Proc Natl Acad Sci USA* 110(37):14984–14989.
- [7] Huang S (2013) Genetic and non-genetic instability in tumor progression: link between the fitness landscape and the epigenetic landscape of cancer cells. *Cancer Metastasis Rev.* 32(3-4):423–448.
- [8] Mann JK et al. (2014) The fitness landscape of HIV-1 gag: advanced modeling approaches and validation of model predictions by in vitro testing. *PLoS Comput. Biol.* 10(8):e1003776.
- [9] Liu G, Rancati G (2016) Adaptive Evolution: Don’t Fix What’s Broken. *Curr Biol* 26(4):R169–71.
- [10] Szendro IG, Schenk MF, Franke J, Krug J (2013) Quantitative analyses of empirical fitness landscapes. *J. Stat. Mech.*
- [11] Aita T, Iwakura M, Husimi Y (2001) A cross-section of the fitness landscape of dihydrofolate reductase. *Protein Eng.* 14(9):633–638.
- [12] Franke J, Klözer A, de Visser JAGM, Krug J (2011) Evolutionary accessibility of mutational pathways. *PLoS Comput. Biol.* 7(8):e1002134.
- [13] Schenk MF, Szendro IG, Salverda MLM, Krug J, de Visser JAGM (2013) Patterns of Epistasis between Beneficial Mutations in an Antibiotic Resistance Gene. *Mol Biol Evol* 30(8):1779–1787.
- [14] Ferretti L et al. (2016) Measuring epistasis in fitness landscapes: The correlation of fitness effects of mutations. *J Theor Biol* 396:132–143.
- [15] Kryazhimskiy S, Rice DP, Jerison ER, Desai MM (2014) Microbial evolution. Global epistasis makes adaptation predictable despite sequence-level stochasticity. *Science* 344(6191):1519–1522.

- [16] Khan AI, Dinh DM, Schneider D, Lenski RE, Cooper TF (2011) Negative epistasis between beneficial mutations in an evolving bacterial population. *Science* 332(6034):1193–1196.
- [17] Schoustra SE, Bataillon T, Gifford DR, Kassen R (2009) The properties of adaptive walks in evolving populations of fungus. *PLoS Biol* 7(11):e1000250.
- [18] Weinreich D, Delaney N, DePristo M (2006) Darwinian Evolution Can Follow Only Very Few Mutational Paths to Fitter Proteins. *Science* 312(5770):111–114.
- [19] Dickinson WJ (2008) Synergistic fitness interactions and a high frequency of beneficial changes among mutations accumulated under relaxed selection in *Saccharomyces cerevisiae*. *Genetics* 178(3):1571–1578.
- [20] Bank C, Hietpas RT, Jensen JD, Bolon DNA (2015) A systematic survey of an intragenic epistatic landscape. *Mol Biol Evol* 32(1):229–238.
- [21] MacLean RC, Hall AR, Perron GG, Buckling A (2010) The population genetics of antibiotic resistance: integrating molecular mechanisms and treatment contexts. *Nat Rev Genet* 11(6):405–414.
- [22] Jasmin JN, Lenormand T (2016) Accelerating Mutational Load Is Not Due to Synergistic Epistasis or Mutator Alleles in Mutation Accumulation Lines of Yeast. *Genetics* 202(2):751–763.
- [23] Blanquart F, Achaz G, Bataillon T, Tenaillon O (2014) Properties of selected mutations and genotypic landscapes under Fisher’s geometric model. *Evolution* 68(12):3537–3554.
- [24] Hietpas RT, Jensen JD, Bolon DNA (2011) Experimental illumination of a fitness landscape. *Proc Natl Acad Sci USA* 108(19):7896–7901.
- [25] Gietz DR, Woods RA (2002) Transformation of yeast by lithium acetate/single-stranded carrier DNA/polyethylene glycol method. *Methods in Enzymology* 350:87–96.
- [26] Hietpas R, Roscoe B, Jiang L, Bolon DNA (2012) Fitness analyses of all possible point mutations for regions of genes in yeast. *Nat Protoc* 7(7):1382–1396.
- [27] Ewing B, Green P (1998) Base-calling of automated sequencer traces using Phred. II. Error probabilities. *Genome Research* 8:186–194.
- [28] Ewing B, Hillier L, Wendl MC, Green P (1998) Base-Calling of Automated Sequencer Traces Using Phred. I. Accuracy Assessment. *Genome Research* 8:175–185.
- [29] Hietpas RT, Bank C, Jensen JD, Bolon DNA (2013) Shifting fitness landscapes in response to altered environments. *Evolution* 67(12):3512–3522.
- [30] Bank C, Hietpas RT, Wong A, Bolon DN, Jensen JD (2014) A Bayesian MCMC approach to assess the complete distribution of fitness effects of new mutations: uncovering the potential for adaptive walks in challenging environments. *Genetics* 196(3):841–852.

- [31] Boone EL, Merrick JRW, Krachey MJ (2012) A Hellinger distance approach to MCMC diagnostics. *Journal of Statistical Computation and Simulation* 84(4):833–849.
- [32] Gillespie JH (1983) A simple stochastic gene substitution model. *Theor Popul Biol* 23(2):202–215.
- [33] Kemeny JG, Snell JL (1960) Finite markov chains (van Nostrand Princeton, NJ).
- [34] Weinreich DM, Watson RA, Chao L (2005) Perspective: Sign Epistasis and Genetic Constraint on Evolutionary Trajectories. *Evolution* 59(6):1165–1174.
- [35] Poelwijk FJ, Kiviet DJ, Weinreich DM, Tans SJ (2007) Empirical fitness landscapes reveal accessible evolutionary paths. *Nature* 445(7126):383–386.
- [36] Brouillet S, Annoni H, Ferretti L, Achaz G (2015) MAGELLAN: a tool to explore small fitness landscapes. *bioRxiv* p. 031583.
- [37] Weinreich DM, Knies JL (2013) Fisher’s geometric model of adaptation meets the functional synthesis: data on pairwise epistasis for fitness yields insights into the shape and size of phenotypic space. *Evolution* 67(10):2957–2972.
- [38] Kimura M, Maruyama T (1966) Mutational Load with Epistatic Gene Interactions in Fitness. *Genetics* 54(6):1337–1351.
- [39] Soskine M, Tawfik DS (2010) Mutational effects and the evolution of new protein functions. *Nat Rev Genet* 11(8):572–582.
- [40] Trindade S et al. (2009) Positive epistasis drives the acquisition of multidrug resistance. *PLoS Genet.* 5(7):e1000578.
- [41] de Visser JA, Hoekstra RF, van den Ende H (1997) An experimental test for synergistic epistasis and its application in *Chlamydomonas*. *Genetics* 145(3):815–819.
- [42] Draghi JA, Plotkin JB (2013) Selection biases the prevalence and type of epistasis along adaptive trajectories. *Evolution* 67(11):3120–3131.
- [43] Gillespie JH (1984) Molecular Evolution Over the Mutational Landscape. *Evolution* 38(5):1116–1129.
- [44] Geisser S (1993) *Predictive Inference*. (CRC Press, New York, NY, USA) Vol. 55.
- [45] Bajaj K et al. (2007) Stereochemical criteria for prediction of the effects of proline mutations on protein stability. *PLoS Comput. Biol.* 3(12):e241.
- [46] Wylie CS, Shakhnovich EI (2011) A biophysical protein folding model accounts for most mutational fitness effects in viruses. *Proc Natl Acad Sci USA* 108(24):9916–9921.
- [47] Henikoff S, Henikoff JG (1992) Amino acid substitution matrices from protein blocks. *Proc Natl Acad Sci USA* 89(22):10915–10919.

- [48] Wang S, Ma J, Peng J, Xu J (2013) Protein structure alignment beyond spatial proximity. *Sci Rep* 3:1448.
- [49] Fisher RA (1930) *The genetical theory of natural selection*. (Clarendon Press, Oxford, UK).
- [50] Wright S (1931) Evolution in Mendelian Populations. *Genetics* 16(2):97.
- [51] McCandlish DM (2011) Visualizing fitness landscapes. *Evolution* 65(6):1544–1558.
- [52] Fisher RA (1918) The correlation between relatives on the supposition of Mendelian inheritance. *Trans R Soc Edinburgh* 52:399–433.
- [53] Kingman JFC (1978) A simple model for the balance between selection and mutation. *J Appl Probab* 15(1):1–12.
- [54] Neidhart J, Szendro IG, Krug J (2014) Adaptation in Tunably Rugged Fitness Landscapes: The Rough Mount Fuji Model. *Genetics* 198(2):699–721.
- [55] Kauffman S, Levin S (1987) Towards a general theory of adaptive walks on rugged landscapes. *J Theor Biol* 128(1):11–45.
- [56] Kauffman SA (1993) *The origins of order: Self organization and selection in evolution*. (Oxford University Press, USA).
- [57] Aita T et al. (2000) Analysis of a local fitness landscape with a model of the rough Mt. Fuji-type landscape: application to prolyl endopeptidase and thermolysin. *Biopolymers* 54(1):64–79.
- [58] Schmiegelt B, Krug J (2014) Evolutionary Accessibility of Modular Fitness Landscapes. *J Stat Phys* 154(1):334–355.

Supporting Information 1: Extended Materials and Methods

Adaptive walks

Under the strong selection weak mutation (SSWM) limit [43], adaptation follows an absorbing Markovian process characterized by a series of fitness-increasing substitutions along one-mutant neighbours until reaching a fitness optimum (forming a so-called adaptive walk), with a total of n different states (i.e., mutants), consisting of k absorbing (i.e., optima) and $n - k$ transient states (i.e., non-optima).

Defining $w(g)$ as the fitness of genotype g , and $g_{[j]}$ as the genotype g carrying a mutant allele at locus j , the selection coefficient is denoted by

$$s_j(g) = w(g_{[j]}) - w(g), \quad (\text{S1_1})$$

such that the transition probabilities for going from any given genotype g to any mutant genotype \tilde{g} are given by the transition matrix \mathbf{P}

$$\mathbf{P} = \begin{cases} 0 & \mathcal{D}_{\text{HD}}(g, \tilde{g}) > 1 \text{ or } s_j(g) \leq 0 \text{ with } \tilde{g} = g_{[j]} \\ \frac{1 - \exp[-2s_j(g)]}{\sum_{i \in M(g)} (1 - \exp[2s_i(g)])} & \text{if } \tilde{g} \in M(g) \text{ with } \tilde{g} = g_{[j]} \\ 1 & \text{if } M(g) = \emptyset \text{ and } \tilde{g} = g, \end{cases} \quad (\text{S1_2})$$

where $\mathcal{D}_{\text{HD}}(g, \tilde{g})$ marks the Hamming distance between two genotypes g and \tilde{g} (i.e., the sum of pairwise allele differences across all loci), and $M(g) := \{j : s_j(g) > 0, \mathcal{D}_{\text{HD}}(g, g_{[j]}) = 1\}$ denotes the set of all adaptive, one-mutant neighbours of the current genotype g . Note that when considering more than two alleles per locus, $g_{[j]}$ (and accordingly $s_j(g)$) are not uniquely determined and – apart from the focal genotype g – depend on the (mutant) allele found at locus j . However, since both entities become explicit once \tilde{g} is specified (and for keeping notation simple), we prefer to define a single selection coefficient (i.e., eq. S1_1) only, that is used consistently in all our approaches. Furthermore note, that transition probabilities are given by the classical fixation probability of a given genotype [49, 50] renormalized over the fixation probability of all beneficial genotypes in a one-mutant neighbourhood of the current genotype g . [51] recently used an even more general form of \mathbf{P} which also took into account differences in mutation rates and also allowed for the fixation of slightly deleterious and neutral mutations. However, under the assumption that there is no mutational bias (i.e., all single-step mutants are equally likely to occur) and that selection is strong as compared to mutation (i.e., the SSWM regime), both definitions are identical.

The canonical form of \mathbf{P} can then be obtained by permutation, such that

$$\mathbf{P} = \begin{pmatrix} \mathbf{Q} & \mathbf{R} \\ \mathbf{0} & \mathbf{I}_k \end{pmatrix}, \quad (\text{S1_3})$$

where \mathbf{Q} is a $(n - k) \times (n - k)$ matrix which contains the transition probabilities between transient states; \mathbf{R} is a $(n - k) \times k$ matrix which gives the transition probabilities from any

transient to any absorbing state; $\mathbf{0}$ is the $k \times (n - k)$ zero matrix; and \mathbf{I}_k is the $k \times k$ identity matrix [33].

Using the above representation, all basic properties of the absorbing Markov chain can be calculated from the fundamental matrix

$$\mathbf{N} = \sum_{i=0}^{\infty} \mathbf{Q}^i = (\mathbf{I}_{(n-k)} - \mathbf{Q})^{-1}. \quad (\text{S1_4})$$

In particular, the expected number of steps before absorption (i.e., the expected number of steps on the fitness landscape before reaching any optimum) is given by

$$\mathbf{E}[\mathbf{t}] = \mathbf{N}\mathbf{1}, \quad (\text{S1_5})$$

where $\mathbf{1}$ is a column vector of length $(n - k)$ with all entries being 1, and the i^{th} entry of $\mathbf{E}[\mathbf{t}]$ gives the expected number of steps when starting from state (mutant) i .

Similarly, the variance in the number of steps before being absorbed can be computed as

$$\text{Var}[\mathbf{t}] = (2\mathbf{N} - \mathbf{I}_t) \mathbf{E}[\mathbf{t}] - \mathbf{E}[\mathbf{t}] \odot \mathbf{E}[\mathbf{t}], \quad (\text{S1_6})$$

where \odot denotes the Hadamard product.

Finally, the probability of being absorbed in state j when starting from transient state i (i.e., reaching optimum j when the initial genotype is i), is given by the $(n - k) \times k$ -matrix

$$\mathbf{B} = \mathbf{N}\mathbf{R}. \quad (\text{S1_7})$$

Thus, these methods give an easy and computationally fast way of quantifying and predicting adaptive walks on fitness landscapes. Furthermore, the robustness of these results and the influence of particular mutants can be assessed by deleting the i^{th} column and row of \mathbf{P} – i.e., by essentially treating mutant i as unobserved –, recalculating the above statistics and comparing these to the statistics obtained from the full data set. Similarly, entire mutations (i.e., amino acids) can be left out to assess their relative effect on the fitness landscape and the generality of our analysis (and the statistics used).

Measuring epistasis

We applied different metrics for quantifying epistasis over the entire fitness landscape as well as for particular mutations and assessed their consistency in capturing the strength of gene×gene interactions. In particular, we follow the definition of epistasis by [52] (originally termed epistacy), as the deviation from additivity when combining two genetic effects which is measured by the difference in log-fitness between the effects of the double mutant and the single mutant relative to the wild-type fitness.

Correlation of fitness effects of mutations: γ

The first measure has recently been introduced by [14] and is defined as the single-step correlation of fitness effects for mutations between neighbouring genotypes. It quantifies how the selective effect of a focal mutation is altered when it occurs in a different genetic background averaged over all genotypes of the fitness landscape. Geometrically, γ measures the correlation between slopes (with respect to genotype-fitness hypercube) of the same mutation put into different genetic backgrounds. Thus, if the fitness effect of a mutation is independent of its genetic background (i.e., if there is no epistasis), the correlation in slopes will be perfect ($\gamma = 1$), whereas it will be zero if the fitness slopes of each genotype are independent of the fitnesses of other genotypes (as in the House-of-Cards model; 53). Depending on the scale γ can either be used to quantify the strength of gene \times gene interactions between specific mutations or as an overall measure for the entire fitness landscape. However, in its original form γ was defined for di-allelic data only and thus needs to be extended by considering pairs of specific alleles at different loci [14].

Let $\mathcal{A}_i = \{1_i, 2_i, \dots, m_i\}$ denote the set of different alleles present at locus i for all polymorphic loci $i \in \{1, 2, \dots, n\}$ such that $|\mathcal{A}_i| \geq 2$ for all loci i . Further, let \mathcal{G} be the set of all genotypes that can be formed by combining all alleles such that the total number of genotype is $|\mathcal{G}| = \prod_{i=1}^n m_i$.

Then the matrix of epistatic effects between loci i and j carrying alleles $A_i, B_i \in \mathcal{A}_i$ and $A_j, B_j \in \mathcal{A}_j$ is given by

$$\gamma_{(A_i, B_i) \rightarrow (A_j, B_j)} = \text{Cor}[s_{(A_j, B_j)}(g), s_{(A_j, B_j)}(g_{[(A_i, B_i)]})] = \frac{\sum_g s_j(g) s_j(g_{[i]})}{\sum_g (s_j(g))^2}. \quad (\text{S1}_8)$$

where $g := \{x \in \mathcal{G} | x_i = A_i \text{ or } x_i = B_i \text{ and } x_j = A_j \text{ or } x_j = B_j\} \subseteq \mathcal{G}$ such that the sum is only calculated over the subset of genotypes carrying one of the two focal alleles at each focal locus. Thus, $\gamma_{(A_i, B_i) \rightarrow (A_j, B_j)}$ is a quadratic matrix of dimension $\left(\sum_{i=1}^n \frac{|\mathcal{A}_i|(|\mathcal{A}_i|-1)}{2}\right)$. Note that in the case where $|\mathcal{A}_i| = 2$ for all loci i , we obtain equation 9 in [14].

Likewise, the epistatic effect of a mutation in locus i with alleles (A_i, B_i) on other loci (and pairs of alleles) can be calculated as

$$\gamma_{(A_i, B_i) \rightarrow} = \text{Cor}[s(g), s(g_{[(A_i, B_i)]})] = \frac{\sum_{j \neq i} \sum_{\mathbf{a}_j} \sum_g s_j(g) s_j(g_{[i]})}{\sum_{j \neq i} \sum_{\mathbf{a}_j} \sum_g (s_j(g))^2}, \quad (\text{S1}_9)$$

where the summation index $\mathbf{a}_j = \{(A_j, B_j) | A_j, B_j \in \mathcal{A}_j \text{ and } A_j \neq B_j\}$ is over the set of subsets of size two that can be constructed from all alleles found at locus j . Note that the third summation index g changes depending on \mathbf{a}_j .

An additional summation allows calculation of the epistatic effect of a mutation in locus i carrying allele (A_i) on other loci (and pairs of alleles) can be calculated as

$$\gamma_{A_i \rightarrow} = \text{Cor}[s(g), s(g_{[A_i]})] = \frac{\sum_{j \neq i} \sum_{\mathbf{f}_i} \sum_{\mathbf{a}_j} \sum_g s_j(g) s_j(g_{[i]})}{\sum_{j \neq i} \sum_{\mathbf{f}_i} \sum_{\mathbf{a}_j} \sum_g (s_j(g))^2}, \quad (\text{S1}_{10})$$

where $\mathfrak{f}_i = \{(A_i, B_i) \mid B_i \in \mathcal{A}_i \text{ and } A_i \neq B_i\}$ such that the sum is only calculated over the elements of the set of subsets of size two that can be constructed from all alleles found at locus i that contain allele A_i .

Then, summing over $\mathfrak{l}_i = \{(A_i, B_i) \mid A_i, B_i \in \mathcal{A}_i \text{ and } A_i \neq B_i\}$, i.e., the elements of the set of subsets of size two that can be constructed from all alleles found at locus i , gives the epistatic effect of a mutation in locus i

$$\gamma_{i \rightarrow} = \text{Cor}[s(g), s(g_{[i]})] = \frac{\sum_{j \neq i} \sum_{\mathfrak{l}_i} \sum_{\mathfrak{a}_j} \sum_g s_j(g) s_j(g_{[i]})}{\sum_{j \neq i} \sum_{\mathfrak{l}_i} \sum_{\mathfrak{a}_j} \sum_g (s_j(g))^2}. \quad (\text{S1_11})$$

Similarly, the epistatic effect of other mutations (again considering pairs of alleles first) on locus j with alleles (A_i, B_i) can be calculated as

$$\gamma_{\rightarrow(A_j, B_j)} = \text{Cor}[s_{(A_j, B_j)}(g), s_{(A_j, B_j)}(g_1)] = \frac{\sum_{i \neq j} \sum_{\mathfrak{a}_i} \sum_g s_j(g) s_j(g_{[i]})}{\sum_{i \neq j} \sum_{\mathfrak{a}_i} \sum_g (s_j(g))^2}, \quad (\text{S1_12})$$

the epistatic effect of other mutations on locus j carrying allele (A_j) is given by

$$\gamma_{\rightarrow A_j} = \text{Cor}[s_{(A_j)}(g), s_{(A_j)}(g_1)] = \frac{\sum_{i \neq j} \sum_{\mathfrak{f}_j} \sum_{\mathfrak{a}_i} \sum_g s_j(g) s_j(g_{[i]})}{\sum_{i \neq j} \sum_{\mathfrak{f}_j} \sum_{\mathfrak{a}_i} \sum_g (s_j(g))^2}, \quad (\text{S1_13})$$

and the epistatic effect of other mutations on locus j becomes

$$\gamma_{\rightarrow j} = \text{Cor}[s_j(g), s_j(g_1)] = \frac{\sum_{i \neq j} \sum_{\mathfrak{l}_j} \sum_{\mathfrak{a}_i} \sum_g s_j(g) s_j(g_{[i]})}{\sum_{i \neq j} \sum_{\mathfrak{l}_j} \sum_{\mathfrak{a}_i} \sum_g (s_j(g))^2}. \quad (\text{S1_14})$$

Finally, γ_d , that is the decay of correlation of fitness effects with Hamming distance d , (i.e., the cumulative epistatic effect of d mutations averaged over the entire fitness landscape) is calculated as

$$\gamma_d = \text{Cor}[s(g), s_j(g_d)] = \frac{\sum_g \sum_{g_d} \sum_{j \neq i_1, i_2, \dots, i_d} \sum_{\mathcal{A}_j \setminus \{A_j\}} s_j(g) s_j(g_{[i_1 i_2 \dots i_d]})}{\sum_g \sum_{\mathcal{A}_j \setminus \{A_j\}} (s_j(g))^2}, \quad (\text{S1_15})$$

where the last summation is over all different alleles present at locus j except the one carried by genotype g at locus j . Note that there is unfortunately no multi-allelic analog to equation (14) of [14]. Furthermore, as desired when there are only two alleles at a given locus, equations (S1_9 – S1_11) and equations (S1_12 – S1_14) collapse and give identical values, and reduce to their di-allelic counterparts (i.e., eq. 7-8 in 14).

Fraction of epistasis

The second statistic quantifies whether specific pairs of alleles between two loci interact epistatically, and if so whether they display magnitude epistasis (i.e., fitness effects are non-additive, but fitness increases with the number of mutations), sign epistasis (i.e., one of

the two mutations considered has an opposite effect in both backgrounds) or reciprocal sign epistasis (i.e., if both mutations show sign epistasis; [34, 35]; for an implementation see [36]).

Using equation (S1_1) the type of epistatic interaction between a mutations at locus i and j (with $i \neq j$) when introduced on some reference genotype g , $\mathbf{e}(g, i, j)$, can formally be given as

$$\mathbf{e}(g, i, j) = \begin{cases} \text{none} & \text{if } |s_i(g_{[j]}) - s_i(g)| = 0 \\ \text{magnitude} & \text{if } s_j(g)s_j(g_{[i]}) \geq 0 \text{ and } s_i(g)s_i(g_{[j]}) \geq 0 \\ \text{reciprocal sign} & \text{if } s_j(g)s_j(g_{[i]}) < 0 \text{ and } s_i(g)s_i(g_{[j]}) < 0 \\ \text{sign} & \text{else} \end{cases}. \quad (\text{S1_16})$$

Note that for numerical purposes we allowed for a very small deviation $\varepsilon = 10^{-6}$ in the case of no epistatic interaction. For ease of notation, we treat the elements in \mathcal{A}_i as fixed and ordered, and define $I(g_{[i]})$ as the index of the allele present at locus i in genotype g in \mathcal{A}_i such that the fraction of epistasis over all \mathcal{G} can be calculated as

$$\mathbf{e}(x) = c^{-1} \sum_g \sum_{k=I(g_{[i]})+1}^{m_i} \sum_{l=k+1}^{m_j} \mathbb{1}_x(\mathbf{e}(g, k, l)), \quad (\text{S1_17})$$

where $x \in \{\text{none, magnitude, sign, reciprocal sign}\}$, $\mathbb{1}_x(y)$ is the indicator function that is 1 if $x = y$ and 0 otherwise, and $c = \left(\sum_g \sum_{k=I(g_{[i]})+1}^{m_i} \sum_{l=k+1}^{m_j} 1 \right)$ is a normalization constant.

Supporting Information 2: Overview of different fitness landscape models introduced in the main text

In the following, we briefly introduce the various fitness landscape models introduced and discussed in the main text and provide references to selected relevant literature. For an excellent general overview, we refer the reader to a review by [10]

The additive model

In the additive model the log-fitness of each genotype is given by the sum over the individual per locus log-fitness effects. Thus, the fitness effect of a specific allele – drawn from a Normal distribution with mean μ_a and variance σ_a^2 [14] [see also 54] – is independent of its genetic background (i.e., it is constant across all genetic backgrounds), such that all mutations are non-interacting (i.e., there is no epistasis) and the resulting single-peaked fitness landscape is (maximally) smooth. In particular, the roughness-to-slope ratio is 0 and $\mathbb{E}[\gamma_d] = 1$ for the entire range of mutational distances d . Note that when fitnesses are not measured on log-scale, the fitness of a genotype is simply the product over the individual fitness effects, which is why this model is sometimes also referred to as the multiplicative model.

The House-of-cards model

On the other extreme, in the House-of-cards (HoC) model [53] the fitness of each genotype is an i.i.d. normally distributed random variable with zero mean and variance σ_{HoC}^2 resulting in an uncorrelated, maximally rugged fitness landscape that is characterized by multiple fitness maxima and minima [55, 56]. In particular, the fitness effect of an allele entirely depends on its genetic background such that there is complete interaction between all loci (i.e., full epistasis) which is also reflected in the roughness-to-slope ratio and $\mathbb{E}[\gamma_d]$ that become infinity and zero (for $d > 0$), respectively.

The Rough Mount Fuji model

Introduced by [57], the Rough-Mount-Fuji (RMF) model – named after the eponymous mountain in Japan –, which was initially formulated in the context of protein evolution [see also 54, for a simplified version], interpolates between the former two extremes. The fitness of a genotype is composed by an additive component (parametrized by μ_a and σ_a^2 ; see above) and a HoC component (parametrized by σ_{HoC}^2) such that the extent of epistatic interactions ranges between none (additive model) to complete (HoC model) depending on the relative sizes of these three parameters. In particular, when $\sigma_{\text{HoC}}^2 \ll \mu_a^2 + \sigma_a^2$ the RMF model becomes an additive model whereas for $\sigma_{\text{HoC}}^2 \gg \mu_a^2, \sigma_a^2$ it essentially behaves like a HoC model [14]. Accordingly, $0 \leq \mathbb{E}[\gamma_d] = \text{const} \leq 1$ for $d > 0$ (see Figure 4a) and the roughness-to-slope ratio ranges from zero to infinity.

The Kauffman NK model

Another frequently used fitness landscape model that, as the RMF model, also interpolates between the additive and the HoC model [55, 56] is the Kauffman NK model, where N diallelic loci interact with $K \in \{0, 1, \dots, L - 1\}$ randomly assigned other loci. In particular, for $K = 0$ the NK model collapses to an additive model whereas for $K = L - 1$ it approaches the HoC model. Although there are different ways how groups of interacting loci can be chosen [58], properties such as the mean number and height of local optima tend only to be weakly dependent on the exact choice being made. For the NK model, $\mathbb{E}[\gamma_d]$ is a non-negative monotonically decreasing function in d , and the roughness-to-slope ratio can again range from zero to infinity.

The eggbox model

Introduced by [14], the eggbox model is a maximally epistatic, anticorrelated fitness landscape model (i.e., all loci interact with each other up to interactions of order L), in which the fitness effect of an allele switches from the highest to lowest value (or vice versa) between genetic backgrounds one step apart. Accordingly, depending on whether two genotypes are separated by an odd or even Hamming distance, their absolute fitness difference is either twice the mean allelic fitness effect or zero. Thus, this model generates an extreme case of reciprocal sign epistasis in which each mutation is either deleterious or compensatory, multiple local optima exist, and γ_d accordingly oscillates between -1 and 1.

Supporting Information 3: Supporting Figures

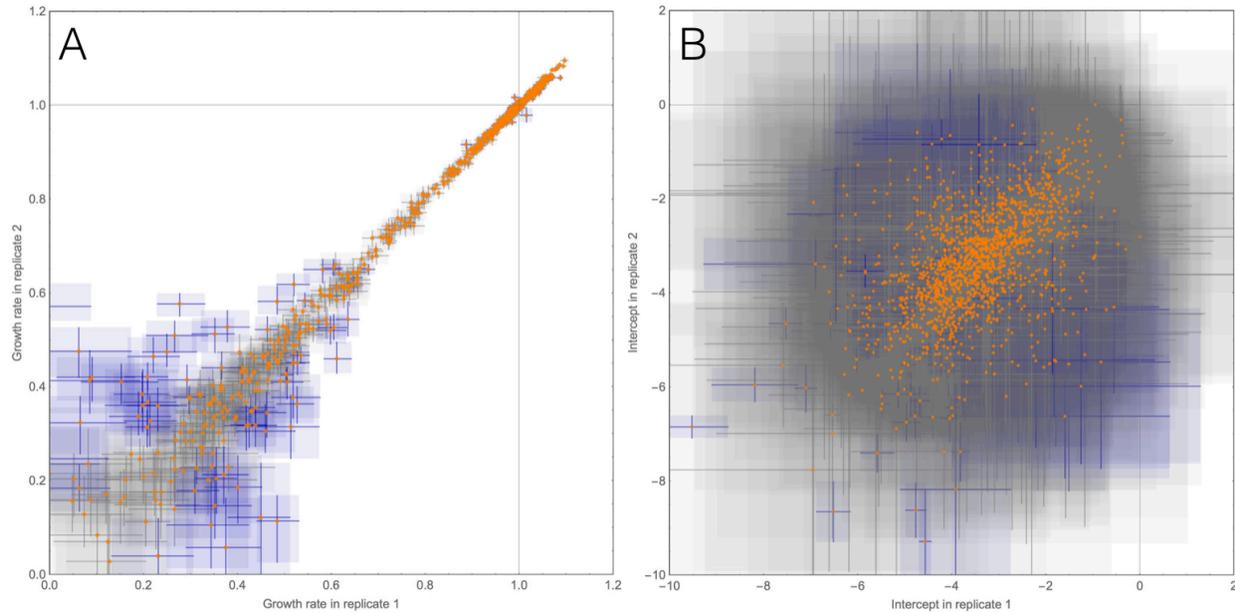


Figure S3_1 – (A) Scatterplot demonstrating the correlation of estimated growth rates ($R^2 = 0.944$). Orange dots indicate median growth rates, rectangles indicate the limits of the 95% credibility intervals (in blue if they do not overlap between replicates). (B) In contrast, estimated initial population sizes are much more variable between replicates, demonstrating that initial representation in the mutant library had no effect on the estimated growth rate.

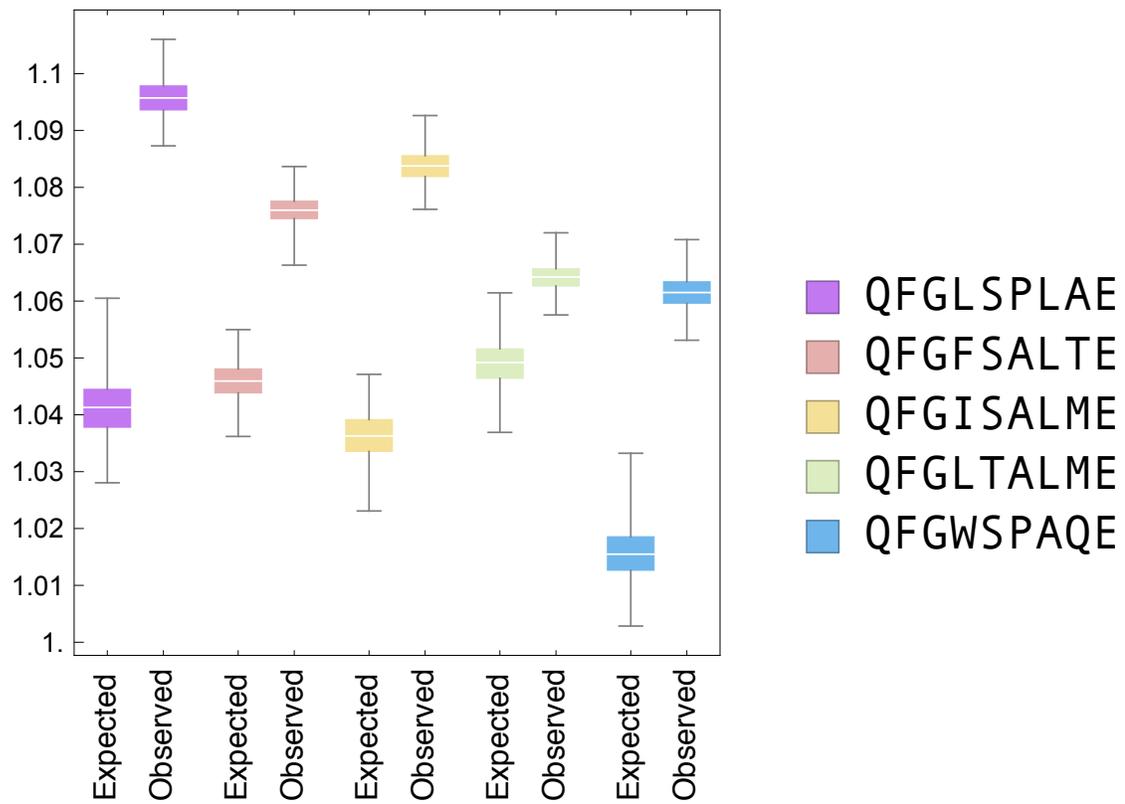


Figure S3_2 – Expected versus observed fitness for all 5 beneficial local optima of the fitness landscape, obtained from 1000 posterior samples.

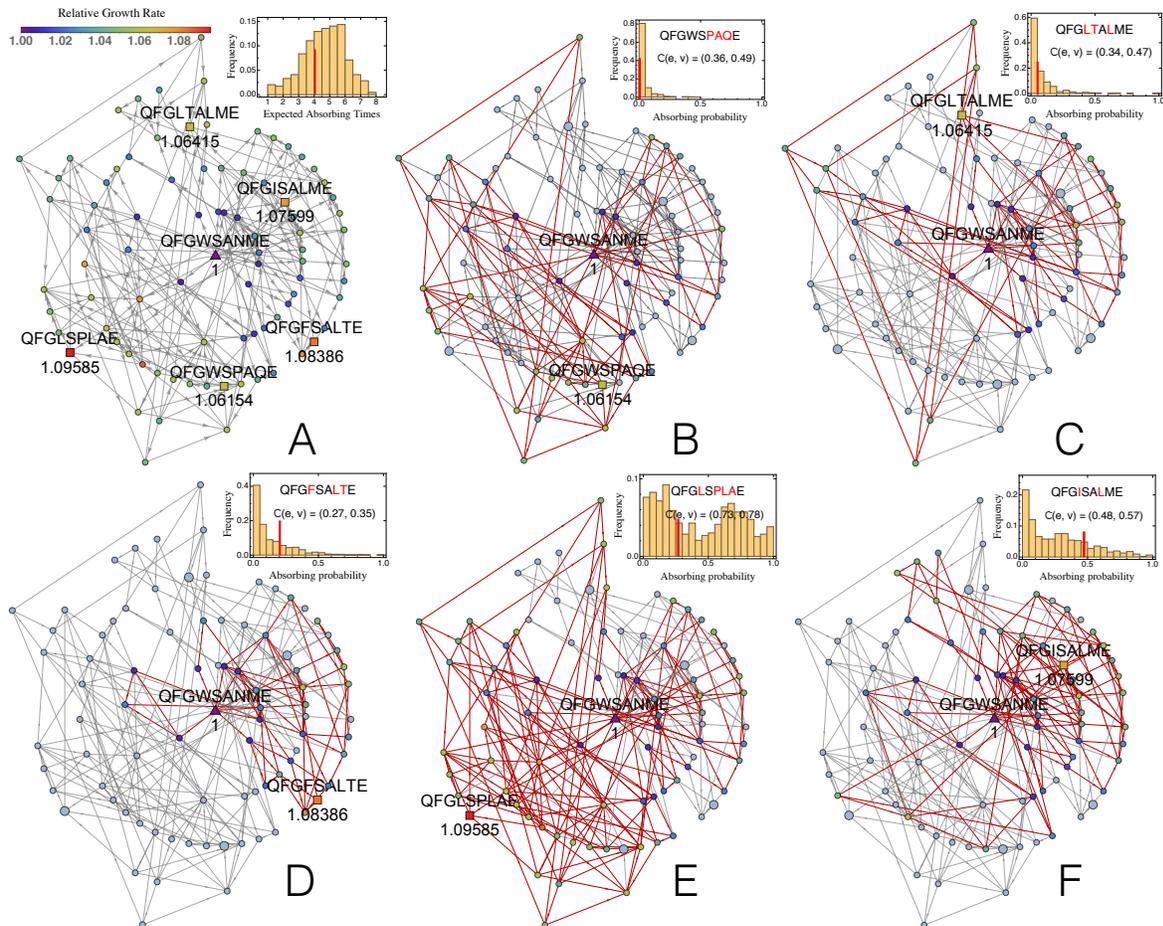


Figure S3_3 – Graphical illustration of the sub-landscape of all beneficial mutants with respect to the parental type. (A) Each vertex (circle) corresponds to a mutant that is beneficial with respect to the parental type (triangle), colored according to their fitness. Arrows connect mutants that differ by a single substitution, with their direction indicating an increase in fitness. The five local optima in this sub-landscape are indicated as squares. Inset: distribution of average lengths of adaptive walks starting from any type in the full landscape (i.e., absorbing times of the Markov chain). The red line indicates mean path lengths for adaptive walks from the parental genotype. (B)-(F) Paths leading towards the different optima – highlighted by their sequence name – when beginning an adaptive walk at the parental genotype. Inset (B) - (F): Distribution of absorbing probabilities, that is, the probability to reach a specific optimum starting from a given mutant computed for all mutants in the data set. The red line corresponds to the respective probability when starting from the parental sequence. The global optimum (panel E) is in general reached with a very high probability, but there are starting points from which it is poorly accessible. $C(e, v)$ quantifies the landscape coverage: although 73% of all edges and 78% of all vertices can be included in walks from the parental genotype to the global optimum, it is reached in only 26% of all possible walks. Conversely, 47% of walks will reach the local optimum in panel C, which is lower in fitness, but closer to the parental type in terms of its Hamming distance. This demonstrates that evolution on this landscape is not well predicted by fitness only, and that a population may become stuck in a sub-optimal fitness peak.

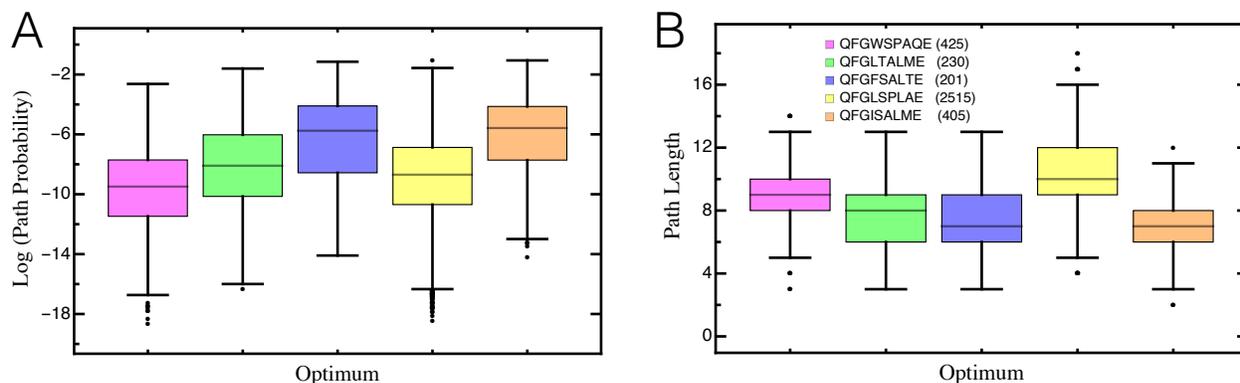


Figure S3_4 – (A) Path probability of reaching a specific local optimum when beginning an adaptive walk at the parental genotype. Results are presented on log scale. Boxes represent the interquartile range (i.e., the 50% C.I.), whiskers extend to the highest/lowest data point within the box ± 1.5 times the interquartile range, and black circles represent close outliers. (B) Path length (i.e., the number of substitutions) to reach a specific local optimum. Inset: Color coding for the different optima with the total number of different paths leading to the respective optimum in brackets. There is a total of 3776 different paths that can be taken from the parental genotype, 66% of which lead to the global optimum (yellow). Most of these, however, require multiple substitutions, such that the probability to reach the global optimum is generally low. In contrast, the local optimum close to the parental genotype (orange) can be reached with only a few substitutions, and thus, with relatively high probability.

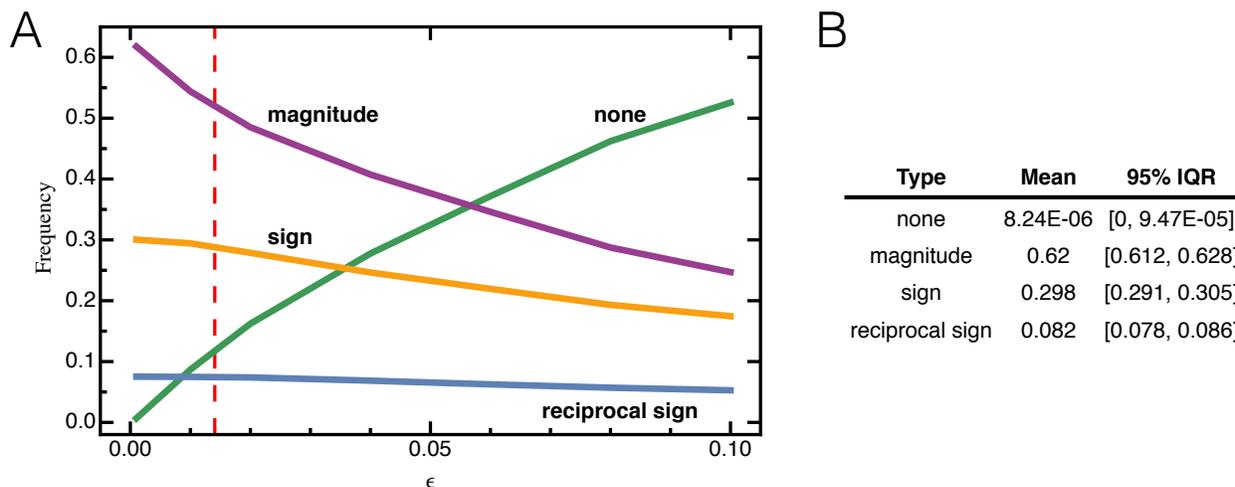


Figure S3_5 – (A) The fraction of epistasis (eq. S1_17) calculated for different values of ϵ , quantifying the error threshold beyond which interactions between different alleles are considered to be epistatic. Expectedly, as ϵ increases the number of non-epistatic interactions between alleles increases, which in the majority of cases were previously classified as magnitude epistatic interactions. Likewise, the fraction of sign and reciprocal sign epistasis decreases. The red dashed line shows the mean absolute difference between median growth rates obtained from the two independent replicates calculated over all mutants with median growth rate larger than the mean median growth rate estimated for the substitution F583* (i.e., when substituting a stop codon) between the two replicates. Using this threshold for ϵ , we find that approximately 10% of all interactions are additive, approximately 50% show magnitude epistasis, whereas the sign and reciprocal sign epistasis remains roughly constant at 30% and 8%, respectively. (B) The mean and the 95% interquartile range (IQR) of the fraction of epistasis calculated over 10,000 posterior samples obtained from the MCMC simulations using the standard threshold $\epsilon = 10^{-6}$. There is only little to no variation in the fraction of epistasis and the mean value is essentially identical to the proportions reported in the main text, indicating robustness of the inferred fraction of epistasis, and confirming that variation in posterior samples is typically small (i.e., growth rates are estimated with high precision).

Supporting Information 4: Addressing measurement uncertainty in fitness landscape statistics

Table S4_1 – Epistasis measure $\gamma_{(A_i, B_i) \rightarrow (A_j, B_j)}$ (eq. S1_8) between any two substitutions, averaged across the entire landscape calculated over 1,000 posterior samples. Shown are the 2.5%, 50%, 97.5% confidence intervals and the corresponding value calculated from the median growth rates. The majority of interactions are small to moderate.

$A_i B_i$	$A_j B_j$	2.5%C.I.	50%C.I.	97.5%C.I.	Main text
F583H	F583H	1	1	1	1
F583H	L585W	0.781186	0.7957	0.808314	0.798262
F583H	F585W	0.656044	0.676274	0.695468	0.680326
F583H	I585W	0.429508	0.457521	0.482397	0.459904
F583H	F585L	0.777136	0.794864	0.812339	0.7982
F583H	I585L	0.569344	0.591398	0.612607	0.59463
F583H	F585I	0.773181	0.797178	0.81682	0.802641
F583H	S586T	0.693191	0.708917	0.724002	0.713048
F583H	A587P	0.754603	0.766899	0.779039	0.771487
F583H	H588N	0.842544	0.854133	0.864508	0.856289
F583H	N588P	0.190576	0.229818	0.269909	0.232421
F583H	L588N	0.925346	0.933872	0.941742	0.936301
F583H	A588N	0.763268	0.778255	0.790182	0.779749
F583H	H588P	0.233406	0.269719	0.310512	0.272072
F583H	H588L	0.897587	0.906249	0.913168	0.908253
F583H	A588H	0.935011	0.94343	0.95087	0.945953
F583H	L588P	0.164165	0.203443	0.244933	0.206263
F583H	A588P	0.302983	0.339205	0.378535	0.341537
F583H	A588L	0.829686	0.839552	0.848894	0.841511
F583H	M589T	0.36478	0.388584	0.410715	0.390504
F583H	M589Q	0.767691	0.787224	0.805853	0.791224
F583H	A589M	0.475742	0.501599	0.524351	0.505354
F583H	Q589T	0.561673	0.582565	0.599189	0.584278
F583H	A589T	0.719254	0.743653	0.766629	0.748196
F583H	A589Q	0.660341	0.68148	0.704744	0.685819
L585W	F583H	0.454182	0.487632	0.518399	0.491896
L585W	L585W	1	1	1	1
L585W	F585W	1	1	1	1
L585W	I585W	1	1	1	1
L585W	F585L	1	1	1	1
L585W	I585L	1	1	1	1

L585W	F585I	1	1	1	1
L585W	S586T	0.473765	0.514234	0.547457	0.517321
L585W	A587P	0.518897	0.554512	0.583423	0.559365
L585W	H588N	0.726887	0.743854	0.760751	0.746688
L585W	N588P	-0.0963246	-0.0408707	0.0188961	-0.0403157
L585W	L588N	0.773517	0.797603	0.818312	0.799818
L585W	A588N	0.637654	0.658128	0.678935	0.660951
L585W	H588P	-0.00537098	0.054329	0.114804	0.0547264
L585W	H588L	0.790838	0.807674	0.82184	0.80912
L585W	A588H	0.942444	0.954765	0.964412	0.95773
L585W	L588P	-0.0629087	-3.80E-05	0.0654933	0.000262741
L585W	A588P	0.0306047	0.0885322	0.153985	0.0907307
L585W	A588L	0.712324	0.737825	0.762529	0.740075
L585W	M589T	-0.0422936	-0.0160635	0.0106293	-0.0159503
L585W	M589Q	0.182411	0.255928	0.320006	0.259131
L585W	A589M	-0.310251	-0.215613	-0.123525	-0.219263
L585W	Q589T	-0.0942194	-0.0693456	-0.0443903	-0.0696156
L585W	A589T	-0.407908	-0.361778	-0.305441	-0.363797
L585W	A589Q	0.275792	0.386399	0.479367	0.393986
F585W	F583H	0.575435	0.60054	0.624705	0.604693
F585W	L585W	1	1	1	1
F585W	F585W	1	1	1	1
F585W	I585W	1	1	1	1
F585W	F585L	1	1	1	1
F585W	I585L	1	1	1	1
F585W	F585I	1	1	1	1
F585W	S586T	0.563464	0.592818	0.617245	0.597256
F585W	A587P	0.63104	0.655021	0.675156	0.659723
F585W	H588N	0.849929	0.873172	0.890695	0.876316
F585W	N588P	0.0748116	0.124129	0.168615	0.126382
F585W	L588N	0.82937	0.853201	0.871674	0.855305
F585W	A588N	0.853412	0.86723	0.879036	0.87048
F585W	H588P	0.127938	0.175822	0.222216	0.178053
F585W	H588L	0.822823	0.842117	0.857598	0.844528
F585W	A588H	0.924808	0.941599	0.954542	0.944652
F585W	L588P	0.0147719	0.0596892	0.100846	0.0609615
F585W	A588P	0.125846	0.178084	0.222375	0.179708
F585W	A588L	0.770335	0.789736	0.806575	0.791355

F585W	M589T	0.420734	0.451691	0.48096	0.453511
F585W	M589Q	0.619419	0.664335	0.70375	0.667214
F585W	A589M	0.17335	0.259591	0.328658	0.262461
F585W	Q589T	0.618766	0.640466	0.659139	0.643393
F585W	A589T	0.72921	0.760218	0.788596	0.767587
F585W	A589Q	0.515032	0.570298	0.616933	0.576526
I585W	F583H	0.565929	0.585825	0.606133	0.588829
I585W	L585W	1	1	1	1
I585W	F585W	1	1	1	1
I585W	I585W	1	1	1	1
I585W	F585L	1	1	1	1
I585W	I585L	1	1	1	1
I585W	F585I	1	1	1	1
I585W	S586T	0.718259	0.74288	0.762948	0.746601
I585W	A587P	0.642102	0.661945	0.680028	0.66543
I585W	H588N	0.734221	0.749666	0.764296	0.751656
I585W	N588P	-0.0953089	-0.0524529	-0.0117559	-0.0529523
I585W	L588N	0.841038	0.854263	0.865488	0.856459
I585W	A588N	0.636411	0.655251	0.674846	0.657185
I585W	H588P	0.0813317	0.121835	0.163185	0.123579
I585W	H588L	0.907622	0.919595	0.928475	0.921124
I585W	A588H	0.943274	0.953209	0.960734	0.955383
I585W	L588P	0.007348	0.0529695	0.0958002	0.0534012
I585W	A588P	0.130452	0.173724	0.215956	0.17604
I585W	A588L	0.877774	0.88869	0.898551	0.890557
I585W	M589T	0.340237	0.366168	0.390183	0.367498
I585W	M589Q	0.67767	0.711942	0.742175	0.715303
I585W	A589M	0.196538	0.241079	0.287418	0.244517
I585W	Q589T	0.606682	0.621816	0.634968	0.623444
I585W	A589T	0.773898	0.799966	0.8209	0.804039
I585W	A589Q	0.509127	0.546985	0.583251	0.552331
F585L	F583H	0.498713	0.537351	0.578225	0.542494
F585L	L585W	1	1	1	1
F585L	F585W	1	1	1	1
F585L	I585W	1	1	1	1
F585L	F585L	1	1	1	1
F585L	I585L	1	1	1	1
F585L	F585I	1	1	1	1

F585L	S586T	0.508648	0.544855	0.577364	0.550048
F585L	A587P	0.748831	0.771728	0.792241	0.78029
F585L	H588N	0.783446	0.81465	0.838701	0.818782
F585L	N588P	0.174674	0.231065	0.283916	0.234195
F585L	L588N	0.842643	0.859868	0.873231	0.864198
F585L	A588N	0.758322	0.781194	0.802305	0.786606
F585L	H588P	0.234263	0.287353	0.344957	0.290631
F585L	H588L	0.867885	0.893601	0.912608	0.898159
F585L	A588H	0.902161	0.926496	0.944877	0.932376
F585L	L588P	0.119219	0.174994	0.225747	0.177166
F585L	A588P	0.239477	0.293336	0.340646	0.297365
F585L	A588L	0.867397	0.883835	0.897475	0.888359
F585L	M589T	0.222748	0.324079	0.417104	0.331449
F585L	M589Q	0.731643	0.775209	0.807377	0.77954
F585L	A589M	0.508923	0.538641	0.566798	0.544187
F585L	Q589T	0.014926	0.0878362	0.16612	0.0910583
F585L	A589T	0.136648	0.183999	0.228006	0.186528
F585L	A589Q	0.617723	0.654863	0.683122	0.658132
I585L	F583H	0.440601	0.466005	0.493341	0.469596
I585L	L585W	1	1	1	1
I585L	F585W	1	1	1	1
I585L	I585W	1	1	1	1
I585L	F585L	1	1	1	1
I585L	I585L	1	1	1	1
I585L	F585I	1	1	1	1
I585L	S586T	0.731721	0.751609	0.772371	0.757651
I585L	A587P	0.722903	0.740117	0.755216	0.745156
I585L	H588N	0.703994	0.725282	0.746375	0.728149
I585L	N588P	-0.0147815	0.0390647	0.0912781	0.0384516
I585L	L588N	0.780189	0.798246	0.813711	0.801102
I585L	A588N	0.601557	0.627081	0.650434	0.629603
I585L	H588P	0.131625	0.184265	0.236204	0.185243
I585L	H588L	0.897078	0.912996	0.923876	0.91505
I585L	A588H	0.916351	0.932322	0.946543	0.936478
I585L	L588P	0.0589946	0.118835	0.171694	0.118318
I585L	A588P	0.192648	0.24165	0.290474	0.244484
I585L	A588L	0.832471	0.851941	0.868587	0.854602
I585L	M589T	0.326097	0.388791	0.443753	0.393248

I585L	M589Q	0.713169	0.744019	0.772596	0.748478
I585L	A589M	0.414194	0.439381	0.461654	0.441654
I585L	Q589T	0.378748	0.425656	0.466665	0.427833
I585L	A589T	0.628706	0.659398	0.688126	0.665311
I585L	A589Q	0.580186	0.605879	0.630692	0.609325
F585I	F583H	0.184445	0.249108	0.306907	0.255675
F585I	L585W	1	1	1	1
F585I	F585W	1	1	1	1
F585I	I585W	1	1	1	1
F585I	F585L	1	1	1	1
F585I	I585L	1	1	1	1
F585I	F585I	1	1	1	1
F585I	S586T	0.375036	0.441937	0.506261	0.45574
F585I	A587P	0.237367	0.294618	0.348139	0.302799
F585I	H588N	0.0905469	0.163255	0.226801	0.162977
F585I	N588P	-0.207737	-0.119257	-0.0320937	-0.124733
F585I	L588N	0.296632	0.3553	0.409815	0.361256
F585I	A588N	0.0843689	0.148095	0.212808	0.150577
F585I	H588P	-0.0721975	0.0221204	0.123655	0.0217359
F585I	H588L	0.682545	0.739801	0.786965	0.748623
F585I	A588H	0.746857	0.80704	0.857383	0.820367
F585I	L588P	-0.244073	-0.134591	-0.0286072	-0.141615
F585I	A588P	-0.0773802	0.0199233	0.130122	0.0208728
F585I	A588L	0.679159	0.739456	0.788322	0.746859
F585I	M589T	-0.0223809	0.0369155	0.0939654	0.0384758
F585I	M589Q	0.181643	0.304356	0.438138	0.316988
F585I	A589M	0.115825	0.207888	0.302134	0.219138
F585I	Q589T	0.258418	0.331378	0.392724	0.337253
F585I	A589T	0.408696	0.498139	0.575123	0.512365
F585I	A589Q	0.192987	0.305332	0.413546	0.314907
S586T	F583H	0.558533	0.578932	0.600615	0.583796
S586T	L585W	0.668146	0.692647	0.715473	0.695892
S586T	F585W	0.60444	0.629865	0.653122	0.634002
S586T	I585W	0.405514	0.450451	0.493272	0.454801
S586T	F585L	0.715555	0.737153	0.757711	0.74183
S586T	I585L	0.626532	0.653608	0.682645	0.661327
S586T	F585I	0.76627	0.796508	0.823164	0.80464
S586T	S586T	1	1	1	1

S586T	A587P	0.756743	0.777474	0.79646	0.784579
S586T	H588N	0.757426	0.77784	0.794093	0.781394
S586T	N588P	0.117798	0.163729	0.20692	0.166125
S586T	L588N	0.803608	0.814496	0.826797	0.81762
S586T	A588N	0.669584	0.692896	0.714201	0.695518
S586T	H588P	0.183005	0.226778	0.271477	0.229452
S586T	H588L	0.905321	0.919734	0.931172	0.922732
S586T	A588H	0.895396	0.912135	0.925907	0.916249
S586T	L588P	0.156165	0.200199	0.244447	0.202201
S586T	A588P	0.210221	0.252071	0.299017	0.25654
S586T	A588L	0.848712	0.864269	0.877965	0.866903
S586T	M589T	0.320181	0.34948	0.376496	0.350684
S586T	M589Q	0.444158	0.500164	0.546745	0.503497
S586T	A589M	0.501336	0.528305	0.555114	0.53348
S586T	Q589T	0.283318	0.304245	0.325068	0.306711
S586T	A589T	0.657295	0.686278	0.711797	0.690561
S586T	A589Q	0.330932	0.363657	0.390418	0.366934
A587P	F583H	0.444399	0.471354	0.495931	0.477261
A587P	L585W	0.458245	0.500674	0.532394	0.504322
A587P	F585W	0.40003	0.441218	0.477764	0.445334
A587P	I585W	0.132874	0.177005	0.216795	0.178225
A587P	F585L	0.674785	0.70362	0.727819	0.714168
A587P	I585L	0.472926	0.500006	0.52672	0.507174
A587P	F585I	0.591916	0.626418	0.658595	0.635448
A587P	S586T	0.619421	0.650491	0.681307	0.660246
A587P	A587P	1	1	1	1
A587P	H588N	0.638158	0.671336	0.701392	0.673741
A587P	N588P	0.153513	0.207187	0.252698	0.209571
A587P	L588N	0.766936	0.785239	0.802774	0.78839
A587P	A588N	0.62863	0.661831	0.688638	0.663819
A587P	H588P	0.106974	0.152807	0.191954	0.154351
A587P	H588L	0.711105	0.747364	0.778275	0.752304
A587P	A588H	0.830963	0.862226	0.887217	0.870824
A587P	L588P	-0.0440905	0.00351999	0.0467686	0.00313926
A587P	A588P	0.0369484	0.0817756	0.127242	0.0829486
A587P	A588L	0.679179	0.709999	0.735459	0.715062
A587P	M589T	0.214566	0.254008	0.290491	0.25677
A587P	M589Q	0.558199	0.594929	0.625845	0.600397

A587P	A589M	0.269509	0.317502	0.357892	0.322202
A587P	Q589T	0.525878	0.554338	0.580558	0.560294
A587P	A589T	0.652057	0.692982	0.728145	0.703486
A587P	A589Q	0.584218	0.623677	0.65867	0.632985
H588N	F583H	0.364167	0.40803	0.447776	0.411761
H588N	L585W	0.305853	0.347263	0.385203	0.350732
H588N	F585W	0.388132	0.45727	0.513089	0.464171
H588N	I585W	0.119186	0.155796	0.188542	0.157692
H588N	F585L	0.561127	0.615694	0.661239	0.622816
H588N	I585L	0.41906	0.450719	0.480376	0.454573
H588N	F585I	0.421243	0.477581	0.525674	0.481796
H588N	S586T	0.459634	0.498636	0.533117	0.504342
H588N	A587P	0.326995	0.386713	0.441572	0.388548
H588N	H588N	1	1	1	1
H588N	N588P	1	1	1	1
H588N	L588N	1	1	1	1
H588N	A588N	1	1	1	1
H588N	H588P	1	1	1	1
H588N	H588L	1	1	1	1
H588N	A588H	1	1	1	1
H588N	L588P	1	1	1	1
H588N	A588P	1	1	1	1
H588N	A588L	1	1	1	1
H588N	M589T	0.1649	0.214481	0.262116	0.214107
H588N	M589Q	0.646526	0.68125	0.711678	0.685033
H588N	A589M	0.260622	0.36808	0.459375	0.370694
H588N	Q589T	0.355734	0.402642	0.446682	0.404114
H588N	A589T	0.541341	0.613113	0.66742	0.619917
H588N	A589Q	0.372683	0.471628	0.558053	0.475109
N588P	F583H	0.796886	0.807261	0.817995	0.809733
N588P	L585W	0.864462	0.874946	0.88532	0.876739
N588P	F585W	0.780449	0.798435	0.811425	0.800614
N588P	I585W	0.658127	0.676268	0.692689	0.678165
N588P	F585L	0.87595	0.885417	0.894828	0.88751
N588P	I585L	0.800502	0.812417	0.823228	0.814426
N588P	F585I	0.921288	0.931104	0.940546	0.934299
N588P	S586T	0.8529	0.862946	0.870969	0.864982
N588P	A587P	0.852285	0.862262	0.870824	0.86468

N588P	H588N	1	1	1	1
N588P	N588P	1	1	1	1
N588P	L588N	1	1	1	1
N588P	A588N	1	1	1	1
N588P	H588P	1	1	1	1
N588P	H588L	1	1	1	1
N588P	A588H	1	1	1	1
N588P	L588P	1	1	1	1
N588P	A588P	1	1	1	1
N588P	A588L	1	1	1	1
N588P	M589T	0.558056	0.58033	0.603979	0.582521
N588P	M589Q	0.88279	0.893628	0.903445	0.895108
N588P	A589M	0.733958	0.755138	0.775567	0.757921
N588P	Q589T	0.746034	0.762406	0.78065	0.76479
N588P	A589T	0.83446	0.852405	0.8691	0.855846
N588P	A589Q	0.857413	0.871084	0.882708	0.873541
L588N	F583H	0.572777	0.623287	0.667224	0.632426
L588N	L585W	0.496956	0.543582	0.586391	0.546829
L588N	F585W	0.55716	0.612614	0.658266	0.615578
L588N	I585W	0.169691	0.231451	0.289624	0.235697
L588N	F585L	0.585495	0.624308	0.657367	0.634131
L588N	I585L	-0.128987	-0.0709644	-0.00350338	-0.0698335
L588N	F585I	0.0869328	0.149603	0.210072	0.15148
L588N	S586T	0.163505	0.207983	0.252605	0.213155
L588N	A587P	0.213638	0.269562	0.323857	0.273392
L588N	H588N	1	1	1	1
L588N	N588P	1	1	1	1
L588N	L588N	1	1	1	1
L588N	A588N	1	1	1	1
L588N	H588P	1	1	1	1
L588N	H588L	1	1	1	1
L588N	A588H	1	1	1	1
L588N	L588P	1	1	1	1
L588N	A588P	1	1	1	1
L588N	A588L	1	1	1	1
L588N	M589T	0.473429	0.531185	0.578612	0.536087
L588N	M589Q	0.634864	0.672167	0.706915	0.677271
L588N	A589M	0.420305	0.47321	0.518072	0.479182

L588N	Q589T	0.317629	0.374117	0.419175	0.378957
L588N	A589T	0.640882	0.684853	0.717536	0.694917
L588N	A589Q	0.266508	0.310993	0.350187	0.314844
A588N	F583H	0.276645	0.314086	0.349309	0.315059
A588N	L585W	0.420779	0.452762	0.486716	0.45531
A588N	F585W	0.529207	0.577347	0.613812	0.582731
A588N	I585W	0.0747905	0.104877	0.13589	0.105309
A588N	F585L	0.652158	0.687974	0.720333	0.694435
A588N	I585L	0.41337	0.443434	0.470715	0.445503
A588N	F585I	0.569462	0.60663	0.637953	0.611501
A588N	S586T	0.445508	0.478259	0.509802	0.481663
A588N	A587P	0.50433	0.545568	0.584122	0.548711
A588N	H588N	1	1	1	1
A588N	N588P	1	1	1	1
A588N	L588N	1	1	1	1
A588N	A588N	1	1	1	1
A588N	H588P	1	1	1	1
A588N	H588L	1	1	1	1
A588N	A588H	1	1	1	1
A588N	L588P	1	1	1	1
A588N	A588P	1	1	1	1
A588N	A588L	1	1	1	1
A588N	M589T	0.0758398	0.140718	0.197795	0.138998
A588N	M589Q	0.68905	0.718399	0.744794	0.721058
A588N	A589M	0.242384	0.309005	0.375651	0.310863
A588N	Q589T	0.242963	0.302766	0.35675	0.302634
A588N	A589T	0.690495	0.729725	0.759972	0.737069
A588N	A589Q	0.450311	0.51089	0.562515	0.514914
H588P	F583H	0.722663	0.736607	0.751248	0.739238
H588P	L585W	0.833589	0.846533	0.857924	0.848363
H588P	F585W	0.730656	0.74906	0.764905	0.751802
H588P	I585W	0.503165	0.527783	0.551007	0.52969
H588P	F585L	0.846354	0.85907	0.871503	0.861465
H588P	I585L	0.696712	0.716426	0.735834	0.719191
H588P	F585I	0.873775	0.889833	0.904735	0.89347
H588P	S586T	0.79044	0.80333	0.815052	0.806012
H588P	A587P	0.852984	0.861531	0.868901	0.864671
H588P	H588N	1	1	1	1

H588P	N588P	1	1	1	1
H588P	L588N	1	1	1	1
H588P	A588N	1	1	1	1
H588P	H588P	1	1	1	1
H588P	H588L	1	1	1	1
H588P	A588H	1	1	1	1
H588P	L588P	1	1	1	1
H588P	A588P	1	1	1	1
H588P	A588L	1	1	1	1
H588P	M589T	0.428294	0.457786	0.488056	0.460445
H588P	M589Q	0.859507	0.872401	0.88332	0.874244
H588P	A589M	0.67024	0.696139	0.718054	0.699549
H588P	Q589T	0.675904	0.697057	0.718344	0.699143
H588P	A589T	0.735399	0.760289	0.783452	0.763774
H588P	A589Q	0.824131	0.840084	0.855654	0.843174
H588L	F583H	0.537322	0.570348	0.59877	0.575948
H588L	L585W	0.483839	0.522418	0.557248	0.52512
H588L	F585W	0.505674	0.548758	0.58806	0.552972
H588L	I585W	0.322656	0.387631	0.440227	0.391864
H588L	F585L	0.798026	0.837371	0.865515	0.843904
H588L	I585L	0.573102	0.631645	0.676372	0.637841
H588L	F585I	0.637739	0.701911	0.754867	0.710492
H588L	S586T	0.659783	0.709983	0.751646	0.718148
H588L	A587P	0.573428	0.628092	0.673632	0.633629
H588L	H588N	1	1	1	1
H588L	N588P	1	1	1	1
H588L	L588N	1	1	1	1
H588L	A588N	1	1	1	1
H588L	H588P	1	1	1	1
H588L	H588L	1	1	1	1
H588L	A588H	1	1	1	1
H588L	L588P	1	1	1	1
H588L	A588P	1	1	1	1
H588L	A588L	1	1	1	1
H588L	M589T	0.355372	0.402987	0.450246	0.406109
H588L	M589Q	0.61995	0.653075	0.682368	0.656049
H588L	A589M	0.30928	0.422855	0.529679	0.42886
H588L	Q589T	0.57079	0.608733	0.639655	0.612123

H588L	A589T	0.490726	0.551847	0.605044	0.561098
H588L	A589Q	0.599976	0.690501	0.763127	0.699435
A588H	F583H	-0.121131	-0.0311957	0.0587075	-0.0342472
A588H	L585W	0.461349	0.554032	0.637626	0.571732
A588H	F585W	0.125621	0.262657	0.385727	0.271563
A588H	I585W	-0.228101	-0.123826	-0.0220694	-0.132762
A588H	F585L	0.421178	0.540025	0.645104	0.563555
A588H	I585L	-0.0200111	0.113126	0.24589	0.121353
A588H	F585I	0.157248	0.327582	0.490841	0.349158
A588H	S586T	-0.098815	0.0239002	0.140459	0.0252088
A588H	A587P	0.256828	0.375819	0.481963	0.39213
A588H	H588N	1	1	1	1
A588H	N588P	1	1	1	1
A588H	L588N	1	1	1	1
A588H	A588N	1	1	1	1
A588H	H588P	1	1	1	1
A588H	H588L	1	1	1	1
A588H	A588H	1	1	1	1
A588H	L588P	1	1	1	1
A588H	A588P	1	1	1	1
A588H	A588L	1	1	1	1
A588H	M589T	-0.287865	-0.152514	-0.00945207	-0.164418
A588H	M589Q	0.524019	0.600639	0.667946	0.61576
A588H	A589M	-0.0516285	0.0519124	0.14294	0.0522902
A588H	Q589T	-0.455615	-0.328814	-0.172035	-0.346725
A588H	A589T	-0.162622	0.00386973	0.176045	0.00539306
A588H	A589Q	0.283386	0.397965	0.50059	0.415179
L588P	F583H	0.815504	0.824543	0.833441	0.826696
L588P	L585W	0.879053	0.887793	0.896158	0.889284
L588P	F585W	0.809015	0.822135	0.832573	0.824236
L588P	I585W	0.629276	0.648119	0.666832	0.650521
L588P	F585L	0.885131	0.894142	0.902543	0.89599
L588P	I585L	0.762746	0.77789	0.791381	0.779594
L588P	F585I	0.897556	0.9096	0.920582	0.912231
L588P	S586T	0.832312	0.841767	0.851157	0.843883
L588P	A587P	0.869341	0.876453	0.883001	0.878686
L588P	H588N	1	1	1	1
L588P	N588P	1	1	1	1

L588P	L588N	1	1	1	1
L588P	A588N	1	1	1	1
L588P	H588P	1	1	1	1
L588P	H588L	1	1	1	1
L588P	A588H	1	1	1	1
L588P	L588P	1	1	1	1
L588P	A588P	1	1	1	1
L588P	A588L	1	1	1	1
L588P	M589T	0.615462	0.635788	0.655433	0.637349
L588P	M589Q	0.891889	0.901547	0.910663	0.903
L588P	A589M	0.753209	0.774391	0.792867	0.776988
L588P	Q589T	0.778945	0.79193	0.805796	0.793852
L588P	A589T	0.844111	0.859023	0.872826	0.862033
L588P	A589Q	0.865193	0.877239	0.888757	0.879972
A588P	F583H	0.708604	0.723912	0.740762	0.726943
A588P	L585W	0.821464	0.83457	0.847312	0.836936
A588P	F585W	0.71429	0.735461	0.753399	0.738473
A588P	I585W	0.47634	0.504271	0.53045	0.505953
A588P	F585L	0.833113	0.84668	0.857802	0.849264
A588P	I585L	0.667854	0.689099	0.709141	0.692123
A588P	F585I	0.86272	0.879768	0.896084	0.884655
A588P	S586T	0.771306	0.784702	0.798082	0.788293
A588P	A587P	0.825476	0.835171	0.844167	0.838456
A588P	H588N	1	1	1	1
A588P	N588P	1	1	1	1
A588P	L588N	1	1	1	1
A588P	A588N	1	1	1	1
A588P	H588P	1	1	1	1
A588P	H588L	1	1	1	1
A588P	A588H	1	1	1	1
A588P	L588P	1	1	1	1
A588P	A588P	1	1	1	1
A588P	A588L	1	1	1	1
A588P	M589T	0.519883	0.547342	0.574208	0.550427
A588P	M589Q	0.839046	0.854109	0.867391	0.856051
A588P	A589M	0.597928	0.627067	0.653776	0.630781
A588P	Q589T	0.748195	0.76731	0.784661	0.770048
A588P	A589T	0.765165	0.789067	0.810309	0.793702

A588P	A589Q	0.778221	0.796749	0.816004	0.80059
A588L	F583H	0.424192	0.454056	0.481585	0.457444
A588L	L585W	0.523416	0.56267	0.598961	0.564757
A588L	F585W	0.463087	0.504819	0.539237	0.507222
A588L	I585W	0.335864	0.378697	0.42374	0.382689
A588L	F585L	0.860671	0.878812	0.894559	0.883591
A588L	I585L	0.546684	0.597053	0.641602	0.602019
A588L	F585I	0.737578	0.789117	0.827323	0.795144
A588L	S586T	0.608846	0.646751	0.682133	0.65238
A588L	A587P	0.664308	0.697305	0.726469	0.70294
A588L	H588N	1	1	1	1
A588L	N588P	1	1	1	1
A588L	L588N	1	1	1	1
A588L	A588N	1	1	1	1
A588L	H588P	1	1	1	1
A588L	H588L	1	1	1	1
A588L	A588H	1	1	1	1
A588L	L588P	1	1	1	1
A588L	A588P	1	1	1	1
A588L	A588L	1	1	1	1
A588L	M589T	0.228406	0.289766	0.344052	0.28894
A588L	M589Q	0.726248	0.753999	0.778689	0.756972
A588L	A589M	0.407792	0.481079	0.544559	0.485841
A588L	Q589T	0.424406	0.477608	0.525877	0.479107
A588L	A589T	0.646349	0.697867	0.73463	0.704303
A588L	A589Q	0.648952	0.706927	0.755811	0.712652
M589T	F583H	0.650735	0.664927	0.677971	0.666804
M589T	L585W	0.54703	0.56318	0.577438	0.564
M589T	F585W	0.480907	0.51288	0.539057	0.515301
M589T	I585W	0.287422	0.316458	0.341994	0.317671
M589T	F585L	0.911228	0.922846	0.933026	0.92536
M589T	I585L	0.853407	0.868474	0.880963	0.870945
M589T	F585I	0.826379	0.840009	0.851789	0.843199
M589T	S586T	0.671977	0.688704	0.70288	0.690386
M589T	A587P	0.796758	0.808119	0.819254	0.810731
M589T	H588N	0.863383	0.874391	0.883686	0.875236
M589T	N588P	-0.0778001	-0.0268903	0.0207394	-0.026676
M589T	L588N	0.949011	0.95541	0.960753	0.9563

M589T	A588N	0.826234	0.840939	0.853505	0.841498
M589T	H588P	0.0750677	0.119964	0.165821	0.122608
M589T	H588L	0.905195	0.914454	0.921855	0.91537
M589T	A588H	0.970463	0.977123	0.982399	0.978197
M589T	L588P	-0.0777429	-0.0284708	0.0193453	-0.0279023
M589T	A588P	0.180031	0.223405	0.266025	0.225898
M589T	A588L	0.867986	0.881186	0.892501	0.881907
M589T	M589T	1	1	1	1
M589T	M589Q	1	1	1	1
M589T	A589M	1	1	1	1
M589T	Q589T	1	1	1	1
M589T	A589T	1	1	1	1
M589T	A589Q	1	1	1	1
M589Q	F583H	0.276666	0.331453	0.388945	0.338601
M589Q	L585W	0.366534	0.423993	0.47329	0.427614
M589Q	F585W	0.230867	0.3144	0.389001	0.318897
M589Q	I585W	0.0347747	0.11596	0.200697	0.116472
M589Q	F585L	0.423183	0.502226	0.568661	0.509794
M589Q	I585L	0.0673483	0.13933	0.212408	0.142196
M589Q	F585I	0.502054	0.585854	0.669046	0.596272
M589Q	S586T	0.29428	0.366017	0.426458	0.368788
M589Q	A587P	0.355792	0.404533	0.446143	0.410554
M589Q	H588N	0.539006	0.582224	0.623371	0.585245
M589Q	N588P	-0.214526	-0.168297	-0.121626	-0.169783
M589Q	L588N	0.757063	0.787239	0.817217	0.791389
M589Q	A588N	0.423492	0.479945	0.525762	0.481975
M589Q	H588P	-0.140508	-0.0813242	-0.0190897	-0.0811569
M589Q	H588L	0.62041	0.657049	0.686359	0.659036
M589Q	A588H	0.888364	0.910151	0.926381	0.91563
M589Q	L588P	-0.159805	-0.107828	-0.0562832	-0.108894
M589Q	A588P	-0.161762	-0.0932533	-0.0222508	-0.0925286
M589Q	A588L	0.610999	0.650268	0.68451	0.65374
M589Q	M589T	1	1	1	1
M589Q	M589Q	1	1	1	1
M589Q	A589M	1	1	1	1
M589Q	Q589T	1	1	1	1
M589Q	A589T	1	1	1	1
M589Q	A589Q	1	1	1	1

A589M	F583H	0.620828	0.639578	0.657849	0.643421
A589M	L585W	0.777034	0.806384	0.830731	0.809089
A589M	F585W	0.677662	0.716196	0.744622	0.719383
A589M	I585W	0.51177	0.544357	0.573395	0.546935
A589M	F585L	0.682592	0.702334	0.72173	0.707208
A589M	I585L	0.483193	0.50437	0.524455	0.507441
A589M	F585I	0.87299	0.890156	0.905071	0.895888
A589M	S586T	0.694659	0.712919	0.729043	0.717051
A589M	A587P	0.774798	0.789417	0.802622	0.79411
A589M	H588N	0.893239	0.910023	0.923302	0.911445
A589M	N588P	0.197911	0.264884	0.323634	0.26732
A589M	L588N	0.910349	0.918635	0.92615	0.920393
A589M	A588N	0.814431	0.834317	0.852222	0.835999
A589M	H588P	0.196481	0.256983	0.310352	0.259128
A589M	H588L	0.920937	0.935692	0.947719	0.93737
A589M	A588H	0.930429	0.942942	0.952734	0.945349
A589M	L588P	0.242241	0.306007	0.363388	0.309128
A589M	A588P	0.20626	0.264019	0.315201	0.267348
A589M	A588L	0.889554	0.904615	0.916889	0.906003
A589M	M589T	1	1	1	1
A589M	M589Q	1	1	1	1
A589M	A589M	1	1	1	1
A589M	Q589T	1	1	1	1
A589M	A589T	1	1	1	1
A589M	A589Q	1	1	1	1
Q589T	F583H	0.597285	0.615061	0.63193	0.617503
Q589T	L585W	0.438987	0.453116	0.467275	0.454343
Q589T	F585W	0.529435	0.553269	0.574216	0.556833
Q589T	I585W	0.272024	0.295169	0.31707	0.296767
Q589T	F585L	0.790603	0.810881	0.830013	0.813257
Q589T	I585L	0.719291	0.739849	0.760291	0.742441
Q589T	F585I	0.777364	0.800068	0.821324	0.804513
Q589T	S586T	0.624019	0.636605	0.649058	0.639424
Q589T	A587P	0.802956	0.814153	0.82618	0.817933
Q589T	H588N	0.855997	0.867725	0.877767	0.868739
Q589T	N588P	0.0525764	0.106028	0.167998	0.109564
Q589T	L588N	0.942627	0.949059	0.953995	0.950424
Q589T	A588N	0.77013	0.789992	0.807899	0.790866

Q589T	H588P	0.196399	0.24484	0.299509	0.247488
Q589T	H588L	0.913361	0.922015	0.928741	0.923086
Q589T	A588H	0.938762	0.950138	0.960523	0.95204
Q589T	L588P	0.0311872	0.0855178	0.143456	0.0868968
Q589T	A588P	0.29697	0.345975	0.396535	0.349837
Q589T	A588L	0.828309	0.846433	0.861905	0.847347
Q589T	M589T	1	1	1	1
Q589T	M589Q	1	1	1	1
Q589T	A589M	1	1	1	1
Q589T	Q589T	1	1	1	1
Q589T	A589T	1	1	1	1
Q589T	A589Q	1	1	1	1
A589T	F583H	0.218973	0.267318	0.320835	0.273015
A589T	L585W	-0.466912	-0.400761	-0.334711	-0.404836
A589T	F585W	0.173136	0.271737	0.368094	0.282318
A589T	I585W	-0.0826207	0.0444774	0.157416	0.0433943
A589T	F585L	0.161457	0.221944	0.276948	0.225876
A589T	I585L	0.365603	0.422121	0.476749	0.430058
A589T	F585I	0.16008	0.287982	0.406716	0.300457
A589T	S586T	0.161759	0.219948	0.278359	0.224516
A589T	A587P	0.573639	0.629198	0.6743	0.640881
A589T	H588N	0.561835	0.6262	0.675585	0.632318
A589T	N588P	0.013092	0.0796778	0.150175	0.0818501
A589T	L588N	0.708985	0.744835	0.770778	0.753502
A589T	A588N	0.549862	0.599588	0.642558	0.60824
A589T	H588P	-0.0347897	0.0301399	0.102947	0.0301491
A589T	H588L	0.627012	0.671714	0.709238	0.679601
A589T	A588H	0.714481	0.768505	0.819705	0.779044
A589T	L588P	0.0145005	0.0767062	0.146671	0.0791503
A589T	A588P	0.0839389	0.151211	0.217324	0.154786
A589T	A588L	0.596823	0.651984	0.695319	0.658814
A589T	M589T	1	1	1	1
A589T	M589Q	1	1	1	1
A589T	A589M	1	1	1	1
A589T	Q589T	1	1	1	1
A589T	A589T	1	1	1	1
A589T	A589Q	1	1	1	1
A589Q	F583H	0.566362	0.594292	0.622859	0.599141

A589Q	L585W	0.842518	0.87691	0.900334	0.880724
A589Q	F585W	0.757333	0.784199	0.80782	0.789586
A589Q	I585W	0.511474	0.550548	0.584331	0.553989
A589Q	F585L	0.647221	0.678017	0.704434	0.682002
A589Q	I585L	0.34506	0.382892	0.417525	0.385933
A589Q	F585I	0.778609	0.817476	0.850239	0.824274
A589Q	S586T	0.527521	0.555718	0.578013	0.559076
A589Q	A587P	0.779712	0.799686	0.819184	0.805849
A589Q	H588N	0.850867	0.875346	0.896006	0.876859
A589Q	N588P	0.368269	0.427761	0.482045	0.433317
A589Q	L588N	0.88703	0.89749	0.906085	0.8993
A589Q	A588N	0.759905	0.788345	0.812009	0.790563
A589Q	H588P	0.334314	0.394742	0.452035	0.400742
A589Q	H588L	0.926399	0.942833	0.956197	0.945271
A589Q	A588H	0.916844	0.932468	0.946369	0.937113
A589Q	L588P	0.391347	0.447345	0.495941	0.452695
A589Q	A588P	0.295553	0.356449	0.414503	0.361675
A589Q	A588L	0.873165	0.89395	0.911214	0.896538
A589Q	M589T	1	1	1	1
A589Q	M589Q	1	1	1	1
A589Q	A589M	1	1	1	1
A589Q	Q589T	1	1	1	1
A589Q	A589T	1	1	1	1
A589Q	A589Q	1	1	1	1

Table S4_2 – The average epistatic effect $\gamma_{A_i \rightarrow}$ (eq. S1_9) of a mutation occurring on any background calculated over 1,000 posterior samples. Shown are the 2.5%, 50%, 97.5% confidence intervals and the corresponding value calculated from the median growth rates.

A_i	2.5%C.I.	50%C.I.	97.5%C.I.	Main text
F583	0.645879	0.655648	0.665972	0.659633
H583	0.645879	0.655648	0.665972	0.659633
W585	0.509098	0.519484	0.529889	0.523148
L585	0.494295	0.504828	0.515661	0.509236
F585	0.480383	0.493608	0.506724	0.500503
T585	0.524727	0.537596	0.549722	0.542512
S586	0.574784	0.58707	0.598905	0.591946
T586	0.574784	0.58707	0.598905	0.591946
A587	0.470531	0.484783	0.498591	0.491348

P587	0.470531	0.484783	0.498591	0.491348
N588	0.753967	0.760283	0.766723	0.763474
H588	0.714414	0.722585	0.72996	0.726474
P588	0.790154	0.79631	0.802035	0.799044
L588	0.78632	0.792616	0.798058	0.795508
A588	0.69515	0.702644	0.709753	0.706728
M589	0.625289	0.637087	0.648515	0.640628
T589	0.601966	0.610839	0.620123	0.614661
Q589	0.614382	0.625208	0.635169	0.629363
A589	0.59604	0.608972	0.621588	0.614412

Table S4_3 – The average epistatic effect $\gamma_{\rightarrow A_j}$ (eq. S1_13) of a background on any new mutation calculated over 1,000 posterior samples. Shown are the 2.5%, 50%, 97.5% confidence intervals and the corresponding value calculated from the median growth rates. In accordance with our results from the main text the average epistatic effect of a background on any new mutation is usually small, except for background P588*, which shows a strong magnitude effect.

A_j	2.5%C.I.	50%C.I.	97.5%C.I.	Main text
583F	0.643547	0.654621	0.665255	0.658462
583H	0.643547	0.654621	0.665255	0.658462
585W	0.623883	0.633104	0.641805	0.636432
585L	0.733728	0.740188	0.746721	0.743645
585F	0.777099	0.78529	0.792513	0.7899
585T	0.659718	0.67033	0.680289	0.674306
586S	0.709667	0.720978	0.730423	0.724845
586T	0.709667	0.720978	0.730423	0.724845
587A	0.774192	0.782478	0.789768	0.78691
587P	0.774192	0.782478	0.789768	0.78691
588N	0.60486	0.614512	0.62454	0.618562
588H	0.673788	0.682904	0.692253	0.687369
588P	0.14836	0.172296	0.194468	0.17391
588L	0.650123	0.659805	0.669129	0.663803
588A	0.658737	0.668566	0.677671	0.672761
589M	0.601843	0.612576	0.622367	0.615955
589T	0.587559	0.599117	0.61039	0.602947
589Q	0.706356	0.714376	0.722292	0.717983
589A	0.667143	0.678345	0.689501	0.683378

Table S4_4 – Observed decay of landscape-wide epistasis measure γ_d (eq. S1_15) with mutational distance d calculated over 1,00 posterior samples. Shown are the 2.5%, 50%, 97.5% confidence intervals and the corresponding value calculated from the median growth rates. Result based on the posterior samples support our finding from the main text, i.e., that the general topography of the fitness landscape resembles that of a RMF landscape with intermediate ruggedness

Mutational distance d	2.5%C.I.	50%C.I.	97.5%C.I.	Main text
0	1	1	1	1
1	0.656709	0.663385	0.668302	0.667961943
2	0.565145	0.571478	0.576907	0.575378659
3	0.529712	0.535176	0.540732	0.539030991
4	0.512834	0.517746	0.522954	0.521536863
5	0.509929	0.51507	0.519867	0.518957681

Table S4_5 – Roughness-to-slope ratio calculated over 1,000 posterior samples. Shown are the 2.5%, 50%, 97.5% confidence intervals and the corresponding value calculated from the median growth rates. Variation between measures is very low, all supporting a fitness landscape consisting of a mixture of a random HoC component and an additive component (i.e., a RMF landscape).

2.5%C.I.	50%C.I.	97.5%C.I.	Main text
0.910872	0.92082	0.930658	0.915688

Table S4_6 – The type and frequency of the global optimum sequence for the 1,000 posterior fitness landscapes. Colored cells represent the mutant that was found to be the global optimum in the median growth rate fitness landscape (i.e., in the main text).

Sequence	Frequency
QFGWSPLAE	3
QFGFSALTE	1
QFGLSPLAE	996

Table S4_7 – The type and frequency of the local optima sequences for the 1,000 posterior fitness landscapes. Colored cells represent those mutants that were found to be the local optima in the median growth rate fitness landscape (i.e., in the main text).

Sequence	Frequency
QFGWSPLME	7
QFGWSPLAE	4
QFGWTPAME	234
QFGWSPAQE	423
QFGLTPHME	7

QFGLTPLME	5
QFGLTALME	831
QFGLSALTE	158
QFGFSALTE	842
QFGLTPLQE	8
QFGLTAHQE	6
QFGFSPLQE	11
QFGLSPLAE	996
QFGLSPAME	393
QFGFSPAME	5
QFGLSPAQE	119
QFGLSAAAE	3
QFGISANTE	5
QFGITANQE	57
QFGISALME	1000
QFGITALQE	182
QFGISAHQE	6
QHGLTPPTE	76
QHGLSALME	32
QHGITPPTE	922

Table S4_8 – The number of local optima for the 1,000 posterior fitness landscapes. The colored cell represents the number of local optima that was found in the median growth rate fitness landscape (i.e., in the main text).

No. of optima	Frequency
4	27
5	184
6	370
7	285
8	119
9	13
10	2