

5           **Species level resolution of 16S rRNA gene amplicons sequenced  
                  through MinION™ portable nanopore sequencer**

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Running Title: Species level resolution of 16S amplicon by using MinION sequencer

25 **Abstract**

The miniaturized and portable DNA sequencer MinION™ has been released to the scientific community in the framework of an early access programme to evaluate its scope in a wide variety of genetic approaches. Although this technology is under constant development to completely deliver error-free and high quality reads, it has demonstrated a great potential especially in wide-genome analyses. In this study, we tested the ability of the MinION™ to perform amplicon sequencing in order to design new approaches to study microbial diversity using nearly full-length 16S rDNA sequences. Using the R7.3 chemistry, we have generated more than 3.8 million events (nt) during a sequencing run. Such data was enough to reconstruct more than 90% of the 16S rRNA gene sequences for 20 different species present in the mock community used as a reference. After read mapping and 16S rRNA gene assembly, we could recover consensus sequences useful to make taxonomy assignments at the species level. Additionally, we could measure the relative abundance of all the species present in the mock community by detecting a homogeneous distribution for most of the species as expected. Despite the nanopore-based sequencing produces reads with lower per-base accuracy in comparison with platforms such as Illumina and 454, promising results were obtained from MinION™, indicating that this technology is helpful to perform microbial diversity analysis. With the imminent improvement of the nanopore chemistry, better results and global performance of the platform are expected to contribute to the specific detection of microbial species and strains in complex ecosystems.

**Keywords:** MinION, nanopore sequencer, 16S rDNA amplicon sequencing, microbial diversity, long-read sequencing

## 50 **Introduction**

The third generation of DNA sequencers is based on single-molecule analysis technology that constantly is under development to deliver error-free and high quality reads. Oxford Nanopore Technologies (ONT) released the first miniaturized and portable DNA sequencer to researchers in early 2014 in the framework of the

55 MinION™ Access Programme. The MinION™ is a USB-stick size device operated from a computer via USB 3.0. Real-time data analysis can be visualized in terms of number of reads and length distribution. Nucleotide basecalling and quality assessment of reads require a further processing where data exchange of Hierarchical Data Format (HDF5) files, containing a large amount of numerical data, is indispensable. This data

60 exchange is done via the Internet through the Metrichor platform, process that can optionally be launched after sequencing process itself. According to its theoretical capabilities, the MinION™ brings out new alternatives for genomic analyses of which the accomplishing of completely finished bacterial genomes is some of the most attractive, as it has been demonstrated recently by Quick and co-workers (Quick et al

65 2014). The moderate throughput that MinION™ exhibits in terms of number of reads when compared with other massive sequencing platforms is definitively compensated with its performance in terms of read length, thus making it possible to obtain reads of thousands nucleotides in length. Short-read length sequencing approaches have permitted delivering high quality but partial genome sequences with unsolved repetitive

70 elements, thus making it impossible to study genetic variation or molecular evolution directly or indirectly associated to such elements. Therefore, long-read approaches offer new insights into genomic analyses facilitating consecution of finished genomes through hybrid assembly strategies (Utturkar et al 2014). Additionally to genome sequencing analysis, microbial diversity and taxonomy approaches are also deeply

75 limited by short-read strategies. Early massive sequencing approaches producing  
effective 50nt (Genome Analyzer, Solexa/Illumina) to 200nt (454 Roche) reads only  
permitted to accurately explore diversity at the phylum level. However, and thanks to  
the chemistry improvement of most common sequencing platforms, in recent years  
dozens of studies have presented a vast inventory of human- or environment-associated  
80 microbial communities reaching detail at the family or even genus level. To date, the  
paired-end short reads approaches for massive sequencing permits the analysis of  
sequence information of roughly 30% (~500nt) of the 16S rDNA, leaving taxonomic  
assignment of reads at the species level elusive. Therefore, implementation of long-read  
sequencing approaches to study the 16S rRNA genes should be determinant to design  
85 new studies conducted to evidence central role of precise bacterial species in a great  
variety of microbial consortia. As a consequence, we present a preliminary study of 16S  
rDNA amplicon sequencing from a mock microbial community composed of genomic  
DNA from 20 different bacterial species (BEI Resources) using MinION™ to evaluate  
the scope of nanopore technology on bacterial diversity and taxonomic analysis  
90 performing sequence analysis of the nearly-full length bacterial 16S rRNA genes.

## Methods

### *Bacterial DNA and 16S rDNA Amplicons*

Genomic DNA for the reference mock microbial community was kindly donated by  
95 BEI Resources (<http://www.beiresources.org>). This mock community (HM-782D) is  
composed of a genomic DNA mix from 20 bacterial strains containing equimolar  
ribosomal RNA operon counts (100,000 copies per organism per  $\mu\text{L}$ ) as indicated by the  
manufacturer. According to BEI Resources instructions, 1  $\mu\text{L}$  of mock community DNA  
was used for amplification of 16S rDNA genes. A 30 cycles PCR of 95° for 20 sec,

100 47°C for 30 seg, and 72°C for 60 seg; was setup using with Phusion High-Fidelity Taq  
Polymerase (Thermo Scientific) and S-D-Bact-0008-c-S-20 and S-D-Bact-1391-a-A-17  
primers targeting a high range of bacterial 16S rDNA (Klindworth et al 2012, Loy et al  
2007). PCR reactions produced ~1.5Kbp blunt-end fragments, which were purified  
using Illustra GFX PCR DNA and Gel Band Purification Kit (GE Healthcare).  
105 Amplicon DNA was quantified using a Qubit 3.0 fluorometer (Life Technologies).

#### Amplicon DNA library preparation

The Genomic DNA Sequencing Kit SQK-MAP-005 was used to prepare the amplicon  
library to be loaded in MinION™. Approximately, 250 ng of amplicon DNA (0.25  
110 pmol) was processed for end-repair using NEBNext End Repair Module (New England  
Biolabs) followed by purification using Agencourt AMPure XP beads (Beckman  
Coulter). Subsequently, we used 200 ng of the purified amplicon DNA (~0.2 pmol) to  
perform dA-tailing using the NEBNextdA-tailing module (New England Biolabs) in a  
total volume of 30  $\mu$ l according to the manufacturer's instructions during 15 minutes at  
115 37°C. To the 30  $\mu$ l dA-tailed ampliconDNA, 50  $\mu$ l Blunt/TA ligase master mix (New  
England Biolabs), 10  $\mu$ l of Adapter mix, and 2  $\mu$ l HP adapter were added and the  
reaction was incubated at 16°C for 15 minutes. The adaptor-ligated amplicon was  
recovered by using Dynabeads® His-Tag (Life Technologies) and washing buffer  
provided with the Genomic DNA Sequencing Kit SQK-MAP-005 (Oxford Nanopore  
120 Technologies). Finally, the sample was eluted from the Dynabeads® by adding 25  $\mu$ l of  
elution buffer and incubating for 10 minutes before pelleting in a magnetic rack.

#### Flowcellset-up

The brand sealed R7.3 flowcell was stored at 4°C until usage. The R7.3 flowcell was  
125 fitted to the MinION™ ensuring a good thermal contact with plastic screws. Priming of  
the R7.3 flow cell was done two times with premixed 71  $\mu$ l nuclease-free water, 75  $\mu$ L  
2x Running Buffer, and 4  $\mu$ L Fuel Mix. At least 10 minutes were needed to equilibrate  
the flowcell before every priming round and final DNA library loading.

### 130 Amplicon DNasequencing

The sequencing mix was prepared with 63  $\mu$ l nuclease-free water, 75  $\mu$ l 2x Running  
Buffer, , 8  $\mu$ L DNA library, and 4 $\mu$ L Fuel Mix. A standard 48-hour sequencing protocol  
was initiated using the MinKNOW™ v0.50.1.15. Base-calling was performed through  
data transference using the Metrichor™ agent v2.29.1 and 2D basecalling workflow  
135 v1.16. During the sequencing run, one additional freshly diluted aliquot of DNA library  
was loaded after 12 hours of initial input.

### Data analysis

Quality assessment of read data and conversion to fasta format was performed with  
140 *porertools*(Quick et al 2014) and *poRe*(Watson et al 2014) packages. Read mapping was  
performed against the reference 16S ribosomal RNA sequences (accessions  
NC\_009085, NZ\_ACYT00000000, NC\_003909, NC\_009614, NC\_009617,  
NC\_001263, NC\_017316, NC\_000913, NC\_000915, NC\_008530, NC\_003210,  
NC\_003112, NC\_006085, NC\_002516, NC\_007493, NC\_010079, NC\_004461,  
145 NC\_004116, NC\_004350, and NC\_003028) using the LAST aligner v.189 with  
parameters -q1 -b1 -Q0 -a1 -e45 which were configured to give the best balance  
between 16S rDNA assembly length and variants. LAST outputs were converted to sam  
files and processed with *samtools* (Li et al 2009) to build indexed bam files and obtain

consensus sequences from alignments and variant calling. Read mapping stats from sam  
150 files were calculated with the *ea-utils* package and its *sam-stats* function (Aronesty  
2011). Different comparisons, GC content correlations, and plots were performed and  
designed in R v3.2.0 (<http://cran.r-project.org>). The species coverage was calculated by  
obtaining fold-change ( $\text{Log}_2$ ) of species-specific read counting against the expected  
average for the entire community. A coverage bias was assumed when coverage  
155 deviation was lower than -1 or higher than 1. The Simpson's reciprocal index was  
calculated with the general formula,  $D = 1 / \sum p_i^2$ , where  $p_i$  is the proportion of reads  
belonging to the  $i$ th species.

## Results

160 The raw data collected in this experiment was obtained from MinKNOW software  
v0.50.1.15 (Oxford Nanopore Technologies) as fast5 files after conversion of electric  
signals into base calls via Metrichor Agent v2.29 and the 2D Basecalling workflow  
v1.16. Base called data passing quality control and filtering was downloaded and basic  
statistics of the experiment's data was assessed with *poretools* (Loman and Quinlan  
165 2014) and *poRe* (Watson et al 2014) packages. Fasta sequences were filtered by size and  
then mapped against reference 16S rRNA gene sequences using common and publicly  
available sequence analysis tools (see methods). Mapping stats are depicted in [Table 1](#)  
and fast5 raw data can be accessed at ENA (European Nucleotide Archive) under  
project PRJEB8730 (sample ERS760633). Only one data set was generated after a  
170 sequencing run of MinION™. After the sequencing process, we obtained 3,404 reads of  
which 58.5% were "template" reads (1,991), 23.8% were "complement" reads (812),  
and 17.7% were "2d" reads (601). Read lengths had a wide distribution ranging from  
reads with 12 nt to more than 50,000 nt in length with median of 1,100 nt. We

hypothesize that extremely large reads could be products of multiple amplicon ligation.

175 However, when we tried to perform alignment among reference sequences and large reads, we could not detect any matches (data not shown). Accordingly, we performed a filtering step only retaining 97% of the original dataset (3,297 reads) with a size range between 100 to 2,000nt in length for downstream analysis. When we performed initial analysis using separate sets of reads being "template", "complement", and "2d", we

180 observed a detrimental effect of 2d reads in the quality of assembled sequences, thus obtaining a higher number of unnatural variants along several 16S sequences (~14%). This could be explained because more than 86% of 2d reads were considered as low quality, a fact critical for aims addressed in this study and regarding study of sequence variants after assembly to distinguish very close species as those included in the mock

185 community and belonging to the *Streptococcus* (3 species) and *Staphylococcus* (2 species) genus. Given that this effect was even present for some species when the full set of "template", "complement", and "2d" reads were combined and used to accomplish the sequence analyses, , the dataset was finally reduced to contain information from "template" and "complement" reads (2,696 in total). Using reference sequence

190 information for the mock microbial community analyzed, we reconstructed more than 90% of 16S rRNA gene sequences for all organisms included in the mock community (Table 1). We observed that even at very low coverage as that retrieved for *Bacteroides vulgatus* 16S rRNA gene (Figure 1), it is possible to reconstruct almost 94% of the entire molecule; therefore, MinION™ sequencing shows no size limitations additional

195 to that associated to the PCR process itself. Indeed, the maximum size of amplicons sequenced in all cases was that expected according to PCR design (Table 1). In terms of coverage, we retrieved a notable lower number of 16S reads than expected from *B. vulgatus* species (Figure 1). We could not define if such lower coverage could be caused

during PCR amplification even using high coverage primers (Klindworth et al 2012,  
200 Loy et al 2007), or if that bias coverage is the result of the sequencing process itself.  
Despite this, *B. vulgatus* 16S rDNA amplicon sequences were almost fully assembled  
with low amounts of variants after DNA read alignment and pileup (Table 1). Data  
produced by MinION™ was further assessed to theoretically calculate the level of  
diversity observed using the Simpson's reciprocal index. This diversity index was  
205 estimated to be 17.785, a value very close of the maximum expected, 20 for the mock  
community analyzed. Read mapping stats were analyzed in order to further measure the  
performance of MinION™ sequencing in microbial diversity analysis based on 16S  
rDNA sequences. GC content of reads produced by MinION™ showed an important  
and significant correlation (Pearson's  $r = 0.47$ ,  $p\text{-value} \leq 0.0376$ ) against GC content of  
210 reference values (Figure 2A) which indicates that 16S rDNA GC content is fairly well  
replicated during sequencing. However, we found a 16S rDNA GC content bias to some  
extent in the reads obtained from MinION™ that in all cases exceeds the GC content of  
reference (Figure 2A). To test the probable influence of such bias and GC content itself  
in basecalling accuracy, we performed linear comparisons against mismatch rate, indels  
215 rate, and coverage deviation. We observed that coverage deviation ( $p\text{-value} \leq 0.00003$ )  
and mismatch rate ( $p\text{-value} \leq 0.00004$ ) are influenced by read GC content (Figure 2B  
and 2C, respectively). In the first case, the influence of GC content on coverage  
deviation could have minimal effect because 95% of species analyzed show no more  
than 1-fold deviation. However, with GC bias detected in reads from the MinION™  
220 sequencer, this effect could be magnified, especially in species where GC content is  
high. On the other hand, we found a strong correlation between GC content of reads and  
the mismatch rate retrieved from alignments which would insinuate again the GC  
content as a factor that influences 16S amplicon sequencing in the MinION™ platform.

Conversely, GC content did not appear to profoundly affect indel rate (Figure 2D). The  
225 complete assembly of the amplified 16S rRNA gene permitted the quantification of the  
level of sequence variants in the consensus sequence. These variants were recovered  
after a pileup of reads against reference sequences and they were variable in number  
with a median of 8 variants per 16S rRNA gene (Table 1). Such number of nucleotide  
substitutions means that approximately 0.5% of the 16S rDNA sequence assembled  
230 from MinION<sup>TM</sup> reads retained unnatural genetic variants directly generated from the  
sequencing process itself, theoretically leaving a bona fide identification and taxonomy  
assignment of 16S rDNA sequences at species level. In the worst cases where number  
of variants are meaningfully (~2.3% of the full assembly), like those observed for the  
*Acinetobacter baumannii* and *Bacillus cereus* (Table 1), direct BLAST comparisons of  
235 these assembled 16S rDNA sequences against the non-redundant reference database  
only matched those sequences with homologue sequences belonging to the same  
species, respectively. As expected, a homogenous distribution for strand mapping was  
observed roughly obtaining 50% of reads mapped against the forward strand and 50%  
of reads mapped against the complement strand, on average (Table 1).

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## Discussion

The microbial diversity analyses based on 16S rDNA sequencing are frequently used in  
biomedical research to determine dysbiosis associated in a great variety of gut-related  
human diseases. Identification of microbial species inhabiting different organs and  
245 cavities of human body relies on handling and processing of millions of DNA sequences  
obtained through the second generation of massive and parallel sequencing methods  
which still present limitations mainly in terms of DNA read length. Inability to fully  
determine 16S rDNA sequences during massive sequencing has led to the development

of multiple algorithms dedicated to theoretically discern microbial species present in  
250 samples according to the sequence similarity degree, the Operational Taxonomy Units  
(OTUs). Despite high accuracy and a constant update of methods used in OTU-based  
approaches, available algorithms produce no consensus outputs leaving a high degree of  
uncertainty when the number of theoretical species and their abundance is being subject  
of study (He et al 2015, Koskinen et al 2014, Schmidt et al 2014a, Schmidt et al 2014b).

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The third generation of sequencing methods based on single-molecule technology offers  
a new fashion to study the microbial diversity and taxonomic composition thanks to  
overcoming DNA read limitations at the expense of decreasing their throughput.  
MinION<sup>TM</sup> is one of these single-molecule methodologies which has demonstrated its  
260 capacity in genome sequencing (Ashton et al 2015, Quick et al 2014). Very recent  
reports have shown application of this technology in medical microbiology by using  
amplicon sequencing to potentially determine bacterial and viral infections (Kilianski et  
al 2015, Quick et al 2015). In this study, we have further explored the scope of  
MinION<sup>TM</sup> into microbial diversity studies by using amplicon sequencing of nearly full-  
265 size 16S rDNA from a mock bacterial community, obtaining a Simpson's reciprocal  
index diversity index close to expected. Our results indicate that MinION<sup>TM</sup> per-base  
accuracy (65-70%) is in concordance with previous results (Kilianski et al 2015,  
Mikheyev and Tin 2014, Quick et al 2014). We found that sequence coverage was close  
to expected in most of cases with only one exceptions, *B. vulgatus* (gene GC = 52%)  
270 which presented 1.84-fold less of the expected coverage. Although we could not  
demonstrate the definitive implication of the sequencing process itself in this coverage  
bias, this effect could be associated with the PCR process despite using "universal"  
primers with higher coverage among bacteria species during amplicon synthesis

(Klindworth et al 2012). In any case, such coverage was enough to reconstruct 93% of  
275 the 16S rRNA gene with a low proportion of unnatural variants. We observed a general  
influence of GC content in the mismatch rate but not in the indel rate, suggesting that  
base miscalling could be associated with the amplicon GC content. Moreover, a slight  
correlation between the amplicon GC content observed and coverage bias was  
evidenced, indicating that GC content could be negatively affecting amplicon coverage  
280 to some extent. Although MinION<sup>TM</sup> replicated fairly well the GC content expected for  
every amplicon sequenced, we observed a slight overrepresentation of GC in all reads  
obtained. This over-calling of GC bases in 16S rDNA amplicons could additionally  
influence the issues stated above in a negative manner.

285 The R7.3 chemistry used in MinION<sup>TM</sup> allowed the acquisition of reads of moderate  
quality which were enough to reconstruct more than 90% of the 16S rDNA molecule in  
all 20 bacterial species analyzed. None of the 20 16S rDNA consensus sequences  
analyzed showed more than 3% of sequence variation, which can be considered as a  
threshold for canonical species identification. Therefore, the consensus sequence  
290 assembled were useful to get a reliable taxonomic identification at the species level. As  
expected, unnatural variants were associated with low coverage regions; therefore,  
increasing the sequencing coverage will reduce drastically the ambiguities at the  
assembled sequences. Notwithstanding, further analysis could help to understand if  
some coverage bias might be associated with certain taxonomic groups.

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We have obtained promising results regarding the study of microbial communities by  
using 16S rDNA amplicon sequencing through MinION<sup>TM</sup> device. Despite the observed  
modest per-base accuracy of this sequencing platform, we were able to reconstruct

nearly full-length 16S rDNA sequences for 20 different species analyzed from a mock  
300 bacterial community while obtaining a modest coverage for some species. To date,  
MinION™ and nanopore technology have demonstrated a great potential in DNA  
sequencing allowing one to retrieve bacterial whole genome sequences with a minimum  
level of variation (Quick et al 2014). With the results presented here, we postulate that  
the MinION™ platform is a reliable methodology to study diversity of microbial  
305 communities permitting: i) a taxonomy identification at the species level through 16S  
rDNA sequence comparisons; and ii) a quantitative method to determine the relative  
species abundance. This type of analysis will likely become more accurate over time as  
the nanopore chemistry is improved in future releases together the implementation of  
the "What's In My Pot" (WIMP) Metrichor workflow aiming real-time taxonomic  
310 identification of sequences by comparison against different bacterial references  
databases (i.e. NCBI, SILVA, GreenGenes). Accordingly, sequence studies of the entire  
16S rDNA molecule could permit a bypass of OTU-based analysis, thus making it  
feasible to obtain a direct inventory of bacterial species and relative abundance, as well  
as determine the key players at the species and/or strain level in different microbial  
315 communities of interest. Implications of the primary and secondary structure of 16S  
rDNA amplicons in the MinION™ sequencing performance must be further explored in  
order to evaluate, minimize, and correct for technical bias regarding quantitative  
approaches of microbial diversity studies.

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### **Conflict of Interest**

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**Table 1. Statistics of the mapping process using 16S rDNA reads produced by MinION™.**

Organism	Mapped Reads	Mapped Bases	Strand Mapping <sup>a</sup>	Max Length <sup>b</sup>	Mean Length	Variants	Consensus	rRNA gene <sup>c</sup>	Assembled 16S
<i>A. baumannii</i>	98	99,352	0.46:0.54	1,390	1,013	32	1,415	1,529	0.93
<i>A. odontolyticus</i>	79	73,480	0.49:0.51	1,377	930	8	1,407	1,528	0.92
<i>B. cereus</i>	144	151,668	0.46:0.54	1,419	1,053	31	1,415	1,508	0.94
<i>B. vulgatus</i>	33	29,499	0.58:0.42	1,346	893	25	1,403	1,510	0.93
<i>C. beijerinckii</i>	97	99,476	0.46:0.54	1,393	1,025	13	1,408	1,505	0.94
<i>D. radiodurans</i>	73	69,940	0.45:0.55	1,390	958	8	1,398	1,502	0.93
<i>E. faecalis</i>	149	153,581	0.50:0.50	1,398	1,030	8	1,444	1,549	0.93
<i>E. coli</i>	167	181,084	0.45:0.55	1,398	1,084	0	1,434	1,542	0.93
<i>H. pylori</i>	67	62,838	0.46:0.54	1,390	937	11	1,411	1,498	0.94
<i>L. gasseri</i>	123	128,120	0.51:0.49	1,407	1,041	0	1,467	1,579	0.93
<i>L. monocytogenes</i>	139	140,478	0.50:0.50	1,343	1,010	13	1,374	1,486	0.92
<i>N. meningitidis</i>	87	86,916	0.48:0.52	1,390	999	11	1,433	1,544	0.93
<i>P. acnes</i>	75	70,160	0.48:0.52	1,375	935	21	1,401	1,525	0.92
<i>P. aeruginosa</i>	113	120,520	0.55:0.45	1,398	1,066	14	1,425	1,536	0.93
<i>R. sphaeroides</i>	95	89,750	0.52:0.48	1,416	944	5	1,352	1,463	0.92
<i>S. epidermidis</i>	164	177,084	0.51:0.49	1,423	1,079	0	1,443	1,540	0.94
<i>S. aureus</i>	163	179,477	0.51:0.49	1,423	1,101	1	1,435	1,554	0.92
<i>S. agalactiae</i>	156	166,420	0.52:0.48	1,411	1,066	5	1,439	1,551	0.93
<i>S. mutans</i>	196	221,682	0.47:0.53	1,411	1,131	2	1,440	1,552	0.93
<i>S. pneumoniae</i>	154	168,657	0.52:0.48	1,411	1,095	2	1,442	1,560	0.92

395 a Proportion of reads mapped against the forward and complemented strand, respectively.

b Maximum length of reads mapped.

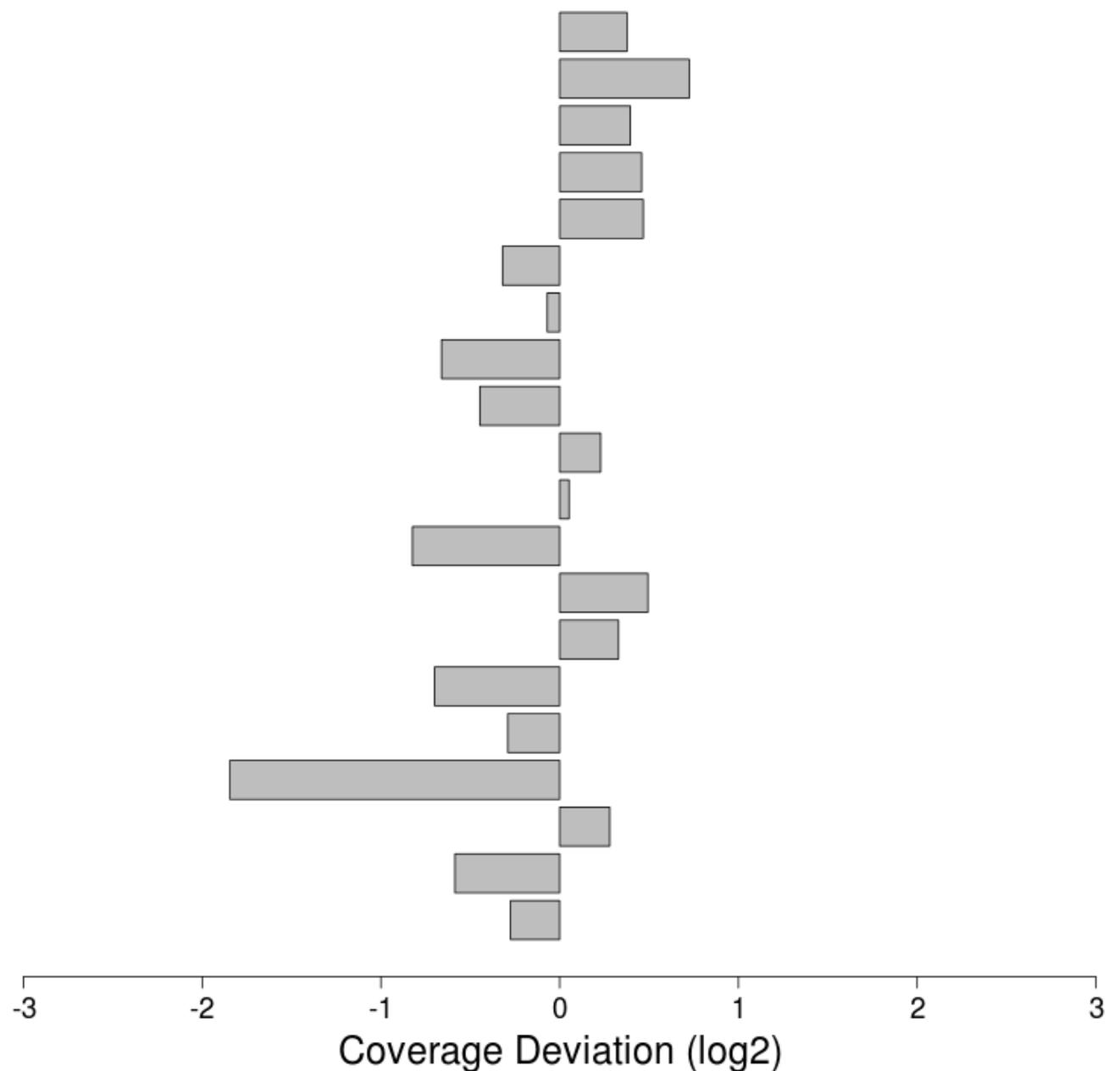
c Length of the 16S sequence used as reference.

## Figure Legends

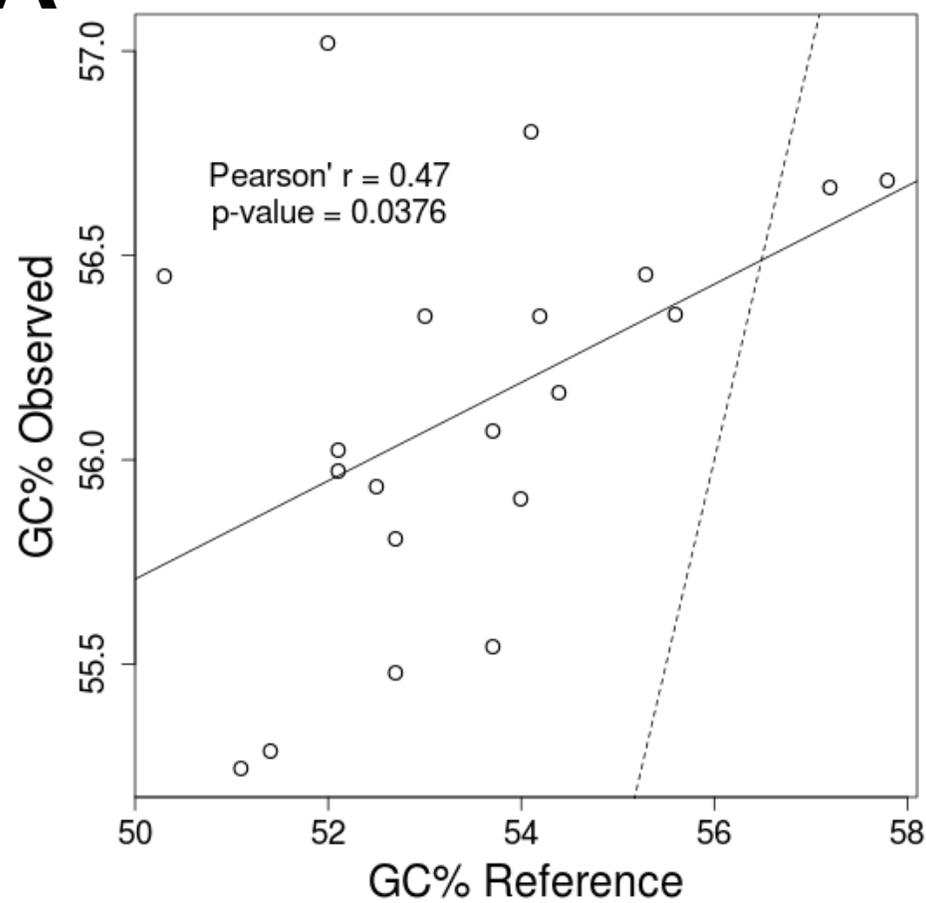
**Figure 1.** Species abundance in the mock community. Species coverage was calculated by obtaining fold-change ( $\text{Log}_2$ ) of species-specific read counting against the expected average for the entire community. A coverage bias was assumed when coverage deviation was lower than -1 or higher than 1.

**Figure 2.** Per-base accuracy of the mapped reads. A - Scatter plot of the GC content observed in mapped reads against the GC obtained from the references sequence. The dashed line indicates correlation with a Pearson's  $r = 1$ . B - Correlation between GC content observed in mapped reads and coverage bias observed in Figure 1. C - Influence of the GC content observed in mapped reads on mismatch rates calculated after mapping. D - Scatter plot of the observed GC content of mapped reads and indels rates calculated after mapping. In all cases the Pearson's  $r$  coefficients and p-values supporting such correlations are presented inside the scatter plots and solid lines indicate the tendency of correlations.

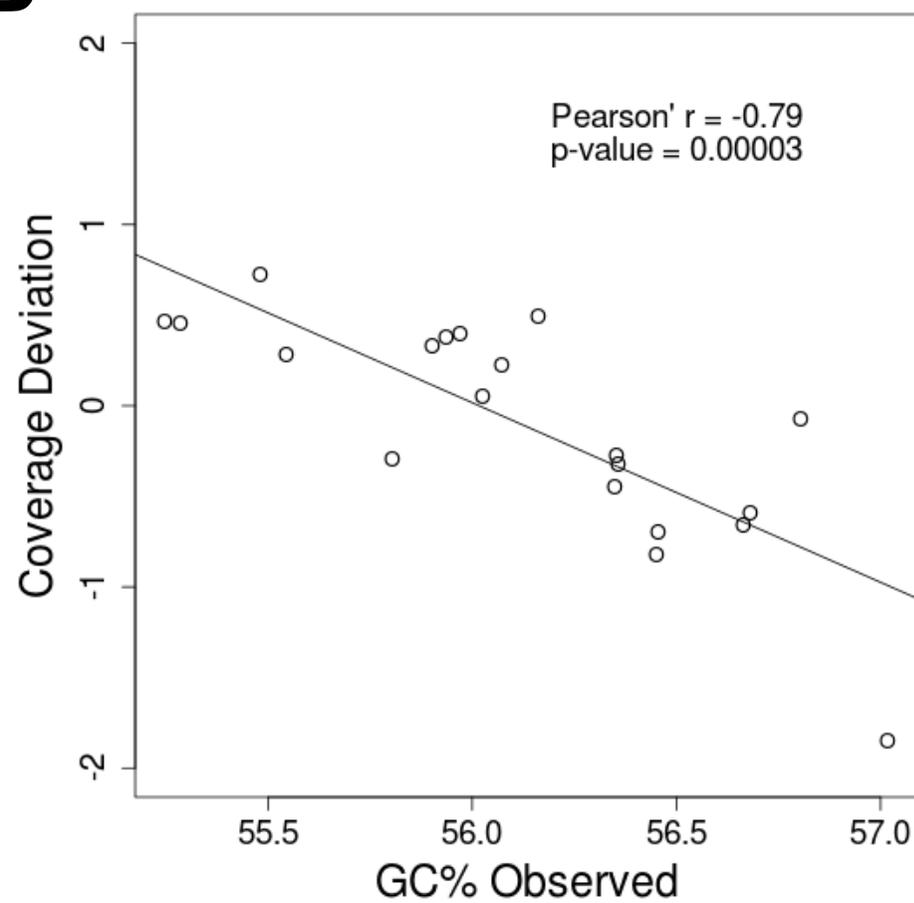
*Streptococcus pneumoniae*  
*Streptococcus mutans*  
*Streptococcus agalactiae*  
*Staphylococcus aureus*  
*Staphylococcus epidermidis*  
*Rhodobacter sphaeroides*  
*Pseudomonas aeruginosa*  
*Propionibacterium acnes*  
*Neisseria meningitidis*  
*Listeria monocytogenes*  
*Lactobacillus gasseri*  
*Helicobacter pylori*  
*Escherichia coli*  
*Enterococcus faecalis*  
*Deinococcus radiodurans*  
*Clostridium beijerinckii*  
*Bacteroides vulgatus*  
*Bacillus cereus*  
*Actinomyces odontolyticus*  
*Acinetobacter baumannii*



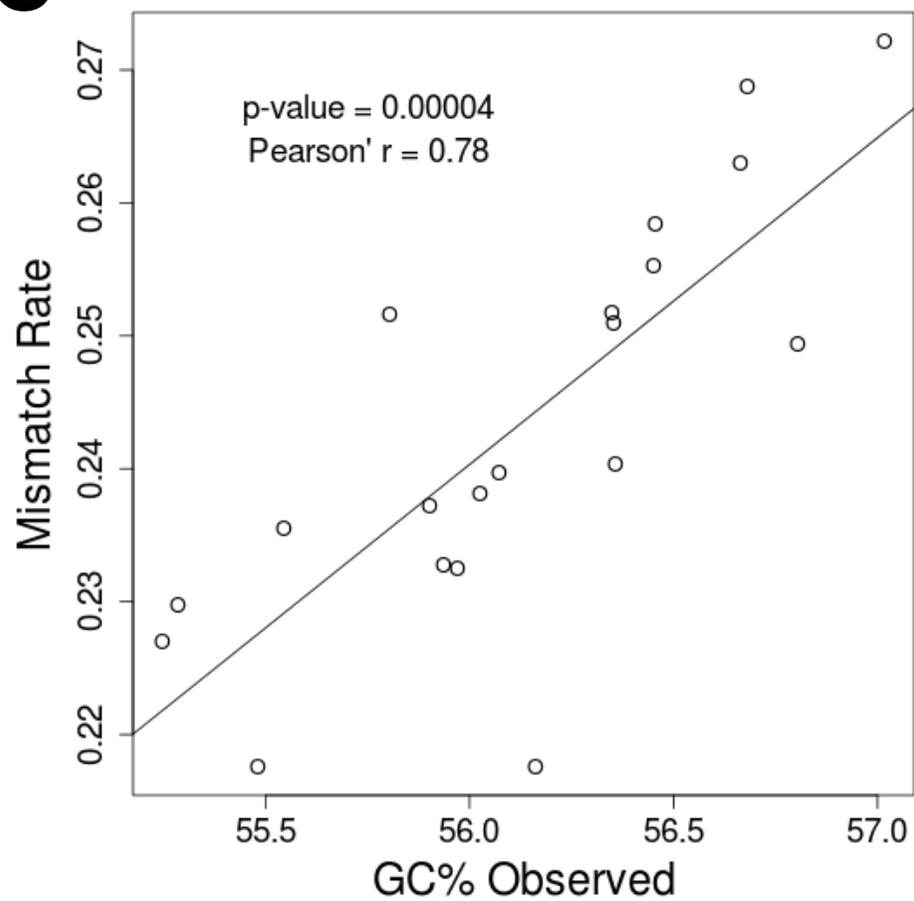
**A**



**B**



**C**



**D**

