

Selection constrains phenotypic evolution in a functionally important plant trait

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Abstract

A long-standing idea is that the macroevolutionary adaptive landscape – a ‘map’ of phenotype to fitness – constrains evolution because certain phenotypes are fit, while others are universally unfit. Such constraints should be evident in traits that, across many species, cluster around particular modal values, with few intermediates between modes. Here, I compile a new global database of 599 species from 94 plant families showing that stomatal ratio, an important functional trait affecting photosynthesis, is multimodal, hinting at distinct peaks in the adaptive landscape. The dataset confirms that most plants have all their stomata on the lower leaf surface (hypostomy), but shows for the first time that species with roughly half their stomata on each leaf surface (amphistomy) form a distinct mode in the trait distribution. Based on a new evolutionary process model, this multimodal pattern is unlikely without constraint. Further, multimodality has evolved repeatedly across disparate families, evincing long-term constraint on the adaptive landscape. A simple cost-benefit model of stomatal ratio demonstrates that selection alone is sufficient to generate an adaptive landscape with multiple peaks. Finally, phylogenetic comparative methods indicate that life history evolution drives shifts between peaks. This implies that the adaptive benefit conferred by amphistomy – increased photosynthesis – is most important in plants with fast life histories, challenging existing ideas that amphistomy is an adaptation to thick leaves and open habitats. I conclude that peaks in the adaptive landscape have been constrained by selection over much of land plant evolution, leading to predictable, repeatable patterns of evolution.

1 The topography of the macroevolutionary adaptive landscape is thought to shape
2 the broad patterns of life's diversity [1]. Adaptive landscapes with multiple peaks
3 are manifest in convergent evolution of similar phenotypes across independent evo-
4 lutionary lineages. In such cases, surveys across species should reveal a multimodal
5 trait distribution in which the modes point to the underlying peaks in the landscape.
6 Multimodality has been observed frequently among plants and animals, including
7 traits such as self-incompatibility [2], the precocial-altricial spectrum [3], pollination
8 syndromes [4], ecomorphology in *Anolis* [5], and plant height [6]. That such disparate
9 classes of traits show broadly similar patterns suggests that divergence on a multi-
10 peaked adaptive landscape may be a general feature of macroevolution. However, we
11 rarely know whether multimodality reflects constraints imposed by the underlying
12 adaptive landscape and not some other constraint on phenotypic evolution.

13 In particular, certain phenotypes may be common not because they are more fit,
14 but rather because they are genetically, developmentally, or functionally accessible.
15 Conversely, rare phenotypes might be inaccessible. I use the definitions given by
16 Arnold [7]: genetic constraints are limitations set by the "pattern of genetic variation
17 and covariation for a set of traits"; developmental constraints are limitations on "pos-
18 sible developmental states"; and functional constraints are imposed by "time, energy,
19 or the laws of physics". Arnold contrasts these with selective constraints determined
20 by the adaptive landscape. There are examples of genetic [8], developmental [9],
21 and functional [10] constraints on phenotypic evolution acting in nature, meaning
22 that we cannot assume selection alone shapes trait evolution. Compelling evidence
23 from cross species comparisons that selection constrains phenotypic evolution re-
24 quires showing that phenotypic evolution is constrained, that selection is sufficient
25 to explain the inferred constraint, and that nonselective constraints are inconsistent

26 with these observations.

27 Here, I evaluate evidence for selective constraints on a functionally important plant
28 trait, stomatal ratio, using comparative methods and theory. Stomatal ratio, defined
29 as the ratio of upper to lower stomatal density, impacts how plants ‘eat’ (i.e. as-
30 similate CO₂ from the atmosphere via photosynthesis). Physiological experiments
31 and biophysical theory demonstrate that amphistomatous leaves, those that have
32 equal stomatal densities on both upper and lower surfaces, maximize photosynthetic
33 rate by minimizing the distance between substomatal cavities and chloroplasts, fa-
34 cilitating rapid CO₂ diffusion [11, 12, 13, 14]. Hence, nearly all plants should be
35 amphistomatous to maximize photosynthesis, yet paradoxically up to 90% of plant
36 species in some communities are hypostomatous [15, 16, 17, 18], meaning that most
37 stomata are on the lower surface. In rare cases, most stomata are on the upper sur-
38 face (hyperstomy). I use upper and lower rather than abaxial and adaxial, because
39 the former applies to ‘upside-down’ (i.e. resupinate) leaves. Stomatal ratio is a quan-
40 titative metric that describes continuous variation between hypo- and hyperstomy.

41 Multiple lines of evidence indicate selection on stomatal ratio, but there is little
42 consensus on the adaptive significance. Stomatal ratio varies widely, but nonran-
43 domly [15, 17, 11, 19, 20, 21] and evolves rapidly in some taxa, possibly due to
44 selection [22, 23, 24]. Several environmental and anatomical factors have been hy-
45 pothesized to favour amphistomy (Table 1). The mechanistic details and literature
46 underlying these hypotheses and predictions are described in Text S1. The prepon-
47 derance of hypostomy almost certainly reflects a cost of upper stomata. For example,
48 hypostomy has evolved anew in resupinate leaves [25]. Upper stomata might be costly
49 because they increase susceptibility to foliar pathogens (e.g. rust fungi) that infect
50 through stomata [13], suggesting that stomatal ratio mediates a tradeoff between

51 photosynthetic rate and defence [23], but other costs have been proposed (Text S1).
52 Identifying the selective forces (i.e. fitness benefits and costs) shaping stomatal ratio
53 have been hampered by four methodological limitations. Namely, previous studies
54 were typically qualitative rather than quantitative, confined to specific geographic re-
55 gions or clades, did not account for phylogenetic nonindependence, and did not take
56 into account multiple confounding factors. To overcome these limitations, I assem-
57 bled a quantitative, global, and phylogenetically extensive database that disentangles
58 correlated predictor variables (e.g. light level and leaf thickness).

59 This new dataset revealed that stomatal ratio is a multimodal trait (Fig. 1). To
60 test whether the observed pattern is consistent with constraint, I modified previous
61 evolutionary process models to accommodate bounded traits like stomatal ratio. Fit-
62 ting this model to the data indicates that stomatal ratio is highly constrained by a
63 rugged adaptive landscape with multiple selective regimes (for discussion of selective
64 regimes, see [26, 27, 5]). This led me to evaluate whether selection is sufficient to
65 account for inferred constraints using theoretical and empirical approaches. First, I
66 constructed a simple cost-benefit model consistent with the underlying physics and
67 a minimum of additional assumptions. This model indicates that distinct peaks in
68 the adaptive landscape can result from selective constraints, even when the under-
69 lying environmental gradients are smooth. In contrast, a review of the literature
70 does not support a large role for genetic, developmental, and functional constraints.
71 Finally, phylogenetic multiple regression identifies life history evolution as the pri-
72 mary selective agent underlying peak shifts, but anatomical and climatic factors are
73 also important. By merging theory and data, this study adduces compelling new
74 evidence that selection is the primary constraint on phenotypic evolution, at least
75 for stomatal ratio. There is no reason to believe this trait is exceptional among func-

76 tional traits, and hence the inferences drawn here could be generalizable to many
77 other phenotypes that exhibit similar patterns indicative of evolutionary constraint.

78 Results

79 Stomatal ratio evolution is constrained by multiple selective 80 regimes

81 I compiled a new, global dataset from 25 previously published studies (Text S2)
82 containing trait data (stomatal ratio and leaf thickness) on 599 species across 94
83 plant families; the dataset with trait and climate data comprised a 552 species subset
84 covering 90 families. The most striking feature of the data is that stomatal ratio (SR)
85 is highly multimodal (Fig. 1), with apparent modes at 0 (hypostomatous), ≈ 0.5
86 (amphistomatous), and 1 (hyperstomatous). Note that here I am reporting stomatal
87 ratio as the ratio of upper density to total density so that the distinct hypo- and
88 hyperstomatous modes can be seen. Stomatal ratio does not conform to a nonmodal,
89 uniform distribution (Komologrov-Smirnov test, $D = 0.433$, $P = 1.11 \times 10^{-15}$),
90 even after removing all hypostomatous (SR = 0) species (K-S test, $D = 0.293$,
91 $P = 1.33 \times 10^{-15}$). The data are also inconsistent with a unimodal, truncated
92 exponential distribution bounded by 0 and 1 (K-S test, $D = 0.429$, $P = 1.11 \times 10^{-15}$).

93 In contrast, the distribution of stomatal ratio values across species is consistent
94 with an evolutionary process model that includes constraints imposed by multiple
95 selective regimes, indicating a rugged adaptive landscape. Although the results pre-
96 sented in this section only identify constraint, not necessarily selective constraint,

97 I use selective regime because evidence in the following sections indicates that se-
98 lection is the primary constraint. To infer regimes, I augmented a commonly used
99 model of selective regimes, the Ornstein-Uhlenbeck process [28], to account for traits
100 like SR that are bounded by 0 and 1 (see Materials and Methods and Text S3 for
101 further detail and mathematical derivation). Under a bounded Ornstein-Uhlenbeck
102 process model, the stationary distribution of stomatal ratio (or any proportion trait)
103 r follows a Beta distribution:

$$f(r) = \frac{r^{2\phi\theta-1}(1-r)^{2\phi(1-\theta)-1}}{B(2\phi\theta, 2\phi(1-\theta))} \quad (1)$$

104 $B(\cdot)$ refers to the Beta function. A selective regime at stationarity is characterized
105 by two parameters, a long-run average or ‘optimum’ in the adaptive landscape, θ ,
106 and a precision, ϕ , around the optimum. Greater values of ϕ produce distributions
107 that are more tightly constrained around the optimum.

108 If a trait evolves on an adaptive landscape with multiple peaks, then a model
109 with multiple selective regimes should fit the data better than a model with a single
110 regime [27, 5]. I used finite mixture model analysis (Text S4) to estimate the number
111 of selective regimes. This approach differs from conceptually similar methods, but
112 can be applied to non-Gaussian traits like SR (see [29, 30] for alternative methods
113 with Gaussian traits). I inferred three selective regimes (Table 2), but note that the
114 mapping between modes and regimes is not always one-to-one. In particular, one
115 regime produces modes at both 0 and 1 (Fig. S1). Nevertheless, the data clearly
116 support the large number of hypostomatous (SR = 0) species as a distinct mode

117 (Fig. S1). There was also strong support for an amphistomatous regime (compare
118 Fig. S1A to Fig. S1B). Finally, the best-supported model also included a small mode
119 for hyperstomatous species and a separate, smaller regime for species intermediate
120 between hypo- and amphistomy (Fig. S1C).

121 The same general pattern seen at the global scale – multiple selective regimes lead-
122 ing to distinct modes – is recapitulated within nine of ten families best-represented in
123 the global dataset (Fig. 2). Two regimes are supported in most (8 of 9) multi-regime
124 families, except Asteraceae, in which three regimes are favoured (Fig. 2A). In one
125 family, Rubiaceae, all species were inferred as members of a hypostomatous regime.
126 In all mutli-regime families except Poaceae, there are distinct regimes associated with
127 hypo- and amphistomy; in Poaceae, there are hyper- and amphistomous regimes in-
128 stead (Fig. 2E). However, the hyperstomatous species of Poaceae in this study may
129 not be representative of family since they are wetland specialists in the genus *Spartina*
130 [31]. Generally, the internal (i.e. amphistomatous) mode is closely centered around
131 0.5, as predicted from biophysical theory [11, 13], except in in the Rosaceae, where
132 the inferred optimum is closer to 0.25. Although I was unable to account for phyloge-
133 netic nonindependence in these analyses (see Materials and Methods), that a similar
134 pattern – species are either amphistomatous or hypo/hyperstomatous, but rarely in-
135 termediate – emerges independently in multiple families indicates the conclusions are
136 unlikely to change qualitatively once fully phylogenetic methods can be extended to
137 bounded traits. In summary, the apparent pattern of constraint on stomatal ratio is
138 strikingly similar across multiple disparate families and at a global scale, suggesting
139 convergent evolution because of shared phenotypic constraint.

140 Selection is sufficient to accommodate constraint

141 I analyzed a simple cost-benefit model of stomatal ratio to ask whether selection is
142 sufficient to account for apparent phenotypic constraint. Not surprisingly, selection
143 favours greater stomatal ratio (S_{fit}) as the fitness benefit of greater photosynthesis
144 increases relative to the cost of upper stomata (Fig. 3A-C), but the shape of the
145 function is highly sensitive to one parameter in the model, σ^2 . In particular, the
146 adaptive landscape goes from being smooth when σ^2 is high to rugged when σ^2 is
147 low (Fig. 3D-F). When the landscape is smooth, intermediate phenotypes between
148 complete hypostomy and amphistomy are best when the benefit:cost ratio itself is
149 intermediate. In contrast, when the landscape is rugged, intermediates are univer-
150 sally less fit than either of the boundary phenotypes. In a rugged landscape, as the
151 benefit:cost ratio decreases there is a sudden shift from amphistomy being favoured
152 to hypostomy being favoured. The dearth of species with intermediate SR in nature,
153 especially within families, therefore suggests that the adaptive landscape for stomatal
154 ratio is generally rugged. Numerical simulations based on smooth variation in the
155 benefit:cost ratio indicate that the simple, yet realistic assumptions of this model are
156 sufficient to generate qualitatively similar patterns of multimodality to those seen in
157 nature (Fig. 3G-H).

158 Growth form, leaf thickness, and precipitation shape stomatal 159 ratio evolution

160 If stomatal ratio is strongly associated with other traits or climatic factors, especially
161 if there are compelling *a priori* hypotheses (Table 1) supporting such associations,

162 then it suggests that trait variation is shaped by selection. Phylogenetic multiple
163 regression consistently identified growth form and, to a lesser extent, leaf thickness
164 and precipitation as the best predictors of stomatal ratio (Table 3). Amphistomy
165 was strongly associated with fast growth forms (herbaceous plants), whereas hypo-
166 tomy was most common in slower growing shrubs and trees (Fig. 4). As predicted
167 by biophysical theory [11, 13], thicker leaves also tended to be amphistomatous, al-
168 though the correlation was weak (Fig. S2A). Finally, amphistomy was more common
169 in dry environments, whereas hypo/hyperstomy were associated with higher precip-
170 itation (Fig. S2B). Elevation and leaf area index, a proxy for open habitat, were
171 not significantly associated with stomatal ratio in this dataset (Table 3). In single
172 regressions, amphistomy was more common more open environments, as in previous
173 studies [12, 18, 19, 21], but this correlation was not significant after precipitation
174 was factored into multiple regression (precipitation and leaf area index are positively
175 correlated).

176 Discussion

177 Phenotypic evolution is often constrained, but the relative role of selective versus
178 nonselective constraints is unclear. This study posits that multimodal traits reveal
179 distinct peaks of high fitness in a rugged adaptive landscape. Hence, the prevalence
180 of certain phenotypes and the dearth of others directly reflects selective constraints
181 on phenotypic evolution. Evidence from a new, global dataset clearly shows that
182 stomatal ratio is a multimodal trait (Fig. 1) and that multimodality has evolved
183 repeatedly in land plants (Fig. 2). These patterns are difficult to reconcile with
184 models omitting constraint, but are consistent with a rugged adaptive landscape

185 comprised of multiple selective regimes (Table 2). A simple cost-benefit model of
186 stomatal ratio shows that selection is a sufficient explanation, particularly when the
187 underlying adaptive landscape is predominantly rugged. Adaptive evolution from
188 one peak in the landscape to another (i.e. hypo- to amphistomy or *vice versa*)
189 appears to be primarily driven by growth form, suggesting that the fitness benefit
190 of amphistomy – faster diffusion of CO₂ to chloroplasts – is greatest in species with
191 ‘fast’ life histories.

192 **Multimodality implies constraint on the macroevolutionary adap-** 193 **tive landscape**

194 Just as water is only found as ice, liquid, and steam, despite continuous variation
195 in temperature, stomatal ratio comes in partially discrete clusters corresponding
196 to hypo-, amphi-, and hyperstomy, but less often intermediate (Fig. 1). In fact,
197 the modes identified here correspond remarkably with traditional botanical classi-
198 fications [32], suggesting that these workers recognized the pattern even without
199 quantitative analyses. The multimodal pattern in the dataset cannot be explained
200 by an evolutionary process model neglecting constraint (Text S3). However, appar-
201 ent clustering could occur by systematic underrepresentation of intermediate trait
202 values [33] or nonrandom taxon sampling. It is highly improbable that intermediate
203 phenotypes exist at greater frequency in nature but are rarely reported, as most
204 studies have no *a priori* hypothesis about stomatal ratio in their study organisms.
205 If anything, by omitting many studies that report only qualitative data, I might
206 have enriched the frequency of intermediate phenotypes, as these are the most likely
207 to be reported quantitatively. Nonrandom taxon sampling, without accounting for

208 phylogeny, could also give the appearance of multimodality. To give an extreme ex-
209 ample, if there had been a single transition from hypo- to amphistomy followed by
210 stasis, then sampling the tips of the phylogeny would produce a multimodal pattern
211 with apparently strong statistical support, even though it only represents a single
212 evolutionary event. Methodological limitations prevented me from fully accounting
213 for phylogeny (see Materials and Methods), but the fact that multimodality reap-
214 pears in multiple distantly-related families (Fig. 2) makes nonrandom taxon sampling
215 alone an unlikely explanation, though it might accentuate the pattern. Future work
216 is needed to extend regime-inference methods [27, 29, 30] to non-Gaussian traits, as
217 this study begins to do with a new evolutionary process model for proportion traits.

218 **Selection is the most likely explanation for phenotypic con-** 219 **straint**

220 In principle, constraint could reflect a mix of selective, genetic, developmental, and
221 functional factors [7]. However, the preponderance of available theory and data on
222 stomatal ratio suggests selection is responsible for most if not all of the phenotypic
223 constraint. Genetic, developmental, and functional constraints cannot explain the
224 dearth of intermediate phenotypes because intermediates are genetically accessible
225 as well as developmentally and functionally possible. The appropriate mutations
226 to generate intermediate phenotypes occur spontaneously during mutagenesis [34],
227 segregate among natural populations [35, 36, 37, 23], and are fixed between closely
228 related species [38, 24].

229 In contrast, the cost-benefit model presented here shows that with a small number

230 of realistic, evidence-based assumptions, selection is sufficient to accommodate the
231 data and helps clarify why discrete modes form even when the underlying environ-
232 mental gradients are smooth (environmental gradients need not be smooth, but it
233 is unnecessary to assume otherwise). Stomata are often distributed equally on both
234 surfaces (amphistomy) because this arrangement optimizes photosynthetic rate. This
235 was an assumption of the model based on biophysical theory [11, 13]. More often,
236 all stomata are on the lower surface because the costs of upper stomata outweigh
237 the benefits. A dearth of intermediates between hypo- and amphistomy occurs when
238 the landscape is rugged, making these phenotypes often fall in a fitness valley. How-
239 ever, the best mixture model includes a small peak of these intermediates (Table 2,
240 Fig. S1). This suggests that although the adaptive landscape is constrained and often
241 rugged, it may shift from rugged to smooth over macroevolutionary time. However,
242 the fact that most species, especially within families (Fig. 2), cluster around partic-
243 ular modes suggests that the landscape is predominantly rugged. Finally, the small
244 number of hyperstomatous species indicates that there are occasionally situations in
245 which upper stomata are favoured, such as in aquatic plants or those with unusual
246 epidermal or spongy mesophyll anatomy.

247 **Life history, more than anatomy and climate, determines stom-** 248 **atal ratio**

249 Nonrandom association between stomatal ratio, other ecologically important traits,
250 and climate also supports a significant role for selection in shaping trait evolution.
251 To my knowledge, this is the first study to rigorously demonstrate a strong associ-
252 ation between growth form and stomatal ratio, although it had been suggested by

253 earlier ecological surveys [15, 39]. Two hypotheses that might explain the relation-
254 ship between growth form and stomatal ratio are: 1) herbaceous plants have shorter
255 leaf lifespans [40], requiring higher photosynthetic rates to pay their construction
256 costs in a shorter time [41]; 2) herbaceous plants have faster life histories, leading to
257 stronger selection on high growth rates, mediated in part by higher leaf-level pho-
258 tosynthetic rate [42]. That the relationship between stomatal ratio and whole-plant
259 lifespan holds within herbaceous (annuals vs. perennials) and woody (shrubs vs.
260 trees), supports the second hypothesis (selection on faster life history favours am-
261 phistomy). Although this hypothesis requires further testing, if correct, it implies
262 remarkably strong selection on leaf-level photosynthesis, as the photosynthetic ad-
263 vantage of amphistomy over hypostomy is only a few percent in a typical herbaceous
264 leaf [11].

265 Surprisingly, I found little evidence supporting the most common adaptive expla-
266 nation for amphistomy, that thicker leaves ‘need’ stomata on both sides to facilitate
267 CO₂ diffusion [11]. In actuality, support for this hypothesis is mixed (Text S1), espe-
268 cially when phylogenetic nonindependence is taken into account [43, 39] (but see [44]).
269 It is now clear why previous studies came to different conclusions: thicker leaves do
270 tend to be amphistomatous, even once phylogeny is accounted for, but the trend is
271 weak (Fig. S2A). Less powerful studies than this one could easily have failed to de-
272 tect a significant relationship. Hence, leaf thickness, by constraining CO₂ diffusion,
273 imposes selection for amphistomy. I also found that amphistomy was more common
274 in plants from low precipitation environments. For a given stomatal conductance,
275 which is proportional to evaporative water loss, amphistomy improves water-use ef-
276 ficiency by increasing photosynthetic rate [11], suggesting a plausible mechanism for
277 selection on amphistomy in dry environments. Although low precipitation was cor-

278 related with habitat openness, measured using leaf area index, multiple phylogenetic
279 regression indicated that precipitation was causal, in contrast to previous studies
280 [18, 21]. These studies used finer scale (plant-level) descriptions of light environment
281 that might have been missed by the coarser, satellite-based measurements of canopy
282 cover used here. Alternatively, patterns at the global scale might differ from those
283 within particular families or biomes. Finally, I was unable to test the effects of leaf
284 orientation and stomatal packing on stomatal ratio, though these are likely to be
285 important factors in many plants [20]. The evidence from this and previous studies
286 shows that stomatal ratio is an ecologically relevant functional trait that could be
287 valuable in physiological ecological and evolution [45].

288 That many ecologically important traits, like stomatal ratio, cluster around par-
289 ticular values but not others suggests pervasive constraint on phenotypic evolution.
290 How can we seek a general explanation for this pattern when any particular instance
291 requires specific mechanistic and ecological knowledge about a focal trait? For ex-
292 ample, the emerging evidence from this and other recent studies on stomatal ratio
293 (see especially [23]) is that peaks of high fitness are constrained by a tradeoff be-
294 tween photosynthetic rate and defence against foliar pathogens that preferentially
295 infect though upper stomata. In particular, the cost-benefit model analyzed here
296 predicts that even a small change in the fitness costs or benefits are sufficient to
297 shift fitness peaks into qualitatively different selective regimes. If it is generally true
298 that multimodal traits are associated with rapid regime shifts, then one way forward
299 is to look for signatures of such shifts in closely-related species that sit astride dif-
300 ferent regimes. For example, one signature of regime shifts could be the presence
301 of quantitative trait loci large enough to pass over valleys separating fitness peaks.
302 Consistent with this, [24] recently identified two large effect loci that together are

303 capable of making a hypostomatous leaf amphistomatous, perhaps suggesting that
304 these loci enabled a regime shift. Integrating comparative biology, mechanistic stud-
305 ies of organismal function, and the genetics of adaptation, as this and others studies
306 [46] have begun to do, points to a general approach for evaluating the common fea-
307 tures of macroevolutionary adaptive landscapes and, hence, the role of selection in
308 constraining phenotypic evolution.

309 **Materials and Methods**

310 **Assembling a comparative data set**

311 **Stomatal ratio and leaf thickness** I collected quantitative data on stomatal
312 ratio and leaf thickness from previously published studies (see Text S2 for full list of
313 sources). These data are spread across a large and diverse literature, including func-
314 tional ecology, taxonomy, agriculture, and physiology. Hence, neither a standardized
315 nor exhaustive search was possible. I started by using Web of Knowledge to locate
316 studies that cited seminal papers on the adaptive significance of amphistomy, specif-
317 ically [11] and [12]. Once I found a paper with data, I examined papers that cited
318 those ones. Finally, I found additional data sources in comprehensive reviews of
319 plant anatomy [47, 32, 48]. For all data papers, I recorded the mean leaf thickness,
320 abaxial (lower) and adaxial (upper) stomatal density for each species. Where only
321 ranges were given, I used the midpoint. If the study included a treatment, I col-
322 lected only data from the control treatment. If studies measured both juvenile and
323 adult leaves, I used only adult leaves (no study reported only juvenile leaves). Usu-
324 ally data were given in a table, but occasionally I used ImageJ [49] to extract data

325 from figures or contacted authors for data. I only included data from studies that
326 intentionally examined both surfaces for stomata; I excluded data from studies that
327 described species categorically as “hypostomatous”, or “amphistomatous”, or “hyper-
328 stomatous”. Excluding qualitative data was necessary because there is no standard
329 definition of “amphistomy” – it has sometimes been used to describe species that
330 have approximately equal densities on each side [11] and at other times for species
331 that have any stomata on the both surfaces [16, 15].

332 **Climate and elevation** Based on the *a priori* hypotheses, I extracted data on
333 mean annual precipitation (average 1950 – 2000), elevation (Worldclim [50]), and
334 light environment (average leaf area index between 1982 – 1998 based on remote
335 sensing [51]). For light environment, I used a satellite indicator of leaf area index, the
336 number of leaf layers between the ground and top of the canopy. Lower leaf area index
337 is interpreted as a more open light environment. The strength of these global data
338 sources is that I was able to obtain data for every species from the same dataset. A
339 limitation of these data is that even the highest resolution (≈ 1 km) data might miss
340 important temporal and microsite variation. I discuss these limitations in light of the
341 findings in the Discussion. For climate and elevation, geographic coordinates for each
342 species are needed. For this, I downloaded all georeferenced herbarium specimens
343 for a given species from GBIF (last accessed Jan 15–18, 2015) using the `occ_search`
344 function in `rgbif` [52]. I filtered out or manually edited clearly erroneous locations (e.g.
345 `lat = 0` or `lon = 0` or where `lat` and `lon` were clearly reversed). The mean and median
346 number of GBIF georeferenced occurrences per species was 737 and 194, respectively.
347 I calculated the trimmed-mean (10% trim) mean annual precipitation, elevation, and
348 leaf area index to further remove specimens well outside the species’ range, possibly

349 because they were, say, misidentified, cultivated, or improperly georeferenced.

350 **Growth Form** I partitioned species by growth form into the following categories:
351 trees, small trees/shrubs, shrubs, and herbaceous species (forbs and grasses). Herba-
352 ceous species were further subdivided into annuals, biennials, and perennials. Species
353 that were variable or intermediate (e.g. annual/biennial, annual/perennial, bien-
354 nial/perennial, or annual/biennial/perennial) were classified as ‘biennial’. Subshrubs
355 with some woody growth were lumped with perennials rather than shrubs. Where
356 possible, I obtained growth form data from associated data papers. When this infor-
357 mation was not given, I used regional floras, supplemented by online trait databases
358 such as USDA Plants [53] and Encyclopedia of Life [54]. When these sources were
359 unavailable or ambiguous for a given species, I checked the primary taxonomic liter-
360 ature by searching the species name in Google Scholar.

361 **Taxonomic name resolution** I submitted taxonomic names in the database to
362 the Taxonomic Name Resolution Service (TNRS) [55]. I used names given by TNRS
363 when it returned an accepted name or synonym with overall score greater than
364 0.97 (scores are between 0 to 1). I scrutinized names where TNRS deemed the
365 name illegitimate, gave no opinion, or was otherwise ambiguous. At that point, I
366 consulted additional plant taxonomic repositories: The Plant List [56], International
367 Plant Names Index [57], and the Euro+Med PlantBase [58]. When no accepted
368 names were identified, I used original name given by the authors. For two very
369 recent papers [59, 60], I used the names given by those authors.

370 **Pattern to process: connecting multimodality to phenotypic**
371 **constraint**

372 Comparative methods often infer constraint by comparing the fit of evolutionary
373 process models with and without constraint. Constraint, usually interpreted as a se-
374 lective regime, is typically modelled as an Ornstein-Uhlenbeck process [28, 27, 5], but
375 this model is inappropriate for proportion traits like stomatal ratio. I therefore devel-
376 oped a new evolutionary process model that is analogous to an Ornstein-Uhlenbeck
377 process except that traits are bounded by 0 and 1. A full description of model
378 assumptions and a derivation of the stationary distribution under a given selective
379 regime are available in Text S3. The key result is that a trait evolving under a single
380 selective regime should conform to a Beta distribution at stationarity.

381 Multimodality suggests the presence of multiple selective regimes associated with
382 different modes. I tested for multiple regimes using a conceptually similar but some-
383 what different approach than previous studies. Current methods for inferring mul-
384 tiple selective regimes are in their infancy [27, 29, 30] and cannot yet accommodate
385 Beta-distributed traits because I could not obtain a general solution to the stochastic
386 differential equation in Text S3. Future work is needed to develop numerical meth-
387 ods, such as Approximate Bayesian Computation [61], to integrate the bounded
388 Ornstein-Uhlenbeck process model elaborated here into existing statistical frame-
389 works for multi-regime inference. However, a few lines of reasoning I discuss below
390 indicate that the main conclusions of this study are robust.

391 I used finite mixture models to infer the number of selective regimes shaping
392 stomatal ratio evolution (see [6] for a similar approach). That is, I assume the current

393 distribution of trait values across species can be represented as a mixture of multiple
394 selective regimes at stationarity, each of which is modelled as a Beta-distributed
395 variable. To fit models, I used an expectation-maximization algorithm to find the
396 maximum likelihood mixture model from the data. A complete derivation of the
397 likelihood function and a description of the fitting algorithm are given in Text S4. R
398 code to implement the algorithm is available on Dryad [62]. I selected the best model
399 using the more conservative Bayesian Information Criterion (BIC) to compensate for
400 the fact that I am not accounting for phylogenetic nonindependence in this analysis
401 (see below). I accepted models with an additional selective regime if they decreased
402 BIC by 2 or more. By fitting the data to the stationary distribution, I implicitly
403 assume that evolution is sufficiently rapid to ignore phylogenetic signal. Numerical
404 simulations of the diffusion indicate that the transitory distribution is also Beta (data
405 not shown), meaning that evidence for multiple regimes (i.e. a better fit of a mixture
406 model with multiple Beta components) cannot be an artifact of transitory behaviour
407 within a single regime. I also tested for multiple regimes within families where there
408 was sufficient data ($n \geq 15$). Ten families met this criterion. For each family, I
409 compared the fit of mixtures with $k = 1, 2$, or 3 regimes, accepting models with an
410 additional regime if they decreased BIC by 2 or more. Further, I rejected additional
411 regimes supported by BIC if one of those regimes contained fewer than 3 species
412 (this affected Poaceae and Salicaceae). Although testing for multiple regimes within
413 families using the stationary distribution is an imperfect substitute for fitting the
414 process model to the entire tree, it is nevertheless informative. If multiple regimes
415 are found repeatedly in disparate families, this provides compelling evidence for
416 convergent evolution because of phenotypic constraints imposed by similar adaptive
417 landscapes.

418 **Is selection sufficient to account for multimodality?**

419 In this section, I use theory to ask under what conditions selection can explain the
420 rugged adaptive landscape implied by fitting the evolutionary process model to the
421 data. First, I ask whether a model with simple fitness costs and benefits of upper
422 stomata produces multiple fitness peaks (Text S1 discusses the fitness benefits and
423 costs associated with stomatal ratio). Next, I examine whether such a landscape
424 generates a trait distribution that qualitatively resembles the data, even when the
425 underlying environmental gradients are smooth. I specifically focus on the pattern
426 observed within families, where there was generally one mode of amphistomatous
427 species and another mode of hypostomatous species (hyperstomatous in the case of
428 Poaceae). I also opted to tradeoff the precision of a biophysical diffusion model for a
429 more general, albeit realistic, model with fewer parameters. Hence, the cost-benefit
430 model of stomatal ratio is true to the underlying physics but otherwise not strongly
431 dependent on specific assumptions. Future work will be needed to test if this more
432 general model is consistent with mechanistic biophysical models. The symbols used
433 in the model are summarized in Table 4.

434 I model selection on the logit of stomatal ratio (upper:total), which I denote $S =$
435 $\text{logit}(SR) = \log(SR/(1-SR))$, so that feasible trait variation (SR is constrained from
436 0 to 1) is continuous and unbounded. Fitness as a function of stomatal ratio depends
437 on the difference between the benefits ($f(S)$) minus the costs ($g(S)$). Therefore,
438 fitness as a function of stomatal ratio is:

$$W(S) = 1 + f(S) - g(S) \quad (2)$$

439 Based on biophysical theory [11, 13], I assume that there is an intermediate op-
440 timal stomatal ratio (S_{opt}) at which photosynthetic rate is maximized. Above and
441 below that optimum, photosynthetic rate decreases, which I modelled as a Gaussian
442 function:

$$f(S) = B_{\text{max}} e^{-\frac{(S-S_{\text{opt}})^2}{2\sigma^2}} \quad (3)$$

443 B_{max} defines the maximum fitness when $S = S_{\text{opt}}$. σ^2 acts akin to a shape factor
444 when the function is viewed from a logit scale. When σ^2 is large, the benefit function
445 has an inverted-U shape. There are increasing returns to fitness of the first few upper
446 stomata, but diminishing returns to further increases in SR (Fig. 3A). In contrast,
447 when σ^2 is small, the benefit function is more bell-shaped; the fitness benefit of the
448 first few upper stomata is large, but with diminishing returns (Fig. 3C).

449 I assumed a linear cost (e.g. increased susceptibility to foliar pathogens [23]) for
450 each additional upper stomate. The total cost as a function of stomatal ratio is the
451 product of the total stomatal density, the stomatal ratio (upper:total density), and
452 the cost per upper stomate. I define the slope of the cost function as C_{max} , which is
453 equal to the total stomatal density times the cost per upper stomate:

$$h(SR) = C_{\text{max}} SR \quad (4)$$

454 On a logit scale, the total cost asymptotically approaches C_{max} :

$$g(S) = \frac{C_{\max}}{1 + e^{-S}} \quad (5)$$

455 If more were known about the cost of having upper stomata, a more realistic model
456 could be constructed. Without such knowledge, I believe it is judicious to start with
457 the simplest model that makes few assumptions and therefore could apply to a large
458 number of particular underlying mechanisms. Substituting Eqs 3 and 5 into Eq 2,
459 fitness as a function of S is:

$$W(S) = 1 + B_{\max} e^{-\frac{(S-S_{\text{opt}})^2}{2\sigma^2}} - \frac{C_{\max}}{1 + e^{-S}} \quad (6)$$

460 Note that if the cost function were applied to lower rather than upper stomata, as
461 might be the case for specialized taxa such as aquatic plants, then one could obtain
462 the same results, except that hyper- rather than hypostomy would prevail, as in the
463 Poaceae data. The fitness function is maximized where the marginal benefit of the
464 next upper stomate is equal to the marginal cost:

$$\frac{df(S)}{dS} = \frac{dg(S)}{dS} \quad (7)$$

465 I did not obtain an analytical solution, so instead I used the optim function in R
466 [63] to numerically solve for the stomatal ratio that maximized fitness (S_{fit}) under
467 varying ratios of fitness cost (C_{\max}) to benefit (B_{\max}). I tuned the benefit:cost ratio
468 by fixing B_{\max} to 1 and varying C_{\max} between 0.01 and 100. I also varied the
469 shape factor σ^2 between 0.1 and 10, which appeared to capture the full range of
470 relevant model behaviour. For all numerical solutions, I assumed that the optimal

471 stomatal ratio for photosynthesis was 0.5, hence $S_{\text{opt}} = 0$ on a logit scale. Next,
472 I generated hypothetical trait distributions under a scenario where the benefit:cost
473 ratio varies uniformly from 10^{-2} to 10^2 . I solved for S_{fit} with 10^4 evenly spaced
474 values of $B_{\text{max}} : C_{\text{max}}$ under low, medium, and high values of σ^2 . R code for finding
475 numerical solutions is available from Dryad [62].

476 **Testing adaptive hypotheses for stomatal ratio using phyloge-** 477 **netic regression**

478 I tested for an association between stomatal ratio, leaf thickness, mean annual pre-
479 cipitation, elevation, leaf area index, and growth form using type 2 phylogenetic
480 ANOVA with both categorical (Growth form) and continuous (e.g. leaf thickness)
481 predictor variables. For this analysis I quantified stomatal ratio as $\min(\text{upper den-}$
482 $\text{sity, lower density}) : \max(\text{upper density, lower density})$. In this form, stomatal ratio
483 equals 1 when the densities on each surface are the same, and goes to 0 as the distri-
484 bution become more asymmetrical (hypostomy or hyperstomy). Note that this form
485 differs from what I use in analyzing multimodality because I wanted to specifically
486 test which factors favour the photosynthetically optimal distribution (amphistomy)
487 versus suboptimal distributions (either hypo- or hyperstomy). I accounted for phy-
488 logeny using a Phylomatic [64] megatree for this relatively large and phylogenet-
489 ically extensive dataset. To examine whether results were robust to phylogenetic
490 correction, I analyzed the data using three methods: Brownian motion (high phy-
491 logenetic signal), Pagel's λ (intermediate phylogenetic signal), and no phylogenetic
492 signal (normal ANOVA). For the intermediate signal model, I estimated Pagel's λ
493 using maximum likelihood. Phylogenetic models were fit using phylogenetic least

494 squares in the R package ‘caper’ [65]. The trait dataset and phylogeny used in these
495 analyses are available on Dryad [62].

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501 **Figures**

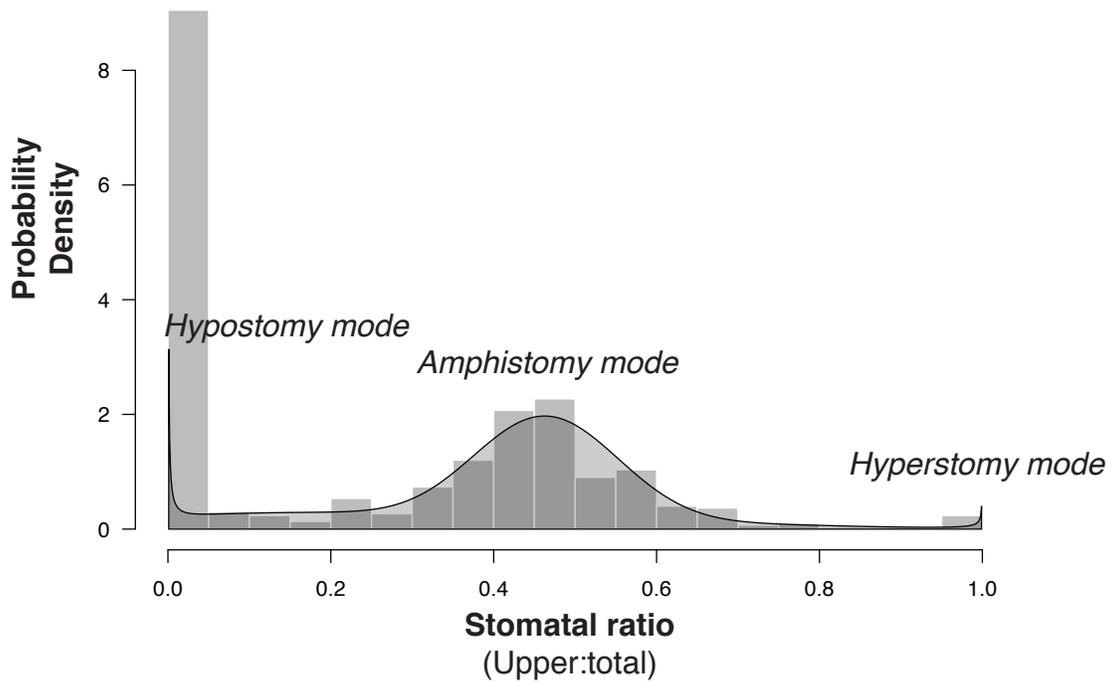


Fig. 1. Stomatal ratio is a multimodal trait. A density histogram of stomatal ratio across 599 species (light grey bars in background) displays three noticeable modes. The plurality of species are completely hypostomatous (all stomata on the lower surface; stomatal ratio equals zero). There is a smaller, broader mode of amphistomatous species (approximately equal density of stomata on upper and lower surfaces; stomatal ratio equals approximately one-half). Finally, there are a small number of hyperstomatous species (all stomata on the upper surface; stomatal ratio equals one). A mixture of selective regimes (shaded grey polygon) manifests these three modes, indicating that they are real features of constrained trait evolution rather than random noise.

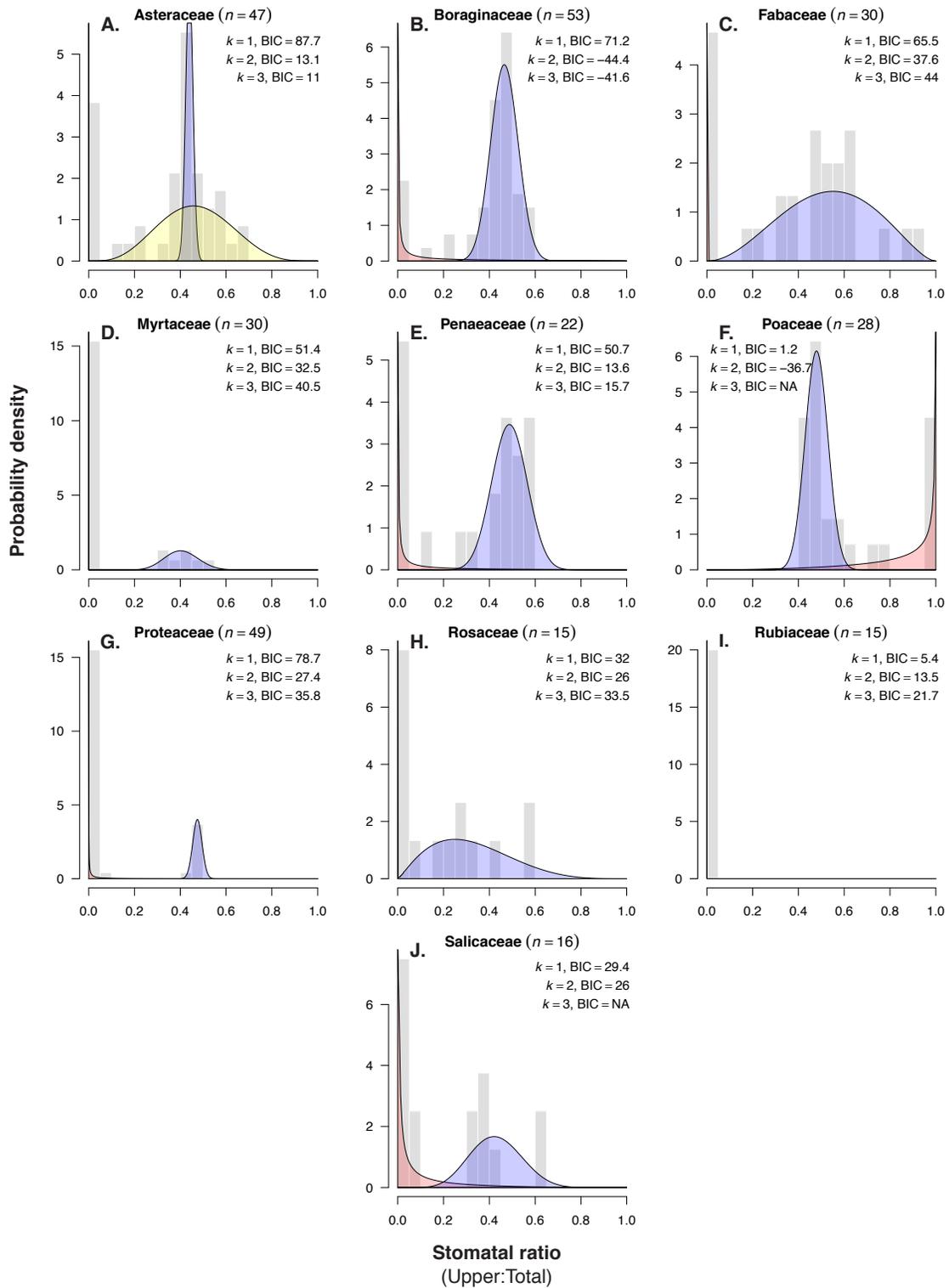


Fig. 2. Repeated evolution of multimodality suggests that the adaptive landscape is conserved across land plants. Shaded polygons of inferred regimes are plotted atop a histogram (grey bar) of stomatal ratio from a given plant family (grey bars). Note that some distributions are very narrow spikes near the origin. The title gives the family name and number of species sampled n from that family. Three regimes were inferred for Asteraceae (panel **A.**); two regimes were inferred for other families except the Rubiaceae (panels **B.-J.**). The number of regimes was inferred from information theoretic comparisons of finite mixture models with Beta-distributed components. The Bayesian Information Criterion (BIC) for models with $k = 1, 2,$ and 3 components is given in the top. I accepted models with additional regimes (higher k) if they decreased BIC by two or more. In Poaceae and Salicaceae, I rejected models with $k = 3$ because some components had very low membership.

Fitness as a function of stomatal ratio

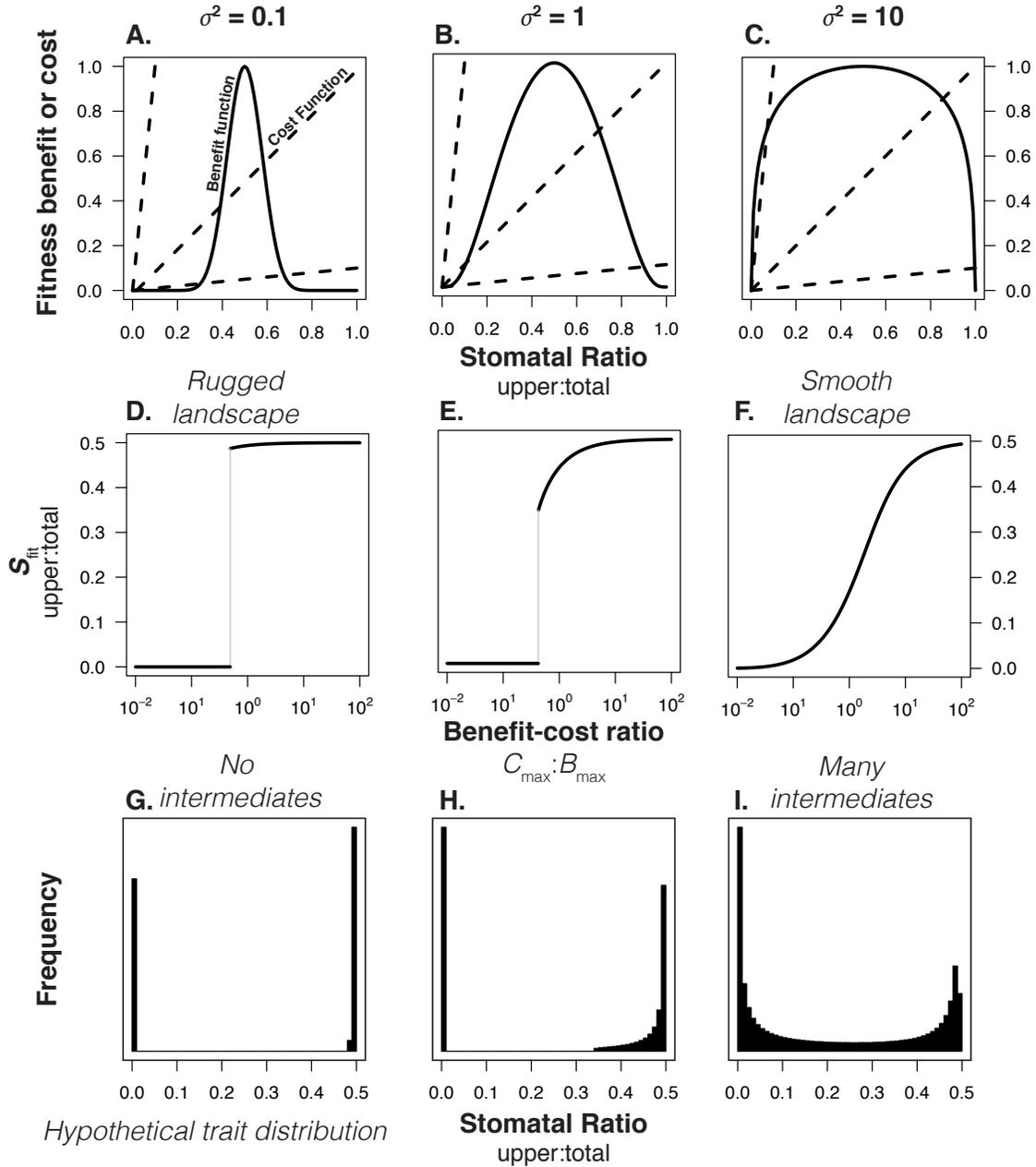


Fig. 3. Selection is sufficient to explain why intermediate phenotypes are universally unfit and the adaptive landscape is rugged. Panels **A.-C.**: In each panel, a benefit function (solid line, see Eq 3) is shown with three different cost functions (dashed line, see Eq 5). In all panels, B_{\max} is fixed at 1 and three slopes of the cost function, C_{\max} are illustrated: 0.1 (shallow slope), 1 (medium slope), and 10 (steep slope). The fitness benefit is always maximized when stomatal ratio is 0.5 (amphistomy), corresponding to $S_{\text{opt}} = 0$ on a logit scale. The shape factor σ^2 changes the benefit function from bell-shaped in **A.** to an inverted-U shape in **C.** Panels **D.-F.** show that the shape of the benefit function affects the topography of the adaptive landscape. Solid lines are the stomatal ratio that optimizes fitness (S_{fit}) as a function of the benefit:cost ratio ($B_{\max} : C_{\max}$). When the benefits are high compared to costs, amphistomy (stomatal ratio = 0.5) is favoured; when the costs are high, hypostomy is favoured (stomatal ratio = 0). However, the transition between these extremes can be abrupt when the landscape is rugged (panel **D.**) or gradual when the landscape is smooth (panel **F.**). The light gray line indicates the range of universally unfit phenotypes. Panels **G.-I.** show hypothetical trait distributions assuming that the benefit:cost ratio varies uniformly from 10^{-2} to 10^2 . Histograms were generated by solving for S_{fit} with 10^4 evenly spaced values of $B_{\max} : C_{\max}$. Note that the trait values range from hypostomatous to amphistomatous (stomatal ratio = 0.5), but a mirror image distribution with hyperstomatous species would be seen if fitness costs accrued to lower stomata.

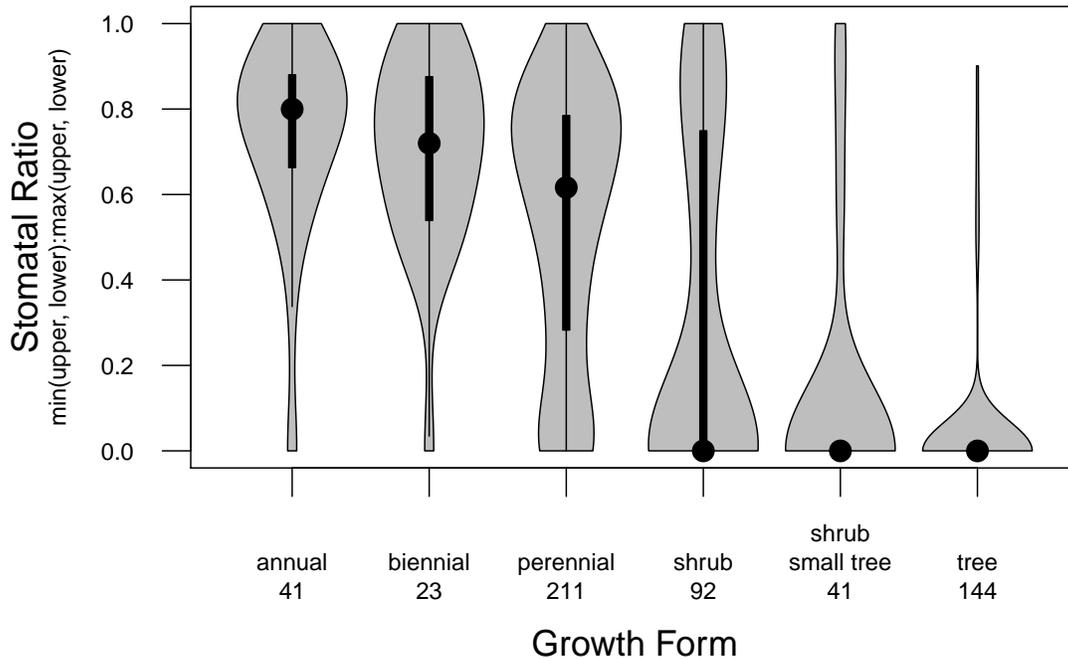


Fig. 4. Association between amphistomy and fast growth forms points to selection on life history shaping stomatal ratio evolution. The violin plot shows of stomatal ratio as a function of growth form across all species in the dataset. The width of the grey polygons indicates the density of data. Length of grey polygon indicate the range of the data; the point indicates the median; the thick lines indicate the 0.25 and 0.75 quantiles. Sample sizes per growth form in the dataset are given below the label.

502 Tables

Table 1. Adaptive hypotheses and predictions for stomatal ratio. The first and second columns indicate the hypothesized ecological factors and the predicted direction of association with amphistomy, respectively. References to key studies are provided, but see Text S1 for additional detail.

Hypothesized factor	Predicted association with amphistomy	References
Leaf thickness	thicker leaves	[11]
Light	greater light intensity	[71, 12, 21]
Precipitation	lower precipitation	[17, 19]
Altitude	higher altitude	[73, 74, 75]
Growth form	herbaceous growth form	[15, 39]

Table 2. Multiple selective regimes are manifest in a multimodal trait distribution. Models with multiple components (k) corresponding to distinct selective regimes under a bounded Ornstein-Uhlenbeck process fit the data significantly better than models with a single regime (lower Bayesian Information Criterion [BIC]). In particular, the model with with three regimes is much more strongly supported than models with one or two regimes (see Fig. S1 for a visual representation of regimes). A mixture of multiple regimes, in turn, gives rise to a multimodal distribution with hypo-, amphi-, and hyperstomatous modes. For a given mixture, each of k regimes is represented as a component i parameterized by the strength of constraint (ϕ_i) around the long-term average (θ_i) and a mixture weight w_i .

k	Parameters	log-likelihood	df	BIC
1	$\phi_1 = 0.4$ $\theta_1 = 0.17$ $w_1 = 1$	-604	2	1220.9
2	$\phi_1 = 0.25$ $\theta_1 = 0.04$ $w_1 = 0.52$ $\phi_2 = 9.98$ $\theta_2 = 0.46$ $w_2 = 0.48$	-252.5	5	536.9
3	$\phi_1 = 0.16$ $\theta_1 = 0.02$ $w_1 = 0.47$ $\phi_2 = 17.24$ $\theta_2 = 0.47$ $w_2 = 0.38$ $\phi_3 = 2.04$ $\theta_3 = 0.35$ $w_3 = 0.16$	-237.7	8	526.6
4	$\phi_1 = 6.99$ $\theta_1 = 0$ $w_1 = 0.44$ $\phi_2 = 1.6$ $\theta_2 = 0.35$ $w_2 = 0.17$ $\phi_3 = 16.85$ $\theta_3 = 0.47$ $w_3 = 0.38$ $\phi_4 = 181.8$ $\theta_4 = 0.99$ $w_4 = 0$	-235.6	11	541.6

Table 3. Growth form, anatomy, and precipitation jointly determine stomatal ratio. Three models with varying levels of phylogenetic signal (Brownian motion [top], Pagel’s λ [middle], and nonphylogenetic [bottom]) identify growth form, leaf thickness, and mean annual precipitation as significantly associated with stomatal ratio.

Stomatal Ratio ~	df	SS	MS	F	<i>P</i>
<i>Brownian Motion</i>					
log(Leaf Thickness)	1	0.017	0.017	20.31	8.08×10^{-6}
Mean Annual Precipitation	1	0.021	0.021	24.11	1.21×10^{-6}
Elevation	1	0	0	0.08	0.78
Leaf Area Index	1	0	0	0.05	0.82
Growth Form	5	0.039	0.008	9.06	2.74×10^{-8}
<i>Pagel’s $\lambda = 0.64$</i>					
log(Leaf Thickness)	1	0.008	0.008	24.38	1.05×10^{-6}
Mean Annual Precipitation	1	0.009	0.009	26.03	4.67×10^{-7}
Elevation	1	0	0	0.26	0.61
Leaf Area Index	1	0	0	0	1
Growth Form	5	0.027	0.005	15.52	2.77×10^{-14}
<i>Nonphylogenetic</i>					
log(Leaf Thickness)	1	2.376	2.376	31.67	2.94×10^{-8}
Mean Annual Precipitation	1	1.711	1.711	22.81	2.31×10^{-6}
Elevation	1	0.009	0.009	0.12	0.72
Leaf Area Index	1	0.031	0.031	0.41	0.52
Growth Form	5	15.897	3.179	42.38	7.36×10^{-37}

Table 4. Glossary of symbols used in the cost-benefit model.

Symbol	Description
SR	Stomatal ratio: ratio of upper to total stomatal density
S	logit of stomatal ratio (SR)
S_{opt}	Stomatal ratio (logit scale) that maximizes fitness benefits
B_{max}	Maximum fitness benefit when $S = S_{\text{opt}}$
σ^2	Shape factor of benefit function
C_{max}	Maximum fitness cost of when all stomata are on the upper side ($SR = 1$)
S_{fit}	Stomatal ratio maximizes fitness (benefits minus costs)

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780 Supporting Information

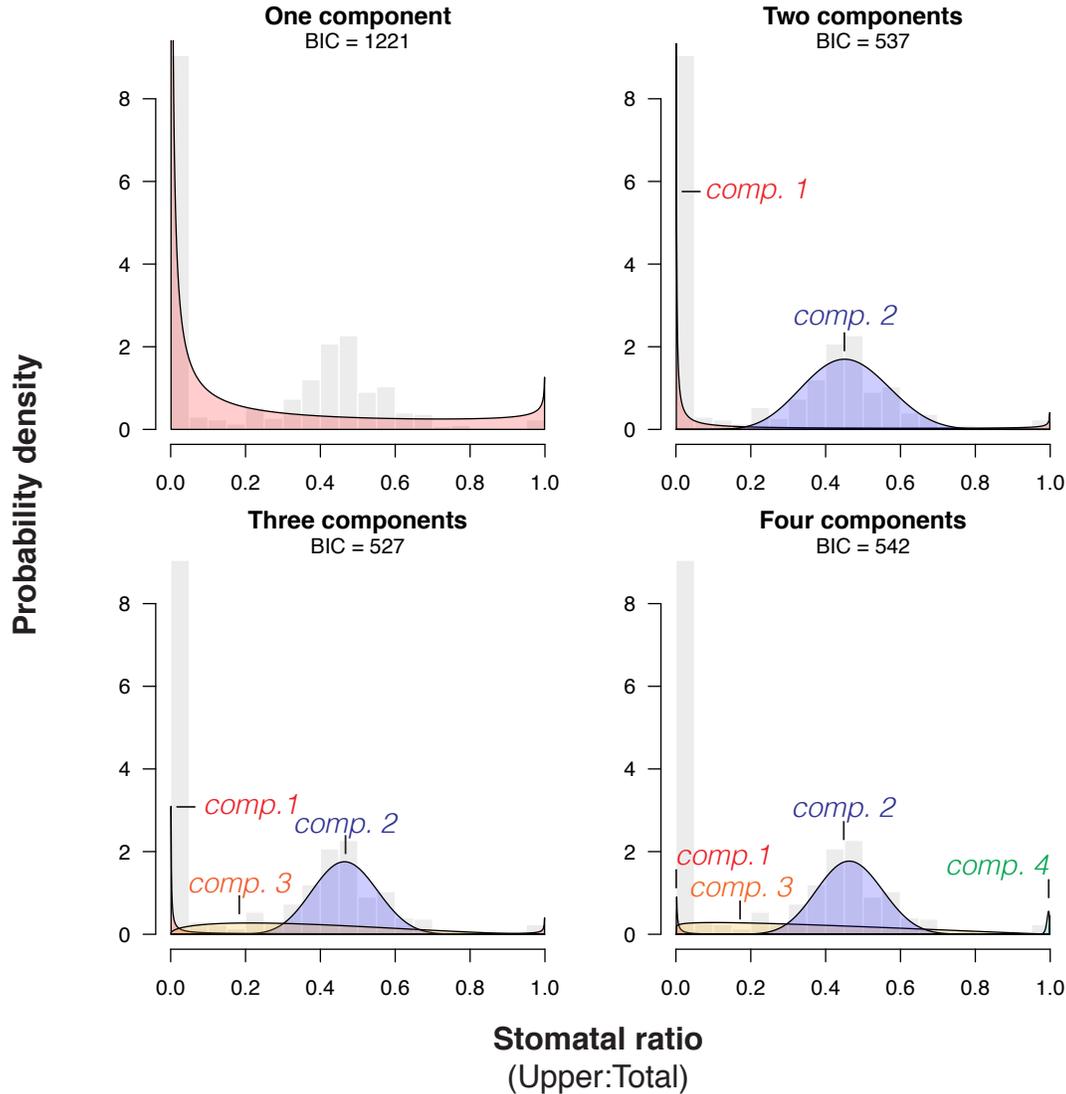


Fig. S1. A multimodal trait distribution implies multiple selective regimes. Model selection using Bayesian Information Criterion (BIC) favoured models that were mixtures of composed multiple selective regimes. **A.** A model with one component was a poor fit because it cannot account for the large peak of amphistomatous species. **B.** A model with two components fit the data much better because it incorporates separate selective regimes for amphistomatous species (blue polygon) and hypo-/hyperstomatous species (red polygon). **C.** An additional selective regime (orange polygon) for species with stomatal ratios between 0 and 0.5 improved model fit, suggesting that intermediate phenotypes are favoured in some circumstances. **D.** Finally, a model with a fourth component (green polygon) did not significantly improve the fit (higher BIC).

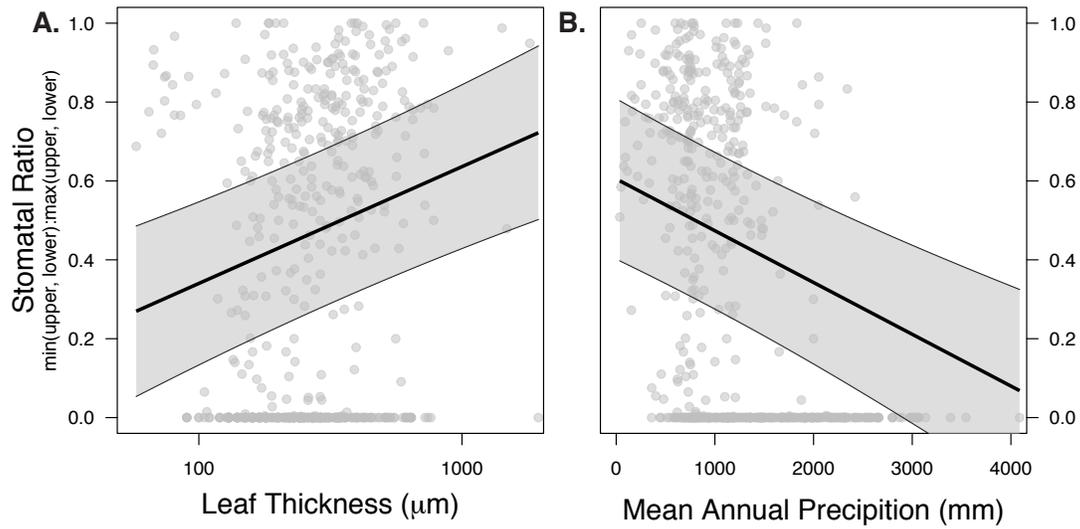


Fig. S2. Amphistomy is weakly associated with thicker leaves and drier habitats. Each point represents a species from the global dataset. The thick line and gray polygon are the median and 95% confidence intervals from the posterior distribution of predicted stomatal ratio as a function of leaf thickness based on phylogenetic regression. The fitted lines and confidence intervals are drawn with growth form set to perennial and other continuous predictor variables set to their median.

781 **Text S1: Hypothesized benefits and costs of amphis-** 782 **tomy**

783 There are at least seven viable, non-mutually exclusive hypotheses for on the adaptive
784 significance of amphistomy, five of which I evaluate here.

785 **H1: Leaf thickness**

786 The most widely cited and frequently tested diffusional limitation hypothesis is that
787 amphistomy is adaptive in thick leaves. Models [11, 13] and experiments [14] demon-
788 strate that the path length from substomatal cavities to chloroplasts can impose a
789 large constraint on photosynthesis, especially when leaf thickness exceeds approx-
790 imately 300 μm . Several studies have found a positive correlation between leaf
791 thickness and amphistomy [11, 20, 66, 67, 68, 59, 44], but the evidence is equiv-
792 ocal [69, 12, 43].

793 **H2: Light**

794 A second hypothesis is that amphistomy is favoured in high light, open environments
795 because CO_2 becomes more limiting at high irradiance. H1 and H2 are difficult to
796 disentangle, and could even reinforce one another, because leaf thickness increases
797 under high irradiance [70]. However, several studies have argued that the light en-
798 vironment, rather than leaf thickness, is the primary factor affecting selection on
799 amphistomy [19, 71, 18, 12, 20, 21].

800 **H3: Precipitation**

801 Wood [17] observed that amphistomy was common in Australian deserts. Although
802 amphistomy is sometimes common in dry environments, most studies conclude that
803 precipitation is indirectly correlated with amphistomy because drier habitats also
804 tend to be more open [19, 21]. Nevertheless, the fact that amphistomy can increase
805 water-use efficiency [11, 72] suggests that it might be favoured in dry habitats, inde-
806 pendent of other factors.

807 **H4: Altitude**

808 Anatomical surveys demonstrate that amphistomy is sometimes more common in
809 high elevation communities compared to nearby low elevation communities [73, 74,
810 75], possibly because lower CO₂ partial pressures place a greater premium on effi-
811 cient diffusion. However, this hypothesis is complicated by the fact that diffusion
812 coefficients are higher at elevation because the air is thinner [76], meaning that CO₂
813 diffusion could actually be less limiting.

814 **H5: Growth form**

815 Independent of leaf anatomy and the abiotic environment, the strength of selection
816 on photosynthetic rate might be stronger among certain growth forms (e.g. forbs
817 vs. trees) because of their different life history strategies. Salisbury (1927) noted
818 qualitatively that herbs tended to amphistomatous, an observation later confirmed
819 by Peat and Fitter (1994). However, other reviews have argued that stomatal ratio

820 is not closely connected with any particular growth form [32, 12].

821 Two hypotheses I have not considered because of methodological limitations are
822 that amphistomy is associated with vertically-oriented, isobilateral leaves [32] and
823 that amphistomy, by doubling the conductive leaf surface area, relieves a constraint
824 the stomatal size-density tradeoff [77, 59]. I did not have sufficient, reliable informa-
825 tion on leaf orientation and guard cell size to evaluate these hypotheses.

826 **Costs of upper stomata**

827 This study reaffirms at a global scale that most species are hypostomatous. The
828 most parsimonious explanation for the preponderance of hypostomy is that there
829 is cost to having stomata on the upper surface of the leaf. A fitness cost associ-
830 ated with increased evaporation [78] cannot explain the dearth of stomata on the
831 upper leaf surface, though this explanation occasionally appears in the literature
832 [79]. In fact, amphistomy is common in some dry habitats [17, 11, 19, 20] and am-
833 phistomatous plants can be functionally hypostomatous when stressed by regulating
834 stomatal aperture differentially on each surface [80, 81, 82, 72]. Although amphis-
835 tomatus plants can be functionally hypostomatous, the reverse is not true. Hence,
836 anatomical amphistomy should be favoured whenever the capacity to be functionally
837 amphistomatous is advantageous.

838 Besides evaporation, several fitness costs have been suggested, including decreased
839 water-use efficiency of amphistomy in large leaves [11], photodamage to guard cell
840 chloroplasts (W.K. Smith, pers. comm.), occlusion of upper stomata by water block-
841 age [83], and increased susceptibility to foliar pathogens [13]. Increased evaporation

842 is an unlikely explanation since so many desert species are anatomically amphis-
843 tomatous (see above), but to my knowledge, most other hypotheses have not been
844 rigorously tested. However, [23] showed that adaxial (upper) stomata pore area, but
845 not abaxial (lower) pore area, was strongly correlated with susceptibility to a rust
846 pathogen. Hence, the pathogen susceptibility hypothesis is best supported by the
847 current data.

848 Text S2: Data Sources

- 849 1. Boeger and Gluzezak 2006 [84]
- 850 2. Brodribb *et al.* 2013 [59]
- 851 3. Camargo and Marengo 2011 [85]
- 852 4. Cooper and Cass 2003 [86]; Cooper *et al.* 2004 [87]
- 853 5. Dickie and Gasson 1999 [88]
- 854 6. Dunbar-Co *et al.* 2009 [89]
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- 856 8. Fahmy *et al.* 2007 [91]
- 857 9. Fontenelle *et al.* 1994 [92]
- 858 10. Giuliani *et al.* 2013 [60]
- 859 11. Holbrook and Putz 1996 [93]
- 860 12. Körner *et al.* 1989 [75]
- 861 13. Lohr 1919 [71]
- 862 14. Loranger and Shipley 2010 [94]
- 863 15. Malaisse and Colonval-Elenkov 1982 [95]
- 864 16. Maricle *et al.* 2009 [31]

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- 866 18. Parkin and Pearson 1903 [96]
- 867 19. Peace and MacDonald 1981 [97]
- 868 20. Rao and Tan 1980 [98]
- 869 21. Reed *et al.* 2000 [99]
- 870 22. Ridge *et al.* 1984 [100]
- 871 23. Selvi and Bigazzi 2001 [101]
- 872 24. Seshavatharam and Srivalli 1989 [102]
- 873 25. Sobrado and Medina 1980 [103]

874 **Text S3: An evolutionary process model for propor-** 875 **tion traits**

876 Making evolutionary sense of a biological pattern requires an underlying process
877 model to provide the theoretical foundation on which data analysis rests. A powerful
878 approach in macroevolution involves modelling trait evolution on adaptive landscapes
879 where the peaks of high fitness evolve with or without constraint [28, 104, 105]. If
880 models with constraint describe the data better than those without, then there is
881 compelling evidence that the adaptive landscape is shaped by some combination of
882 selective, genetic, functional, or developmental constraints. Furthermore, the adap-
883 tive landscape may change under multiple selective regimes, meaning that a trait
884 is best described by a mixture of distributions, each generated under separate se-
885 lective regimes [27, 29, 30]. Current evolutionary process models such as Brownian
886 motion and Ornstein-Uhlenbeck assume that traits follow a Gaussian distribution,
887 but this is clearly inappropriate for traits like stomatal ratio. In this text, I modify
888 previous evolutionary process models to accommodate proportion traits and derive
889 the expected pattern given adaptive landscapes that are constrained versus those
890 that are unconstrained. This model provides a strong theoretical foundation for the
891 model-based statistical inference described in Text S4. A glossary of symbols used
892 in this text are provided in Table S1.

893 In both models with and without constraint, I assume that *total* stomatal density
894 follows a random walk over macroevolutionary time, though the exact process is
895 irrelevant here. Imagine for a set area (A_{leaf}) of leaf (e.g. $1 \mu\text{m}^2$) there are $N_T(t) =$
896 $A_{\text{leaf}}D_T(t) = A_{\text{leaf}}(D_U(t) + D_L(t))$, where $N_T(t)$ is the total number of stomata in

Table S1. Glossary of symbols used in process models of stomatal trait evolution.

Symbol	Description
r	Stomatal ratio: ratio of upper to total stomatal density
N_T, N_U, N_L	Number of stomata in a focal leaf area A_L The total number N_T is the sum of upper N_U and lower N_L stomata
D_T, D_U, D_L	Density of stomata in total, upper, and lower surfaces
A_{leaf}	Focal leaf area
ν	Diffusion coefficient of stomatal ratio
θ	Long-run average stomatal ratio
α	Return rate to long-run average ratio
ϕ	Defined as $\nu\alpha$
$M_{\delta x}$	Drift function of stomatal ratio r in diffusion approximation
$V_{\delta x}$	Diffusion function of stomatal ratio r in diffusion approximation

897 that area at time t . Total stomatal number $N_T(t)$ is the sum of upper ($N_U(t)$)
 898 and lower ($N_L(t)$) stomata. Let $\Delta N_{T,t} = N_T(t+1) - N_T(t)$ be the change in total
 899 stomatal number that must be made up of changes in upper stomata, lower stomata,
 900 or some combination of both. I assume that the contribution to $\Delta N_{T,t}$ from upper
 901 and lower stomata is proportional to their density. For reasons explained below, I
 902 define $\nu = N_T(t+1)$ as the total stomata at time $t+1$. The transition rate u_{ij} from
 903 $N_U = i$ upper stomata at time t to $N_U = j$ upper stomata at time $t+1$ is binomially
 904 distributed with a rate determined by the stomatal ratio r :

$$u_{ij} = \binom{\nu}{j} r^j (1-r)^{\nu-j} \quad j \in \{0, 1, 2, \dots, \nu\} \quad (\text{S1})$$

905 Note that stomatal ratio here is defined as the proportion of upper stomata, $r =$
 906 $N_U/(N_U + N_L) = N_U/N_T = N_U/\nu$. The mean and variance of stomatal ratio in the
 907 next time step is therefore:

$$\mu(r) = \mathbb{E} \left[\frac{N_U}{\nu} \right] = r \quad (\text{S2})$$

$$\sigma^2(r) = \mathbb{E} \left[\left(\frac{N_U}{\nu} \right)^2 \right] - \left(\mathbb{E} \left[\frac{N_U}{\nu} \right] \right)^2 = \frac{r(1-r)}{\nu} \quad (\text{S3})$$

908 In other words, the average stomatal ratio does not change, but the variance
909 increases each time step. When ν is large, the distribution can be approximated with
910 a normal distribution and a diffusion approximation can be used to model the long
911 term evolution of the trait. This diffusion process is analogous to Brownian motion,
912 except that the trait is bounded by 0 and 1. It is also mathematically equivalent
913 to one-locus, two-allele population genetic models of neutral evolution (see [106] for
914 a detailed derivation). I will make reference to results from this literature without
915 rigorously deriving them here. In particular, it has been shown that the stationary
916 distribution of the diffusion is:

$$f(r) = \frac{e^{A(r)} (c_1 \int e^{-A(r)} dr + c_2)}{V_{\delta x}} \quad (\text{S4})$$

917 where

$$A(r) = \int \frac{2M_{\delta x}}{V_{\delta x}} dr \quad (\text{S5})$$

$$M_{\delta x} = 0 \quad (\text{S6})$$

$$V_{\delta x} = \frac{r(1-r)}{\nu} \quad (\text{S7})$$

918 and the time scale is in units of ν^{-1} . Thus, ν can be interpreted as a diffusion
919 coefficient without necessarily specifying a genetic or developmental mechanism that
920 governs the amount of variance in stomatal ratio from one time to the next. Solving
921 for $f(r)$ without selection on stomatal ratio yields:

$$f(r) = \frac{6}{r(1-r)} \quad (\text{S8})$$

922 Thus, without selection on stomatal ratio, most species should be hypo- or hyper-
923 stomatous (Fig. S3). Next, I modify the model to include stabilizing selection around
924 a long-run average θ , which may be interpreted as a peak in the adaptive landscape
925 under a single selective regime. This process model is analogous to an Ornstein-
926 Uhlenbeck process for a bounded trait. I again use the diffusion approximation, but
927 this time the drift and diffusion coefficients are:

$$M_{\delta x} = \alpha(\theta - r) \quad (\text{S9})$$

$$V_{\delta x} = \frac{r(1-r)}{\nu} \quad (\text{S10})$$

928 α is the return rate to θ . Greater values of α constrain trait variation more tightly
929 around θ . With these coefficients and setting the first constant of integration c_1 to 0
930 yields:

$$f(r) = c_2 \nu r^{2\alpha\nu\theta-1} (1-r)^{2\alpha\nu(1-\theta)-1} \quad (\text{S11})$$

931 where:

$$c_2 = 1 / \int_0^1 \nu r^{2\alpha\nu\theta-1} (1-r)^{2\alpha\nu(1-\theta)-1} dr \quad (\text{S12})$$

$$= \frac{1}{\nu B(2\alpha\nu\theta, 2\alpha\nu(1-\theta))} \quad (\text{S13})$$

932 $B(\cdot)$ is the beta function. Setting c_1 to 0 can be justified by recognizing that the
933 distribution should be symmetrical ($x = 1 - x$) when $\theta = 0.5$, which only occurs if
934 $c_1 = 0$ (S.P. Otto pers. comm.). Further, I confirmed the accuracy of the analytically-
935 derived stationary distribution using stochastic simulations (data not shown).

936 Defining $\phi = \alpha\nu$, the stationary distribution simplifies somewhat to:

$$f(r) = \frac{r^{2\phi\theta-1} (1-r)^{2\phi(1-\theta)-1}}{B(2\phi\theta, 2\phi(1-\theta))} \quad (\text{S14})$$

937 This is the Beta(α, β) distribution with $\alpha = 2\phi\theta$ and $\beta = 2\phi(1-\theta)$. Note that,
938 following standard notation, α here refers to the first shape parameter of the Beta
939 distribution, not the constraint factor of the evolutionary process model. This result
940 means that the well-known statistical properties of the Beta distribution can be
941 leveraged to understand the stationary distribution of a proportion trait under a
942 constrained adaptive landscape. For example, the Beta distribution takes on a variety
943 of shapes that begin to resemble the distribution of proportional traits like stomatal
944 ratio (Fig. S4). Hence, the evolutionary process model developed here provides

945 a strong theoretical justification for fitting the stomatal ratio data to a mixture of
946 Beta distributions in order to infer the selective regimes shaping this trait across plant
947 species. Although I have derived the model with stomatal ratio in mind, it should
948 be applicable to wide variety of proportional traits evolving under a constrained
949 adaptive landscape.

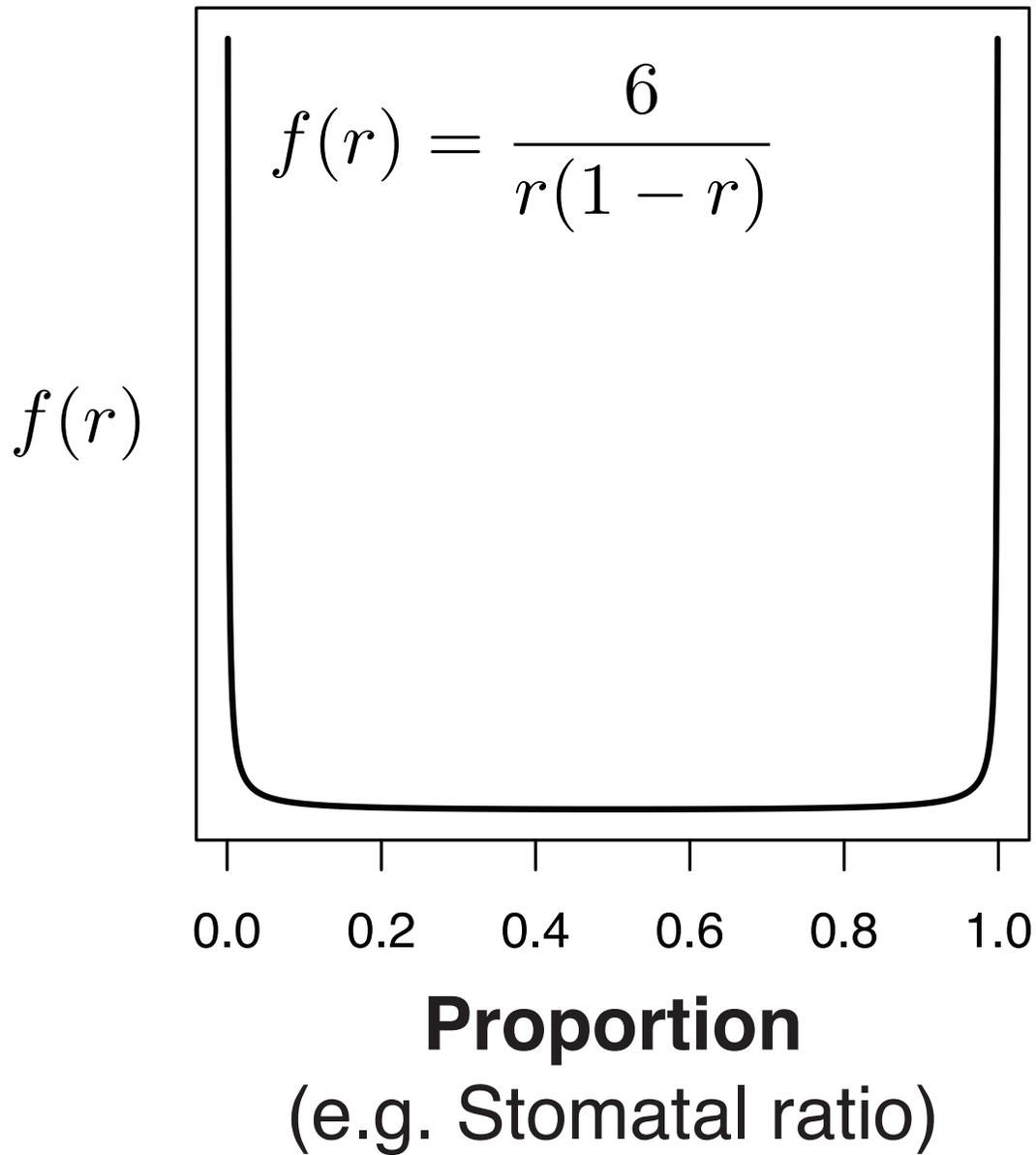


Fig. S3. Without constraint, a proportion trait like stomatal ratio (r) will evolve toward a distribution in which most species are 0 or 1.

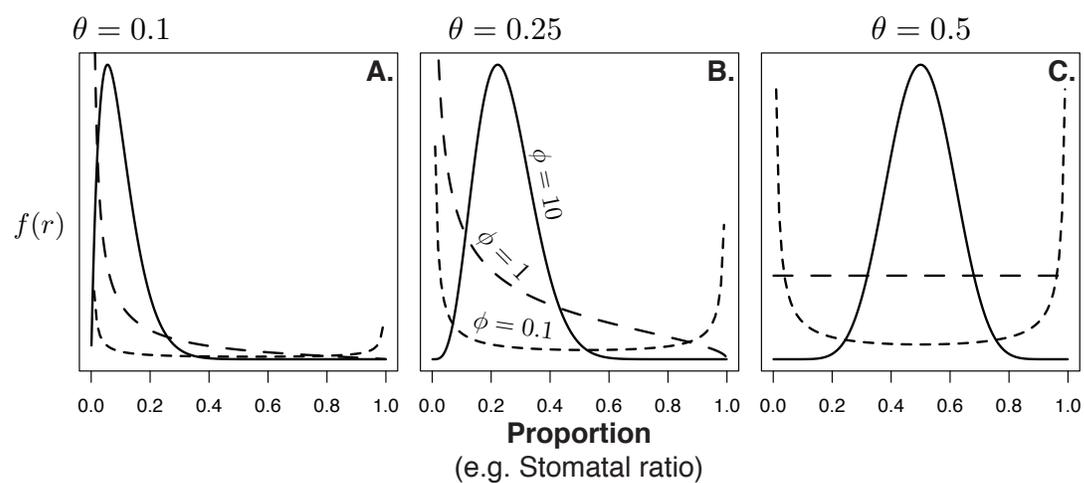


Fig. S4. A proportion trait like stomatal ratio evolving under a constrained adaptive landscape is Beta distributed. The Beta distribution can take on a wide variety of shapes depends on the long-run average θ and the levels of constraint ϕ (greater ϕ equals greater constraint).

950 **Text S4: Fitting evolutionary process to pattern us-**
951 **ing finite mixture models estimated with maximum**
952 **likelihood**

953 In this paper, I infer the number of selective regimes acting on stomatal ratio by
954 fitting a mixture of stationary distributions derived from the process model above to
955 the data. In this section I derive the likelihood functions and describe an expectation-
956 maximization algorithm to find the maximum likelihood mixture model given the
957 data. R code to implement these methods is available on Dryad [62]. In general,
958 finite mixture distributions are the summation of $k \geq 2$ mixture components (i.e.
959 probability distributions) with density $f_i(x)$ and mixture weight w_i :

$$g(x; k) = \sum_{i=1}^k w_i f_i(x) \quad (\text{S15})$$

960 Here the i -th mixture component has a probability density $f_i(x)$ given by the
961 stationary distribution in Eq S14 with parameters θ_i, ϕ_i . The likelihood of a mixture
962 distribution given k mixture components and a data vector \mathbf{x} with sample size n is
963 the weighted sum of the likelihoods of each component:

$$\mathcal{L}(\mathbf{w}, \boldsymbol{\phi}, \boldsymbol{\theta}; \mathbf{x}, k) = \sum_{i=1}^k w_i \mathcal{L}_i(\phi_i, \theta_i; \mathbf{x}) \quad (\text{S16})$$

964 The parameter vectors \mathbf{w} , $\boldsymbol{\phi}$, and $\boldsymbol{\theta}$ are defined as:

$$\mathbf{w} := \{w_1, \dots, w_k\} \quad (\text{S17})$$

$$\boldsymbol{\phi} := \{\phi_1, \dots, \phi_k\} \quad (\text{S18})$$

$$\boldsymbol{\theta} := \{\theta_1, \dots, \theta_k\} \quad (\text{S19})$$

965 For the i -th component, the likelihood of parameters ϕ_i and θ_i given the data is
966 the product of the probability densities of each datum (x_1, x_2, \dots, x_n) :

$$\mathcal{L}_i(\phi_i, \theta_i; \mathbf{x}) = \prod_{j=1}^n f_i(x_j; \phi_i, \theta_i) \quad (\text{S20})$$

967 To obtain reasonable fits, I found it necessary to modify the likelihood to incorpo-
968 rate left- and right-censored data. This is because the stomatal ratio dataset contains
969 many 0's (all stomata are on the lower surface of the leaf) and 1's (all stomata on the
970 upper surface). Under most parameterizations of the Beta distribution, the proba-
971 bility density of 0 and 1 is ∞ or 0. I left- and right-censored the data at $x_l = 0.001$
972 and $x_r = 0.999$ as these were very close to the lowest and highest values reported
973 in the dataset (except 0 and 1), respectively. This means that any datum reported
974 as 0 was statistically interpreted as falling anywhere between 0 and 0.001. Likewise,
975 a datum reported as 1 was assumed to fall between 0.999 and 1. A reasonable in-
976 terpretation is that a stomatal ratio so close to 0 or 1 would be practically difficult
977 to measure. Biologically, a stomatal ratio less than 0.001 or greater than 0.999 are

978 indistinguishable from 0 and 1. With censoring, the likelihood of the i -th component
979 becomes:

$$\mathcal{L}_i(\phi_i, \theta_i; \mathbf{x}) = \prod_{j=1}^n f(x_j; \phi_i, \theta_i)^{I_l(x_j)I_r(x_j)} F(x_l; \phi_i, \theta_i)^{1-I_l(x_j)} (1 - F(x_r; \phi_i, \theta_i))^{1-I_r(x_j)} \quad (\text{S21})$$

980 $F(x; \phi_i, \theta_i)$ is the cumulative density function of the Beta distribution; $I_l(x)$ and
981 $I_r(x)$ are indicator functions:

$$I_l(x) = \begin{cases} 0 & \text{if } x = x_l \\ 1 & \text{if } x \neq x_l \end{cases} \quad (\text{S22})$$

$$I_r(x) = \begin{cases} 0 & \text{if } x = x_r \\ 1 & \text{if } x \neq x_r \end{cases} \quad (\text{S23})$$

982 To find the maximum likelihood mixture distribution, I used an expectation-
983 maximization (EM) algorithm similar to [107]. EM algorithms are particularly well-
984 suited to fitting mixture distributions. Here, I describe the initialization, expectation
985 (E-step), and maximization (M-step) procedure.

986 Initialization

987 The data were divided into k evenly-sized components. For example, if $k = 2$,
988 data below the median were assigned to component 1; data above the median were

989 assigned to component 2. For each component, the initial weight was therefore
990 $w_{i,\text{init}} = 1/k$. Within each component, I used the `optim` function in R to estimate the
991 maximum likelihood parameters ($\hat{\phi}_i^{(\text{init})}$ and $\hat{\theta}_i^{(\text{init})}$) of a Beta distribution. Note that I
992 am using parenthetical superscript to indicate the iteration of the algorithm, starting
993 with the initial parameterization, followed by $t = 1, 2, 3, \dots$ until the likelihood
994 converges. The initial parameter vectors are therefore:

$$\mathbf{w}^{(\text{init})} := \{1/k, \dots, 1/k\} \quad (\text{S24})$$

$$\boldsymbol{\phi}^{(\text{init})} := \{\hat{\phi}_1^{(\text{init})}, \dots, \hat{\phi}_k^{(\text{init})}\} \quad (\text{S25})$$

$$\boldsymbol{\theta}^{(\text{init})} := \{\hat{\theta}_1^{(\text{init})}, \dots, \hat{\theta}_k^{(\text{init})}\} \quad (\text{S26})$$

995 Expectation

996 In the E-step, the expected likelihood is calculated under the parameters estimated
997 from the previous iteration. The mixture weights are then updated and carried
998 forward to the M-step. For the first iteration following initialization, the mixture
999 weights $\mathbf{w}^{(1)}$ conditional on the initial parameterization are:

$$w_i^{(1)} = \frac{\sum_{j=1}^N y_{ij}^{(\text{init})}}{n} \quad (\text{S27})$$

1000 where $y_{ij}^{(\text{init})}$ is the probability that x_j belongs to component i given initial param-
1001 eters:

$$y_{ij}^{(\text{init})} = \frac{w_i^{(\text{init})} f(x_j; \hat{\phi}_i^{(\text{init})}, \hat{\theta}_i^{(\text{init})})}{g(x_j; k, \mathbf{w}^{(\text{init})}, \boldsymbol{\phi}^{(\text{init})}, \boldsymbol{\theta}^{(\text{init})})} \quad (\text{S28})$$

In subsequent iterations, the equations are similarly:

$$w_i^{(t+1)} = \frac{\sum_{j=1}^N y_{ij}^{(t)}}{N} \quad (\text{S29})$$

$$y_{ij}^{(t)} = \frac{w_i^{(t)} f(x_j, \phi_i^{(t)}, \theta_i^{(t)})}{g(x_j; k, \mathbf{w}^{(t)}, \boldsymbol{\phi}^{(t)}, \boldsymbol{\theta}^{(t)})} \quad (\text{S30})$$

1002 Maximization

1003 During the M-step, estimates of $\boldsymbol{\phi}$ and $\boldsymbol{\theta}$ are updated using maximum likelihood
1004 conditional on mixture weights calculated in the E-step:

$$\{\boldsymbol{\phi}^{(t+1)}, \boldsymbol{\theta}^{(t+1)}\} = \arg \max_{\boldsymbol{\phi}, \boldsymbol{\theta}} \mathcal{L}(\boldsymbol{\phi}, \boldsymbol{\theta}; \mathbf{x}, k, \mathbf{w}^{(t)}) \quad (\text{S31})$$

1005 I used the `optim` function in R to find the parameters that maximized the likelihood
1006 function. After the M-step, the next iteration begins at the E-step and continues
1007 until the likelihood converges to a stable value. As with other hill-climbing likelihood
1008 searches, EM does not guarantee convergence at the maximum likelihood. With the
1009 stomatal ratio data, I found that multiple initialization procedures yielded the same
1010 final parameter estimates, suggesting that the algorithm was successfully converging
1011 on the maximum likelihood solution.