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3 Heritability and relationship of oxytocin receptor gene variants with social behavior and
4 central oxytocin in colony-reared adult female rhesus macaques

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24

26 **Abstract**

27 The genetic contributions to sociality are an important research focus for understanding
28 individual variation in social function and risk of social deficits in neurodevelopmental disorders
29 (e.g. autism). The neuropeptide oxytocin (OXT) and its receptor, OXTR, influence social
30 behavior across species. In humans and animals, common variants within the OXTR gene
31 (*OXTR*) have been associated with varying socio-behavioral traits. However, the reported
32 magnitude of influence of individual variants on complex behavior has been inconsistent.
33 Compared to human studies, non-human primate (NHP) studies in controlled environments
34 have the potential to result in robust effects detectable in relatively small samples. Here we
35 estimate heritability of social behavior and central OXT concentrations in 214 socially-housed
36 adult female rhesus macaques, a species sharing high similarity with humans in genetics,
37 physiology, brain and social complexity. We present a bioinformatically-informed approach for
38 identifying single nucleotide polymorphisms (SNPs) with likely biological relevance. We tested
39 13 common SNPs in regulatory and coding regions of *OXTR* for associations with behavior (pro-
40 social, anxiety-like, and aggressive) and OXT concentration in cerebrospinal fluid (CSF). We
41 found moderate rates of heritability for both social behavior and CSF OXT concentrations. No
42 tested SNPs showed significant associations with behaviors or CSF in this sample. Associations
43 between OXT CSF and social behavior were not significant either. SNP effect sizes were
44 generally comparable to those reported in human studies of complex traits. While environmental
45 control and a socio-biological similarity with humans is an advantage of rhesus models for
46 detecting smaller genetic effects, it is insufficient to obviate large sample sizes necessary for
47 appropriate statistical power.

48

49 **Introduction**

50 Oxytocin is an evolutionarily conserved neuropeptide involved in orchestrating
51 reproductive, maternal, and social behaviors across species [1]. Individual variability in these
52 traits is attributable to genetic variation, apart from the role of experience. As such, variants
53 such as single nucleotide polymorphisms (SNPs) within the OXT receptor gene (*OXTR*) may
54 contribute to the diversity of complex social repertoires. Genetic association studies suggest
55 that variation in *OXTR* may explain part of the variance in social phenotypes including pair-
56 bonding behavior, social recognition, prosocial temperament, and sensitivity to social stressors
57 [1–3]. Multiple *OXTR* SNPs have also been linked to social impairments in individuals with
58 autism [4].

59
60 Despite reports linking human *OXTR* variants to social phenotypes, other studies,
61 including a meta-analysis [5], have failed to find effects of consistent magnitude and direction of
62 these variants on social domains [see 6 for review]. Inconsistent results may be related to
63 biases which disproportionately suppress non-significant results from publication (e.g.
64 publication bias, selective reporting), and inflated effect sizes due to low statistical power [7].

65
66 The use of animal models, however, has allowed the underlying mechanisms for SNP-
67 behavior associations to be probed in the brain, including assessing the functional effects of
68 differential *OXTR* gene expression caused by genetic variants in brain tissue. For example, in
69 the prairie vole rodent model, a single *Oxtr* SNP predicted more than 70% of the variability in
70 expression in the nucleus accumbens, a region critical for social reward [8]. This suggests that
71 individual variants have the potential to profoundly impact brain phenotypes which can mediate
72 downstream behavior differences.

73

74 Non-human primates (NHPs), such as the rhesus macaque, have been widely used to
75 study social behavior. Both rhesus and humans display rich behavioral repertoires, complex
76 social hierarchies, and are very similar in their physiology and neuroanatomy. Additionally,
77 rhesus have frequently been used to investigate the role of OXT on sociality [9], and as such, an
78 understanding of the genetic contributions to the oxytocinergic system in this NHP model is
79 valuable for translation to humans.

80

81 While human genetic association studies necessitate extremely large samples to detect
82 small effects reliably among the multitude of factors influencing complex traits, it is possible that
83 NHP studies conducted in experimentally controlled, semi-naturalistic settings with known
84 pedigrees can detect larger effects due to the reduction in environmental noise (e.g. standard
85 diets and perinatal environments). The goal of the present study was to use an informed
86 approach to examine the associations of novel rhesus *OXTR* (*rhOXTR*) SNPs with social
87 behavior (pro-social, anxiety-like, and aggressive) and OXT in cerebrospinal fluid (CSF) in 214
88 adult female rhesus macaques housed in large social groups. We also report estimates of
89 heritability and examine the effects of OXT CSF on each behavior.

90

91 **Materials and methods**

92 **Subjects and housing**

93 Subjects were 214 adult female rhesus monkeys (4-24 years in age; median: 7,
94 interquartile range: 6-11) of known pedigrees living in complex social groups at the Yerkes
95 National Primate Research Center (YNPRC) Field Station (Lawrenceville, GA) as described in
96 [10]. Subjects came from five social groups consisting of 28-94 adult females, their kin, and 2-10
97 adult males. Animals were selected from matrilineal lines across all social status ranks (high, medium

98 and low) and with varying degrees of relatedness for heritability analyses. Animals were housed
99 in outdoor enclosures (3/4 to 1 acre areas) with access to climate-controlled indoor facilities and
100 provided a standard commercial low-fat, high-fiber diet (Purina Mills International, LabDiets, St.
101 Louis, MO) and water *ad libitum*, supplemented with seasonal fruits or vegetables twice per day.
102 All procedures complied with the Animal Welfare Act and U.S. DHHS “Guide for the Care and
103 Use of Laboratory Animals” and were approved by the Emory IACUC.

104

105 Behavioral data collection

106 Focal observations were collected in real-time following published protocols [10,11]
107 based on established ethograms (Altmann, 1962). An average behavioral observation of 80
108 min/animal was collected during the mating season to reduce seasonal variability in behavior.
109 We focused on representative behaviors: pro-social (percent of time spent in proximity to other
110 adult females), anxiety-like (frequency of self-scratches) [12], and aggressive (non-contact,
111 subject-initiated) behaviors (Table 1).

112

113 **Table 1. Specific behaviors analyzed for this study**

Variable	Definition	Data Collected
<i>Pro-social Behavior</i>		
Proximity to other adult females ^a	Subject is within 1 foot of one or more other adult females (including physical contact)	Duration
<i>Aggressive Behavior</i>		
Non-contact aggression	Counts of agonistic chases, open-mouth threats, barks, and lunges of/by another animal	Frequency
<i>Anxiety-like / Solitary Behavior</i>		
Anxiety/self-directed	Counts of self-scratches ^b	Frequency

114 ^a Given the matrilineal structure of rhesus troops [19], proximity to adult females (as opposed to other
115 members in the group) comprise the most frequent and relevant social interaction observed

116 ^b In order for the same frequency behavior to be scored a second time, at least 3 seconds had to have
117 passed from the first instance of the behavior

118

119 **CSF and blood samples**

120 Animals were habituated to experimental procedures to facilitate blood and CSF
121 collection using methods minimizing arousal [10]. Each subject was accessed once, shortly after
122 sunrise and during mating season, but never on the same day as the behavioral data collection.
123 Animals were anesthetized with Telazol (3 mg/kg, IM). CSF samples were collected in a subset
124 of the subjects (n=166) to examine central OXT concentrations. CSF samples were collected (2
125 ml/subject) from the *cisterna magna* by gravity through a 22 G needle and placed immediately
126 on dry ice. Two 3 ml blood samples were collected in EDTA tubes for DNA extraction and
127 immediately placed on ice. Samples were stored at -80°C until time of processing. CSF samples
128 were processed by the YNPRC Biomarkers Core Laboratory using commercially available
129 ELISA kits produced by Assay Designs (Ann Arbor, MI), following manufacturer's
130 recommendations. Sensitivity of the OXT assay was 15.6 pg/ml and the inter- and intra- assay
131 CVs were 7.48% and 10.2%, respectively.

132

133 **Covariates**

134 Social rank and age (and no other variables) were included as covariates for all models.
135 OXT concentrations were assayed in three batches; observations were mean-centered per
136 batch to control for inter-batch variation.

137

138 **SNP selection**

139 SNP discovery efforts were completed in a separate group of rhesus and focused on 5'
140 regulatory and coding regions of *rhOXTR*. Within these regions, candidate SNPs were identified
141 in positions analogous to those in humans that have been cited as being involved in *OXTR*

142 expression [13] or suggested to predict individual differences in behavior and disease
143 vulnerability. While SNPs are not conserved across species, comparably located SNPs in
144 macaques may confer functionally similar effects and therefore provide translational potential.
145 Additional SNPs in highly conserved areas within the region of interest were also included as
146 candidates. Candidates were narrowed down based on their presence with sufficient minor
147 allele frequency in that separate group of macaques, comprising representatives from multiple
148 genetically-distinct populations. This resulted in 13 loci to be genotyped for this study's sample
149 (Fig 1).

150
151 **Fig 1. Schematic of *OXTR* SNPs probed in this study.** Untranslated portions of the exons are
152 indicated by the white bars. Genomic coordinates correspond to chromosome 3 of the MacaM
153 reference genome (SNP 1) 140481032, (SNP 2) 140497572, (SNP 3) 140497358, (SNP 4)
154 140497003, (SNP 5) 140496605, (SNP 6) 140496348, (SNP 7) 140496344, (SNP 8)
155 140496196, (SNP 9) 140495645, (SNP 10) 140495542, (SNP 11) 140495303, (SNP 12)
156 140495244 G/A, (SNP 13) 140495203.

157

158 **Genotyping**

159 Three different genotyping techniques were used. Archived next-generation sequencing
160 data targeting *OXTR* exons was available for loci 4-13. Libraries were generated using the
161 Illumina NexteraXT DNA kit and sequenced on an Illumina HiSeq1000. The remaining markers
162 were genotyped using TaqMan or Sanger Sequencing. Cycle sequencing was performed using
163 the Big Dye Terminator, version 3.1, reaction in 96-well optical plates (Applied Biosystems,
164 Foster City, California). Variants were detected by visualization of electropherograms generated
165 by ABI Sequencing Analysis software. Relevant assay and primer information are in
166 Supplemental Table S1.

167

168 **Statistical analysis**

169 All statistical analyses were performed in R version 3.4.1 (R Project for Statistical
170 Computing, Vienna, Austria). Associations of SNPs with behavioral and CSF measures, as well
171 as heritability, were examined using the "animal model" with the "MCMCglmm" package [14].
172 Narrow-sense heritability was calculated as the proportion of variance explained by additive
173 genetic factors (via the pedigree) out of all phenotypic variance. SNPs and covariates were
174 included as fixed effects, and relatedness was accounted for by including pedigree as a random
175 effect. To account for differences in total time observed, observation length was included as an
176 offset for models including frequency behaviors (i.e. raw counts). The prior distribution for
177 additive genetic variance (σ^2_A) was defined by commonly used non-informative parameters for
178 the inverse-gamma distribution, IG(0.001, 0.001). Models were run with a minimum of 5 million
179 iterations, a burn-in of 5,000, and thinning interval of 1,000 until adequate mixing was ensured.

180

181 A Gaussian distribution was specified for pro-social behavior and log-transformed OXT
182 CSF measures. For anxiety-like behavior, a Poisson specification was used. Heritability was not
183 estimable for aggression, even after trying multiple distributions and stronger priors on σ^2_A . This
184 was most likely due to a combination of low occurrence of this behavior (i.e. low information)
185 and a small sample size [15]. Because controlling for relatedness was essential for all models,
186 this outcome was excluded from subsequent analyses.

187

188 In post-hoc analyses, a likelihood-ratio test was used to compare whether inclusion of all
189 SNPs (versus none) significantly improved model fit. To do so, frequentist versions of each

190 model were run using R packages “pedigreemm” and “regress” to extract maximum log-
191 likelihood values.

192

193 **Results**

194 **Heritability estimates**

195 We report moderate heritability for pro-social behavior ($h^2 = 0.312$), anxiety-like behavior
196 ($h^2 = 0.283$), and OXT CSF ($h^2 = 0.183$), though all 95% credible intervals were wide (Table 2).

197

198

199 **Table 2. Heritability and SNP estimates**

200

Narrow sense heritability (h^2)	Proximity to Adult Females (Pro-social)			Self-Scratches (Anxiety-like)			OXT in CSF		
	Point Estimate	Lower 95% CI	Upper 95% CI	Point Estimate	Lower 95% CI	Upper 95% CI	Point Estimate	Lower 95% CI	Upper 95% CI
Without fixed effects	0.312	0.046	0.591	0.213	4.98×10^{-4}	0.670	0.183	0.002	0.478
With fixed effects	0.306	0.038	0.587	0.297	3.95×10^{-4}	0.679	0.193	7.93×10^{-4}	0.500
SNPs	β	Lower 95% CI	Upper 95% CI	β	Lower 95% CI	Upper 95% CI	β	Lower 95% CI	Upper 95% CI
1) 140481032 G/A	-0.011	-0.043	0.023	0.003	-0.141	0.148	0.039	-0.056	0.136
2) 140497572 G/C	0.003	-0.028	0.037	0.058	-0.089	0.224	0.023	-0.072	0.123
3) 140497358 G/A	0.003	-0.028	0.038	0.079	-0.076	0.238	0.025	-0.074	0.122
4) 140497003 G/T	-0.037	-0.132	0.063	-0.481	-0.912	0.015	0.123	-0.177	0.431
5) 140496605 T/C	-0.006	-0.038	0.027	0.043	-0.126	0.19	0.015	-0.073	0.108
6) 140496348 A/G	-0.004	-0.038	0.027	0.031	-0.111	0.188	0.019	-0.073	0.108
7) 140496344 A/C	-0.005	-0.038	0.028	0.032	-0.126	0.181	0.018	-0.076	0.115
8) 140496196 G/T	0.014	-0.026	0.049	0.043	-0.128	0.22	-0.032	-0.131	0.072
9) 140495645 C/G	-0.007	-0.036	0.021	0.107	-0.037	0.242	0.038	-0.048	0.131
10) 140495542 A/T	-0.005	-0.096	0.087	0.056	-0.386	0.526	0.126	-0.117	0.387
11) 140495303 C/T	0.004	-0.028	0.034	-0.064	-0.21	0.084	0.06	-0.035	0.157
12) 140495244 G/A	-0.005	-0.052	0.037	-0.176	-0.381	0.04	0.044	-0.104	0.17
13) 140495203 T/G	-0.001	-0.033	0.03	-0.076	-0.229	0.072	0.068	-0.027	0.154
	β	Lower 95% CI	Upper 95% CI	β	Lower 95% CI	Upper 95% CI			
OXT CSF vs Behavior	-0.024	-0.080	0.032	-0.273	-0.543	0.006	-	-	-

201

202 **SNP associations**

203 All 13 markers investigated (Fig 1) conformed to Hardy-Weinberg equilibrium, except
 204 two (SNPs 1 and 9, Table 3). No significant associations were detected between individual
 205 markers and behavioral outcomes or OXT CSF in our sample, as all uncorrected 95% credible
 206 intervals overlapped zero (Table 2). When all markers were included in the model, no significant

207 improvement in model fit was observed, indicating that the cumulative effect of all SNPs still
208 results in negligible effects on the investigated traits.

209

210 **Table 3. OXTR SNP Characteristics**

	SNP	Hardy-Weinberg (χ^2)	p	Minor Allele Frequency
1	chr3:140481032 G/A	80.365	<0.001 ***	0.204
2	chr3:140497572 G/C	0.001	0.981	0.315
3	chr3:140497358 G/A	0.132	0.716	0.314
4	chr3:140497003 G/T	0.150	0.698	0.026
5	chr3:140496605 T/C	0.494	0.482	0.462
6	chr3:140496348 A/G	0.054	0.817	0.479
7	chr3:140496344 A/C	0.054	0.817	0.479
8	chr3:140496196 G/T	1.103	0.294	0.265
9	chr3:140495645 C/G	4.628	0.031 *	0.430
10	chr3:140495542 A/T	0.246	0.620	0.033
11	chr3:140495303 C/T	0.494	0.482	0.462
12	chr3:140495244 G/A	0.042	0.837	0.143
13	chr3:140495203 T/G	0.051	0.821	0.448

211 *indicates significant deviation from Hardy-Weinberg equilibrium, as tested with a chi-square test

212

213

214 **Associations between OXT CSF and behavior**

215 Finally, the relationship between OXT CSF and pro-social and anxiety-like behavior
216 yielded no significant effects (Table 2).

217

218 **Discussion**

219 Rhesus are a species with great translational value due to their behavioral and biological
220 parallels to humans. Animals in this particular study also experienced complex social housing,
221 and experimentally controlled environments (similar living, housing, and dietary conditions),
222 attenuating the variability introduced by environmental factors that impact human studies. These

223 benefits might suggest that rhesus behavioral genetic studies, which aim to detect subtle
224 genetic signals among various sources of environmental noise, are exempt from the large
225 samples sizes necessary in analogous studies conducted in humans. However, the use of the
226 rhesus model did not produce effects large enough to be detected in this sample. Specifically,
227 we report no significant associations between the 13 *rhOXTR* SNPs on pro-social or anxiety-like
228 behavior, or on OXT concentrations in CSF. Further, OXT CSF was not associated with any
229 behavioral outcomes.

230

231 Although we did not find significant effects of individual markers, our heritability analysis
232 showed moderate additive genetic effects on all investigated traits, underscoring the existence
233 of substantial genetic influence on these outcomes. This is unsurprising given extensive
234 research in humans demonstrating that most complex traits are on average 50% heritable [16].

235

236 Regarding SNP associations, explanations for our null results include: (1) Despite our
237 informed strategy for selecting markers, it is possible we identified *rhOXTR* variants that do not
238 affect expression (nor subsequent downstream normative behavior). This is a limitation of
239 candidate-gene approach, which narrows the breadth of testable markers. Whole-genome
240 association methods would be needed to comprehensively identify any and all robust markers
241 (while acknowledging the non-trivial challenges to statistical power this introduces). Alternatively
242 (2), the contributions of individual SNPs across the genome, including these 13, may have
243 small, incremental effects on polygenic, complex traits that studies like this one are too
244 underpowered to detect. Decades of research in humans indicates the latter is more plausible
245 [17]. As such, the experimental control afforded by NHP studies such as this one is not sufficient
246 to generate effects robust enough to override the necessity for large samples required in human
247 studies.

248

249 Our results are consistent with [18], who also investigated *rhOXTR* markers and social
250 behavior in a similarly-sized sample of rhesus and likewise found no significant effects. We
251 agree with their conclusions that behavioral genetic studies in NHPs likely face the same
252 challenges as in humans: First, samples upwards of tens of thousands would be required to be
253 adequately powered. Second, reported associations with complex traits resulting from small
254 genetic studies are more likely to be false-positives and fail to replicate, which has been well-
255 documented in human literature.

256

257 While our approach to select influential *rhOXTR* markers was not effective for our
258 specific behavioral outcomes in the context of this sample size, it could be suitable for
259 identifying endophenotypes such as brain expression and neural activity, which bear a closer
260 biological relationship to the proximate consequences of genetic variation. Future genetic
261 associations studies of social behavior in NHPs should parallel the human genetics field in
262 shifting to extensive collaborative efforts, resulting in the possibility of acquiring large sample
263 sizes, surpassing what would be feasible for individual research groups. Importantly, our results
264 do not negate the wealth of OXT research demonstrating OXT's role in regulating social
265 behavior, nor do they refute the possibility that *OXTR* SNPs may have measurable effects in
266 larger samples, or under different experimental conditions (e.g. stress challenges). Whole-
267 genome approaches, when appropriate sample sizes can be attained, can address the relative
268 influence of *OXTR* variants on shaping primate social behavior.

269

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271

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276
277

278 **References**

279

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334

335

336 **Supporting information**

337 **Table S1. Assay and Primer information**

338

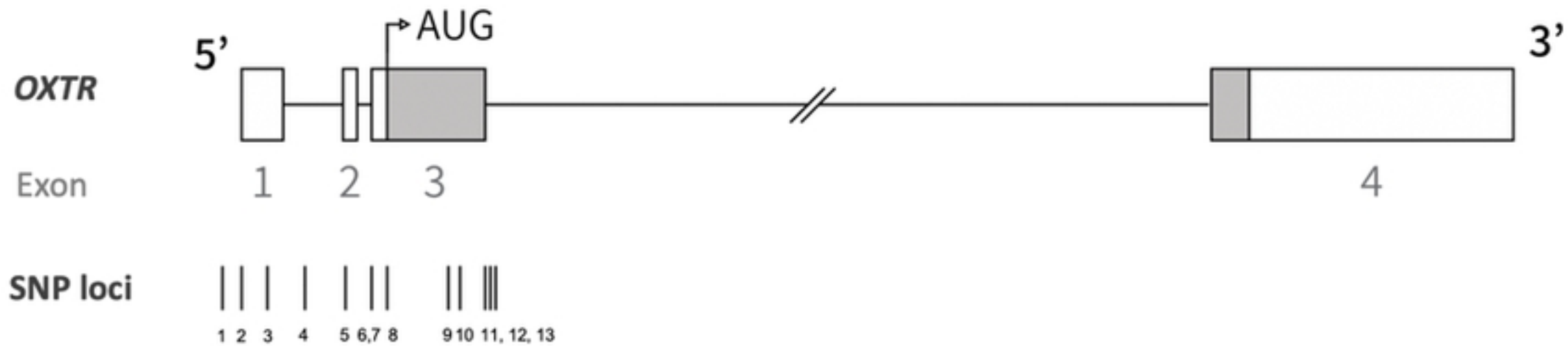


Fig1