

1 **Machine learning guided association of adverse drug reactions with *in vitro* target-based**  
2 **pharmacology**

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## 24 **Abstract**

25 Adverse drug reactions (ADRs) are one of the leading causes of morbidity and mortality in health  
26 care. Understanding which drug targets are linked to ADRs can lead to the development of safer  
27 medicines. Here, we analyze *in vitro* secondary pharmacology of common (off) targets for 2134  
28 marketed drugs. To associate these drugs with human ADRs, we utilized FDA Adverse Event  
29 Reports and developed random forest models that predict ADR occurrences from *in vitro*  
30 pharmacological profiles. By evaluating Gini importance scores of model features, we identify 221  
31 target-ADR associations, which co-occur in PubMed abstracts to a greater extent than expected  
32 by chance. Among these are established relations, such as the association of *in vitro* hERG  
33 binding with cardiac arrhythmias, which further validate our machine learning approach. Evidence  
34 on bile acid metabolism supports our identification of associations between the Bile Salt Export  
35 Pump and renal, thyroid, lipid metabolism, respiratory tract and central nervous system disorders.  
36 Unexpectedly, our model suggests PDE3 is associated with 40 ADRs. These associations provide  
37 a comprehensive resource to support drug development and human biology studies.

## 38 **Keywords**

39 Adverse drug reactions, adverse event report, FAERS, secondary pharmacology, machine  
40 learning, statistical modeling, drug discovery & development, drug safety.

41 Toxicity is one of the major causes of termination, withdrawal, or labeling of a drug candidate or  
42 drug, other than lack of efficacy<sup>1-3</sup>. There is an urgent need to better identify toxic on- and off-  
43 target effects on vital organ systems especially for cardiovascular, renal, hepatic and central  
44 nervous system (CNS)-related toxicities; furthermore, there is a desire to reduce cost and labor  
45 in preclinical assays and drug testing on non-human species<sup>4-6</sup>. *In vitro* pharmacological assays  
46 have been widely used to screen for possible off-targets and potential adverse effects and  
47 eliminate compounds that are not safe enough in the drug development stage as early as possible  
48<sup>5,7</sup>. However, systematic prediction of compound safety and potential adverse events associated  
49 with a compound is still a challenge for the pharmaceutical industry.

50  
51 Machine learning can be very insightful for many different stages of drug discovery and  
52 development, such as automation in pharmacology assays, clinical trials, and basic science  
53 research. Previous studies have focused on predicting structure-function relationships based on  
54 chemical structure of small molecules and potency assays that probe the physicochemical  
55 properties of compounds to estimate associations with off-targets<sup>8</sup>. However, the diversity of  
56 structures that interact with targets, even when they are well described like human Ether-a-go-  
57 go-related gene (hERG), make it challenging to produce reliable models<sup>9</sup>. Several papers provide  
58 small, hand-curated databases providing up to 70 pharmacological targets (i.e. receptors, ion  
59 channels, transporters, etc.) with established links to adverse side effects based on a scientific  
60 literature search<sup>5,7,10,11</sup>. Mirams et al. recently described how integration of data from multiple ion  
61 channels (e.g. hERG, sodium, L-type calcium) provided improved *in silico* prediction of  
62 torsadogenic risk<sup>12</sup>. Chen et al. proposed a machine learning approach to predict adverse drug  
63 reaction (ADR) outcomes for given patient characteristics and drug usage<sup>13</sup>. Another study  
64 highlights importance of predicting the likelihood of clinical trial side effects using human genetic  
65 studies of drug-targeted proteins<sup>14</sup>. From a pharmacogenomics perspective, predicting drug-  
66 target interactions using pharmacological similarities of drugs and the US Food and Drug  
67 Administration (FDA) Adverse Event Reporting System (FAERS<sup>15</sup>) can be beneficial for drug  
68 repositioning and repurposing<sup>16</sup>.

69  
70 FAERS is a voluntary, post-marketing pharmacovigilance tool that can be used to monitor the  
71 clinical performance of drugs. In this study, we explore an alternative use of FAERS data to predict  
72 compound safety using Medical Dictionary for Regulatory Activities (MedDRA<sup>®</sup><sup>17</sup>) terms, which  
73 we envision to be useful for future preclinical studies. Our machine learning approach is different  
74 from the aforementioned approaches because we not only predict adverse drug reaction

75 occurrences of drugs but most importantly also extract biologically meaningful target-ADR links.  
76 Using an *in vitro* secondary pharmacology database of more than 2,000 marketed or withdrawn  
77 drugs (see Methods), we built a random forest model to predict drug-ADR and target-ADR  
78 associations. We validate drug-ADR predictions through systematic Side Effect Resource  
79 (SIDER) drug label analysis and 221 target-ADR predictions through systematic literature co-  
80 occurrence analysis. Furthermore, we find canonical target-ADR associations, such as hERG  
81 binding causing cardiac arrhythmias. We also encountered unexpected associations which  
82 warrant further investigations, such as a link between Phosphodiesterase 3 (PDE3) and several  
83 ADRs, including congenital renal and urinary tract disorders. We conclude our study with potential  
84 targets that are associated with cardiovascular and renal ADRs to demonstrate the utility and  
85 possible impact of this method in drug development and preclinical safety sciences by enabling  
86 prediction of ADRs from *in vitro* pharmacological profiles.

## 87 **Results**

### 88 **Systematic *in vitro* pharmacology of marketed and withdrawn drugs**

89 To link gene targets to ADR occurrence, we utilized *in vitro* pharmacology assay data for 2134  
90 marketed or withdrawn drugs, generated by Novartis, and ADR reports from FAERS (Figure 1A,  
91 Supplementary Table 1). Withdrawn drugs and their assay data are also included due to the fact  
92 that they are associated with a plethora of ADRs, and thereby constitute an important resource  
93 for our predictive approach. Figure 1B summarizes the top 50% of frequently occurring primary  
94 indications, classified by the Anatomical Therapeutic Chemical (ATC) codes, of the 2134 drugs  
95 using a word cloud. The categories that have the highest number of compounds are antibacterial,  
96 ophthalmological, and antineoplastic drugs. The *in vitro* pharmacology assay data includes  $AC_{50}$   
97 values for each drug at up to 218 different assays for 184 gene targets (see Supplementary Table  
98 2 for a list of target assays). There are 6 classes of these 184 gene targets, with the majority  
99 (47%) of targets falling into G protein-coupled receptors (GPCRs) (Figure 1C), which is a  
100 dominant, widely studied drug target family, broadly represented by marketed drugs<sup>18</sup>. Figure 1D  
101 is a heatmap visualization of the *in vitro* pharmacology assay data, where each row is a drug,  
102 grouped by their ATC anatomical main group terms<sup>19</sup>, and each column is a target assay, grouped  
103 by target class. It consists of  $AC_{50}$  values of drugs for target assays. The heatmap is not a  
104 complete data matrix; 70% of drug-assay combinations have not been tested, i.e. these

105 combinations have NA value for AC<sub>50</sub>. Nevertheless, our data indicate relatively uniform assaying  
106 with respect to the different drug classes.

### 107 **Analysis of adverse event reports from FAERS connects drugs with human ADRs**

108 We queried FAERS<sup>15</sup> using openFDA<sup>20</sup> for 2134 marketed or withdrawn drugs in October 2018  
109 (FAERS Q4\_2018 version; covering all reports from January 2004 to October 2018) and retrieved  
110 671143 adverse event reports using our data extraction criteria (Figure 2A). We only included  
111 reports which were submitted by physicians and were annotated as the primary suspect drug<sup>21</sup>.  
112 There are 464 drugs that did not have a matching name in FAERS, 341 drugs that did not have  
113 any adverse event reports, and 1329 drugs with at least 1 adverse event report. We developed a  
114 significance test based on a binomial null distribution and false discovery rate (FDR) multiple  
115 testing correction to determine if the observed ADR occurrence was significantly high to be  
116 classified as an association (or alternatively no association) between ADR and drug (see Methods  
117 for detail). The resulting drug-ADR associations corresponded strongly (odds ratio = 11,  $\chi^2$ -test,  
118 p-value < 10<sup>-16</sup>) with those identified with ERAM (Empirical-Bayes Regression-adjusted Arithmetic  
119 Mean), an established Bayesian method based on the proportional reporting ratio adjusted for  
120 covariates and concomitant drugs<sup>22,23</sup>. Overall, we observe a positive trend between the number  
121 of adverse event reports and the number of ADR associations (Figure 2B). Antineoplastic and  
122 immunomodulatory drugs (Figure 2B, blue, N=155) have many ADR associations while the extent  
123 of ADR association for antihypertensive drugs (Figure 2B, red, N=35) varies more widely. As an  
124 example, we visualized our drug-ADR associations (Figure 2C), in which ADRs are grouped by  
125 MedDRA System Organ Class (SOC) level terms and drugs are grouped by ATC anatomical main  
126 group terms<sup>19</sup>, revealing that ADRs are widespread across organs caused by antineoplastic and  
127 immunomodulating agents (Figure 2C, label L), as well as nervous system drugs (Figure 2C, label  
128 N).

129

### 130 **Random forest model learns relationship between *in vitro* pharmacology and reported 131 ADRs in humans**

132 We deployed a machine learning approach to predict ADRs for a given drug from their *in vitro*  
133 secondary pharmacology profiles (Figure 3A). We consider this a multi-label classification  
134 problem because a given drug can cause multiple ADRs based on its possible engagement with  
135 multiple targets and because a single target may be associated with multiple ADRs. We  
136 discretized and one-hot encoded our *in vitro* pharmacology assay data (AC<sub>50</sub> values) into 3

137 classes: highly active ( $AC_{50} < 3 \mu\text{M}$ ), active ( $3 \mu\text{M} \leq AC_{50} \leq 30 \mu\text{M}$ ) and inactive ( $AC_{50} > 30 \mu\text{M}$ ),  
138 which reflect commonly used ranges in the field<sup>4</sup>. In total, 413 features (assay information) were  
139 used to predict 321 High Level Group Term (HLGT) ADRs or 26 System Organ Class (SOC)  
140 ADRs for each drug. The observed drug-ADR associations from FAERS, as described above,  
141 constitute the dependent variable that the model is learning. We constructed a unifying binary  
142 relevance random forest model, which consists of 321 random forest HLGT ADR models. The  
143 models were first trained and tested, using 5-fold cross validation where each fold is selected  
144 sequentially (Figure 3B). We used 1329 drugs for model construction because these drugs had  
145 at least 1 adverse event report in FAERS Q4\_2018. The remaining 805 drugs, which did not have  
146 any ADR reports, were excluded for training or cross-validation. The model predictions are in  
147 probability format, which is used later for target-ADR predictions, and in boolean format (Figure  
148 3A), to enable assessment of model performance via accuracy; macro-precision; macro-recall;  
149 Matthew's correlation coefficient (MCC), a performance measure that takes class imbalance into  
150 account; and area under the receiver operating characteristic curve (macro-AUROC) (Figure 3B).  
151 The unifying random forest model performance of SOC ADRs and HLGT ADRs using the full  
152 training set (1329 drugs) and the 5-fold cross validation sets (266 drugs, averaged) are depicted  
153 in Figure 3B. Accuracy ranges from 0.82 to 0.98, macro-precision ranges from 0.5 to 0.85, macro-  
154 recall ranges from 0.29 to 0.74, MCC ranges from 0.37 to 0.83, and macro-AUROC ranges from  
155 0.80 to 0.96. Compared to SOC level (21 ADR terms), the finer grain HLGT level (321 ADR terms)  
156 had proportionally fewer drug-ADR associations; additionally, the performance of the HLGT and  
157 SOC models are comparable. We therefore proceeded with the HLGT level models for further  
158 investigation.

159  
160 For 55 of the 321 HLGT ADRs, the corresponding random forest models simply predicted zero  
161 for all drugs as mostly none (and at most 4) of the 1329 drugs with adverse event reports were  
162 associated with those ADRs (Supplemental Table 3). Since these models were not predictive, we  
163 did not consider them for further analyses. For the remaining 266 ADRs, we could determine  
164 performance metrics (Figure 3C). Accuracy and precision were high, ranging between 0.9 and 1,  
165 whilst the recall and MCC range more widely (Figure 3C). This variability occurs for ADRs that  
166 have only a few drugs associated with them (Figure 3D). As the number of associated drugs  
167 increases, the models learn to better distinguish true positives from false negatives so that their  
168 recall and MCC values increase (Figure 3D).

169

## 170 **Predictive power of the random forest model for multiple FAERS reporting time periods**

171 To test if our random forest model framework is sensitively dependent on the FAERS reporting  
172 period, we constructed new random forest models and performed 5-fold cross validations for both  
173 SOC and HLGT levels using FAERS data from 2 different time points: Q4\_2014 (including all  
174 reports from January 2004 to December 2014) and Q2\_2019 (including all reports from January  
175 2004 to June 2019). For proper comparison, the model constructions and cross validations were  
176 identical to our above described “main” model based on FAERS Q4\_2018. Overall, the  
177 performance metrics (accuracy, MCC, macro-precision, macro-recall, macro-AUROC) of both  
178 SOC and HLGT level models are comparable between Q4\_2014, Q4\_2018 and Q2\_2019  
179 (Supplementary Table 4). This analysis demonstrates that our random forest modeling framework  
180 has a comparable predictive power despite changes in the FAERS reporting time period;  
181 therefore, it is not sensitive to different versions of FAERS.

182

## 183 **Chronological validation of predicted drug-ADR associations**

184 To validate the predictive power of our random forest modeling framework further, we performed  
185 a chronological validation analysis, through identification of initial false predictions (false positives  
186 and false negatives) from the random forest model trained on FAERS Q4\_2014 that become  
187 validated in the subsequent time period 2015-2019. The random forest model trained on Q4\_2014  
188 data has 421 (0.1% of a total of N=433671 model predictions) false positive drug - ADR  
189 associations, i.e. based on a drug’s pharmacology profile the model predicted a probability > 0.5  
190 (Figure 3A) for an ADR even though there was no association observed from the adverse event  
191 reports up until 2014. However, when compared to the observed Q2\_2019 FAERS data, which  
192 also include adverse event reports from the time period 2014-2019, 3.1% (13) of the false  
193 positives turned into (true positive) observed drug-ADR associations, which is 4.4-fold more than  
194 expected by chance ( $\chi^2$ -test: p-value =  $2 \times 10^{-5}$ ). Similarly, the Q4\_2014 random forest model made  
195 8519 false negative predictions, of which 2.2% (184), 40-fold more than expected by chance ( $\chi^2$ -  
196 test: p-value <  $10^{-16}$ ), turned into true negative predictions when compared to the Q2\_2019  
197 observed drug-ADR associations. This analysis indicates that significant proportions of our model  
198 predictions on drug-ADR associations that were initially “false predictions” are chronologically  
199 validated through accumulation of new adverse events reports over time.

200

## 201 **Random forest model predicts expected ADR profiles for anti-hypertensive drugs**

202 As another demonstration of model validation, we analyzed the ADR profiles of 6 subclasses of  
203 antihypertensive drugs: adrenergic alpha, adrenergic beta, ACE inhibitors, angiotensin AT2



204 inhibitors, calcium channel blockers and diuretics (Supplementary Table 5). The signature of the  
205 anti-hypertensive drug subclass represents a set of ADRs that were common to all drugs in this  
206 subclass. Each antihypertensive drug subclass has a unique ADR fingerprint in the Q4\_2018  
207 FAERS version which was closely predicted by our random forest model (Figure 3E). The  
208 accuracy ranged from 0.984 to 1, with perfect specificity and precision (Supplementary Table 6).  
209 The sensitivity ranged from 0.882 to 1, except for the diuretics sub-class, which had a sensitivity  
210 of 0.167. This may be because diuretics target the kidney, and not the cardiovascular system as  
211 the rest of the anti-hypertensive drugs do. Of note, the adrenergic alpha and adrenergic beta  
212 receptor subclasses maintain distinct profiles in the predicted data. Specifically, the model  
213 correctly predicts that adrenergic alpha receptor drugs are associated with suicidal and self  
214 injurious behaviors, which has been reported in the literature<sup>24,25</sup>.

215

### 216 **Random forest model validation through comparison with drug label ADRs**

217 To demonstrate the predictive power of our random forest model on a test set of drugs that were  
218 not used for model construction, we utilized the model to predict drug-ADR associations for 805  
219 drugs that did not have any reported ADRs in the FAERS Q4\_2018 version, either because there  
220 was no match with the drug name or there were no ADR reports for that drug submitted to FAERS.  
221 For validation, we queried the Side Effect Resource (SIDER) database<sup>26</sup>, which is independent  
222 from FAERS and contains drug-ADR pairs extracted from FDA drug labels by text mining<sup>26</sup>. For  
223 these 805 drugs, we obtained 95 drug matches, which were further reduced to 75 drugs that did  
224 not share active ingredients with drugs in the training set. Overall, 57% of positive drug-ADR pairs  
225 (i.e. drugs where the model predicts ADRs) were reported in SIDER, compared to 9% of negative  
226 pairs (N = 24075;  $\chi^2$ -test: p-value <  $10^{-16}$ ; Supplementary Table 7). For instance, methysergide,  
227 a 5-HT receptor antagonist used to treat migraine and cluster headaches, has predicted ADRs  
228 from 6 HLG categories, all of which are supported by specific ADRs from SIDER (Figure 3F).  
229 "Cardiovascular disorders with murmurs" appears in the Warnings and Precautions section of the  
230 label. Other adverse events under gastrointestinal symptoms and CNS symptoms from SIDER  
231 were confirmed in the Adverse Events section. Oxprenolol, a lipophilic beta blocker used for  
232 treating angina pectoris, abnormal heart rhythms and high blood pressure, has predicted ADRs  
233 from 3 HLG categories. The specific SIDER ADRs of bradycardia, dizziness and asthenia were  
234 also confirmed in the label from the Electronic Medicines Compendium  
235 (<https://www.medicines.org.uk/emc/product/3235>; accessed 09/11/2019). Overall, our random



236 forest model proves to be a powerful tool to predict both on- and off-target related drug-ADR  
237 associations from *in vitro* pharmacological drug profiles.

238

### 239 **Random forest model predicts 221 target-ADR associations**

240 To predict which target genes are associated with which ADRs, we utilized the Gini importance  
241 score to rank features for their importance in random forest models for each ADR (Figure 4A).  
242 For a given ADR, we selected assays that had multiple AC<sub>50</sub> features represented in the top 5%  
243 of Gini scores ranking (see Methods for detail). We then generated ADR probability predictions  
244 for an *in silico* compound that is assumed to target only the selected assay with an AC<sub>50</sub> value  
245 corresponding to a represented feature. We also assumed no available data for all other assays.  
246 Using this *in silico* AC<sub>50</sub> profile as an input to the ADR model, we could generate the ADR  
247 probability. By assessing differences in ADR probabilities (two sample t-test, FDR corrected p-  
248 value < 0.1) between different AC<sub>50</sub> classes, e.g. highly active (0-3 μM) vs inactive (>30 μM), we  
249 predict positive or negative correlations, collectively termed associations, between the selected  
250 target assay and ADR. Unsurprisingly, some ADRs did not generate any target associations.

251

252 To find biologically meaningful associations, we first filtered out HLGTT terms belonging to SOC  
253 classes that are not specific to human body parts or only procedural or intervention related (see  
254 Methods for detail). Secondly, we filtered out terms that fall under the SOC class neoplasms,  
255 since genes are often severely misregulated in cancers and therefore not representative of  
256 neoplasm-related ADRs in the organ where the tumor resides. After filtering, we found 221  
257 statistically significant target-ADR associations (Figure 4B, full details including p-values in  
258 Supplementary Table 8); 51 out of 184 target assays and 132 out of 321 ADRs are represented  
259 (Figure 4B). In the following sections we investigate these associations in more detail.

260

### 261 **Systematic literature validation of target-ADR associations**

262 To validate our ADR-target predictions, we performed a systematic literature co-occurrence  
263 analysis. First, we mapped all genes corresponding to the assays and HLGTT level ADRs to their  
264 respective MeSH terms (Supplementary Table 9). Next, we queried PubMed for the publication  
265 identifiers linked to these MeSH terms and determined the number of publications that  
266 corresponded to both a gene and HLGTT term (i.e. co-occurrence). We found at least one co-

267 occurrence publication for 66% (145) of 219 predicted unique gene-HLGT Mesh pairs, which was  
268 higher (Fisher Exact test: odds ratio=1.8, p-value= $6 \times 10^{-5}$ ) than for all possible negative unique  
269 gene-HLGT pairs (N=26705). In order to control for the fact that some ADRs and genes are  
270 studied more intensively than others, we also compared our set of positive predictions to a  
271 negative control set (N=4890) formed by permuted pairs from the positive set and obtained similar  
272 results (Fisher Exact test: odds ratio=1.5, p-value= $3 \times 10^{-3}$ ). Furthermore, as quantified by the co-  
273 occurrence “lift” over the reporting probability when assuming independence,  $\frac{P(A,T_{co-occurrence})}{P(A)P(T)}$   
274 (see Methods for details), we found 4-fold higher co-occurrence median lift values for our  
275 predictions compared to all negative pairs (Mann Whitney U-test: p-value= $2 \times 10^{-5}$ ), and 3-fold  
276 higher lift than permuted negative pairs (Mann Whitney U-test: p-value= $3 \times 10^{-4}$ ). We conclude that  
277 our target-ADR identification method provides association predictions that are supported by the  
278 literature in higher proportion than random selection of target-ADR pairs.

279

## 280 **Evidence for targets that are predicted to cause cardiovascular-related ADRs**

281 To further validate our model’s ability to predict target-ADR associations, we investigated a group  
282 of cardiovascular ADRs. We found that hERG binding was associated with cardiac arrhythmias  
283 and heart failure (Table 1). hERG encodes for a subunit of the cardiac potassium ion channel and  
284 contributes to cardiac electrical activity, which is necessary to regulate the heartbeat. The  
285 mechanism of action for drug-induced arrhythmias by blocking hERG has been described in  
286 numerous human <sup>27</sup> and animal studies <sup>28</sup>, as well as structural modeling <sup>29</sup> studies (Table 1).  
287 Consistently, our systematic PubMed queries found 753 co-occurrence publications in support of  
288 this predicted association and 6 co-occurrences for hERG binding increasing the risk of heart  
289 failure. We did not find an ADR probability associated within the range of 0-3  $\mu\text{M}$   $\text{AC}_{50}$  of hERG  
290 binding, likely because such strong binding to hERG is a common reason for deprioritizing drug  
291 candidates in development <sup>30</sup>.

292

293 The model predictions also suggest that PDE3 inhibition is associated with cardiac valve disorders  
294 (Table 1, 3 co-occurrence publications). PDE3 inhibition is used clinically to treat dilated  
295 cardiomyopathy <sup>31</sup>, which encapsulates valvular heart disorder. However, the PDE3 therapeutic  
296 window is narrow, partially due to complex signaling networks <sup>32</sup>, and careful dosing is required  
297 to avoid increased mortality in response to treatment.

298

299 Furthermore, our model predicts that adenosine transporter (AdT) inhibition increases the risk of  
300 pericardial disorders (Table 1). For this scenario, we did not find direct supporting evidence in the  
301 literature, however there is evidence that disturbed adenosine homeostasis in pathological  
302 cardiac conditions could result in pericardial effusion or pericarditis <sup>33</sup>.

303

304 The model suggests that glucocorticoid receptor (GR) binding is more likely to lead to myocardial  
305 disorders if the drug has high affinity for GR (Table 1, 8 co-occurrence publications). This is  
306 supported by the finding that glucocorticoid treatment of patients with rheumatoid arthritis  
307 increased the risk of myocardial infarction <sup>34</sup>. Furthermore, it is known that dysregulation of  
308 glucocorticoids can give rise to cardiotoxicity <sup>35</sup>.

309

310 Taken together, this investigation of genes associated with cardiovascular ADRs confirms the  
311 well-known association of hERG with cardiac arrhythmia, and also highlights ADR associations  
312 that would merit further experimental investigation.

313

#### 314 **COX-2, PDE3, and hERG associations with kidney related ADRs**

315 Another important class of ADRs are related to the kidney (Figure 4B, label: renal). We found  
316 COX-2 associated with nephropathies (Table 2), which has been well recognized (398 co-  
317 occurrence publications) and evidenced previously <sup>36-38</sup>. Interestingly, another model prediction  
318 is PDE3 sensitivity correlating with congenital renal and urinary tract disorders (Table 2).  
319 According to a mouse model study <sup>39</sup>, PDE3 inhibition could be a contributing factor in Polycystic  
320 Kidney Disease (PKD), as PDE3 protein levels are already lower in PKD than WT kidneys. Lastly,  
321 we found an unexpected association between hERG and renal disorders (excluding nephropathy)  
322 (Table 2). One study has found a loss of hERG function in renal cell carcinoma <sup>40</sup>. In humans,  
323 hERG expression in the kidney is much lower than in the heart <sup>41</sup>. Therefore, we conclude that a  
324 link between hERG and renal disorders remains a prediction that warrants further investigation.

325

#### 326 **PDE3 and nuclear hormone receptors AR, ERα, and PR are overrepresented in ADR** 327 **associations**

328 To investigate if the number of different drugs tested for a target assay is predictive to the number  
329 of ADRs associated with that target (Figure 4C), we calculated their Spearman correlation

330 coefficient and found a moderate correlation ( $\rho=0.5$ ; Figure 4C). However, some targets had  
331 considerably more associated ADRs than other targets that were tested a similar number of times,  
332 indicating that more frequently performed assays do not necessarily result in a higher number of  
333 associated ADRs (Figure 4C). Out of all target assays, PDE3 was associated with the most ADRs  
334 (40) (Figure 4C), falling in a wide range of SOC classes (Figure 4B, Supplementary Table 8).  
335 Furthermore, nuclear hormone receptors for androgen (AR), estrogen (ERa) and progesterone  
336 (PR) binding assays also have disproportionately many ADR associations, compared to their  
337 frequency of testing (Figure 4C). As expected, AR (7/14 ADRs), ERa (9/10 ADRs) but not PR  
338 (0/17 ADRs) are associated with sexual reproductive organ and pregnancy-related ADRs (Figure  
339 4B, Supplementary Table 8). Androgen is produced in the adrenal gland<sup>42</sup> and we predict a link  
340 between AR with adrenal gland disorders, with evidence in mouse studies<sup>43</sup>. Interestingly, the  
341 model predicted 6 ocular ADRs associated to PR, including vision disorders, anterior eye  
342 structural change (deposit and degeneration), infections, irritations and inflammations and  
343 structural changes (Figure 4B, Supplementary Table 8), for which we could find supporting  
344 evidence<sup>44</sup>.

345

#### 346 **GABA<sub>A</sub> receptor associations with psychoactive ADRs**

347 GABA<sub>A</sub> receptor is the primary target of benzodiazepines (BZD), a drug class known to be  
348 psychoactive with potential of addiction<sup>45</sup>. Consistently, our model predicts that this ligand-gated  
349 chloride ion channel assay is associated with 14 ADRs, 13 of which are neurologically and  
350 psychiatrically related, including disturbances in thinking and perception, sleep disorders,  
351 depression and suicidal behaviors (Figure 4B, Supplementary Table 8).

352

#### 353 **Bile salt export pump BSEP associations with ADRs in various organs**

354 BSEP, encoded by *ABCB11* and a member of the superfamily of ATP-binding cassette (ABC)  
355 transporters, is most highly expressed in the liver<sup>41</sup>. Drugs that target BSEP are often associated  
356 with hepatotoxicity<sup>46</sup>. However, initially, we did not find a BSEP association with hepatic and  
357 hepatobiliary disorders. To investigate this false negative prediction, we noted that the dynamic  
358 range of the BSEP assay specifically extends up to 300  $\mu\text{M}$  because the first pass effect for orally  
359 delivered drugs results in high concentrations in the liver<sup>47</sup>; as a result, most of our data falls into  
360 the 'inactive' (>30  $\mu\text{M}$  class). Consistently, the BSEP inactive feature had the highest Gini score  
361 for this HLG term, while its two active features had much lower Gini scores, falling outside of the

362 top 5%. To take the extended dynamic range into account, we altered the BSEP assay class  
363 boundaries to 0-30  $\mu\text{M}$ , 30-300  $\mu\text{M}$  and >300  $\mu\text{M}$  and retrained the random forest model. In this  
364 case, we did find BSEP associated with hepatic and hepatobiliary disorders (Table 3, 354  
365 publication co-occurrences), according to our association criteria (Figure 4A). We repeated this  
366 procedure whilst replacing the first class boundary (30  $\mu\text{M}$ ) with 100  $\mu\text{M}$  and found the same  
367 association again, indicating the robustness of our results. Interestingly, with our original  $\text{AC}_{50}$   
368 discretizations (Figure 1D), we found BSEP associated with 7 other ADRs from various organ  
369 classes (Table 3), much more than other targets that were assayed at a similar frequency (Figure  
370 4C). This suggests that compounds potent against BSEP ( $\text{AC}_{50} < 30 \mu\text{M}$ ) could cause adverse  
371 effects in addition to hepatotoxicity, which already occurs at lower potency. We found BSEP  
372 associated with urolithiasis and with disorders of the thyroid gland, upper respiratory tract  
373 disorders (excl infections), lipid metabolism and central nervous system (Table 3). Since BSEP  
374 expression is much lower in these organs <sup>41</sup>, we searched the literature for evidence including a  
375 substrate of BSEP, bile acid. We could find previous studies linking bile acid to these disorders  
376 (Table 3), which suggests an indirect relation between BSEP and these ADRs through bile acid  
377 metabolism. Lastly, we found BSEP associated with foetal complications and pregnancy  
378 conditions (Table 3), both supported through prior studies that link BSEP with transient neonatal  
379 cholestasis and intrahepatic cholestasis of pregnancy, respectively <sup>48,49</sup>.

380

## 381 Discussion

382 In this study we have taken a machine learning approach to predict human ADRs from the *in vitro*  
383 secondary pharmacology profiles of a large number of marketed and withdrawn drugs. Several  
384 prior studies focus on predicting ADRs directly from chemical drug structure <sup>50,51</sup>. However,  
385 utilizing functional information such as *in vitro* pharmacological targeting of common (off) targets  
386 represents a viable alternative to bridge the complex relationship between drugs and their effects  
387 in the human body <sup>4</sup>.

388

389 Our random forest model performance metrics are good considering the sparse coverage (2134  
390 drugs) over a large input space ( $3^{184}$  possibilities) and partial overlap with ADR reporting for these  
391 drugs, making ADR occurrence prediction effectively a one shot learning task. Importantly,  
392 optimizing test performance was not the main objective of this study. Instead, we endeavored to  
393 find biologically meaningful target-ADR associations. To achieve this without relying on test  
394 performance, we trained on all data and made use of Gini scores to robustly select relevant

395 features for ADR probability predictions. Our novel method for target-ADR associations was able  
396 to recapitulate well recognized causal relations, such as hERG with cardiac arrhythmias. For  
397 others, we were able to find literature evidence in animal or *in vitro* studies but our study is, to our  
398 knowledge, a first in human report. Another fraction of target-ADR associations represents  
399 predictions of novel, unexpected or little known associations, such as Adenosine Transporter  
400 (AdT) and pericardial disorders, for which we could find little evidence other than our analysis of  
401 adverse event reports. Similar to genome-wide association studies, our quantitative methodology  
402 extracts statistically significant relations from human population data. With this framework in mind,  
403 our 221 associations form a rich resource that can be used for further mechanistic studies in the  
404 drug discovery process.

405  
406 Our random forest model is agnostic to molecular mechanisms; therefore, resulting associations  
407 could arise from indirect regulation. A likely example is the bile transporter BSEP, which is  
408 associated with numerous ADRs, although it is most highly expressed in the liver and kidney. We  
409 have related our findings to evidence that misregulation of its substrate, bile acid, could result in  
410 disorders related to kidney stones, lipid metabolism, thyroid gland, respiratory system, and central  
411 nervous system. This also indicates the strength of our approach, which can relate genes to  
412 physiological processes unbiasedly in humans, without any interventions or large scale genome-  
413 wide association studies, but solely with voluntary adverse event reporting.

414  
415 While we recommend this approach to find target-ADR associations to impact safety awareness  
416 in drug discovery, we are also aware of the limitations. Firstly, the presented analyses are limited  
417 by the input data. The *in vitro* data matrix is incomplete (targets in the *in vitro* pharmacology panel  
418 cover a small fraction of the biological target space and not all drugs were tested in all assays).  
419 We recognize that the present set of targets is biased towards the GPCR target family with limited  
420 representation of other therapeutic or ADR-associated targets such as ion channels and kinases.  
421 Also, data are influenced by prior knowledge; for example, more than 87% of all drugs in the set  
422 were tested for hERG activity. High affinity (lower  $AC_{50}$  value) for hERG is associated with higher  
423 probability for QT prolongation for human and non-human preclinical species<sup>27,28</sup>. As discussed  
424 earlier, there are not many drugs with a hERG  $AC_{50}$  value in the highly active class (0-3  $\mu$ M),  
425 which is a commonly encountered roadblock for drug candidates to progress towards clinical trials  
426<sup>30</sup>. Only about 10% of all drugs fall into the highly active class in our assay data. To limit feature  
427 engineering, our  $AC_{50}$  discretization into three classes (Figure 1D) was kept uniform across all  
428 assays. Notably for the BSEP assay only, the dynamic range extends up to 300  $\mu$ M and as a



429 result most of our data falls into the 'inactive' (>30  $\mu$ M) class. Consequently, we initially did not  
430 find the expected association with hepatotoxicity. We rectified this by reclassifying the BSEP  
431 assay data according to levels required for hepatotoxicity of BSEP inhibition<sup>52,53</sup> and indeed  
432 recovered the expected association.

433  
434 Secondly, *in vitro* potency is a very simplified marker of clinical effect, and does not take into  
435 account prolonged dosing, comorbidity or pharmacokinetic/pharmacodynamic relationships (e.g.  
436 therapeutic window). For 9 of 184 assays, non-human proteins were assayed (e.g. rat brain was  
437 used as a source for the benzodiazepine receptor) which may not be a direct correlate of the  
438 human protein. Further development of the model would require addition of parameters on  
439 occupancy and pharmacodynamic components for more precision and enhanced predictive  
440 value.

441  
442 Thirdly, the FAERS database has limitations. For example, drug-ADR associations may be  
443 mislabeled, e.g. anti-hypertensives are often reported as associated with hypertension as an  
444 ADR, rather than as the indication. This and other limitations are discussed by Maciejewski et al.  
445<sup>21</sup> with suggestions and methodology for further refinement of the method. Additionally, the  
446 FAERS database does not contain information on the total number of patients exposed to a  
447 particular drug, nor is it necessarily a reflection of the true incidence or frequency of ADRs.

448  
449 This work retains several uncertainties. One of the most critical might be the prediction of  
450 congenital ailments, which are hard to prove. The one example we would like to highlight is the  
451 PDE3 enzyme association with congenital renal disorders association. While the association is  
452 correct, the modality has to be clarified: PDE3 inhibitors are proposed to ameliorate certain forms  
453 of chronic kidney disease<sup>54</sup>, instead of causing it. Thus, predictions of congenital disorders should  
454 be considered but confirmed by checking the modality of the effects.

455  
456 We investigated one-to-one associations between targets and ADRs because these relationships  
457 are biologically meaningful and have utility in preclinical drug development. However, in some  
458 cases, a given ADR can be a prerequisite for others (e.g. hypotension leading to reflex  
459 tachycardia). We leave a model extension to incorporate these dependencies as future work. For  
460 target-ADR associations, we utilized our random forest model for a single drug at a time. One can  
461 repurpose our model to predict possible ADRs from combination drug therapies and likelihood of  
462 drug-drug interactions. In principle, this can be extended for combination therapies by merging



463 the *in vitro* data from the individual compounds. Offside and Twosides databases can be used for  
464 validation <sup>55</sup>. Similarly, our model can be utilized for drug repositioning and repurposing, using  
465 similar target-ADR profiles. In conclusion, our random forest model and the target ADR  
466 associations provide a validated, comprehensive resource to support drug development and  
467 future human biology studies.

468

## 469 **Methods**

### 470 ***In vitro* secondary pharmacology assays for marketed drugs**

471 AC<sub>50</sub> values of 2134 marketed drugs (Supplementary Table 1) were measured in up to 218  
472 different *in vitro* secondary pharmacology assays. Compounds were obtained from the Novartis  
473 Institutes of Biomedical Research (NIBR) compound library and tested in a panel of *in vitro*  
474 biochemical and cell-based assays at Eurofins and at NIBR in concentration-response (8  
475 concentrations, half-log dilutions starting at 30 μM). Assay formats varied from radioligand binding  
476 to isolated protein to cellular assays. Example protocols may be found at  
477 <https://www.eurofinsdiscoveryservices.com/cms/cms-content/services/in-vitro-assays/>.

478 Normalized concentration response curves were fitted using a four parameter logistic equation  
479 with internally developed software (Helios). The equation used is for a one site sigmoidal dose  
480 response curve Y as a function of tested concentrations X:  $Y(X)=A+(B-A)/(1+(X/C)^D)$ , with fitted  
481 parameters A=min(Y), B=max(Y), C=AC<sub>50</sub> and exponent D. By default, A is fixed at 0, whereas B  
482 is not fixed.

483

484 If a drug was not tested against a specific assay, the AC<sub>50</sub> value was set to NA (not available).  
485 AC<sub>50</sub> values from similar assays with the same gene target were merged to reduce the NA data  
486 and features in the random forest model; this procedure resulted in 184 different target assays  
487 (Supplementary Table 2). In case any merged assays had multiple AC<sub>50</sub> values for the same drug,  
488 we averaged these geometrically to take into account variation over orders of magnitudes. In  
489 figures 1D and 2C, the drugs are classified according to their annotated Anatomical Therapeutic  
490 Chemical (ATC) code <sup>19</sup>. In case of multiple ATC codes, we assigned the most frequent level 1  
491 code.

492

### 493 **Mining adverse event reports of marketed drugs using OpenFDA**

494 In this study, we utilized openFDA to acquire FAERS reports related to the query compounds<sup>15,20</sup>.  
495 This Elasticsearch-based API provides a raw download access to a large volume of structured  
496 datasets, including adverse events reports from FAERS.

497  
498 We used generic compound names (e.g. “Amoxicillin”) to query through the openFDA interface,  
499 accessed programmatically using Python. In order to maximize the coverage over FDA datasets,  
500 we normalized generic names to uppercase format followed by a name similarity metric to filter  
501 out unrelated records in our analysis. We included reports when the Jaro similarity between the  
502 query generic name and reported compound name was equal or greater than 0.8. To illustrate, to  
503 query “3alpha-Androstanediol”, we acquired reports including “3 $\alpha$ -Androstanediol”,  
504 “Androstanediol”, “3-alpha-Androstanediol” as different lexical variations of the generic name and  
505 collated the resulting adverse event reports.

506  
507 As the FAERS database contains information voluntarily submitted by healthcare professionals,  
508 consumers, lawyers and manufacturers, adverse event reports may be duplicated by multiple  
509 parties per event, and may be more likely to contain incorrect information if submitted by a non-  
510 medical professional. To reduce reporting bias and increase report information accuracy, we only  
511 analyzed reports submitted by physicians (data field: ‘qualification’ = 1). In this subset of adverse  
512 event reports, the data were further filtered by reported drug characterization, which indicates  
513 how the physician characterized the role of the drug in the patient’s adverse event. A drug can be  
514 characterized as a primary suspect drug, holding a primary role in the cause of the adverse event  
515 (data field: ‘drugcharacterization’ = 1); a concomitant drug (‘drugcharacterization’ = 2); or an  
516 interacting drug (‘drugcharacterization’ = 3). Here, we included only primary suspect drug reports.  
517 Without this restriction, model performances did not improve. We obtained all adverse events  
518 reports corresponding to the query compound that passed through the aforementioned filters.

519  
520 Adverse event report descriptions are coded as medical terms of MedDRA terminology<sup>17</sup>. Medical  
521 observations can be reported using 5 hierarchical levels of medical terminology, ranging from a  
522 very general System Organ Class term (e.g. gastrointestinal disorders) to a very specific Lowest  
523 Level Term (e.g. feeling queasy). Each term is linked to only one term on a higher level. For each  
524 report, we recorded all MedDRA Reaction terms (data field: “reactionmeddrapt”) at the Preferred  
525 Term level and mapped these Preferred Terms to Higher Level Group Term and System Organ  
526 Class level. For each (ADR term, drug) tuple, we then calculated the ADR occurrence, defined as

527 the following fraction: number of adverse event reports containing that ADR term relative to the  
528 total number of ADR reports for that drug.

529  
530 For different FAERS versions (Q4\_2014, Q4\_2018 and Q2\_2019), we used the same query  
531 except the time parameter TO, which was set to 12/30/2014 for the Q4\_2014 query. For other  
532 two queries, we didn't set the limit parameter which was filled with the query time by default (query  
533 date was 10/10/2018 for Q4\_2018 and 08/12/2019 for Q2\_2019).

### 534 535 **Random forest models**

536 To construct and train our models (Figure 3A), we used  $AC_{50}$  values for a panel of target assays  
537 for marketed drugs (model input; independent variable) and ADR occurrences of the compounds  
538 (model output/predictions; dependent variable). Since there may be several ADRs associated  
539 with any given drug, we considered this a multi-label learning problem. We took a “first-order  
540 strategy”, i.e. we assume there is no correlation between different ADRs, and a “divide and  
541 conquer” strategy, i.e. we decompose our multi-label learning task into  $n$  independent binary  
542 classification problems, where  $n$  is the number of different ADR terms in our output data ( $n = 26$   
543 for SOC and  $n = 321$  for HLG T level respectively). We built a random forest<sup>56</sup> binary classifier for  
544 each ADR using Binary Relevance with the random forest modeling option in mlr package<sup>57</sup> and  
545 utiml package in R<sup>58</sup>.

546  
547 To define the features for the random forest models, we discretized and one-hot encoded our  
548 input  $AC_{50}$  values. Discretization was essential to limit the number of features and enhance the  
549 predictive power of the model. We defined 3 classes (levels) of  $AC_{50}$  ranges for each target assay.

- 550 ● Highly active class:  $AC_{50}$  in  $[0, 3 \mu\text{M})$
- 551 ● Active class:  $AC_{50}$  in  $[3 \mu\text{M}, 30 \mu\text{M}]$
- 552 ● Inactive class:  $AC_{50}$  greater than  $30 \mu\text{M}$

553 If the  $AC_{50}$  value is NA, the values for all Classes are 0. Each drug has  $AC_{50}$  values for 184  
554 (merged) assays, so there are  $184 \times 3 = 552$  binary features to represent our input data. Features  
555 consisting of only 0 values were removed, resulting in 413 input features used for model  
556 construction.

557  
558 The observed ADR occurrences were discretized into binary dependent variables. To achieve  
559 this, first let  $N_d$  be the total number of ADR reports for a given drug. The probability to observe an  
560 ADR occurrence  $O^{\text{ADR}} = X / N_d$  at random is equivalent to choosing that ADR  $X$  times out of  $N_d$

561 with  $X$  distributed binomially:  $X \sim \text{bin}(N_d, p=1/n)$ . Here,  $n$  represents the total number of ADRs as  
562 defined above. Under this null distribution, we calculate the  $p$ -values for all observed ADR  
563 occurrences  $O^{\text{ADR}}$  for a given drug, and then perform a Benjamini-Hochberg False Discovery Rate  
564 (FDR) correction (using the Python statsmodels package). If an FDR-corrected  $p$ -value is  $< 0.01$ ,  
565 then the ADR value for that drug is 1, reflecting an association; 0 otherwise.

566  
567 All random forest models were first trained using 5-fold cross validation and each fold is selected  
568 sequentially. 1063 drugs were used for training and 266 drugs were used for testing in each fold.  
569 Then, the drugs with at least 1 ADR report are used as a training set. For a given (drug) input of  
570  $AC_{50}$  values and ADR, the random forest model output, termed ADR probability, can be  
571 interpreted as the probability that the ADR is associated with the drug. To enable direct  
572 comparison of model predictions with binarized ADR occurrences, we binarized these ADR  
573 probabilities with a simple threshold value of 0.5. These binary values were used for training,  
574 cross validation and to calculate classification performance metrics (Figure 3B,C). All models  
575 have been constructed the same way regardless of different FAERS versions.

576  
577 We evaluated our models based on five metrics: accuracy, Matthew's correlation coefficient  
578 (MCC), macro-precision, macro-recall and area under the receiver operating characteristic curve  
579 (macro-AUROC). These metrics are calculated using their definitions below, except 2 metrics: (1)  
580 MCC, which is calculated using mltools package in R (<https://github.com/ben519/mltools>) and (2)  
581 AUROC, which is calculated using precrec package in R<sup>59</sup>.

- 582 • Accuracy =  $(TP + TN) / (TP + TN + FP + FN)$
- 583 • Precision =  $TP / (TP + FP)$
- 584 • Recall =  $TP / (TP + FN)$
- 585 • MCC =  $(TP * TN - FP * FN) / \text{SQRT}((TP + FP) * (TP + FN) * (TN + FP) * (TN + FN))$
- 586 • AUROC =  $\int_{x=0}^1 TPR(FPR^{-1}(x)) dx$

587 where TPR (true positive rate =  $TP / (TP + FN)$ ) and FPR (false positive rate =  $FP / (FP + TN)$ ).  
588 The corresponding metrics for each ADR model (Figure 3C, 3D) are accuracy, precision, recall,  
589 and MCC, which is calculated using mltools package in R (<https://github.com/ben519/mltools>).

590  
591 **Determination of target-ADR associations**  
592 To find associations between gene target assays and ADRs (Figure 4), we first generated ADR  
593 probabilities specific to a given assay. As a model input, one out of its three random forest input  
594 features' value was set to 1 and all others to 0. This simulates the scenario of an *in silico*

595 compound that is potent with an  $AC_{50}$  value in the range corresponding to the positive feature  
596 only. We then utilized the ADR's random forest model, pre-trained on all available marketed drug  
597 data (see previous section), to calculate the resulting ADR probability. We repeated this  
598 procedure for each feature of all assays and each ADR.

599  
600 To select the predictive features for a given ADR, we ordered the pre-trained random forest  
601 model's input features according to their Gini importance score<sup>60</sup> and denote the top 5% as  
602 significant features. Our criteria for a gene (target assay) - ADR pair were:

- 603 ● For a given ADR: at least 2 out of 3 assay features need to be significant in order to make  
604 a reliable comparison between the ADR probabilities with respect to  $AC_{50}$  values.
- 605 ● At least one of the ADR probabilities of the significant features has to be larger than zero.

606  
607 We filtered out target-ADR pairs if the ADR term maps to the following SOC classes, which are  
608 not specific to body parts or underlying human biology:

- 609 ● general disorders and administration site conditions
- 610 ● injury, poisoning and procedural complications
- 611 ● investigations
- 612 ● neoplasms benign, malignant and unspecified (incl cysts and polyps)
- 613 ● poisoning and procedural complications
- 614 ● social circumstances
- 615 ● surgical and medical procedures

616 To ensure the reproducibility of the target-ADR pair selection procedure, we repeated the random  
617 forest model training with different seeds for a total of 5 times. We then took the union of the 5  
618 sets of target-ADR pairs and discarded pairs that were only found once out of 5 runs. Finally, to  
619 determine if the mean ADR probabilities between the selected  $AC_{50}$  classes were statistically  
620 significantly different, we performed a two-sample t-test with sample sizes equal to the number of  
621 times a class was selected (ranging from 2 to 5 times) using the Python scikit.stats package. In  
622 case all three  $AC_{50}$  classes were represented, we tested the highly active versus inactive class.  
623 We then performed a Benjamini-Hochberg FDR correction. If the FDR-corrected p-value is  $< 0.1$ ,  
624 then the target-ADR pair is considered a statistically significant association (Figure 4B,  
625 Supplementary Table 8).

626

627 To evaluate the relation between the HLGT level ADR term hepatic and hepatobiliary disorders  
628 and target assay BSEP, we also trained and analyzed two random forest models as described  
629 above to find target-ADR pairs but with only the BSEP assay data discretized with class  
630 boundaries [0, 30  $\mu$ M), [30, 300  $\mu$ M] and >300  $\mu$ M or [0, 100  $\mu$ M), [100, 300  $\mu$ M] and >300  $\mu$ M.

631  
632 **Side Effect Resource (SIDER) analysis**  
633 The Side Effect Resource (SIDER; version 4.1) was downloaded  
634 (<http://sideeffects.embl.de/download/>; accessed 09/16/2019). The file meddra\_all\_se.tsv.gz  
635 contains drug-ADR pairs extracted from drug labels using text mining<sup>26</sup>. The supplied MedDRA  
636 preferred term (PT) was mapped to HLGT used for the random forest modeling. The file  
637 drug\_atc.txt provides mappings from drug names as used in SIDER to Anatomical Therapeutic  
638 Chemical (ATC) codes. ATC codes for the 805 drugs in the test set were obtained from the NIBR  
639 compound database, and matched to ATC codes from SIDER. For drugs that could not be  
640 matched via ATC codes, additional matches were obtained by mapping the compound name, first  
641 trying the name in its entirety (e.g. “butriptyline hydrochloride”, then on the first word in the drug  
642 name (e.g. “butriptyline”). All matches, whether obtained on ATC codes or by drug name, were  
643 reviewed manually for accuracy.

644  
645 **Systematic validation of predicted target-ADR association using PubMed database**  
646 We built a query based on 254 unique HLGT level ADR terms and 106 unique target genes  
647 (corresponding to the assays), for which we could find a corresponding MeSH term  
648 (Supplementary Table 9), to retrieve linked publication identifiers (PMIDs) from the PubMed  
649 database. All PMIDs were acquired by submitting a query for every MeSH entity separately via  
650 the PubMed API engine, a search engine that provides access to the MEDLINE database of  
651 references and abstracts on life sciences and biomedical articles. Next, we determined the PMIDs  
652 for a gene-ADR pair as the intersection of the two PMID sets of each corresponding MeSH term  
653 query. Furthermore, for each possible gene-ADR pair we determined whether it was part of the  
654 221 predicted associations from the Random Forest model or not. In this way, we obtained 219  
655 unique positive gene-ADR pairs and a total 26705 unique negative pairs. Lastly, we generated a  
656 set of negative pairs corresponding to all permutation pairs from the 39 unique genes and 131  
657 unique ADRs that are part of the positive set, resulting in 4890 unique negative pairs in this  
658 negative control set. To assess any statistical overrepresentation, we calculated the number of  
659 pairs with at least one co-occurrence publication for both negative and positive sets and assessed  
660 significance with a Fisher Exact test (Python function `scipy.stats.fisher_exact`). Furthermore, we

661 calculated the co-occurrence “lift” over the reporting probability when assuming independence,  
662 defined as  $lift := \frac{P(A,T_{co-occurrence})}{P(A)P(T)} = \frac{N(A,T)*N_{pubmed}}{N(A)N(T)}$ , with  $N_{pubmed} = 29138919$  the total  
663 number of PMIDs in the Pubmed database in 2019  
664 ([https://www.nlm.nih.gov/bsd/licensee/2019\\_stats/2019\\_LO.html](https://www.nlm.nih.gov/bsd/licensee/2019_stats/2019_LO.html)).  $N(A, T)$ ,  $N(A)$ , and  $N(T)$  are  
665 respectively the number of retrieved PMIDs for a unique gene-ADR pair, ADR, or gene target  
666 separately. To assess the location differences of the above described positive versus negative  
667 distribution of lift values, we performed a Mann Whitney U test (Python function  
668 `scipy.stats.mannwhitneyu`, `two-sided`, `continuity correction=True`).



## 669 **Figure Legends**

670

### 671 **Figure 1. Major elements of the target-ADR association analysis**

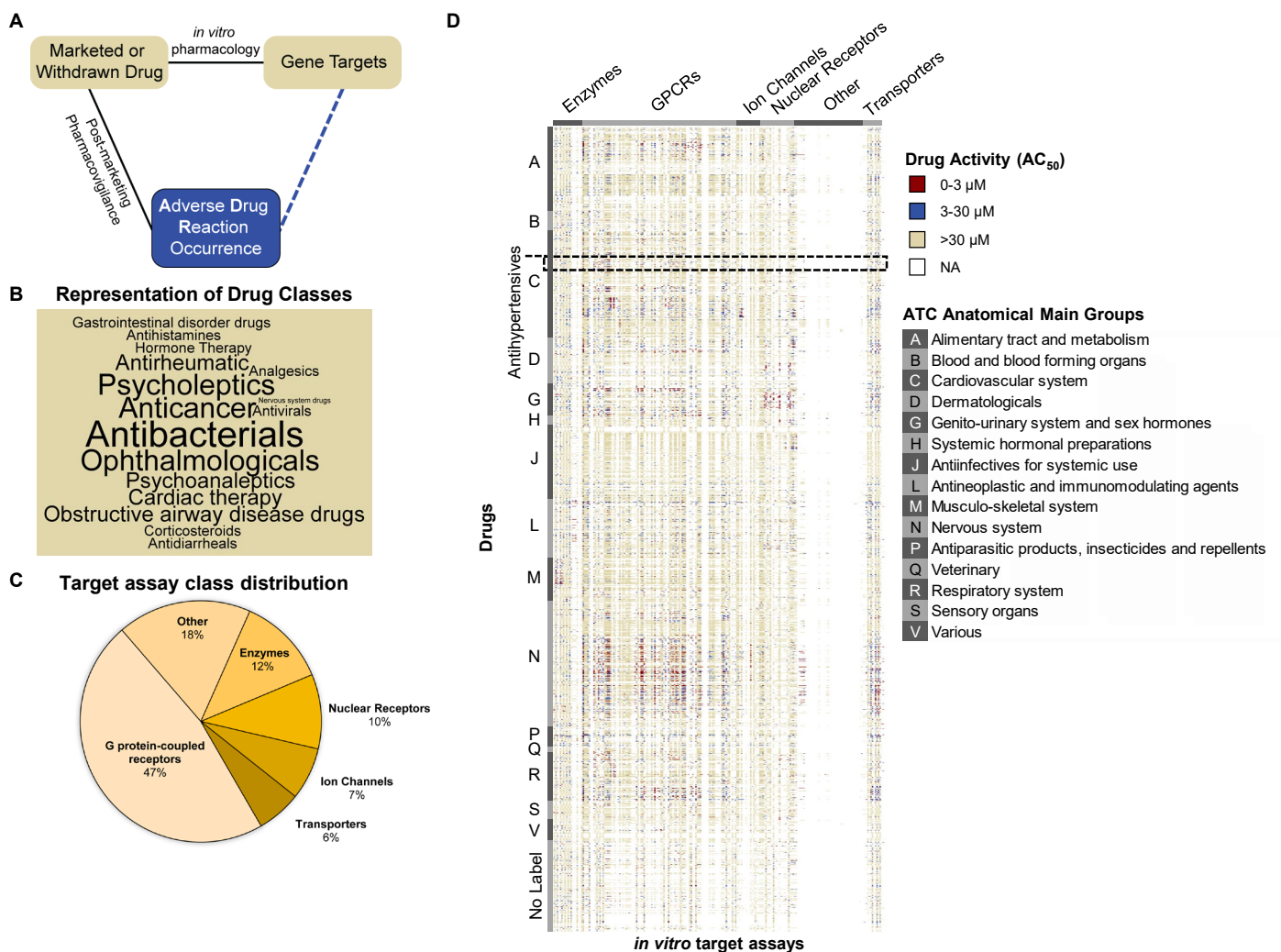
672 **A. Schematic outline of target-ADR pair determinations.** The observed relations (solid lines)  
673 between drugs and adverse drug reactions (ADRs) are determined by post-marketing  
674 pharmacovigilance and between drugs and their (off) targets by *in vitro* pharmacology. This  
675 approach enables prediction of associations (dashed line) between targets and ADRs through  
676 random forest modeling.

677 **B. Representation of drug classes in word cloud.** The cloud displays the top 50% most  
678 frequently occurring drug classes, representing 2134 drugs, in the Novartis *in vitro* pharmacology  
679 data warehouse. Size of the font of the drug class reflects the number of associated drugs.

680 **C. Target class distribution in the Novartis *in vitro* secondary pharmacology assay panel.**  
681 The 184 targets in the Novartis assay panel cover 6 target classes. Almost half of the target  
682 assays belong to the G protein-coupled receptor (GPCR) class.

683 **D. Novartis target panel potency (AC<sub>50</sub>) heatmap.** The profile consists of the AC<sub>50</sub> values of  
684 184 target assays for 2134 drugs. We considered an AC<sub>50</sub> value less than 3 μM as highly active  
685 (red), between 3 μM and 30 μM as active (blue), and greater than 30 μM as inactive (yellow). No  
686 data for a drug-target pair is labeled as NA (white). Drugs are grouped (vertically) by their  
687 Anatomical Therapeutic Chemical (ATC) codes. Assays are grouped (horizontally) by target class.

## FIGURE 1



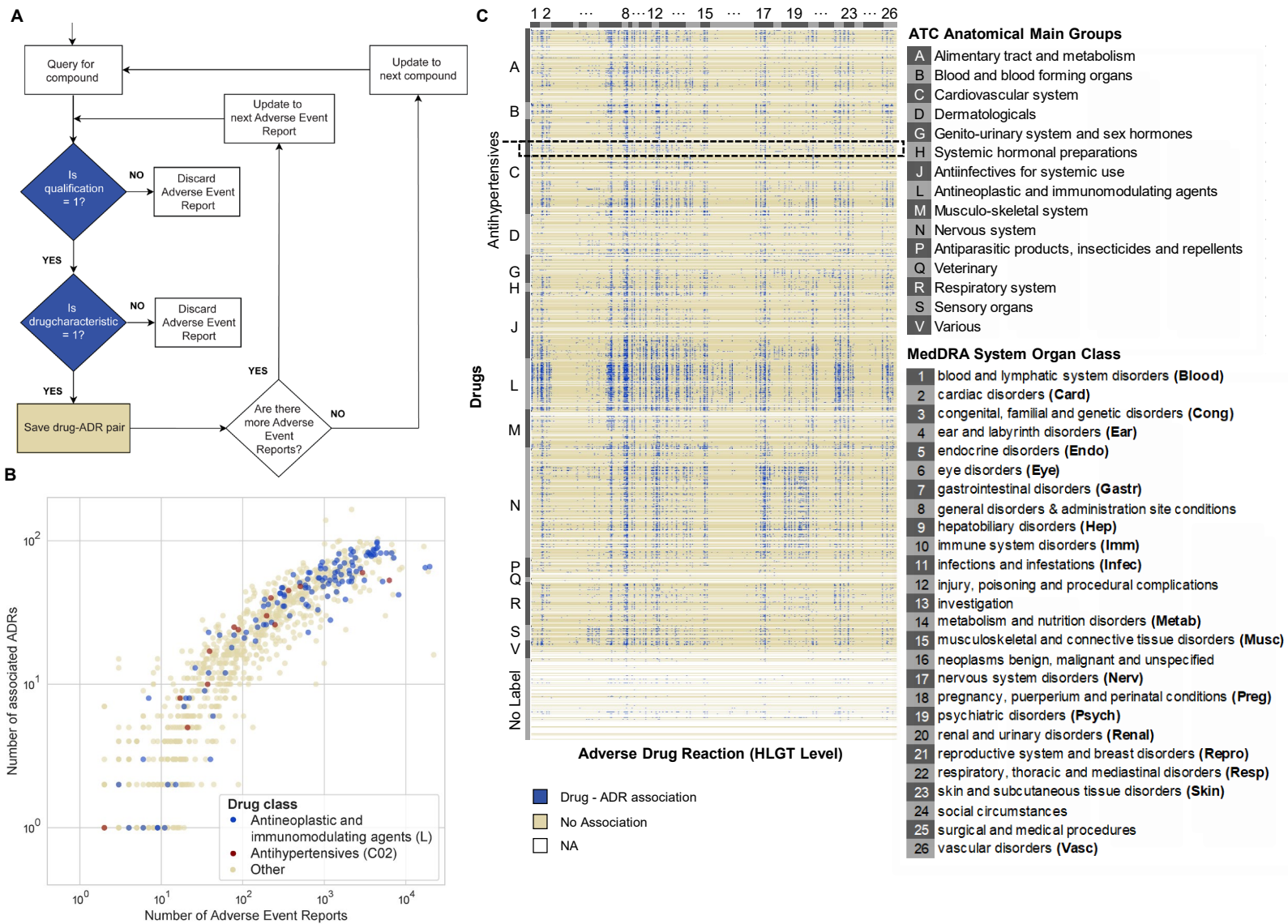
688 **Figure 2. Retrieval of Adverse Event Reports from the FDA Adverse Event Reporting**  
689 **System (FAERS) database**

690 **A. Flow chart of the programmatic strategy for Adverse Event Report retrieval from FAERS**  
691 **by using openFDA.** 'is qualification = 1' is a positive filter for adverse event reports that were  
692 reported by physicians. 'is drugcharacterization' = 1 is a positive filter for drugs that are annotated  
693 as the primary suspect drug, which hold a primary role in the cause of the adverse event.

694 **B. Scatter plot of the number of associated ADRs for drugs as a function of the number of**  
695 **adverse event reports retrieved for each drug ( $N_{\text{drugs}} = 1329$ ).** Drugs without any reported ADR  
696 are not shown.

697 **C. Heatmap of ADR profiles (discretized as used for input of random forest model) for all**  
698 **marketed drugs used in this study ( $N_{\text{drugs}} = 2134$ ).** Drugs are clustered (vertically) according to  
699 their ATC drug classes (A-V, or No label if without any ATC code) and HLG (high level group  
700 term) ADRs are grouped (horizontally) according to the parent System Organ Class (SOC) level  
701 listed in the legend.

## FIGURE 2



702 **Figure 3. Application of the random forest model to characterize drug-ADR associations**

703 **A. Schematic representation of the machine learning approach.** Using input data, which is a  
704 discretized  $AC_{50}$  *in vitro* pharmacological profile, we built a separate random forest model for each  
705 adverse drug reaction (ADR) that predicts the probability of a drug causing that ADR. For training  
706 we used all drugs for which we could retrieve FAERS Q4\_2018 adverse event reports ( $N_{\text{drugs}} =$   
707 1329).

708 **B. Summary statistics of overall model performance.** We developed two unified random forest  
709 models based on two hierarchical levels of organ class specifications. The high level group term  
710 (HLGT; blue) unified random forest model consists of 321 ADR random forest models whereas  
711 the system organ class (SOC; yellow) unified random forest model consists of 26 ADR random  
712 forest models. The performance of the HLGT and SOC models is similar, except in few cases  
713 when the HLGT model outperforms the SOC model. (MCC: Matthew's correlation coefficient,  
714 AUROC: area under receiver operating characteristic). Training reflects performance after model  
715 training on all 1329 drugs (see A). 5-fold cross validation results are averaged over each fold (all  
716 metrics for each fold are detailed in Supplementary Table 4).

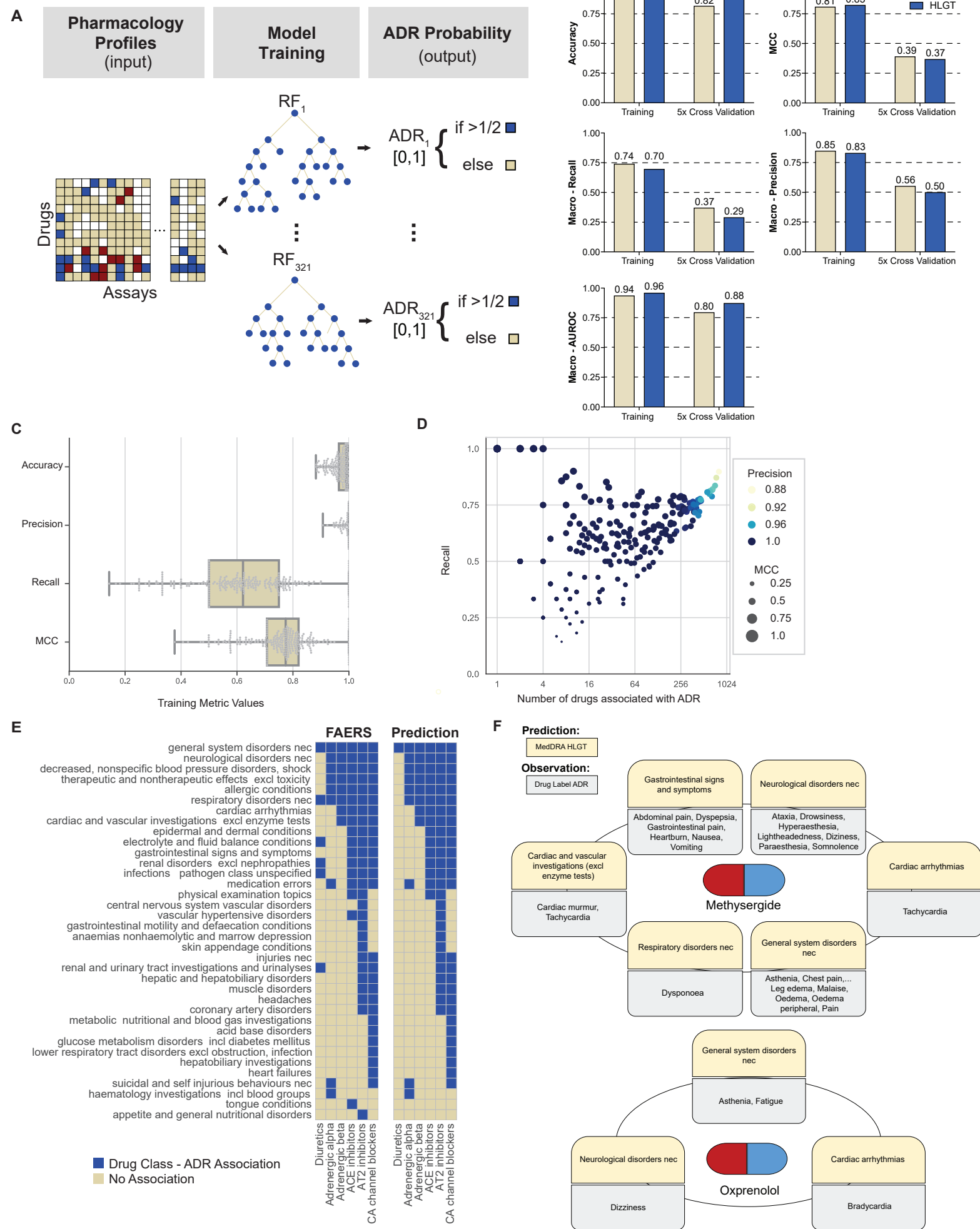
717 **C. Box plots indicating the distributions of the training performance metrics** (as in B) for all  
718 random forest models of each individual HLGT ADR ( $N_{\text{ADRs}} = 266$ ).

719 **D. Scatter plot of the random forest models' recall (all metrics as in C) as a function of**  
720 **number of associated ADRs**, which served as positive training examples. Colors indicate model  
721 precision and circle size reflects the MCC.

722 **E. ADR predictions for anti-hypertensive drugs with different pharmacological targets.** For  
723 a set of 22 antihypertensive drugs, we visualized the association between the drugs and HLGT-  
724 level ADRs (left). Using the ADR random forest models, we predicted the differences in ADR  
725 associations between antihypertensive drugs representing various pharmacological targets (right;  
726 overall 36 of the HLGT terms are visualized). True negative predictions (285 HLGT-level ADRs)  
727 were omitted from this visualization.

728 **F. Examples of model validation using methysergide and oxprenolol.** The random forest  
729 model predicted associations of methysergide with 6 of 321 HLGTs (yellow) which were validated  
730 by comparison of ADRs from its drug label (grey) using the SIDER database. One or more of the  
731 ADRs corresponding to each HLGT category were confirmed in the drug label.





732 **Figure 4. Random forest model predicts target-ADR associations**

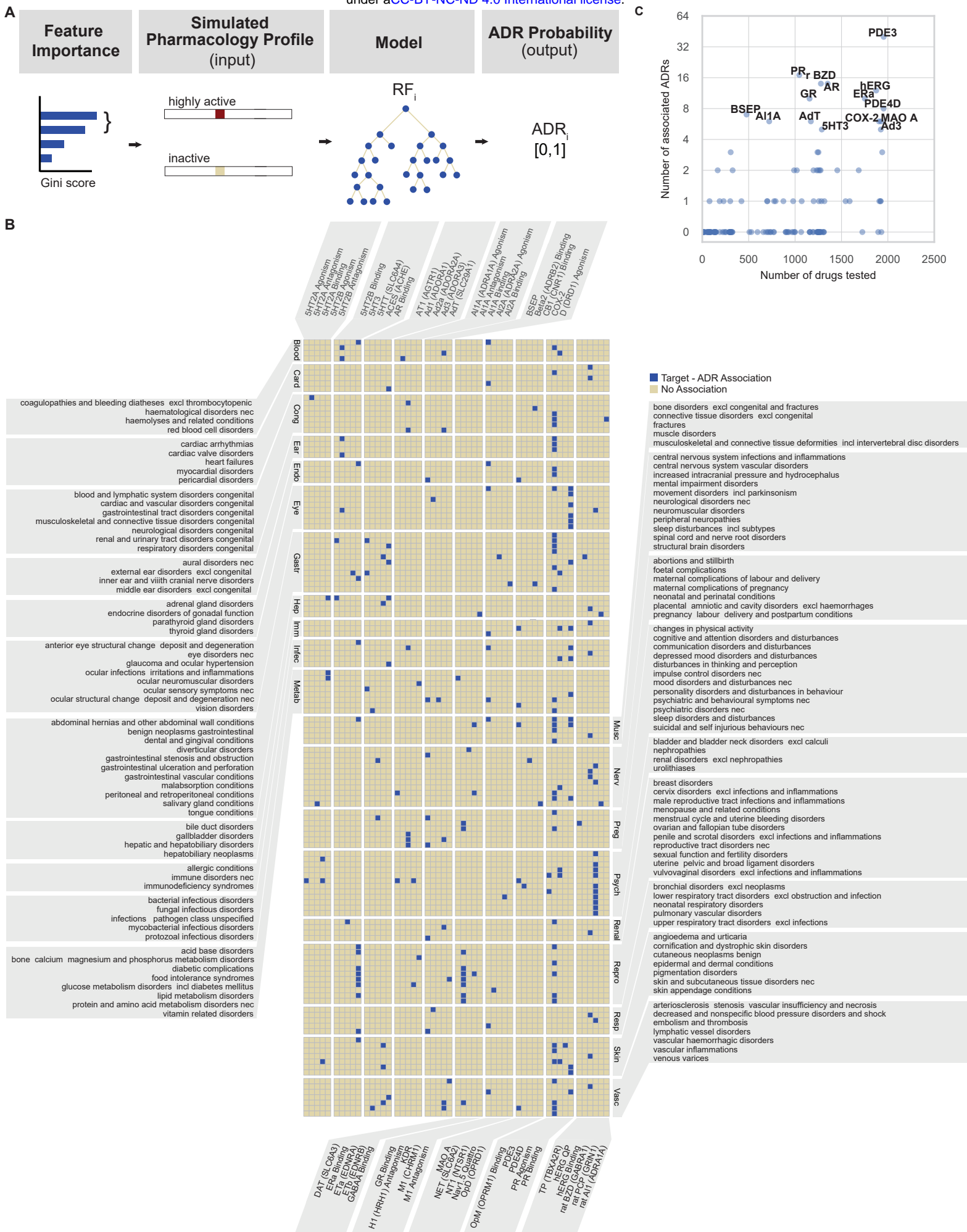
733 **A. Schematic outline of the *in silico* ADR-target predictions.** For an ADR of interest, we  
734 determined the top 5% of features from the corresponding trained random forest model, ranked  
735 according to their Gini importance scores, which measures their contribution to the predictive  
736 power of the model. If at least two features (e.g. as depicted: highly active and inactive) from the  
737 same target assay are within that top 5%, we determined the ADR probabilities for the simulated  
738 cases where an *in silico* compound would target those assay AC<sub>50</sub> classes only. The ADR  
739 probabilities of those simulated cases can then be compared to determine the concentration  
740 dependence of the ADR probability. If there is a non-zero correlation between AC<sub>50</sub> values and  
741 ADR probabilities, we conclude that there is an association between the respective ADR and  
742 target. For full details, see the Methods.

743 **B. Heatmap showing the resulting 221 predicted target-ADR associations (blue).** Target  
744 (gene symbol) assays are listed alphabetically (horizontal), and HLGT ADRs (vertical) are  
745 grouped according to their parent SOC level (as detailed in Figure 2C). For a full description of all  
746 target-ADR associations and their ADR probabilities, see Supplementary Table 8.

747 **C. Scatter plot of each target (assay, N=184) showing the number of ADR associations as**  
748 **a function of number of assayed drugs.**



# FIGURE 4



749 **Table 1** Predicted associations between targets and cardiac ADRs.

750 High Level Group Terms (HLGT; MedDRA) associations with targets and Adverse Drug Reaction  
 751 (ADR) probability in three concentration ranges (third column). Evidence of the ADR-target pairs  
 752 were obtained from peer reviewed publications (fourth column). The number of publications linked  
 753 to both an HLGT ADR and target gene was obtained via a systematic literature co-occurrence  
 754 analysis (fifth column). hERG: human Ether-a-go-go-Related Gene associated potassium  
 755 channel; PDE3: phosphodiesterase-3 enzyme; GR: glucocorticoid receptor; AdT: Adenosine  
 756 transporter; COX-2: cyclooxygenase enzyme, type 2.

Cardiac Disorder HLGT	Target	ADR Probability			Literature evidence human (h), animal (a), <i>in vitro</i> (v)	Co-occurrence (number)
		0-3 $\mu\text{M}$	3-30 $\mu\text{M}$	>30 $\mu\text{M}$		
cardiac arrhythmias	hERG (Binding)	-	0.03	0.002	h <sup>27</sup> a <sup>28</sup> v <sup>29</sup>	753
cardiac valve disorders	PDE3	0.05	-	0	h <sup>31,32</sup>	3
heart failures	hERG (Binding)	-	0.005	0	h <sup>61</sup>	6
myocardial disorders	GR (Binding)	0.02	-	0.005	h <sup>34,35</sup>	8
pericardial disorders	AdT	-	0.01	0	a <sup>33</sup>	0

757

758 **Table 2** Predicted renal ADR - target associations (detailed legend in Table 1).

Renal Disorder HLGT	Target	ADR Probability			Literature evidence human (h); animal (a), <i>in vitro</i> (v)	Co-occurrence (number)
		0-3 $\mu\text{M}$	3-30 $\mu\text{M}$	>30 $\mu\text{M}$		
nephropathies	COX-2	0.003	-	0	h <sup>36</sup> a <sup>37,38</sup>	398
renal and urinary tract disorders congenital	PDE3	0.004	-	0	h <sup>54,62</sup> a <sup>39,63</sup>	0
renal disorders excl nephropathies	hERG (Binding)	-	0.01	0.0007	h <sup>40</sup> a <sup>64</sup>	2

759

760 **Table 3** Predicted ADR associations with inhibition of the Bile Salt Export Pump (BSEP)  
 761 transporter (detailed legend in Table 1).

HLGT	Target	ADR Probability			Literature evidence	Co-occurrence (number)
		0-3 $\mu\text{M}$	3-30 $\mu\text{M}$	>30 $\mu\text{M}$	human (h); animal (a)	
central nervous system vascular disorders	BSEP	-	0.09	0.008	for BSEP and bile acid) a <sup>65</sup>	2
foetal complications	BSEP	0.01	-	0	h <sup>48</sup>	7
pregnancy labour delivery and postpartum conditions	BSEP	-	0.1	0	h <sup>49</sup>	0
lipid metabolism disorders	BSEP	-	0.2	0	h <sup>66,67</sup>	5
thyroid gland disorders	BSEP	-	0.07	0	a <sup>68,69</sup>	1
upper respiratory tract disorders excl infections	BSEP	0.1	-	0	h <sup>70</sup> a <sup>71</sup>	0
urolithiasis	BSEP	-	0.07	0	h <sup>72</sup>	0
		<b>0-30 <math>\mu\text{M}</math></b>	<b>30-300 <math>\mu\text{M}</math></b>	<b>&gt;300 <math>\mu\text{M}</math></b>		
hepatic and hepatobiliary disorders	BSEP	-	0.2	0.09	h <sup>46</sup>	354

762

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768

769 **Author contributions**

770 R.I., S.A., A.X.C., S.F., B.K., D.A. and L.U. conceived the study. D.A., A.F. and L.U. provided  
771 the Novartis *in vitro* pharmacology data, advice and mentorship. S.A., R.I., A.X.C., S.F., B.K.,  
772 W.D.M. and J.S. performed data analysis. S.A. developed the random forest modeling. R.I.  
773 developed the formalism for target-ADR association inference. S.F., R.I. and A.X.C. developed  
774 the query of OpenFDA. J.S. performed the SIDER analysis. S.F., J.S. and R.I. performed the  
775 systematic PubMed query. S.A., R.I., and A.X.C. wrote the paper and designed the figures with  
776 input from all the authors.

777

778 **Conflict of interest**

779 Authors declare no conflict of interest.

780

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