## 1 HERV-K and HERV-W transcriptional activity in Myalgic

## 2 Encephalomyelitis/ Chronic Fatigue Syndrome

3

4	Short	title: Endogenous retroviruses in ME/CFS
5		
6	Lucas	S Rodrigues <sup>1#</sup> , Luiz H da Silva Nali <sup>1,2#</sup> , Cibele O D Leal <sup>1</sup> , Ester C Sabino <sup>1,3</sup> ,
7	Eliana	M Lacerda <sup>4</sup> , Caroline C Kingdon <sup>4</sup> , Luis Nacul <sup>4</sup> , Camila M Romano <sup>1,5*</sup>
8		
9	1.	Instituto de Medicina Tropical de São Paulo, Universidade de São Paulo, São
10		Paulo, Brazil
11	2.	Universidade Santo Amaro, Pós-Graduação em Ciências da Saúde
12	3.	Departamento de Moléstias Infecciosas e Parasitárias, Faculdade de Medicina da
13		Universidade de São Paulo, São Paulo, Brazil
14	4.	Department of Clinical Research, Faculty of Infectious and Tropical Diseases,
15		London School of Hygiene and Tropical Medicine, London, United Kingdom
16	5.	Hospital das Clinicas HCFMUSP (LIM52), Faculdade de Medicina,
17		Universidade de São Paulo, São Paulo, Brazil
18		
19	# Thes	se authors contributed equally to this manuscript
20	* Corr	espondence to: CMR, Laboratório de Virologia, Instituto de Medicina Tropical de
21	São Pa	aulo, Universidade de São Paulo, Rua Dr. Enéas de Carvalho Aguiar, 470, ZIP.
22	05403	-000 São Paulo, SP, Brazil. phone: +55 11 3061-8668; cmromano@usp.br
23		
24		
25	Keyw	ords: Endogenous retroviruses; HERV-W; HERV-K; myalgic encephalomyelitis;
26	chroni	c fatigue

28

#### 29 Abstract

30 Chronic Fatigue Syndrome / Myalgic Encephalomyelitis (CFS / MS) is an incapacitating chronic disease that dramatically compromise the life quality. The 31 32 CFS/ME pathogenesis is multifactorial, and it is believed that immunological, metabolic 33 and environmental factors play a role. It is well documented an increased activity of 34 Human endogenous retroviruses (HERVs) from different families in autoimmune and 35 neurological diseases, making these elements good candidates for biomarkers or even 36 triggers for such diseases. Here the expression of Endogenous retroviruses K and W 37 (HERV-K and HERV-W) was determined in blood from moderately and severely affected ME/CFS patients. HERV-K was overexpressed only in moderately affected 38 39 individuals and HERV-W showed no difference. This is the first report about HERV-K differential expression in moderate ME/CFS. 40

- 41
- 42
- 43
- 44

45

- 46
- 47

48

50

### 51 Introduction

52 Myalgic Encephalomyelitis/ Chronic Fatigue Syndrome (ME/CFS) is a chronic and 53 debilitating disease with unknown etiology [1]. Affected individuals have compromised motor and cognitive capacities. There is a wide variation in the symptoms of this 54 disease, which include joint pains, mood disturbance, and malaise and worsening of 55 56 symptoms following minimal physical or mental exertion. More severe symptoms can be also present including extreme exhaustion, severe joint pains with no apparent cause, 57 58 non-restorative sleep and a range of immune and neurological symptoms. These 59 symptoms may lead to depression and social isolation in the person with ME/CFS [1]. 60 The pathophysiology of the ME/CFS is not understood and there is no diagnostic 61 biomarker available. There is still controversy over the etiology of the disease; however, it is widely accepted that several immunological alterations are present in ME/CFS 62 63 patients [2]. In addition, accumulated evidence for an association of ME/CFS with viral infections also exists and many patients report the onset of their symptoms during or 64 65 right after a flu-like illness [3]. Thereafter, an unusual autoimmune response against the 66 infection would be responsible for the perpetuation of the ME/CFS symptoms. Viral 67 participation is finally supported by the evidences of clinical benefit of patients treated with valganciclovir [4]. Unfortunately, the absence of large cohort studies that 68 69 investigate at the molecular level the participation of infectious agents on the ME/CFS 70 pathogenesis impairs our understanding of this disease.

Human Endogenous Retroviruses (HERVs) are derived from exogenous retroviral infections, which occurred early in the evolution of vertebrates. Due to active replication and transposition events, HERVs are extensively distributed through the host

74 genome and constitute about 8% of the human genome [5]. Due to accumulated 75 mutations over the primate and human evolution, most HERVs are non-functional, but 76 intact open reading frames of some HERVs persist and can be reactivated in response to 77 systemic and environmental factors such as hormones, stress, and infection by exogenous viruses including almost all human herpesviruses, HIV and others [6,7]. 78 79 Given their potential pathogenic effects, which include molecular mimicry and immune 80 deregulation, HERVs are often postulated as possible causes of autoimmune diseases. 81 Among the more than 30 families, the K and W families are the most recently 82 integrated, the most active, and have been frequently associated with neurological and 83 autoimmune diseases such as multiple sclerosis, diabetes mellitus SLE, ALS and 84 rheumatoid arthritis [8]. 85 To our knowledge, only two studies have investigated the participation of endogenous 86 retroviruses in ME/CFS with contrasting results [9,10]. 87 Given the extensively described altered patterns of HERVs in several diseases and the

gap in knowledge of its expression in ME/CFS, we investigated the expression of the
HERVs K and W in patients diagnosed with ME/CFS.

90

91 Methods

92

### 93 **Participants**

94

95 We used PBMC samples from a hundred patients diagnosed with ME/CFS and stored

96 in the UK ME/CFS Biobank (UKMEB) at the London School of Hygiene and Tropical

97 Medicine in this study. The UKMEB is among the few biorepositories worldwide with

98	advanced storage and linked research infrastructure dedicated to research into ME/CFS
99	[11]. Seventy five samples were requested from participants diagnosed with moderate
100	fatigue (ME/CFSm), and 25 from participants with severe fatigue (ME/CFSs).
101	Participants with ME/CFS were defined as moderate or severely affected based on their
102	mobility: those described as severely affected were house-bound or bed-bound, while
103	those described as having mild/moderate ME/CFS were ambulatory [11]. Samples from
104	70 healthy controls (also provided by the UKMEB) were included. This study was
105	approved by LSHTM and University of São Paulo ethical committees [#EC.2017.02
106	and #2728254 respectively].
107	
108	RNA extraction and Real Time PCR
109	
110	RNA extraction from the PBMC samples was performed by the Trizol-chloroform
111	method, with 1ml Trizol and subsequent addition of chloroform to solubilize lipids

allowing its removal. The samples were centrifuged at 15,000 rpm for 15 minutes and

the upper phase containing the RNA was further used. The material was precipitated

114 with Isopropanol 100% and washed with 75% Ethanol. In both steps the material was

centrifuged at 15,000 RPM for 10 minutes at 4 °C. After this process, the pellets were

dried at room temperature for 10 minutes, and the RNA was eluted in 45µl of Nuclease-

117 Free H2O. The decontamination of remnant DNA was performed using two rounds

118 DNAse treatment (Turbo DNA-Free (Ambion) following the manufacturer's

119 instruction. The absence of DNA was confirmed by real time PCR without reverse

120 transcriptase using primers for HERV-K or HERV-W (see primers description below).

- 121 After this procedure, cDNA was synthesized using the High capacity cDNA Reverse
- 122 Transcription kit (Ambion, USA) according the manufacturer's instructions. Real-time
- 123 PCRs were performed for the HERV-W, K and the endogenous gene using the primers

- and conditions used previously by Nali et al [12] using the Sybr Green method. The
- 125 primers used are described in Table 1.
- 126
- 127
- 128
- 129 Table 1. Primers used in real-time PCR assays

Oligo	Sense	Antisense
HERV-W	CCAATGCATCAGGTGGGTAAC	GAGGTACCACAGACAAAAAATATTCCT
HERV-K	TCCCCTTGGAATACTCCTGTTTT	CATTCCTTGTGGTAAAACTTTCCA
GAPDH	ACCCACTCCTCCACCTTTGAC	TGTTGCTGTAGCCAAATTCGTT

130

131

132 The cycling conditions for both HERVs detection were: 50 °C for 2 minutes, 95 °C for 10 minutes, followed by 40 cycles of 95 °C for 15 seconds, 50 °C for 1 minute, 60 °C 133 134 for 1 minute. HERV activity was qualitatively (referred as presence/absence) and 135 quantitatively (level of expression) evaluated. As positive controls we used a plasmid containing both HERV-W envelope and HERV-K polymerase fragments correspondent 136 to the region covered by the primers used. The level of expression was determined by 137 calculation of  $2^{-\Delta\Delta_{Ct}}$ , and the results were represented as fold changes. Statistical 138 139 analysis was performed using the Wilcox test in the GraphPad Prism program v.6.04. Samples were only considered positive for HERVs and included in the analyses if 140 expression of the endogenous control was also detected. 141 142 143

144 **Results** 

145	General description of individuals included in the study is described in Table 2. As
146	expected, women were 4 times more prevalent than men. Therefore, we adjusted the
147	control group to the same gender prevalence. HERV-K and W expression were
148	evaluated in ME/CFS patients and healthy controls; and some level of expression of
149	HERV-W was detected in all patients with severe fatigue and in 72/75 ME/CFSm
150	(96%). HERV-K was also detected in all severe cases but in 65/75 of moderate cases
151	(86.6%). The healthy control group was very similar to the moderate group, with $68/70$
152	(97%) and 60/70 (85.7%) presenting expression of HERV-W and HERV-K respectively
153	(Table 2). Only one patient with moderate fatigue and one control individual had no
154	HERV activity at all. No relation was observed regarding HERV detection and duration
155	of disease.

- 156 Table 2. Main characteristics of individuals included in the study.
- 157

Participants (#)	Age median (max, min)	Gender (%)		Time of disease median (max, min)	Detection activ	
		М	F		HK	HW
ME/CFSs (25)	43.3 (25-62)	24%	76%	16.8 (2.8-40)	100%	100%
ME/CFSm (75)	22.9 (18-64)	25,4%	74,6%	11.2 (0.2 – 33.7)	86.6%	96%
Controls (70)	42.8 (19-63)	25.7%	74.3%	-	85.7%	97%
* Qualitativaly						

158 \* Qualitatively

160 Regarding to the level of expression (quantitative analysis), real time results revealed

that HERV-W did not present significant differences when the healthy controls (HCs)

162 or the two ME/CFS groups were compared between each other (Figure 1 A), i.e.

<sup>159</sup> 

- 163 ME/CFSs vs HCs (p = 0.89), ME/CFSm vs HCs (p = 0.77), ME/CFSs vs ME/CFSm (p
- 164 = 0.95), all patients ME/CFS vs HCs (p = 0.78).
- 165 On the other hand, the HERV-K expression differed significantly between ME/CFSm
- group and the HCs (p = 0.050). HERV-K activity was not distinct between the ME/CFS
- 167 groups: ME/CFSs vs ME/CFSm (p = 0.12), ME/CFS vs HCs (p = 0.17). ME/CFSs vs
- 168 HCs (p = 0.97) (Figure 1 B).
- 169 [FIGURE 1]
- 170

### 171 Discussion

The most recognized and widely-used case definitions (Fukuda [13] and Canadian Consensus criteria [1]) are based on self-reported symptoms. Studies of energy metabolism, oxidative stress and immunological alterations in ME/CFS have demonstrated imbalance in all these pathways, but the use of such information for diagnostic purposes is still far from reality.

Here, HERV-K and W transcripts were detected in all groups investigated, and we found that HERV-K was overexpressed in moderate ME/CFS. It is possible that the immunological, genic expression and metabolic alterations are different according to disease severity.

The interplay between endogenous retroviruses and the immune system is complex. ERVs are part of the host genome and in theory, they are supposed to be recognized as self-antigens and an immune tolerance should be established during the early stages of the organism development [14]. However, HERV products can interact with components of the innate immune system leading to the activation of pro-inflammatory

pathways or, in some particular cases, their suppression [15]. The syncytin -2 protein 186 187 for example, is a product of the ERV-FRD Env gene that has an immunosuppressive role by preventing maternal immune response against the fetus [16]. In a distinct 188 scenario, it was demonstrated using psoriasis model that a pro-inflammatory 189 190 environment could be able to suppress the expression of repetitive elements, including 191 HERVs [17]. It would be reasonable to suggest that the immunological enhancement 192 seen in more severe ME/CFS works by silencing the HERV transactivation that occurs in moderate cases. Such transactivation could be caused by exogenous viral replication 193 194 or another as yet unknown factor. In line with this, Montoya and colleagues (2017) 195 found a cytokine signature of severity in people with ME/CFS [18]. They demonstrated 196 that from the 17 cytokines related to severity, 13 are pro-inflammatory, and (in addition to the worsening of the symptoms) may cause the reversion of the HERV-K activity to 197 198 levels similar to those seen in healthy individuals. It may similarly occur with HERV-199 W, which, despite not being at significant levels, there was a slight decrease in people 200 severely affected by ME/CFS when compared to those who are moderately affected.

201

202 Infection has often been considered as a trigger to ME/CFS. Many patients report that 203 the fatigue began during or short after an episode of infectious disease. A number of 204 pathogens including viruses have been associated with this disease [3]. And, due to its 205 life long persistence and broad cell tropism, the herpesviridae family, particularly HHV-206 6 has been considered to be a possible trigger for ME/CFS for many, even though such 207 relationship has not been consistent [3,9]. Interestingly, HHV-6 as some other 208 herpesviruses, is also capable of transactivating HERVs, particularly, HERV-K [6]. 209 Such transactivation may be either direct (through LTR activation by viral products) or 210 indirect (via transcriptional binding factors and cytokines produced by viral replication)

[3,6]. It is possible that as the disease progresses, whatever the exogenous infection that
would have act as the trigger factor is controlled, and consequently, the HERVs
transactivation decrease. Unfortunately, we did not perform serological or molecular
tests for exogenous viruses.

215 Recently, it was suggested that differential methylation patterns of promoters in 216 ME/CFS would impact on the expression of nearby transposable elements, including 217 HERVs [19]. Two reports of HERV activity in ME/CFS were published some years ago 218 but the results were conflicting. In 2013 Oakes and his team found no difference on the 219 expression of HERV-K18 envelope in people with ME/CFS when compared with HCs 220 [9]. In the same year De Meirleir and colleagues, using immunohistochemical methods, 221 found immunoreactivity to HERV proteins (HERV-K, HERV-18, HERV-R and HERV-222 FRD) in dendritic cells of the duodenum of individuals diagnosed with the syndrome 223 [10], suggesting that alterations in endogenous retroviruses expression pattern may 224 occur in ME/CFS. The differences between the results of Oakes and colleagues and ours 225 may be due to the methods used to detect HERV-K. While the present work used 226 generic primers for HERV-K that allow the detection of hundreds of elements from 227 most HML subfamilies the Oakes team searched for the HERV-K 18 envelope 228 transcripts only, using a method specific to this particular element, while neglecting all 229 the remaining proviruses from the K family. On the other hand, we are unable to 230 determine which K family proviruses are involved in the differential expression 231 observed.

The molecular method used here to detect HERV-W was also generic and was widely used in several studies that found differential expression of this element in pathological conditions, including in the blood, brain and CSF of multiple sclerosis (MS) patients [20]. Therefore, despite the similarity of a number of symptoms and the strong

236 immunological component of ME/CFS and MS, the mechanisms responsible for HERV

237 reactivation in such diseases are likely distinct.

In conclusion, this is the first report that demonstrates increased expression of an endogenous retrovirus in the blood of individuals with moderate ME/CFS. While the increased expression of these retroelements can't be directly associated to the ME/CFS pathogeny, the observation of this phenomenon cannot be ignored.

242

243 Funding: This work was supported by Fundação de Amparo à Pesquisa do Estado de

244 São Paulo (FAPESP) [grant number 15/05958-3]. LN, EL and CK have been funded by

the National Institutes of Health (NIH USA) under award number [2R01AI103629],

however the content is solely the responsibility of the authors and does not necessarily

represent the official views of the NIH.

248

249

250

251

252

253

254

255

256

257

258

260		
261		
262		
263	Refer	rences
264 265 266	1.	Carruthers BM, Jain AK, De Meirleir KL, Peterson DL, Klimas NG, Lerner AM, et al. Myalgic Encephalomyelitis/Chronic Fatigue Syndrome. J Chronic Fatigue Syndr. 2003;11(1):7–115.
267 268 269	2.	Lorusso L, Mikhaylova S V., Capelli E, Ferrari D, Ngonga GK, Ricevuti G. Immunological aspects of chronic fatigue syndrome. Autoimmun Rev. 2009;8(4):287–291.
270 271 272	3.	Rasa S, Nora-Krukle Z, Henning N, Eliassen E, Shikova E, Harrer T, et al. Chronic viral infections in myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS). J Transl Med. 2018;16(1):268.
273 274 275	4.	Watt T, Oberfoell S, Balise R, Lunn MR, Kar AK, Merrihew L, et al. Response to valganciclovir in chronic fatigue syndrome patients with human herpesvirus 6 and Epstein-Barr virus IgG antibody titers. J Med Virol. 2012;84(12):1967–1974.
276 277	5.	Weiss RA. The discovery of endogenous retroviruses. Retrovirology. 2006;3(1):67.
278 279 280	6.	Chen J, Foroozesh M, Qin Z. Transactivation of human endogenous retroviruses by tumor viruses and their functions in virus-associated malignancies. Oncogenesis. 2019;8(1):6.
281 282 283	7.	Nellåker C, Yao Y, Jones-Brando L, Mallet F, Yolken RH, Karlsson H. Transactivation of elements in the human endogenous retrovirus W family by viral infection. Retrovirology. 2006;3(1):44.
284 285 286	8.	Gröger V, Cynis H. Human Endogenous Retroviruses and Their Putative Role in the Development of Autoimmune Disorders Such as Multiple Sclerosis. Front Microbiol. 2018 ;9:265.
287 288 289 290	9.	Oakes B, Hoagland-Henefield M, Komaroff AL, Erickson JL, Huber BT. Human Endogenous Retrovirus-K18 Superantigen Expression and Human Herpesvirus-6 and Human Herpesvirus-7 Viral Loads in Chronic Fatigue Patients. Clin Infect Dis. 2013;56(10):1394–1400.
291 292 293 294	10.	De Meirleir KL, Khaiboullina SF, Frémont M, Hulstaert J, Rizvanov AA, Palotás A, et al. Plasmacytoid dendritic cells in the duodenum of individuals diagnosed with myalgic encephalomyelitis are uniquely immunoreactive to antibodies to human endogenous retroviral proteins. In Vivo. 2013;27(2):177–187.

295 296 297 298	11.	Lacerda EM, Mudie K, Kingdon CC, Butterworth JD, O'Boyle S, Nacul L. The UK ME/CFS Biobank: A Disease-Specific Biobank for Advancing Clinical Research Into Myalgic Encephalomyelitis/Chronic Fatigue Syndrome. Front Neurol. 2018 Dec;9:1026.
299 300 301	12.	Nali LHS, Oliveira ACS, Alves DO, Caleiro GS, Nunes CF, Gerhardt D, et al. Expression of human endogenous retrovirus K and W in babies. Arch Virol. 2017;162(3):857–861.
302 303 304 305	13.	Fukuda K, Straus SE, Hickie I, Sharpe MC, Dobbins JG, Komaroff A. The chronic fatigue syndrome: a comprehensive approach to its definition and study. International Chronic Fatigue Syndrome Study Group. Ann Intern Med. 1994;121(12):953–959.
306 307 308	14.	Tugnet N, Rylance P, Roden D, Trela M, Nelson P. Human Endogenous Retroviruses (HERVs) and Autoimmune Rheumatic Disease: Is There a Link? Open Rheumatol J. 2013;7:13–21.
309 310	15.	Magiorkinis G, Hurst TP. Activation of the innate immune response by endogenous retroviruses. J Gen Virol. 2015;96(6):1207–1218.
311 312 313 314	16.	Mangeney M, Renard M, Schlecht-Louf G, Bouallaga I, Heidmann O, Letzelter C, et al. Placental syncytins: Genetic disjunction between the fusogenic and immunosuppressive activity of retroviral envelope proteins. Proc Natl Acad Sci. 2007;104(51):20534–20539.
315 316 317	17.	Lättekivi F, Kõks S, Keermann M, Reimann E, Prans E, Abram K, et al. Transcriptional landscape of human endogenous retroviruses (HERVs) and other repetitive elements in psoriatic skin. Sci Rep. 2018;8(1):4358.
318 319 320	18.	Montoya JG, Holmes TH, Anderson JN, Maecker HT, Rosenberg-Hasson Y, Valencia IJ, et al. Cytokine signature associated with disease severity in chronic fatigue syndrome patients. Proc Natl Acad Sci. 2017;114(34):E7150–8.
321 322 323 324	19.	Almenar-Pérez E, Ovejero T, Sánchez-Fito T, Espejo JA, Nathanson L, Oltra E. Epigenetic Components of Myalgic Encephalomyelitis/Chronic Fatigue Syndrome Uncover Potential Transposable Element Activation. Clin Ther. 2019;41(4):675-698.
325 326 327 328	20.	Morandi E, Tanasescu R, Tarlinton RE, Constantinescu CS, Zhang W, Tench C, et al. The association between human endogenous retroviruses and multiple sclerosis: A systematic review and meta-analysis. PLoS One. 2017;12(2):e0172415.
329		
330		
331		

- Figure 1. Boxplot of expression levels (in fold change) of HERVs among the groups. A)
- HERV-K and B) HERV-W. Significance between the groups (obtained by Wilcox test)
- is evidenced by an asterisk.

337





