

1 **Title**

2 Axial variation of deoxyhemoglobin density as a source of the low-frequency time lag
3 structure in blood oxygenation level-dependent signals

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23 coupling, Resting-state fMRI, Vasomotion

24

25

26 **Abstract**

27 Perfusion-related information is reportedly embedded in the low-frequency component of a
28 blood oxygen level-dependent (BOLD) functional magnetic resonance imaging (fMRI)
29 signal. The blood-propagation pattern through the cerebral vascular tree is detected as an
30 interregional lag variation of spontaneous low-frequency oscillations (sLFOs). Mapping of
31 this lag, or phase, has been implicitly treated as a projection of the vascular tree structure
32 onto real space. While accumulating evidence supports the biological significance of this
33 signal component, the physiological basis of the “perfusion lag structure,” a requirement for
34 an integrative resting-state fMRI-signal model, is lacking. In this study, we conducted
35 analyses furthering the hypothesis that the sLFO is not only largely of systemic origin, but
36 also essentially intrinsic to blood, and hence behaves as a virtual tracer. By summing the
37 small fluctuations of instantaneous phase differences between adjacent vascular regions, a
38 velocity response to respiratory challenges was detected. Regarding the relationship to
39 neurovascular coupling, the removal of the whole lag structure, which can be considered as
40 an optimized global-signal regression, resulted in a reduction of inter-individual variance
41 while preserving the fMRI response. Examination of the T2* and S₀, or non-BOLD,
42 components of the fMRI signal revealed that the lag structure is deoxyhemoglobin
43 dependent, while paradoxically presenting a signal-magnitude reduction in the venous side
44 of the cerebral vasculature. These findings provide insight into the origin of BOLD sLFOs,
45 suggesting that they are highly intrinsic to the circulating blood.

46

47 **1 Introduction**

48 In functional magnetic resonance imaging (fMRI), there are 2 established physiological
49 bases of signal change: neurovascular coupling (NVC) and autoregulation. The former
50 involves a local blood flow increase of 30–70% which gives rise to a 0.5–2% blood

51 oxygenation level-dependent (BOLD) signal increase with around a 5-s delay (Buxton,
52 2013). This is the target phenomenon of fMRI as a tool for brain mapping, due to its limited
53 spatial extent mainly involving local arterioles (Hillman, 2014). In contrast, autoregulatory
54 responses in vessel diameter are found in a wide range of arteries including the internal
55 carotid or middle cerebral arteries (Hoiland *et al.*, 2016). Detection of a compromised
56 response in vascular disorders has proven useful for clinical purposes (Murphy *et al.*, 2011).
57 Importantly, there is no clear physiological distinction between these 2 phenomena as each
58 involves multiple pathways (Willie *et al.*, 2014). Their traces in the fMRI signal are also
59 uniformly postulated to reflect the increased cerebral blood flow and eventual dilution of
60 deoxy-hemoglobin (Hb) in the postcapillary part of the vasculature, with the additional
61 effect of a local blood volume increase (Kim & Ress, 2016).

62 In efforts to improve the efficiency of fMRI, studies have revealed various systemic
63 physiological components in BOLD signal fluctuations. Physiological parameters, such as
64 cardiac pulsation (Chang *et al.*, 2009), blood pressure, and end-tidal carbon dioxide (CO₂)
65 (Wise *et al.*, 2004; Murphy *et al.*, 2013), are considered artifact sources. The contamination
66 is expected to be emphasized in resting-state fMRI (rs-fMRI), where signals are evaluated
67 without trial averaging (Winder *et al.*, 2017). However, discrimination between neuronal
68 and non-neuronal components has been a major challenge due to the lack of validation
69 techniques with spatial and temporal precision comparable to that of fMRI. Another source
70 of difficulty is the fact that many neural and non-neural parameters are intercorrelated in
71 this low-frequency range (Murphy *et al.*, 2013).

72 A focus of recent studies exploring this matter is the spontaneous BOLD low-
73 frequency oscillation (sLFO, < 0.1 Hz) possibly encompassing multiple artifact sources
74 (Zhu *et al.*, 2015; Tong *et al.*, 2017). One well-known sLFO source is the respiratory
75 volume fluctuation involving a chemoreflex loop (Birn *et al.*, 2008). An sLFO in systemic
76 blood pressure, known as the Traube–Hering–Mayer wave, has been shown to originate
77 from another autonomic loop (Guyton & Harris, 1951; Julien, 2006). Moreover, associated
78 sLFOs in blood flow and velocity have been found (Killip, 1962; Fagrell *et al.*, 1977) and,
79 later, transcranial Doppler ultrasonography and optical methods confirmed their traces

80 within the brain (Giller *et al.*, 1999; Obrig *et al.*, 2000). This optically detected sLFO was
81 found in both oxy- and deoxy-Hb with an interesting phase difference exclusively observed
82 in the brain (Rayshubskiy *et al.*, 2014; Tgavalekos *et al.*, 2016). Additionally, sLFOs were
83 found in electroencephalographic recordings, for which arterial vasomotion was suggested
84 as the origin (Nikulin *et al.*, 2014), although it is unclear how the vasomotion accounts for
85 the fMRI signal mainly from the capillary bed.

86 As mentioned above, Hb-sLFOs are postulated to be of systemic origin (Katura *et al.*,
87 2006; Tian *et al.*, 2011; Sassaroli *et al.*, 2012). Hence, it was not surprising to find a
88 correlation between the global fMRI signal and extra-cerebral signals (near-infrared
89 spectroscopy [NIRS] or MRI), but the constant time shift across body/brain parts was
90 unexpected (Anderson *et al.*, 2011; Tong *et al.*, 2012). The similarity between the low-
91 frequency phase map and perfusion MRI in healthy participants was a milestone in this
92 direction, as it presented the perfusion time lag embedded in the BOLD signal (Tong *et al.*,
93 2017). The resilient nature of the lag map against the fMRI task condition was shown,
94 further supporting its non-neuronal origin (Aso *et al.*, 2017a). In parallel, a number of
95 clinical studies have established the phase delays as a marker of cerebrovascular disorders
96 (Amemiya *et al.*, 2013; Lv *et al.*, 2013; Christen *et al.*, 2015; Ni *et al.*, 2017; Nishida *et al.*,
97 2018; Khalil *et al.*, 2018). Moreover, gross vascular anatomy has been detected consistently
98 in these studies, replicating the results from respiratory challenges (Chang & Glover,
99 2009a; Blockley *et al.*, 2011), which importantly suggests an equivalence between the
100 sLFO and manipulated circulatory turbulences. Apart from patient data, a recent study
101 involving healthy participants revealed changes in venous drainage patterns with normal
102 aging (Satow *et al.*, 2017). A body of evidence thus empirically supports the biological
103 significance of the low-frequency lag structure and its underlying principles.

104 The current analytical model of the BOLD lag structure assumes the presence of this
105 signal variation from the very early stages of cerebral perfusion (Tong *et al.*, 2018) (**Fig.**
106 **1A**). Such synchronized variation should naturally affect the global mean signal, and the
107 model is hence related to the unresolved fMRI global signal problem (Liu *et al.*, 2017;
108 Power *et al.*, 2017). This view is not only compatible with the occasional favorable effect

109 of global signal regression (GSR) in task fMRI (Aguirre *et al.*, 1998) but has led some
 110 scientists to propose the removal of the lag structure as an approach for noise elimination
 111 (Erdoğan *et al.*, 2016; Amemiya *et al.*, 2016; Byrge & Kennedy, 2018). Under this model,
 112 the elimination of the lag structure can be viewed as an optimized GSR (of the low-
 113 frequency component) for each voxel group. Conversely, with the presence of the BOLD
 114 lag structure, a simple GSR should retain a residual correlation between in-phase voxels
 115 that confounds fMRI analyses (Taylor Webb *et al.*, 2013; Erdoğan *et al.*, 2016).

116 The fundamental and critical question remaining is the mechanism by which sLFOs
 117 (or respiratory maneuvers) create the BOLD lag structure. The BOLD response to neural
 118 activity via NVC is a well-documented passive process, involving the expansion of the
 119 intravascular compartment (Herman *et al.*, 2009). A typical model of the BOLD signal
 120 change is described as follows (Hoge *et al.*, 1999):

$$121 \quad \frac{\Delta \text{BOLD}}{\text{BOLD}_0} = M \left(1 - \left(\frac{\text{CMRO}_2}{\text{CMRO}_{2|0}} \right)^\beta \left(\frac{\text{CBV}}{\text{CBV}_0} \right) \left(\frac{\text{CBF}}{\text{CBF}_0} \right)^{-\beta} \right) \quad [1],$$

122 where CMRO_2 stands for the cerebral metabolic rate of oxygen in a voxel and CBV/CBF
 123 represents the cerebral blood volume and flow, respectively. Beta (β) is an exponent of the
 124 power-law describing the relationship between $T2^*$ and the deoxy-Hb amount that only
 125 depends on CMRO_2 and CBF in a reciprocal manner, under the assumption of a negligible
 126 inflow of deoxy-Hb. M is the factor for the BOLD susceptibility effect, defined as:

$$127 \quad M \equiv TE \cdot B_0 \cdot \text{CBV}_0 \cdot [\text{deoxy-Hb}]_{V0}^\beta \quad [2],$$

128 where B_0 is the main magnetic field strength and TE represents the echo time. Triggered by
 129 vasodilation of the arteriole, this effect is diminished by an inflow of fresh blood, which
 130 increases the MR signal. This baseline BOLD effect has been modeled in the formula for
 131 the off-resonance frequency shift ($\delta\omega$) as follows (Yablonskiy & Haacke, 1994; An & Lin,
 132 2002):

$$133 \quad \delta\omega = \gamma \cdot \frac{4}{3} \cdot \pi \cdot \Delta\chi_0 \cdot \text{Hct} \cdot \text{OEF} \cdot B_0 \quad [3],$$

134 where γ is the gyromagnetic ratio (42.58 MHz/T), $\Delta\chi_0$ is the susceptibility difference
 135 between the fully oxygenated and fully deoxygenated blood, and Hct is the hematocrit

136 (volume fraction of erythrocytes to the blood volume, typically around 40%). The oxygen
 137 extraction fraction (OEF) represents the only source of deoxy-Hb under the assumption of
 138 100% oxygen saturation (SaO_2) in the inflow.

139 Variations of this base susceptibility can occur due to the local Hct and SaO_2 changes
 140 and, in fact, have been shown to cause intersession variabilities (Cohen *et al.*, 2002;
 141 Tuunanen & Kauppinen, 2006); however, within-session fluctuations have rarely been
 142 considered (Thomas *et al.*, 2000). For example, even at a constant Hb density and oxygen
 143 partial pressure, CO_2 fluctuation, a driving factor of vasomotion, alone can modify SaO_2
 144 through pH changes (Collins *et al.*, 2015a). Although the assumption of constant base
 145 deoxy-Hb concentration may be sufficient for modeling its dilution by NVC (Ogawa *et al.*,
 146 1998), other effects might not be negligible in non-neuronal fluctuations (**Fig. 1b**).

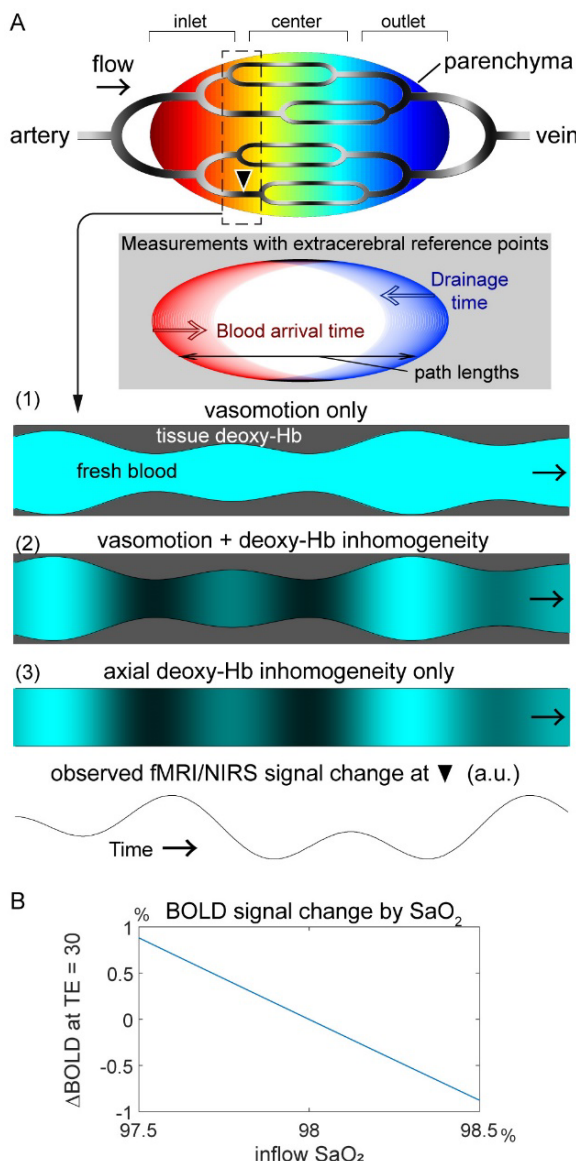


Fig. 1. A working model of BOLD lag mapping. (A) A schematic of the cerebral vascular tree presenting possible physiological models that account for the fMRI and NIRS observations. Each brain voxel has its lag (phase) of the spontaneous low frequency oscillation (sLFO) relative to the reference point, depicted in warm and cool colors. The inlet, center, and outlet parts of the gross vasculature are determined purely by the temporal relationship and not by vessel types; all signals should originate from the capillary bed. Another schematic is inserted to illustrate the effect of the reference point selection in relation to the vascular path length, which accounts for the disagreement with other blood-tracking techniques. Below are 3 models with different physiological signal sources

167 that are moving along the vasculature. The current model with constant deoxy-Hb
168 requires vasomotion to account for the sLFO (1). While the axial inhomogeneity of
169 the deoxy-Hb density may be linked with vasomotion (2), it can persist without it (3).
170 The moving axial inhomogeneity creates a similar temporal profile with varying
171 phases across regions. (B) Dependence of the BOLD signal on inflow oxygen
172 saturation (SaO_2) at 3 T, TE = 30 ms. All other parameters were held constant at
173 physiological values: oxygen extraction fraction = 45%, tissue blood volume fraction
174 = 3%, and hematocrit = 40% (see Eqs. [1] and [2]).

175 BOLD, blood oxygen level-dependent; fMRI, functional magnetic resonance
176 imaging; NIRS, near-infrared spectroscopy; TE, echo time; Hb, hemoglobin

177

178 In this exploratory study, we conducted 3 investigations to further advance our
179 knowledge on the BOLD lag structure and its underlying physiology. We first evaluated if
180 changes in blood transit velocity are embedded in the BOLD low-frequency phase to
181 confirm its behavior as a virtual tracer [a preliminary analysis of these data was presented
182 previously as a poster (Aso *et al.*, 2017b)]. Next, we investigated the effect of eliminating
183 the lag structure from task-based fMRI, which had not been tested previously. Finally, we
184 used multi-echo imaging to assess the components of the BOLD signal that determine the
185 lag structure. One of the recent approaches toward fMRI denoising has focused on S_0
186 fluctuations (signal at TE = 0, which is the baseline MR signal from the fluid
187 compartments), as it is weakly associated with neural activity (Posse *et al.*, 1999; Wu *et al.*,
188 2012; Kundu *et al.*, 2012; Yen *et al.*, 2017). In contrast, the total-Hb sLFO, detected by
189 NIRS, is interpreted as a local CBV change under the assumption of a constant Hct, which
190 should also affect the non-BOLD component via changes in plasma volume and inflow
191 (Rostrup *et al.*, 2005). Notably, the contributions of the T2* and S_0 components may differ
192 from NVC contributions to the BOLD lag structure; hence their impact may have been
193 overlooked in studies based on trial averaging. From the influences of 3 different fMRI task
194 paradigms, including a simple reaction-time visuomotor task, a short breath-holding task,
195 and a hyperventilation task on the neural and non-neural components of the fMRI signal,
196 we sought further validation of our hypothetical model of the BOLD lag structure.

197 **2 Materials and methods**

198 **2.1 Participants and experimental procedures**

199 Twenty-one healthy participants (8 women, 19–26 years of age) participated in Experiment
200 1; only 1 person was excluded from the analysis because of an abrupt head motion, which
201 prevented BOLD lag mapping (see Data processing). The remaining 20 participants
202 performed the sparse visuomotor and 10-s breath-holding tasks, but the hyperventilation
203 task was only performed by 18 participants, as it was introduced after the first 2 individuals
204 had concluded their participation. Another 21 participants were recruited for Experiment 2,
205 involving multi-echo acquisition, all of whom performed the above 3 tasks but with an 18-s
206 version of the breath-holding task. To avoid vigilance level fluctuations, all MRI sessions
207 were scheduled in the morning and the participants were encouraged to sleep well the
208 previous night.

209 The protocol for this study was approved by the internal ethics review board of Kyoto
210 university. The participants provided written informed consent in advance, according to the
211 Declaration of Helsinki, for the analysis of anonymized MRI scans and simultaneously
212 acquired physiological data.

213 **2.2 Image acquisition**

214 A Tim-Trio 3-Tesla scanner (Siemens, Erlangen, Germany) with a 32-channel phased-array
215 head coil was used for MRI acquisition. For Experiment 1, T2*-weighted echo-planar
216 images were acquired using multiband gradient-echo echo-planar imaging (EPI) (Feinberg
217 *et al.*, 2010) with the following parameters to cover the entire cerebrum: 64×64 pixels, 35-
218 slice interleave, 192-mm field of view (FOV), 3.5-mm slice thickness, repetition time
219 (TR)/ TE = 500/35 ms, flip angle = 40° , and a multiband factor of 5. Three 9-min runs
220 (1,080 volumes) were acquired for each of the 3 task conditions. The same pulse sequence
221 program was used in Experiment 2 but with multi-echo settings: TE1 = 7.76 ms and TE2 =
222 25.82 ms for the first 6 participants and TE1 = 11.2 ms and TE2 = 32.78 ms for the
223 remaining 15 participants. A smaller multiband factor of 2 was selected to allow for the

224 short TE in combination with parallel imaging using GeneRalized Autocalibrating Partial
225 Parallel Acquisition. Other acquisition parameters were: TR = 1,300 ms, flip angle = 65°;
226 36-slice interleave, FOV = 256 × 192 mm², 64 × 48 matrix, and 3.5-mm slice thickness.
227 Three 7-min (323 TR) runs were acquired. Seven participants in Experiment 2 underwent 2
228 additional runs with a shorter TR of 700 ms and a flip angle of 45° to examine the
229 sensitivity of the respiration-related signal component to inflow modulation. At the end of
230 every experimental session, a 3-dimensional (3D) magnetization-prepared rapid acquisition
231 with gradient echo (MPRAGE) T1-weighted image was acquired for obtaining anatomical
232 information (Aso *et al.*, 2017a). A dual-echo gradient-echo dataset for B₀ field mapping
233 was also acquired after the BOLD scan in the same orientation.

234 **2.3 Task conditions**

235 Throughout the experimental session, task instructions were presented via a liquid crystal
236 display (LCD) monitor inside the scanner room, viewed through a mirror. Beat-to-beat
237 fluctuations in the mean arterial pressure and heart rate were obtained via a non-invasive
238 MR-compatible device (Caretaker, BIOPAC Systems, Inc., Goleta, CA, USA). Careful
239 instructions were provided to the participants on how to avoid motion, especially during the
240 respiratory challenges.

241 **2.3.1 Sparse visuomotor task**

242 A simple visuomotor task with a varying intertrial interval of 6 to 24 s was performed
243 during the first run. Participants were instructed to press a button with their right index
244 finger as soon as the computer screen changed from “Please hold still” to “Press the button.”
245 The screen returned to “Please hold still” at the button press or after 3 s, if the participant
246 had not pressed the button.

247 **2.3.2 Breath-holding task**

248 To minimize head motion induced by the tasks, the 2 respiratory challenges were adapted
249 to be less strenuous than in earlier studies. In Experiment 1, the breath-holding task was
250 cued by a “Hold your breath” instruction on the screen, at which point the participants were
251 asked to immediately hold their breath, irrespective of the respiration phase. The holding

252 periods lasted 10 s and were separated by 90-s intervals. This short duration was selected to
253 minimize strain that can cause body movements, while evoking a detectable autoregulatory
254 response (Murphy *et al.*, 2011). In Experiment 2, a longer holding period of 18 s after a
255 brief inhalation for 2 s was used to evoke a more pronounced vasodilation.

256 **2.3.3 Hyperventilation task**

257 The hyperventilation task involved paced breathing at 0.2 Hz for 25 s, separated by a 30-s
258 rest. Each 5-s cycle began with the screen presenting “Please inhale” for 1.5 s, followed by
259 “Please exhale slowly,” lasting 3.5 s. Participants were instructed to breathe as deeply as
260 possible, while avoiding head movement. A short inspiration period was selected to
261 suppress motion by minimizing movements in the thoracic cage and spine.

262 **2.4 Data processing**

263 For image processing, SPM12 (Wellcome Department of Cognitive Neurology, London,
264 United Kingdom) and FSL5 (FMRIB Software Library, www.fmrib.ox.ac.uk/fsl) (Smith *et al.*, 2004) were used in combination with in-house MATLAB scripts. Off-resonance
265 geometric distortions in the EPI data were corrected using FUGUE/FSL with B_0 field maps.
266 After inter-scan slice-timing correction, head motion was compensated by 3D motion
267 correction and data repair (Mazaika *et al.*, 2009). The repairing procedure aimed to remove
268 motion-related signal dropout and involved searching for time points satisfying 2 stringent
269 criteria: (1) global signal changes between consecutive volumes exceeding 1% and (2) head
270 displacement exceeding a Euclidian distance of ± 1 mm or $\pm 1^\circ$ rotation per TR. The
271 affected time points were replaced with linearly interpolated values, but this procedure was
272 required in only 6 of the 41 participants.

274 The data were further cleaned by regressing out 24 head motion-related parameters.
275 Unlike in previous studies, the 6 rigid-body parameter time series were not directly used
276 because of the possible contamination of the motion parameters with the global signal when
277 the participants were immobile (Freire & Mangin, 2001). We used the first temporal
278 derivatives of the motion parameters, their versions after being shifted by 1 TR, and the
279 squares of those 12 time series (Satterthwaite *et al.*, 2013). Images were spatially

280 normalized to the template space using the T1-weighted anatomical image and resliced to a
281 4-mm isotropic voxel size to achieve a high voxel temporal signal-to-noise ratio.

282 **2.4.1 Lag mapping**

283 A recursive technique was used (Aso *et al.*, 2017a) after temporal bandpass filtering at
284 0.008–0.07 Hz to ensure that the phase was uniquely determined within the cross-
285 correlation range. Whereas lag was tracked up to 7 s for most analyses, it was limited to 4 s
286 in both directions, upstream and downstream, for the calculation of the relative BOLD
287 transit time (rBTT, see below). This shorter tracking range allowed a higher cutoff
288 frequency (0.12 Hz) to preserve the high-frequency component in the velocity change
289 profiles.

290 The global mean signal was used to select the initial seed that defined the reference
291 phase (lag = 0). First, voxels presenting a cross-correlogram peak at 0 with the global signal
292 were determined. The time course averaged over this set of voxels served as the initial
293 reference. In each step of the recursive procedure toward up- and downstream, a cross-
294 correlogram was calculated between the time series obtained from the previous seed voxels
295 and every undetermined voxel to find a set of voxels with a peak at ± 0.5 s, which then
296 served as the new seed. This tracking part retained some voxels without any lag values
297 because cross-correlogram peaks < 0.3 were not used following earlier works (Tong *et al.*,
298 2017). These voxels were later filled 1 by 1 with average phases from voxels with similar
299 time courses and correlation coefficients > 0.3 . There were single isolated holes even after
300 this procedure, which were filled by linear interpolation, using the 6 neighbors. There is a
301 concern that this correlation coefficient threshold is too low to accurately claim a
302 significant correlation. However, recursive tracking involves finding the cross-correlogram
303 peak precisely at ± 0.5 s, which conveys different information from its height. Besides,
304 most earlier works empirically supporting the biological significance of this phenomenon
305 involved no thresholding. When the threshold correlation coefficient was increased to 0.6,
306 most brain voxels required the hole-filling procedure, but we still obtained lag maps (by
307 between-voxel intra-class correlation (2,1) > 0.4) in 16 out of 20 participants during resting
308 state and 17 during 10-s breath holding.

309 **2.4.2 fMRI analysis on “cleaned” BOLD datasets**

310 Individual BOLD data from Experiment 1, after the above pre-processing steps including
311 the motion parameter regression, served as the reference or “raw” dataset. GSR with a low-
312 pass filtered global signal, a normal GSR, global scaling implemented in SPM12, and
313 without the perfusion lag structure (“deperfused”) were compared with the raw dataset.

314 The GSR involves regression by the global signal and extracting the residuals in each
315 voxel. In deperfusing, instead of the uniform regressor, a corresponding time series was
316 used for each voxel group by the lag value (“dynamic” GSR) (Erdoğan *et al.*, 2016). For
317 global scaling, the raw dataset was entered into the same SPM pipeline, except for the
318 option of internally dividing each volume by its global mean signal instead of the constant
319 session mean. The normal GSR and global scaling thus affected all frequency ranges,
320 whereas the GSR and deperfusing removed only the low-frequency components.

321 Random effects analysis (Friston *et al.*, 2002) was used to evaluate the effect of these
322 procedures on the fMRI results from the standard analysis framework. For the visuomotor
323 task, the neural response to each trial was modeled as an event of 0.5 s in duration. For the
324 hyperventilation condition, a boxcar with a 25-s duration modeled the activation related to
325 volitional respiratory control. Similarly, both the onset and offset timing of breath holding
326 were used to model the time-locked neural activity. The canonical hemodynamic response
327 function was convolved to the model time series to create the regressors of interest. The
328 threshold for all activation maps was $p = 0.05$ after correcting for multiple comparisons,
329 using family-wise error across the whole brain (Poline *et al.*, 1997).

330 **2.4.3 Relative BOLD transit time**

331 **Fig. 3A** illustrates the method. This analysis was performed on data from Experiment 1,
332 acquired at a short TR of 0.5 s. The lag structure consisted of a lag map and the set of time
333 series averaged over the voxels with the same phase. This structure represents the
334 propagation of the sLFO phase along the vessels. The phase is expected to move across
335 adjacent regions of the vascular tree every 0.5 s, the lag tracking step, on average. However,
336 if there is variation in propagation velocity, there would be a deviation of the instantaneous

337 phase difference from 0.5. Here, the phase difference is supposed to reflect the time the
338 blood requires to cross the boundary between the neighboring voxel groups.

339 Based on this supposition, the phase difference fluctuation was calculated from each
340 of the 16 pairs of seeds corresponding to lags of -4 to +4 s at 0.5-s intervals. Due to the
341 broad frequency range of the fluctuations of interest, we chose a smooth sliding window
342 algorithm (window length = 30 s, Kaiser window with a β value of 4) instead of using
343 analytic methods (e.g., Hilbert transform). The region time series were resampled to a 0.02-
344 s resolution to capture minute phase difference fluctuations (4%) from 0.5 s. Instantaneous
345 phase differences from each pair of seeds were averaged over respiratory task events and
346 divided by 0.5 to obtain the time course of the rBTT. The rBTTs from the 16 pairs of
347 neighboring regions were then averaged to obtain the regional or global rBTT. According
348 to this model, the inverse of the rBTT corresponded to the instantaneous velocity (relative
349 to the baseline average velocity), as the rBTT should reflect the average time required to
350 traverse fixed distances.

351 **2.4.4 Multi-echo combination**

352 For Experiment 2, involving multi-echo acquisition, S_0 and T_2^* datasets were created by a
353 simple estimation used in earlier works (Posse *et al.*, 1999; Yen *et al.*, 2017). We assumed
354 a single compartment monoexponential decay of the MR signal as follows:

355

$$356 \quad S(TE) = S_0 \exp(-TE/T_2^*) \quad [4],$$

357

358 where T_2^* and S_0 were calculated for each TR as follows (Kundu *et al.*, 2012; Posse *et al.*,
359 1999):

360

$$361 \quad T_2^* = (TE_2 - TE_1)/\ln(S_1/S_2) \quad [5], \text{ and}$$

362

363
$$S_0 = S_1^{TE2/(TE2-TE1)} / S_2^{TE1/(TE2-TE1)} \quad [6],$$

364

365 where S_1 and S_2 are the acquired signals at TE1 and TE2, respectively. Negative or T2*
366 values exceeding 100 ms were considered as noise and were ignored. All four datasets (S_0 ,
367 S_1 , S_2 or BOLD, and T2*) were entered into the same analysis pipeline used for Experiment
368 1 while accounting for the different TR value of 1.3 s. The following analyses were
369 performed on a resampled time course with a TR of 0.5 s.

370 The interaction between the signal component and vascular anatomy was examined
371 by extracting the signal time series from the inlet, center, and outlet parts of the vascular
372 tree, based on the individual lag map created from the BOLD (i.e., at TE2) dataset. By
373 exploiting the longer lag tracking range (± 7 s) than the one used in the rBTT analysis in
374 Experiment 1, the center region in this analysis covered a wider range (± 2.5 s). Using the
375 JMP12 software (SAS Institute, Cary, NC), the magnitude and phase of the regional signals
376 were analyzed by repeated-measures ANOVA followed by post hoc analyses, using
377 Tukey's honestly significant difference (HSD) test. Statistical significance was set at $p <$
378 0.05. Additional analyses were performed to examine the origins of the signal components,
379 including a region-of-interest analysis and an SPM analysis of respiration phase-related
380 small S_0 fluctuations.

381 **3 Results**

382 The average root mean square of the head motion was measured as the framewise
383 displacement (i.e., the shift in the position of the brain in 1 volume compared to the
384 previous volume), was 0.039 ± 0.007 mm (mean \pm standard deviation (SD) over
385 participants) for Experiment 1, with a maximum displacement of 0.37 ± 0.18 mm, and
386 0.034 ± 0.015 mm for Experiment 2, with a maximum of 0.24 ± 0.12 mm (Van Dijk et al.,
387 2012).

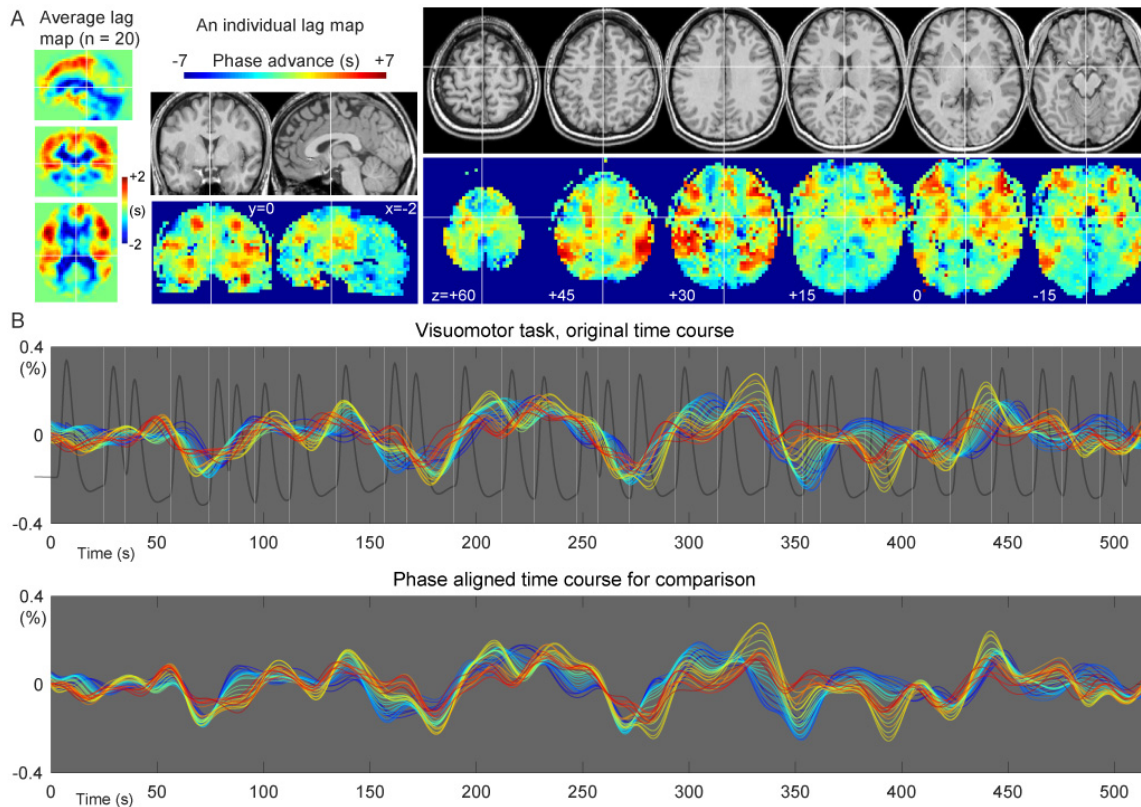


Fig. 2. Representative spatial and temporal profiles of the BOLD lag structure. (A) A lag map created by tracking the BOLD sLFO phase up to 7 s toward both up- and downstream is shown. The group average map shows the gross vascular anatomy with the early phase (positive values) distributed in the middle cerebral artery territories. The individual map provides more detailed information. (B) Seed time courses updated at each step (0.5 s) of the recursive lag tracking procedure, representing the temporal aspect of the lag structure, are shown. The warm-colored traces with advanced phases originate from the voxels with the same colors in the lag map, corresponding to the inlet or arterial side of the vasculature. A gradual change in the temporal profile is noted on top of the slow component, which is stable across regions. The white vertical lines indicate the timing of the visuomotor task, whose NVC was modeled by the hemodynamic response function (dark gray trace). The detected LFO was poorly correlated with the task-related fluctuation of neuronal origin (see main text).

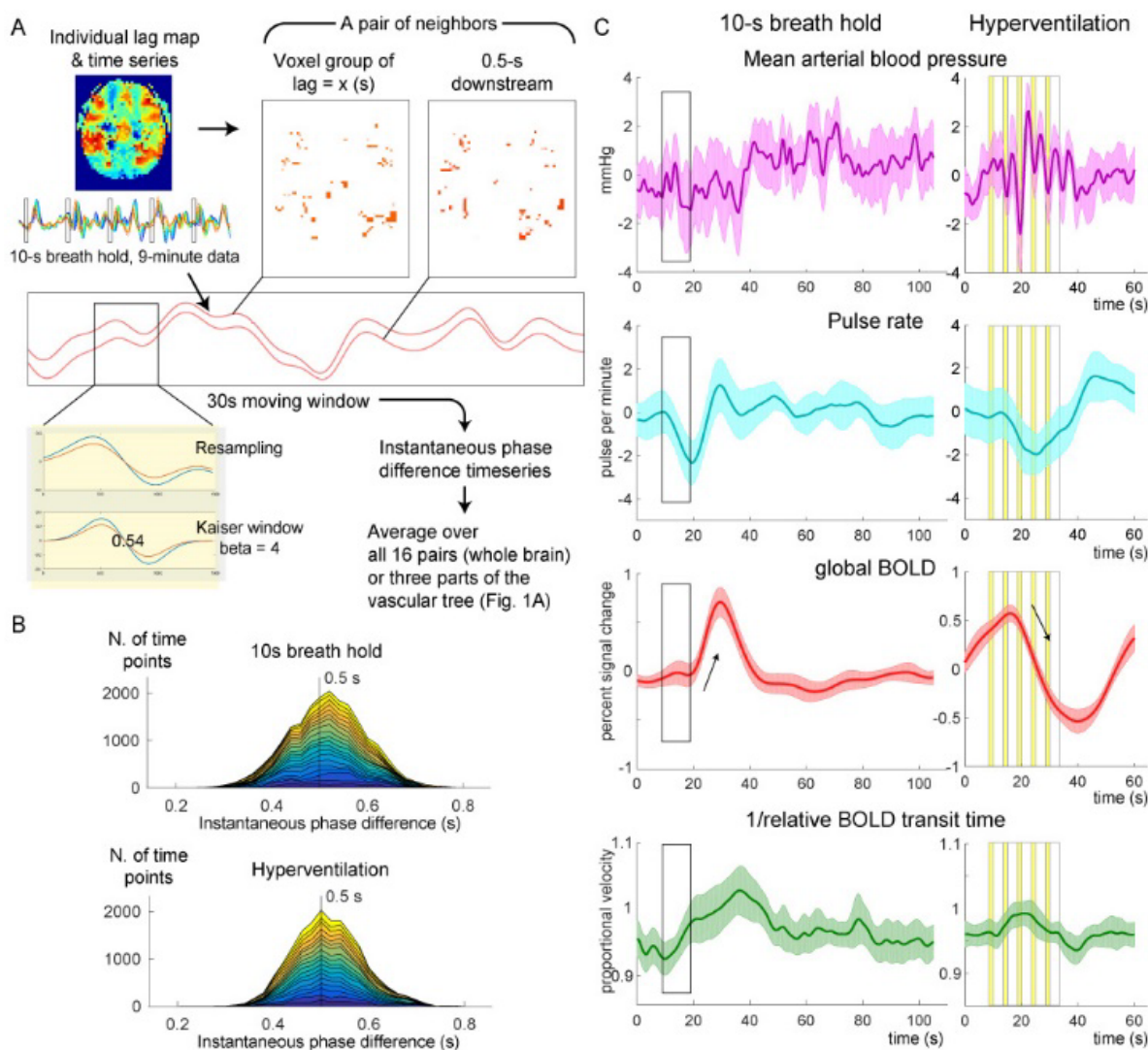
BOLD, blood oxygen level-dependent; NVC, neurovascular coupling

404

405 **Fig. 2A** shows an individual and the average BOLD lag maps during the sparse visuomotor
406 task. Warm colors indicate a positive travel time from those voxels to the phase of the
407 global LFO, signifying that the voxels are considered “upstream.” Most brain voxels fell
408 into the -4 to +4 s range (mean \pm SD, $93.8 \pm 2.9\%$). The recursively defined seed time
409 courses are shown in **Fig. 2B** with warm colors indicating averaged time series from the
410 upstream voxels. This time course of the lag structure was poorly correlated with the
411 visuomotor task (white vertical lines) or the evoked NVC response (trace in dark gray),
412 supporting a non-neuronal origin of the sLFO. This was confirmed by the correlation
413 coefficient between the global mean signal and the modeled response that was not
414 significantly different from 0 (0.0265 ± 0.140 , mean and SD over 20 participants), although
415 still slightly positive in some participants and its regression affected the activation maps
416 (see section 3.2). In addition to the constant phase shift across regions, the lag structure
417 time courses presented minute fluctuations of the phase difference (i.e., the temporal
418 relationship between the lines) over time.

419 **3.1 Flow velocity information in the instantaneous phase**

420 A transient change in the propagation velocity of the low-frequency phase, obtained as the
421 inverse of the global rBTT, was found in response to the respiratory challenges. **Fig. 3B**
422 shows the stacked histograms of the distribution of the instantaneous phase difference
423 measured by the moving window analysis. This instantaneous phase difference is
424 interpreted as the time the blood takes to move over a unit distance that requires 0.5 s on
425 average. The time courses are presented in **Fig. 3C**. After approximately 10 s of delay, an
426 increase and a decrease of the BOLD signal was found during breath holding and
427 hyperventilation, respectively (arrows).



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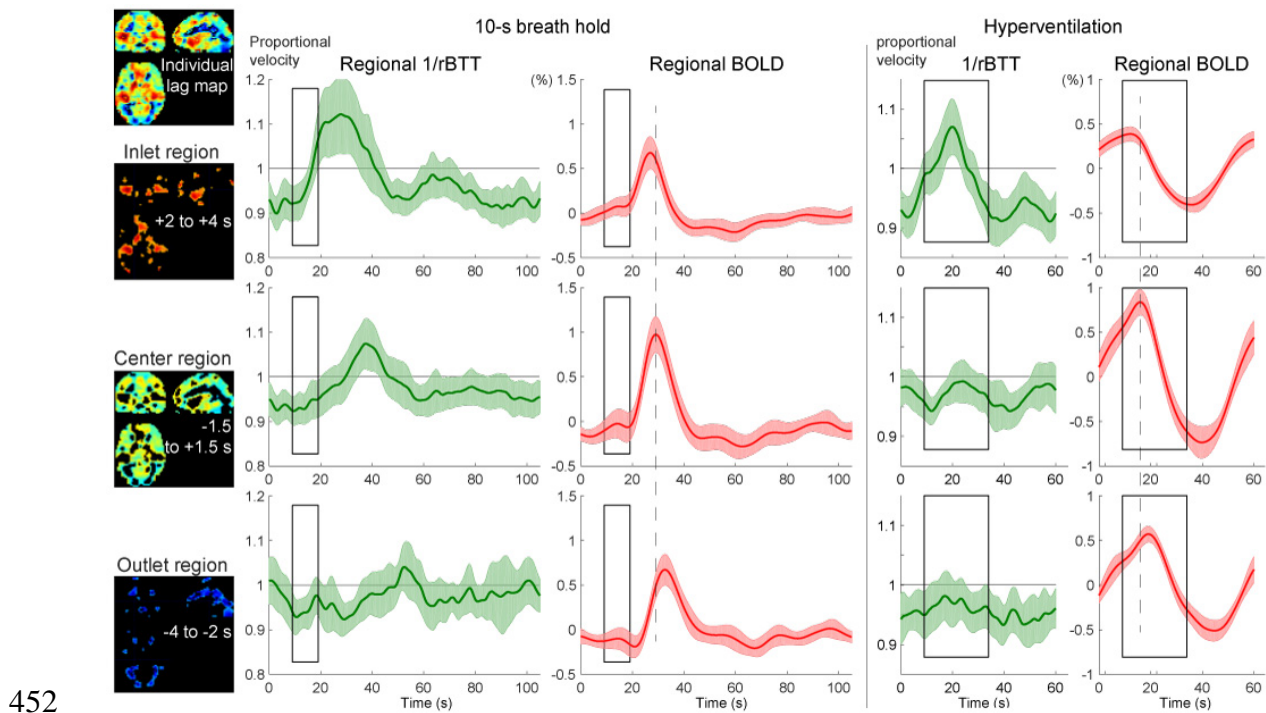
Fig. 3. Measurements of the relative BOLD transit time (rBTT). (A) A schematic illustration of the analysis is presented. For each pair of voxel groups representing neighboring regions of the vascular tree, the 2 average time series were fed into a moving window analysis of instantaneous phase difference. (B) Time point histograms of the instantaneous phase difference averaged over 16 pairs of neighboring regions in the lag structure are shown. Twenty participants are stacked. Deviation of the phase difference from 0.5 s reflects a fluctuation of the flow velocity, although the slight shift of the peak is observed in the breath hold data presumably due to windowing. (C) Responses to the respiratory challenges of the physiological recordings, raw BOLD signal, and inverse of the rBTT that can be interpreted as relative velocity are shown. Thin arrows indicate the autoregulatory response of the raw BOLD signal. The 5

440 cycles of volitional breathing for hyperventilation are indicated with yellow bands
441 representing the inspiration phase. Shaded areas indicate the 95% confidence interval
442 of the mean across participants.

443 BOLD, blood oxygen level-dependent

444

445 The temporal profile of the instantaneous velocity (green curves) was roughly in
446 phase with the global BOLD response but had different onset and peak timings, indicating
447 different physiological bases. To evaluate the relationship of this phenomenon with the
448 vascular structure, the inlet, center, and outlet regions were separately analyzed (**Fig. 4**). In
449 both respiratory challenges, there was a clear asymmetry over the vasculature, with
450 pronounced velocity responses in the inlet region. Despite similar BOLD response profiles,
451 the velocity response was not clearly found in the outlet region.



452

453 **Fig. 4.** Autoregulatory responses to respiratory challenges in the 3 vascular regions,
454 using the same conventions as in **Fig. 3**. The inlet (arterial side), center, and outlet
455 (venous side) of the parenchymal vasculature were defined for each participant, using
456 the lag map created from the session data. The relative changes in propagation velocity

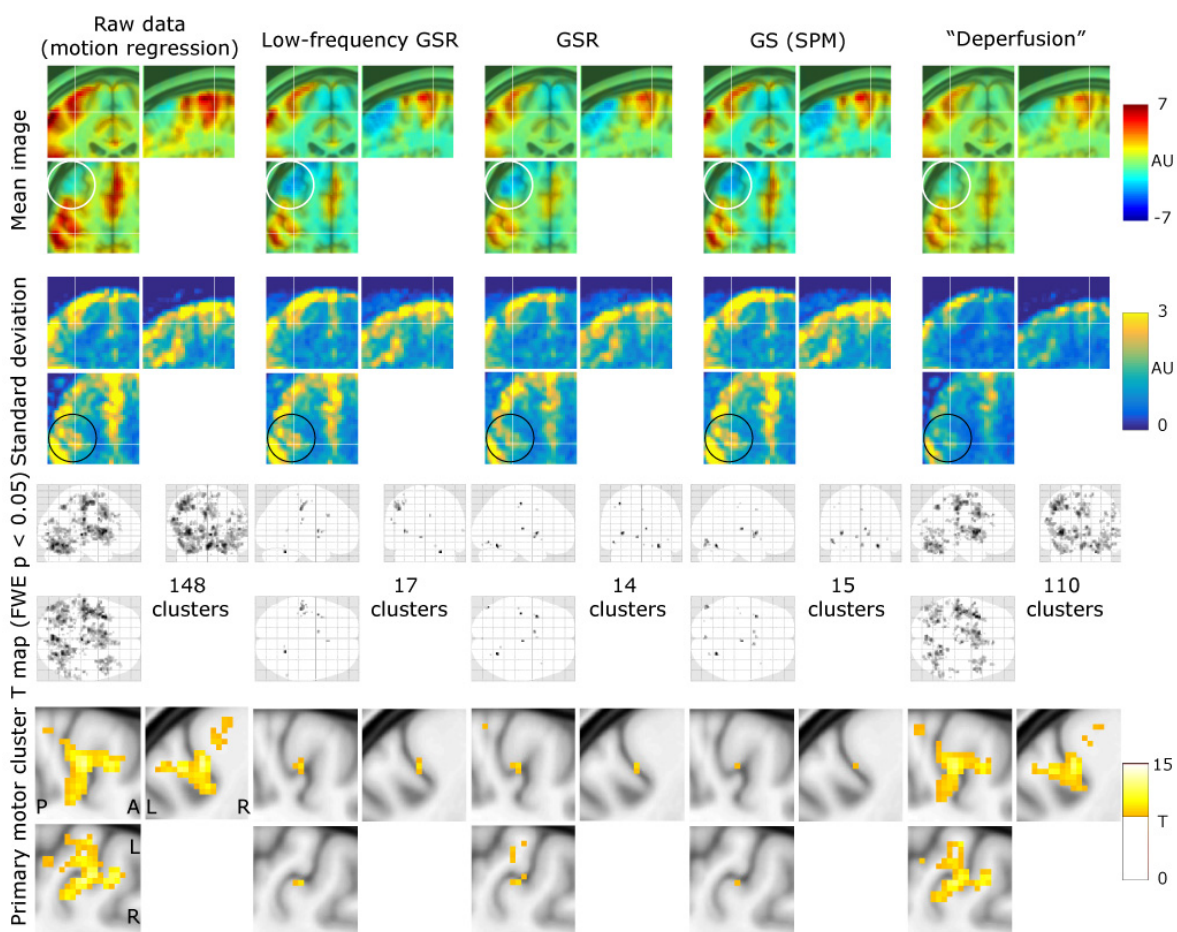
457 were detected as the inverse of the instantaneous transit time deviation from 0.5 s
458 (rBTT), since the lag mapping was performed in 0.5-s increments. The BOLD signal
459 time courses present different peak latencies for the 3 regions (broken lines are aligned
460 to the center region peak), directly reflecting the lag structure, but with similar profiles,
461 ruling out its effects on the rBTT measurement. Shaded regions represent the 95%
462 confidence intervals (N = 20).

463 BOLD, blood oxygen level-dependent

464

465 **3.2 Effect of deperfusioning on the detection of neurovascular coupling**

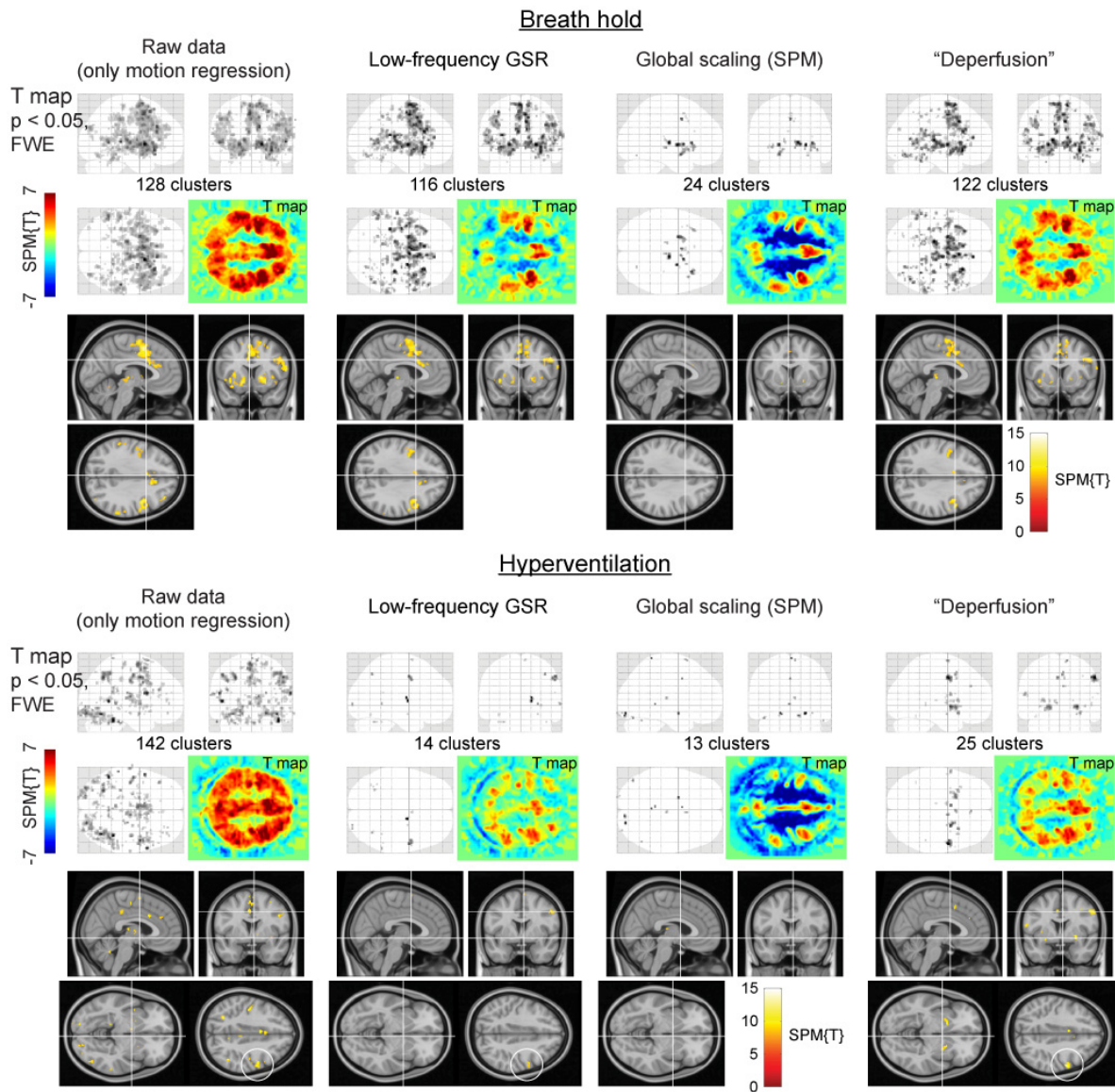
466 Removal of the whole lag structure exhibited unique effects on the fMRI analysis. While
467 the mean and SD images from the individual activation maps showed similar spatial
468 distributions, all procedures reduced the sensitivity compared to the raw dataset where only
469 the motion-related variances were removed (**Fig. 5**). However, after the deperfusioning
470 procedure, primary motor cortex activation was successfully detected by decreased
471 interindividual variances (black circles). The effect of these procedures was not uniform
472 between the respiratory challenges (**Fig. 6**), but some interpretable clusters were selectively
473 captured by the "deperfused" signals despite the reduced cluster number. For example,
474 the laryngeal motor cortices were detected at the onset and offset of the 10-s breath-holding
475 sessions (Kumar *et al.*, 2016). In the hyperventilation condition, bilateral recruitment of the
476 putamen was noted. Additionally, a premotor peak was found at coordinates [+56, 0, 40].



477

478 **Fig. 5.** The effects of the denoising procedures on the detection of neurovascular
 479 coupling during the visuomotor task. Mean and standard deviation maps from 20
 480 individual activation maps (contrast images) illustrate the spurious negative responses
 481 by the global-signal based methods (white circle) and the reduced between-participant
 482 variation by the lag structure removal or deperfusioning (black circle). The threshold
 483 for the activation maps was $p = 0.05$, corrected for FWE of multiple comparisons over
 484 the entire brain, and zoomed in to show the "hand-knob" of the left primary
 485 sensorimotor cortex.

486 FWE, family-wise error corrected; GSR, global signal regression; GS, global scaling;
 487 SPM, SPM12 software



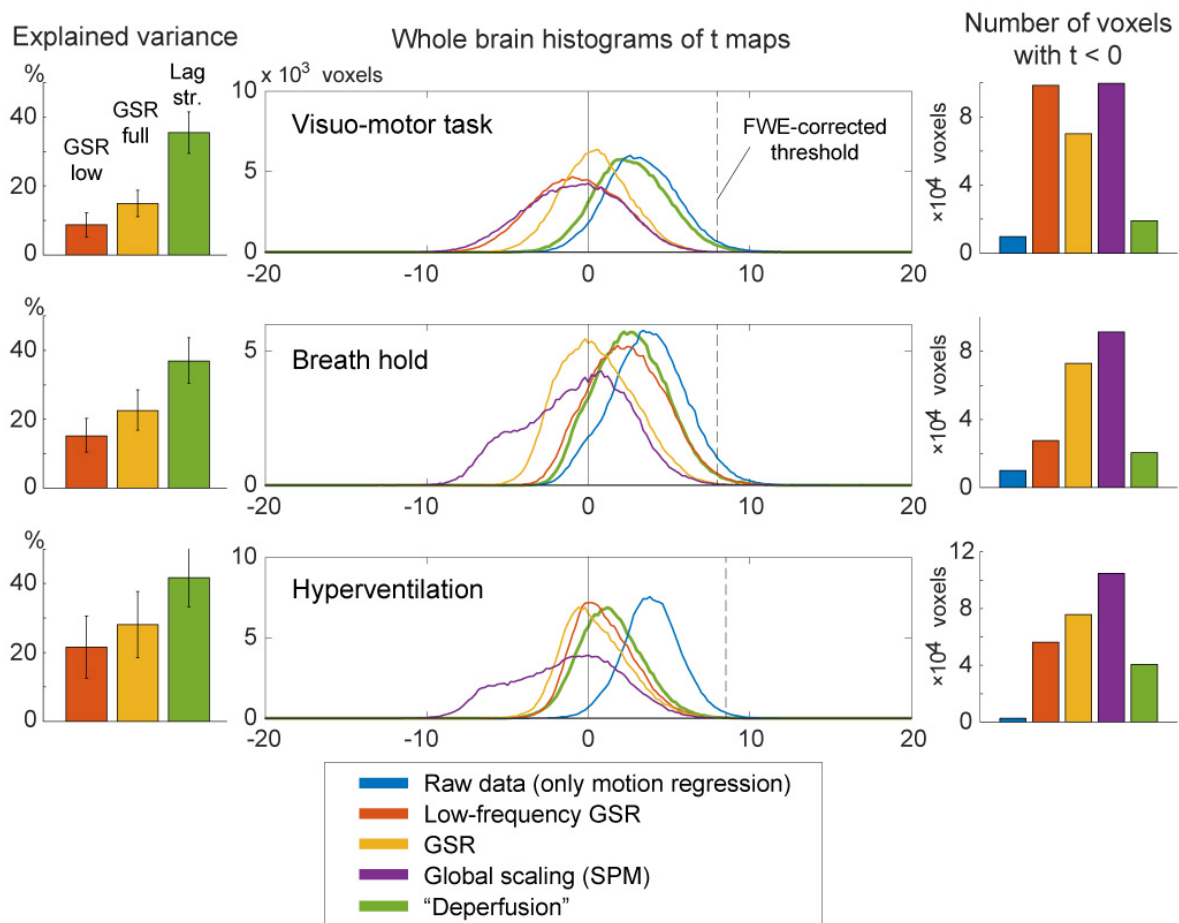
488

489 **Fig. 6.** SPM results of the respiratory challenges. Color panels show un-thresholded t
 490 map slices at the height of the laryngeal motor cortex ($z = 35$), revealing spurious
 491 negative responses (cool colors) after global signal removal but not after the
 492 deperfusioning procedure. In the hyperventilation condition, the “deperfusioned” data
 493 revealed clusters in the bilateral putamen and premotor cortex (white circle),
 494 consistently with the findings of earlier reports.

495 FWE, family-wise error corrected; GSR, global signal regression; GS, global scaling;
 496 SPM, SPM12 software

497

498 Voxel histograms from the group SPM analyses showed clear leftward shifts by the
499 global signal removal, indicating spurious deactivations (**Fig. 7**). The correlation of the
500 neuronal response with the global signal (or the extracted sLFO) was near 0, as described
501 above, but it varied across participants and tended to be positive. This trace of NVC may
502 have created the spurious deactivation after regression. Notably, this effect was very weak
503 after deperfusioning across all 3 conditions (green plots), despite a large amount of variance
504 removed by the procedure.



505

506 **Fig. 7.** The effects of preprocessing on group SPM analyses. Removal of the lag
507 structure or deperfusioning resulted in the greatest reduction of the signal variance
508 under all 3 tasks (left panels). Nevertheless, the spurious negative task responses were
509 attenuated in comparison to the removal of global fluctuation (middle and right panels).

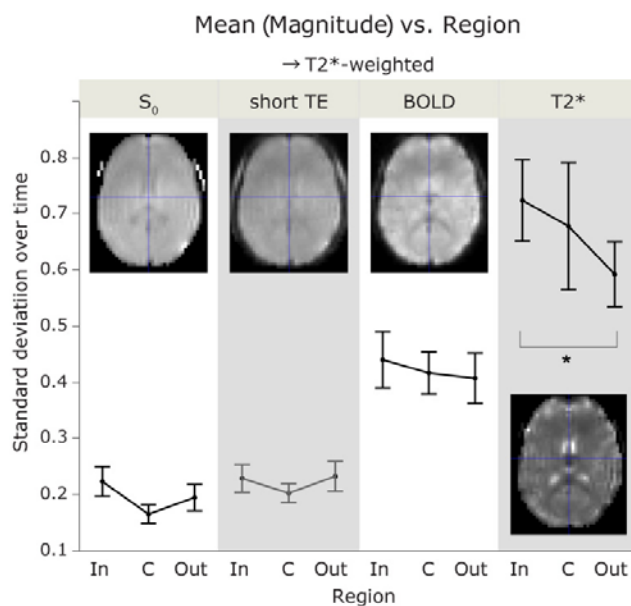
510 Error bars indicate the standard deviation. The vertical broken line in the histogram
511 indicates the statistical height threshold of $p = 0.05$, corrected for multiple comparisons
512 by the family-wise error (FWE) rate.

513 GSR, global signal regression; SPM, SPM12 software

514

515 3.3 Magnetic resonance signal components of the lag structure

516 As depicted in **Fig. 8**, the percent signal change of the sLFO, or the lag structure amplitude,
517 revealed a clear T2*-dependence with a significant reduction of amplitude in the outlet (i.e.,
518 the venous side of the gross vasculature [post hoc Tukey's HSD between T2* signals from
519 the inlet and outlet regions, following a repeated-measures ANOVA]).



520

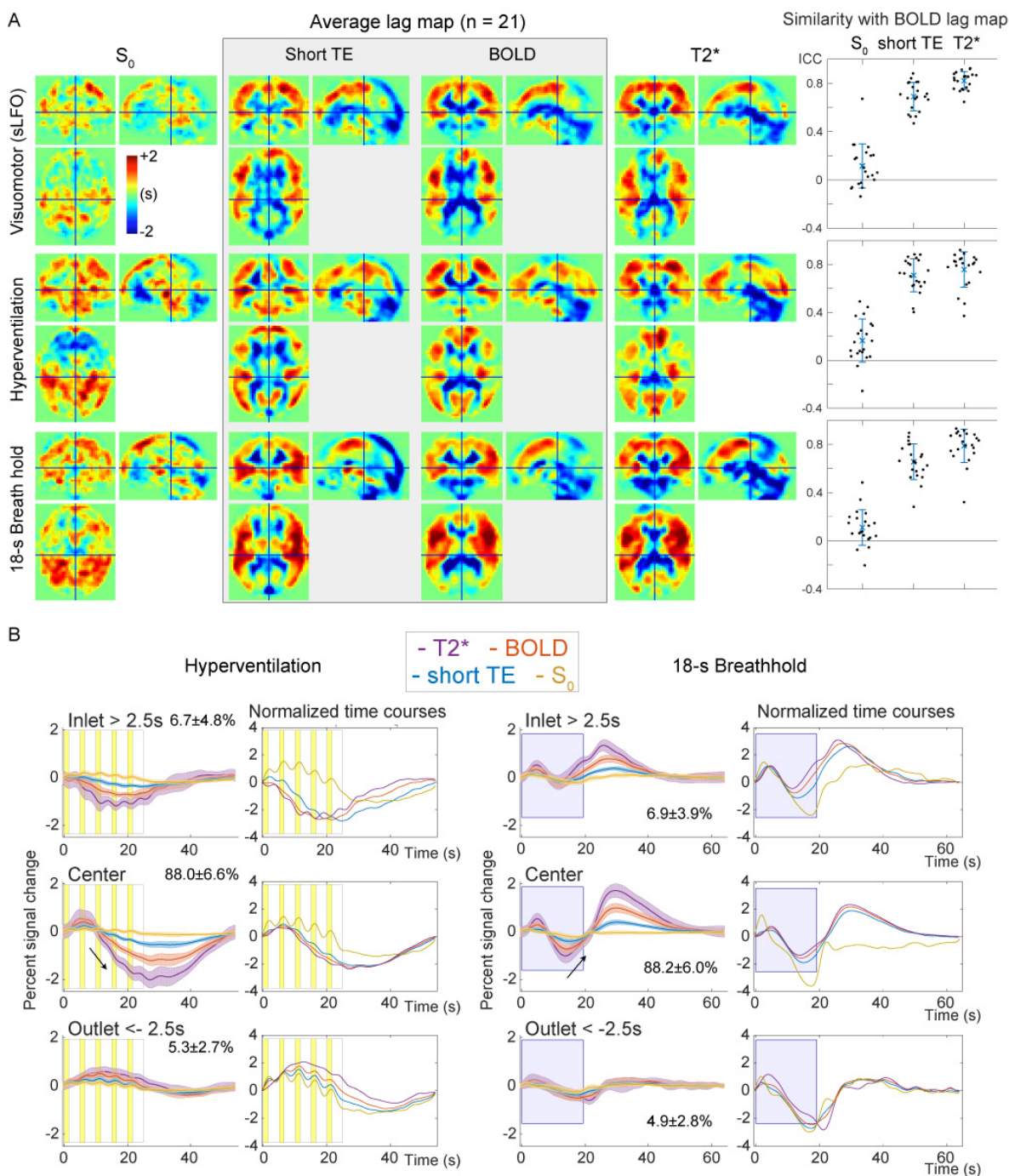
521 **Fig. 8.** The standard deviation of the percent signal change as a measure of sLFO
522 magnitude. No main effect of vascular region was observed by repeated-measures
523 ANOVA, but the T2* magnitude was significantly different between the inlet and
524 outlet sides ($p < 0.05$, Tukey HSD). The short-TE image was not included in the
525 ANOVA to avoid data redundancy. Each error bar is constructed using a 95%
526 confidence interval of the mean.

527 sLFO, spontaneous low-frequency oscillation; ANOVA, analysis of variance; TE, echo
528 time; In, inlet; C, center; Out, outlet

529

530 In lag maps from the 3 T2*-weighted images, there were some effects of the
531 respiratory challenges, but the gross cerebral vascular structure was preserved across tasks;
532 the periventricular regions and major venous sinuses were uniformly found downstream
533 (i.e., with negative arrival time) of the global signal phase, while the cortical territory of the
534 middle cerebral arteries exhibited earlier arrival (**Fig. 9A**). Only the S₀ image presented a
535 different lag structure, according to the image similarity (**Fig. 9A**, right panels).

536 Temporal analysis of the signal components revealed significant main effects of both
537 region [F (2,532) = 16.877, p < 10⁻⁶] and T2* weighting [F (2,532) = 280.786, p < 10⁻⁶]
538 (**Supp. Fig. 1A**). The S₀ time series failed to show a correlation with the T2*-weighted
539 signals, but the z-value in the inlet (i.e., the arterial side) differed significantly from that in
540 other regions (p < 0.05, Tukey's HSD). A region effect was also found for phase
541 relationships (**Supp. Fig. 1B**). Similarly, the main effects for both region [F (2,532) =
542 27.548, p < 10⁻⁶] and T2* weighting [F (2,532) = 44.679, p < 10⁻⁶], as well as their
543 interaction [F (4,532) = 24.902, p < 10⁻⁶], were significant. The phase of the TE1 signal,
544 which is less T2*-weighted than that of the BOLD signal, gradually advanced, finally
545 showing a phase lead in the outlet region, further supporting an interaction between the
546 signal components and vascular regions. These signal phase dissociations within regions
547 are displayed in **Supp. Fig. 1C**. Significant differences among the 3 T2*-weighted signals
548 were also found after the post-hoc test (p < 0.05). The signal-region interaction was evident
549 in the signal response to the respiratory challenges shown in **Fig. 9B**. Note that the traces
550 contain higher frequency components that were eliminated prior to lag mapping. In contrast
551 to the changes in T2* responses for both phase and magnitude, the S₀ component was
552 stable across vascular regions.



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Fig. 9. Analysis of the signal components and the lag structure. (A) S_0 and $T2^*$ signals were interpolated based on multi-echo acquisitions at short and typical TEs for BOLD fMRI. The lag map created from the S_0 image shows a unique structure but fails to reflect the arterial and venous structures that are consistently found in the BOLD lag map. Using the $T2^*$ -weighted signals, the lag map changes upon respiratory challenges,

559 which should primarily reflect the modification of the perfusion pattern; however, an
560 interaction with the signal component is not excluded. The right panels show intraclass
561 correlation coefficients as a quantitative measure of within-participant image similarity
562 with the BOLD lag map, with error bars indicating 95% confidence intervals. (B)
563 Temporal profiles from the 3 vascular regions indicate an absence of region effects in
564 the S_0 signal, suggesting a globally uniform mechanism underlying the S_0 response.
565 BOLD, blood oxygen level-dependent; ICC, interclass coefficient; TE, echo time;
566 fMRI, functional magnetic resonance imaging; sLFO, spontaneous low-frequency
567 oscillation

568

569 To further investigate the signal origin, we extracted the response in the
570 motor/premotor area activation peak, where NVC was expected to dominate (**Supp. Fig.**
571 **2A**). For this analysis, the group activation map from Experiment 1 was used to define the
572 regions-of-interest in order to avoid bias. We found high-frequency components
573 dominating the S_0 responses in comparison to the responses from larger regions shown in
574 **Figure 9B**. During hyperventilation, respiratory phase-related fluctuations were observed
575 with a signal decrease initiated by inhalation and followed by a positive deflection during
576 exhalation. The $T2^*$ signal also exhibited small fluctuations but with a different phase,
577 possibly dominated by the NVC component.

578 We conducted an additional analysis for the fast respiration-related non-BOLD
579 components. The spatial distribution of this response is shown in **Supp. Fig. 2B**.
580 Interestingly, there was a clear anterior-posterior segmentation of response polarity, with
581 the posterior regions presenting the opposite phase of the fast S_0 deflection by respiratory
582 maneuvers. Some additional symmetrical structures were found in deep-brain regions, near
583 the deep middle cerebral and inferior ventricular veins, implying a unique vascular
584 involvement. Importantly, this spatial pattern was not that of typical motion artifacts that
585 can accompany volitional respiration in spite of the careful instruction.

586 Finally, **Supp. Fig. 2C** shows the data from a subset of participants, obtained using a
587 different TR/flip-angle setting to manipulate the inflow effect. The $T2^*$ response was

588 smaller than that shown in **Fig. 2A**, presumably due to the short TR. The different TR also
589 contributed to the rich high-frequency components by the fast sampling rate. The slow S_0
590 change was also diminished, but the respiration-related fast component was relatively
591 preserved suggesting the absence of a strong inflow effect.

592 **4 Discussion**

593 The principal findings of this study are summarized as follows. First, based on the
594 instantaneous phase difference within the BOLD lag structure, we observed a small blood
595 flow velocity change selectively in the inlet region of the vasculature. Next, the complete
596 elimination of the lag structure reduced interindividual variance and spurious deactivation,
597 supporting our hypothesis that NVC could be observed more specifically by this
598 deperfusioning procedure. This finding is in agreement with the results of earlier work on
599 resting-state fMRI (Erdoğan *et al.*, 2016). Finally, the lag structures in the S_0 (or non-
600 BOLD) component did not correlate with that from $T2^*$, either spatially or temporally. We
601 also found a vascular region-dependent change in the $T2^*$ sLFO, with a decreased
602 amplitude in the outlet part close to major veins, in contrast to the S_0 response that
603 remained constant; this finding replicates a previous observation in the raw BOLD signal
604 (Aso *et al.*, 2017a). The S_0 component exhibited a unique brain region-dependent response
605 to the respiratory phase, suggesting that certain perfusion parameters specifically contribute
606 to this component, but not the perfusion lag. Overall, the BOLD low-frequency phase
607 behaved as a deoxy-Hb-based virtual contrast agent in the present data, leaving a global
608 noise component for the fMRI analysis.

609 The observation that the velocity on the arterial side exhibits changes alongside
610 respiratory variations is consistent with the findings of previous reports using transcranial
611 Doppler ultrasonography (Malatino *et al.*, 1992; Poulin *et al.*, 1996). This information was
612 extracted from the BOLD lag structure, which itself presented autoregulatory response
613 consistent with earlier work (Murphy *et al.*, 2011). During the initial whole-brain CBV
614 increase in response to autoregulatory vasodilation, a sole increase in the inflow should first
615 occur to meet the volume demand. It is therefore reasonable that this effect is absent in the

616 outlet (i.e., the venous side of the gross vasculature). This observation seems to support the
617 model in which the BOLD lag structure is derived from an axial non-uniformity in the
618 vessels, already present in the inflow (Tong *et al.*, 2018). A distinct mechanism of the lag
619 structure was also suggested by the diminished magnitude observed in the outlet side of the
620 vasculature, since a CBF increase should evoke larger response in the downstream (Krings
621 *et al.*, 1999).

622 It is unclear what proportion of this axial variation is systemic, i.e., originates from
623 the autonomic loops, mediated by peripheral baro- or chemoreceptors. However, even when
624 the neural activity is contributing to the sLFO time course as demonstrated previously (Aso
625 *et al.*, 2017), the resulting lag structure largely reflects the vasculature. In this work, we
626 focused on non-neuronal mechanisms to account for the BOLD lag structure as much as
627 possible, in the hope that it may ultimately help achieve a better understanding and provide
628 improved modeling approaches of the fMRI signal.

629 **4.1 Source of BOLD low-frequency oscillation signals**

630 Previous studies on sLFOs have reported that both Hb species fluctuate, but with varying
631 phase differences that are selectively found in the brain (Obrig *et al.*, 2000; Rayshubskiy *et al.*
632 *et al.*, 2014; Tgavalekos *et al.*, 2016). The observed fluctuations of total Hb density have been
633 linked to CBV changes (Boas & Dale, 2005; Kennerley *et al.*, 2005; Kim & Ogawa, 2012),
634 but interpretations for that of deoxy-Hb have rarely been provided. Only 1 series of studies
635 by Fantini and colleagues directly addressed the possible axial variation of blood content
636 such as oxygen saturation (Fantini, 2014). In support of the conventional theory,
637 Rayshubskiy and colleagues reported, in their human intraoperative study, that slow Hb
638 oscillations correlated with vasomotion in the superficial arteries (Rayshubskiy *et al.*, 2014).
639 However, it remains unclear whether an equivalent vasomotion exists in the non-arterial
640 vessels to fully account for the observed lag structure. Hence, it is worth considering other
641 sources of deoxy-Hb variation.

642 The concept of vasomotion stems from an active diameter change in the precapillary
643 vessels, driving local velocity fluctuations, termed “flowmotion” (Intaglietta, 1990). This

644 flowmotion can reportedly accompany the fluctuation of local Hct that should affect deoxy-
645 Hb density (Fagrell *et al.*, 1980; Hudetz *et al.*, 1999). Another possible source for the
646 deoxy-Hb fluctuations is a change in SaO₂ that ranges from 94–98% in the artery
647 (Intaglietta *et al.*, 1996; Collins *et al.*, 2015). For example, the respiration-related BOLD
648 signal component is supposed to be mediated by the blood CO₂ level and pH, which can
649 shift the oxygen dissociation curve (Birn *et al.*, 2006; Chang & Glover, 2009a). These
650 parameters are considered to fluctuate in the blood as part of the autonomic loop, possibly
651 driving local vasomotion, which persists after denervation (Morita *et al.*, 1995). As
652 mentioned above, the phase difference between the 2 Hb species remains to be elucidated
653 (Sassaroli *et al.*, 2012), but those observations do substantiate an unstable deoxy-Hb supply
654 in brain tissues. Besides, such a signal component would have escaped detection in NVC
655 studies using trial averaging.

656 A signal origin intrinsic to the flowing blood may, as suggested by Tong and
657 colleagues, thus explain the constant phase difference among signals from different body
658 parts (Tong *et al.*, 2012, 2017). In the literature, an axial variation of the Hct in the brain
659 has indeed been suggested, in relation to both NVC (Kleinschmidt *et al.*, 1996; Siegel *et al.*,
660 2003; Chen & Pike, 2009) and sLFOs (Fagrell *et al.*, 1980; Mayhew *et al.*, 1996).
661 Furthermore, the reduction in T2* LFO amplitude in the outlet side can be explained by the
662 high tissue deoxy-Hb density, which likely diminishes the proportional effect of intrinsic
663 deoxy-Hb fluctuations, unless the OEF is completely coupled to this variation. Importantly,
664 temporal dispersion alone would not fully account for the amplitude reduction, as it was
665 only found in the venous side, despite the fact that the lag structure was tracked both up-
666 and downstream from the global phase. These results provide good contrast with the stable
667 S₀ response, reflecting its insensitivity to oxygen saturation. Although it would be too
668 challenging to incorporate complex rheological parameters, a reconsideration of the
669 constant deoxy-Hb assumption may help improve BOLD signal modelling.

670 In the hyperventilation condition, we observed a fast response to each ventilation
671 cycle, accompanying blood pressure changes. This is consistent with reports using
672 optimized acquisition techniques (Dresel *et al.*, 2005; Pattinson *et al.*, 2009), supported by

673 anatomical (Simonyan & Jürgens, 2003), as well as electrophysiological (Radna &
674 MacLean, 1981) studies. The premotor peak at coordinates [+56, 0, 40] was also very close
675 to the reported activation site for volitional respiration (McKay *et al.*, 2008). Although the
676 effect of respiratory movement cannot be fully excluded, the spatial pattern in **Supp. Fig.**
677 **2B** is not that of a typical motion artifact centered on the brain surface (Krings *et al.*, 2001).
678 In healthy participants, inhalation increases systemic venous return through decreased
679 intrathoracic pressure, causing an elevation of cardiac output with some delay. In contrast,
680 exhalation is considered to cause CBV increases through elevated cerebral venous pressure.
681 To our knowledge, the timing order of these events has not been studied at the precision of
682 the current data; further studies are needed to determine the source of this S_0 fluctuation
683 (Yen *et al.*, 2017). The only available clue in our results is the spatial distribution, such as
684 the interesting anterior-posterior segmentation (resembling the unique “ S_0 lag structure” in
685 **Fig. 9a**) or the symmetrical pattern in the deep brain structures. Nonetheless, some
686 mechanical effects of the respiratory act on the fluid dynamics likely exist, causing this
687 spatially heterogeneous S_0 deflection.

688 **4.2 Lag structure as noise**

689 Based on the assumption that the global signal fluctuation is the sum of all variations by
690 NVC, its elimination by GSR has been considered to negatively bias the results (Caballero-
691 Gaudes & Reynolds, 2017). However, as noted by Aguirre and others, there are cases
692 where GSR yields interpretable results even in the absence of global motion artifacts
693 (Aguirre *et al.*, 1998). The rise in popularity of rs-fMRI since 2005 has led to this issue
694 resurfacing in a different form. A variation in GSR, in which the time series extracted from
695 a set of regions-of-interest (whole brain, white matter, and cerebrospinal fluid) are removed,
696 has become a *de facto* standard. It is indeed computationally closer to our deperfusioning in
697 that the regional phase difference is somehow tolerated. However, because this practice
698 lacks a strong theoretical background (Chang *et al.*, 2009; Liu *et al.*, 2017), currently, the
699 identification and elimination of bodily movements and physiological noise are more
700 widely recommended. There are various approaches to this end, such as simultaneous
701 physiological measurements (Chang & Glover, 2009b), as well as data-driven methods that

702 only use fMRI data (Smith *et al.*, 2013). To date, however, objective criteria for
703 distinguishing neural activity from noise components remains an issue (Salimi-Khorshidi *et*
704 *al.*, 2014).

705 In the present study, the lag structure was treated as a broadly distributed, structured
706 noise for fMRI. Indeed, it can be partly eliminated by sophisticated denoising techniques
707 (Aso *et al.*, 2017a). However, the specificity of lag mapping in isolating information on a
708 purely vascular origin remains unclear. For example, measurements of velocity changes
709 critically depend on a recursive lag-tracking method that incorporates the gradual change in
710 LFO over regions (Tong & Frederick, 2014). Adaptation for changes in the waveform that
711 may arise from different paths of the blood was demonstrated to increase the
712 reproducibility of the lag map (Aso *et al.*, 2017a). However, as the changes in waveform
713 can also reflect NVC, removing the whole lag structure may lead to type II errors in the
714 fMRI results. Hence, the favorable impacts of the deperfusioning procedure that we
715 observed on the fMRI results are clearly insufficient to prove the advantage of this
716 technique and require further confirmation.

717 Importantly, the detection of the lag structure itself largely depends on the data
718 quality, especially in terms of head movement. When a head movement results in a
719 synchronized deflection that exceeds the LFO amplitude, it would obscure the phase
720 variation. However, it can be also questioned if the correlational structure of neural activity
721 is reliably detected from such motion-contaminated data. In general, NVC should have a
722 limited spatial extent and signal magnitude without time-locked averaging (Power *et al.*,
723 2012). In turn, successful tracking of a lag structure may even be considered as evidence of
724 “clean” data. The elimination of this identified lag structure can be a relatively
725 straightforward approach to reduce structured physiological noise (Caballero-Gaudes &
726 Reynolds, 2017).

727 In conclusion, by investigating various aspects of the BOLD sLFO, we compiled
728 supporting evidence for a component intrinsic to flowing blood that has been a focus of
729 interest in earlier works (Tong *et al.*, 2017). To establish a framework by which the fMRI

730 signal can be fully modeled, more detailed characterization of the lag structure as part of
731 the “global noise” is needed (Glasser *et al.*, 2018).

732 **5 Conflict of Interest**

733 The authors declare that the research was conducted in the absence of any commercial or
734 financial relationships that could be construed as a potential conflict of interest.

735 **6 Author Contributions**

736 All authors provided approval of the final version of the manuscript to be published. TA
737 contributed to concept and design of this research, data analysis and interpretation, and
738 drafting of the manuscript. SU contributed to data acquisition and drafting of the
739 manuscript. SU, HF, and TM contributed to data interpretation and review of the final draft.

740 **7 Funding**

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742 (Non-linear Neuro-oscillology: Towards Integrative Understanding of Human Nature,
743 JP15H05875) to TA and JP16H06395 and JP16H06397 to TM from the Japan Society for
744 the Promotion of Science.

745 **8 Acknowledgments**

746 We would like to thank Editage (www.editage.jp) for English language editing.

747 **9 Data Availability Statement**

748 The datasets analyzed for this study can be provided upon request.

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1046 **11 Additional Information**

1047 **11.1 Competing Interests**

1048 None of the authors has any conflicts of interests.

1049