

1 **Micro- and nanostructural characteristics of rat masseter muscle entheses**

2 **-Characteristics of masseter muscle entheses-**

3

4 Keitaro Arakawa¹⁾²⁾, Satoru Matsunaga¹⁾³⁾, Kunihiko Nojima¹⁾²⁾, Takayoshi Nakano⁴⁾,

5 Shinichi Abe³⁾, Masao Yoshinari¹⁾, Kenji Sueishi¹⁾²⁾

6

7 1) Oral Health Science Center, Tokyo Dental College

8 2) Department of Orthodontics, Tokyo Dental College

9 3) Department of Anatomy, Tokyo Dental College

10 4) Division of Materials & Manufacturing Science, Graduate School of Engineering,

11 Osaka University

12

13 Corresponding author: Satoru Matsunaga

14 Department of Anatomy, Tokyo Dental College, 2-9-18 Kandamisaki-cho, Chiyoda-ku,

15 Tokyo 101-0061, Japan

16 Tel.: +81-3-6380-9592; Fax: +81-3-6380-9664

17 E-mail: matsuna@tdc.ac.jp

18

19 Abstract

20 The entheses of the masticatory muscles differ slightly from those of the trunk and limb
21 muscles. However, the bones of the skull are subject to various functional pressures,
22 including masticatory force, resulting in a complex relationship between bone structure
23 and muscle function that remains to be fully elucidated. The present study aimed to
24 clarify aspects of masseter muscle-tendon-bone morphological characteristics and local
25 load environment through quantitative analysis of biological apatite (BAP) crystallite
26 alignment and collagen fiber orientation together with histological examination of the
27 entheses.

28 Result of histological observation, the present findings show that, in the entheses of the
29 masseter muscle in the first molar region, tendon attaches to bone via unmineralized
30 fibrocartilage, while some tendon collagen fibers insert directly into the bone, running
31 parallel to the muscle fibers. Furthermore, BAP crystallites in the same region show
32 uniaxial preferential alignment at an angle that matches the insertion angle of the tendon
33 fibers. Conversely, in the entheses of the masseter muscle in the third molar region, the
34 tendon attaches to the bone via a layer of thickened periosteum and chondrocytes. As in
35 the first molar region, the results of bone quality analysis in the third molar region
36 showed BAP crystallite alignment parallel to the orientation of the tendon fibers. This

37 indicates that the local mechanical environment generates differences in entheses
38 morphology.

39 The present study showed a greater degree of uniaxial BAp crystallite alignment in
40 entheses with direct insertion rather than indirect tendon-bone attachment and the
41 direction of alignment was parallel to the orientation of tendon fibers. These findings
42 suggest that functional pressure from the masseter muscle greatly affects bone quality as
43 well as the morphological characteristics of the entheses, specifically causing micro- and
44 nanostructural anisotropy in the direction of resistance to the applied pressure.

45 (292 words)

46

47 Keywords: collagen fiber, biological apatite (BAp) crystallite, bone quality, microbeam
48 X-ray diffraction, SHG imaging, entheses

49

50

51

52

53

54

55 1. Introduction

56 The tendons of the trunk and limb muscles attach to bones via entheses [1-3].
57 These attachment sites are broadly categorized as either fibrous entheses, composed of
58 perforating mineralized collagen fibers, or fibrocartilaginous entheses, comprising a
59 multitissue interface involving the following four tissues of tendon, unmineralized
60 fibrocartilage, mineralized fibrocartilage, and bone [4]. The composition, structure, and
61 mechanical properties of these multitissue interfaces vary widely, creating spatial
62 gradients that mediate load transfer between soft and hard tissues and minimize stress
63 concentration [5]. Muscle loading is extremely important for healthy enthesis formation
64 and suppression of muscle function greatly diminishes the biomechanical performance
65 of the enthesis [6].

66 Histological examination by Hems et al. revealed that the entheses of the
67 masticatory muscles differ slightly from those of the trunk and limb muscles [7].
68 Specifically, the masticatory muscles contain three types of enthesis, including sites of
69 direct tendon insertion into the bone. The authors concluded that these different types
70 contribute to the unique biomechanical function of the masticatory muscles, enabling
71 them to work as an “angle and stretching brake”. However, the bones of the skull are
72 subject to various functional pressures, including masticatory force, resulting in a

73 complex relationship between bone structure and muscle function that remains to be
74 fully elucidated. Clarification of the relationships between the micro- and nanostructural
75 characteristics of the muscles, tendons, and bones in the maxillofacial area and the
76 mechanical environment is required [8, 9].

77 The relevance of bone quality in addition to bone density with regard to bone
78 strength was proposed National Institutes of Health Consensus Development
79 Conference in 2000. Since then, studies on the relationship between bone structural
80 characteristics and bone strength have focused on bone quality factors [10]. Collagen
81 fibers and biological apatite (BAp) crystallites have been identified as dominant bone
82 quality factors that are resistant to tensile and compressive stress, respectively, on bone
83 tissue [11, 12].

84 Biological apatite is a hexagonal, ionic crystal that has a highly anisotropic
85 nanostructure with preferential alignment along the c axis in the loading direction [13].
86 Using microbeam X-ray diffraction analysis, Nakano et al. quantitatively analyzed BAp
87 crystallite alignment in animal trunk and limb bones, demonstrating a high correlation
88 between mechanical stress and BAp crystallite alignment [14, 15]. With regard to
89 collagen fibers as a bone quality factor, ongoing research by Vashishth et al. has shown
90 that collagen crosslinks are a factor in age-related reductions in bone quality [16].

91 Meanwhile, Kawagoe et al. reported a relationship between bone strength and
92 orientational anisotropy of collagen fibers for the masticatory muscles [17]. Quantitative
93 analysis of the jaw bone, particularly at entheses, should enable accurate prediction of
94 the effects of the masseter muscles on the load environment of the mandible.

95 The present study aimed to clarify aspects of masseter muscle-tendon-bone
96 morphological characteristics and local load environment through quantitative analysis
97 of BAp crystallite alignment and collagen fiber orientation together with histological
98 examination of the entheses.

99

100 2. Materials and Methods

101 2.1 Samples

102 The present study was approved by the Ethics Committee of Tokyo Dental
103 College (Ethics Application No. 282807). Samples were prepared from the skulls of
104 five 24-week-old male Wistar rats euthanized after deep anesthesia with ethyl ether.

105

106 2.2 Tissue slice preparation

107 To obtain suitable samples for bone quality analysis, the left skull was
108 embedded in autopolymerizing acrylic resin and sagittally sectioned using a saw

109 microtome (SP1600; Leica, Wetzlar, Germany) with a blade width of 300 μm . Samples
110 were then sanded using wet/dry sandpaper of increasing grit (400, 800, and 1200) to
111 prepare thin, 200 μm slices. The right skull was fixed in 4% paraformaldehyde
112 phosphate buffer solution and demineralized in 10% ethylenediaminetetraacetic acid
113 (EDTA) for 4 weeks. Using standard methods, samples were embedded in paraffin
114 embedding and sliced about 5 μm thick in the coronal plane to enable observation of the
115 masseter muscle entheses. Masson's trichrome staining were performed to observe the
116 structural morphology of masseter muscle entheses in the first and third molar regions.
117 And Toluidine blue staining were used to make the acidic mucus polysaccharide present
118 in the cartilage metachromatic.

119

120 2.3 Second harmonic generation (SHG) imaging

121 SHG images were acquired using a multiphoton confocal microscopy system
122 (A1R+MP, Nikon, Japan) with an excitation laser (Mai Tai eHP, wavelengths: 690-
123 1040 nm; repetition rate: 80 MHz; pulse width: 70 fs; Spectra-Physics, CA, US) and a
124 water-immersion objective lens (CFI75 Apo 25 \times W MP, numerical aperture: 1.1; Nikon,
125 Tokyo, Japan). The excitation wavelength for the observation of collagen fibers was 880
126 nm. Image acquisition, processing for orthogonal views and cropping were performed

127 using NIS-Elements version 4.0 (Nikon). Brightness and contrast of some images were
128 adjusted using look-up tables (LUTs) of this software by the same parameters among
129 relevant images to facilitate visibility.

130

131 2.4 Micro-computed tomography (micro-CT) imaging

132 Samples were examined using micro-computed tomography (micro-CT;
133 HMX225 Actis4; Tesco, Tokyo, Japan) under the following imaging conditions: tube
134 voltage, 140 kV; tube current, 100 μ A; matrix size, 512 \times 512; magnification, \times 2.5; slice
135 width, 50 μ m; and slice pitch, 50 μ m. Three-dimensional reconstruction was performed
136 using TRI/3D-BON software (RATOC System Engineering, Tokyo, Japan).

137

138 2.5 BAp crystallite alignment

139 Quantitative analysis of BAp crystallite alignment was conducted using an
140 optical curved imaging plate (IP) X-ray diffraction system (XRD; D/MAX PAPID II -
141 CMF; Rigaku, Tokyo, Japan). Measurements were performed in reflection and
142 transmission modes with Cu-K α as the radiation source at a tube voltage of 40 kV and
143 tube current of 30 mA. Reference axes were established in X axis, Y axis, and Z axis for
144 each sample (Fig. 1) [18, 19]. Regions of interest in the mandible comprised masseter

145 muscle entheses (Fig. 2). The radiation site was determined using the light microscope
146 of the XRD system (magnification, $\times 0.6-4.8$), then an incident beam (diameter, $50\ \mu\text{m}$)
147 was applied. Using reflection mode in the X-axis direction and transmission mode in the
148 Y-axis and Z-axis directions, the diffracted X-ray beam was detected using a curved IP
149 based on the conditions described by Nakano et al. [20]. The diffracted X-ray beam was
150 detected as a diffraction ring on the IP. Using 2-dimensional data-processing software
151 (Rigaku), X-ray diffraction intensity ratios were calculated for the two diffraction peaks
152 corresponding to planes 002 and 310.

153

154 **Fig 1. Reference points, plane and axes: point a the lowest point in anterior**
155 **thickening area of mandible.** Point p the lowest point in posterior thickening area of
156 mandible; Mandibular plane passing through a-a' and p-p' lines; X-axis passing through
157 the mid-point of a-a' and p-p'; Y-axis the vertical axis against the mandibular plane; Z-
158 axis the vertical axis against the X-Y plane.

159 **Fig 2. Measurement points.** (A) First molar region. (B) Third molar region.

160

161 2.6 Statistical analysis

162 Mean values for the five samples for each measurement point were calculated

163 and compared using Tukey's multiple comparison test. Significance was set at $P < 0.05$.

164

165 3. Results

166 3.1 Histological observation of entheses

167 The results of Masson's trichrome staining and toluidine blue staining are
168 showed of a coronal section in the first molar region of 24-week-old Wistar rat skulls
169 (Fig. 3). Thick masseter muscle tendons with fibers largely grouped into bundles could be
170 seen directly integrating into the buccal cortical bone. In the enthesis, the periosteum was
171 fragmented and aggregation of chondrocytes was observed at the tendon-bone interface
172 (Fig. 3B, C,D,E). The results of Masson's trichrome staining and toluidine blue staining
173 are showed of a coronal section in the third molar region of 24-week-old Wistar rat skulls
174 (Fig. 4).

175

176 **Fig 3. Masson's trichrome and toluidine blue staining of masseter muscle enthesis in**
177 **the first molar region.** MA: masseter muscle. T: tongue. 1st Molar: first molar region.

178 **Fig 4. Masson's trichrome and toluidine blue staining of masseter muscle enthesis in**
179 **the third molar region.** MA: masseter muscle. T: tongue. 3rd Molar: third molar region.

180

181 In the masseter muscle enthesis, muscle fibers could be seen covering the area
182 from the lateral aspect to the base of the mandible, while thin tendons ran toward the
183 cortical bone protuberance on the buccal side of the mandibular body (Fig. 4A). The
184 tendon was attached to the periosteum without rupturing (Fig. 4B,D) and thickened
185 chondrocytes were observed on the bone surface in the enthesis (Fig. 4C,E).

186

187 3.2 Orientational anisotropy of collagen fibers

188 SHG images from the first molar region are showed in Figure 5. The masseter
189 muscle tendon in the first molar region comprised thick collagen fibers extending through
190 the enthesis, some of which penetrated the cortical bone. As in the first molar region,
191 collagen fibers in the third molar region ran toward the bone. However, these fibers were
192 interrupted at the thickened periosteum. In the vicinity of the enthesis, many collagen
193 fibers inside the bone were observed running parallel to the orientation of the tendon (Fig.
194 6). In the first and third molar regions, tendon fibers ran tangentially to the bone at 31.1°
195 [standard deviation (SD), 4.6°] and 40.4° [SD, 3.5°], respectively (Fig. 7).

196

197 **Fig 5. Coronal section in first molar region.** (A) Hematoxylin and eosin staining. (B)
198 Second harmonic generation imaging. The masseter muscle tendon in the first molar

199 region comprised thick collagen fibers extending through the enthesis, some of which
200 penetrated the cortical bone.

201 **Fig 6. Coronal section in third molar region.** (A) Hematoxylin and eosin staining. (B)
202 Second harmonic generation imaging. In the vicinity of the enthesis, many collagen
203 fibers inside the bone were observed running parallel to the orientation of the tendon.

204 **Fig 7.** Orientation of tendon fibers in relation to bone. (A) First molar region. (B) Third
205 molar region.

206

207 3.3 BAp crystallite alignment

208 The angles of preferential alignment of BAp crystals are showed in an enthesis
209 in relation to tooth axis in the first and third molar regions (28° [SD, 10.95°]) and 36°
210 [SD, 8.94°], respectively). X-ray diffraction intensity ratios calculated for the three
211 reference axes for quantitative analysis are showed in Figure 8. The intensity ratios for
212 hydroxyapatite powder were 1.4 and 5.6 in reflection and transmission modes,
213 respectively. In both the first and third molar regions, strong uniaxial preferential
214 alignment was noted in the Y-axis direction in the masseter muscle entheses.
215 Furthermore, X-ray diffraction intensity ratios in the Y-axis direction were significantly
216 higher in the first compared to the third molar region.

217

218 4. Discussion

219 According to Huang et al., as the superior digital flexor tendon develops before
220 and then joins with the subsequently formed digital bone, the related entheses penetrate
221 the periosteum [21]. Conversely, the entheses of many of the muscles in the trunk and
222 limbs do not involve the periosteum, suggesting that these attachments are already
223 formed at the stage of periosteal development [22]. The present findings also show that,
224 in the entheses of the masseter muscle in the first molar region, tendon attaches to bone
225 via unmineralized fibrocartilage, while some tendon collagen fibers insert directly into
226 the bone, running parallel to the muscle fibers. Furthermore, BAp crystallites in the
227 same region show uniaxial preferential alignment at an angle that matches the insertion
228 angle of the tendon fibers. This suggests that both the anisotropy of the collagen fibers
229 and BAp crystallite alignment confer high resistance in the direction of the masseter
230 muscle tendon. As the masseter muscle in the first molar region is directly attached to
231 the bone via the tendon, these structural characteristics may optimize bone quality to
232 enable the high load generated by muscle contraction to be efficiently transmitted from
233 tendon to bone [23].

234 Conversely, in the entheses of the masseter muscle in the third molar region,

235 the tendon attaches to the bone via a layer of thickened periosteum and chondrocytes.
236 Muscles with entheses that indirectly attach to the bone via the periosteum do not
237 produce large functional pressures; rather, they are responsible for precise movements
238 [24]. As in the first molar region, the results of bone quality analysis in the third molar
239 region showed BAp crystallite alignment parallel to the orientation of the tendon fibers.
240 However, the intensity ratio values were significantly lower. This indicates that the local
241 mechanical environment generates differences in enthesis morphology.

242 Matsumoto et al. analyzed bone quality in human jaw bones and found uniaxial
243 preferential alignment of BAp crystallites and high bone strength in the tooth axis
244 direction in specific alveolar bone [25]. Meanwhile, Nakano et al. demonstrated a strong
245 positive correlation between bone quality factors and bone strength, indicating that the
246 mechanical environment of entheses determines the orientation of the collagen fibers,
247 which is linked to the preferential alignment of BAp crystallites. Changing the amount
248 and direction of functional pressure from the muscles may thus affect not only bone
249 density, but also bone density.

250

251 5. Conclusion

252 In the entheses of rat masseter muscle, some tendons attach to the bone directly

253 and others attach indirectly via the periosteum. The present study showed a greater
254 degree of uniaxial BAp crystallite alignment in entheses with direct insertion rather than
255 indirect tendon-bone attachment and the direction of alignment was parallel to the
256 orientation of tendon fibers. These findings suggest that functional pressure from the
257 masseter muscle greatly affects bone quality as well as the morphological characteristics
258 of the enthesis, specifically causing micro- and nanostructural anisotropy in the
259 direction of resistance to the applied pressure.

260

261 6. Acknowledgement

262 This research was supported by a Grant-in-Aid for Scientific Research (C)
263 (17K11808 and 25463055) from the Japan Society for the Promotion of Science. This
264 study was supported by a research grant under the Private University Research
265 Branding Project from the Ministry of Education, Culture, Sports, Science, and
266 Technology of Japan (2018).

267

268 7. Conflict of Interest

269 No potential conflicts of interest to disclose.

270

271 References

272 1. Benjamin M, Evans EJ, Copp L. The histology of tendon attachments to bone in man.

273 J Anat. 1986;149: 89-100.

274 2. Holger P, Martin S. Histological evidence for muscle insertion in extant amniote

275 femora: implications for muscle reconstruction in fossils. J Anat. 2013;222: 419-436.

276 3. Evans E J, Benjamin M, Pemberton DJ. Variations in the amount of calcified tissue at

277 the attachments of the quadriceps tendon and patellar ligament in man. J Anat.

278 1991;174: 145-151.

279 4. Apostolakos J, Durant TJ, Dwyer CR, Russell RP, Weinreb JH, Alae F, Beitzel K,

280 McCarthy MB, Cote MP, Mazzocca AD. The enthesis: a review of the tendon-to-bone

281 insertion. Muscles Ligaments Tendons J. 2014;4: 33-42.

282 5. Thomopoulos S, Williams GR, Gimbel JA, Favata M, Soslowsky LJ. Variations of

283 biomechanical, structural, and compositional properties along the tendon to bone

284 insertion site. J Orthop Res.2003; 1: 13-19.

285 6. Schwartz AG, Lipner JH, Pasteris JD, Genin GM, Thomopoulos S. Muscle loading is

286 necessary for the formation of a functional tendon enthesis. Bone.2013;55: 44-51.

287 7. Hems T, Tillmann B. Tendon entheses of the human masticatory muscles. Anat

288 Embryol (Berl). 2000;202: 201-208.

- 289 8. Yamada K. Histological study on the migration mechanism of the attachment of the
290 deep layer of the masseter muscle to the rat mandible during growth. *Jpn J Oral Biol.*
291 1985;27: 254-271.
- 292 9. Doreen A, Carla A. Histologic study of the attachment of muscle to the rat mandible.
293 *Arch Oral Biol.* 1982;27: 519-527.
- 294 10. NIH consensus development panel on osteoporosis prevention, diagnosis, and
295 therapy. Osteoporosis prevention, diagnosis, and therapy. *JAMA.* 2001;285, 785-795.
- 296 11. Sasaki N, Sudoh, Y. X-ray pole figure analysis of apatite crystals and collagen
297 molecules in bone. *Calcif Tissue Int.* 1997;60: 361-367.
- 298 12. Wang J, Ishimoto T, Nakano T. Unloading-induced degradation of the anisotropic
299 arrangement of collagen/apatite in rat femurs. *Calcif Tissue Int.* 2017;100: 87-94.
- 300 13. Elliot JC. Structure and Chemistry of the Apatites and Other Calcium
301 Orthophosphates. Amsterdam: Elsevier Science; 1994.
- 302 14. Jimbo R , Nakano T , Sawase T. Alignment of Biological Apatite c-Axis Under
303 Functional Loading:A Preliminary Report. *Implant Dent.* 2016;25:594-598
- 304 15. Nakano T , Kaibara K , Ishimoto T, Tabata Y, Umakoshi Y. Biological apatite
305 (BAp) crystallographic orientation and texture as a new index for assessing the
306 microstructure and function of bone regenerated by tissue engineering. *Bone.* 2012;51:

- 307 741-747.
- 308 16. Vashishth D. The role of the collagen matrix in skeletal fragility. *Curr Osteoporos*
- 309 *Rep.* 2007;5: 62-66.
- 310 17. Benjamin M, Kumai T, Milz S, Boszczyk BM, Boszczyk AA, Ralphs JR. The
- 311 skeletal attachment of tendons--tendon "entheses". *Comp Biochem Physiol A Mol*
- 312 *Integr Physiol.* 2002;133: 931-945.
- 313 18. Xiao F, Hayashi H, Fujita T, Shirakura M, Tsuka Y, Fujii E, Tanne K, Tanimoto K.
- 314 Role of articular disc in cartilaginous growth of the mandible in rats. *APOC.*2017;7: 29-
- 315 34.
- 316 19. Fujita T, Ohtani J, Shigekawa M, Kawata T, Kaku M, Kohno S, Tsutsui K, Tenjo K,
- 317 Motokawa M, Tohma Y, Tanne K. Effects of Sex Hormone Disturbances on
- 318 Craniofacial Growth in Newborn Mice. *J Dent Res.*2004;83: 250-254.
- 319 20. Nakano T, Kaibara K, Tabata Y, Nagata N, Enomoto S, Marukawa E, Umakoshi Y.
- 320 Unique alignment and texture of biological apatite crystallites in typical calcified tissues
- 321 analyzed by microbeam X-ray diffractometer system. *Bone.* 2002;31: 479-487.
- 322 21. Huang AH, Riordan TJ, Wang L, Eyal S, Zelzer E, Brigande JV, Schweitzer R.
- 323 Repositioning forelimb superficialis muscles: tendon attachment and muscle activity
- 324 enable active relocation of functional myofibers. *Dev Cell.* 2013;26: 544-551.

- 325 22. Berendsen AD, Olsen BR. Bone development. *Bone*. 2015;80: 14-18.
- 326 23. Apostolakos J, Durant TJ, Dwyer CR, Russell RP, Weinreb JH, Alae F, Beitzel
327 K, McCarthy MB, Cote MP, Mazzocca AD. The enthesis: a review of the tendon-to-
328 bone insertion. *Muscles Ligaments Tendons J*. 2014;4: 333-342.
- 329 24. Lu HH, Thomopoulos S. Functional attachment of soft tissues to bone:
330 development, healing, and tissue engineering. *Annu Rev Biomed Eng*. 2013;15: 201-
331 226.
- 332 25. Matsumoto T, Matsunaga S, Morioka T, Nakano T, Yoshinari M, Yajima Y.
333 Relationship between preferential alignment of biological apatite and Young's modulus
334 at first molar in human mandible cortical bone. *J Hard Tissue Biol*. 2013;22: 163-170.

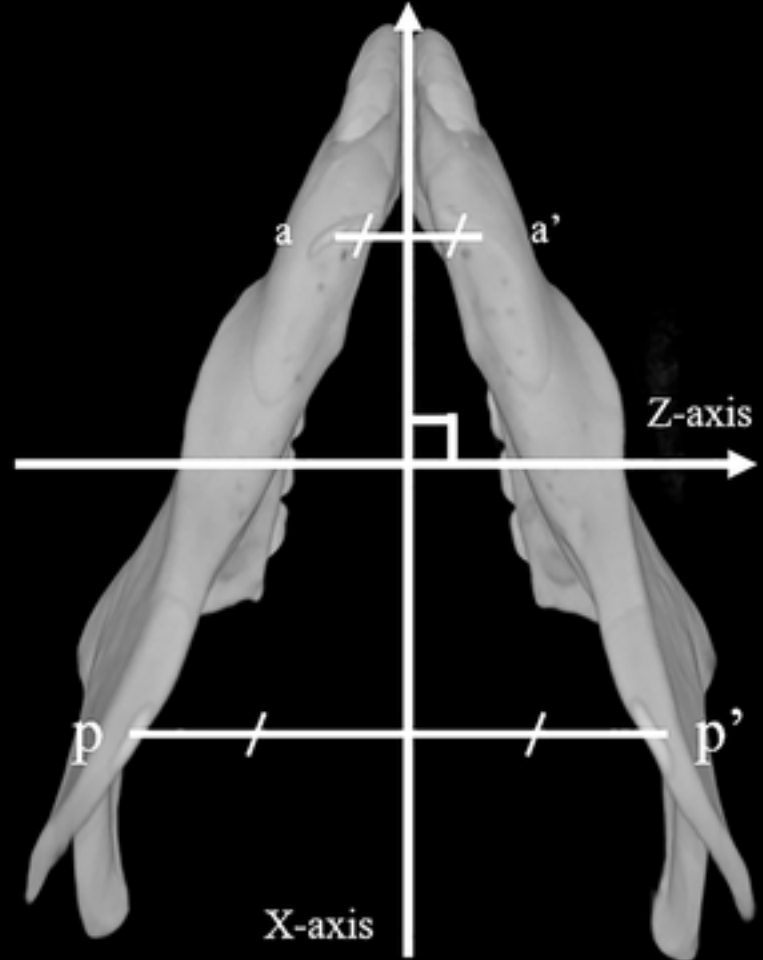
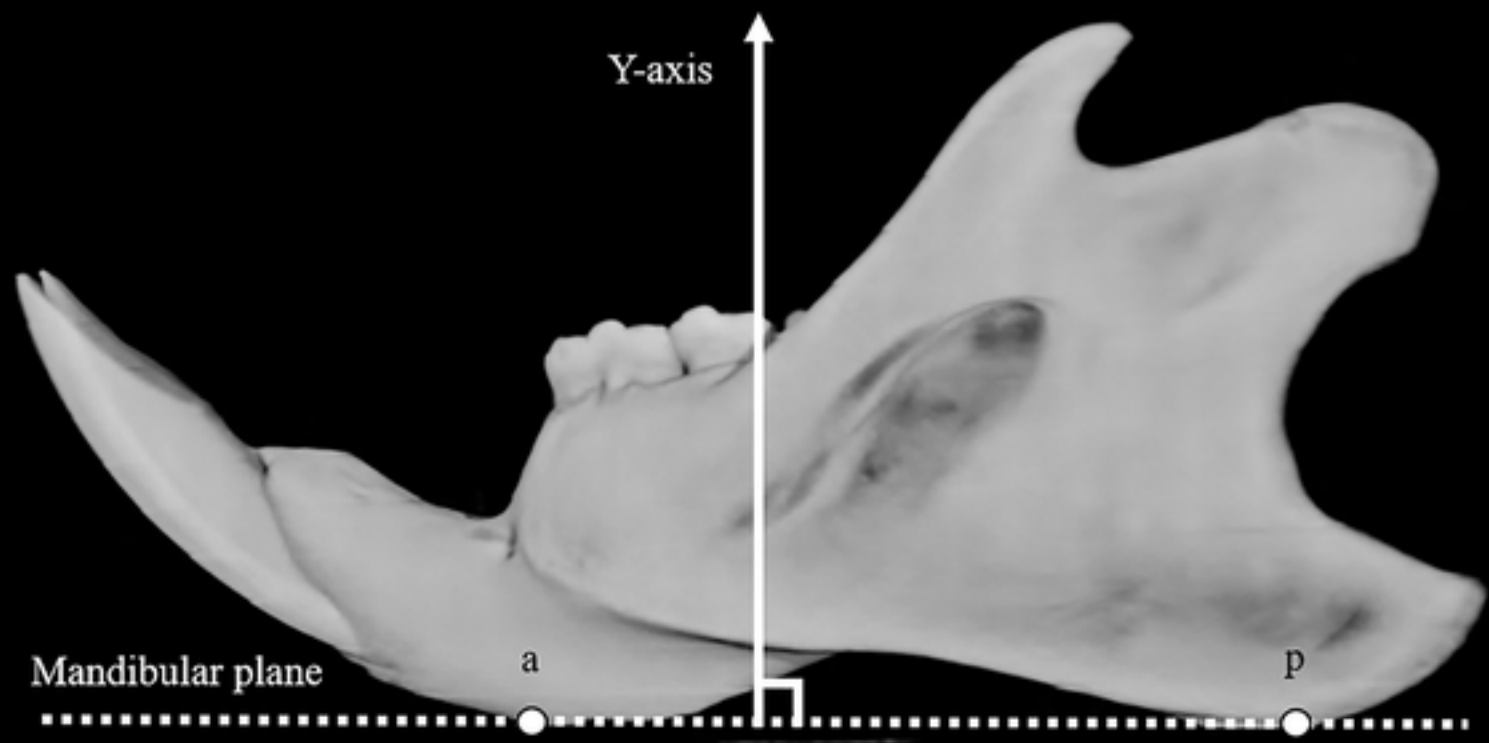


Fig 1

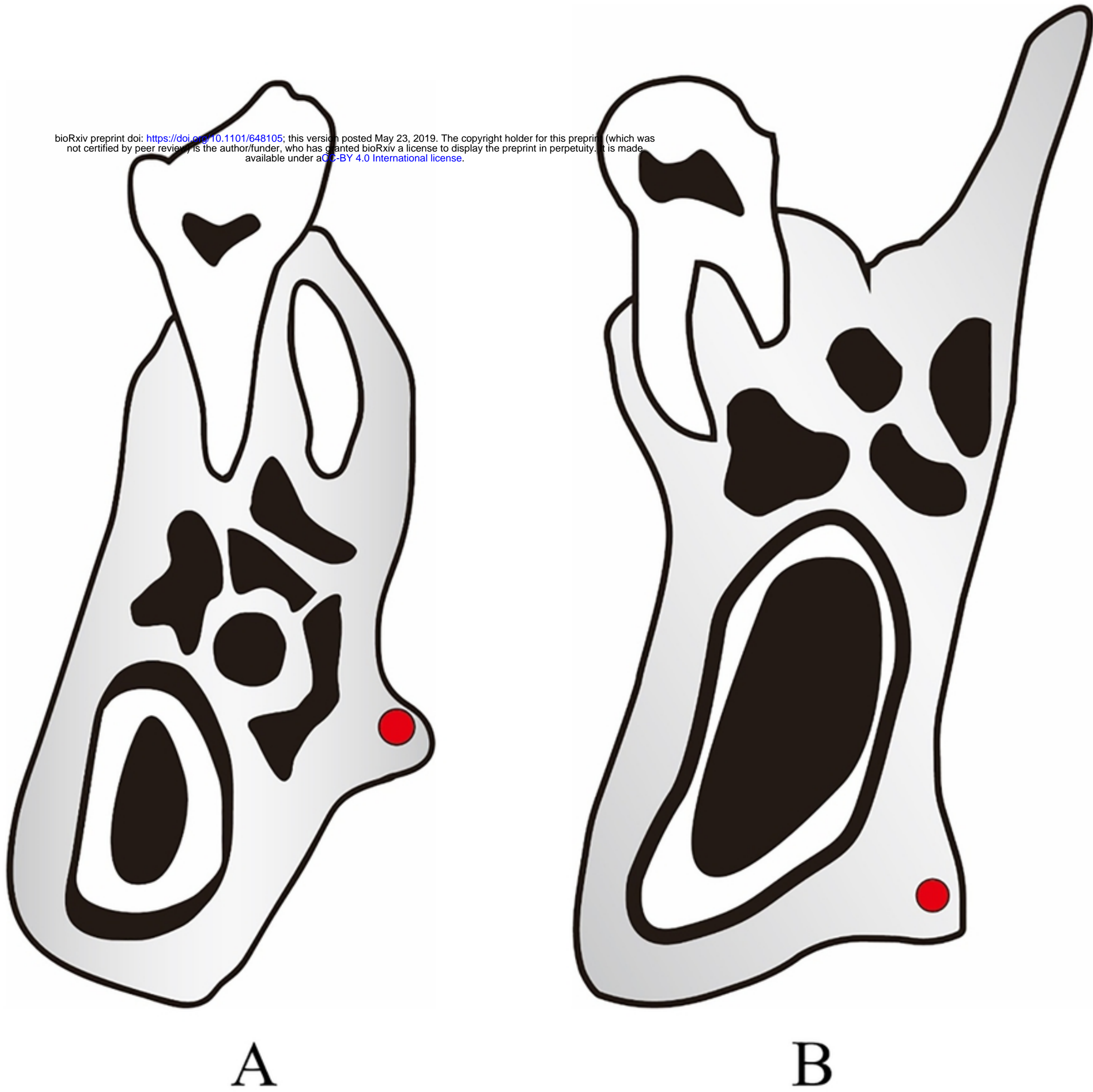


Fig 2

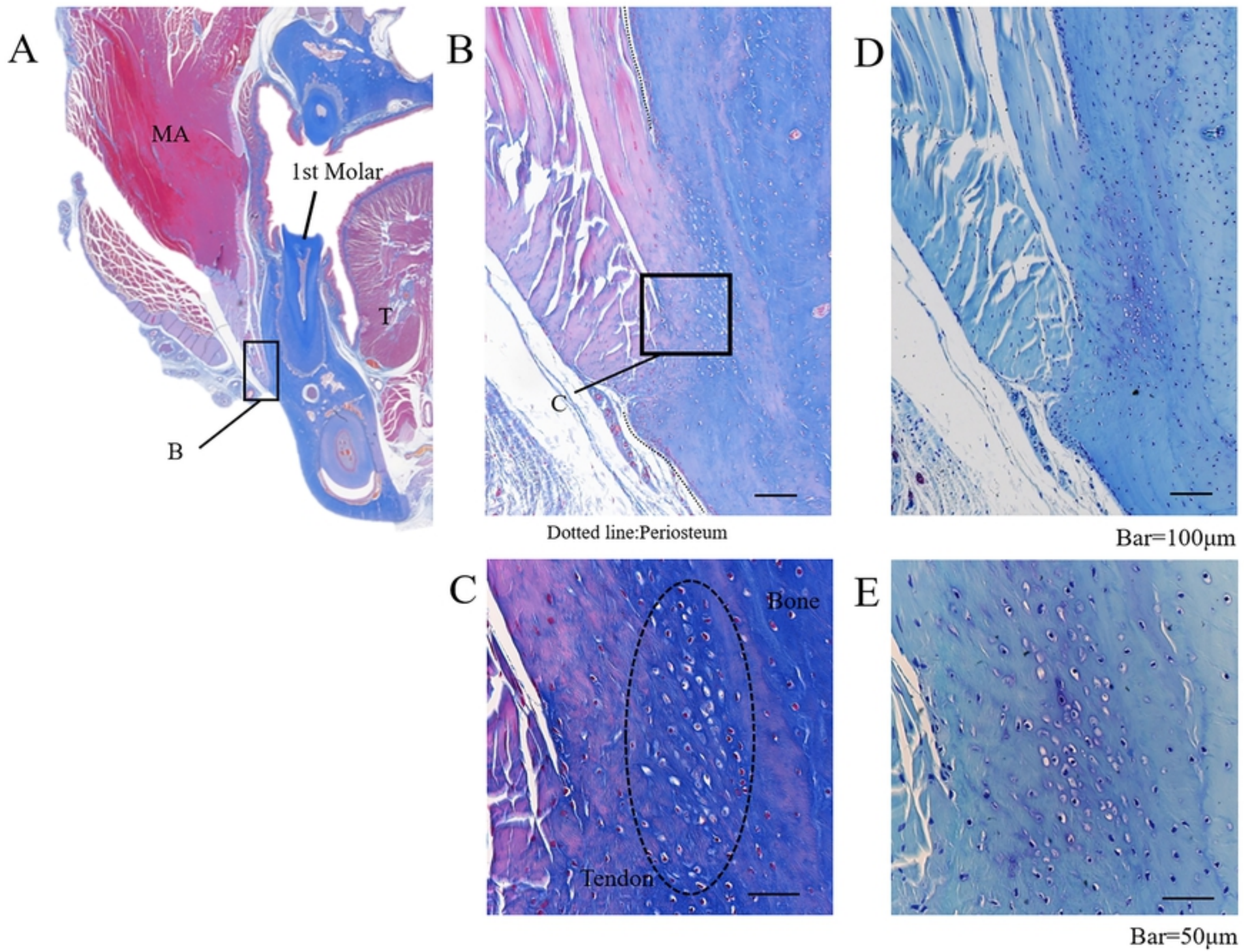


Fig 3

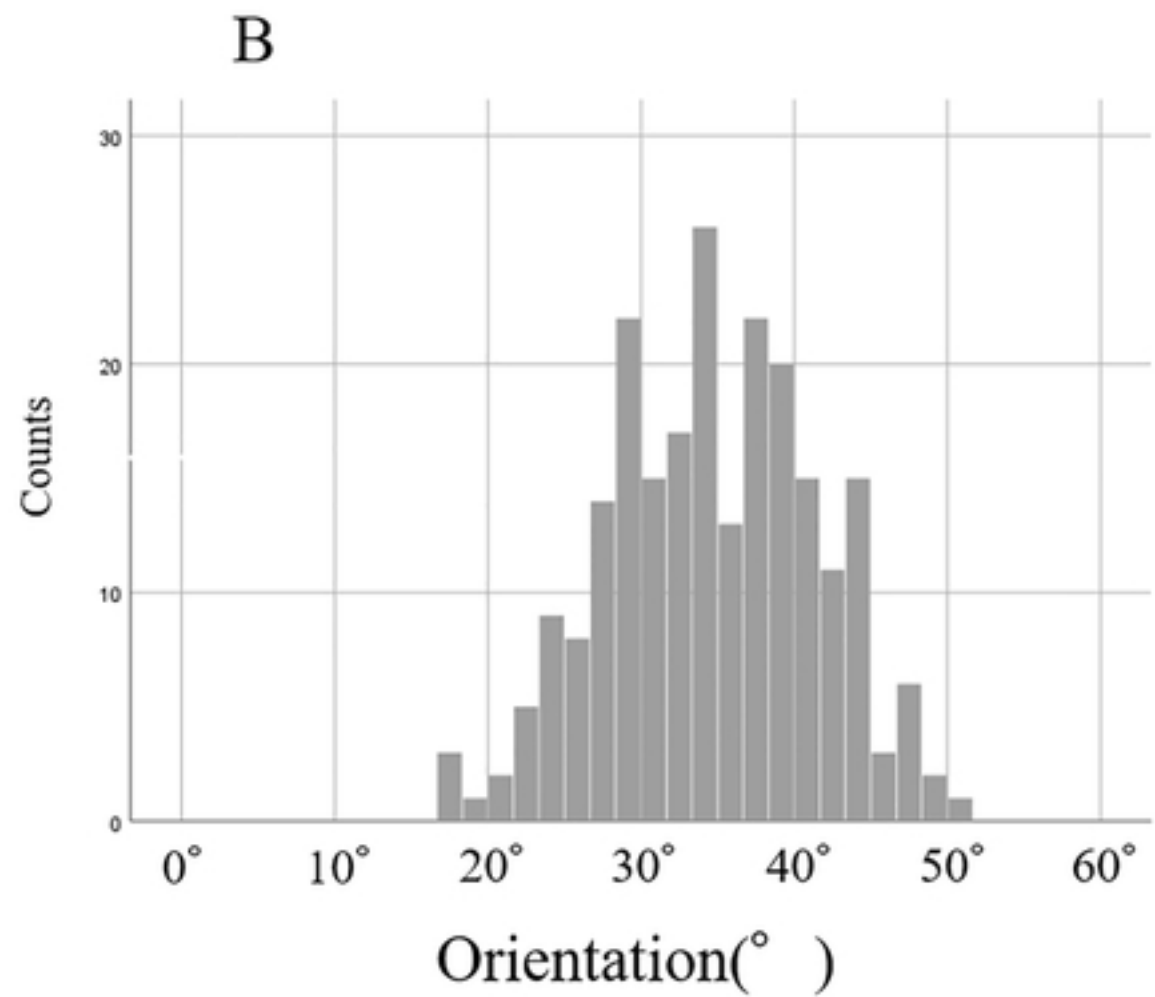
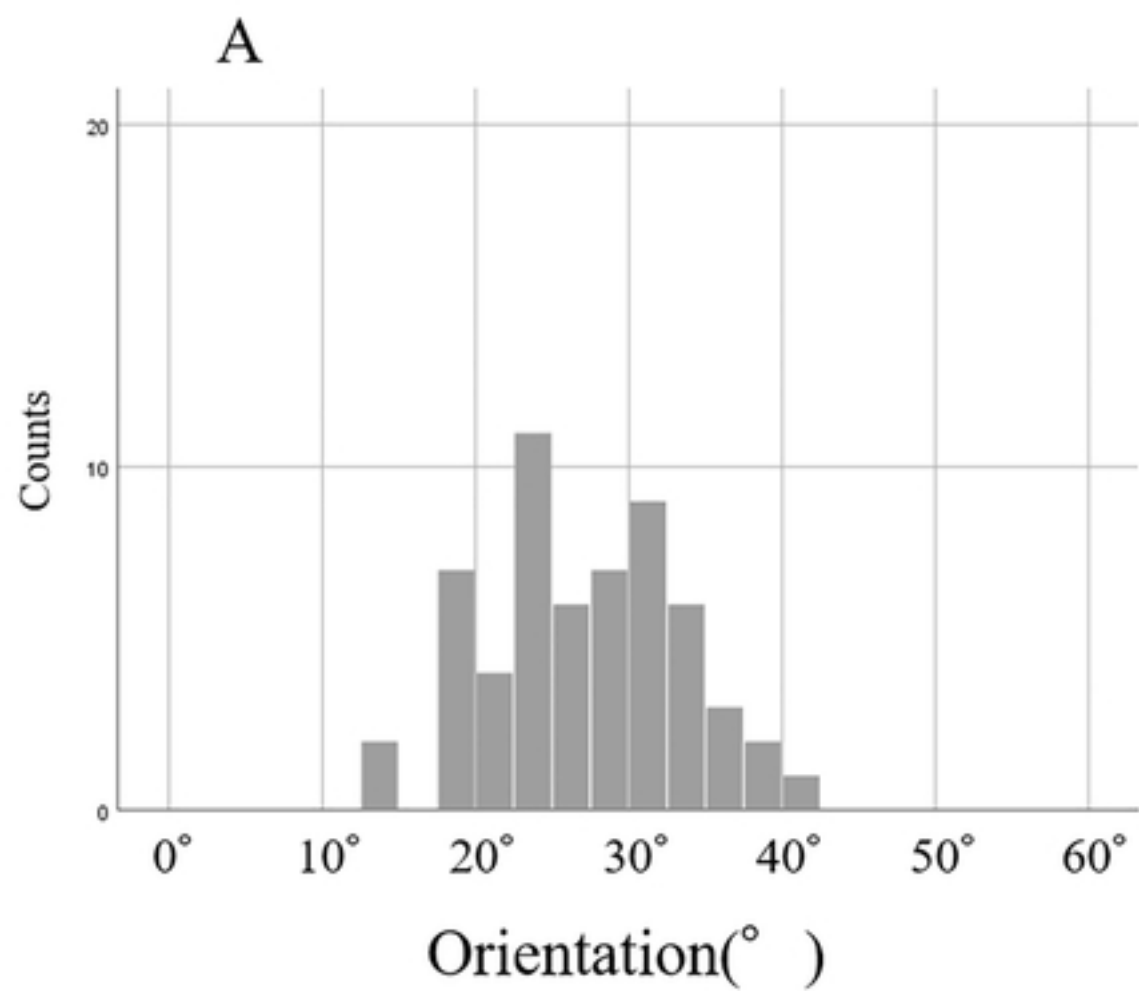
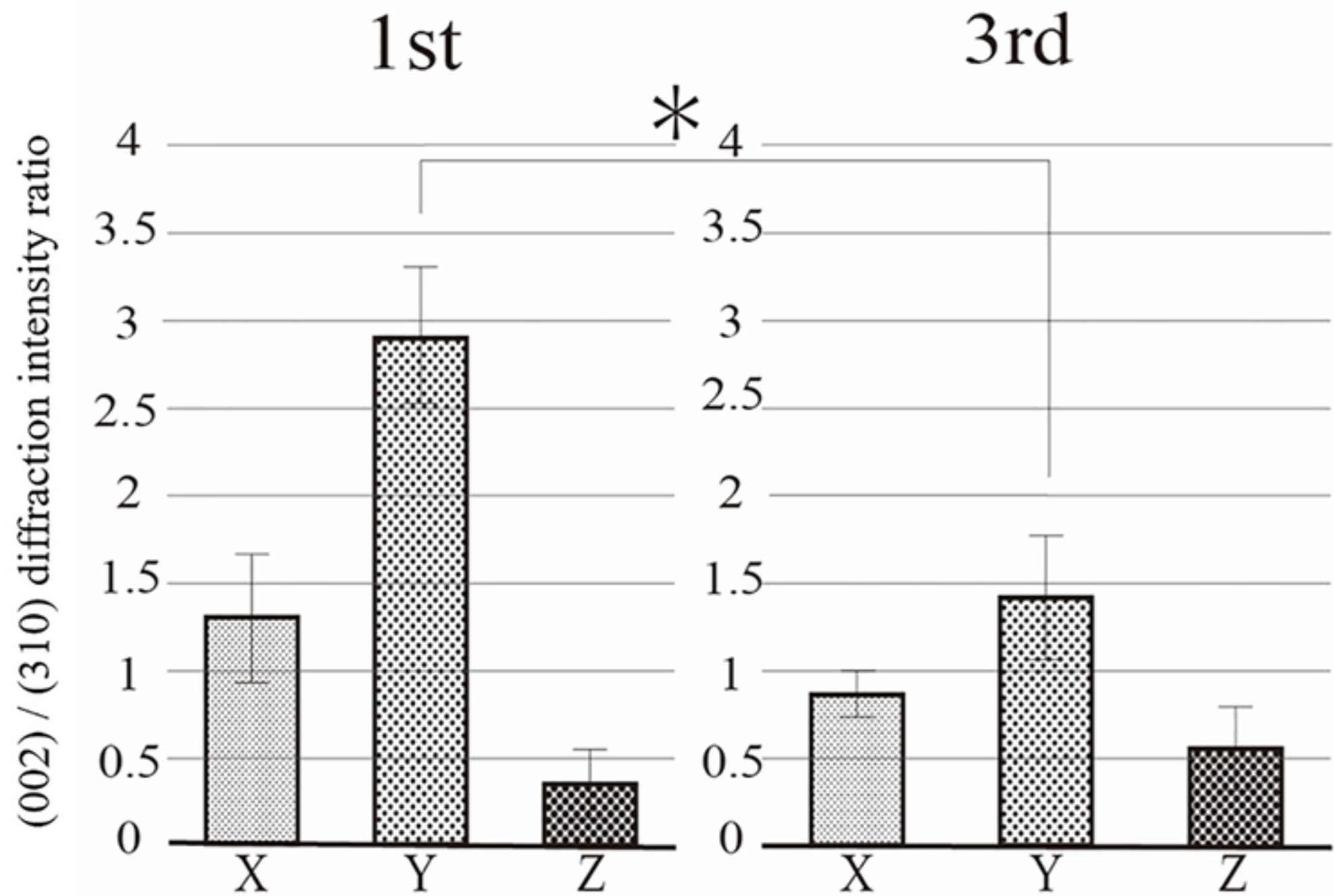


Fig 7



* : P<0.05

Randomly oriented hydroxyapatite powder; reflection mode:1.4
transmission mode:5.6

Fig 8

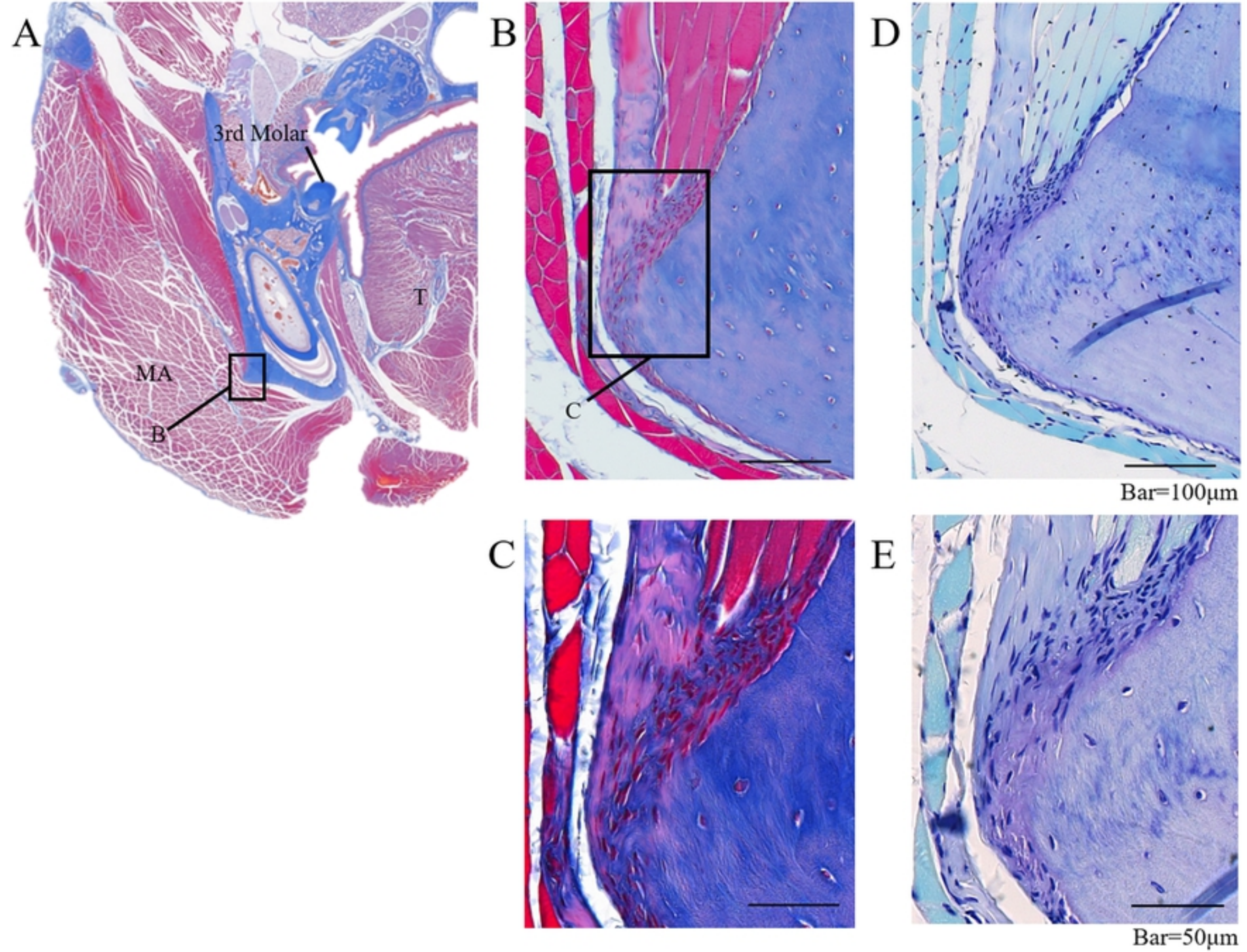


Fig 4

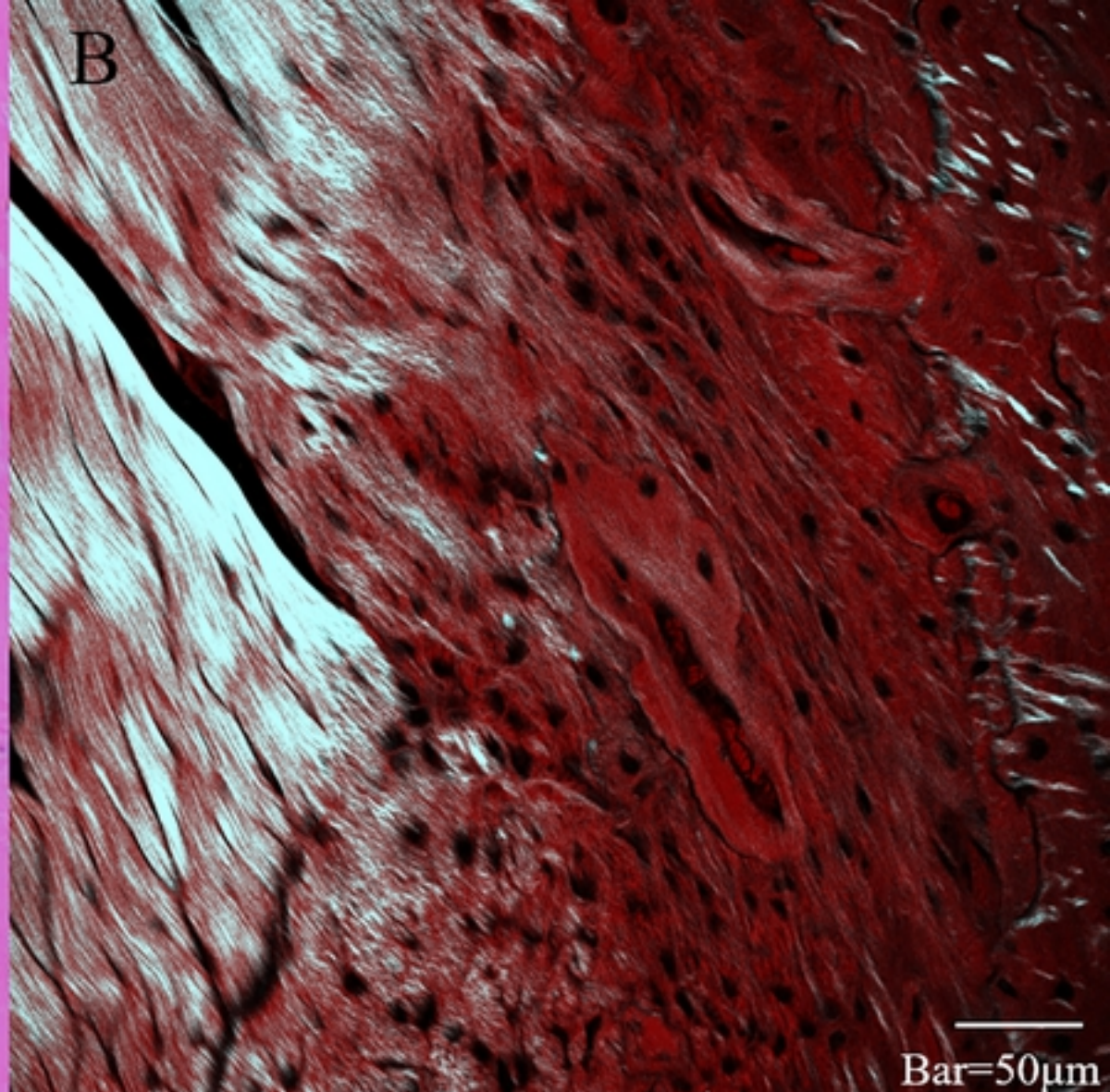
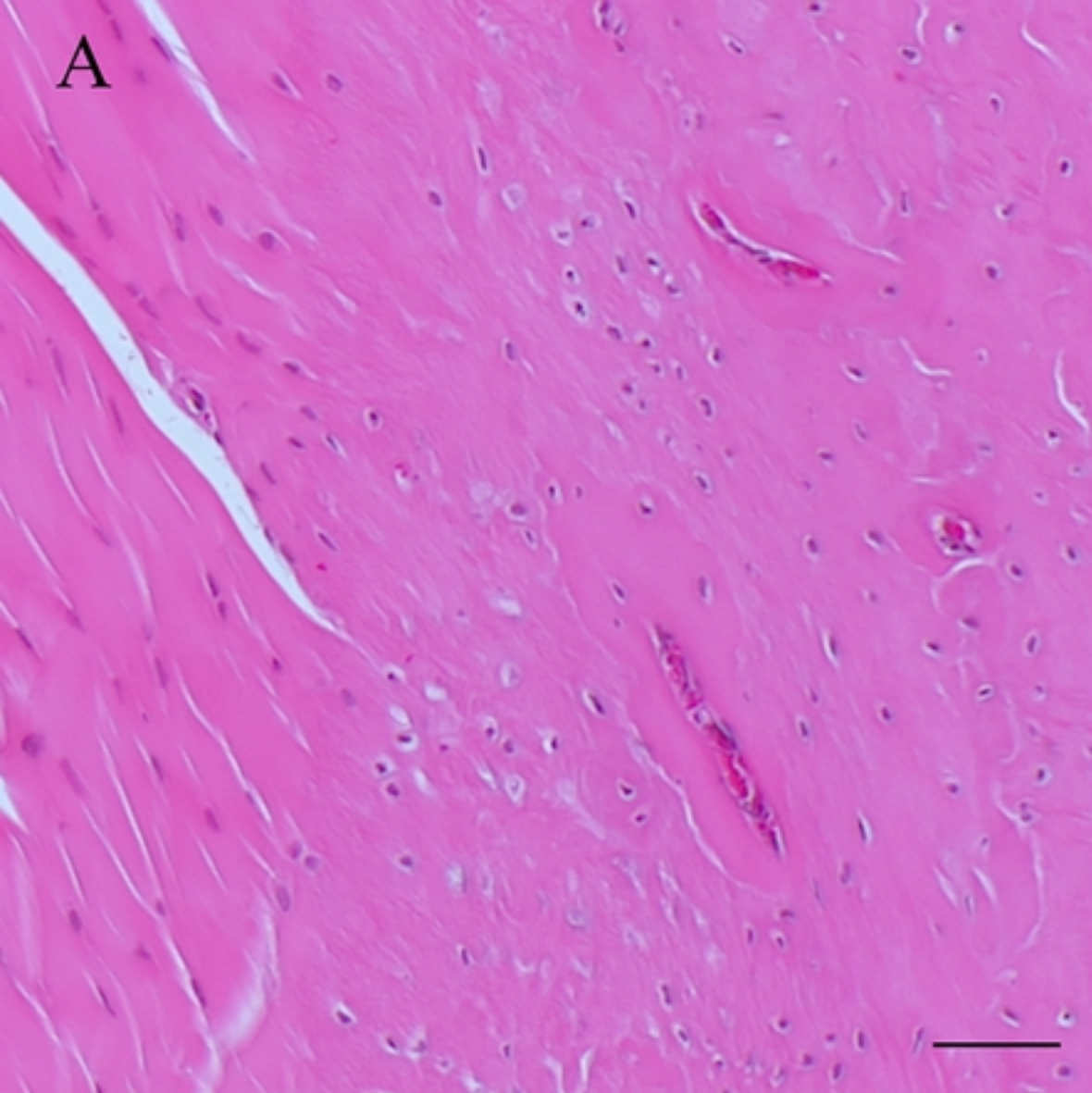


Fig 5

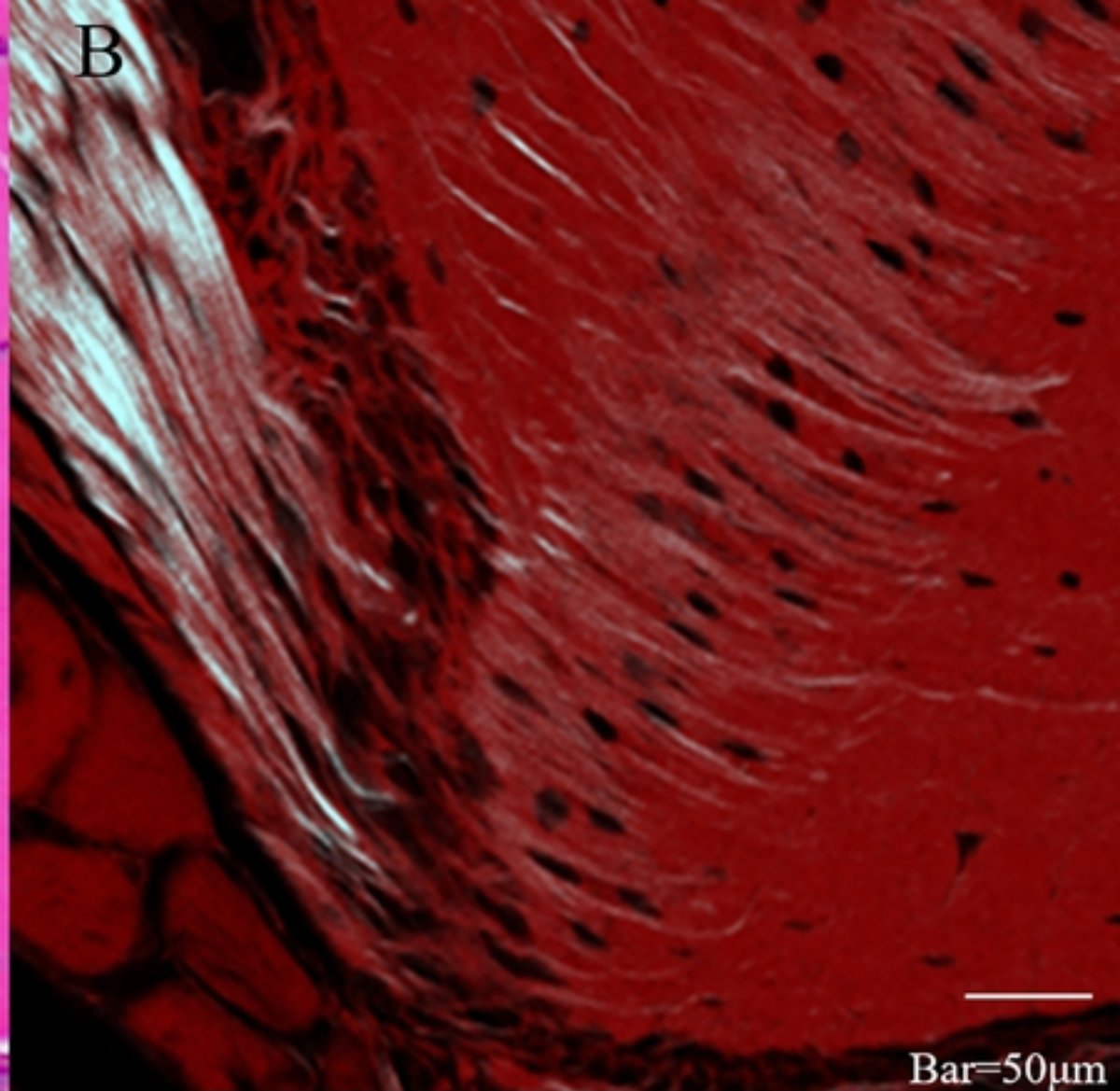
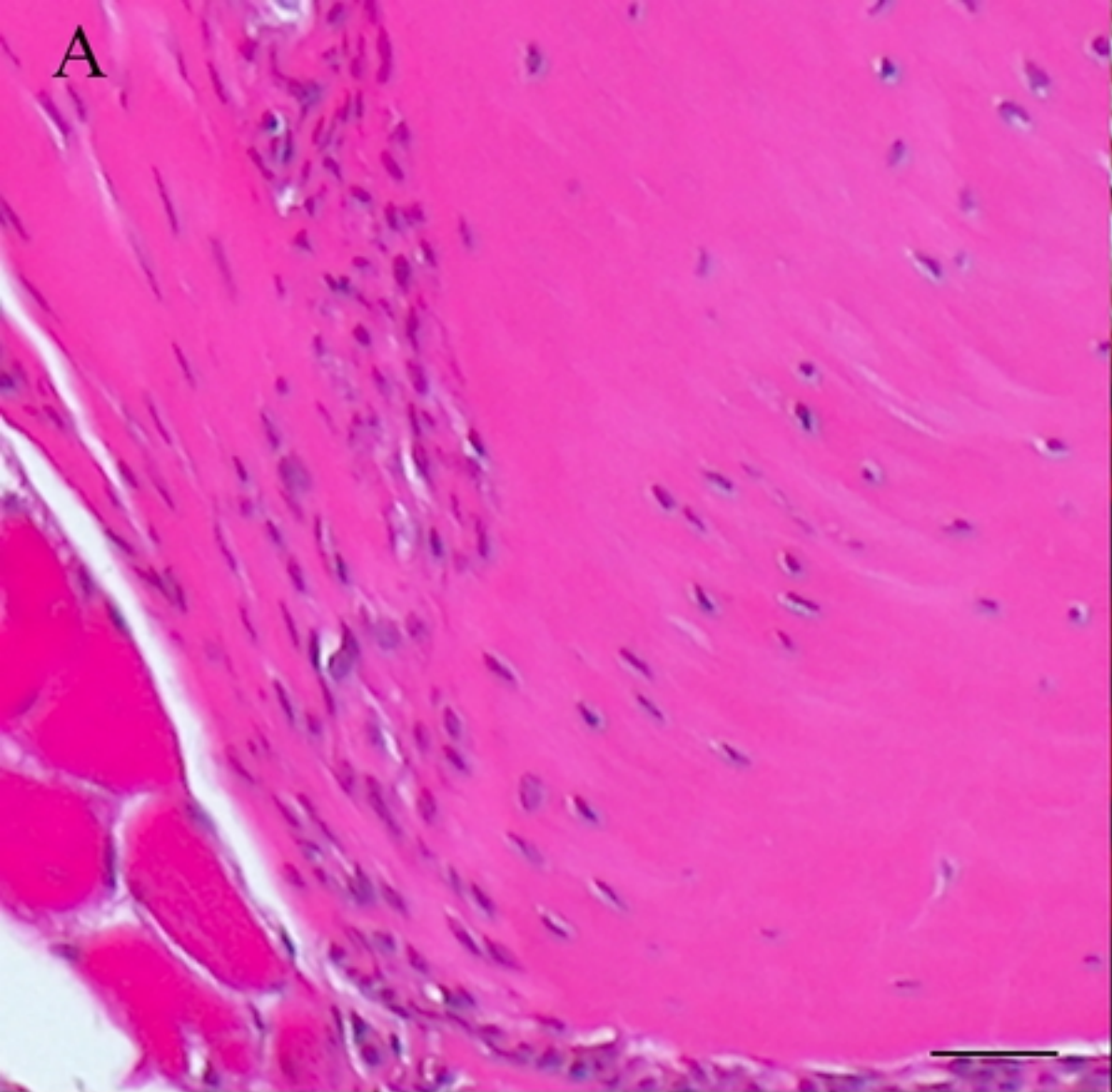


Fig 6