

1 **Land Use Change Increases Wildlife Parasite Diversity in Anamalai Hills, Western Ghats,**
2 **India**

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10 **ABSTRACT**

11 Anthropogenic landscape change such as land use change and habitat fragmentation are
12 known to alter wildlife diversity. Since host and parasite diversities are strongly connected,
13 landscape changes are also likely to change wildlife parasite diversity with implication for
14 wildlife health. However, research linking anthropogenic landscape change and wildlife
15 parasite diversity is limited, especially comparing effects of land use change and habitat
16 fragmentation, which often cooccur but may affect parasite diversity substantially
17 differently. Here, we assessed how anthropogenic land use change (presence of plantation,
18 livestock foraging and human settlement) and habitat fragmentation may change the
19 gastrointestinal parasite diversity of wild mammalian host species (n=23) in Anamalai hills,
20 India. We found that presence of plantations, and potentially livestock, significantly
21 increased parasite diversity due possibly to spillover of parasites from livestock to wildlife.
22 However, effect of habitat fragmentation on parasite diversity was not significant. Together,
23 our results showed how human activities may increase wildlife parasite diversity within
24 human-dominated landscape and highlighted the complex pattern of parasite diversity
25 distribution as a result of cooccurrence of multiple anthropogenic landscape changes.

26

27 **Keywords:** Parasite diversity, Land use change, Plantation, Livestock, Human settlement,
28 Anamalai hills, Habitat fragmentation, Rainforest

29

30 **INTRODUCTION:**

31 Land use change and habitat fragmentation are two major landscape-level outcomes of
32 human activities that significantly impact biodiversity¹⁻³. Consequently, considerable
33 research on biodiversity change in human-dominated landscape have been conducted,
34 which has resulted in improved understanding of how these two human impacts on
35 landscape can impact biodiversity^{1,4,5}. These anthropogenic factors can also modify host-
36 parasite interactions, which, in turn, can lead to either increase or decrease in parasite
37 diversity^{6-8,8}. Understanding how these factors may influence parasite diversity is
38 ecologically important for multiple reasons. For instance, parasites regulate host population
39 dynamics⁹, alter species communities¹⁰ and constitute a significant proportion of total
40 biomass of any ecosystem¹¹, which is not surprising considering parasites comprise at least
41 40% of all animal species on earth¹². Despite their ecological importance, our knowledge on
42 parasite diversity is limited^{13,14}, particularly in the context of increasing human impact on
43 environment, underlining a significant research gap^{15,16}. The gap is specifically wide for
44 wildlife hosts and urgent research is required in the face of recent increased emergence of
45 novel pathogens of wildlife origin^{7,17,18}. It is, thus, crucial to answer how anthropogenic land
46 use change and habitat fragmentation may impact parasite diversity in the wild.

47 Land use change can affect parasites both directly and indirectly. By altering
48 environment (for example, through pollution), land use change may render transmission of
49 environmentally-transmitted parasites difficult. This is particularly true for parasites that has
50 life stages outside host body. However, land use change can indirectly impact parasite
51 diversity by altering host diversity as it is one of the strongest predictors of parasite diversity
52¹⁹⁻²². By decreasing host diversity and abundance, land use change can deplete richness of
53 parasites particularly those that require multiple obligatory hosts²³. This is evident when
54 many host species that are threatened in their natural habitat appear to harbour fewer
55 parasites²⁴. On the other hand, land use change can also increase parasite diversity in
56 multiple ways. Land use change can increase parasite diversity by increasing host diversity.
57 For instance, land use change such as agricultural field or land-fill can act as resource traps
58 and amplify host diversity artificially²⁵. Land use change can also increase parasite diversity
59 by introducing non-native parasites such as parasites of domestic and feral animals and
60 even from humans²⁶.

61 It is also important to distinguish between different types of land use change and
62 their effects on parasites²⁷. One type of land use change that has not been studied well is
63 the effect of plantation on wildlife parasites²⁷. Plantations are usually monocultures of
64 exotic or native plant species grown as timber or fuel wood or as cash crops and have a
65 large and increasing footprint in wildlife habitats worldwide²⁸. They can sometime act as
66 refuge to wildlife but usually with a biotic homogenising effect^{29,30}. Consequently, plantation
67 may also increase but homogenise parasite community. Plantations are often accompanied
68 by settlement of labourers and livestock foraging^{31,32}. These changes within a wildlife
69 habitat can both increase or decrease parasite diversity. Parasite diversity may decrease if
70 wildlife hosts avoid human areas to lessen confrontation with humans and resource
71 competition with livestock. On the other hand, generalist species may actually thrive in
72 human settlements by utilizing novel resources^{33,34}. Herbivore species may also prefer to
73 stay closer to human settlements and livestock (“spatial refugia”) that may displace
74 predators^{35–38}. Moreover, many wildlife, over time, may actually get habituated to humans
75 and livestock and aggregate near human-dominated landscape^{39,40}. These aggregations may
76 eventually increase parasite diversity by increasing contact between native host species.
77 Such situations may also increasingly expose wildlife to humans and human-associated
78 animals such as livestock and commensals, increasing chance of spillover of non-native
79 parasites to wildlife.

80 Habitat fragmentation may lead to higher parasite diversity because heavily
81 fragmented habitats may disrupt wildlife dispersal and increase host diversity in smaller
82 fragments. Such increase in host diversity in a smaller patch may alter host characteristics
83 such as home range, abundance and intra and interspecific contacts thus increasing overlap
84 among host species making host individuals exposed to higher parasite infections^{41,42}.
85 These effects are likely to be greatest in the smallest and most isolated of the fragments^{3,43}.
86 By disrupting host dispersal, fragmentations can also adversely affect parasite diversity. This
87 could be especially true for parasites who require multiple host species to complete its life
88 cycle, such as those that are transmitted trophically⁴⁴. So far, many studies looked into this
89 effect but the results have been mixed^{6,41,45–47}.

90 The Anamalai (Elephant hills in *Tamil*) hills of southern India is a highly biodiverse
91 rainforest habitat of Western Ghats, which holds about 30% of India's plant and vertebrate

92 species diversity in less than 6% of the country's area ⁴⁸. It is also one of the most altered
93 natural habitats in India and typifies different levels of land use change and habitat
94 fragmentation rampant in Indian wildlife habitats. Large section of the habitat is highly
95 modified due to land use change, bordered by large, relatively undisturbed tropical
96 rainforests. The landscape is a matrix of over 40 rainforest fragments (1-2,500 ha in
97 size), often surrounded by plantations (coffee, tea and cardamom), roads, hydroelectric
98 dams and settlements ⁴⁹. Highly-modified fragments contain within them human
99 settlements and have higher livestock pressures than other remote, less disturbed
100 fragments. In spite of such high levels of land use change and habitat fragmentation, the
101 Anamalai hills still harbour a large number of wildlife whose ranges often unavoidably
102 overlap with humans and livestock ⁵⁰⁻⁵³. In fact, large number of wildlife species are
103 regularly observed within human-dominated habitats and this concurrence with humans
104 often precipitates into wildlife-human conflicts ^{49,50,54-56}. It is possible that many of the
105 wildlife are important reservoirs of multiple environmentally-transmitted parasites. In fact,
106 recent studies have recorded important parasite groups within certain host species
107 populations that may cause Ascariasis, Trichuriasis and Strongyloidiasis in humans ^{45,46,57,58}.

108 To assess the effect of land use change (plantation, livestock foraging and human
109 settlements) and habitat fragmentation on parasite diversity, we studied gastrointestinal
110 parasites of wild mammalian hosts across rainforest fragments in Anamalai hills. Using
111 statistical models, we tested effects of land use change and habitat fragmentation on
112 parasite diversity. We predicted a positive impact of land use change on parasite diversity
113 due to increased host diversity and an increased exposure of wildlife to humans and
114 livestock. For habitat fragmentation too, we predicted an increase in parasite diversity with
115 decrease in habitat size and increase in habitat isolation. Our alternative predictions were
116 that land use change and habitat fragmentation could actually deplete parasite diversity by
117 decreasing host diversity in disturbed fragments. Finally, land use change and habitat
118 fragmentation may not significantly impact parasite diversity either by not impacting host
119 community or by not spillover from non-native hosts such as livestock and humans.

120 **MATERIALS AND METHODS:**

121 **Ethical statement:** For this study, faecal samples were collected only noninvasively. As a
122 result, no animal was sacrificed or harmed during sampling. Part of the sampling was done
123 within Anamalai Tiger Reserve, which is a protected area. Hence, appropriate written
124 permission was taken from the Tamil Nadu Forest Department (Letter Ref. No. WL
125 5/58890/2008, dated 2nd September 2009).

126 **Study site:** Located south of the Palghat gap (11° N) of the Western Ghats, Anamalai hills
127 once had large tracts of tropical rainforest dotted with few tribal settlements. Between
128 1860 and 1930, British colonisers started clearing the rainforests extensively for cultivation
129 of tea and coffee and developing teak and *Eucalyptus* plantations, particularly in the
130 Valparai Plateau⁵⁹. As a result, the Anamalais today consists of both a relatively
131 undisturbed, large (958.59 km²) tropical rainforest within the protected Anamalai Tiger
132 Reserve (ATR; 10°12'–10°35'N and 76°49'–77°24'E) and about 1,000 ha highly degraded
133 Valparai Plateau (Figure 1). The plateau consists of many tea estates and other plantations,
134 which are surrounded by four protected areas—ATR in Tamil Nadu state and three others in
135 Kerala state. The major vegetation types include scrub forests in the rain-shadow areas in
136 the eastern foothills, dry and moist deciduous forests (<800 m), mid-elevation tropical wet
137 evergreen forest (600-1,500 m) and high-altitude shola-grassland ecosystems (>1,500 m)⁶⁰.
138 Although a large part of the tropical wet evergreen forests occurs within ATR, many of the
139 smaller (< 200 ha) fragments are found in private estates in the Valparai plateau. These
140 small fragments are highly degraded and disturbed due to fuel-wood collection and
141 livestock grazing. Valparai town also is a part of the plateau and around 200,000 people live
142 across the town and plantations⁶⁰. Due to the ongoing habitat fragmentation, the whole
143 landscape is a matrix of over 40 rainforest fragments, ranging 1 ha-2,500 ha in size and
144 often surrounded by plantations (coffee, tea and cardamom), roads, hydroelectric dams and
145 settlements⁴⁹. Based on size range (2-2,500 ha), level of perceived human disturbance and
146 access, we selected 19 mid-elevation tropical rainforest and three low-elevation dry and
147 moist deciduous forest fragments for sampling (Figure 1).

148 **Host sampling:** Between Oct 2013 and Oct 2015, faecal samples were collected from
149 populations of mammalian wildlife. We collected fresh faecal samples, non-invasively during
150 the day, on transects (400 m-3 km in length). For large and medium herbivores and

151 primates, we followed individuals and collected fresh faeces when animals defaecated. For
152 elusive species such as carnivores, we identified home range based on secondary
153 information and faecal samples were identified based on morphology and also using nearby
154 secondary signs such as pug-marks or hoof-prints. To avoid sampling the same individual
155 repeatedly, only one sample of a host species was collected from each spot and the sample
156 source was either marked or removed. To avoid contamination from soil, samples were
157 collected from the inside of the bolus or only top pellet was collected from a heap. We
158 immediately fixed each sample in 10% formaldehyde solution (50 ml), labelled the
159 containers with the information of origin (fragment name, date, Time and host species) and
160 stored them at room temperature until parasitological screening. Differences in sampling
161 effort can confound the comparison of diversity among replicates. We accounted for
162 differences in number of host species encountered by calculating richness estimates with
163 the assumption that each faecal sample represents single individual. We used bootstrap,
164 which is a resampling method for estimating the whole sampling distribution of richness by
165 sampling with replacement from the original sample and can offer greater precision than
166 jackknife estimates, especially when sample sizes are small ⁶¹.

167 **Parasite sampling:** Employing both the flotation and sedimentation techniques (NaNO₃
168 solution), we screened the faecal samples for the presence of helminth eggs, larvae and
169 protozoan cysts ⁶². For each parasite concentration technique, we examined two slides per
170 sample. Slides were examined under a light microscope (400X). Eggs and cysts were first
171 examined at 10× magnification and then their size was measured with a micrometre
172 eyepiece (0.1 µm) at 40× magnification. To facilitate identification of parasite eggs, we often
173 added a drop of Lugol's iodine solution to the slides, which highlighted detailed structures.
174 In addition, photographs of each parasite species have been archived and are available for
175 examination by request to the corresponding author. We identified parasites to the lowest
176 possible taxonomic level using published keys ^{63,64}. Differences in sampling effort can
177 confound the comparison of diversity among replicates. We accounted for differences in
178 number of parasite taxon encountered by calculating richness estimates with the
179 assumption that each faecal sample represents single host individual. We used bootstrap,
180 which is a resampling method for estimating the whole sampling distribution of richness by
181 sampling with replacement from the original sample. Bootstrap can offer greater precision

182 over jackknife estimator, especially when sample sizes are small⁶¹. This method is
183 particularly recommended for parasite richness estimation⁶⁵.

184 **Land use data:** In Anamalai hills, land use change manifests in largely three forms—
185 presence of human settlements, plantations and livestock foraging. There are only few large
186 (>1000 ha) fragments that are legally protected and thus undisturbed. Many of the studied
187 fragments share more than one type of land use change. For instance, some fragments with
188 human settlements may also have livestock present. For the current study, we identified 18
189 fragments with land use change, out of which 15 (83.3%) had plantation, in contrast to three
190 (16.7%). Eleven (61.1%) of the fragments have significant livestock foraging pressure, in
191 contrast to seven (38.9%) fragments without livestock. Finally, ten (55.6%) of the fragments
192 had human settlements within them, in contrast to eight (44.4%) without settlements.

193 **Habitat fragmentation data:** To measure effect of habitat fragmentation, we used fragment
194 size and isolation distance between fragments. According to the equilibrium theory of island
195 biogeography, organism dispersal probability declines as distance between islands
196 increases, reducing rates of immigration and, in turn, reducing diversity (MacArthur &
197 Wilson, 1963, 1967; Whittaker & Fernandez-Palacios, 2007). Assuming each forest fragment
198 as an island, their isolation was summarized with an isolation index which was calculated as
199 the sum of the square root of the distances to the nearest equivalent (no smaller than 80%
200 of size) or larger fragment (Dahl, 2004). Data on fragment size, distance between fragments
201 and presence of human settlements, plantations and livestock were collected from earlier
202 studies from Anamalai hills^{45,60}.

203 **Data analyses:** To assess the effects of land use change and habitat fragmentations on
204 bootstrap estimate of parasite taxon richness, we created two different linear mixed effects
205 models⁶⁶. Each model included random effects of host species and fragments to account for
206 multiple observations within each fragment (across host species) and across fragments. In
207 the land use model, the predictor variables (fixed effects) were presence of plantation,
208 human settlement and livestock. The predictor variables for the habitat fragmentation
209 model were fragment size and fragment isolation index. In both the models, we
210 incorporated both bootstrap estimates of host species richness and host body mass as co-
211 predictors as these were known to effect parasite richness. We retrieved host body mass
212 data from online ecological database⁶⁷. After fitting these model to the data, we also

213 compared and selected the best fit model using lowest AIC value⁶⁸. At the end, diagnostics
214 were run to check distribution of the residuals for each model. This analysis was conducted
215 in the lme4 package⁶⁹. We also assessed the effects of land use change and habitat
216 fragmentation on bootstrap estimates of host species richness using two linear models. In
217 the land use model, the predictor variables were presence of plantation, human settlement
218 and livestock. The predictor variables for the habitat fragmentation model were fragment
219 size and fragment isolation index. We followed the same strategy as described above for
220 model fitting, fitting diagnostics and model selection. Finally, we tested whether land use
221 change homogenized the composition of the parasite community. We used a multivariate
222 nonparametric Analysis of Variance (permAnoVa; 1,000 permutations) based on the Jaccard
223 dissimilarity index for a matrix of parasite presence/absence. We calculated the variance of
224 homogeneity of parasite communities within each fragment based on disturbed vs.
225 undisturbed divisions using the betadisper function of the vegan package in R⁷⁰.

226

227 **RESULTS:**

228 **Sample diversity:** From 19 forest fragments, we collected 4,056 mammalian faecal samples
229 belonging to 23 mammalian wildlife species and two livestock species—domestic goats
230 (*Capra aegagrus*) and cattle (*Bos taurus*). Analyses were done only on wildlife samples.
231 Number of samples varied from 41 in Uralikal to 495 in Puthuthottam (Table 1). Number of
232 samples for each host species varied between six in Otter (*Lutra lutra*) and 623 in gaur (*B.*
233 *gaurus*). In total, seven protozoa (18.42%) and 32 helminth (81.58%) species were recorded,
234 including five trematodes, five cestodes and 20 nematodes. At least seven different
235 parasites, belonging to different parasite groups, were recorded in ≥ 20 different host
236 species—protozoa *Coccidia sp.* (23 hosts); cestodes *Hymenolepis nana* (20 hosts) and
237 *Moniezia sp.* (22 hosts); and nematodes *Gongylonema sp.* (20 hosts), *Strongyloides sp.* (23
238 hosts), *Trichuris sp.* (24 hosts) and *Ascaris sp.* (26 hosts). On the other hand, cestode
239 *Dipylidium sp.* and nematode *Parascaris sp.* were found only in civet and Indian porcupine
240 (*Hystrix indica*) samples, respectively.

241 **Host and parasite diversity and disturbance:** For parasite diversity analysis the human
242 disturbance model was the best fit (Table 2). Parasite diversity was significantly driven by

243 presence of plantation (estimate = 4.779, $CI_{Profile} = 0.326 - 9.232$, $t = 2.103$, $p < 0.05$).
244 Presence of livestock had a substantial but not significant positive effect (estimate = 3.209,
245 $CI_{Profile} = -0.052 - 6.366$, $t = 1.992$, $p > 0.05$). Effects of settlement, host richness and host body
246 mass on parasite richness were not significant (Figure 2). For host diversity analysis the
247 human disturbance model was again the best fit (Table 3). Presence of plantation was the
248 only predictor that had a significant positive effect on host diversity (estimate = 10.798,
249 $CI_{Profile} = 2.302 - 19.294$, $t = 2.726$, $p < 0.05$)—almost half of all host species occur in
250 plantations. Although presence of livestock did not have a significant effect, its wide
251 confidence interval was mostly on the positive side suggesting potential positive impact—
252 limited by sample size—on host richness (estimate = 5.602, $CI_{Profile} = -0.639 - 11.843$, $t =$
253 1.925 , $p > 0.05$). Similarly, presence of human settlement did not significantly affect host
254 richness, however, the substantial effect was mostly on the negative side, suggesting
255 potential negative effect on host diversity (estimate = -4.112, $CI_{Profile} = -10.717 - 2.492$, $t = -$
256 1.335 , $p > 0.05$).

257 We recorded 12 parasites (ten helminths and two protozoa) that occurred only in
258 plantations. Six of the ten helminths were nematodes (60%), while rest were trematodes
259 (30%) and one cestode (10%). Fragments without plantations did not harbour any parasite
260 taxon exclusively, which means parasites in those undisturbed fragments also occurred in
261 plantations. Fragments with livestock harboured three parasite taxa (two nematodes and
262 one cestode) exclusively relative to their undisturbed counterpart. However, only one
263 parasite taxon (*Taenia sp.*) exclusively occurred in livestock disturbed fragments, while other
264 two nematodes also occurred in the plantations. Its counterpart undisturbed fragments only
265 harboured one taxon exclusively (*Paragonimus sp.*), which however also occurred in
266 plantations. Finally, settlements harboured three nematode taxa exclusively in comparison
267 to their undisturbed counterpart. Only one of these taxa (*Uncinaria sp.*) were exclusive to
268 settlements across all fragments. Undisturbed counterpart of settlements harboured only
269 one parasite taxon (*Sarcocystis sp.*) exclusively.

270 **Parasite and host homogeneity:** Parasite communities within disturbed forest fragments
271 were not significantly more homogeneous than the undisturbed ones due to presence of
272 either plantations ($F = 2.58$, $p > 0.05$), livestock ($F = 0.04$, $p > 0.05$) or settlements ($F = 3.55$,
273 $p > 0.05$). Host communities within plantations (TukeyHSD; $p < 0.05$; Figure 4a) and human

274 settlements (TukeyHSD; $p < 0.05$; Figure 4b) were, however, significantly more homogeneous
275 than undisturbed fragments. Finally, we did not find any of the disturbance variables to
276 significantly alter the parasite community composition between undisturbed and disturbed
277 fragments.

278

279 **DISCUSSION:**

280 Our results reveal that rainforest fragments with plantations (and potentially with livestock)
281 in Anamalai hills harbour significantly higher parasite diversity than undisturbed fragments.
282 Interestingly, some of the modified fragments (at least, fragments with plantations) also has
283 significantly more host diversity than the undisturbed fragments, however host diversity
284 was not found to significantly affect parasite diversity.

285 In Anamalai hills, plantations (coffee, tea and cardamom) had more mammalian
286 wildlife species richness than the undisturbed fragments. This was not particularly a
287 surprising result because studies have reported similar high richness in vertebrate species
288 from plantation within wildlife habitats^{71,72}. In fact, earlier studies from Western Ghats also
289 found high vertebrate richness within or around plantations with large variations depending
290 on plantation types, from open tea to more shaded coffee and cardamom plantations
291^{30,73,74}. The reason for such increased host diversity is thought to be an increase in habitat
292 heterogeneity within plantations. Increased habitat heterogeneity is thought to generate
293 greater diversity of niches consequently facilitating cooccurrence of many species^{75,76}.
294 However, such increase in species richness is often accompanied by more generalist and
295 wide-ranging species being more abundant within the plantations and a loss of community
296 heterogeneity relative to undisturbed habitats⁷⁷⁻⁷⁹. We found similar loss of heterogeneity
297 for host species in disturbed habitats with plantations and settlements (Figure 4).

298 Effect of livestock presence on host species richness was positive but not statistically
299 significant at $\alpha = 0.05$. The effect, however, was significant at $\alpha = 0.10$, which suggested
300 potential, but weak effect, which was reflected by the almost equal number of wildlife
301 species recorded from these two groups of fragments ($n_{\text{Livestock}} = 20$ and $n_{\text{Undisturbed}} = 22$).
302 Interaction between livestock and wildlife is complicated. For instance, while a number of
303 studies found evidence of competitive exclusion between livestock and large herbivore⁸⁰,

304 may other recorded resource sharing between these two groups^{81,82}. Yet still, may other
305 studies did not find any relationship between the two⁸³. The outcome of the interaction
306 may depend on the ecological similarity between the two groups (Niche overlap),
307 availability of natural resources that may vary between habitats (between low to high
308 productivity) and also degree of behavioural habituation by the wildlife. The wildlife
309 community that we studied was an ecologically broad one consisting of wildlife with very
310 different ecology. Therefore, while some of the species—such as spotted deer and sambar
311 deer, who were found only in the undisturbed fragments—may face resource competition
312 from livestock grazing, others (for example, small carnivores and primates) may not face any
313 competition. In addition, many large herbivores, such as gaur and elephants, who despite
314 resource competition may still use the disturbed fragments as corridors contributing to host
315 richness. These processes together may explain almost similar host species richness
316 between fragments with and without livestock grazing.

317 We did not find any significant effect of human settlement on host diversity but the
318 trend is negative (Figure 3). While human settlement may attract and facilitate generalist
319 and weedy species with high tolerance for disturbance (for example, rodents, which were
320 not sampled in the present study), many elusive species such as carnivores may be
321 adversely affected and may prefer to avoid fragments with settlements⁸⁴. Still, we recorded
322 overall a large host species richness ($\text{host richness}_{\text{Settlement}} = 19$, $\text{host richness}_{\text{Undisturbed}} = 23$)
323 from around the settlement in Anamalai hills. This could be explained by the facts that many
324 of these settlements may attract wildlife with unintentionally supplemented resources such
325 as planted fruit trees⁶⁰. Additionally, the high level of fragmentation of the landscape
326 meant large herbivores and carnivores may not have much choice but to disperse through
327 human settlements^{54,56}. We did not find evidence of habitat fragmentation (fragment size
328 and isolation) influencing host species richness in Anamalai hills (Figure 3). This is in line with
329 findings from across studies that effects of fragmentation on species communities are often
330 weak⁸⁵. Effects of habitat fragmentation on species diversity is highly context-specific and
331 varies considerably between animal groups, ecosystems and kinds of human activities
332 prevalent in the landscape^{85–89}. In Anamalai hills, habitat fragmentation is widespread,
333 which likely disrupt animal movement to some extent but, in the absence of hunting,
334 perhaps not substantially. For instance, studies recorded use of certain plantation as

335 corridors to connect with isolated undisturbed habitats^{51,52,74}. However, the adverse outcome
336 of these movement through human-dominated habitats is the increase in wildlife-human
337 conflict^{54,56}.

338 Among the different types of land use change in Anamalai hills, plantations had the
339 strongest positive effect on parasite diversity (Figure. 2). Increase in number of parasite taxa
340 in modified fragments ranged between one to ten, with eight parasite taxa that were
341 recorded exclusively in these fragments (Table 4). However, this increased richness in
342 disturbed fragments were likely not driven by host richness as host richness had a small and
343 statistically not significant effect on parasite richness in the disturbance model (Figure 2).
344 This is in contrast to the predominant patterns across most studies on parasite diversity that
345 found host richness to be the strongest predictor of parasite richness¹⁹⁻²². However, there
346 could be potential deviations from this rule, particularly due to human impacts^{21,90,91}. For
347 instance, many human parasites may spillover to wildlife (anthropozoonoses) as humans
348 regularly come in contact with wildlife⁹²⁻⁹⁷. Humans may also introduce many non-wildlife
349 species such as feral dogs, cats in addition to livestock into wildlife habitats and these
350 species may share parasites with wildlife²⁶. In such cases, parasite richness in wildlife would
351 be more than in the undisturbed fragments. Indeed, all but one (*Schistosoma sp.*) of the
352 parasites that we found exclusively in plantations also occurred in cattle (Table 4).
353 Surprisingly, wildlife parasite taxa that were present in the livestock foraging fragments did
354 not occur in cattle samples from the same fragments. This was also the case for the wildlife
355 parasites that only occurred in settlement but not in undisturbed fragments. We did not find
356 any significant effect of host body mass on parasite diversity (Figure 2). This is in contrast
357 many studies that found a significant relationship between these two variables^{98,99}. On the
358 other hand, many other empirical studies that did not find any relationship between body
359 mass and parasite richness when accounting for host phylogenetic relationships^{100,101}. Such
360 contradictory results may suggest that relationship between host body mass and parasite
361 diversity is a factor of body mass and life history traits, which vary between ecologically
362 different groups of hosts²². Thus, the broad ecological diversity among host species in the
363 present study might have confounded this relationship.

364 Our results did not find any significant effect of habitat fragmentation on parasite
365 richness (Figure 2). This was expected as we did not find any effect of fragmentation on host

366 richness either. This lack of relationship between fragmentation and parasite diversity could
367 also be an outcome of large home ranges and low habitat specialisation of most of the host
368 species in our study. Many of the species that we sampled were large herbivores or
369 carnivores (e.g., *Elephas maximus*, *Bos gaurus*, *Panthera tigris*, *Panthera pardus*) with large
370 home range and they disperse across fragments. The level of fragment isolation (Median
371 distance = 30.2 km) may not be a deterrent to their dispersal. Similarly, many host species in
372 the study community such as *Macaca radiata*, *Sus scrofa*, *Viverricula indica*, are habitat
373 generalists. According to the distribution-abundance relationship hypothesis⁶, smaller,
374 fragmented habitats may be conducive for these generalist wildlife with high reproductive
375 rates. These hosts may then spread parasites across habitats, independent of the level of
376 habitat fragmentation.

377

378 **CONCLUSION:**

379 With data on 40 parasites from 23 host species collected from 19 rainforest fragments with
380 different types of land use change, we demonstrated that land use changes increased
381 parasite diversity and presence of potential spillover of parasites from livestock to wildlife.
382 We also showed that the observed pattern of parasite diversity was not driven by habitat
383 fragmentation.

384 One of the limitations of this study was that it could not test the effect of land use change
385 and habitat fragmentation on the relationship between host density and parasite diversity.
386 Host density is an important predictor of parasite diversity and in nature, host density is
387 linked to host ecology (e.g., home range). However, land use change can unpredictably
388 change host density, which may have a complex outcome for parasite diversity. It will thus
389 be worthwhile in future to explore this question in the present system. Additionally, with
390 the present evidence of potential anthroponosis, it will be important in future to compare
391 parasite from the present study to samples from humans, livestock and commensal animals
392 in the fragments. Finally, as far as land use change and habitat fragmentation of wildlife
393 habitats in India are concerned, the present study represents a case study with particular
394 relevance for tropical rainforest habitats. However, there exists a large diversity in habitats
395 and levels of disturbance in India. Given the increased threat to wildlife health from

396 anthropogenic environmental change, it will thus be crucial for wildlife conservation to
397 study the patterns of parasite diversity in other types of habitats, especially those with
398 already threatened wildlife.

399 **Data availability:** The datasets generated and analysed during the current study are
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407 **Author Contributions:** Sampling and data collection were done by DMR, ST and DC. DC
408 conducted data analysis and wrote manuscript. GU conceived, designed and lead the study,
409 organised funding and reviewed the manuscript.

410 **Competing interests:** The authors declare no competing interests.

411 **References:**

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657 **Table 1.** Bootstrap estimate of host richness in each fragment of Anamalai hills, India

<i>Fragment name</i>	<i>Bootstrap estimate of host species richness</i>	<i>SE</i>	<i>Sample size</i>
Aliyar_dam	21.9	0.8	210
Akkamalai	18.9	0.7	199
Anaikundi	20.1	0.8	150
Andiparai	22.6	1	256
Attakatty	11.7	0.6	15
Iyerpadi	25.5	1	398
Karian_shola	25.3	0.9	244
Korangumudi	24.9	0.8	356
Monica_estate	20.4	0.9	124
Monomboly	26.6	1	181
Nirar_dam	24.2	0.9	167
Pannimedu	17.5	1	55
Puthuthottam	24.9	0.8	426
Sethumadai	18.1	0.9	65
Shekkalmudi	16.0	0.8	42
Sirukundra	17.9	0.8	127
Uralikal	11.5	0.5	41
Varagaliyar	21.6	1	162
Varattuparai	23.4	0.9	397

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661 **Table 2.** Comparison between two different models to explain bootstrap estimate of
 662 parasite taxon richness in Anamalai hills, India

<i>Models</i>	<i>K</i>	<i>logLik</i>	<i>AIC</i>	<i>delta</i>	<i>weight</i>
<i>Plantation + Settlement + Livestock + Host richness +Host body size</i>	9	-667.54	1353.07	0	0.887
<i>Fragment size + Isolation index + Host richness +Host body size</i>	8	-670.59	1357.19	4.112	0.113

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669 **Table 3.** Comparison between two different models to explain bootstrap estimate of host
 670 species richness in Anamalai hills, India

<i>Models</i>	<i>K</i>	<i>logLik</i>	<i>AIC</i>	<i>delta</i>	<i>weight</i>
<i>Plantation + Settlement + Livestock</i>	5	-46.73	103.47	0	0.971
<i>Fragment size + Isolation index</i>	4	-51.25	110.5	7.029	0.029

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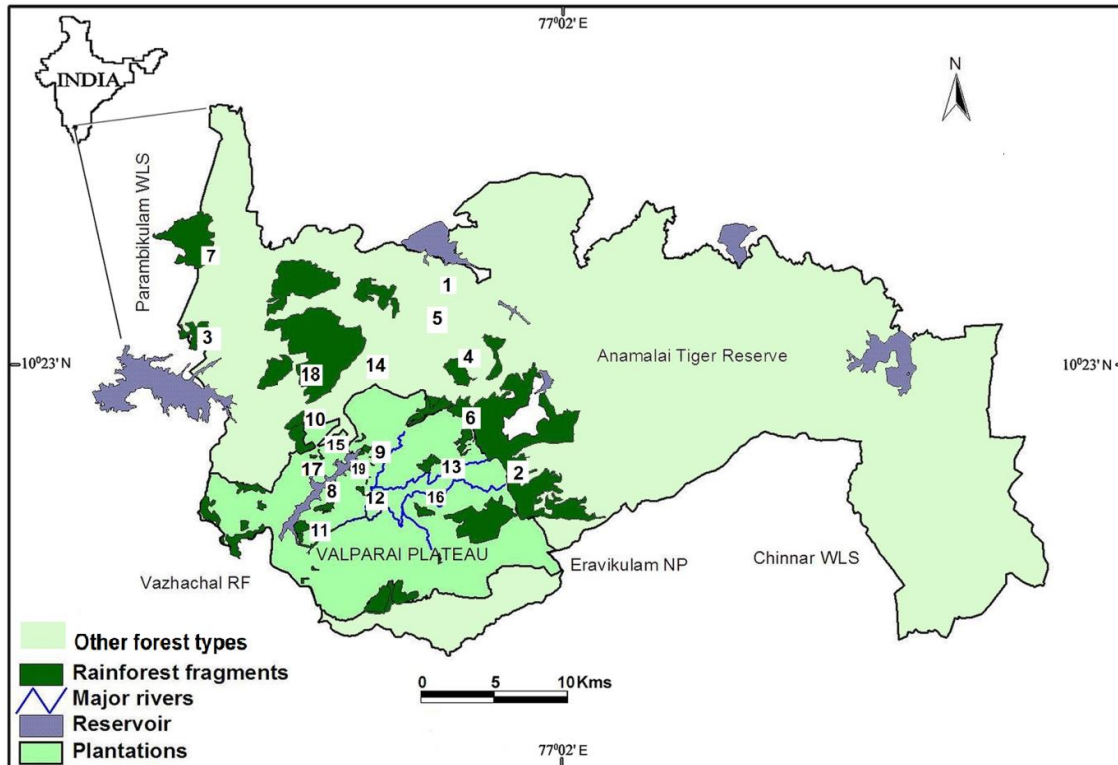
674 **Table 4.** Parasite taxa that were found only in disturbed or undisturbed fragments in
 675 Anamalai hills, India. Highlighted parasite taxa were found in the corresponding fragment
 676 group exclusively. 1. Current study; 2. Natural History Museum parasite database, London,
 677 UK

Parasite taxa	Parasite group	Family	In livestock samples ¹	Known human case ²
Plantation only				
Baylisascaris sp.	Nematodes	Ascaridoidea	Present	present
Nematodirus sp.	Nematodes	Trichostrongyloidea	Present	present
Enterobius sp.	Nematodes	Oxyuroidea	Present	present
Dictyocaulus sp.	Nematodes	Trichostrongyloidea	Absent	absent
Uncinaria sp.	Nematodes	Ancylostomatoidea	Absent	present
Schistosoma sp.	Trematodes	Schistosomatidae	Absent	present
Metastrongylus sp.	Nematodes	Metastrongyloidea	Absent	present
Clonorchis sp.	Trematodes	Opisthorchiidae	Present	present
Toxoplasma sp.	Apicomplexa		Present	present
Isospora sp.	Apicomplexa		Present	present
Paragonimus sp.	Trematodes	Paragonimidae	Present	present
Dipylidium sp.	Cestodes	Dilepididae	Present	present
Livestock presence only				
Dictyocaulus sp.	Nematodes	Trichostrongyloidea	Absent	absent
Taenia sp.	Cestodes	Taeniidae	Absent	present
Metastrongylus sp.	Nematodes	Metastrongyloidea	Absent	present
Undisturbed (Livestock) only				
Paragonimus	Trematodes	Paragonimidae	Present	present
Settlement only				
Dictyocaulus sp.	Nematodes	Trichostrongyloidea	Absent	absent
Uncinaria sp.	Nematodes	Ancylostomatoidea	Absent	present
Metastrongylus sp.	Nematodes	Metastrongyloidea	Absent	present
Undisturbed (Settlement) only				
Sarcocystis sp.	Apicomplexa		Present	present

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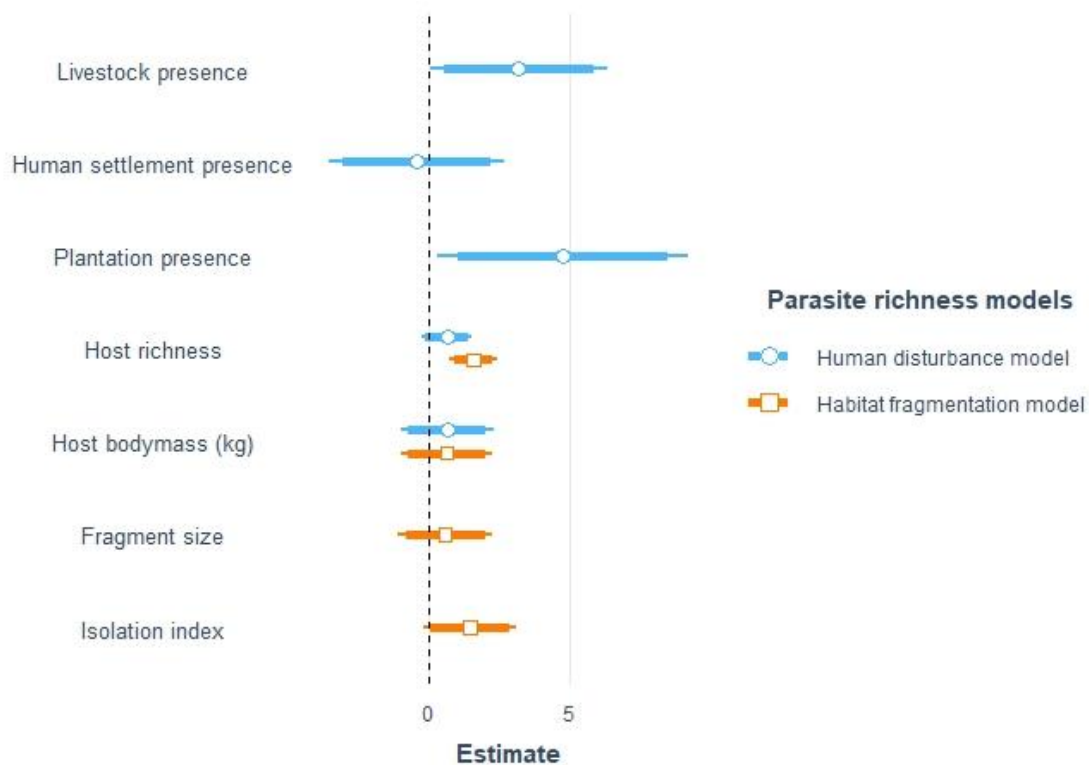


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682 Figure 1. Map of Anamalai hills, Western Ghats, India with numbered study fragments. (1)
683 Aliyar dam, 2) Akkamalai, 3) Anaikundi, 4) Andiparai, 5) Attakatty, 6) Iyerpadi 7)
684 Karian_shola, 8) Korangumudi, 9) Monica_estate, 10) Monomboly, 11) Nirar_dam, 12)
685 Pannimedu, 13) Puthuthottam, 14) Sethumadai 15) Shekkalmudi 16) Sirukundra 17) Uralikal,
686 18) Varagaliyar and 19) Varattuparai WLS: Wildlife sanctuary; RF: Reserve Forest; NP:
687 National Park

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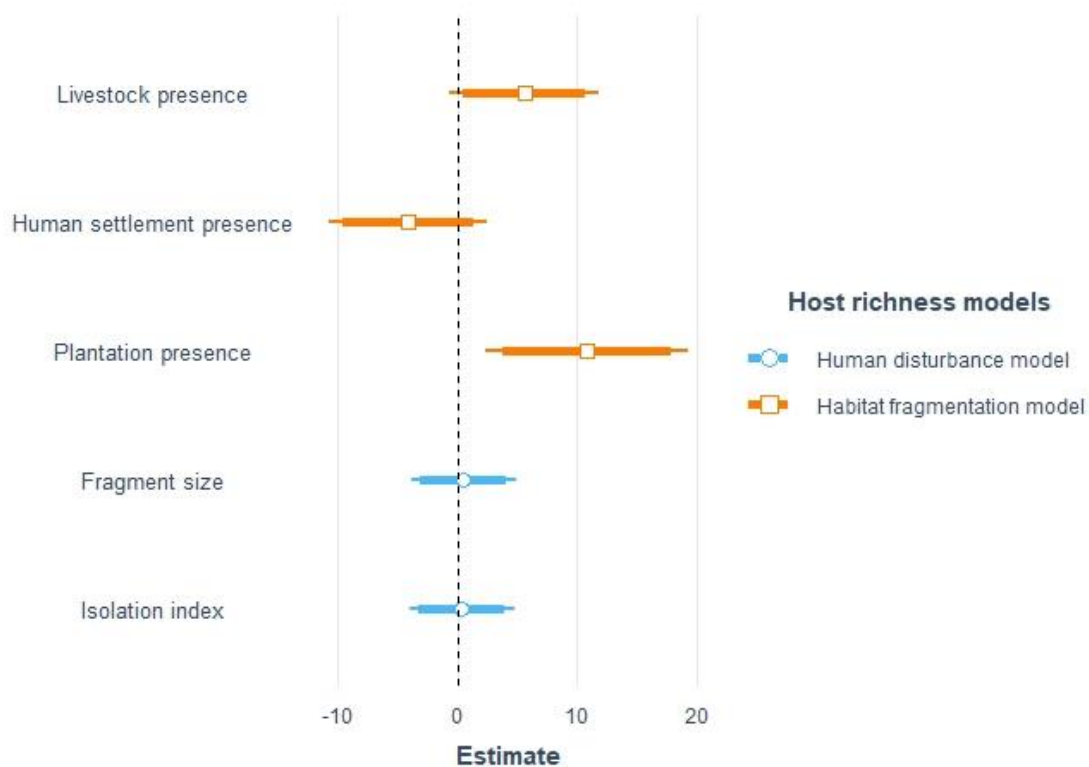


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691 **Figure 2.** Unstandardized effect size of predictor variables on bootstrap estimate of parasite
692 taxon richness in rainforest fragments of Anamalai hills, India. Estimates were plotted to
693 scale. Intercepts were omitted to avoid distortion of scale. Disturbance model was the best
694 fitted model based on AIC. Confidence intervals are represented by the lines around the
695 points— thick ($\alpha = 0.10$) and thin ($\alpha = 0.05$).

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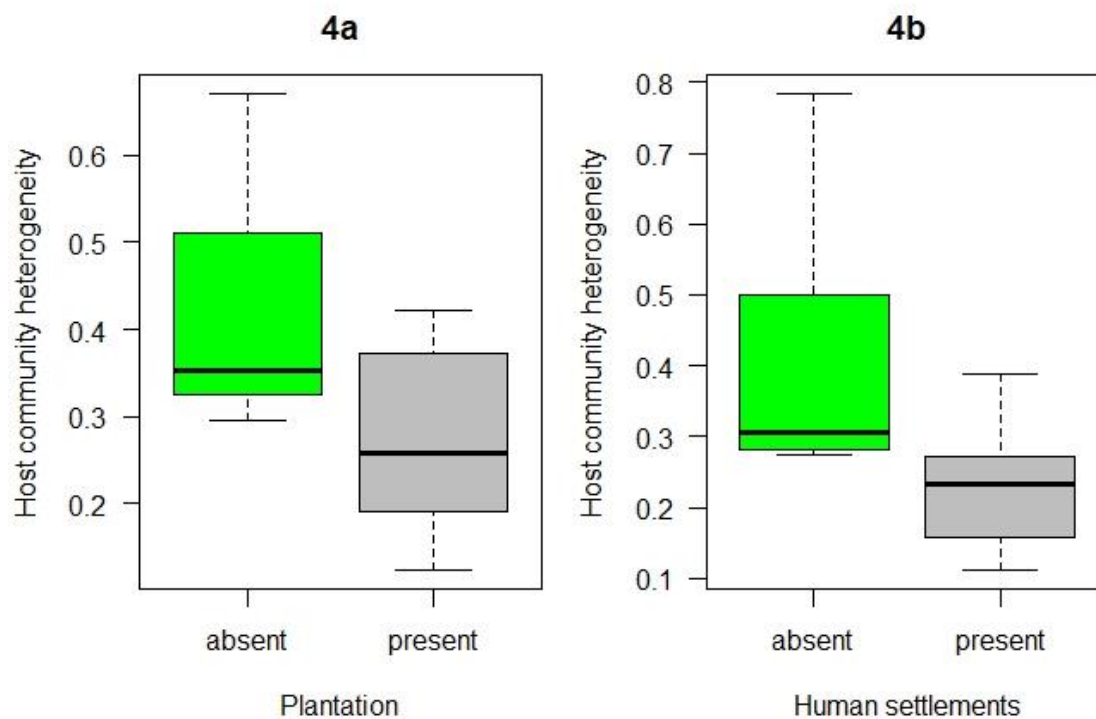
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698

699 **Figure 3.** Unstandardized effect size of predictor variables on bootstrap estimate of host
700 species richness in rainforest fragments of Anamalai hills, India. Estimates were plotted to
701 scale. Intercepts were omitted to avoid distortion of scale. Disturbance model was the best
702 fitted model based on AIC. Confidence intervals are represented by the lines around the
703 points— thick ($\alpha = 0.10$) and thin ($\alpha = 0.05$). Host sample sizes are given in Table 1.

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705

706 **Figure 4.** Host community heterogeneity between undisturbed (absent) and disturbed
707 (present) rainforest fragments of Anamalai hills, India. Community heterogeneity is the
708 within group dispersion values based on Jaccard distance for presence/absence data.

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