

Biological Aging in Childhood and Adolescence Following Experiences of Threat and Deprivation: A Systematic Review and Meta-Analysis

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Abstract

Life history theory argues that exposure to early-life adversity (ELA) accelerates development, although existing evidence for this varies. We present a meta-analysis and systematic review testing the hypothesis that ELA involving threat (e.g., violence exposure) will be associated with accelerated biological aging across multiple metrics, whereas exposure to deprivation (e.g., neglect, institutional rearing) and low-socioeconomic status (SES) will not. We meta-analyze 46 studies (n=64,925) examining associations of ELA with pubertal timing and cellular aging (telomere length and DNA methylation age), systematically review 19 studies (n=2276) examining ELA and neural markers of accelerated development (cortical thickness and amygdala-prefrontal cortex functional connectivity) and evaluate whether associations of ELA with biological aging vary according to the nature of adversity experienced. ELA overall was associated with accelerated pubertal timing (d=-0.12) and cellular aging (d=-0.32). Moderator analysis revealed that ELA characterized by threat (d=-0.26), but not deprivation or SES, was associated with accelerated pubertal development. Similarly, exposure to threat-related ELA was associated with accelerated cellular aging (d=-0.43), but not deprivation or SES. Systematic review revealed associations between ELA and accelerated cortical thinning, with threat-related ELA consistently associated with thinning in ventromedial prefrontal cortex, and deprivation and SES associated with thinning in frontoparietal, default, and visual networks. There was no consistent association of ELA with amygdala-PFC connectivity. These findings suggest specificity in the types of early environmental experiences associated with accelerated biological aging and highlight the importance of evaluating how accelerated aging contributes to health disparities and whether this process can be mitigated through early intervention.

Keywords: Early-Life Adversity; Threat; Deprivation; Pubertal Timing; Telomere Length; Methylation Age, Cortical Thickness; Amygdala-PFC

Introduction

Exposure to early-life adversity (ELA)—including exposure to child abuse, sexual assault, neglect, and chronic poverty—is associated with elevated risk for numerous mental and physical health problems, including depression, anxiety disorders, substance abuse, suicide, and cardiovascular disease (Felitti et al., 1998; Green et al., 2010; Heim & Binder, 2012; Kessler et al., 2010; McLaughlin et al., 2010, 2012; Norman et al., 2012; Scott et al., 2011). The associations of ELA with mental and physical health problems are observable beginning in childhood and adolescence (Boynton-Jarrett, Ryan, Berkman, & Wright, 2008; Halpern et al., 2013; McLaughlin et al., 2012; McLaughlin, Basu, et al., 2016) and persist into adulthood (Dong et al., 2004; Felitti et al., 1998; Green et al., 2010; Kessler et al., 2010). Recent evidence from longitudinal and population-based studies indicates that exposure to ELA is even associated with elevated risk for premature mortality (Brown et al., 2009; E. Chen, Turiano, Mroczek, & Miller, 2016).

Accelerated Development / Biological Aging

One potential mechanism linking exposure to ELA with this wide range of physical and mental health problems is accelerated biological aging. Specifically, exposure to adversity early in life may alter the pace of development, resulting in faster aging. Most conceptual models on the link between ELA and accelerated development are based in Life History Theory (J. Belsky, Steinberg, & Draper, 1991; Ellis, Figueredo, Brumbach, & Schlomer, 2009; Ellis & Garber, 2000), which postulates that experiences in early-life can program an individual's developmental trajectory in order to respond most effectively to the environmental demands they are likely to encounter later in life. The pattern and timing of life history events—such as age of sexual maturation, gestational period, number of offspring, birth spacing, length of parental investment, longevity, and others—is determined by the relative prioritization of time and energy invested in growth, reproduction, and longevity (Del Giudice, Gangestad, & Kaplan, 2016; Hill & Kaplan, 1999). For instance, in a safe, predictable and enriched environment, a slow and protracted

development may be optimal, as it allows for maximal parental investment prior to offspring independence. However, in a harsh or unpredictable environment, a faster pace of development in which individuals reach adult-like capabilities at an earlier age may be favored in order to maximize reproduction prior to potential mortality. Life history theories of human development argue that early environments characterized by harshness (e.g. trauma, violence exposure) may accelerate the onset of puberty in order to maximize the opportunity for reproduction prior to mortality (J. Belsky, 2012; Ellis et al., 2009; Rickard, Frankenhuis, & Nettle, 2014). However, in unpredictable environments, where there is large variation in harshness, it may be optimal to delay reproductive milestones, depending upon various features of the environment including population density and resource availability (J. Belsky, 2012; Ellis et al., 2009).

More recently, life history theories regarding the pace of development following ELA have been extended to focus on additional measures of biological aging. First, predictive adaptive response models (Nettle, Frankenhuis, & Rickard, 2013; Rickard et al., 2014) focus on cellular and molecular development and how it relates to an individual's morbidity and mortality across the lifespan. These models propose that ELA negatively influences physical health, through altered cellular development as a result of reduced energy to build or repair cellular tissue. This advanced cellular aging forecast may reduce longevity and contribute to acceleration in reproductive maturity (Nettle et al., 2013; Rickard et al., 2014). Second, the stress acceleration hypothesis (Callaghan & Tottenham, 2016) suggests that ELA accelerates development of neural circuitry underlying emotional processing, specifically, development of the amygdala-prefrontal cortex (PFC) circuit thought to underlie emotion regulation capabilities. This accelerated development in the context of unreliable or absent caregiving may occur in order to allow for independent emotion regulation at an earlier age (Callaghan & Tottenham, 2016). Each of these theories rest on the assumption that ELA impacts the pace of development across multiple domains and metrics of biological aging.

ELA and Biological Aging

Biological aging following ELA has been measured with a variety of different metrics. By far the most commonly used metric is the timing and pace of pubertal development, including age of menarche in females (Boynton-Jarrett & Harville, 2012; Deardorff, Abrams, Ekwaru, & Rehkopf, 2014; Graber, Brooks-Gunn, & Warren, 1995b) and pubertal stage controlling for chronological age (Colich et al., 2019; Mendle, Leve, Van Ryzin, Natsuaki, & Ge, 2011; Negri, Blankson, & Trickett, 2015; Noll et al., 2017; Sumner, Colich, Uddin, Armstrong, & McLaughlin, 2019). A second line of work has examined measures of cellular aging, including leukocyte telomere length (Coimbra, Carvalho, Moretti, Mello, & Belangero, 2017; Drury et al., 2014; Price, Kao, Burgers, Carpenter, & Tyrka, 2013) and DNA methylation (DNAm) age (Gassen, Chrousos, Binder, & Zannas, 2017; Wolf et al., 2017). A separate literature has examined markers of neural maturation such as amygdala-PFC connectivity (Callaghan & Tottenham, 2016; Gee, Gabard-Durnam, et al., 2013) and cortical thickness (McLaughlin, Sheridan, Winter, et al., 2014).

Evidence for accelerated biological aging following ELA has been found across all of these metrics. For example, numerous studies have found that ELA is associated with earlier pubertal timing (Graber, Brooks-Gunn, & Warren, 1995a; Hartman, Li, Nettle, & Belsky, 2017; Mendle et al., 2011; Negri, Blankson, et al., 2015). Similarly, a small but increasing number of studies have reported accelerated cellular aging following ELA, including shorter telomere length (Drury et al., 2012, 2014; Mitchell et al., 2014; Shalev et al., 2013), and advanced DNAm age relative to chronological age (Jovanovic et al., 2017a; Sumner et al., 2019). Finally, much of the evidence for accelerated neural development following ELA comes from studies examining amygdala-PFC functional connectivity (Colich et al., 2017; Gee, Gabard-Durnam, et al., 2013; Keding & Herringa, 2016) and cortical thinning across development (McLaughlin, Sheridan, Winter, et al., 2014). However, other studies have found no associations between ELA and pubertal timing (Negri, Saxbe, & Trickett, 2015; Negri & Trickett, 2012) or cortical thinning (McLaughlin, Sheridan, et al., 2016; Rosen, Sheridan, Sambrook, Meltzoff, & McLaughlin,

2018). Some studies have even found that ELA is associated with slower or delayed pubertal timing (Johnson et al., 2018; Negriff, Blankson, et al., 2015; Sumner et al., 2019) and a more immature pattern of amygdala-PFC connectivity (Cisler, James, et al., 2013; Marusak, Martin, Etkin, & Thomason, 2015; Silvers, Lumian, et al., 2016). The strength and direction of the association between ELA and markers of biological aging varies widely across studies, and to date no systematic review or meta-analysis on this topic has been conducted.

We argue, and test through meta-analysis and systematic review, that the wide variability in the association of ELA with accelerated development might be explained—at least in part—by differences in how distinct types of ELA influence the pace of development. Existing studies have focused on a wide range of adversity experiences, ranging from physical abuse and violence exposure to physical and emotional neglect and institutional rearing, and provide some clues about the types of ELA that might be particularly likely to produce a pattern of accelerated development. For example, physical and sexual abuse have been consistently associated with accelerated pubertal development in females (Mendle, Ryan, & McKone, 2016; Natsuaki, Leve, & Mendle, 2011; Noll et al., 2017; Trickett, Noll, & Putnam, 2011; Trickett & Putnam, 1993). In contrast, studies of war and famine suggest that severe material deprivation can delay pubertal development (Prebeg & Bralic, 2000; van Noord & Kaaks, 1991). Although less work has examined the effects of neglect and psychosocial deprivation on biological aging, existing studies typically find no association of neglect or early institutional rearing with pubertal timing (Johnson et al., 2018; Mendle et al., 2011; Reid et al., 2017; Ryan, Mendle, & Markowitz, 2015). In contrast, early institutionalization is associated with an accelerated pattern of cellular aging (Drury et al., 2012) and maturation of the amygdala-PFC circuit (Gee, Gabard-Durnam, et al., 2013), suggesting that accelerated biological aging might not occur in a uniform manner across various neurobiological systems.

Discrepancies in these findings may be due to the treatment of ELA as a monolithic construct with equifinality across all metrics of biological aging. To date, no attempt has been

made to consider how associations of ELA with accelerated biological aging might vary according to the nature of the adversity experienced. Systematic investigation into variability in the association of ELA with biological aging across adversity types may help to reconcile inconsistent findings and advance theoretical models of how early experiences alter the pace of development at reproductive, cellular, and neural levels of analysis. This meta-analysis aims to do so by: 1) examining how *differing dimensions* of ELA influence biological aging, distinguishing between experiences characterized by threat versus deprivation; and 2) evaluating whether the associations of these different types of adverse early experience with biological aging are global or specific to particular domains of aging—including pubertal timing, cellular aging and brain development.

Conceptual Model of Early-Life Adversity and Accelerated Development

Many prior studies examining the effects of ELA on accelerated biological aging have focused on a limited range of ELA experiences, typically focusing on relative extreme exposures like sexual abuse or institutional rearing. Other studies have utilized a cumulative-risk approach, which tallies the number of distinct forms of ELA experienced to create a risk score without regard to the type, chronicity, or severity of the experience and uses this risk score as a predictor of outcomes, with the assumption that all forms of ELA have equal and additive effects on developmental outcomes (Evans, Li, & Whipple, 2013). Very few studies attempt to address the high co-occurrence of varying forms of ELA (Green et al., 2010; McLaughlin et al., 2012) or examine the differential influences of particular adversity types on biological aging, with some notable exceptions (Mendle et al., 2011, 2016; Negri, Saxbe, et al., 2015; Sumner et al., 2019).

The dimensional model of adversity and psychopathology (DMAP) argues that the wide range of experiences currently classified as ELA can be organized into core underlying dimensions that have unique influences on cognitive, emotional, and neural development (McLaughlin, Sheridan, & Lambert, 2014; McLaughlin & Sheridan, 2016; Sheridan & McLaughlin, 2014). This model attempts to distill complex adverse experiences into core

underlying dimensions that cut across multiple forms of ELA that share common features. Two such dimensions are *threat*, which encompasses experiences involving harm or threat of harm to the child, and *deprivation*, which involves an absence of expected inputs from the environment during development, such as cognitive and social stimulation (e.g., complex language directed at the child) as well as emotional nurturance (e.g., emotional neglect). In addition, the DMAP model argues that these dimensions of adversity have influences on emotional, cognitive, and neural development that are at least partially distinct. Increasing evidence has demonstrated the unique developmental consequences of threat and deprivation on developmental outcomes (Busso, McLaughlin, & Sheridan, 2016; Dennison et al., 2019; Everaerd et al., 2016; Lambert, King, Monahan, & McLaughlin, 2017; Rosen et al., 2018; Sheridan, Peverill, Finn, & McLaughlin, 2017). Determining whether all forms of ELA are associated with accelerated development across multiple metrics of biological aging or whether only particular dimensions of ELA are associated with this pattern is critical for identifying the mechanisms linking ELA to health outcomes and to better inform early interventions.

The threat dimension of ELA is conceptually similar to the life history theory dimension of environmental harshness, and involves experiences of trauma and violence exposure. We expect that experiences characterized by threat will be associated with accelerated biological aging, potentially in order to maximize the opportunity for reproduction prior to mortality (J. Belsky, Schlomer, & Ellis, 2012; Ellis et al., 2009). However, it is unclear how experiences of deprivation align with life history theory; whereas nutritional deprivation and food insecurity are thought to delay pubertal timing to ensure maximal bioenergetic resources should reproduction occur (Rogol, Clark, & Roemmich, 2000), specific predictions about physical and emotional neglect are lacking in life history models. Preliminary evidence suggests that accelerated development following ELA may vary across different dimensions of adversity. For instance, we have found that experiences characterized by threat, but not deprivation, were associated with accelerated pubertal stage relative to chronological age and accelerated DNAm age in a

community-based sample of children and adolescents (Sumner et al., 2019). In contrast, experiences characterized by deprivation were associated with *delayed* pubertal timing, after controlling for co-occurring threat experiences. We recently replicated this work in a nationally-representative sample of adolescent females, using age of menarche as our metric of accelerated aging (Colich et al., 2019). Determining whether accelerated biological aging is associated with exposure to ELA generally or with particular dimensions of ELA may help to elucidate the specific psychological and biological mechanisms underlying these associations.

Metrics of Biological Aging

Accelerated biological aging has been conceptualized in many ways, across multiple domains of biological development. Historically, these domains have been examined in isolation, independent of other domains of biological aging. Only two studies to our knowledge have explored the effects of ELA on multiple domains of accelerated development in adolescence (J. Belsky & Shalev, 2016; Sumner et al., 2019) and recent work suggests that accelerated telomere erosion and accelerated pubertal development represent similar biological processes as a consequence of ELA (Shalev & Belsky, 2016).

Pubertal timing. The most consistently examined marker of accelerated development in relation to ELA is pubertal timing, typically operationalized as the age of onset of pubertal development, or the age of achieving a reproductive milestone such as menarche. Puberty begins as early as ages 8-14 in females and 9-15 in males with the activation of the hypothalamic-pituitary-gonadal (HPG) axis. This ultimately initiates the start of gonadarche, in which the gonads mature and produce gonadal hormones or sex steroids. This in turn, leads to breast development and eventually menarche in girls, and in increased testicle size and the onset of spermarche in males. Typical measures of pubertal development use secondary sex characteristics as a metric of pubertal stage (Carskadon & Acebo, 1993; Marshall & Tanner, 1969, 1970). For the purposes of this meta-analysis, we have included studies that explore how ELA is associated with three commonly used metrics of pubertal timing—pubertal stage relative

to chronological age, age at the achievement of the onset of secondary sex characteristics, and age of menarche. Although menarche occurs relatively late in the pubertal process, participants are relatively reliable in their reporting of this milestone, particularly in adolescence (Dorn, Sontag-Padilla, Pabst, Tissot, & Susman, 2013).

Cellular aging. Predictive adaptive response models of accelerated aging following adversity postulate that the early environment influences an individual's somatic state, which in turn influences reproductive timing and other life history events (Nettle et al., 2013; Rickard et al., 2014). One pathway linking ELA to somatic states is cellular aging. Some argue that if the body detects a shortened cellular lifespan, mechanisms may exist to accelerate the development of the reproductive system in order to maximize the chances of reproduction prior to mortality (Nettle et al., 2013; Rickard et al., 2014). Cellular aging in the context of ELA has been measured in two different ways – telomere length and metrics of epigenetic aging using DNAm patterns.

Telomeres are nucleoprotein complexes that sit at the end of chromosomes and protect the chromosome from degradation (Chan & Blackburn, 2004). Telomeres shorten due to both cell replication and exposure to oxidative stress and inflammation. In normal aging, telomeres shorten in all cell types, which allows for the use of telomere length as a biological marker of cellular age (Frenck, Blackburn, & Shannon, 1998). Chronic stress has been shown to accelerate the shortening of telomeres in adults (Epel et al., 2004), and several studies have demonstrated associations between ELA and telomere length in children (Coimbra et al., 2017; Essex et al., 2013; Price et al., 2013). Shortened telomere length has been implicated in the pathogenesis of both physical and mental health problems in adulthood (Gotlib et al., 2015; Hoen et al., 2013; Needham, Mezuk, et al., 2015; Tyrka et al., 2016), suggesting a potential mechanism linking ELA and maladaptive health outcomes in adolescence and adulthood.

A second recently established metric of cellular aging is an epigenetic clock that considers genome-wide DNAm patterns (both increased and decreased methylation of select

CpG sites) to quantify biological age independent from chronological age (DNAm age) (Hannum et al., 2013; Horvath, 2013). This metric correlates strongly with chronological age in both adolescents and adults (Horvath & Raj, 2018; Suarez et al., 2018) and shows strong positive associations with age of death (B. H. Chen et al., 2016; Marioni, Shah, McRae, Chen, et al., 2015), suggesting it is a valid metric of cellular aging. Deviations between DNAm age and chronological age have been used as a metric of accelerated development (Davis et al., 2017; Jovanovic et al., 2017b; Sumner et al., 2019) and are associated with exposure to ELA (Jovanovic et al., 2017b; Sumner et al., 2019). Advanced DNAm age has been associated with increased risk of cardiovascular disease, cancer, and obesity (Horvath et al., 2014; Marioni, Shah, McRae, Ritchie, et al., 2015; Perna et al., 2016), again potentially highlighting a mechanism linking ELA and physical health problems.

For the purpose of this meta-analysis, we have included studies that explore how ELA impacts cellular aging, as measured by telomere length and DNAm age.

Brain Development. Numerous studies have investigated the neural consequences of ELA. Here, we focus specifically on neural markers of maturation. As such, we focus on two metrics for which patterns of development have been well characterized: cortical thickness and functional connectivity between the amygdala and prefrontal cortex (PFC). We focus on cortical thickness as a metric of structural development because the pattern of development is well characterized, replicated across many studies, and shows a clear linear association with age, such that cortical thickness steadily decreases from middle childhood to early adulthood (Ducharme et al., 2016; LeWinn, Sheridan, Keyes, Hamilton, & McLaughlin, 2017; Vijayakumar et al., 2016; Wierenga, Langen, Oranje, & Durston, 2014). Second, we focus on functional connectivity between the amygdala and PFC as a metric of maturation because it serves as a key component in the stress acceleration hypothesis, which posits that the amygdala-PFC circuit supporting emotional processing and regulation matures more rapidly among children exposed to ELA (Callaghan & Tottenham, 2016).

Cortical thickness declines steadily from childhood to early adulthood (Ducharme et al., 2016; LeWinn et al., 2017; Wierenga et al., 2014), as a result of developmentally-appropriate pruning of synapses and increases in myelination of connections between neurons (Natu et al., 2018; Sowell et al., 2004). This linear pattern of development enables assessment of whether development is accelerated or delayed among children with ELA relative to their peers. Cortical structure can be measured in a variety of ways including surface area, thickness, and volume (for review, see Vijayakumar et al., 2016). However, cortical thickness is the only metric that has a linear developmental trajectory, declining steadily from early childhood through early adulthood (Ducharme et al., 2016; LeWinn et al., 2017; Walhovd, Fjell, Giedd, Dale, & Brown, 2017; Wierenga et al., 2014). In contrast, cortical surface area and volume exhibit non-linear associations with age and the inflection points of these trajectories vary across samples and remain a source of debate (Ducharme et al., 2016; Giedd et al., 1999; Lenroot et al., 2007; LeWinn et al., 2017; Mills et al., 2016; Vijayakumar et al., 2016). These non-linear patterns of development make assessing deviations from the expected pattern more difficult. Therefore, we focus only on studies that use cortical thickness—including both whole cortex and specific regions—as an outcome.

The stress acceleration hypothesis focuses on the impact of ELA on the developmental trajectory of neural circuits supporting emotion processing and regulation, particularly on connectivity between the amygdala and medial PFC (mPFC; (Callaghan & Tottenham, 2016). Animal tracing studies demonstrate that feedforward connections between amygdala and PFC exist early in life, but feedback connections emerge later in development (Barbas & García-Cabezas, 2016). It has been proposed that in humans, changes in functional connectivity between the mPFC and amygdala may reflect the maturation of these feedback connections (Gee, Humphreys, et al., 2013). The pattern of functional connectivity between the amygdala and the mPFC shifts from positive to negative across development in the context of emotional processing tasks (Gee, Humphreys, et al., 2013; Silvers, Lumian, et al., 2016; Wu et al., 2016).

For the purpose of this systematic review, we have included studies that explore how ELA impacts both cortical thickness and amygdala-mPFC functional connectivity.

The Current Study

We aimed to test the hypothesis that experiences characterized by threat, but not deprivation, would accelerate biological aging. Applying this theoretical framework may help to reconcile discrepant findings in the literature by evaluating how different dimensions of ELA influence biological aging. In addition, we aimed to integrate disparate literatures by examining whether different dimension of adversity have general or specific effects on multiple domains of biological aging—including pubertal timing, cellular aging and brain development. We expected that threat and deprivation will have differing effects on biological aging, with threat associated with accelerated biological aging across all metrics and deprivation associated with delayed pubertal development. We did not have specific hypotheses about how deprivation would influence cellular aging or brain development. We also separately examined the associations of socioeconomic status (SES) with biological aging, as SES is a commonly used global measure of early experience that is associated with increased risk of exposure to both threat and deprivation (e.g., Green et al., 2010; McLaughlin et al., 2012). We had no a priori hypotheses about SES, given these associations with both threat and deprivation. A final guiding question was whether the associations of ELA characterized by threat and deprivation with biological aging would be consistent across all metrics. Whereas pubertal development reflects a more global measure of aging, cellular aging is a metric of biological aging most relevant to physical health, and cortical thickness and development in the amygdala-PFC circuit may reflect learning or adaptation to a stressful early environment, but not aging in a global way.

Methods

Information Sources and Search Strategy

This meta-analysis and systematic review was conducted in line with the PRISMA guidelines for meta-analyses (Moher, Liberati, Tetzlaff, & Altman, 2009; Figure 1). To identify

studies with relevant data, literature searches were conducted using internet databases (PubMed, SCOPUS, PsycINFO, Web of Science and Google Scholar) through January 2018. To ensure a thorough search, search terms encompassed various forms of ELA (e.g., violence, trauma, neglect, maltreatment, institutional rearing, deprivation, SES, poverty, early adversity, early life stress) as well as our dependent measures of interest (e.g., puberty, cell aging, methylation, menarche, telomere length, methylation, neural) and our targeted study population (e.g., infant, child, adolescent, pediatric) (see Supplemental Information for all search terms). All included studies were published in English and from peer-reviewed journals. To further identify eligible studies, we reviewed references of identified papers for additional studies using forward and backward searching.

Study inclusion criteria. In order to be included in the meta-analysis, studies had to meet the following criteria. First, studies had to examine an association between ELA and one of our dependent measures (pubertal timing, cellular aging, or brain development) and report sufficient statistics to calculate an effect size. Second, studies had to have been conducted in children and adolescent human samples (participants under age 18), rather than using retrospective reports of ELA and development in adults given the well documented recall biases associated with retrospective reporting of childhood experiences in adulthood (Hardt & Rutter, 2004; Widom, Raphael, & DuMont, 2004).

Inclusion criteria for ELA. We draw on a recent definition of ELA as experiences that were either chronic or severe in nature that require psychological or neurobiological adaptation by an average child and that represent a deviation from the expectable environment (McLaughlin, 2016). As detailed above, we used a wide range of search terms for ELA encompassing maltreatment experiences (e.g., physical, sexual, and emotional abuse; physical and emotional neglect), exposure to traumatic events (e.g., observing domestic violence, being the victim of interpersonal violence), institutional rearing, material deprivation (e.g., food insecurity), and childhood SES. We did not consider biological father absence as a form of ELA

given that: a) it is not clearly a form of ELA based on prevailing definitions (McLaughlin, 2016); and b) a meta-analysis on father absence and pubertal timing was recently conducted (Webster, Graber, Gesselman, Crosier, & Schember, 2014). We did not include other early experiences or more global stressful life events that did not clearly meet our definition of ELA (e.g., parental psychopathology, peer victimization).

Inclusion criteria for studies of pubertal timing. To retain as many studies as possible, we included studies that used self-report, parent-report and physician-rated Tanner/PDS stage (controlling for age) or age of menarche. Physician-rated Tanner stage and interview-based assessments of age of menarche in adolescence have been shown to be acceptably reliable (Coleman & Coleman, 2002; Dorn & Biro, 2011; Dorn et al., 2013). We examined whether the specific measure of pubertal timing was a moderator of ELA-puberty associations.

Inclusion criteria for studies of cellular aging. Although there have been prior reviews and meta-analyses exploring the effects of ELA on telomere length (Coimbra et al., 2017; Price et al., 2013) or DNAm age (Gershon & High, 2015; Lewis & Olive, 2014; Silberman, Acosta, & Zorrilla Zubilete, 2016; Vinkers et al., 2015; Wolf et al., 2018), none has focused on differences across distinct adversity types or restricted the focus to studies measuring ELA and cellular aging in childhood or adolescence. Telomere length and DNAm age can be assessed through both blood and saliva samples (Wren, Shirtcliff, & Drury, 2015); we have included both measures in our analyses.

Inclusion criteria for studies of brain development. We included only studies that assessed cortical thickness—including both whole cortex and specific regions—as an outcome and not other measures of cortical structure (e.g., volume and surface area) where age-related patterns are non-linear and thus more difficult to interpret with regard to acceleration of development. If ELA-exposed youth exhibit thinner cortex than non-exposed youths of the same age, this will be interpreted as accelerated maturation; if ELA-exposed youths exhibit thicker cortex than non-exposed youths of the same age, this will be interpreted as delayed

development. Similarly, we focus only on studies exploring task-related amygdala-mPFC functional connectivity, where a developmental shift from positive to negative in task-related amygdala-mPFC connectivity has been documented (Callaghan & Tottenham, 2016; Gee, Gabard-Durnam, et al., 2013). We will evaluate studies of ELA with this normative developmental pattern in mind; if children who have experienced adversity demonstrate greater negative connectivity for their age than children who have not, this would reflect accelerated development and if children who have experienced adversity exhibit more positive or less negative connectivity than comparison children, this would reflect delayed development. In contrast, studies investigating developmental patterns of connectivity at rest have been more mixed, with some studies demonstrating an increase in connectivity with age (e.g., Gabard-Durnam et al., 2014) and others demonstrating a decrease (Jalbrzikowski et al., 2017). Because a consensus has not been reached on the normative developmental pattern of amygdala-mPFC connectivity during rest, we focus only on papers that explore the associations of ELA with amygdala-mPFC connectivity using task-related functional connectivity.

We conducted a systematic review for metrics of brain development rather than a meta-analysis for the following reasons. First, whole-brain fMRI meta-analyses focus on the spatial nature of associations across the brain as opposed to the strength of effect sizes within a designated region. Given our focus on a specific measure of functional connectivity (amygdala-mPFC), spatial maps do not sufficiently address our research question regarding this metric of neural development. Second, the use of heterogeneous ROIs in studies of cortical thickness and amygdala-mPFC connectivity (e.g., different regions of mPFC), make it difficult to quantitatively compare results across studies. Third, meaningful differences in task design and task demands make it difficult to directly compare results of amygdala-mPFC connectivity using meta-analysis.

Measuring and Coding Adversity

In order to directly compare results from the present study with the majority of the existing literature, we first conducted an analysis in which we include studies defining ELA

broadly, regardless of adversity type. We then examined whether associations of ELA with biological aging metrics exhibited significant heterogeneity, and evaluated whether adversity type (i.e., threat, deprivation, SES) was a moderator of these associations (see Analysis Methods for details). Consistent with previous work from our group (Colich et al., 2019; Dennison et al., 2019; McLaughlin, Sheridan, Lambert, et al., 2014; Sheridan & McLaughlin, 2014), we conceptualized threat-related adversities to include experiences of physical abuse, domestic violence, sexual assault, witnessing or being the victim of violence in the community, and emotional abuse. Deprivation-related adversities included physical neglect, low cognitive stimulation, food insecurity, and early institutionalization/international adoption. We also examined the effects of SES, including family income and parental education. Although low SES is associated with reductions in cognitive stimulation among children (Bradley, Corwyn, Burchinal, McAdoo, & Garcia Coll, 2001; Duncan & Magnuson, 2012; Gilkerson et al., 2017), SES is a proxy for deprivation rather than a direct measure. This is especially true when studies examine the effects of SES without controlling for co-occurring experiences of threat or violence. To ensure that we had not diluted our deprivation composite by including SES as an indicator, we chose to examine studies using SES as a metric of ELA separately.

Study exclusion criteria. The literature search yielded a total of 7081 studies. Studies were first excluded based on their title or abstract (k=6515) with exclusion decisions made by one of the authors (NLC, EAW or MLR) and confirmed by another. Exclusion criteria included any publication that was not an analysis of primary data (i.e. a review, book chapter, etc.). We also excluded any studies conducted outside of the United States, Western Europe or Australia, given well-documented effects of ethnicity, nutritional status, and SES on timing of development (Parent et al., 2003) and difficulties assessing SES consistently across different countries. A subset of studies were examined more thoroughly for eligibility (k=519). After a careful review of the methods, studies were excluded if they didn't include a relevant independent or dependent variable (i.e. single gene methylation patterns rather than a measure of epigenetic aging or

resting state amygdala-PFC connectivity rather than task-based connectivity; k=136), if they were a review paper or book chapter (k=134), if they were conducted outside of the US/Europe/Australia (k=81), if the study was conducted in infants, adults or nonhuman animals (k=56), if we were unable to access the manuscript (n=24), if the data were from a conference abstract (k=8), published in a foreign language only (k=3), if the study was not sufficiently powered (i.e. less than or equal to 5 participants per group; k=2), or the study was later retracted (k=1). Given our focus on understanding deviations in developmental timing following ELA, studies were also excluded if the exposed and control group differed significantly in age (k=1; Humphreys et al., 2016). Finally, studies were excluded if they did not include data that we were able to convert into an effect size after multiple attempts to contact the study authors for original data; (k=9). Overall, the current study included a total of 64 studies: 37 studies contributing to our meta-analysis exploring the effects of ELA on pubertal timing, 9 studies contributing to our meta-analysis exploring the effects of ELA on cellular aging, and 19 studies contributing to our systematic review exploring the effects of ELA on brain development (with one study [Sumner et al., 2019] contributing to both pubertal timing and cellular aging analyses.

Management of Non-Independent Samples

In many cases, we extracted multiple effect sizes from the same sample. For example, some studies included multiple measures of pubertal timing (e.g. pubertal stage and age of menarche) or multiple measures of ELA (e.g. sexual abuse and physical abuse). Similarly, associations between ELA and developmental timing from a single study were sometimes examined across multiple publications using the same sample. To deal with this non-independence, we conducted multilevel mixed effects analyses with restricted maximum likelihood estimation, including study nested within sample as a random effect, such that multiple effect size estimates are nested within a higher-level grouping variable (e.g. study or sample). In the case of longitudinal data, we always included data from Wave I, as this wave tends to have the lowest attrition rate and in turn, the largest sample size (Borenstein, Hedges,

Higgins, & Rothstein, 2009). If a separate paper included data from a later wave, we included that data and did not report data from Wave I a second time (as these associations were included in the analysis from the Wave I paper; for example, (Mendle, Leve, Van Ryzin, & Natsuaki, 2014; Mendle et al., 2011).

Data Extraction

Three trained raters (NLC, MLR, EAW) coded individual studies. We screened each study and coded variables for study year, authors, participant composition/sample, mean age of participants, number of males and female participants, ethnicity, pubertal timing measure and informant, ELA measure and informant, as well as whether analyses controlled for other types of adversity, parent psychopathology, child psychopathology, father absence, mother's age at menarche, and BMI. All disagreements in coding were resolved via discussion amongst the three raters until consensus was achieved.

Data Analysis

To ensure consistency in the directionality of the effect sizes, in all cases, metrics of developmental timing were coded to indicate that numerically lower values (negative values) indicated accelerated development, to be consistent with age at menarche or age at pubertal attainment (the most commonly used metrics). Similarly, adversities were coded so that a numerically higher value indicates greater adversity. To be consistent with other variables, SES was coded to indicate that numerically higher SES values indicate lower SES.

For each study we calculated an effect size d and corresponding sampling variance (Cohen's d ; Cohen, 1988) for each relevant analysis. Following Cohen's suggestion, we interpret a $d=0.2$ to be a 'small' effect, $d=0.5$ to be a 'medium' effect and $d=0.8$ to be a 'large' effect. A positive d indicates that exposure to ELA is associated with delayed development (later age at a developmental milestone), whereas a negative d indicates that greater adversity is associated with accelerated development (earlier age at a developmental milestone). We derived d s from multiple reported statistics including: unadjusted or adjusted correlations

between two variables, odds ratios, mean differences and standard deviations, t statistics, F statistics and associated N s and p values, as well as unstandardized and standardized regression coefficients. All effect sizes were computed in R (version 3.4.1) using the `escalc` function in the “metafor” package (Viechtbauer, 2010) and converted to Cohen’s d using established formulas (Borenstein et al., 2009). Authors were contacted when published manuscripts met criteria for inclusion but did not include the necessary data to calculate an effect size amenable to our analyses, which occurred in thirteen cases (of which 4 provided necessary data and were included in our analyses).

All meta-analysis were conducted using three-level mixed-effects models and the `rma.mv` function in the “metafor” package (Viechtbauer, 2010) in R (version 3.4.1) including both study and sample as random effect (study nested within sample in order to deal with potentially non-independent effect sizes coming from the same manuscript or the same sample of participants (Assink & Wibbelink, 2016; Konstantopoulos, 2011). Publication bias was assessed using the Rank correlation test (Begg & Mazumdar, 1994). This approaches may be less appropriate for mixed-effects meta-analysis which include non-independent data points (Assink & Wibbelink, 2016). However, we provide the results of this tests to be consistent with prior meta-analyses. Heterogeneity was assessed using the Brestlow-day test (Cochran, 1954) and the method proposed by Higgins et al. (termed I^2 -squared; Higgins & Thompson, 2002).

We conducted separate sets of analyses to explore the associations of ELA with two metrics of biological aging: pubertal timing and cellular aging. As described above, data on neural development was not reported in a manner across studies that permitted meta-analysis; instead, these results are systematically reviewed. Within each set of analyses, we began by exploring the association of all adversity types (regardless of dimension) with our two domains of biological aging, then examined whether adversity type was a moderator of these associations. If the moderator analysis was significant, we then ran separate sensitivity

analyses to examine associations separately by threat, deprivation, and SES to assess associations of each adversity type with biological aging outcomes.

Moderator Analyses

In cases where effect sizes showed significant heterogeneity, we tested whether demographic or methodological factors moderated the associations between ELA and biological aging. These factors were based on prior literature, and included sample race/ethnicity (% white), sex composition of the sample (% male), metric of pubertal timing (age of menarche OR measure of secondary sex characteristics), and whether the study controlled for BMI (0/1) or other forms of adversity (0/1). For telomere length and DNAm age we examined the use of blood vs. saliva as a potential moderator in our analyses. If there was no information given by the manuscript regarding a specific moderator then they were marked as missing and not included in the moderator analysis. We tested each moderator separately using the moderator flag in the `rma.mv` function.

Results

Pubertal Timing

The 37 studies included in this meta-analysis produced 86 effect sizes and a total of 63,914 participants. Sample sizes ranged from 25 to 16,202 (Median=480). Of the 86 effect sizes, 20 focused on ELA characterized as threat, 21 focused on deprivation, 33 on SES, and 12 used a cumulative approach of summing across multiple forms of adversity. Table 1 presents descriptive demographic information for each study.

All Adversities. We first examined the effect of all forms of adversity on pubertal timing across all 37 studies included in the meta-analysis. Greater exposure to ELA was associated with earlier pubertal timing ($d=-0.12$, 95% CI [-0.22, -0.02]) and significantly differed from zero ($Z=-2.41$, $p=0.02$; Figure 2). Significant heterogeneity was observed across studies ($Q(85)=625.50$, $p<0.0001$; $I^2=96.04$). The result of Begg's publication bias test was not

significant (Kendall's tau=-0.10, $p=0.18$), suggesting no publication bias in our sample of studies.

Using adversity type as a moderator, we tested our hypothesis that threat would have a significant negative effect on pubertal timing (suggesting accelerated development), whereas deprivation would have a significant positive effect on pubertal timing (suggesting delayed development). The random-effects meta-analysis including 4 adversity types as a moderator (threat, deprivation, SES, and any studies using only a composite measure of adversity) revealed that adversity type significantly moderated the association between ELA and pubertal timing ($QM(3)=50.89$, $p<0.0001$). Given significant differences across adversity type, we explored the effect of adversity on pubertal timing separately for each category of adversity.

Threat. In studies that specifically explored the association of threat exposure with pubertal timing (11 studies; 20 effect sizes, $N=20,401$), greater exposure to threat was associated with earlier pubertal timing ($d=-0.26$, 95% CI [-0.41, -0.11]). The effect size was small and significantly differed from zero ($Z=-3.44$, $p<0.001$; Figure 3). Significant heterogeneity was observed across studies ($Q(19)=226.32$, $p<0.001$; $I^2=95.15$). The result of Begg's publication bias test was not significant (Kendall's tau=-0.31, $p=0.07$), suggesting no publication bias in our sample of studies.

Given significant heterogeneity in our studies examining the association of threat-related adversities with pubertal timing, we conducted a series of moderator analyses. None of the five moderators (metric of pubertal timing, sex, race/ethnicity, BMI, controls for other ELA types) were significantly associated with variations in effect size.

Deprivation. In studies that specifically focused on the association of deprivation exposure with pubertal timing (12 studies; 21 effect sizes, $N=34,193$), deprivation was not associated with pubertal timing ($d=0.05$, 95% CI [-0.07, 0.18]) and did not significantly differ from zero ($Z=0.81$, $p=0.42$; Figure 3). Significant heterogeneity was observed across studies

($Q(20)=51.17$, $p<0.001$; $I^2=89.34$). The result of Begg's publication bias test was not significant (Kendall's tau= 0.24. $p=0.15$), suggesting no publication bias in our sample of studies.

Given significant heterogeneity in our studies examining the association of deprivation-related adversities with pubertal timing, we conducted moderator analyses. For the association between deprivation and pubertal timing, sex was significantly associated with variation in effect sizes (Estimate=0.01, SE=0.00, $Z=3.50$, $p<0.001$), suggesting that the more males included in the sample, the more positive the association between deprivation exposure and pubertal timing (i.e., the more delayed the pattern of maturation).

SES. When looking at studies that explored the effect of SES specifically on pubertal timing (21 studies; 33 effect sizes, $N=34,489$), the random-effects meta-analysis found that SES was associated with earlier pubertal timing ($d=-0.15$, 95% CI [-0.30, 0.01]), but the effect size was not significantly different than zero ($Z=-1.90$ $p=0.06$; Figure 3). Significant heterogeneity was observed across studies ($Q(32)=211.66$, $p<0.001$; $I^2=96.22$). The result of Begg's publication bias test was significant (Kendall's tau=-0.27, $p=0.03$), suggesting publication bias in our sample of studies.

Given significant heterogeneity in our studies examining the association of SES with pubertal timing, we conducted a series of moderator analyses. None of the five moderators were significantly associated with variation in effect sizes.

Cellular Aging

A total of 9 studies (7 examining telomere length, 2 examining DNA methylation age) produced 14 effect sizes across a total of 1,011 participants. Sample sizes for included studies ranged from 38 to 247 (Median=99). Of the 14 effect sizes, 4 focused on the effect of threat on cellular aging, 2 focused on deprivation, 5 on SES, and 3 used a cumulative approach of summing across multiple forms of adversity. Table 2 presents descriptive demographic information for each study.

All Adversities. We first examined the effect of all forms of adversity on cellular aging across all 9 studies included in the meta-analysis. The random-effects meta-analysis found that greater exposure to early-life adversity was associated with accelerated cellular aging ($d=-0.30$, 95% CI [-0.46, -0.15]) and significantly differed from zero ($Z=-3.83$, $p<0.0001$; Figure 4). There was no significant heterogeneity observed across studies ($Q(13)=20.30$, $p=0.09$; $I^2=31.64$). The result of Begg's publication bias test was not significant (Kendall's tau=-0.27, $p=0.19$), suggesting no publication bias in our sample of studies.

Although there was limited heterogeneity across studies, given our theoretical interests and hypotheses about differences in associations by adversity type, we examined adversity type as a moderator of these associations to evaluate our hypothesis that threat would be associated with advanced cellular aging and determine whether these effects were similar for other adversity types. Adversity type significantly moderated the association between ELA and cellular aging ($QM(3)=9.68$, $p=0.02$). We additionally explored the associations of ELA with cellular aging separately for each adversity type.

Threat. In studies that explored the association of threat exposure with cellular aging (4 studies; 4 effect sizes, $N=664$), greater exposure to threat was associated with accelerated cellular aging ($d=-0.43$, 95% CI [-0.61, -0.25]). The effect size was moderate in magnitude and differed from zero ($Z=-4.65$, $p<0.0001$; Figure 5). Significant heterogeneity was not observed across studies ($Q(3)=0.39$, $p=0.94$; $I^2=0.00000001$). The result of Begg's publication bias test was not significant (Kendall's tau=-0.33, $p=0.75$), suggesting no publication bias in our sample of studies. We did not explore moderators given the lack of heterogeneity in effect sizes.

Deprivation. In studies that explored the association of deprivation exposure with cellular aging (2 studies; 2 effect sizes, $N=347$), the random-effects meta-analysis found that deprivation was not associated with cellular aging ($d=-0.06$, 95% CI [-0.30, 0.17]), with an effect size that did not significantly differ from zero ($Z=-0.51$ $p=0.61$; Figure 5). Significant heterogeneity was not observed across studies ($Q(1)=0.060$, $p=0.81$; $I^2=0.00000001$). The

result of Begg's publication bias test was not significant (Kendall's tau=-1.000, $p=1.000$), suggesting no publication bias in our sample of studies. However, given this analysis only contained two effect sizes, this is not a reliable estimate of publication bias. We did not explore moderators given the lack of heterogeneity in effect sizes.

SES. In studies of SES and cellular aging (4 studies; 5 effect sizes, $N=456$), SES was not associated with cellular aging ($d=-0.16$, 95% CI [-0.53, 0.20]), with an effect size that did not significantly differ from zero ($Z=-0.87$, $p=0.38$; Figure 5). Significant heterogeneity was not observed across studies ($Q(4)=7.64$, $p=0.11$; $I^2=53.89$). The result of Begg's publication bias test was not significant (Kendall's tau=-0.60, $p=0.23$), suggesting no publication bias in our sample of studies. We did not explore moderators given the lack of heterogeneity in effect sizes.

Brain Development

We systematically reviewed the associations between ELA and two metrics of brain development: cortical thickness and task-based amygdala-PFC functional connectivity. Across the two metrics there were 19 studies across a total of 2,276 unique participants (13 cortical thickness papers, $N = 1,848$, 6 amygdala-PFC connectivity papers, $N = 428$).

Cortical Thickness

Threat. We found four papers that investigated the association between experiences of threat and cortical thickness in childhood and adolescence ($N=161$; Busso et al., 2017; Gold et al., 2016; Kelly et al., 2013; McLaughlin, Sheridan, et al., 2016). Of these four studies, three found that children exposed to threat had accelerated thinning of the cortex. Critically, all three of these studies found decreased cortical thickness among children exposed to threat in the ventromedial PFC (Busso et al., 2017; Gold et al., 2016; Kelly et al., 2013), and two found additional associations that follow the same pattern of decreased thickness among threat-exposed youths in regions including the lateral PFC and medial and lateral temporal cortex (Busso et al., 2017; Gold et al., 2016). In contrast, one study that spanned a larger age range found no association between experiences of threat and cortical thickness in regions of interest

in the dorsal anterior cingulate or ventromedial PFC (McLaughlin, Sheridan, et al., 2016). As a whole, these studies provide support for the hypothesis that experiences of threat are associated with accelerated development, especially in the ventromedial PFC.

Deprivation. A total of four studies (N = 353) met inclusion criteria for our review of the association between experiences of deprivation and cortical thickness. This included two studies investigating cortical structure among previously institutionalized children (Hodel et al., 2015; McLaughlin, Sheridan, Winter, et al., 2014) and two investigating the association between cognitive stimulation/deprivation in the home environment and cortical structure (Avants et al., 2015; Rosen et al., 2018). Both studies of institutionalized children demonstrate support for the hypothesis that experiences of deprivation are associated with accelerated cortical thinning. In one study institutionalization was associated with widespread reductions in cortical thickness, including in nodes of the frontoparietal and dorsal attention networks (superior parietal lobule, frontal pole, superior frontal gyrus), default mode network (inferior parietal cortex, precuneus, mid-cingulate), lateral temporal cortex, parahippocampal cortex, and insula at age 8-10 years (McLaughlin, Sheridan, Winter, et al., 2014). In contrast, Hodel and colleagues (2015) found reduced cortical thickness only in the inferior frontal gyrus among previously institutionalized children compared to controls at age 12-14 years.

The other two studies investigated cortical thickness and its association with cognitive stimulation/deprivation in the home environment. In a cross-sectional study spanning children and adolescents, cognitive stimulation (i.e., lower deprivation) was positively associated with cortical thickness in two nodes in the left, but not right frontoparietal network (superior parietal lobule and middle frontal gyrus) (Rosen et al., 2018). These results are consistent with the idea that deprivation (i.e. low cognitive stimulation) is associated with accelerated development. In contrast, in a longitudinal study, results revealed that cognitive stimulation at age 4, but not at age 8, was negatively associated with cortical thickness at age 19, such that lower deprivation was associated with thicker cortex in the ventral temporal cortex and inferior frontal gyrus.

These results suggest that cognitive deprivation is associated with delayed development in these regions.

SES. Six studies met the criteria for inclusion examining SES and cortical thickness (N = 1512) (Jednoróg et al., 2012; Lawson, Duda, Avants, Wu, & Farah, 2013; Mackey et al., 2015; Noble et al., 2015; Piccolo, Merz, He, Sowell, & Noble, 2016; Rosen et al., 2018). Of those, four found that low SES was associated with thinner cortex across large swaths of cortex encompassing the frontoparietal network (lateral prefrontal cortex, superior parietal cortex), default mode network (lateral temporal cortex, precuneus), and the visual system (lateral occipital and ventral temporal cortex), supporting the idea that low SES is associated with accelerated cortical development. Piccolo and colleagues (2016) found that SES moderates the association between age squared and cortical thickness such that low SES individuals show a sharper decline in cortical thickness early in development, which may reflect accelerated development compared to higher SES individuals. Additionally, Lawson and colleagues (2013) demonstrate that low parent education is associated with reduced cortical thickness in the right cingulate gyrus and right superior frontal gyrus, and Mackey and colleagues (2015) demonstrate that low SES individuals demonstrate thinner cortex across much of the brain including the frontoparietal network (right middle frontal gyrus, left superior parietal lobule, right frontal pole), default mode network (left precuneus, bilateral lateral temporal cortex, right frontal pole), and visual system (bilateral occipital and ventral temporal cortex). Two studies spanning larger age ranges (Noble et al., 2015; Rosen et al., 2018) found no association between SES and cortical thickness. Importantly, Noble et al., (2015) and Piccolo et al., (2016) used the same sample and while there were no main effects of SES on cortical thickness and no age x SES interactions (Noble et al., 2015), Piccolo and colleagues demonstrate an age squared x SES interaction such that children from low-income households demonstrate accelerated thinning compared to high-income counterparts.

Amygdala-PFC Connectivity

Threat. Our search yielded four papers that evaluated the association between amygdala-PFC connectivity and threat-related experiences. Of these four studies (N = 250), two support the hypothesis that experiences of threat are associated with accelerated maturation of this network such that threat-exposed children exhibit more negative connectivity between amygdala and PFC during both an implicit dynamic emotion face task, and an explicit affect labeling task, than children of the same age (Colich et al., 2017; Keding & Herringa, 2016). The two other studies demonstrate the opposite pattern of results such that children who have experienced threat demonstrate more positive task-related amygdala-PFC connectivity compared to controls while viewing emotional faces and while performing an emotional conflict task (Cisler, Scott Steele, Smitherman, Lenow, & Kilts, 2013; Marusak et al., 2015). These mixed findings do not provide conclusive evidence that experiences of threat are associated with either accelerated or delayed development of the circuits.

Deprivation. Our search yielded two papers (N = 89) that evaluated the association between experiences of deprivation and task-related connectivity between mPFC and amygdala. Of these two studies, one demonstrated evidence for accelerated development of these circuits such that children who have experienced deprivation exhibit more negative connectivity earlier in development than comparison children in a passive viewing task of facial emotion (Gee, Gabard-Durnam, et al., 2013). The other study found the opposite pattern of results such that children who had experienced deprivation demonstrated more positive amygdala-mPFC connectivity than comparison children in a fear conditioning paradigm (Silvers, Lumian, et al., 2016).

Discussion

Through the use of meta-analysis and systematic review, we provide evidence that ELA accelerates biological aging, as measured by pubertal timing, cellular aging, and cortical thinning in childhood and adolescence. We found no evidence for a consistent effect of ELA on accelerated development of amygdala-mPFC connectivity. First, although we observed an

overall association of ELA with pubertal timing, moderator analysis revealed that ELA characterized by threat, but not deprivation or SES, was associated with accelerated pubertal development with a small effect size, suggesting specificity in the link between ELA and pubertal timing to threat-related adversity. Second, ELA was also associated with accelerated cellular aging as measured by both leukocyte telomere length and DNA methylation age. Again, moderator analyses revealed accelerated aging among children exposed to threat of moderate effect size, but no association with deprivation or SES. Finally, the results of our systematic review of the effects of ELA on brain development revealed a consistent association between ELA and accelerated cortical thinning across multiple types of ELA, although the specific brain regions involved vary by adversity type. Associations of threat with cortical thinning were most consistent in ventromedial PFC, whereas associations of deprivation with cortical thinning were most consistent in the frontoparietal and default mode networks and the ventral visual stream. In contrast, there was no consistent association of ELA with amygdala-mPFC connectivity. These findings suggest both common and specific effects of dimensions of ELA across multiple domains of biological aging.

ELA and Pubertal Timing

ELA was associated with accelerated pubertal timing overall, but significant heterogeneity existed in this effect as a function of adversity type. The strength of the association of ELA with pubertal timing was significantly moderated by adversity type, such that the association between ELA and accelerated pubertal timing was specific to experiences characterized by threat, and showed no association with deprivation or SES. These results are consistent with predictions from life history models that exposure to environmental harshness (i.e. threat) in childhood accelerates sexual maturation, in order to increase chances of reproduction prior to mortality (J. Belsky et al., 2012; Ellis et al., 2009). They are also consistent with recent findings from our lab demonstrating that threat-related adversities are associated with accelerated pubertal development even after adjustment for exposure to co-occurring

deprivation (Colich et al., 2019; Sumner et al., 2019). Some have argued that ELA impacts pubertal timing through influences on the hypothalamic-pituitary-adrenal (HPA) axis (Negriff, Saxbe, et al., 2015; Saxbe, Negriff, Susman, & Trickett, 2015). Given associations between threat-related adversity and altered diurnal patterns of cortisol and cortisol reactivity in childhood (Carpenter, Shattuck, Tyrka, Geraciotti, & Price, 2011; Jaffee et al., 2014; King et al., 2017; Tyrka et al., 2009), it is plausible that trauma-related alterations of the HPA-axis may interact with the HPG-axis to accelerate the onset of pubertal development (Negriff, Saxbe, et al., 2015; Saxbe et al., 2015). It is also important to consider the role of gene-environment correlation in the association between threat-related ELA and pubertal timing (Cousminer, Widén, & Palmert, 2015; Harden, 2014; Rowe, 2002). For instance, mothers who experience earlier onset of puberty may reproduce at an earlier age, and have children who are both more likely to experience trauma and an earlier onset of puberty (de Vries, Kauschansky, Shohat, & Phillip, 2004; Towne et al., 2005). Future research should explore how maternal age at menarche influences the associations of ELA and pubertal timing in their offspring in order to better understand the mechanisms linking threat-related ELA and accelerated pubertal development.

We did not find support for our hypothesis that ELA characterized by deprivation would show an association with delayed pubertal timing. Instead, we found no association between deprivation and pubertal timing. Life history theory posits that deprivation of bioenergetics resources could result in delayed maturation and later pubertal development (J. Belsky et al., 2012; Ellis et al., 2009). In this analysis, we included emotional and physical neglect (Boynton-Jarrett & Harville, 2012; Colich et al., 2019; Mendle et al., 2014, 2011, 2016; Ryan et al., 2015; Sumner et al., 2019), and early institutionalization (G. et al., 2009; Hayes & Tan, 2016; Johnson et al., 2018; Reid et al., 2017; Sonuga-Barke, Schlotz, & Rutter, 2010) as forms of deprivation. It is likely that deprivation in our modern context, represented by the forms of psychosocial deprivation included in our analyses, is qualitatively different from deprivation in our evolutionary past. Whereas there is strong evidence for associations of food insecurity and severe

deprivation associated with war and famine with delayed pubertal timing (Prebeg & Bralic, 2000; van Noord & Kaaks, 1991), there is less support for association of early institutionalization (where most children experience severe emotional deprivation but not necessarily food insecurity) with pubertal timing (Johnson et al., 2018; Reid et al., 2017).

In this meta-analysis, we decided to isolate the effects of deprivation (including neglect and early life institutionalization) from the effects of SES. Low SES, defined as poverty and low parental education, has previously been used as an indicator of deprivation in studies that adjust for co-occurring threat exposure (e.g., Lambert et al., 2017; Sheridan et al., 2017), based on extensive evidence demonstrating that children from families with low parental education and/or income experience reductions in cognitive and social stimulation than children from higher-SES families (Bradley et al., 2001; Duncan & Magnuson, 2012). However, within the DMAP model, poverty is conceptualized as a risk factor for both threat and deprivation, rather than a direct marker of deprivation (McLaughlin, Sheridan, Lambert, et al., 2014; Sheridan & McLaughlin, 2014). Indeed, there is a strong association between SES and exposure to violence (Foster, Brooks-Gunn, & Martin, 2007) in addition to deprivation. Here, the association of SES with accelerated pubertal timing was not significant, potentially reflecting the fact that SES is a risk marker for exposure to other forms of adversity (e.g., trauma) that are associated with accelerated pubertal development rather than having a direct effect on pubertal timing. Overall, these findings highlight the importance of considering the nature of the exposure when exploring the developmental consequences of ELA. Future research should carefully distinguish between the effects of threat- and deprivation-related adversities on pubertal timing.

There was no evidence for the moderating effect of metric of pubertal timing (age of menarche vs. secondary sex characteristics) or race/ethnicity on associations of ELA with pubertal timing, or whether studies controlled for BMI or exposure to other adversity types. However, sex did moderate the association between deprivation-related adversities and pubertal timing such that the more males included in the sample, the more positive the

association between deprivation exposure and pubertal timing, suggesting more delayed pubertal maturation. These results suggest that deprivation may have a differential effect on males and females. Animal models of nutritional challenge (including undernutrition and obesity) have differential effects on pubertal timing in male and female rats (Sánchez-Garrido et al., 2013), suggesting that sex-specific metabolic effects of deprivation may have a significant impact on pubertal timing. Future research in humans studying the impact of food insecurity specifically, should explore this question directly.

ELA and Cellular Aging

ELA was associated with accelerated cellular aging, as measured by both leukocyte telomere length and DNAm age, such that greater exposure to adversity was associated with decreased telomere length and more advanced DNAm age relative to chronological age. These results replicate earlier meta-analyses conducted in adult populations of adversity with DNAm age (Wolf et al., 2018) and telomere length (Hanssen, Schutte, Malouff, & Epel, 2017). These results are also broadly consistent with an earlier meta-analysis exploring the effects of stress exposure (broadly defined) on telomere length (Coimbra et al., 2017). The consistency in findings is striking given significant differences in the approach of these meta-analyses. Whereas Wolf and colleagues (2018) and Hanssen and colleagues (2017) examined the association between ELA and accelerated biological aging in adults, Coimbra et al. (2017) examined a broad range of stressors in childhood and adolescence, including stress reactivity as indexed by cortisol reactivity and parental psychopathology. We did not include cortisol reactivity or parental psychopathology as adversities in the current meta-analysis, yet results are largely consistent with Coimbra and colleagues.

We did not observe heterogeneity in the associations of ELA with cellular aging. However, in stratified analysis, we found that exposure to threat was associated with accelerated cellular aging of moderate magnitude, whereas neither deprivation nor SES was associated with cellular aging. These differential associations should be interpreted with caution, however, as our

analysis of whether type of ELA was a moderator effect size magnitude was only significant at a trend-level and the number of studies examining deprivation and SES with cellular aging were small. Nonetheless, these findings suggest that threat-related adversities are consistently associated with accelerated cellular aging. Greater work is needed to clarify the magnitude and direction of effects for deprivation and childhood SES.

These results are consistent with “internal prediction” models of predictive adaptive response (Nettle et al., 2013; Rickard et al., 2014), which propose that ELA negatively influences physical health through altered cellular development as a result of reduced energy to build or repair cellular tissue. This theory expands earlier models focused on allostatic load, or the accumulation of environmental insults on biological systems (Danese & McEwen, 2012; McEwen, 1998; McEwen & Stellar, 1993), and developmental origins of health and disease models (Barker, 2007) focused on how early experience programs biological development to adapt to later environmental conditions. These theories all suggest that accelerated cellular aging occurs as a result of environmental experiences in development. Accelerated cellular aging following ELA may occur in response to alterations in mitochondrial function, oxidative stress, and inflammation (Shalev, 2012). Although we observed consistent effects of ELA across two metrics of cellular aging (telomere length and DNAm age), some work indicates that exposure to ELA may not have consistent associations across other metrics of biological aging (D. W. Belsky et al., 2015). Future research should explore the effect of distinct forms of ELA on additional metrics of allostatic load that may represent accelerated biological aging, including cardiometabolic risk, inflammation, and respiratory health.

ELA and Brain Development

Cortical Thickness. Consistent with the hypothesis that ELA leads to accelerated development, the majority of studies investigating the association between ELA and cortical thickness found that children exposed to adversity of any kind have thinner cortex than their non-exposed counterparts across threat, deprivation, and SES. However, it is critical to note

that the specific brain regions that exhibited this pattern of thinning varied consistently by adversity type. This specificity may reflect precocious maturation of particular regions of the brain depending on the particular type of adversity experienced, reflecting adaptive experience-related tuning of neural systems to the environment in which they are developing. There was remarkable consistency across studies of threat-related experiences and cortical thickness, with the majority observing thinner cortex in the ventromedial PFC among children exposed to trauma (Busso et al., 2017; Gold et al., 2016; Kelly et al., 2013). The vmPFC is implicated in multiple forms of emotion processing, including recall of extinction learning, appraisal of episodic memories, and appraisal of simulated future events (Dixon, Thiruchselvam, Todd, & Christoff, 2017; Milad & Quirk, 2012; Phelps & LeDoux, 2005). The vmPFC has strong interconnections with the amygdala and modulates amygdala activation based on appraisals and prior learning (Phelps & LeDoux, 2005). Accelerated thinning of this region among children exposed to trauma could reflect earlier or more frequent recruitment of this region to modulate amygdala responses, which are well-established to be elevated in response to threat cues among children exposed to violence (Hein & Monk, 2017; McCrory, De Brito, & Viding, 2011; McLaughlin, Peverill, Gold, Alves, & Sheridan, 2015), ultimately producing more rapid specialization of this region, potentially through more rapid synaptic pruning or increased myelination in this region.

Association between experiences of deprivation and cortical structure were more mixed. While one study of previously institutionalized children demonstrated widespread reductions in cortical thickness across regions of the frontoparietal, default mode, and visual networks (McLaughlin, Sheridan, Winter, et al., 2014), another found reduced cortical thickness only in the inferior frontal gyrus (Hodel et al., 2015). Studies investigating low cognitive stimulation have also been mixed. While one study found that low cognitive stimulation was associated with thinner cortex in the frontoparietal network across childhood and adolescence (Rosen et al., 2018), another found that lower cognitive stimulation was associated with thicker cortex in the

lateral prefrontal cortex and ventral visual stream in late adolescents (Avants et al., 2015).

Differences in the age of the samples and timing of assessment of cognitive stimulation may have contributed to these inconsistent findings.

The studies investigating SES-related differences in cortical thickness also had mixed results. Two studies found widespread positive associations with SES and thickness in the frontoparietal and default mode networks and the visual system (Jednoróg et al., 2012; Mackey et al., 2015). One study focused only on the PFC also found similar reductions in thickness (Lawson et al., 2013). Broadly, these regions are involved in a wide range of cognitive processing including working memory, cognitive control, autobiographical memory, theory of mind, and visual processing (Cole & Schneider, 2007; Corbetta, Kincade, & Shulman, 2002; DiCarlo, Zoccolan, & Rust, 2012; Spreng & Grady, 2010). Given that SES-related differences in many of these domains are well-established (Noble, McCandliss, & Farah, 2007), these findings could represent a neural mechanism explaining these SES-related differences in cognitive function. In contrast, two studies spanning a large age range did not find SES-related differences in thickness (Noble et al., 2015; Rosen et al., 2018). This could be because SES associations with cortical thickness vary across childhood and adolescence. Indeed, using the same sample as Noble and colleagues, Piccolo and colleagues (2016) found an SES by age interaction for average cortical thickness such that lower SES was associated with a more rapid age-related decrease in cortical thinning early in development while higher SES was associated with a less steep linear decline in thickness from childhood to adolescence. These findings are consistent with the hypothesis that low SES is associated with accelerated maturation of the cortex.

Linear decreases in cortical thickness from infancy to adulthood are well-established (LeWinn et al., 2017; Vijayakumar et al., 2016; Wierenga et al., 2014), although the mechanisms by which this pattern emerges remain in question. One interpretation of is that synaptic connections that are underutilized or inefficient are pruned, allowing the brain to adapt

to the environment in which it develops (Huttenlocher, 1979; Petanjek et al., 2011; Rakic, Bourgeois, Eckenhoff, Zecevic, & Goldman-Rakic, 1986). If pruning is the primary mechanism driving cortical thinning, it is possible that ELA-related differences in cortical thickness are due to accelerated pruning. In the case of deprivation-related experiences, this may be due to a lack of experience with socially or cognitively stimulating environments (McLaughlin, Sheridan, & Nelson, 2017). Alternatively, greater pruning could reflect precocious specialization and maturation of circuits utilized more frequently by children exposed to ELA; in the absence of behavioral data associated with specific patterns of cortical thinning, caution is warranted in interpreting these patterns as either adaptive or maladaptive (ME., MD., & IH., 2016). Other work suggests that age-related decreases in cortical thinning may actually be due to increases in myelination across development (Natu et al., 2018; Sowell et al., 2004). Increased myelination, which is most pronounced in deeper cortical layers may increase the intensity of voxels at the grey-white matter border, therefore making the cortex appear thinner across age. If myelination is the primary mechanism by which cortical thinning happens, it is possible that increased cortical thickness in response to ELA may be due to faster development of structural connectivity between regions. Of course, these mechanisms are not mutually exclusive and future longitudinal work measuring multiple forms of ELA utilizing both T1-weighted imaging and diffusion tensor imaging is needed to disentangle the precise mechanisms by which ELA leads to thinner cortex in youths.

Amygdala-PFC Connectivity. Existing work examining ELA and task-related amygdala-PFC connectivity has produced mixed findings. Across both threat and deprivation, approximately half of the studies observed that ELA was associated with more negative functional connectivity, indicating accelerated development (Colich et al., 2017; Gee, Gabard-Durnam, et al., 2013; Keding & Herringa, 2016), while several others showed the opposite pattern of results such that youths exposed to ELA demonstrate more positive amygdala-PFC connectivity than non-exposed youths, indicating delayed development (Cisler, James, et al.,

2013; Marusak et al., 2015; Silvers, Insel, et al., 2016a). Therefore, existing work has yet to provide clear evidence for an association between ELA and accelerated development of these systems. Moreover, there is no clear evidence that specific types of adversity have differential influences on the development of this circuit.

One possibility is that amygdala-PFC functional connectivity is not a reliable marker of neural development. Unlike cortical thickness which has been studied widely across large representative samples (for review see Vijayakumar et al., 2016), research documenting amygdala-PFC connectivity as a marker of maturation is more modest (Gee, Humphreys, et al., 2013; Kujawa et al., 2016; Silvers, Insel, et al., 2016b; Wu et al., 2016), and to our knowledge, all of the studies that have demonstrated a developmental shift in this circuit have been cross-sectional. As such, amygdala-mPFC connectivity may be an unreliable marker of neural maturation. Alternatively, while all these tasks focused on some sort of emotional processing, it is possible that heterogeneity across different tasks may contribute to differences in results. Future longitudinal work with a range of emotional processing tasks will be needed to establish the developmental trajectory of amygdala-mPFC connectivity to determine whether it is a robust metric of development.

Effect of ELA Across Multiple Domains of Biological Aging

Given the range in operationalizing accelerated development and potential mechanisms linking ELA and accelerated biological development, it is surprising that few have attempted to reconcile across these different metrics of maturation. Only three studies to our knowledge have incorporated multiple metrics of accelerated development in adolescence. Belsky and Shalev (2016) put forth a “two-hit” model suggesting that ELA accelerates development first through telomere erosion and second, through earlier reproduction, which can increase oxidative stress and accelerate telomere erosion. Although this model accounts for two forms of accelerated aging, it does not directly compare and contrast the effects of ELA on both metrics of accelerated development – cellular aging and pubertal timing. Sumner and colleagues (Sumner

et al., 2019) examined how exposure to threat and deprivation-related ELA influenced both DNAm age and pubertal timing. They found that exposure to threat, but not deprivation, contributes to both accelerated DNAm age and accelerated pubertal timing. Understanding the effect of ELA across domains of biological aging is important as it reflects a unique or shared mechanism linking ELA and accelerated development. There is also evidence to suggest that pubertal timing and cellular aging are highly correlated (Binder et al., 2018), suggesting a potential shared mechanism contributing to the development of both domains. However, other work demonstrates variation in the rate of change across different metrics of biological aging (D. W. Belsky et al., 2015), indicating that multiple mechanisms might underlie the ELA-accelerated development association, depending upon the metric of accelerated development. Although increased allostatic load has been proposed as a mechanism linking ELA to accelerated pubertal timing (Danese & McEwen, 2012; McEwen, 1998), empirical evidence testing this possibility is currently lacking. Moreover, allostatic load is a multi-dimensional construct involving numerous biological systems, and it is unclear if accelerated weathering occurs across all systems to a similar degree (Geronimus, 1992; Geronimus, Hicken, Keene, & Bound, 2006). If allostatic load is a mechanism contributing to accelerated pubertal development, it could explain our disparate findings regarding threat and deprivation as exposure to early-life trauma has been consistently associated with elevated allostatic load (Danese & McEwen, 2012; Scheuer et al., 2018; Widom, Horan, & Brzustowicz, 2015), whereas recent work indicates that even extreme exposure to deprivation association with institutional rearing is unrelated to allostatic load (Slopen et al., 2019). It is clear that greater work is needed to elucidate the mechanisms underlying accelerated development following exposure to ELA across domains, whether they are global or specific to particular dimensions of early experience, and how these mechanisms ultimately contribute to changes in reproductive function, cellular aging, and brain development.

Implications of Accelerated Development

Accelerated aging across domains has been associated with a host of mental and physical health problems. For instance, accelerated pubertal timing is linked with a range of mental health problems including heightened levels of risk-taking behavior, delinquency and substance abuse problems (Copeland et al., 2013; Harden & Mendle, 2012), as well as depression and anxiety disorders (Hamilton, Hamlat, Stange, Abramson, & Alloy, 2014; Mendle, Harden, Brooks-Gunn, & Graber, 2010; Mendle et al., 2014; Negriff & Susman, 2011; Ullsperger & Nikolas, 2017). Accelerated pubertal timing is also associated with a range of physical health problems, including cardiovascular disease, polycystic ovarian syndrome in females, and testicular cancer in males (Day, Elks, Murray, Ong, & Perry, 2015; Golub et al., 2008; Lakshman et al., 2009; Velie, Nechuta, & Osuch, 2006). Accelerated cellular aging has also been associated with depression (Ridout, Ridout, Price, Sen, & Tyrka, 2016), anxiety (Malouff & Schutte, 2017), posttraumatic stress disorder (Li, Wang, Zhou, Huang, & Li, 2017), cardiovascular disease (Rehkopf et al., 2016), cancer (Zhu et al., 2016), and all-cause mortality (Needham, Rehkopf, et al., 2015). Finally, altered trajectories of cortical development have been linked to variations in general intelligence (Shaw et al., 2006) and attention-deficit hyperactivity disorder (McLaughlin, Sheridan, Winter, et al., 2014). Little research has directly examined whether accelerated development in these systems is a consequence of pre-existing mental and physical disorders, or a mechanism explaining elevated risk for mental and physical health problems in youth who have experienced ELA (see J. Belsky, Ruttle, Boyce, Armstrong, & Essex, 2015; Colich et al., 2019; Mendle et al., 2014; Negriff, Saxbe, et al., 2015 for work that has explored this idea). For instance, some evidence suggests that accelerated pubertal timing explains a significant proportion of the association between threat-related ELA risk for mental health problems in adolescence (Colich et al., 2019), and that telomere shortening occurs prior to the onset of depression in an at-risk population (Gotlib et al., 2015). However, there is also evidence to suggest that early psychosocial difficulties precede early pubertal onset (Mensah et al., 2013) and could potentially accelerate cellular aging as well (Lindqvist et al., 2015). A key

issue for future research will be to determine whether early interventions targeting psychosocial mechanisms linking ELA with mental and physical health problems are capable of altering observed patterns of accelerated biological aging.

It is also important to acknowledge that although there are strong associations among accelerated development and negative mental and physical health outcomes, accelerated development is most likely an adaptation to current and presumably future environmental conditions (J. Belsky, 2019). In a highly dangerous or unpredictable environment, it may be adaptive in the short-term to reach adult-like capabilities at an earlier age, in order to either reach reproductive status earlier, or reach independence from the caregiving situation at an earlier age. This immediate goal may outweigh the longer-term consequences of mental and physical health problems. If the environment is signaling imminent mortality, then this trade-off is one that is evolutionarily adaptive. It will be important that future work consider the adaptive significance of accelerated development in response to ELA.

Limitations and Future Directions

Several limitations of this work highlight key directions for future research. First, we examined a relatively small number of studies for some domains and within each dimension of adversity, particularly with regard to cellular aging and brain development. More research is needed to evaluate whether all forms of adversity influence cellular aging or whether these associations are stronger for experiences of threat. Similarly, due to the small number of studies published on these variables, we collapsed across measures of cellular aging, including telomere length and DNAm age. These markers reflect distinct biological processes with differing molecular signatures, and we recognize that combining across these two metrics of cellular aging is most likely an over-simplification of the effects of ELA on cellular aging. These findings should be replicated when more studies have been published on the associations of ELA with both telomere length and DNAm age. Similarly, it is important to note that we did not include studies examining the effects of ELA on methylation patterns of single genes due

difficulties in understanding what typical developmental patterns of specific gene methylation would be. For a systematic review of the effects of ELA on gene-specific methylation patterns see Lang et al. (2019). Second, although we show associations among ELA, pubertal timing, cellular aging, and cortical thinning, we do not have the data to speak to the underlying mechanisms driving these associations. Although we speculate that accelerated biological development following ELA is most likely due to the effects of ELA on allostatic load, future experimental work should investigate the underlying biological mechanisms supporting these associations. Finally, in only examining samples in childhood and adolescence, we hoped to examine the associations between ELA and accelerated development, independent of psychopathology. However, given strong links among ELA, accelerated development and psychopathology, it is impossible to confidently conclude that psychopathology is not driving the effects of ELA on accelerated biological development. Future longitudinal work using at-risk samples should address the directionality of these associations to determine with confidence the direct effect of ELA on accelerated biological aging.

CONCLUSIONS

Through meta-analysis and systematic review, we support the idea that ELA accelerates biological aging, as measured by pubertal timing, cellular aging, and cortical thinning in childhood and adolescence. However, these associations varied systematically as a function of adversity type. Specifically ELA characterized by threat, but not deprivation or SES, was associated with accelerated pubertal development, suggesting specificity in the link between ELA and pubertal timing. ELA was also associated with accelerated cellular aging as measured by both leukocyte telomere length and DNA methylation age, with the strongest evidence for threat-related ELA being associated with cellular aging. ELA was consistently associated with accelerated cortical thinning, with threat-related ELA associated with ventromedial PFC thinning and deprivation and SES more consistently associated with thinning in the frontoparietal, default mode, and visual networks. We found no consistent association of ELA with amygdala-mPFC

functional connectivity. These findings suggest both common and specific associations of dimensions of ELA with multiple domains of biological aging and highlight the importance of delineating the mechanisms through which specific types of early environmental experiences influence different aspects of biological aging in childhood and adolescence and determining how these pathways ultimately contribute to health disparities.

References

- Assink, M., & Wibbelink, C. J. M. (2016). Fitting three-level meta-analytic models in R: A step-by-step tutorial. *The Quantitative Methods for Psychology*, *12*(3), 154–174. <https://doi.org/10.20982/tqmp.12.3.p154>
- Avants, B. B., Hackman, D. A., Betancourt, L. M., Lawson, G. M., Hurt, H., & Farah, M. J. (2015). Relation of childhood home environment to cortical thickness in late adolescence: Specificity of experience and timing. *PLoS ONE*, *10*(10), 1–10. <https://doi.org/10.1371/journal.pone.0138217>
- Barbas, H., & García-Cabezas, M. Á. (2016). How the prefrontal executive got its stripes. *Current Opinion in Neurobiology*, *40*(4), 125–134. <https://doi.org/10.1016/j.conb.2016.07.003>
- Barker, D. J. P. (2007). The origins of the developmental origins theory. *Journal of Internal Medicine*, *261*(5), 412–417. <https://doi.org/10.1111/j.1365-2796.2007.01809.x>
- Begg, C. B., & Mazumdar, M. (1994). Operating Characteristics of a Rank Correlation Test for Publication Bias. *Biometrics*, *50*(4), 1088. <https://doi.org/10.2307/2533446>
- Belsky, D. W., Caspi, A., Houts, R., Cohen, H. J., Corcoran, D. L., Danese, A., ... Moffitt, T. E. (2015). Quantification of biological aging in young adults. *Proceedings of the National Academy of Sciences of the United States of America*, *112*(30), E4104–10. <https://doi.org/10.1073/pnas.1506264112>
- Belsky, J. (2012). *An Evolutionary Perspective on Child Development in the Context of War and Political Violence. The Oxford Handbook of Evolutionary Perspectives on Violence, Homicide, and War*. Oxford University Press. <https://doi.org/10.1093/oxfordhb/9780199738403.013.0023>
- Belsky, J. (2019). Early-Life Adversity Accelerates Child and Adolescent Development. *Current Directions in Psychological Science*, *096372141983767*. <https://doi.org/10.1177/0963721419837670>
- Belsky, J., Ruttle, P. L., Boyce, W. T., Armstrong, J. M., & Essex, M. J. (2015). Early adversity, elevated stress physiology, accelerated sexual maturation, and poor health in females. *Developmental Psychology*, *51*(6), 816–822. <https://doi.org/10.1037/dev0000017>
- Belsky, J., Schlomer, G. L., & Ellis, B. J. (2012). “Beyond cumulative risk: Distinguishing harshness and unpredictability as determinants of parenting and early life history strategy”: Correction. *Developmental Psychology*, *48*(3), 686–686. <https://doi.org/10.1037/a0025837>
- Belsky, J., & Shalev, I. (2016). Contextual adversity, telomere erosion, pubertal development, and health: Two models of accelerated aging, or one? *Development and Psychopathology*, *28*(4), 1367–1383. <https://doi.org/10.1017/S0954579416000900>
- Belsky, J., Steinberg, L., & Draper, P. (1991). Childhood Experience, Interpersonal Development, and Reproductive Strategy: An Evolutionary Theory of Socialization. *Child Development*, *62*(4), 647–670. <https://doi.org/10.1111/j.1467-8624.1991.tb01558.x>
- Binder, A. M., Corvalan, C., Mericq, V., Pereira, A., Santos, J. L., Horvath, S., ... Michels, K. B. (2018). Faster ticking rate of the epigenetic clock is associated with faster pubertal development in girls. *Epigenetics*, *13*(1), 85–94. <https://doi.org/10.1080/15592294.2017.1414127>
- Borenstein, M., Hedges, L. V., Higgins, J. P. T., & Rothstein, H. R. (2009). *Introduction to Meta-Analysis*. London, UK: Wiley.
- Boynton-Jarrett, R., & Harville, E. W. (2012). A prospective study of childhood social hardships and age at menarche. *Annals of Epidemiology*, *22*(10), 731–737. <https://doi.org/10.1016/j.annepidem.2012.08.005>
- Boynton-Jarrett, R., Ryan, L. M., Berkman, L. F., & Wright, R. J. (2008). Cumulative Violence Exposure and Self-Rated Health: Longitudinal Study of Adolescents in the United States.

- Pediatrics*, 122(5), 961–970. <https://doi.org/10.1542/peds.2007-3063>
- Bradley, R. H., Corwyn, R. F., Burchinal, M., McAdoo, H. P., & Garcia Coll, C. (2001). The Home Environments of Children in the United States Part II: Relations with Behavioral Development through Age Thirteen. *Child Development*, 72(6), 1868–1886. <https://doi.org/10.1111/1467-8624.t01-1-00383>
- Brown, D. W., Anda, R. F., Tiemeier, H., Felitti, V. J., Edwards, V. J., Croft, J. B., & Giles, W. H. (2009). Adverse Childhood Experiences and the Risk of Premature Mortality. *American Journal of Preventive Medicine*, 37(5), 389–396. <https://doi.org/10.1016/j.amepre.2009.06.021>
- Busso, D. S., McLaughlin, K. A., Brueck, S., Peverill, M., Gold, A. L., & Sheridan, M. A. (2017). Child Abuse, Neural Structure, and Adolescent Psychopathology: A Longitudinal Study. *Journal of the American Academy of Child & Adolescent Psychiatry*, 56(4), 321–328.e1. <https://doi.org/10.1016/j.jaac.2017.01.013>
- Busso, D. S., McLaughlin, K. A., & Sheridan, M. A. (2016). Dimensions of Adversity, Physiological Reactivity, and Externalizing Psychopathology in Adolescence. *Psychosomatic Medicine*, 79(2), 1. <https://doi.org/10.1097/PSY.0000000000000369>
- Callaghan, B., & Tottenham, N. (2016). The Stress Acceleration Hypothesis: effects of early-life adversity on emotion circuits and behavior. *Current Opinion in Behavioral Sciences*, 7, 76–81. <https://doi.org/10.1016/j.cobeha.2015.11.018>
- Carpenter, L. L., Shattuck, T. T., Tyrka, A. R., Geraciotti, T. D., & Price, L. H. (2011). Effect of childhood physical abuse on cortisol stress response. *Psychopharmacology*, 214(1), 367–375. <https://doi.org/10.1007/s00213-010-2007-4>
- Carskadon, M. A., & Acebo, C. (1993). A Self-Administered Rating Scale for Pubertal Development. *Journal of Adolescent Health*, 14, 190–195.
- Chan, S. R. W. L., & Blackburn, E. H. (2004). Telomeres and telomerase. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences*, 359(1441), 109–122. <https://doi.org/10.1098/rstb.2003.1370>
- Chen, B. H., Marioni, R. E., Colicino, E., Peters, M. J., Ward-Caviness, C. K., Tsai, P.-C., ... Horvath, S. (2016). DNA methylation-based measures of biological age: meta-analysis predicting time to death. *Aging*, 8(9), 1844–1865. <https://doi.org/10.18632/aging.101020>
- Chen, E., Turiano, N. A., Mroczek, D. K., & Miller, G. E. (2016). Association of Reports of Childhood Abuse and All-Cause Mortality Rates in Women. *JAMA Psychiatry*, 73(9), 920. <https://doi.org/10.1001/jamapsychiatry.2016.1786>
- Cisler, J. M., James, G. A., Tripathi, S., Mletzko, T., Heim, C., Hu, X. P., ... Kilts, C. D. (2013). Differential functional connectivity within an emotion regulation neural network among individuals resilient and susceptible to the depressogenic effects of early life stress. *Psychological Medicine*, 43(03), 507–518. <https://doi.org/10.1017/S0033291712001390>
- Cisler, J. M., Scott Steele, J., Smitherman, S., Lenow, J. K., & Kilts, C. D. (2013). Neural processing correlates of assaultive violence exposure and PTSD symptoms during implicit threat processing: A network-level analysis among adolescent girls. *Psychiatry Research: Neuroimaging*, 214(3), 238–246. <https://doi.org/10.1016/j.psychres.2013.06.003>
- Cochran, W. G. (1954). Some Methods for Strengthening the Common χ^2 Tests. *Biometrics*, 10(4), 417. <https://doi.org/10.2307/3001616>
- Cohen, J. (1988). *Statistical Power Analysis for the Behavioral Sciences*. New York, NY: Routledge Academic.
- Coimbra, B. M., Carvalho, C. M., Moretti, P. N., Mello, M. F., & Belangero, S. I. (2017). Stress-related telomere length in children: A systematic review. *Journal of Psychiatric Research*, 92(PG-47-54), 47–54. <https://doi.org/10.1016/j.jpsychires.2017.03.023>
- Cole, M. W., & Schneider, W. (2007). The cognitive control network: Integrated cortical regions with dissociable functions. *NeuroImage*, 37(1), 343–360. <https://doi.org/10.1016/j.neuroimage.2007.03.071>

- Coleman, L., & Coleman, J. (2002). The measurement of puberty: a review. *Journal of Adolescence*, 25, 535–550. <https://doi.org/10.1006/jado.2002.0494>
- Colich, N. L., Platt, J. M., Keyes, K. M., Sumner, J. A., Allen, N. B., & McLaughlin, K. A. (2019). Earlier age at menarche as a transdiagnostic mechanism linking childhood trauma with multiple forms of psychopathology in adolescent girls. *Psychological Medicine*.
- Colich, N. L., Williams, E. S., Ho, T. C., King, L. S., Humphreys, K. L., Price, A. N., ... Gotlib, I. H. (2017). The association between early life stress and prefrontal cortex activation during implicit emotion regulation is moderated by sex in early adolescence. *Development and Psychopathology*, 29(05), 1851–1864. <https://doi.org/10.1017/S0954579417001444>
- Copeland, W. E., Adair, C. E., Smetanin, P., Stiff, D., Briante, C., Colman, I., ... Angold, A. (2013). Diagnostic transitions from childhood to adolescence to early adulthood. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*. <https://doi.org/10.1111/jcpp.12062>
- Corbetta, M., Kincade, J. M., & Shulman, G. L. (2002). Neural Systems for Visual Orienting and Their Relationships to Spatial Working Memory. *Journal of Cognitive Neuroscience*, 14(3), 508–523. <https://doi.org/10.1162/089892902317362029>
- Cousminer, D. L., Widén, E., & Palmert, M. R. (2015). The genetics of pubertal timing in the general population. *Current Opinion in Endocrinology & Diabetes and Obesity*, 23(1), 57–65. <https://doi.org/10.1097/med.0000000000000213>
- Danese, A., & McEwen, B. S. (2012). Adverse childhood experiences, allostasis, allostatic load, and age-related disease. *Physiology and Behavior*, 106(1), 29–39. <https://doi.org/10.1016/j.physbeh.2011.08.019>
- Davis, E. G., Humphreys, K. L., McEwen, L. M., Sacchet, M. D., Camacho, M. C., Maclsaac, J. L., ... Gotlib, I. H. (2017). Accelerated DNA methylation age in adolescent girls: associations with elevated diurnal cortisol and reduced hippocampal volume. *Translational Psychiatry*, 7(8), e1223. <https://doi.org/10.1038/tp.2017.188>
- Day, F. R., Elks, C. E., Murray, A., Ong, K. K., & Perry, J. R. B. (2015). Puberty timing associated with diabetes, cardiovascular disease and also diverse health outcomes in men and women: The UK Biobank study. *Scientific Reports*, 5(June), 1–12. <https://doi.org/10.1038/srep11208>
- de Vries, L., Kauschansky, A., Shohat, M., & Phillip, M. (2004). Familial Central Precocious Puberty Suggests Autosomal Dominant Inheritance. *The Journal of Clinical Endocrinology & Metabolism*, 89(4), 1794–1800. <https://doi.org/10.1210/jc.2003-030361>
- Deardorff, J., Abrams, B., Ekwaru, J. P., & Rehkopf, D. H. (2014). Socioeconomic status and age at menarche: an examination of multiple indicators in an ethnically diverse cohort. *Annals of Epidemiology*, 24(10), 727–733. <https://doi.org/10.1016/j.annepidem.2014.07.002>
- Del Giudice, M., Gangestad, S. W., & Kaplan, H. S. (2016). Life History Theory and Evolutionary Psychology. In M. Buss, David (Ed.), *The Handbook of Evolutionary Psychology, Volume 1: Foundations* (Second, pp. 88–114). Hoboken, NJ: John Wiley & Sons.
- Dennison, M. J., Rosen, M. L., Sambrook, K. A., Jenness, J. L., Sheridan, M. A., & McLaughlin, K. A. (2019). Differential Associations of Distinct Forms of Childhood Adversity With Neurobehavioral Measures of Reward Processing: A Developmental Pathway to Depression. *Child Development*, 90(1), e96–e113. <https://doi.org/10.1111/cdev.13011>
- DiCarlo, J. J., Zoccolan, D., & Rust, N. C. (2012). How Does the Brain Solve Visual Object Recognition? *Neuron*, 73(3), 415–434. <https://doi.org/10.1016/j.neuron.2012.01.010>
- Dixon, M. L., Thiruchselvam, R., Todd, R., & Christoff, K. (2017). Emotion and the prefrontal cortex: An integrative review. *Psychological Bulletin*, 143(10), 1033–1081. <https://doi.org/10.1037/bul0000096>
- Dong, M., Giles, W. H., Felitti, V. J., Dube, S. R., Williams, J. E., Chapman, D. P., & Anda, R. F. (2004). Insights Into Causal Pathways for Ischemic Heart Disease. *Circulation*, 110(13),

- 1761–1766. <https://doi.org/10.1161/01.CIR.0000143074.54995.7F>
- Dorn, L. D., & Biro, F. M. (2011). Puberty and Its Measurement: A Decade in Review. *Journal of Research on Adolescence*, 21(1), 180–195. <https://doi.org/10.1111/j.1532-7795.2010.00722.x>
- Dorn, L. D., Sontag-Padilla, L. M., Pabst, S., Tissot, A., & Susman, E. (2013). Longitudinal reliability of self-reported age at menarche in adolescent girls: variability across time and setting. *Developmental Psychology*, 49(6), 1187–1193. <https://doi.org/10.1037/a0029424>
- Drury, S. S., Mabile, E., Brett, Z. H., Esteves, K., Jones, E., Shirtcliff, E. A., & Theall, K. P. (2014). The Association of Telomere Length With Family Violence and Disruption. *PEDIATRICS*, 134(1), e128–e137. <https://doi.org/10.1542/peds.2013-3415>
- Drury, S. S., Theall, K., Gleason, M. M., Smyke, A. T., De Vivo, I., Wong, J. Y. Y., ... Nelson, C. A. (2012). Telomere length and early severe social deprivation: linking early adversity and cellular aging. *Molecular Psychiatry*, 17(7), 719–727. <https://doi.org/10.1038/mp.2011.53>
- Ducharme, S., Albaugh, M. D., Nguyen, T.-V., Hudziak, J. J., Mateos-Pérez, J. M., Labbe, A., ... Karama, S. (2016). Trajectories of cortical thickness maturation in normal brain development — The importance of quality control procedures. *NeuroImage*, 125, 267–279. <https://doi.org/10.1016/j.neuroimage.2015.10.010>
- Duncan, G. J., & Magnuson, K. (2012). Socioeconomic status and cognitive functioning: Moving from correlation to causation. *Wiley Interdisciplinary Reviews: Cognitive Science*, 3(3), 377–386. <https://doi.org/10.1002/wcs.1176>
- Ellis, B. J., Figueredo, A. J., Brumbach, B. H., & Schlomer, G. L. (2009). *Fundamental dimensions of environmental risk: The impact of harsh versus unpredictable environments on the evolution and development of life history strategies*. *Human Nature* (Vol. 20). <https://doi.org/10.1007/s12110-009-9063-7>
- Ellis, B. J., & Garber, J. (2000). Psychosocial Antecedents of Variation in Girls' Pubertal Timing: Maternal Depression, Stepfather Presence, and Marital and Family Stress. *Child Development*, 71(2), 485–501. <https://doi.org/10.1111/1467-8624.00159>
- Epel, E. S., Blackburn, E. H., Lin, J., Dhabhar, F. S., Adler, N. E., Morrow, J. D., & Cawthon, R. M. (2004). Accelerated telomere shortening in response to life stress. *Proceedings of the National Academy of Sciences*, 101(49), 17312–17315. <https://doi.org/10.1073/pnas.0407162101>
- Essex, M. J., Boyce, W. T., Hertzman, C., Lam, L. L., Armstrong, J. M., Neumann, S. M. A., & Kobor, M. S. (2013). Epigenetic vestiges of early developmental adversity: childhood stress exposure and DNA methylation in adolescence. *Child Development*, 84(1), 58–75. <https://doi.org/10.1111/j.1467-8624.2011.01641.x>
- Evans, G. W., Li, D., & Whipple, S. S. (2013). Cumulative risk and child development. *Psychological Bulletin*, 139(6), 1342–1396. <https://doi.org/10.1037/a0031808>
- Everaerd, D., Klumpers, F., Zwieters, M., Guadalupe, T., Franke, B., van Oostrom, I., ... Tendolkar, I. (2016). Childhood abuse and deprivation are associated with distinct sex-dependent differences in brain morphology. *Neuropsychopharmacology*, 41(7), 1716–1723. <https://doi.org/10.1038/npp.2015.344>
- Felitti, V. J., Anda, R. F., Nordenberg, D., Williamson, D. F., Spitz, A. M., Edwards, V., ... Marks, J. S. (1998). Relationship of Childhood Abuse and Household Dysfunction to Many of the Leading Causes of Death in Adults. *American Journal of Preventive Medicine*, 14(4), 245–258. [https://doi.org/10.1016/S0749-3797\(98\)00017-8](https://doi.org/10.1016/S0749-3797(98)00017-8)
- Foster, H., Brooks-Gunn, J., & Martin, A. (2007). Poverty/Socioeconomic Status and Exposure to Violence in the Lives of Children and Adolescents. In D. J. Flannery, A. T. Vazsonyi, & I. D. Waldman (Eds.), *The Cambridge Handbook of Violent Behavior and Aggression* (pp. 664–687). Cambridge: Cambridge University Press. <https://doi.org/10.1017/CBO9780511816840.036>
- Frenck, R. W., Blackburn, E. H., & Shannon, K. M. (1998). The rate of telomere sequence loss

- in human leukocytes varies with age. *Proceedings of the National Academy of Sciences*, 95(10), 5607–5610. <https://doi.org/10.1073/pnas.95.10.5607>
- G., T., JH., P., M., G., K., D., NE., S., & TK., J. (2009). Early puberty in internationally adopted girls: hormonal and clinical markers of puberty in 276 girls examined biannually over two years. *Hormone Research*, 72(4 PG-236-46), 236–246. <https://doi.org/10.1159/000236085>
- Gabard-Durnam, L. J., Flannery, J., Goff, B., Gee, D. G., Humphreys, K., Telzer, E., ... Tottenham, N. (2014). The Development of Human Amygdala Functional Connectivity at Rest from 4 to 23 years: a cross-sectional study. *Neuroimage*, (95), 193–207. <https://doi.org/10.1016/j.neuroimage.2014.03.038>
- Gassen, N. C., Chrousos, G. P., Binder, E. B., & Zannas, A. S. (2017). Life stress, glucocorticoid signaling, and the aging epigenome: Implications for aging-related diseases. *Neuroscience and Biobehavioral Reviews*, 74(PG-356-365), 356–365. <https://doi.org/10.1016/j.neubiorev.2016.06.003>
- Gee, D. G., Gabard-Durnam, L., Flannery, J., Goff, B., Humphreys, K. L., Telzer, E. H., ... Tottenham, N. (2013). Early developmental emergence of human amygdala-prefrontal connectivity after maternal deprivation. *Proceedings of the National Academy of Sciences*, 110(39), 15638–15643. <https://doi.org/10.1073/pnas.1307893110>
- Gee, D. G., Humphreys, K. L., Flannery, J., Goff, B., Telzer, E. H., Shapiro, M., ... Tottenham, N. (2013). A developmental shift from positive to negative connectivity in human amygdala-prefrontal circuitry. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 33(10), 4584–4593. <https://doi.org/10.1523/JNEUROSCI.3446-12.2013>
- Geronimus, A. T. (1992). The weathering hypothesis and the health of African-American women and infants: evidence and speculations. *Ethnicity & Disease*, 2(3), 207–221.
- Geronimus, A. T., Hicken, M., Keene, D., & Bound, J. (2006). “Weathering” and age patterns of allostatic load scores among blacks and whites in the United States. *American Journal of Public Health*, 96(5), 826–833. <https://doi.org/10.2105/AJPH.2004.060749>
- Gershon, N. B., & High, P. C. (2015). Epigenetics and child abuse: Modern-day darwinism - The miraculous ability of the human genome to adapt, and then adapt again. *American Journal of Medical Genetics Part C: Seminars in Medical Genetics*, 169(4), 353–360. <https://doi.org/10.1002/ajmg.c.31467>
- Giedd, J. N., Blumenthal, J. D., Jeffries, N. O., Castellanos, F. X., Liu, H., Zijdenbos, A., ... Rapoport, J. L. (1999). Brain development during childhood and adolescence : a longitudinal MRI study. *Nature Neuroscience*, 2(10), 861–863.
- Gilkerson, J., Richards, J. A., Warren, S. F., Montgomery, J. K., Greenwood, C. R., Oller, D. K., ... Paul, T. D. (2017). Mapping the early language environment using all-day recordings and automated analysis. *American Journal of Speech-Language Pathology*, 26(2), 248–265. https://doi.org/10.1044/2016_AJSLP-15-0169
- Gold, A. L., Sheridan, M. A., Peverill, M., Busso, D. S., Lambert, H. K., Alves, S., ... McLaughlin, K. A. (2016). Childhood abuse and reduced cortical thickness in brain regions involved in emotional processing. *Journal of Child Psychology and Psychiatry*, 57(10), 1154–1164. <https://doi.org/10.1111/jcpp.12630>
- Golub, M. S., Collman, G. W., Foster, P. M. D., Kimmel, C. A., Rajpert-De Meyts, E., Reiter, E. O., ... Toppari, J. (2008). Public Health Implications of Altered Puberty Timing. *Pediatrics*, 121(Supplement 3), S218–S230. <https://doi.org/10.1542/peds.2007-1813g>
- Gotlib, I. H., Lemoult, J., Colich, N. L., Foland-Ross, L. C., Hallmayer, J., Joormann, J., ... Wolkowitz, O. M. (2015). Telomere length and cortisol reactivity in children of depressed mothers. *Molecular Psychiatry*, 20(5). <https://doi.org/10.1038/mp.2014.119>
- Graber, J., Brooks-Gunn, J., & Warren, M. (1995a). The Antecedents of Menarcheal Age: Heredity, Family Environment, and Stressful Life Events. *Child Development*, 66(2), 346. <https://doi.org/10.2307/1131582>
- Graber, J., Brooks-Gunn, J., & Warren, M. P. (1995b). The antecedents of menarcheal age:

- heredity, family environment, and stressful life events. *Child Development*, 66(2), 346–359. <https://doi.org/10.2307/1131582>
- Green, J. G., McLaughlin, K. A., Berglund, P. A., Gruber, M. J., Sampson, N. a, Zaslavsky, A. M., & Kessler, R. C. (2010). Childhood Adversities and Adult Psychiatric Disorders in the National Comorbidity Survey Replication I. *Archives of General Psychiatry*, 67(2), 113. <https://doi.org/10.1001/archgenpsychiatry.2009.186>
- Halpern, C. T., Tucker, C. M., Bengtson, A., Kupper, L. L., McLean, S. A., & Martin, S. L. (2013). Somatic Symptoms Among US Adolescent Females: Associations with Sexual and Physical Violence Exposure. *Maternal and Child Health Journal*, 17(10), 1951–1960. <https://doi.org/10.1007/s10995-013-1221-1>
- Hamilton, J. L., Hamlat, E. J., Stange, J. P., Abramson, L. Y., & Alloy, L. B. (2014). Pubertal timing and vulnerabilities to depression in early adolescence: differential pathways to depressive symptoms by sex. *Journal of Adolescence*, 37(2), 165–174. <https://doi.org/10.1016/j.adolescence.2013.11.010>
- Hannum, G., Guinney, J., Zhao, L., Zhang, L., Hughes, G., Sada, S., ... Zhang, K. (2013). Genome-wide methylation profiles reveal quantitative views of human aging rates. *Molecular Cell*, 49(2), 359–367. <https://doi.org/10.1016/j.molcel.2012.10.016>
- Hanssen, L. M., Schutte, N. S., Malouff, J. M., & Epel, E. S. (2017). The relationship between childhood psychosocial stressor level and telomere length: a meta-analysis. *Health Psychology Research*, 5(1), 6378. <https://doi.org/10.4081/hpr.2017.6378>
- Harden, K. P. (2014). Genetic influences on adolescent sexual behavior: Why genes matter for environmentally oriented researchers. *Psychological Bulletin*, 140(2), 434–465. <https://doi.org/10.1037/a0033564>
- Harden, K. P., & Mendle, J. (2012). Gene-environment interplay in the association between pubertal timing and delinquency in adolescent girls. *Journal of Abnormal Psychology*, 121(1), 73–87. <https://doi.org/10.1037/a0024160>
- Hardt, J., & Rutter, M. (2004). Validity of adult retrospective reports of adverse childhood experiences: review of the evidence. *Journal of Child Psychology and Psychiatry*, 45(2), 260–273. <https://doi.org/10.1111/j.1469-7610.2004.00218.x>
- Hartman, S., Li, Z., Nettle, D., & Belsky, J. (2017). External-environmental and internal-health early life predictors of adolescent development. *Development and Psychopathology*, 29(05), 1839–1849. <https://doi.org/10.1017/S0954579417001432>
- Hayes, P., & Tan, T. X. (2016). Timing of menarche in girls adopted from China: a cohort study. *Child Care Health and Development*, 42(6 PG-859-862), 859–862. <https://doi.org/10.1111/cch.12393>
- Heim, C., & Binder, E. B. (2012). Current research trends in early life stress and depression: Review of human studies on sensitive periods, gene-environment interactions, and epigenetics. *Experimental Neurology*, 233(1 PG-102-111), 102–111. <https://doi.org/10.1016/j.expneurol.2011.10.032>
- Hein, T. C., & Monk, C. S. (2017). Research Review: Neural response to threat in children, adolescents, and adults after child maltreatment ??? a quantitative meta-analysis. *Journal of Child Psychology and Psychiatry and Allied Disciplines*, 58(3), 222–230. <https://doi.org/10.1111/jcpp.12651>
- Higgins, J. P. T., & Thompson, S. G. (2002). Quantifying heterogeneity in a meta-analysis. *Statistics in Medicine*, 21(11), 1539–1558. <https://doi.org/10.1002/sim.1186>
- Hill, K., & Kaplan, H. (1999). Life History Traits in Humans: Theory and Empirical Studies. *Annual Review of Anthropology*, 28(1), 397–430. <https://doi.org/10.1146/annurev.anthro.28.1.397>
- Hodel, A. S., Hunt, R. H., Cowell, R. A., Van Den Heuvel, S. E., Gunnar, M. R., & Thomas, K. M. (2015). Duration of early adversity and structural brain development in post-institutionalized adolescents. *NeuroImage*, 105(PG-112-9), 112–119.

- <https://doi.org/10.1016/j.neuroimage.2014.10.020>
- Hoehn, P. W., Rosmalen, J. G. M., Schoevers, R. A., Huzen, J., van der Harst, P., & de Jonge, P. (2013). Association between anxiety but not depressive disorders and leukocyte telomere length after 2 years of follow-up in a population-based sample. *Psychological Medicine*, 43(04), 689–697. <https://doi.org/10.1017/S0033291712001766>
- Horvath, S. (2013). DNA methylation age of human tissues and cell types. *Genome Biology*, 14(10), R115. <https://doi.org/10.1186/s13059-015-0649-6>
- Horvath, S., Erhart, W., Brosch, M., Ammerpohl, O., von Schonfels, W., Ahrens, M., ... Hampe, J. (2014). Obesity accelerates epigenetic aging of human liver. *Proceedings of the National Academy of Sciences*, 111(43), 15538–15543. <https://doi.org/10.1073/pnas.1412759111>
- Horvath, S., & Raj, K. (2018). DNA methylation-based biomarkers and the epigenetic clock theory of ageing. *Nature Reviews Genetics*. <https://doi.org/10.1038/s41576-018-0004-3>
- Humphreys, K. L., Esteves, K., Zeanah, C. H., Fox, N. A., Nelson, C. A., & Drury, S. S. (2016). Accelerated telomere shortening: Tracking the lasting impact of early institutional care at the cellular level. *Psychiatry Research*, 246(PG-95-100), 95–100. <https://doi.org/10.1016/j.psychres.2016.09.023>
- Humphreys, K. L., & Zeanah, C. H. (2015). Deviations from the Expectable Environment in Early Childhood and Emerging Psychopathology. *Neuropsychopharmacology*, 40(1), 154–170. <https://doi.org/10.1038/npp.2014.165>
- Huttenlocher, P. R. (1979). Synaptic density in human frontal cortex — Developmental changes and effects of aging. *Brain Research*, 163, 195–205. [https://doi.org/10.1016/0006-8993\(79\)90349-4](https://doi.org/10.1016/0006-8993(79)90349-4)
- Jaffee, S. R., Mcfarquhar, T., Stevens, S., Ouellet-Morin, I., Melhuish, E., & Belsky, J. (2014). Interactive effects of early and recent exposure to stressful contexts on cortisol reactivity in middle childhood. *Journal of Child Psychology and Psychiatry and Allied Disciplines*, 2, 138–146. <https://doi.org/10.1111/jcpp.12287>
- Jalbrzikowski, M., Larsen, B., Hallquist, M. N., Foran, W., Calabro, F., & Luna, B. (2017). Development of White Matter Microstructure and Intrinsic Functional Connectivity Between the Amygdala and Ventromedial Prefrontal Cortex: Associations With Anxiety and Depression. *Biological Psychiatry*, 82(7), 511–521. <https://doi.org/10.1016/j.biopsych.2017.01.008>
- Jednoróg, K., Altarelli, I., Monzalvo, K., Fluss, J., Dubois, J., Billard, C., ... Ramus, F. (2012). The Influence of Socioeconomic Status on Children's Brain Structure. *PLoS ONE*, 7(8), e42486. <https://doi.org/10.1371/journal.pone.0042486>
- Johnson, D. E., Tang, A., Almas, A. N., Degnan, K. A., McLaughlin, K. A., Nelson, C. A., ... Drury, S. S. (2018). Caregiving Disruptions Affect Growth and Pubertal Development in Early Adolescence in Institutionalized and Fostered Romanian Children: A Randomized Clinical Trial. *Journal of Pediatrics*, 203, 345–353.e3. <https://doi.org/10.1016/j.jpeds.2018.07.027>
- Jovanovic, T., Vance, A., Cross, D., Knight, A., Kilaru, V., Michopoulos, V., ... Smith, A. (2017a). Experiencing Violence Accelerates Epigenetic Aging in Children. *Biological Psychiatry*, 81(10 PG-S24-S24), S24–S24.
- Jovanovic, T., Vance, L. A., Cross, D., Knight, A. K., Kilaru, V., Michopoulos, V., ... Smith, A. K. (2017b). Exposure to Violence Accelerates Epigenetic Aging in Children. *Scientific Reports*, 7(1), 8962. <https://doi.org/10.1038/s41598-017-09235-9>
- Keding, T. J., & Heringa, R. J. (2016). Paradoxical Prefrontal–Amygdala Recruitment to Angry and Happy Expressions in Pediatric Posttraumatic Stress Disorder. *Neuropsychopharmacology*, 41(12), 2903–2912. <https://doi.org/10.1038/npp.2016.104>
- Kelly, P. A., Viding, E., Wallace, G. L., Schaer, M., De Brito, S. A., Robustelli, B., & McCrory, E. J. (2013). Cortical Thickness, Surface Area, and Gyrfication Abnormalities in Children Exposed to Maltreatment: Neural Markers of Vulnerability? *Biological Psychiatry*, 74(11),

- 845–852. <https://doi.org/10.1016/j.biopsycho.2013.06.020>
- Kessler, R. C., McLaughlin, K. A., Green, J. G., Gruber, M. J., Sampson, N. A., Zaslavsky, A. M., ... Williams, D. R. (2010). Childhood adversities and adult psychopathology in the WHO world mental health surveys. *British Journal of Psychiatry*, *197*(5), 378–385. <https://doi.org/10.1192/bjp.bp.110.080499>
- King, L. S., Colich, N. L., LeMoult, J., Humphreys, K. L., Ordaz, S. J., Price, A. N., & Gotlib, I. H. (2017). The impact of the severity of early life stress on diurnal cortisol: The role of puberty. *Psychoneuroendocrinology*, *77*, 68–74. <https://doi.org/10.1016/j.psyneuen.2016.11.024>
- Konstantopoulos, S. (2011). Fixed effects and variance components estimation in three-level meta-analysis. *Research Synthesis Methods*, *2*(1), 61–76. <https://doi.org/10.1002/jrsm.35>
- Kujawa, A., Wu, M., Klumpp, H., Pine, D. S., Swain, J. E., Fitzgerald, K. D., ... Phan, K. L. (2016). Altered Development of Amygdala-Anterior Cingulate Cortex Connectivity in Anxious Youth and Young Adults. *Biological Psychiatry: Cognitive Neuroscience and Neuroimaging*, *1*(4), 345–352. <https://doi.org/10.1016/j.bpsc.2016.01.006>
- Lakshman, R., Forouhi, N. G., Sharp, S. J., Luben, R., Bingham, S. A., Khaw, K. T., ... Ong, K. K. (2009). Early age at menarche associated with cardiovascular disease and mortality. *Journal of Clinical Endocrinology and Metabolism*, *94*(12), 4953–4960. <https://doi.org/10.1210/jc.2009-1789>
- Lambert, H. K., King, K. M., Monahan, K. C., & McLaughlin, K. A. (2017). Differential associations of threat and deprivation with emotion regulation and cognitive control in adolescence. *Development and Psychopathology*, *29*(03), 929–940. <https://doi.org/10.1017/S0954579416000584>
- Lang, J., McKie, J., Smith, H., McLaughlin, A., Gillberg, C., Shiels, P. G., & Minnis, H. (2019). Adverse childhood experiences, epigenetics and telomere length variation in childhood and beyond: a systematic review of the literature. *European Child & Adolescent Psychiatry*. <https://doi.org/10.1007/s00787-019-01329-1>
- Lawson, G. M., Duda, J. T., Avants, B. B., Wu, J., & Farah, M. J. (2013). Associations between children's socioeconomic status and prefrontal cortical thickness. *Developmental Science*, *16*(5), 641–652. <https://doi.org/10.1111/desc.12096>
- Lenroot, R. K. R., Gogtay, N., Greenstein, D. D. K., Wells, E. M., Wallace, G. L., Clasen, L. S., ... Giedd, J. N. (2007). Sexual dimorphism of brain developmental trajectories during childhood and adolescence. *Neuroimage*, *36*(4), 1065–1073. <https://doi.org/10.1016/j.neuroimage.2007.03.053>
- LeWinn, K. Z., Sheridan, M. A., Keyes, K. M., Hamilton, A., & McLaughlin, K. A. (2017). Sample composition alters associations between age and brain structure. *Nature Communications*, *8*(1), 874. <https://doi.org/10.1038/s41467-017-00908-7>
- Lewis, C. R., & Olive, M. F. (2014). Early-life stress interactions with the epigenome: potential mechanisms driving vulnerability toward psychiatric illness. *Behavioural Pharmacology*, *25*(5-6 PG-341–351), 341–351. <https://doi.org/10.1097/fbp.0000000000000057>
- Li, X., Wang, J., Zhou, J., Huang, P., & Li, J. (2017). The association between post-traumatic stress disorder and shorter telomere length: A systematic review and meta-analysis. *Journal of Affective Disorders*, *218*(October 2016), 322–326. <https://doi.org/10.1016/j.jad.2017.03.048>
- Lindqvist, D., Epel, E. S., Mellon, S. H., Penninx, B. W., Révész, D., Verhoeven, J. E., ... Wolkowitz, O. M. (2015). Psychiatric disorders and leukocyte telomere length: Underlying mechanisms linking mental illness with cellular aging. *Neuroscience and Biobehavioral Reviews*. Elsevier Ltd. <https://doi.org/10.1016/j.neubiorev.2015.05.007>
- Mackey, A. P., Finn, A. S., Leonard, J. A., Jacoby-Senghor, D. S., West, M. R., Gabrieli, C. F. O., & Gabrieli, J. D. E. (2015). Neuroanatomical Correlates of the Income-Achievement Gap. *Psychological Science*, *26*(6), 925–933. <https://doi.org/10.1177/0956797615572233>
- Malouff, J. M., & Schutte, N. S. (2017). A meta-analysis of the relationship between anxiety and

- telomere length. *Anxiety, Stress, & Coping*, 30(3), 264–272.
<https://doi.org/10.1080/10615806.2016.1261286>
- Marioni, R. E., Shah, S., McRae, A. F., Chen, B. H., Colicino, E., Harris, S. E., ... Deary, I. J. (2015). DNA methylation age of blood predicts all-cause mortality in later life. *Genome Biology*, 16(1), 25. <https://doi.org/10.1186/s13059-015-0584-6>
- Marioni, R. E., Shah, S., McRae, A. F., Ritchie, S. J., Muniz-Terrera, G., Harris, S. E., ... Deary, I. J. (2015). The epigenetic clock is correlated with physical and cognitive fitness in the Lothian Birth Cohort 1936. *International Journal of Epidemiology*, 44(4), 1388–1396.
<https://doi.org/10.1093/ije/dyu277>
- Marshall, W., & Tanner, J. (1969). Variations in the Pattern of Pubertal Changes in Girls. *Archives of Disease in Childhood*, 44(291), 291–303. <https://doi.org/10.1136/adc.45.239.13>
- Marshall, W., & Tanner, J. (1970). Variations in the Pattern of Pubertal Changes in Boys. *Archives of Disease in Childhood*, 45(239), 13–23. <https://doi.org/10.1136/adc.45.239.13>
- Marusak, H. A., Martin, K. R., Etkin, A., & Thomason, M. E. (2015). Childhood Trauma Exposure Disrupts the Automatic Regulation of Emotional Processing. *Neuropsychopharmacology*, 40(5), 1250–1258. <https://doi.org/10.1038/npp.2014.311>
- McCrary, E. J., De Brito, S. A., & Viding, E. (2011). The impact of childhood maltreatment: A review of neurobiological and genetic factors. *Frontiers in Psychiatry*, 2(JUL), 1–14.
<https://doi.org/10.3389/fpsy.2011.00048>
- McEwen, B. S. (1998). Stress, Adaptation, and Disease: Allostasis and Allostatic Load. *Annals of the New York Academy of Sciences*, 840(1), 33–44. <https://doi.org/10.1111/j.1749-6632.1998.tb09546.x>
- McEwen, B. S., & Stellar, E. (1993). Stress and the Individual. *Archives of Internal Medicine*, 153(18), 2093. <https://doi.org/10.1001/archinte.1993.00410180039004>
- McLaughlin, K. A. (2016). Future Directions in Childhood Adversity and Youth Psychopathology. *Journal of Clinical Child and Adolescent Psychology*, 45(3), 361–382.
<https://doi.org/10.1080/15374416.2015.1110823>
- McLaughlin, K. A., Basu, A., Walsh, K., Slopen, N., Sumner, J. A., Koenen, K. C., & Keyes, K. M. (2016). Childhood Exposure to Violence and Chronic Physical Conditions in a National Sample of US Adolescents. *Psychosomatic Medicine*, 78(9), 1072–1083.
<https://doi.org/10.1097/PSY.0000000000000366>
- McLaughlin, K. A., Green, J. G., Gruber, M. J., Sampson, N. A., Zaslavsky, A. M., & Kessler, R. C. (2010). Childhood Adversities and Adult Psychiatric Disorders in the National Comorbidity Survey Replication II. *Archives of General Psychiatry*, 67(2), 124.
<https://doi.org/10.1001/archgenpsychiatry.2009.186>
- McLaughlin, K. A., Green, J. G., Gruber, M. J., Sampson, N. A., Zaslavsky, A. M., & Kessler, R. C. (2012). Childhood Adversities and First Onset of Psychiatric Disorders in a National Sample of US Adolescents. *Archives of General Psychiatry*, 69(11), 1151.
<https://doi.org/10.1001/archgenpsychiatry.2011.2277>
- McLaughlin, K. A., Peverill, M., Gold, A. L., Alves, S., & Sheridan, M. A. (2015). Child Maltreatment and Neural Systems Underlying Emotion Regulation. *Journal of the American Academy of Child & Adolescent Psychiatry*, 54(9), 753–762.
<https://doi.org/10.1016/j.jaac.2015.06.010>
- McLaughlin, K. A., Sheridan, M. A., Gold, A. L., Duys, A., Lambert, H. K., Peverill, M., ... Pine, D. S. (2016). Maltreatment Exposure, Brain Structure, and Fear Conditioning in Children and Adolescents. *Neuropsychopharmacology*, 41(8), 1956–1964.
<https://doi.org/10.1038/npp.2015.365>
- McLaughlin, K. A., Sheridan, M. A., Lambert, H. K., KA., M., MA., S., & HK., L. (2014). Childhood adversity and neural development: Deprivation and threat as distinct dimensions of early experience. *Neuroscience and Biobehavioral Reviews*, 47(PG-578-91), 578–591.
<https://doi.org/10.1016/j.neubiorev.2014.10.012>

- McLaughlin, K. A., Sheridan, M. A., & Nelson, C. A. (2017). Neglect as a Violation of Species-Expectant Experience: Neurodevelopmental Consequences. *Biological Psychiatry*, *82*(7), 462–471. <https://doi.org/10.1016/j.biopsych.2017.02.1096>
- McLaughlin, K. A., Sheridan, M. A., Winter, W., Fox, N. A., Zeanah, C. H., & Nelson, C. A. (2014). Widespread Reductions in Cortical Thickness Following Severe Early-Life Deprivation: A Neurodevelopmental Pathway to Attention-Deficit/Hyperactivity Disorder. *Biological Psychiatry*, *76*(8), 629–638. <https://doi.org/10.1016/j.biopsych.2013.08.016>
- ME., E.-L., MD., S., & IH., G. (2016). The application of neuroimaging to social inequity and language disparity: A cautionary examination. *Developmental Cognitive Neuroscience*, *22*(PG-1-8), 1–8. <https://doi.org/10.1016/j.dcn.2016.10.001>
- Mendle, J., Harden, K. P., Brooks-Gunn, J., & Graber, J. (2010). Development's tortoise and hare: pubertal timing, pubertal tempo, and depressive symptoms in boys and girls. *Developmental Psychology*, *46*(5), 1341–1353. <https://doi.org/10.1037/a0020205>
- Mendle, J., Leve, L. D., Van Ryzin, M., & Natsuaki, M. N. (2014). Linking Childhood Maltreatment With Girls' Internalizing Symptoms: Early Puberty as a Tipping Point. *Journal of Research on Adolescence*, *24*(4), 689–702. <https://doi.org/10.1111/jora.12075>
- Mendle, J., Leve, L. D., Van Ryzin, M., Natsuaki, M. N., & Ge, X. (2011). Associations Between Early Life Stress, Child Maltreatment, and Pubertal Development Among Girls in Foster Care. *Journal of Research on Adolescence*, *21*(4), 871–880. <https://doi.org/10.1111/j.1532-7795.2011.00746.x>
- Mendle, J., Ryan, R. M., & McKone, K. M. (2016). Early Childhood Maltreatment and Pubertal Development: Replication in a Population-Based Sample. *Journal of Research on Adolescence*, *26*(3), 595–602. <https://doi.org/10.1111/jora.12201>
- Mensah, F. K., Bayer, J. K., Wake, M., Carlin, J. B., Allen, N. B., & Patton, G. C. (2013). Early puberty and childhood social and behavioral adjustment. *Journal of Adolescent Health*, *53*(1), 118–124. <https://doi.org/10.1016/j.jadohealth.2012.12.018>
- Milad, M. R., & Quirk, G. J. (2012). Fear Extinction as a Model for Translational Neuroscience: Ten Years of Progress. *Annual Review of Psychology*, *63*(1), 129–151. <https://doi.org/10.1146/annurev.psych.121208.131631>
- Mills, K. L., Goddings, A.-L., Herting, M. M., Meuwese, R., Blakemore, S.-J., Crone, E. A., ... Tamnes, C. K. (2016). Structural brain development between childhood and adulthood: Convergence across four longitudinal samples. *NeuroImage*, (July). <https://doi.org/10.1016/j.neuroimage.2016.07.044>
- Mitchell, C., Hobcraft, J., McLanahan, S. S., Siegel, S. R., Berg, A., Brooks-Gunn, J., ... Notterman, D. (2014). Social disadvantage, genetic sensitivity, and children's telomere length. *Proceedings of the National Academy of Sciences of the United States of America*, *111*(16 PG-5944-5949), 5944–5949. <https://doi.org/10.1073/pnas.1404293111>
- Moher, D., Liberati, A., Tetzlaff, J., & Altman, D. G. (2009). Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. *Journal of Clinical Epidemiology*, *62*(10), 1006–1012. <https://doi.org/10.1016/j.jclinepi.2009.06.005>
- Natsuaki, M. N., Leve, L. D., & Mendle, J. (2011). Going through the rites of passage: timing and transition of menarche, childhood sexual abuse, and anxiety symptoms in girls. *Journal of Youth and Adolescence*, *40*(10), 1357–1370. <https://doi.org/10.1007/s10964-010-9622-6>
- Natu, V., Gomez, J., Barnett, M., Jeska, B., Zhen, Z., Kirilina, E., ... Grill-Spector, K. (2018). Gray Matter Thinning in Ventral Temporal Cortex from Childhood to Adulthood is Associated with Increased Myelination. *Journal of Vision*, *18*(10), 542. <https://doi.org/10.1167/18.10.542>
- Needham, B. L., Mezuk, B., Bareis, N., Lin, J., Blackburn, E. H., & Epel, E. S. (2015). Depression, anxiety and telomere length in young adults: Evidence from the National Health and Nutrition Examination Survey. *Molecular Psychiatry*, *20*(4), 520–528. <https://doi.org/10.1038/mp.2014.89>

- Needham, B. L., Rehkopf, D., Adler, N., Gregorich, S., Lin, J., Blackburn, E. H., & Epel, E. S. (2015). Leukocyte Telomere Length and Mortality in the National Health and Nutrition Examination Survey, 1999–2002. *Epidemiology*, *26*(4), 528–535. <https://doi.org/10.1097/EDE.0000000000000299>
- Negriff, S., Blankson, A. N., & Trickett, P. K. (2015). Pubertal Timing and Tempo: Associations With Childhood Maltreatment. *Journal of Research on Adolescence*, *25*(2), 201–213. <https://doi.org/10.1111/jora.12128>
- Negriff, S., Saxbe, D. E., & Trickett, P. K. (2015). Childhood maltreatment, pubertal development, HPA axis functioning, and psychosocial outcomes: An integrative biopsychosocial model. *Developmental Psychobiology*, *57*(8), 984–993. <https://doi.org/10.1002/dev.21340>
- Negriff, S., & Susman, E. J. (2011). Pubertal timing, depression, and externalizing problems: A framework, review, and examination of gender differences. *Journal of Research on Adolescence*, *21*(3), 717–746. <https://doi.org/10.1111/j.1532-7795.2010.00708.x>
- Negriff, S., & Trickett, P. K. (2012). Peer substance use as a mediator between early pubertal timing and adolescent substance use: Longitudinal associations and moderating effect of maltreatment. *Drug and Alcohol Dependence*, *126*(1–2), 95–101. <https://doi.org/10.1016/j.drugalcdep.2012.04.018>
- Nettle, D., Frankenhuys, W. E., & Rickard, I. J. (2013). The evolution of predictive adaptive responses in human life history. *Proceedings Of The Royal Society Of London Series B-Biological Sciences*, *280*(1766), 20131343.
- Noble, K. G., Houston, S. M., Brito, N. H., Bartsch, H., Kan, E., Kuperman, J. M., ... Sowell, E. R. (2015). Family income, parental education and brain structure in children and adolescents. *Nature Neuroscience*, *18*(5), 773–778. <https://doi.org/10.1038/nn.3983>
- Noble, K. G., McCandliss, B. D., & Farah, M. J. (2007). Socioeconomic gradients predict individual differences in neurocognitive abilities. *Developmental Science*, *10*(4), 464–480. <https://doi.org/10.1111/j.1467-7687.2007.00600.x>
- Noll, J. G., Trickett, P. K., Long, J. D., Negriff, S., Susman, E. J., Shalev, I., ... Putnam, F. W. (2017). Childhood Sexual Abuse and Early Timing of Puberty. *Journal of Adolescent Health*, *60*(1), 65–71. <https://doi.org/10.1016/j.jadohealth.2016.09.008>
- Norman, R. E., Byambaa, M., De, R., Butchart, A., Scott, J., & Vos, T. (2012). The Long-Term Health Consequences of Child Physical Abuse, Emotional Abuse, and Neglect: A Systematic Review and Meta-Analysis. *PLoS Medicine*, *9*(11). <https://doi.org/10.1371/journal.pmed.1001349>
- Parent, A.-S., Teilmann, G., Juul, A., Skakkebaek, N. E., Toppari, J., & Bourguignon, J.-P. (2003). The timing of normal puberty and the age limits of sexual precocity: variations around the world, secular trends, and changes after migration. *Endocrine Reviews*, *24*(5), 668–693.
- Perna, L., Zhang, Y., Mons, U., Holleczeck, B., Saum, K.-U., & Brenner, H. (2016). Epigenetic age acceleration predicts cancer, cardiovascular, and all-cause mortality in a German case cohort. *Clinical Epigenetics*, *8*(1), 64. <https://doi.org/10.1186/s13148-016-0228-z>
- Petanjek, Z., Judaš, M., Šimić, G., Rašin, M. R., Uylings, H. B. M., Rakic, P., & Kostović, I. (2011). Extraordinary neoteny of synaptic spines in the human prefrontal cortex. *Proceedings of the National Academy of Sciences*, *108*(32), 13281–13286. <https://doi.org/10.1073/pnas.1105108108>
- Phelps, E. A., & LeDoux, J. E. (2005). Contributions of the amygdala to emotion processing: From animal models to human behavior. *Neuron*. <https://doi.org/10.1016/j.neuron.2005.09.025>
- Piccolo, L. R., Merz, E. C., He, X., Sowell, E. R., & Noble, K. G. (2016). Age-related differences in cortical thickness vary by socioeconomic status. *PLoS ONE*, *11*(9), 1–18. <https://doi.org/10.1371/journal.pone.0162511>

- Prebeg, Z., & Bralic, I. (2000). Changes in menarcheal age in girls exposed to war conditions. *American Journal of Human Biology*, 12(4), 503–508. [https://doi.org/10.1002/1520-6300\(200007/08\)12:4<503::AID-AJHB10>3.0.CO;2-H](https://doi.org/10.1002/1520-6300(200007/08)12:4<503::AID-AJHB10>3.0.CO;2-H)
- Price, L. H., Kao, H.-T., Burgers, D. E., Carpenter, L. L., & Tyrka, A. R. (2013). Telomeres and Early-Life Stress: An Overview. *Biological Psychiatry*, 73(1), 15–23. <https://doi.org/10.1016/j.biopsych.2012.06.025>
- Rakic, P., Bourgeois, J. P., Eckenhoff, M. F., Zecevic, N., & Goldman-Rakic, P. S. (1986). Concurrent overproduction of synapses in diverse regions of the primate cerebral cortex. *Science*, 232(4747), 232–235. <https://doi.org/10.1126/science.3952506>
- Rehkopf, D. H., Needham, B. L., Lin, J., Blackburn, E. H., Zota, A. R., Wojcicki, J. M., & Epel, E. S. (2016). Leukocyte Telomere Length in Relation to 17 Biomarkers of Cardiovascular Disease Risk: A Cross-Sectional Study of US Adults. *PLoS Medicine*, 13(11). <https://doi.org/10.1371/journal.pmed.1002188>
- Reid, B. M., Miller, B. S., Dorn, L. D., Desjardins, C., Donzella, B., & Gunnar, M. (2017). Early growth faltering in post-institutionalized youth and later anthropometric and pubertal development. *Pediatric Research*, 82(2), 278–284. <https://doi.org/10.1038/pr.2017.35>
- Rickard, I. J., Frankenhuis, W. E., & Nettle, D. (2014). Why Are Childhood Family Factors Associated With Timing of Maturation? A Role for Internal Prediction. *Perspectives on Psychological Science*, 9(1), 3–15. <https://doi.org/10.1177/1745691613513467>
- Ridout, K. K., Ridout, S. J., Price, L. H., Sen, S., & Tyrka, A. R. (2016). Depression and telomere length: A meta-analysis. *Journal of Affective Disorders*, 191(5), 237–247. <https://doi.org/10.1016/j.jad.2015.11.052>
- Rogol, A. D., Clark, P. A., & Roemmich, J. N. (2000). Growth and pubertal development in children and adolescents : effects of diet and physical activity 1 – 4. *The American Journal for Clinical Nutrition*, 72(June), 521S–528S. <https://doi.org/10.1093/ajcn/72.2.521S>
- Rosen, M. L., Hagen, M., Miles, Z., Sheridan, M. A., Meltzoff, A. N., & McLaughlin, K. A. (n.d.). Socioeconomic status and executive function: The role of cognitive stimulation in the early home environment.
- Rosen, M. L., Sheridan, M. A., Sambrook, K. A., Meltzoff, A. N., & McLaughlin, K. A. (2018). Socioeconomic disparities in academic achievement: A multi-modal investigation of neural mechanisms in children and adolescents. *NeuroImage*, 173(February), 298–310. <https://doi.org/10.1016/j.neuroimage.2018.02.043>
- Rowe, D. C. (2002). On genetic variation in menarche and age at first sexual intercourse. A critique of the Belsky-Draper hypothesis. *Evolution and Human Behavior*, 23(5), 365–372. [https://doi.org/10.1016/S1090-5138\(02\)00102-2](https://doi.org/10.1016/S1090-5138(02)00102-2)
- Ryan, R. M., Mendle, J., & Markowitz, A. J. (2015). Early Childhood Maltreatment and Girls' Sexual Behavior: The Mediating Role of Pubertal Timing. *Journal of Adolescent Health*, 57(3), 342–347. <https://doi.org/10.1016/j.jadohealth.2015.06.005>
- Sánchez-Garrido, M. A., Castellano, J. M., Ruiz-Pino, F., Garcia-Galiano, D., Manfredi-Lozano, M., Leon, S., ... Tena-Sempere, M. (2013). Metabolic Programming of Puberty: Sexually Dimorphic Responses to Early Nutritional Challenges. *Endocrinology*, 154(9), 3387–3400. <https://doi.org/10.1210/en.2012-2157>
- Saxbe, D. E., Negri, S., Susman, E. J., & Trickett, P. K. (2015). Attenuated hypothalamic–pituitary–adrenal axis functioning predicts accelerated pubertal development in girls 1 year later. *Development and Psychopathology*, 27(03), 819–828. <https://doi.org/10.1017/S0954579414000790>
- Scheuer, S., Wiggert, N., Brückl, T. M., Awaloff, Y., Uhr, M., Lucae, S., ... Wilhelm, F. H. (2018). Childhood abuse and depression in adulthood: The mediating role of allostatic load. *Psychoneuroendocrinology*, 94(April), 134–142. <https://doi.org/10.1016/j.psyneuen.2018.04.020>
- Scott, K. M., Von Korff, M., Angermeyer, M. C., Benjet, C., Bruffaerts, R., de Girolamo, G., ...

- Kessler, R. C. (2011). Association of childhood adversities and early-onset mental disorders with adult-onset chronic physical conditions. *Archives of General Psychiatry*, 68(8), 838–844. <https://doi.org/10.1001/archgenpsychiatry.2011.77>
- Shalev, I. (2012). Early life stress and telomere length: Investigating the connection and possible mechanisms. *BioEssays*, 34(11), 943–952. <https://doi.org/10.1002/bies.201200084>
- Shalev, I., & Belsky, J. (2016). Early-life stress and reproductive cost: A two-hit developmental model of accelerated aging? *Medical Hypotheses*, 90, 41–47. <https://doi.org/10.1016/j.mehy.2016.03.002>
- Shalev, I., Moffitt, T. E., Sugden, K., Williams, B., Houts, R. M., Danese, A., ... Caspi, A. (2013). Exposure to violence during childhood is associated with telomere erosion from 5 to 10 years of age: a longitudinal study. *Molecular Psychiatry*, 18(5), 576–581. <https://doi.org/10.1038/mp.2012.32>
- Shaw, P., Greenstein, D., Lerch, J., Clasen, L., Lenroot, R., Gogtay, N., ... Giedd, J. (2006). Intellectual ability and cortical development in children and adolescents. *Nature*, 440(7084), 676–679. <https://doi.org/10.1038/nature04513>
- Sheridan, M. A., & McLaughlin, K. A. (2014). Dimensions of early experience and neural development: deprivation and threat. *Trends in Cognitive Sciences*, 18(11), 580–585. <https://doi.org/10.1016/j.tics.2014.09.001>
- Sheridan, M. A., Peverill, M., Finn, A. S., & McLaughlin, K. A. (2017). Dimensions of childhood adversity have distinct associations with neural systems underlying executive functioning. *Development and Psychopathology*, 29(05), 1777–1794. <https://doi.org/10.1017/S0954579417001390>
- Silberman, D. M., Acosta, G. B., & Zorrilla Zubilete, M. A. (2016). Long-term effects of early life stress exposure: Role of epigenetic mechanisms. *Pharmacological Research*, 109(PG-64-73), 64–73. <https://doi.org/10.1016/j.phrs.2015.12.033>
- Silvers, J. A., Insel, C., Powers, A., Franz, P., Helion, C., Martin, R. E., ... Ochsner, K. N. (2016a). vIPFC–vmPFC–Amygdala Interactions Underlie Age-Related Differences in Cognitive Regulation of Emotion. *Cerebral Cortex*, bhw073. <https://doi.org/10.1093/cercor/bhw073>
- Silvers, J. A., Insel, C., Powers, A., Franz, P., Helion, C., Martin, R., ... Ochsner, K. N. (2016b). The transition from childhood to adolescence is marked by a general decrease in amygdala reactivity and an affect-specific ventral-to-dorsal shift in medial prefrontal recruitment. *Developmental Cognitive Neuroscience*. <https://doi.org/10.1016/j.dcn.2016.06.005>
- Silvers, J. A., Lumian, D. S., Gabard-Durnam, L., Gee, D. G., Goff, B., Fareri, D. S., ... Tottenham, N. (2016). Previous institutionalization is followed by broader amygdala–hippocampal–PFC network connectivity during aversive learning in human development. *The Journal of Neuroscience*, 36(24), 6420–6430. <https://doi.org/10.1523/JNEUROSCI.0038-16.2016>
- Slopen, N., Tang, A., Nelson, C. A., Zeanah, C. H., McDade, T. W., McLaughlin, K. A., & Fox, N. (2019). The consequences of foster care versus institutional care in early childhood on adolescent cardiometabolic and immune markers. *Psychosomatic Medicine*. <https://doi.org/10.1097/PSY.0000000000000696>
- Sonuga-Barke, E. J., Schlotz, W., & Rutter, M. (2010). Physical growth and maturation following early severe institutional deprivation: Do they mediate specific psychopathological effects? *Monographs of the Society for Research in Child Development*, 75(1 PG-143-166), 143–166. <https://doi.org/http://dx.doi.org/10.1111/j.1540-5834.2010.00554.x>
- Sowell, E. R., Thompson, P. M., Leonard, C. M., Welcome, S. E., Kan, E., & Toga, A. W. (2004). Longitudinal mapping of cortical thickness and brain growth in normal children. *The Journal of Neuroscience : The Official Journal of the Society for Neuroscience*, 24(38), 8223–8231. <https://doi.org/10.1523/JNEUROSCI.1798-04.2004>

- Spreng, R. N., & Grady, C. L. (2010). Patterns of Brain Activity Supporting Autobiographical Memory, Prospection, and Theory of Mind, and Their Relationship to the Default Mode Network. *Journal of Cognitive Neuroscience*, 22(6), 1112–1123. <https://doi.org/10.1162/jocn.2009.21282>
- Suarez, A., Lahti, J., Czamara, D., Lahti-Pulkkinen, M., Girchenko, P., Andersson, S., ... Raikkonen, K. (2018). The epigenetic clock and pubertal, neuroendocrine, psychiatric, and cognitive outcomes in adolescents, 1–12. <https://doi.org/10.1186/s13148-018-0528-6>
- Sumner, J. A., Colich, N. L., Uddin, M., Armstrong, D., & McLaughlin, K. A. (2019). Early Experiences of Threat, but Not Deprivation, Are Associated With Accelerated Biological Aging in Children and Adolescents. *Biological Psychiatry*, 85(3), 268–278. <https://doi.org/10.1016/j.biopsych.2018.09.008>
- Towne, B., Czerwinski, S. A., Demerath, E. W., Blangero, J., Roche, A. F., & Siervogel, R. M. (2005). Heritability of age at menarche in girls from the Fels Longitudinal Study. *American Journal of Physical Anthropology*, 128(1), 210–219. <https://doi.org/10.1002/ajpa.20106>
- Trickett, P. K., Noll, J. G., & Putnam, F. W. (2011). The impact of sexual abuse on female development: Lessons from a multigenerational, longitudinal research study. *Development and Psychopathology*, 23(02), 453–476. <https://doi.org/10.1017/S0954579411000174>
- Trickett, P. K., & Putnam, F. W. (1993). Impact of child sexual abuse on females: Toward a Developmental, Psychobiological Integration. *Psychological Science*, 4(2), 81–87. <https://doi.org/10.1111/j.1467-9280.1993.tb00465.x>
- Tyrka, A. R., Parade, S. H., Price, L. H., Kao, H. T., Porton, B., Philip, N. S., ... Carpenter, L. L. (2016). Alterations of Mitochondrial DNA Copy Number and Telomere Length with Early Adversity and Psychopathology. *Biological Psychiatry*, 79(2), 78–86. <https://doi.org/10.1016/j.biopsych.2014.12.025>
- Tyrka, A. R., Price, L. H., Gelernter, J., Schepker, C., Anderson, G. M., & Carpenter, L. L. (2009). Interaction of Childhood Maltreatment with the Corticotropin-Releasing Hormone Receptor Gene: Effects on Hypothalamic-Pituitary-Adrenal Axis Reactivity. *Biological Psychiatry*, 66(7), 681–685. <https://doi.org/10.1016/j.biopsych.2009.05.012>
- Ullsperger, J. M., & Nikolas, M. A. (2017). A meta-analytic review of the association between pubertal timing and psychopathology in adolescence: Are there sex differences in risk? *Psychological Bulletin*, 143(9), 903–938. <https://doi.org/10.1037/bul0000106>
- van Noord, P. A. H., & Kaaks, R. (1991). The effect of wartime conditions and the 1944-45 “dutch famine” on recalled menarcheal age in participants of the DOM breast cancer screening project. *Annals of Human Biology*, 18(1), 57–70. <https://doi.org/10.1080/03014469100001402>
- Velie, E. M., Nechuta, S., & Osuch, J. R. (2006). Lifetime Reproductive and Anthropometric Risk Factors for Breast Cancer in Postmenopausal Women. *Breast Disease*, 24(1), 17–35. <https://doi.org/10.3233/BD-2006-24103>
- Viechtbauer, W. (2010). Conducting meta-analyses in R with the metafor package. *Journal of Statistical Software*, 36(3), 1–48.
- Vijayakumar, N., Allen, N. B., Youssef, G., Dennison, M., Yücel, M., Simmons, J. G., & Whittle, S. (2016). Brain development during adolescence: A mixed-longitudinal investigation of cortical thickness, surface area, and volume. *Human Brain Mapping*, 37(6), 2027–2038. <https://doi.org/10.1002/hbm.23154>
- Vinkers, C. H., Kalafateli, A. L., Rutten, B. P., Kas, M. J., Kaminsky, Z., Turner, J. D., & Boks, M. P. (2015). Traumatic stress and human DNA methylation: a critical review. *Epigenomics*, 7(4), 593–608. <https://doi.org/10.2217/epi.15.11>
- Walhovd, K. B., Fjell, A. M., Giedd, J., Dale, A. M., & Brown, T. T. (2017). Through Thick and Thin: a Need to Reconcile Contradictory Results on Trajectories in Human Cortical Development. *Cerebral Cortex (New York, N.Y. : 1991)*, 27(2), 1472–1481. <https://doi.org/10.1093/cercor/bhv301>

- Webster, G. D., Graber, J. A., Gesselman, A. N., Crosier, B. S., & Schember, T. O. (2014). A Life History Theory of Father Absence and Menarche: A Meta-Analysis. *Evolutionary Psychology*, 12(2), 147470491401200. <https://doi.org/10.1177/147470491401200202>
- Widom, C. S., Horan, J., & Brzustowicz, L. (2015). Childhood maltreatment predicts allostatic load in adulthood. *Child Abuse and Neglect*, 47, 59–69. <https://doi.org/10.1016/j.chiabu.2015.01.016>
- Widom, C. S., Raphael, K. G., & DuMont, K. A. (2004). The case for prospective longitudinal studies in child maltreatment research: commentary on Dube, Williamson, Thompson, Felitti, and Anda (2004). *Child Abuse & Neglect*, 28(7), 715–722. <https://doi.org/10.1016/j.chiabu.2004.03.009>
- Wierenga, L. M., Langen, M., Oranje, B., & Durston, S. (2014). Unique developmental trajectories of cortical thickness and surface area. *NeuroImage*, 87, 120–126. <https://doi.org/10.1016/j.neuroimage.2013.11.010>
- Wolf, E. J., Maniates, H., Nugent, N., Maihofer, A. X., Armstrong, D., Ratanatharathorn, A., ... Logue, M. W. (2018). Traumatic stress and accelerated DNA methylation age: A meta-analysis. *Psychoneuroendocrinology*, 92(November 2017), 123–134. <https://doi.org/10.1016/j.psyneuen.2017.12.007>
- Wren, M. E., Shirtcliff, E. A., & Drury, S. S. (2015). Not All Biofluids Are Created Equal: Chewing over Salivary Diagnostics and the Epigenome. *Clinical Therapeutics*, 37(3), 529–539. <https://doi.org/10.1016/j.clinthera.2015.02.022>
- Wu, M., Kujawa, A., Lu, L. H., Fitzgerald, D. A., Klumpp, H., Fitzgerald, K. D., ... Phan, K. L. (2016). Age-related changes in amygdala-frontal connectivity during emotional face processing from childhood into young adulthood. *Human Brain Mapping*, 37(5), 1684–1695. <https://doi.org/10.1002/hbm.23129>
- Zhu, X., Han, W., Xue, W., Zou, Y., Xie, C., Du, J., & Jin, G. (2016). The association between telomere length and cancer risk in population studies. *Scientific Reports*, 6, 1–10. <https://doi.org/10.1038/srep22243>

Figure Legends

Figure 1. 2009 PRISMA flow diagram of literature search and screening process.

Figure 2. Association of all adversities and pubertal timing.

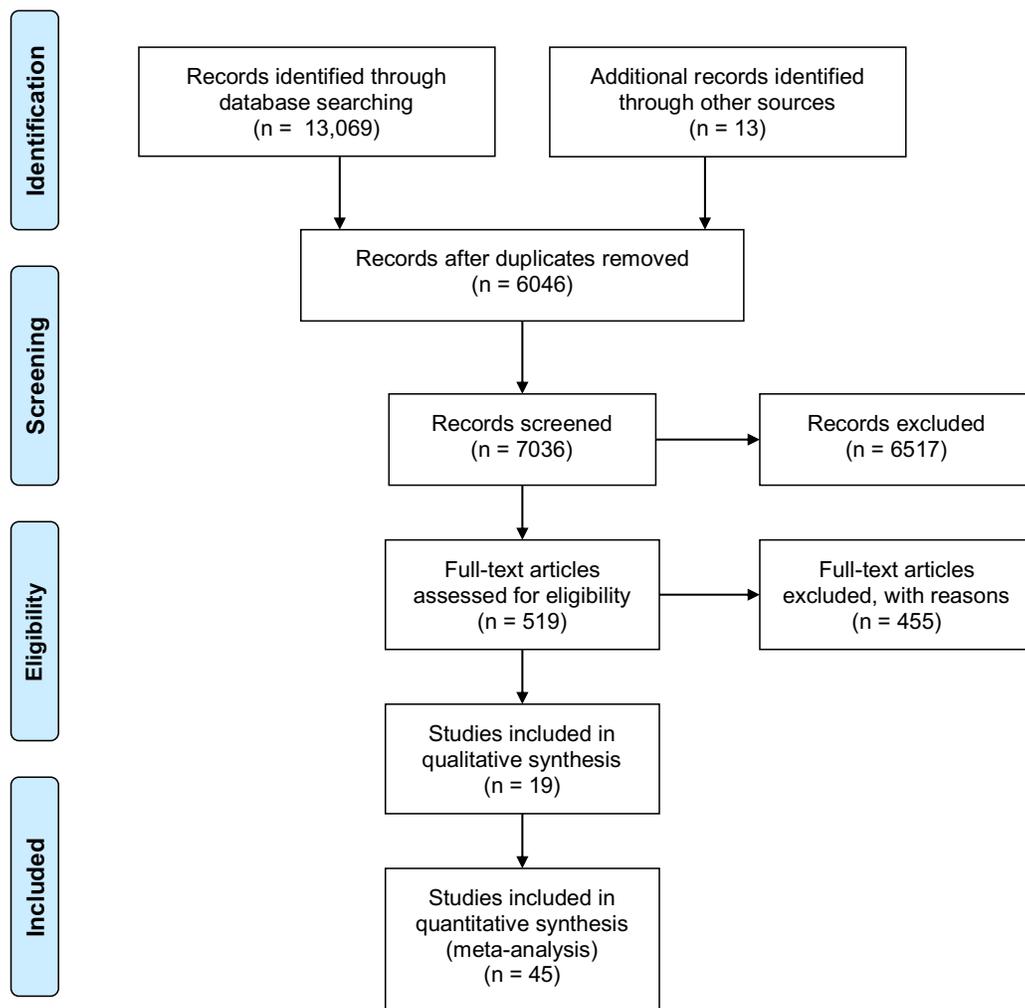
Figure 3. Association of adversity and pubertal timing by adversity type.

Figure 4. Association of all adversities and cellular aging.

Figure 5. Association of adversity and cellular aging by adversity type.

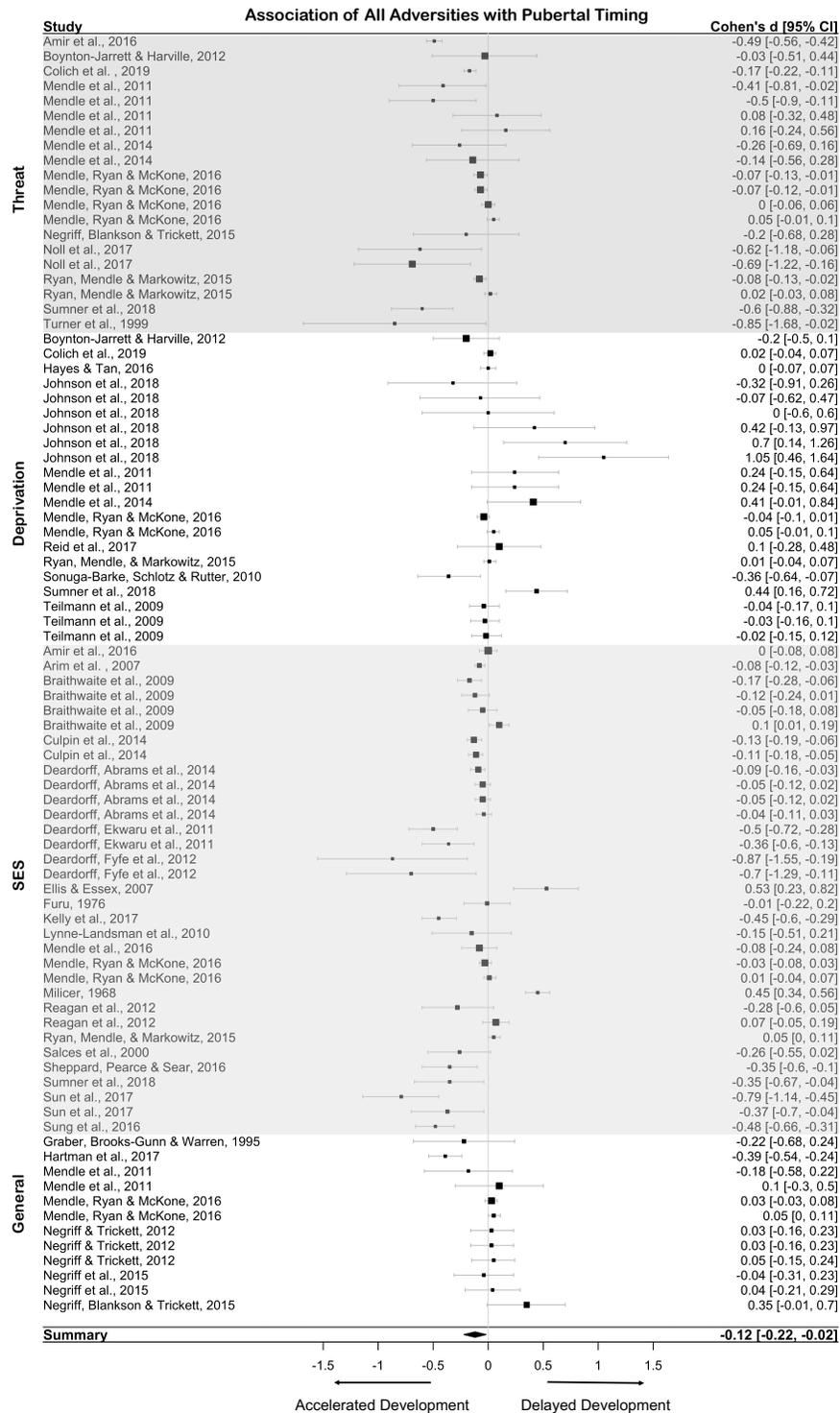


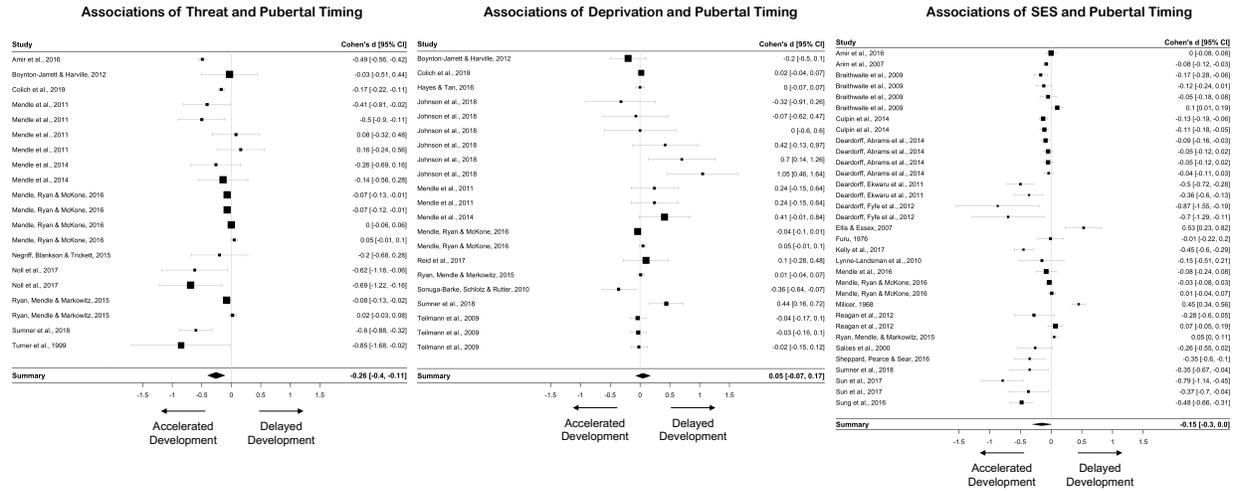
PRISMA 2009 Flow Diagram

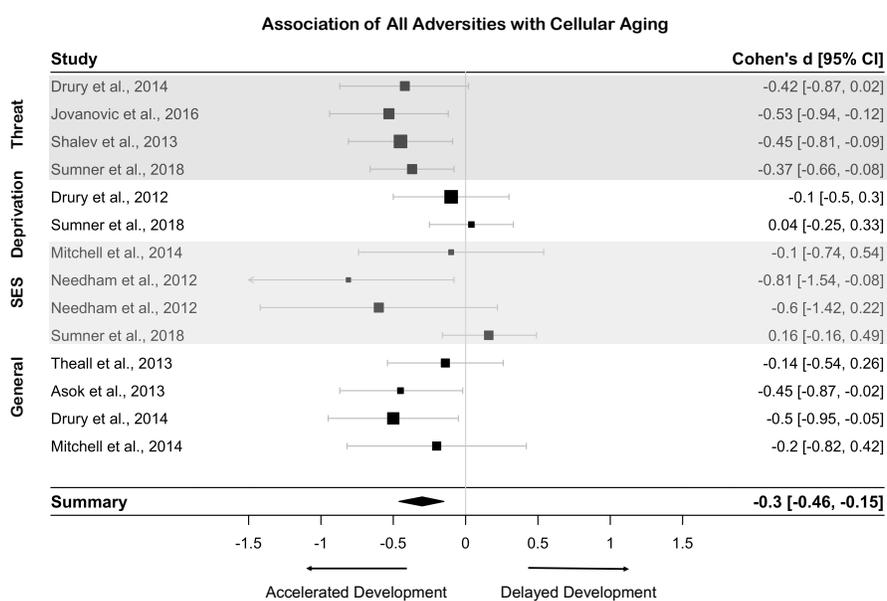


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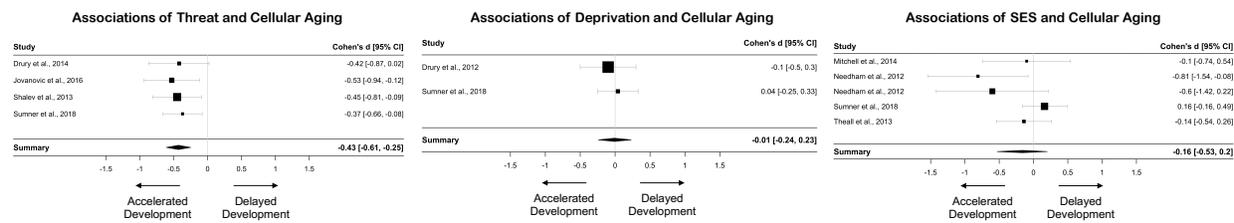


Table 1. Characteristics of Individual Studies Included in the Pubertal Timing Meta-Analysis								
Study	Sample	Sample Size (N)	Age Range	Sex Distribution	Early Life Adversity	Adversity Details	Pubertal Timing Metric	Covariates
Amir, Jordan & Bribiescas, 2016	Nationally Representative Sample (US); Add Health Survey Participants	3606	12-20	Female only	Threat/SES	Subjective ratings of the availability of resources (family income) and environmental safety	Age at Menarche	None
Arim, Shapka, Dahinten, Willms, 2007	Nationally Representative Sample (Canada); National Longitudinal Survey of Children and Youth	7,977	10-17	50% Female	SES	Family SES (composite of household income, parental education and occupation levels, weighted by the factor loadings obtained through principal components factor analysis with varimax rotation)	PDS	Sex
Boynton-Jarrett & Harville, 2012	Nationally Representative Sample (UK); National Child Development Study	4,524	11-16	Female only	Threat/Deprivation	Exposure to violence/mental health issues; Lack of supportive caregiving	Menarche < 11 years	None
Braithwaite, Moore, Lustig, Epel, Ong, Rehkopf, Wang, Miller, & Hiatt, 2009	Nationally Representative Sample (US); National Growth and Health Study	2,077	9-19	Female only	SES	SES (parent education level and household income)	Age at Menarche < 12 years	None

Colich, Platt, Keyes, Sumner, Allen & McLaughlin	Nationally Representative Sample (US); National Comorbidity Study - Adolescent Sample	4,937	13-18	Female only	Threat/Deprivation	Threat exposure composite, Deprivation exposure composite	Age at Menarche	None
Culpin, Heron, Araya, Melotti, Lewis, & Joinson, 2014	Nationally Representative Sample (UK); Avon Longitudinal Study of Parents and Children	3,776	8-18	Female only	SES	Presence of financial problems at age 6 and age 8 and Mother's education status	Age at Menarche	None
Deardorff, Abrams, Ekwuru & Rehkopf, 2014	Nationally Representative Sample (US); National Longitudinal Study of Youth	4,851	9-16	Female only	SES	SES before age 7 (Family income; Family wealth [income - total debt]; mother's education level; mother's employment status)	Age at Menarche	None
Deardorff, Ekwuru, Kushi, Ellis, Greenspan, Mirabedi, Landaverde, & Hiatt, 2011	Cohort Study of Young Girls' Nutrition, Environment, and Transitions	444	6-10	Female only	SES	Family Income (<75,000)	Tanner Stage	None
Deardorff, Fyfe, Ekwuru, et al 2012	Cohort Study of Young Girls' Nutrition, Environment, and Transitions	213	6-12	Female only	SES	Family Income (<50,000)	Tanner Stage	None
Ellis & Essex, 2007	Wisconsin Study of Families and Work	180	0-11	Female only	SES	SES	PDS and Tanner	None

Furu, 1976	Community sample from Sweden	917	10-17	Female only	SES	Father's or Mother's Occupation	Age at Menarche	General Health; BMI
Graber, Brooks-Gunn & Warren, 1995	Premenarcheal subset from a larger longitudinal sample	75	10-14	Female only	General	Number of stressful life events	Age at Menarche and Tanner Stage	None
Hartman, Li, Nettle, Belsky, 2017	Nationally Representative Sample; NICHD Study of Early Child Care and Youth Development (SECCYD)	659	1-15	Female only	General	Composite of maternal sensitivity, maternal harshness, unpredictability, income harshness	Age at Menarche	General Health; BMI
Hayes & Tan, 2016	Cohort of adopted Chinese girls	16,202	11-13	Female only	Deprivation	International Adoption (from China to North America)	Age at Menarche	None
Johnson, Tang, Almas et al., 2018	Bucharest Early Intervention Project (BEIP)	107	0-12	49% Female	Deprivation	Institutionalization	Age at Menarche and Tanner Stage	None
Kelly, Zilanawala, Sacker, Hiatt & Viner, 2017	Longitudinal cohort from UK Millennium Cohort Study	5,839	0-11	Female only	SES	SES (Income Quartiles)	Menarche status at age 11	None
Lynne-Landsman, Graber & Andrews, 2010	Longitudinal cohort from Oregon Youth Substance Use Project; 538 females; 537 males	1,075	Grades 1-12	52% female	SES	Household resources, conflict and stability	PDS	None
Mendle, Leve, Van Ryzin, & Natsuaki, 2014	Longitudinal intervention cohort of girls in foster care	100	10-12	Female only	Threat/Deprivation	Sexual Abuse, Neglect, Physical Abuse, Foster Care Placement	PDS	None

Mendle, Leve, Van Ryzin, Natsuaki, & Ge, 2011	Longitudinal intervention cohort of girls in foster care	100	10-12	Female only	Threat/Deprivation	Sexual Abuse, Neglect, Physical Abuse, Foster Care Placement	Age at Menarche and PDS	All forms of adversity, child's age, child's race
Mendle, Moore, Briley & Harden, 2016	Nationally Representative Sample (US); Add Health Survey Participants	1,260	11-21	Female only	SES	SES (parental education)	Age at Menarche	None
Mendle, Ryan, McKone, 2016	Nationally Representative Sample (US); Add Health Survey Participants	6,273	11-34	Female only	Threat/Deprivation/SES	Sexual Abuse, Physical Abuse, Physical Neglect, Undetermined Maltreatment, SES (income to needs ratio)	Age at Menarche and Secondary Sex Characteristics (Breast and Curve Development)	None
Milicer, 1968	Community sample in Wroclaw, Poland	5,859	9-17	Female only	SES	SES (parental education, occupation and income)	Age at Menarche	age, BMI, race
Negriff & Trickett 2012	Longitudinal cohort in US	454	9-13	46.9% female	General	Active case within Department of Child and Family Services	Tanner Stage and PDS	None
Negriff, Blankson & Trickett, 2015	Longitudinal cohort in US	454	9-13	47% female	Threat	Sexual Abuse, Other Maltreatment	PDS	ethnicity, obesity, biological father absence
Negriff, Saxbe, & Trickett, 2015	Longitudinal cohort in US	454	9-13	47% female	General	Active case within Department of Child and Family Services	PDS	None

Noll, Trickett, Long et al., 2017	Longitudinal cohort in US	173	11-20	Female only	Threat	Sexual Abuse	Tanner Stage	None
Reagan, Salsberry, Fang, Gardner & Pajer, 2012	Nationally Representative Sample (US); National Longitudinal Study of Youth	2,337	12-17	Female only	SES	% time in poverty between ages 0-5	Age at Menarche	All forms of adversity, child's age, child's race
Reid, Miller, Dorn, Desjardins, Donzella & Gunnar, 2017	Cohort of interntionally adopted youth	165	7-14	Female only	Deprivation	Institutionalizati on	% Menstruating	BMI, people in household, father's occupation
Ryan, Mendle & Markowitz, 2015	Nationally Representative Sample (US); Add Health Survey Participants	6,273	11-34	Female only	Threat/Depriva tion/SES	Sexual Abuse, Phycial Abuse, Physical Neglect, SES (income to needs ratio)	Age at Menarche	None
Salces, Rebato, Susanne, San Martin & Rosique, 2001	Community sample from Basque Country, Spain	196	9-17	Female only	SES	SES (father's occupation)	Age at Menarche	None
Sheppard, Pearce & Sear, 2016	Longitudinal sample (UK); Newcastle Thousand Families Study	251	0-15	Female only	SES	SES (father's occupation)	Age at Menarche	Sex, race/ethnicity, family poverty status, other exposure composite

Sonuga-Barke, Schlotz, & Rutter, 2010	Cohort of adopted Romanian youth	197	6-15	~ 50% Female	Deprivation	Intitutionaliziati on	Tanner Stage	Parental early puberty history, perinatal factors, childhood and current adiposity, physical activity, sleep duration, presence of stepfather, stressful life events
Sumner, Colich, Uddin, Armstrong & McLaughlin, 2018	Community sample of maltreated children and adolescents from Seattle, WA	247	8-16	48% female	Threat/Deprivation/SES	Threat exposure composite, Deprivation exposure composite, SES (below or above poverty line)	Tanner Stage	Sex, Race/Ethnicity, Family Poverty Status, Other Adversity Exposure Composite
Sun, Mensah, Azzopardi, Patton & Wake, 2017	Nationally Representative Sample (Australia); The Longitudinal Study of Australian Children	3,764	0-11	48% female	SES	SES (family income, education and occupation)	PDS	None

Sung, Simpson, Griskevicius, Kuo, Schlomer & Belsky, 2016	Nationally Representative Sample (US); NICHD Study of Early Child Care and Youth Development (SECCYD)	492	0-15	Female only	SES	SES (income to needs ratio) in the first 5 years of life	Age at Menarche	BMI, race, age, other adversity
Teilmann, Pedersen, Gormsen, Damgaard, Skakkebaek & Jensen, 2009	Cohort of Internationally adopted individuals in Denmark	1,376	4-13	Female only	Deprivation	International Adoption (from Asia, South America, Eastern Europe and Africa to Denmark)	Tanner Stage	None
Turner, Runtz & Galambos, 1999	Canadian cohort of girls recruited from outpatient clinical agencies and public schools	44	12-19	Female only	Threat	Sexual Abuse	Age at Menarche	None

Table 2. Characteristics of Individual Studies Included in the Cellular Aging Meta-Analysis								
Study	Sample	Sample Size (N)	Age Range	Sex Distribution	Early Life Adversity	Adversity Details	Cellular Aging Metric	Covariates
Asok, Bernard, Roth, Rosen & Dozier, 2013	Longitudinal cohort examining efficacy of attachment-based intervention for infants in Child Welfare System	38	4-7	58% Female	General	Identified by Child Welfare System as being at-risk for maltreatment (conditions noted most often include child neglect, domestic violence, parental substance use, inadequate housing)	Telomere Length (buccal swabs)	Household Income, Sex, Minority Status
Drury, Mabile, Brett et al. 2014	Community sample from New Orleans, Louisiana	80	5-15	49% Female	Threat/General	Family Instability (witnessing family violence, suicide, incarceration)	Telomere Length (buccal swabs)	None
Drury, Theall, Gleason et al., 2012	Bucharest Early Intervention Project (BEIP)	100	6 months - 10	41% Female	Deprivation	Early Institutionalization (% of time spend in institution at 54 months)	Telomere Length (buccal swabs)	Group, sex, ethnicity, age at telomere collection and low birth weight
Jovanovic, Vance, Cross et al., 2016	Longitudinal cohort of trauma exposed children and adolescents	101	6-13	54% Female	Threat	Violence Exposure	Horvath DNAm age (saliva)	None
Mitchell, Hobcraft, McLanahan, et al., 2014	2 cohorts including Fragile Families and Child Wellbeing Study and an independent cohort	40	9	Male only	General/SES	Harsh Environment determined by equally weighted combination of (i) family economic conditions, (ii) parenting	Telomere Length (saliva)	None

						practices, and (iii) family structure/stability -- then incorporating maternal depression; Income to Needs Ratio		
Needham, Fernandez, Lin, Epel & Blackburn, 2012	Admixture Mapping for Ethnic and Racial Insulin Complex Outcomes (AMERICO) cohort	70	7-13	53% Female	SES	SES (family income and parental education)	Telomere Length (blood)	None
Shalev, Moffitt, Sugden et al., 2013	Environmental-Risk Longitudinal Twin Study	236	5-12	49% Female	Threat	Domestic Violence, Bullying Victimization, Physical Maltreatment	Telomere Length (buccal swabs)	None
Sumner, Colich, Uddin, Armstrong & McLaughlin, 2018	Community sample of maltreated children and adolescents from Seattle, WA	247	8-16	48% Female	Threat/Deprivation/SES	Threat exposure composite, Deprivation exposure composite, SES (below or above poverty line)	Horvath DNAm age (saliva)	Sex, Race/Ethnicity, Family Poverty Status, Other Adversity Exposure Composite
Theall, Brett, Shirtcliff, Dunn & Drury, 2013	Community sample from New Orleans, Louisiana	99	4-14	53% Female	SES	Household socioeconomic position	Telomere Length (saliva)	None

Table 3. Characteristics of Individual Studies Included in the Cortical Thickness Systematic Review										
Authors	Sample	Sample Size (N)	Age Range ¹	Sex Distribution	Early-Life Adversity	Adversity Details	ROI or Whole Brain?	Brain regions with significant associations with adversity)	Covariates	Group Matching Characteristics
Busso, McLaughlin, Brueck, Peverill, Gold & Sheridan, 2017	Community based sample of children in Boston, MA exposed to high rates of maltreatment	51	13-20	60% Female	Threat	Childhood Trauma Questionnaire and Childhood Experiences of Care and Abuse to assess emotional, sexual, and physical abuse	ROI (vmPFC, lateral OFC, anterior and posterior cingulate cortices, ventrolateral PFC (including inferior frontal gyrus [IFG]), dorsolateral PFC (including middle and superior frontal gyri), insular cortex, parahippocampal gyrus, temporal pole, and lateral temporal cortex (spanning inferior, middle, and superior	Thinner cortex among abused in vmPFC, right inferior frontal gyrus, left and right parahippocampal gyri, right inferior temporal gyrus, and right middle temporal gyrus compared to non-exposed controls	age, gender, parental education	race, IQ, handedness

							temporal gyri)			
Gold, Sheridan, Peverill, Busso, Lambert, Alves, Pine & McLaughlin, 2016	Community based sample of children in Boston, MA exposed to high rates of maltreatment	58	13-20	60% Female	Threat	Childhood Trauma Questionnaire and Childhood Experiences of Care and Abuse to assess emotional, sexual, and physical abuse	ROIs in medial and lateral prefrontal cortex, and medial and lateral temporal cortex	Thinner cortex among abused sample in bilateral ventromedial prefrontal cortex, right orbitofrontal cortex, left temporal pole, right inferior frontal gyrus, left and right parahippocampal gyri, left and right inferior temporal gyrus, right superior temporal gyrus, and right middle temporal gyrus	race, age, sex, and parental education	
McLaughlin, Sheridan, Gold, Duys, Lambert, Peverill, Heleniak, Shechner, Wojcieszak	Community based sample of children in Seattle, WA exposed to high rates of	60	18-Jun	48% Female	Threat	Childhood Trauma Questionnaire and Childhood Experiences of Care and Abuse to assess	ROI in dorsal anterior cingulate and ventromedial prefrontal cortex	No significant associations with cortical thickness	age, sex	

& Pine, 2016	maltreatment					emotional, sexual, and physical abuse				
Kelly, Viding, Wallace, Schaer, De Brito, Robustelli & McCrory, 2013	Community based sample recruited from London Social Services department	43	12.77(1.19) for controls, 12.27(1.41) for maltreated	56% Female	Threat	Social worker report using Kaufman's 4 point scale for physical abuse, emotional abuse, sexual abuse, and neglect	Whole brain	decreased cortical thickness in mPFC in maltreated,	age and sex	Tanner stage, handedness, cognitive ability, ethnicity
Lawson, Duda, Avants, Wu & Farah, 2013	Participants from NIH Pediatric MRI Data Repository	283	11.47 (3.50)	53% Female	SES	Family income, parental education summed from two parents. Parental education variable was square-root transformed	ROIs in the prefrontal cortex	Lower parental education was associated with decreased cortical thickness in right anterior cingulate cortex and left superior frontal gyrus. No associations emerged with parental income.	age, sex, total brain volume, race, body mass index	N/A
Mackey, Finn, Leonard, Jacoby-Senghor, West,	Community sample from Boston, Massachusetts area	58	Lower income: 14.47 (0.38), Higher income:	53% Female	SES	Free or reduced-price lunch status obtained from the	Whole brain	Thinner cortex in lower income group in bilateral	sex, intracranial volume. Associations also hold when	None

Gabrieli, Gabrieli, 2015			14.35 (0.47)			Massachusetts Department of Elementary and Secondary Education		temporal and occipital lobes and in right lateral prefrontal cortex	controlling for race and ethnicity	
Jednoróg, Altarelli, Monzalvo, Fluss, Dubois, Billard, Dehaene-Lambertz & Ramus, 2012	Community sample from Paris, France	23	8-10	57% Female	SES	Hollingshead Two-Factor Index of Social Position using maternal report of maternal occupation and education	Whole Brain	Positive associations between SES and thickness in L angular gyrus, L heschl's gyrus, L post-ventral cingulate, and L anterior occipital sulcus. Negative associations between SES and thickness in L fronto-maginal gyrus/sulcus, L middle frontal sulcus, and L intraparietal sulcus.	None	N/A

Noble, Houston, Brito, Bartsch, Kan, Kuperman, Akshoomoff, Amaral, Bloss, Libiger, Schork, Murray, Casey, Chang, Ernst, Frazier, Gruen, Kennedy, Van Zijl, Mostofsky, Kaufmann, Kenet, Dale, Jernigan & Sowell, 2015	Pediatric Imaging, Neurocognition, Genetics Study (PING)	1148	3-20	48% Female	SES	parent report of income and education	Whole Brain	No significant associations between cortical thickness and SES or SES x Age interactions	age, age-squared, scanner, sex, genetic ancestry	N/A
Piccolo, Merz, He, Sowell, Noble KG; Pediatric Imaging, Neurocognition, Genetics Study, 2016	Pediatric Imaging, Neurocognition, Genetics Study (PING)	1148	3-20	48% Female	SES	parent report of income and education	Whole Brain	Age(square d) X SES (both income and education) interaction in average cortical thickness, such that lower SES children demonstrate faster thinning across the whole cortex. No specific	SES measure, age, age-squared, scanner, sex, genetic ancestry, and parental SES measure x age	N/A

								regions survive correction for multiple comparisons		
Rosen, Sheridan, Sambrook, Meltzoff & McLaughlin, 2018	Community based sample of children in Seattle, WA exposed to high rates of maltreatment (t)	53	6-19	48% Female	Deprivation/ SES	SES-Family income divided by the federal poverty line for a family of that size; Cognitive stimulation-Parent report using Home Observation Measurement of the Environment (HOME)-short form	Whole Brain and ROI	Lower levels of cognitive stimulation were associated with thinner cortex in left superior parietal lobule and left middle frontal gyrus. There were no associations with SES and no significant whole brain associations.	age, sex, violence exposure	N/A
Avants, Hackman, Betancourt, Lawson, Hurt & Farah, 2015	52 young adults	52	19.16 years (SD = 1.32)	52% female	Deprivation	Environmental Stimulation as measured by the HOME at ages 4 and 8	Whole Brain	Thicker cortex among children with lower levels of cognitive stimulation at age 4 (but not 8) in L lateral inferior temporal—posterior,	age at scan, maternal IQ, gender, foster care and prenatal cocaine	N/A

								L lateral Inferior temporal -anterior, Bilateral fusiform, R lateral inferior temporal		
Hodel, Hunt, Cowell, Van Den Heuvel, Gunnar & Thomas, 2015	110 post- institutionali zed children, from Minnesota Internationa l Adoption Project	172	12-14	61% female	Deprivation	previously institutionali zed children adopted into the United States	Whole Brain	Thinner cortex in right inferior frontal gyrus among previously institutionali zed children compared to controls	age, sex	
McLaughlin , Sheridan, Winter, Fox, Zeanah & Nelson, 2014	Bucharest Early Intervention Project (BEIP)	80	8-10	50% female	Deprivation	part of a randomized control trial where 1/2 of the institutionali zed children were placed in high quality forster care. There were no differences between the care as usual group and foster care group, so all analyses compared the Ever	Whole Brain	widespread reductions in cortical thickness across frontal, parietal and temporal lobes in children who were ever institutionali zed	age, sex, total brain volume, and TR length (bc different acquisition parameters)	

						Institutionalized Group (EIG) to the Never Institutionalized Group (NIG)			
¹ Age range (in. years) was the most commonly used descriptor of the full sample. When this was not available, means and standard deviations are given.									

Table 4. Characteristics of Individual Studies Included in the amygdala-mPFC Review											
Authors	Sample	Sample Size (N)	Age Range	Sex Distribution	Early-Life Adversity	Adversity Details	Seed-to-seed or Whole Brain?	Task	Associations with adversity and amygdala-mPFC connectivity	Covariates	Group Matching Characteristics
Cisler, Scott, Smitherman, Lenow & Kilts, 2013	Community based sample in Arkansas	30	12-16	100% female	Threat	Assessed using the trauma assessment section of the National Survey of Adolescents (sexual assault, physical assault, severe abuse from caregiver)	whole brain	implicit emotion processing task	greater connectivity between right amygdala and left middle frontal gyrus, and right amygdala and left perigenual anteriorcingulate	none	Age, ethnicity, anxiety and depression diagnoses
Colich, Williams, Ho et al., 2017	Community based sample in the San Francisco Bay Area	137	9-13	57% female	Threat	Based on modified version of Traumatic Events Screening Inventory for Children	seed-to-seed	implicit emotion regulation task	ELS was associated with increased negative connectivity between the right ventrolateral prefrontal cortex and bilateral amygdala	age	N/A

Gee, Gabard-Durnam, Flannery et al., 2013	Post institutionalized children adopted into the United States and comparison children*	89	6-18	56% female	Deprivation	Sample of previously institutionalized children adopted into the United States	whole brain	Emotional face viewing task	Comparison group children showed positive amygdala-mPFC connectivity during viewing of fearful faces while adolescents showed comparison group adolescents showed negative connectivity. In contrasts, both PI children and adolescents showed negative amygdala-mPFC connectivity	amygdala reactivity	Pubertal status
Keding & Herringa, 2016	Community based sample in Wisconsin. Sampled specifically for PTSD	53	8-18	58% female	Threat	All trauma exposed participants also had PTSD. Trauma exposures were Sexual abuse, traumatic death of loved one, witnessing violence, accident,	whole brain	implicit dynamic emotion face	PTSD sample showed reduced connectivity between amygdala and both dorsal anterior cingulate and ventrolateral PFC during angry trials and increases	age, sex	age, pubertal status, IQ, handedness

						physical abuse			connectivity to happy faces. Control participants showed the opposite pattern of results.		
Marusak, Martin, Etkin & Thomason, 2015	Community based sample in Detroit, MI	30	9-17	80% female	Threat	At least one traumatic life event endorsed on parent or child report of Children's Trauma Assessment Center Screen Checklist. These included sexual abuse, physical abuse, neglectful home environment, exposure to domestic violence, exposure to other violence, multiple separations from caregiver	whole brain	Emotional Stroop	During emotional conflict regulation, trauma exposed group demonstrated positive amygdala - pregenual cingulate cortex connectivity while the control group demonstrated negative connectivity.	none	age, sex, IQ

Silvers, Lumian, Gabard-Durnam et al., 2016	Post institutionalized children adopted into the United States and comparison children*	89	7-16	62% female	Deprivation	Sample of previously institutionalized children adopted into the United States	whole brain	Aversive learning paradigm	During a fear conditioning task, PI youth demonstrate positive connectivity between the amygdala and vmPFC, dmPFC, and vIPFC, while the comparison group demonstrates negative connectivity	average head motion,	age, IQ
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