Identification of novel key biomarkers in Simpson-Golabi-Behmel Syndrome: Evidence from bioinformatics analysis

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Abstract:

Background: The Simpson-Golabi-Behmel Syndrome (SGBS) or overgrowth Syndrome is a rare inherited X-linked condition characterized by pre- and postnatal overgrowth. The aim of the present study is to identify functional non-synonymous SNPs of GPC3 gene using various in silico approaches. These SNPs are supposed to have a direct effect on protein stability through conformation changes. **Material and methods:** The SNPs were retrieved from the Single Nucleotide Polymorphism database (dbSNP) and further used to investigate a damaging effect using SIFT, PolyPhen, PROVEAN, SNAP2, SNPs&GO, PHD-SNP and P-mut, While we used I-mutant and MUPro to study the effect of SNPs on GPC3 protein structure. The 3D structure of human GPC3 protein is not available in the Protein Data Bank, so we used RaptorX to generate a 3D structural model for wild-type GPC3 to visualize the amino acids changes by UCSF Chimera. For biophysical validation we used project HOPE. Lastly we run conservational analysis by BioEdit and Consurf web server respectively. **Results**: our results revealed three novel missense mutations (rs1460413167, rs1295603457 and rs757475450) that are found to be the most deleterious which effect on the GPC3 structure and function. **Conclusion:** This present study could provide a novel insight into the molecular basis of overgrowth Syndrome.

Keywords: Simpson-Golabi-Behmel Syndrome (SGBS), overgrowth Syndrome, *GPC3* bioinformatics analysis, SNPs, diagnostic markers.

1. Introduction:

The Simpson-Golabi-Behmel Syndrome (SGBS) or overgrowth Syndrome is a rare inherited X-linked condition characterized by pre- and postnatal overgrowth, Affected individuals have several dysmorphisms that can include a distinct facial appearance, macroglossia, cleft palate, cardiac defects, enlarged and dysplastic kidneys, cryptorchidism, hypospadia, hernias, supernumerary nipples, vertebral and rib anomalies, coccygeal bony appendage, syndactyly, and polydactyly.[1-4] the first case was reported around 1940.[5] To date, two different clinical subtypes of SGBS have been defined. The typical SBGS (SGBS type I) [2-8] and a lethal and rare form, probably with less than 10 cases described known as SGBS type II.[9-11] In addition, these patients have an increased risk for the development of Wilms' tumors.[12] Premature death is also very frequent.[13] Different mutations have been reported in SGBS type I.[14-22]

This dysplasia syndrome caused by loss-of-function mutations of the X-linked *GPC3* gene is localized on Xq26.1 [23, 24] which encodes a developmentally regulated cell membrane proteoglycan, glypican-3. [17, 19, 20, 25-29] that apparently plays a negative role in growth control by an unknown mechanism, However, outcomes from a detailed comparative analysis of growth patterns in dual mutants lacking GPC3 provided conclusive genetic evidence inconsistent with the theory that GPC3 performances as a growth suppressor by downregulating an IGF ligand.[29] Such a proteoglycan is inferred to play an important role in control and diagnosis in mesodermal tissues and in tumors predisposition.[30, 31] Some studies show association between *GPC3* gene and some types of human cancers.[32-36]

The aim of this study was to identify the pathogenic SNPs in the coding region of *GPC3* gene which causes SGBS type I using variant bioinformatics tools, Also to identify the most deleterious nsSNPs that could be used as diagnostic markers. A nsSNP is a single base change in a coding region that causes an amino acid change in its corresponding protein. If nsSNP modifies protein function, the change can have major phenotypic effects which are responsible for the pathology of the disease [37, 38] Genetic testing for mutations often reveals substitutions that are not easily categorized as pathogenic. Therefore a great effort has been done in translational bioinformatic tools for analysis of nsSNPs which have enhanced significantly in recent years and thus become more reliable for SNPs analysis.[39] Translational bioinformatics has become an important discipline in the era of personalized and precision medicine which aims to fill the gap between clinical and academic research by prioritizing the most pathogenic nsSNPs for further studies.[40-44] This is the first in silico analysis in the coding region of *GPC3* gene that prioritized nsSNPs for the larger population-based studies of overgrowth syndrome.

2. Material and methods:

2.1 Data mining:

The data on human *GPC3* gene was collected from National Center for Biological Information (NCBI) web site.[45] The SNP information (protein accession number and SNP ID) of the *GPC3* gene was retrieved from the NCBI dbSNP (http://www.ncbi.nlm.nih.gov/snp/) and the protein sequence was collected from Uniprot database. (<u>https://www.uniprot.org/</u>).

2.2 PolyPhen:

PolyPhen (version 2) We used PolyPhen to study probable impacts of A.A. substitution on structural and functional properties of the protein by considering physical and comparative approaches.[46] It is available at (http://genetics.bwh.harvard.edu/pph2/).

2.3 PROVEAN:

PROVEAN is a software tool which predicts whether an amino acid substitution or indel has an impact on the biological function of a protein. It is useful for filtering sequence variants to identify nonsynonymous or indel variants that are predicted to be functionally important.[47] It is available at (<u>https://rostlab.org/services/snap2web/</u>).

2.4 SNAP2:

SNAP2 is a trained classifier that is based on a machine learning device called "neural network". It distinguishes between effect and neutral variants/non-synonymous SNPs by taking a variety of sequence and variant features into account.[48] It is available at (https://rostlab.org/services/snap2web/).

2.5 SNPs&GO:

SNP&GO is a method for the prediction of deleterious SNPs using protein functional annotation. The server is based on SVM classifier that discriminates between disease related and neutral SNPs. It has two components, one is sequence based and the other is structure based. The other method were used too (PHD-SNP and PANTHER) .[49] It is available at (http://snps.biofold.org/snps-and-go/snps-and-go.html).

2.6 Stability Analysis:

2.6.1 I-Mutant 3.0:

I-Mutant is an SVM-based tool for the automatic prediction of protein stability changes upon single point mutations. The predictions are performed starting either from the protein structure or, more importantly, from the protein sequence[50] It is available at (http://gpcr2.biocomp.unibo.it/cgi/predictors/I-Mutant3.0/IMutant3.0.cgi).

2.6.2 MUpro:

MUpro is a SVM-based tool for the prediction of protein stability changes upon nsSNPs. The value of the energy change is predicted, and a confidence score between -1 and 1 for measuring the confidence of the prediction is calculated. The accuracy for SVM using sequence information is 84.2%.[51] It is available at (http://mupro.proteomics.ics.uci.edu/).

2.7 Structural Analysis:

2.7.1 Modeling nsSNP locations on protein structure:

Project hope is a new online web-server to search protein 3D structures (if available) by collecting structural information from a series of sources, including calculations on the 3D coordinates of the protein, sequence annotations from the UniProt database, and predictions by DAS services. Protein sequences were submitted to project hope server in order to analyze the structural and conformational variations that have resulted from single amino acid substitution corresponding to single nucleotide substitution. It is available at (http://www.cmbi.ru.nl/hope).

2.7.2 Modeling Amino Acid Substitution:

UCSF Chimera is a highly extensible program for interactive visualization and analysis of molecular structures and related data, including density maps, supramolecular assemblies, sequence alignments, docking results, conformational analysis Chimera (version 1.8).[52] It is available at (http://www.cgl.ucsf.edu/chimera/).

2.8 Conservational Analysis:

2.8.1 BioEdit:

BioEdit software is intended to supply a single program that can handle most simple sequence and alignment editing and manipulation functions as well as a few basic sequences analyses. It is available for download at (http://www.mbio.ncsu.edu/bioedit/bioedit.html).

2.8.2 Identification of Functional SNPs in Conserved Regions by using ConSurf server:

ConSurf web server provides evolutionary conservation profiles for proteins of known structure in the PDB. Amino acid sequences similar to each sequence in the PDB were collected and multiply aligned using CSI-BLAST and MAFFT, respectively. The evolutionary conservation of each amino acid position in the alignment was calculated using the Rate 4Site algorithm, implemented in the ConSurf web server.[53] It is available at (http://consurf.tau.ac.il/).



Figure (1): Diagrammatic Workflow Software's used in SNPs analysis.

3. Results:

The effect of each SNP has been studied regarding function and stability of the protein using different in analysis tools with different considerations and features, in order to minimize the error to the least percentage possible. (Figure 1)

The total number of nsSNPs in the coding region of the *GPC3* gene in Human is 765 SNPs we retrieved from the dbSNP/NCBI database. There were 256 missense mutations then submitted them to functional analysis by SIFT, PolyPhen-2, PROVEAN and SNAP2 respectively. SIFT predicted 109 damaging SNPs, polyphen-2 predicted 115 damaging SNPs (50 possibly damaging and 65 probably damaging), PROVEAN predicted 82 damaging SNPs and SNAP2 predicted 127 deleterious SNPs. After filtering the Quadra-positive damaging SNPs, the number of SNPs reduced to 7 SNPs (Table 1) and we submitted them to SNP&GO, PANTHER, PhD-SNP and P-mut for further study their effect on the function. PhD-SNP and P-mut predicted 7 disease associated SNPs, SNP&GO predicted 5, and while PANTHER predicted 5 disease associated SNPs. So we filtered the Quadra-positive disease associated SNPs the number reduce to 4 SNPs (Table 2) and submitted them to I-Mutant3.0 and MUPro to study their effect on the stability. All the SNPs were found to cause a decrease in the stability of the protein except for one SNP (G257D) predicted by I-Mutant 3.0 to increase the stability. (Table 3)

Various	Solution								
		SIFT				PROVEAN		SNAP2	
dbSNP rs#	sub	Prediction	Score	Polyphen Prediction	Score	Prediction	Score	Prediction	Score
-	D500Y	Affect	0	probably damaging	1	Deleterious	-4.572	effect	68
rs1203009272	G440R	Affect	0	probably damaging	1	Deleterious	-5.147	effect	90
rs104894854	W296R	Affect	0	probably damaging	1	Deleterious	-13.836	effect	98
rs1460413167	P212H	Affect	0	probably damaging	1	Deleterious	-7.77	effect	73
rs140848049	F208L	Affect	0	probably damaging	1	Deleterious	-5.913	effect	78
rs1295603457	C65Y	Affect	0	probably damaging	1	Deleterious	-7.572	effect	87
rs757475450	R39C	Affect	0	probably damaging	1	Deleterious	-4.575	effect	59

Table(1):DamagingorDeleteriousnsSNPsassociatedpredictedbyvarious softwares:

sub	SNPs&GO Prediction	RI	Probability	PANTHER Prediction	RI	Probability	PHD Prediction	RI	Probability	P-mut Prediction	Probability
W296R	Disease	7	0.858	Disease	10	0.99	Disease	9	0.925	Disease	0.89 (92%)
P212H	Disease	4	0.709	Disease	10	0.994	Disease	6	0.788	Disease	0.81 (89%)
C65Y	Disease	6	0.787	Disease	8	0.878	Disease	7	0.85	Disease	0.80 (89%)
R39C	Disease	3	0.63	Disease	8	0.903	Disease	4	0.687	Disease	0.73 (87%)

Table (2): Disease effect nsSNPs associated variations predicted by predicted by various softwares:

*RI: Reliability Index

 Table (3): stability analysis predicted by I-Mutant version 3.0 and MUPro:

dbSNP rs#	Sub	SVM2 Prediction Effect	RI	DDG Value Prediction	Mupro Prediction	Mupro Score
rs104894854	W296R	Decrease	8	-0.99	Decrease	-0.95856
rs1460413167	P212H	Decrease	8	-1.67	Decrease	-1.30249
rs1295603457	C65Y	Increase	0	-0.09	Decrease	-0.56309
rs757475450	R39C	Decrease	3	-0.74	Decrease	-0.28424

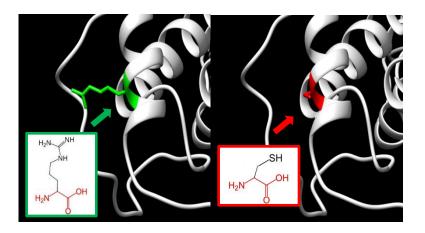


Figure (2): (rs757475450) (R39C) the amino acid Arginine changes to Cysteine at position 39. Illustrated by chimera (v 1.8) and project HOPE.

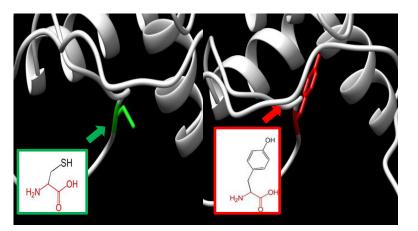


Figure (3): (rs1295603457) (C65Y) the amino acid Cysteine changes to Tyrosine at position 65. Illustrated by chimera (v 1.8) and project HOPE.

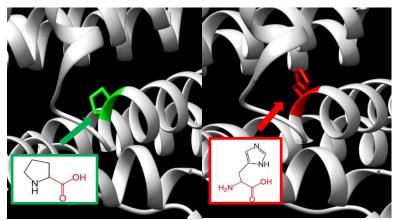


Figure (4): (rs1460413167) (P212H) the amino acid Proline changes to Histidine at position 65. Illustrated by chimera (v 1.8) and project HOPE.

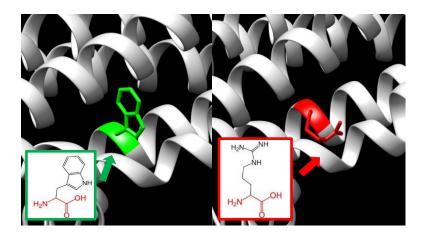
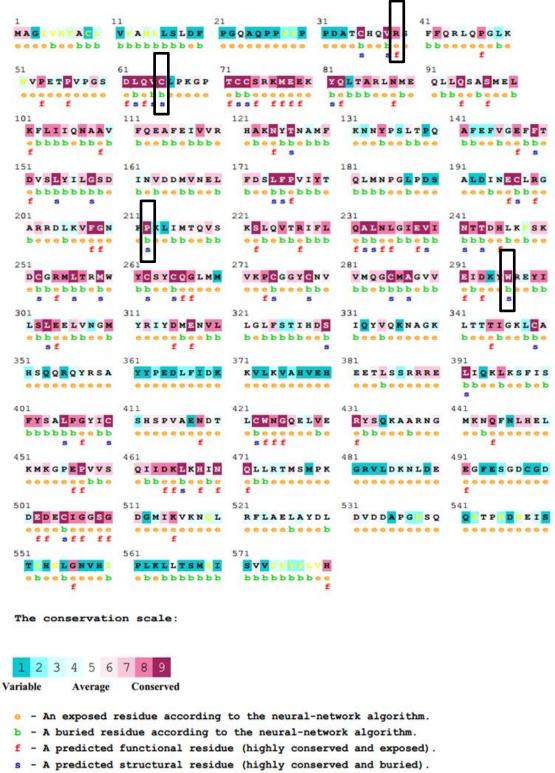


Figure (5): (rs104894854) (W296R) the amino acid Tryptophan changes to Arginine at position 296. Illustrated by chimera (v1.8) and project HOPE.

Hum an Chim panzee Bovine Dog Giant panda Horse Rabbit	R39 40 TCHQVRSFFQ	Human GS Chimpanzee Bovine Dog Giantpanda Horse	
Chim none o	P212	Chim nanzoo	290 VVEIDKYWR

Figure (6): *GPC3* Family of seven amino acid sequences demonstrating that the residues predicted to be mutated (indicated by red arrows) are evolutionarily conserved across species. Sequences Alignment was done by BioEdit (v7.2.5).



> - Insufficient data - the calculation for this site was performed on less than 10% of the sequences.

Figure (7): Unique and conserver regions in GPC3 protein were determined using Consurf web server.

4. Discussion:

Three novel mutations were found to have a damaging effect on the stability and function of the *GPC3* gene using bioinformatics analysis.

There is a study that has been reported which shows a missense mutation that causes SGBS, [19] which matches with this study findings. Some studies show association between *GPC3* gene and some types of liver cancer such as hepatocellular carcinoma.[30, 34, 54, 55] Therefore, this study can open the door for novel diagnostic biomarkers for hepatocellular carcinoma. Combination detection of serum GPC3 and pathogenic SNPs through clinical and genetic testing must be positively matched; this can enhance accuracy and efficiency of hepatocellular carcinoma diagnosis.

In addition it confirms that (W296R) is pathogenic; this result shows similarities with the result found earlier in dbSNPs/NCBI database. Furthermore, all these SNPs (P212H, C65Y, R39C) were retrieved as untested, in this study were found to be all pathogenic.

The most four deleterious SNPs were submitted to project HOPE which revealed that all they are located in a domain in the protein and thus might have a dynamic change in the structure and function of the protein and it may affect its ability to bind with its targets, in (R39C): The wild-type residue charge was positive, the mutant residue charge is neutral; Due to the loss of charge of the wild-type residue, this can cause loss of interactions with other molecules or residues. The mutant residue is more hydrophobic than the wild-type residue; in arrears of this change, this can result in loss of hydrogen bonds and/or disturb correct folding. In (C65Y): The wild-type residue is annotated in UniProt to be involved in a cysteine bridge, which is important for stability of the protein. Only cysteines can make these type of bonds, the mutation causes loss of this interaction and will have a severe effect on the 3D-structure of the protein. In (P212H): The wild-type residue is a proline. Prolines are known to be very rigid and therefore induce a special backbone conformation which might be required at this position. The mutation can disturb this special conformation. While in (W296R): This mutation is located in a region with known splice variants and matches a previously described variant, with the following description: Simpson-Golabi-Behmel syndrome 1 (SGBS1) [MIM: 312870]. The hydrophobicity of the wild-type and mutant residue differs; hydrophobic interactions, either in the core of the protein or on the surface, will be lost. While UCSF Chimera were used to visualize the amino acids change. (figures 2-5)

We also observed that, all the four SNPs were found in conserve region. (Figure 6) The seven amino acid sequences of GPC3 were retrieved from UniProt database.[56] While Sequence Alignments were done by BioEdit (v7.2.5). We also used ConSurf server; the

nsSNPs that are located at highly conserved amino acid positions have a tendency to be the most damaging nsSNPs. (Figure 7)

This study is the first in analysis approach while all other previous studies were in vitro, in vivo and whole exome sequencing. [57-60] It revealed three novel missense mutations that have a potential functional impact and may thus be used as diagnostic markers for overgrowth syndrome.[61] Finally some appreciations of wet lab techniques are recommended to support these findings.

5. Conclusion:

The present study could provide a novel insight into the molecular basis of overgrowth Syndrome by evidence from bioinformatics analysis. Three novel missense mutations (rs757475450, rs1295603457 and rs1460413167) that have a potential functional impact and may consequently can be used as diagnostic markers for overgrowth syndrome. As well as these SNPs can be used for the larger population-based studies of overgrowth syndrome.

Conflict of interest:

The authors declare no conflict of interest.

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