

1 **Mate choice confers direct benefits to females of *Anastrepha fraterculus***

2 **(Diptera: Tephritidae)**

3

4

5 Guillermo E. Bachmann^{1,2}, Francisco Devescovi^{1,2}, Ana L. Nussenbaum^{1,2}, Fabián H.

6 Milla¹, Todd E. Shelly³, Jorge L. Cladera¹, Patricia C. Fernández^{2,4}, María T. Vera^{2,5},

7 Diego F. Segura^{1,2*}

8

9

10 ¹Instituto de Genética “E.A. Favret”, Instituto Nacional de Tecnología Agropecuaria,
11 Hurlingham, Buenos Aires, Argentina.

12 ²Consejo Nacional de Investigaciones Científicas y Técnicas, Ciudad Autónoma de
13 Buenos Aires, Argentina.

14 ³United States Department of Agriculture, Animal and Plant Health Inspection Service,
15 Waimanalo, Hawaii, USA.

16 ⁴Estación Experimental Agropecuaria Delta del Paraná, Instituto Nacional de
17 Tecnología Agropecuaria, Campana, Buenos Aires, Argentina.

18 ⁵Facultad de Agronomía y Zootecnia, Universidad Nacional de Tucumán, San Miguel
19 de Tucumán, Tucumán, Argentina.

20

21 * corresponding author

22 E-mail: segura.diego@inta.gob.ar (DFS)

23

24

25 **Abstract**

26 Exposure to plant compounds and analogues of juvenile hormone (JH) increase male
27 mating success in several species of tephritid fruit flies. Most of these species exhibit a
28 lek mating system, characterized by active female choice. Although the pattern of
29 enhanced male mating success is evident, few studies have investigated what benefits, if
30 any, females gain via choice of exposed males in the lek mating system. In the South
31 American fruit fly, *Anastrepha fraterculus*, females mate preferentially with males that
32 were exposed to volatiles released by guava fruit or treated with methoprene (a JH
33 analogue). Here, we tested the hypothesis that female choice confers direct fitness
34 benefits in terms of fecundity and fertility. We first carried out mate choice experiments
35 presenting females with males treated and non-treated with guava volatiles or,
36 alternatively, treated and non-treated with methoprene. After we confirm female
37 preference for treated males, we compared the fecundity and fertility between females
38 mated with treated males and non-treated ones. We found that *A. fraterculus* females
39 that mated with males exposed to guava volatiles showed higher fecundity than females
40 mated to non-exposed males. On the other hand, females that mated methoprene-treated
41 males showed no evidence of direct benefits. Our findings represent the first evidence of
42 a direct benefit associated to female preference for males that were exposed to host fruit
43 odors in tephritid fruit flies. Differences between the two treatments are discussed in
44 evolutionary and pest management terms.

45 **Introduction**

46 Tephritid fruit flies (Diptera: Tephritidae) infest hundreds of different plant species,
47 including many economically important fruits [1]. One environmentally friendly control
48 strategy used against fruit fly pests is the sterile insect technique (SIT) [2], which
49 requires solid knowledge of the sexual behaviour of the target pest [3]. SIT is based on
50 the ability of mass-released, sterile males to mate with fertile, wild females and hence
51 induce sterility in the pest population [4]. Many species of tephritid fruit flies exhibit lek
52 mating systems characterized by female choice, and this has prompted considerable
53 attention on the factors that influence male mating success [5,6]. The collective outcome
54 of this research has been the identification of several pre-release treatments that boost
55 the sexual competitiveness of male tephritids [7-10] including: pre-release diet
56 containing a proteinaceous source [11-20]; exposure of males to semiochemicals
57 [9,10,21,22]; and the use of methoprene, a juvenile hormone (JH) analogue, to both
58 boost male mating success and accelerate their sexual maturation [7,17,18,23].

59 Although the drive to improve SIT has, in many cases, identified male traits
60 associated with mating success, little is known about the ecological and evolutionary
61 forces that shape female mate preferences, particularly the potential fitness benefits
62 associated with mate choice [24]. Kirkpatrick and Ryan [25] and Wyatt [26] proposed
63 that female choice is generally associated with direct benefits, such as increased
64 fecundity and longevity, reduced risk of predation, or access to material resources under
65 male control [27-29]. Simultaneously, females may gain indirect benefits if mate choice
66 positively affects the fitness of their offspring, such as those explained under the sexy
67 son and good genes hypotheses [30-33]. For tephritids, female choice has been
68 associated with direct and indirect benefits [34,35]. In particular, the association

69 between female benefits and mate selection mediated via feeding or exposure of males
70 to plant-borne compounds has been investigated in four tephritid species: *Bactrocera*
71 *dorsalis* (Hendel) [36,37]; *Bactrocera tryoni* (Froggatt) [38,39]; *Ceratitidis capitata*
72 (Wiedemann) [13,40]; and *Zeugodacus cucurbitae* (Coquillett) [24]. Although females
73 of this species showed a preference for males either exposed or fed with plant-derived
74 compounds, only Kumaran *et al.* [38,39] found evidence of direct and indirect benefits,
75 respectively, for *B. tryoni* females.

76 The South American fruit fly, *Anastrepha fraterculus* (Wiedemann), is a
77 polyphagous species that attacks more than 100 species of fruit plants [41], many of
78 which have a high commercial value. Vera *et al.* [42] found an increase in the mating
79 success of *A. fraterculus* males following exposure to the volatiles of guava fruits
80 (*Psidium guajava* L.), a native plant and one of their main hosts in the wild. This
81 phenomenon was later confirmed and extended by Bachmann *et al.* [43], who also
82 found that males exposed to guava fruit volatiles released larger amounts of sex
83 pheromone and performed courtship behaviours more frequently than non-exposed
84 males. Similarly, topical applications of methoprene conferred a mating advantage to
85 the males which also seems to be associated to larger amounts of pheromone being
86 released by methoprene-treated males when competing with non-treated males [44].

87 Despite evidence showing that guava fruit volatiles and mimics of natural
88 compounds (like methoprene) enhance male mating success in different *Anastrepha*
89 species, no prior investigation has measured the potential fitness benefits gained by
90 females that mate with those enhanced males. Even though guava volatiles are natural
91 compounds and methoprene is synthetic, they both stimulate males to release larger
92 amounts of pheromone and increased male mating success [43,44]. Furthermore, plants

93 produce several natural analogues of the juvenile hormone (termed juvenoids), which
94 protect them from herbivore larvae [45-47]. According to Bede & Tobe [48], some
95 species of insects have adapted to consume these juvenoids to increase their own
96 reproductive output. So, even though methoprene is a synthetic compound, it could still
97 trigger natural responses in males (enhanced signalling) and females (attraction to such
98 signalling). In the present work, we evaluated whether the preference of *A. fraterculus*
99 females for males exposed to guava volatiles and those treated with methoprene derive
100 from direct benefits in terms of increased fecundity and fertility. Because in *A.*
101 *fraterculus* longer copulations result in longer refractory periods [49], which can
102 decrease the need for future mates and consequently the costs in terms of energy and re-
103 exposure to predators [50], we also evaluated copula duration as potential direct benefit
104 to females.

105

106 **Materials and Methods**

107 **Biological material**

108 *Anastrepha fraterculus* flies were obtained from a laboratory colony kept at Instituto de
109 Genética E. A. Favret (IGEAF) which was originally established at Estación
110 Experimental Agroindustrial Obispo Colombres (Tucumán, Argentina) in 1997 with
111 pupae obtained from infested guavas collected in Tafi Viejo (26°43'25''S 65°16'43''W,
112 Tucumán, Argentina) [51]. Rearing followed standard procedures using an artificial diet
113 based on yeast, wheat germ, sugar, and agar for larvae [52] and a mixture of sugar,
114 hydrolysed yeast (MP Biomedicals, San Francisco, CA, USA), hydrolysed corn
115 (ARCOR, Tucumán, Argentina) (4:1:1 ratio) and vitamin E (Parafarm, Buenos Aires,

116 Argentina) for adults [53]. Flies used in the tests were virgin and sexually mature (males
117 were > 10 days-old; females were > 14 days-old) [54] and were kept under controlled
118 environmental conditions (24 ± 2 °C, $70 \pm 10\%$ RH, and a 12L: 12D photoperiod).

119 One day after adult emergence flies were sorted by sex, transferred to plastic
120 containers, and provided food and water. Females were placed in 1 L plastic cylindrical
121 containers (15 cm tall, 12 cm in diameter) in groups of 25 individuals and fed with the
122 standard diet. Males were placed in 21 L plastic containers (37 x 28 x 21 cm) in groups
123 of 100 individuals and fed with sugar and brewer's yeast (CALSA, Tucumán,
124 Argentina) (3:1 ratio).

125

126 **Experiments**

127 **Experiment 1. *Mating and reproductive output of females offered males*** 128 ***exposed or not exposed to guava volatiles***

129 In order to determine whether the preference of *A. fraterculus* females for guava-
130 exposed males [44] is associated with direct fitness benefits in terms of mating and
131 reproductive output, females were first given the choice to mate with guava-exposed or
132 non-exposed males. Then, we determined differences in mating and reproductive
133 parameters between females that selected exposed or non-exposed males.

134 Treated males were exposed to guava volatiles without physical access to the fruit
135 following procedures of Bachmann *et al.* [43]. Non-exposed males were kept under the
136 same environmental conditions but in a different room and had no exposure to guava
137 odours. After exposure, males were kept in the 21 L plastic containers in separate rooms
138 and under the same conditions described above. In the mating test, a total of 401
139 experimental arenas were established, each consisting of three virgin flies: one exposed

140 male, one non-exposed male of the same age, and one female. Males were 12-14 d old; ;
141 whereas females were 14-18 d old. These experimental arenas have been extensively
142 used in *A. fraterculus* as a valid experimental approach to study female mate choice [42-
143 44,55,56]. Males were marked on their thorax with a dot of non-toxic, water-based paint
144 for identification [54]. Randomly assigned colours identified different male treatments.
145 Males and females were released in the experimental arenas early in the morning under
146 semi-darkness. Males were released 15 min before females. Once all arenas were set up,
147 fluorescent lights were turned on, and the occurrence of mating pairs was monitored
148 continuously for 2 h during the natural period of mating activity for Argentinean
149 populations of *A. fraterculus* (9 – 11 am) [54,57]. Whenever a couple was detected,
150 male colour and time at which copulation started and ended were recorded. The
151 recording of copula start and end times was done continuously, and each mating couple
152 was checked every 2-3 minutes. Due to inadvertent errors in recording times, 4
153 replicates were excluded from the analysis of mating duration. Experiments were
154 conducted under laboratory conditions (25 ± 1 °C and $70 \pm 20\%$ RH). Illumination was
155 provided by fluorescent tubes and natural light coming from a window.

156 After the mating test, mated females were transferred to 3 L glass flasks (with
157 water and food) in groups of three according to the type of male mated. We used groups
158 of females instead of solitary individuals because several studies indicate that the
159 presence of conspecific females stimulates the oviposition [58-60]. Each flask with
160 three females was considered a replicate, with 25 and 22 replicates for females mated to
161 exposed or non-exposed males, respectively. Every 3 days each replicate was provided
162 an oviposition substrate (hereafter, oviposition unit) that consisted of a cylindrical
163 plastic vial (2 cm tall, 2.5 cm in diameter) filled with water coloured with edible red dye
164 (Fleibor, Tablada, Buenos Aires, Argentina) and covered with Parafilm M (Pechiney

165 Plastic Packaging, Chicago, Illinois, USA). This oviposition unit is normally used in the
166 laboratory rearing of *A. fraterculus* at IGFAF. After 24 h, the oviposition unit was
167 removed, and the eggs were recovered using a plastic pipette and then transferred onto a
168 black filter paper. This was, in turn, placed inside a plastic Petri dish (2 cm tall, 8 cm in
169 diameter) on top of a wet cloth and then covered with its lid. The number of eggs from
170 each oviposition unit was counted under a stereoscopic microscope (20x). Petri dishes
171 were then placed inside an incubator at 25 ± 1 °C and $70 \pm 10\%$ RH for 4 days to allow
172 embryonic development. After this time, the numbers of hatched and unhatched eggs
173 were counted under a stereoscopic microscope (20x). This entire process was repeated
174 nine times with each replicated during 27 days, thus covering the peak period of egg-
175 laying in this species [61]. Whenever a dead female was detected, it was removed from
176 the flask and recorded, thus allowing computation of ovipositions *per capita* for each
177 day of egg collection. Due to logistic problems with the incubator, 7 and 1 replicates
178 coming from females mated with exposed and non-exposed males, respectively, were
179 discarded and not considered in the data analysis.

180

181 **Experiment 2. Mating and reproductive output of females offered males**
182 ***treated or not treated with methoprene***

183 To determine whether the preference of *A. fraterculus* females for methoprene-treated
184 males [44] is associated with a direct benefit, we followed a protocol similar to that
185 described for Experiment 1.

186 The procedure for methoprene application followed Teal *et al.* [62]. Briefly, on
187 the day of emergence males were topically treated by applying 1 μ L of a solution of
188 methoprene dissolved in acetone (5 μ g/ μ l) into their torax. Females, treated males, and
189 non-treated males were kept in separate rooms under the same environmental and

190 feeding conditions as described above. In the mating tests a total of 166 experimental
191 arenas were set up (4 replicates were excluded from the analysis of mating duration due
192 to mistakes during the recording of duration times). At the day of the mating test, males
193 were 12 d old and females were 14-18 d old. Afterwards, mated females were
194 transferred in groups of three to cylindrical, 1 L plastic containers. Fecundity and
195 fertility were assessed following the same procedures as described for experiment 1,
196 except that oviposition units were offered seven times to each replicate, covering a total
197 of 21 days. The number of replicates was 24 for females mated to methoprene-treated
198 males and 16 for females mated to non-treated males.

199

200 **Data analysis**

201 The numbers of copulations achieved by guava-exposed and non-exposed males
202 (experiment 1) or methoprene-treated and non-treated males (experiment 2) were
203 compared by means of a G test of goodness of fit to an equal proportion hypothesis. The
204 latency to mate (i.e., time elapsed between female release and mating), the mating
205 duration (i.e., time elapsed between the start and end of mating), the overall fecundity
206 (i.e., number of eggs laid across all the egg collections), and the fertility (i.e., average of
207 hatch rate across all egg collections) were compared between females mated to treated
208 or non-treated males by means of a t-test for independent samples. Assumptions of
209 normality of the residuals and homoscedasticity were checked prior to each test. In
210 order to meet the homoscedasticity assumption, mating duration from experiment 1 and
211 fertility in experiment 2 were ln- and logit-transformed, respectively. Also, fertility was
212 logit-transformed in experiment 2. Finally, we carried out a series of survival analyses
213 (Kaplan-Meier estimation, Log-rank test) to evaluate the temporal pattern of female
214 fecundity after mating with exposed or non-exposed males (experiment 1) or treated or

215 non-treated males (experiment 2). STATISTICA 7 [63] and GraphPad Prism 6 [64]
216 were used for statistical analyses and preparation of figures.

217

218 **Results**

219 **Experiment 1. *Mating and reproductive output of females*** 220 ***offered males exposed or not exposed to guava volatiles***

221 A total of 401 mating arenas were established of which 387 resulted in mating. Females
222 mated significantly more often with males exposed to guava than with non-exposed
223 males ($G = 19.73$, $N = 387$, $d.f. = 1$, $p < 0.001$) (Fig 1). The latency to mate was not
224 affected by the type of male chosen for mating ($t = 0.284$, $d.f. = 385$, $p = 0.776$) (Fig
225 2a). Mating duration, on the other hand, was longer in matings that involved males
226 exposed to guava volatiles ($t = 2.626$, $d.f. = 381$, $p < 0.01$) (Fig 2b). Female fecundity
227 was significantly different between females mated with exposed and non-exposed males
228 ($t = 2.145$, $d.f. = 45$, $p = 0.037$), as females mated with guava exposed males laid
229 significantly more eggs (Fig 2c). Fertility was not statistically different between the two
230 types of females ($t = 0.372$, $d.f. = 37$, $p = 0.712$) (Fig 2d).

231 **Fig 1. Percentage of matings obtained by males exposed or non-exposed to guava**
232 **volatiles. Numbers above bars represent numbers of matings. * statistically**
233 **significant difference ($\alpha = 0.05$).**

234 **Fig 2. Latency to mate (a), mating duration (b), fecundity (*per capita*) (c) and**
235 **fertility (d) for females mated to males exposed or non-exposed to guava volatiles**
236 **(mean and SE). * statistically significant difference ($\alpha = 0.05$); n.s. not statistically**
237 **significant difference.**

238

239 The temporal pattern of egg-laying was independent of the type of male chosen
240 for mating (50% of the total eggs deposited by the 7th irrespective of male type) ($\chi^2 =$
241 1.02, d.f. = 1, $p = 0.312$) (Fig 3).

242 **Fig 3. Temporal patterns of egg-laying by females mated to males exposed or non-**
243 **exposed to guava volatiles.**

244

245 **Experiment 2. Mating and reproductive output of females**
246 ***offered males treated or not treated with methoprene***

247 Successful matings were recorded in 152 out of 166 mating arenas. Females mated
248 significantly more often with males treated with methoprene than with non-treated
249 males ($G = 11.76$, $N = 152$, d.f. = 1, $p < 0.001$) (Fig 4). However, male type had no
250 effect on any of the measured variables (latency to mate: $t = 0.357$, d.f. = 150, $p =$
251 0.722 ; mating duration: $t = 0.257$, d.f. = 146, $p = 0.797$; fecundity: $t = 0.715$, d.f. = 38, p
252 $= 0.476$; fertility: $t = 1.100$, d.f. = 38, $p = 0.278$) (Fig 5a-d). Likewise, the temporal
253 pattern of oviposition was independent of the type of male selected by the female (50%
254 of the total eggs deposited by the 5th egg collection for both groups) ($\chi^2 = 1.61$, d.f. = 1,
255 $p = 0.205$) (Fig 6).

256 **Fig 4. Percentage of matings obtained by males treated or non-treated with**
257 **methoprene. Numbers above bars represent numbers of matings. * statistically**
258 **significant difference ($\alpha = 0.05$).**

259 **Fig 5. Latency to mate (a), mating duration (b), fecundity (*per capita*) (c) and**
260 **fertility (d) for females mated to males treated or non-treated with methoprene**
261 **(mean and SE). n.s. not statistically significant difference ($\alpha = 0.05$).**

262 **Fig 6. Temporal pattern of egg deposition by females mated to males treated or**
263 **non-treated with methoprene.**

264

265 **Discussion**

266 Documenting female preference based on particular male traits is more easily, and thus
267 more frequently, accomplished than demonstrating fitness benefits to females accruing
268 from such preferences. Vera *et al.* [42] and Bachmann *et al.* [43,44] observed that *A.*
269 *fraterculus* females prefer males that had been exposed to volatiles of guava fruit (one
270 of its main hosts) or treated with methoprene (an artificial analogue of JH). Here, we
271 verified these patterns of sexual selection and investigated whether female preferences
272 can be explained on the basis of a direct fitness benefit. We found that females that
273 selected males exposed to guava volatiles had higher fecundity than those selecting non-
274 exposed males. Therefore, such enhanced egg output is consistent with the existence of
275 a direct fitness benefit. In other words, preference for males exposed to guava volatiles
276 represents an adaptive, fitness-based decision. Additionally, matings that involved
277 guava exposed males lasted longer, which, as discussed below, may also be considered
278 beneficial for the females. On the other hand, for methoprene treatment, we did not
279 detect any association between preference and fitness benefits for females. Female
280 latency to mate did not differ with male status for both guava volatile and methoprene
281 treatments. Thus, although exposure to guava volatiles and application of methoprene
282 boosted male mating success, neither treatment stimulated more rapid mating decisions
283 by females.

284 The difference in fecundity between females mated to guava-exposed and non-
285 exposed males was constant over time, which indicates that the benefit derived from

286 mating with exposed males was long-lasting and not restricted to the beginning of the
287 oviposition period. The higher fecundity observed here is consistent with Kumaran *et al.*
288 [38], who recorded increased fecundity in *B. tryoni* females mated to males that were
289 previously exposed to zingerone, a floral compound produced by orchids, that enhances
290 male mating success. Particularly, our findings constitute the first evidence of such
291 mechanism involving volatiles from the host fruit. For tephritids, that study and the
292 present one provide the only clear evidence of a direct benefit associated with female
293 mate choice, where female preference is mediated by male exposure to plant-derived
294 compounds. Other related studies on tephritids found no support for this phenomenon
295 [22]. For example, females of *B. dorsalis* mate preferentially with methyl eugenol-fed
296 males, but they did not show an increase in their fecundity, fertility, or survival [37,65].
297 Similarly, *C. capitata* females did not exhibit any reproductive or survival benefits after
298 mating with preferred, ginger root oil treated males [13].

299 One mechanism by which fecundity might have increased is the acquisition of
300 higher quality male accessory gland products (AGPs) during copulation, which occurs
301 in many insects including tephritids [66-68]. There is a large variety of physiologically
302 active substances, such as proteins and juvenile hormone in the ejaculate, that may
303 inhibit female remating propensity and induce egg maturation [66,69,70]. The
304 perception of guava volatiles by males could act as an indicator of host availability and
305 alter male reproductive physiology, stimulating the production of AGPs that would be
306 transferred in higher amounts to the females with their concomitant impact on
307 fecundity. The possibility that males spend more energetic sources in reproduction (e.g.
308 signalling and AGPs production) when a host is present can be a reasonable
309 explanation, but it is, of course, conjectural, and studies on the effects of guava
310 exposure on male reproductive physiology are required.

311 In contrast to the guava treatment, mating with methoprene-treated males did not
312 result in increased fecundity, fertility, or copulation duration. The lack of evidence of
313 direct benefits can be interpreted in different ways. First, because methoprene is a
314 synthetic compound, it may be possible that looking at the behavior of females from an
315 evolutionary perspective (i.e. relating their preference with direct fitness benefits) is
316 misleading. In this scenario, methoprene would induce a higher rate of pheromone
317 release in males, which triggers female acceptance over non-treated males [44] by
318 exploiting females' sensory channel with no reward in terms of their own potential
319 fecundity. However, juvenile hormone analogues do exist in nature [45-47] and some
320 insects consume them, increasing their reproductive output [48]. Kumaran *et al.* [38]
321 found that females of *B. tryoni* obtained a direct benefit by mating males exposed to
322 zingerone (a natural compound) as well as males exposed to cuelure (a synthetic
323 compound). This argues against the idea that the lack of benefits associated to
324 methoprene lays on the fact that methoprene is a synthetic compound. Methoprene is
325 similar to JH in chemical structure and, more importantly, its role on *A. fraterculus*
326 seems to replicate that of JH [55,71,72]. Therefore, the strong preference of females for
327 methoprene-treated males could, still, be associated to other benefits, yet unidentified.
328 First, because males treated with methoprene release more pheromone than no-treated
329 males, females could be obtaining a direct benefit if treated males are more easily
330 located, consequently reducing predation risks and mate location costs [13]. Second,
331 indirect (genetic) benefits may be involved in female preference for methoprene-treated
332 males. In *B. tryoni*, Kumaran *et al.* [39] found that the sons of males that were treated
333 with cuelure and zingerone detected and located these chemicals more effectively than
334 sons of non-exposed males, thus showing evidence of a potential "sexy son"
335 mechanism. Potential indirect benefits, like those reported for *B. tryoni* [38,39], should

336 also be studied for *A. fraterculus* females mated not only with methoprene-treated males
337 but also in females mated with guava-exposed males, as direct and indirect effects can
338 both be influencing female choice.

339 Copula lasted longer for guava exposed males than for non-exposed ones.
340 *Anastrepha fraterculus* females appear to remate primarily to replenish sperm [73], and
341 this tendency is negatively associated to copula duration [49]. Thus, long copulations
342 may be adaptive, because the transfer of large amounts of sperm would eliminate or
343 delay the expenditure of energy and time to find males. Because predation risk increases
344 during courtship and mating [50] delaying remating would also be beneficial in terms of
345 survival. These benefits may, of course, be offset to some degree by any increase in
346 predation risk during a single but lengthy copulation and the fact that overall genetic
347 variability of the progeny is expected to be lower. In the case of methoprene, copula
348 duration did not differ between treated and non-treated males. This agrees with Haq et
349 al. [19] for *Z. cucurbitae* and shows a general lack of other types of benefits associated
350 to methoprene.

351 The preference of females for males treated with sexual enhancers, together with a
352 resulting increased fecundity could have practical implications in the context of SIT. In
353 a mass rearing facility, an increase in fecundity would mean a higher yield which
354 directly translates into lower costs of maintenance (because fewer reproductive adults
355 would be needed). Also, if enhanced, but sterile, males were able to induce sterility in
356 wild females, then a higher proportion of the reproductive output of the wild population
357 would be unviable. Nonetheless, in order to extend our results to the context of SIT,
358 further research is needed. For instance, the impact of irradiation on both the
359 enhancement of males mating success and females fecundity were not assessed.

360 To conclude, previous work [43] showed that the preference of females for males
361 exposed to guava volatiles could be explained at a proximal level by a higher rate of
362 sexual displays and sex pheromone release by exposed males. Here, at least for guava
363 volatiles, we found evidence that female preference could also be explained at an
364 evolutionary level, because females that mate with guava-exposed males obtained a
365 direct benefit in terms of increased fecundity. Our results contribute to a better
366 understanding of the mechanisms related to mate choice and their evolutionary
367 implications. However, the ultimate (physiological) causes of an increased fecundity
368 after mating with a sexually enhanced male still remain unknown and represent an
369 interesting field of study. It is worth mentioning that, in all the experiments of the
370 present study, females were fed with diet of high nutritional quality and presumably
371 met, to a large extent, their nutritional and physiological needs (e.g., oocytes
372 development). Aluja et al. [11] found that *Anastrepha ludens* and *Anastrepha obliqua*
373 females fed on protein diets developed more oocytes than those fed without protein. It is
374 thus possible that a rich diet actually reduced the positive effects of mating with guava-
375 exposed males and that females feeding on a lower quality diet, as expected in the wild,
376 would show an even greater increase in fecundity than found here. The interaction
377 between female nutritional status and potential benefits gained through mate choice
378 should be considered in future studies.

379

380 **Acknowledgements**

381 The authors are grateful to our colleagues at IGEAF, INTA, for their collaboration
382 during the tests. These studies were funded by the FAO/IAEA and Ministerio de

383 Ciencia, Tecnología e Innovación Productiva of Argentina (through Research Contract
384 16483 and FONCYT PICT 2013 - 054 to JLC and DFS, respectively).

385

386 **References**

387 1. White IM, Elson-Harris MM. Fruit Flies of Economic Significance: Their
388 Identification and Bionomics (1992). CAB International.

389 2. Knipling EF. Possibilities of insect control or eradication through the use of sexually
390 sterile males. *J Econ Entomol.* 1955; 48(4): 459-462.

391 3. Robinson AS, Hendrichs J. Prospects for the future development and application of
392 the sterile insect technique. In: Dyck VA, Hendrichs J, Robinson AS, editors. *Sterile
393 Insect Technique. Principles and Practice in Area-Wide Integrated Pest Management.*
394 Springer; 2005. pp. 727-760.

395 4. Lance DR, McInnis DO. Biological basis of the sterile insect technique. In: Dyck
396 VA, Hendrichs J, Robinson AS, editors. *Sterile Insect Technique. Principles and
397 Practice in Area-Wide Integrated Pest Management.* Springer; 2005. pp. 69-94.

398 5. Hendrichs J, Robinson AS, Cayol JP, Enkerlin W. Medfly areawide sterile insect
399 technique programmes for prevention, suppression or eradication: the importance of
400 mating behavior studies. *Fla Entomol.* 2002; 85(1): 1-13.

401 6. Robinson AS, Cayol JP, Hendrichs J. Recent findings on medfly sexual behavior:
402 implications for SIT. *Fla Entomol.* 2002; 85(1): 171-181.

403 7. Teal PEA, Gomez-Simuta Y, Dueben BD, Holler TC, Olson S. Improving the
404 efficacy of the sterile insect technique for fruit flies by incorporation of hormone and
405 dietary supplements into adult holding protocols. In: Vreysen MJB, Robinson AS,
406 Hendrichs J, editors. *Area-Wide Control of Insect Pests;* 2007. pp. 163-173.

- 407 8. Yuval B, Maor M, Levy K, Kaspi R, Taylor P, Shelly T. Breakfast of champions or
408 kiss of death? Survival and sexual performance of protein-fed, sterile Mediterranean
409 fruit flies (Diptera: Tephritidae). *Fla Entomol.* 2007; 90(1): 115-122.
- 410 9. Papadopoulos NT, Kouloussis NA, Katsoyanos BI. Effect of plant chemicals on the
411 behavior of the Mediterranean fruit fly. In: Sugayama RL, Zucchi RA, Ovruski SM,
412 Sivinski J, editors. Proceedings of the 7th International Symposium on fruit Flies of
413 Economic Importance. Press Color Graficos Especializados, Brotas, Bahia, Brazil. 2008
- 414 10. Shelly TE, Cowan AN, Edu J, Pahio E. Mating success of male Mediterranean fruit
415 flies following exposure to two sources of α -copaene, manuka oil and mango. *Fla*
416 *Entomol.* 2008a; 91(1): 9-15.
- 417 11. Aluja M, Díaz-Fleischer F, Papaj DR, Lagunes G, Sivinski J. Effects of age, diet,
418 female density, and the host resource on egg load in *Anastrepha ludens* and *Anastrepha*
419 *obliqua* (Diptera: Tephritidae). *J Insect Physiol.* 2001; 47(9): 975-988.
- 420 12. Yuval B, Kaspi R, Field SA, Blay S, Taylor P. Effects of post-teneral nutrition on
421 reproductive success of male Mediterranean fruit flies (Diptera: Tephritidae). *Fla*
422 *Entomol.* 2002; 85(1): 165-170.
- 423 13. Shelly TE. Does mating with ginger root oil-exposed males confer fitness benefits to
424 female Mediterranean fruit flies, *Ceratitis capitata* (Diptera: Tephritidae)? *Proc Hawaii*
425 *Entomol Soc.* 2005; 37: 65-71.
- 426 14. Pérez-Staples D, Prabhu V, Taylor PW. Post-teneral protein feeding enhances
427 sexual performance of Queensland fruit flies. *Physiol Entomol.* 2007; 32(3): 225-232.
- 428 15. Pérez-Staples D, Weldon CW, Smallridge C, Taylor PW. Pre-release feeding on
429 yeast hydrolysate enhances sexual competitiveness of sterile male Queensland fruit flies
430 in field cages. *Entomol Exp Appl.* 2009; 131(2): 159-166.

- 431 16. Pereira R, Sivinski J, Teal PE. Influence of methoprene and dietary protein on male
432 *Anastrepha suspensa* (Diptera: Tephritidae) mating aggregations. J Insect Physiol.
433 2009; 55(4): 328-335.
- 434 17. Pereira R, Sivinski J, Teal PE. Influence of a juvenile hormone analog and dietary
435 protein on male *Anastrepha suspensa* (Diptera: Tephritidae) sexual success. J Econ
436 Entomol. 2010; 103(1): 40-46.
- 437 18. Haq I, Cáceres C, Hendrichs J, Teal P, Wornoyaporn V, Stauffer C, et al. Effects of
438 the juvenile hormone analogue methoprene and dietary protein on male melon fly
439 *Bactrocera cucurbitae* (Diptera: Tephritidae) mating success. J Insect Physiol. 2010;
440 56(11): 1503-1509.
- 441 19. Haq I, Cáceres C, Liedo P, Soriano D, Jessup A, Hendrichs J, et al. Effect of
442 methoprene application, adult food and feeding duration on male melon fly starvation
443 survival. Appl Entomol Zool. 2013; 137(s1): 61-68.
- 444 20. Gavriel S, Jurkevitch E, Gazit Y, Yuval B. Bacterially enriched diet improves sexual
445 performance of sterile male Mediterranean fruit flies. Appl Entomol Zool. 2011; 135(7):
446 564-573.
- 447 21. Tan KH, Nishida R, Jang EB, Shelly TE. Pheromones, male lures, and trapping of
448 tephritid fruit flies. In: Shelly T, Epsky N, Jang E, Reyes-Flores J, Vargas R,
449 editors. Trapping and the Detection, Control, and Regulation of Tephritid Fruit
450 Flies. 2014. pp. 15-74. Springer Netherlands.
- 451 22. Segura DF, Belliard SA, Vera MT, Bachmann GE, Ruiz MJ, Jofre-Barud F,
452 Fernández PC, López ML, Shelly TE. Plant Chemicals and the Sexual Behavior of Male
453 Tephritid Fruit Flies. Ann Entomol Soc Am. 2018; 111: 239–264.

- 454 23. Collins SR, Reynolds OL, Taylor PW. Combined effects of dietary yeast
455 supplementation and methoprene treatment on sexual maturation of Queensland fruit
456 fly. *J Insect Physiol.* 2014; 61: 51-57.
- 457 24. Shelly TE, Nishimoto J. Does female mate choice confer direct fitness benefits?
458 Results from a tephritid fruit fly. *Ann Entomol Soc Am.* 2016; 110(2): 204-211.
- 459 25. Kirkpatrick M, Ryan MJ. The evolution of mating preferences and the paradox of
460 the lek. *Nature.* 1991; 350(6313): 33-38.
- 461 26. Wyatt TD. *Pheromones and Animal Behaviour: Communication by Smell and*
462 *Taste.* Cambridge university press (2003).
- 463 27. Gwynne DT. Courtship feeding increases female reproductive success in
464 bushcrickets. *Nature.* 1984; 307(5949): 361-363.
- 465 28. Reinhold K. Paternal investment in *Poecilimon veluchianus* bushcrickets: beneficial
466 effects of nuptial feeding on offspring viability. *Behav Ecol Sociobiol.* 1999; 45(3):
467 293-299.
- 468 29. Arnqvist G, Nilsson T. The evolution of polyandry: multiple mating and female
469 fitness in insects. *Anim Behav.* 2000; 60(2): 145-164.
- 470 30. Andersson MB. *Sexual Selection.* Princeton University Press. 1994.
- 471 31. Johnstone RA. Sexual selection, honest advertisement and the handicap principle:
472 reviewing the evidence. *Biol Rev.* 1995; 70(1): 1-65.
- 473 32. Ryan MJ. Sexual selection and mate choice. In: Krebs JR, Davies NB, editors.
474 *Behavioural ecology: an evolutionary approach.* 4th edition. Oxford: Blackwell Science;
475 1997. pp. 179–202.
- 476 33. Fedorka KM, Mousseau TA. Material and genetic benefits of female multiple
477 mating and polyandry. *Anim Behav.* 2002; 64(3): 361-367.

- 478 34. Kasuya E. Female mate preference and offspring fitness in the melon fly. *Ecol Res.*
479 1992; 7(3): 277-281.
- 480 35. Briceño RD, Eberhard WG. Possible fisherian changes in female mate-choice
481 criteria in a mass-reared strain of *Ceratitis capitata* (Diptera: Tephritidae). *Ann Entomol*
482 *Soc Am.* 2000; 93(2): 343-345.
- 483 36. Shelly TE. Fecundity of female oriental fruit flies (Diptera: Tephritidae): effects of
484 methyl eugenol-fed and multiple mates. *Ann Entomol Soc Am.* 2000b; 93(3): 559-564.
- 485 37. Shelly TE, Nishida R. Larval and adult feeding on methyl eugenol and the mating
486 success of male oriental fruit flies, *Bactrocera dorsalis*. *Entomol Exp Appl.* 2004;
487 112(2): 155-158.
- 488 38. Kumaran N, Balagawi S, Schutze MK, Clarke AR. Evolution of lure response in
489 tephritid fruit flies: phytochemicals as drivers of sexual selection. *Anim Behav.* 2013;
490 85(4): 781-789.
- 491 39. Kumaran N, Clarke AR. Indirect effects of phytochemicals on offspring
492 performance of Queensland fruit fly, *Bactrocera tryoni* (Diptera: Tephritidae). *Appl*
493 *Entomol Zool.* 2014; 138(5): 361-367.
- 494 40. Morelli R, Paranhos BJ, Coelho AM, Castro R, Garziera L, Lopes F, et al. Exposure
495 of sterile Mediterranean fruit fly (Diptera: Tephritidae) males to ginger root oil reduces
496 female remating. *Appl Entomol Zool.* 2013; 137(s1): 75-82.
- 497 41. Norrbom AL. Host plant database for *Anastrepha* and *Toxotrypana* (Diptera:
498 Tephritidae: Toxotrypanini). *Diptera Data Dissemination Disk*, 2. 2004.
- 499 42. Vera MT, Ruiz MJ, Oviedo A, Abraham S, Mendoza M, Segura DF, et al. Fruit
500 compounds affect male sexual success in the South American fruit fly, *Anastrepha*
501 *fraterculus* (Diptera: Tephritidae). *Appl Entomol Zool.* 2013; 137(s1): 2-10.

- 502 43. Bachmann GE, Segura DF, Devescovi F, Juárez ML, Ruiz MJ, Vera MT, et al. Male
503 sexual behavior and pheromone emission is enhanced by exposure to guava fruit
504 volatiles in *Anastrepha fraterculus*. PLOS ONE. 2015; 10(4): e0124250.
505 <https://doi.org/10.1371/journal.pone.0129523>
- 506 44. Bachmann GE, Devescovi F, Nussenbaum AL, Cladera JL, Fernández PC, Vera
507 MT, et al. Male sexual enhancement after methoprene treatment in *Anastrepha*
508 *fraterculus* (Diptera: Tephritidae): A sustained response that does not fade away after
509 sexual maturation. J. Insect. Physiol. 2017; 101: 7-14.
510 <https://doi.org/10.1016/j.jinsphys.2017.06.009>
- 511 45. Arora R, Singh G, Kailey JS, Kalsi SS. Biological activity of terpenoid lactones as
512 juvenile hormone analogues against the mustard aphid, *Lipaphis*
513 *erysimi*. Phytoparasitica. 1982; 10(1): 57-60.
- 514 46. Balandrin MF, Klocke JA, Wurtele ES, Bollinger WH. Natural plant chemicals:
515 sources of industrial and medicinal materials. Science. 1985; 228(4704): 1154-1160.
- 516 47. Singh G, Kalsi PS. 1996. Juvenile hormone activity of some plant oils against the
517 mustard aphid, *Lipaphis erysimi* (Halt.). In: Narwal, Tauro, editors. Allelopathy in Pest
518 Management for Sustainable Agriculture. Scientific publishers, Jodhpur, India. 1996.
519 pp. 175-178.
- 520 48. Bede JC, Tobe SS. Insect Juvenile Hormones in Plants. In: Rahman AU, editor.
521 Studies in Natural Products Chemistry. Elsevier Press, Amsterdam, Netherlands. 2000;
522 22: 369-418.
- 523 49. Abraham S, Goane L, Cladera J, Vera MT. Effects of male nutrition on sperm
524 storage and remating behavior in wild and laboratory *Anastrepha fraterculus* (Diptera:
525 Tephritidae) females. J Insect Physiol. 2011a; 57(11): 1501-1509.

- 526 50. Hendrichs J, Katsoyannos BI, Wornoayporn V, Hendrichs MA. Odour-mediated
527 foraging by yellowjacket wasps (Hymenoptera: Vespidae): predation on leks of
528 pheromone-calling Mediterranean fruit fly males (Diptera: Tephritidae). *Oecol.*
529 1994; 99(1-2): 88-94.
- 530 51. Jaldo HE, Gramajo MC, Willink E. Mass rearing of *Anastrepha fraterculus*
531 (Diptera: Tephritidae): a preliminary strategy. *The Fla. Entomol.* 2001; 84(4): 716-718.
- 532 52. Salles LAB. Bioecologia e Controle das Moscas das Frutas Sul-Americanas.
533 Embrapa Clima Temperado-Livros técnicos (INFOTECA-E). Pelotas, Brazil. 1995
- 534 53. Jaldo HE, Willink E, Liedo P. Demographic analysis of mass-reared *Anastrepha*
535 *fraterculus* (Diptera: Tephritidae) in Tucumán, Argentina. *Revista Industrial y Agrícola*
536 *de Tucumán.* 2007; 84: 15-20.
- 537 54. Petit-Marty N, Vera MT, Calcagno G, Cladera JL, Segura DF, Allinghi A, et al.
538 Sexual behavior and mating compatibility among four populations of *Anastrepha*
539 *fraterculus* (Diptera: Tephritidae) from Argentina. *Ann Entomol Soc Am.* 2004; 97(6),
540 1320-1327.
- 541 55. Segura DF, Utgés ME, Liendo MC, Rodríguez MF, DevescoviF, Vera MT, et al.
542 Methoprene treatment reduces the pre-copulatory period in *Anastrepha fraterculus*
543 (Diptera: Tephritidae) sterile males. *J Appl Entomol.* 2013; 137: 19-29.
- 544 56. Juárez ML, Pimper LE, Bachmann GE, Conte CA, Ruiz MJ, Goane L, et al. Gut
545 bacterial diversity and physiological traits of *Anastrepha fraterculus* Brazilian-1
546 morphotype males are affected by antibiotic treatment. *BMC Microbiol*, in press.
- 547 57. Vera MT, Cáceres C, Wornoayporn V, Islam A, Robinson AS, De la Vega MH, et
548 al. Mating incompatibility among populations of the South American fruit fly
549 *Anastrepha fraterculus* (Diptera: Tephritidae). *Ann Entomol Soc Am.* 2006; 99(2): 387-
550 397.

- 551 58. Clayton DA. Socially facilitated behavior. *Q Rev Biol.* 1978; 53(4): 373-392.
- 552 59. Prokopy RJ, Duan JJ. Socially facilitated egg-laying behavior in Mediterranean
553 fruit flies. *Behav Ecol Sociobiol.* 1998; 42: 117-122.
- 554 60. Rull J, Prokopy RJ, Vargas RI. Effects of conspecific presence on arrival and use
555 of host in *Ceratitidis capitata* flies. *J Insect Behav.* 2003; 16: 329-346.
- 556 61. Oviedo A, Nestel D, Papadoupulos NT, Ruiz MJ, Prieto SC, Willink E, et al.
557 Management of protein intake in the fruit fly *Anastrepha fraterculus*. *J Insect Physiol.*
558 2011; 57(12): 1622-1630.
- 559 62. Teal PEA, Gomez-Simuta Y, Proveaux AT. Mating experience and juvenile
560 hormone enhance sexual signaling and mating in male Caribbean fruit flies. *Proc Natl*
561 *Acad Sci.* 2000; 97(7): 3708-3712.
- 562 63. StatSoft, Inc. STATISTICA (data analysis software system), version 7. 2004.
563 www.statsoft.com.
- 564 64. GraphPad Software, Inc. 1992-2012. Prism 6 for Windows, Version 6.1.
- 565 65. Shelly TE. Effects of raspberry ketone on the mating success of male melon flies
566 (Diptera: Tephritidae). *Proc Hawaii Entomol Soc.* 2000a; 34: 143-147.
- 567 66. Gillott C. Male accessory gland secretions: modulators of female reproductive
568 physiology and behavior. *Annu Rev Entomol.* 2003; 48(1): 163-184.
- 569 67. Avila FW, Sirot LK, LaFlamme BA., Rubinstein CD, Wolfner MF. Insect seminal
570 fluid proteins: identification and function. *Annu Rev Entomol.* 2011; 56: 21-40.
- 571 68. Abraham S, Cladera J, Goane L, Vera MT. Factors affecting *Anastrepha fraterculus*
572 female receptivity modulation by accessory gland products. *J Insect Physiol.*
573 2012; 58(1): 1-6.

- 574 69. Pszczolkowski MA, Tucker A, Srinivasan A, Ramaswamy SB. On the functional
575 significance of juvenile hormone in the accessory sex glands of male *Heliothis*
576 *virescens*. *J Insect Physiol.* 2006; 52(8): 786-794.
- 577 70. Clifton ME, Correa S, Rivera-Perez C, Nouzova M, Noriega FG. Male *Aedes*
578 *aegypti* mosquitoes use JH III transferred during copulation to influence previtellogenic
579 ovary physiology and affect the reproductive output of female mosquitoes. *J Insect*
580 *Physiol.* 2014; 64: 40-47.
- 581 71. Segura DF, Cáceres C, Vera MT, Wornoayporn V, Islam A, Teal PEA, et al.
582 Enhancing mating performance after juvenile hormone treatment in *Anastrepha*
583 *fraterculus*: a differential response in males and females acts as a physiological sexing
584 system. *Entomol Exp Appl.* 2009; 131(1): 75-84.
- 585 72. Liendo MC, Devescovi F, Bachmann GE, Utgés ME, Abraham S, Vera MT, et al.
586 Precocious sexual signalling and mating in *Anastrepha fraterculus* (Diptera:
587 Tephritidae) sterile males achieved through juvenile hormone treatment and protein
588 supplements. *Bull Entomol Res.* 2013; 103(1): 1-13.
- 589 73. Abraham S, Goane L, Rull J, Cladera J, Willink E, Vera MT. Multiple mating in
590 *Anastrepha fraterculus* females and its relationship with fecundity and fertility. *Entomol*
591 *Exp Appl.* 2011b; 141(1): 15-24.
- 592
- 593

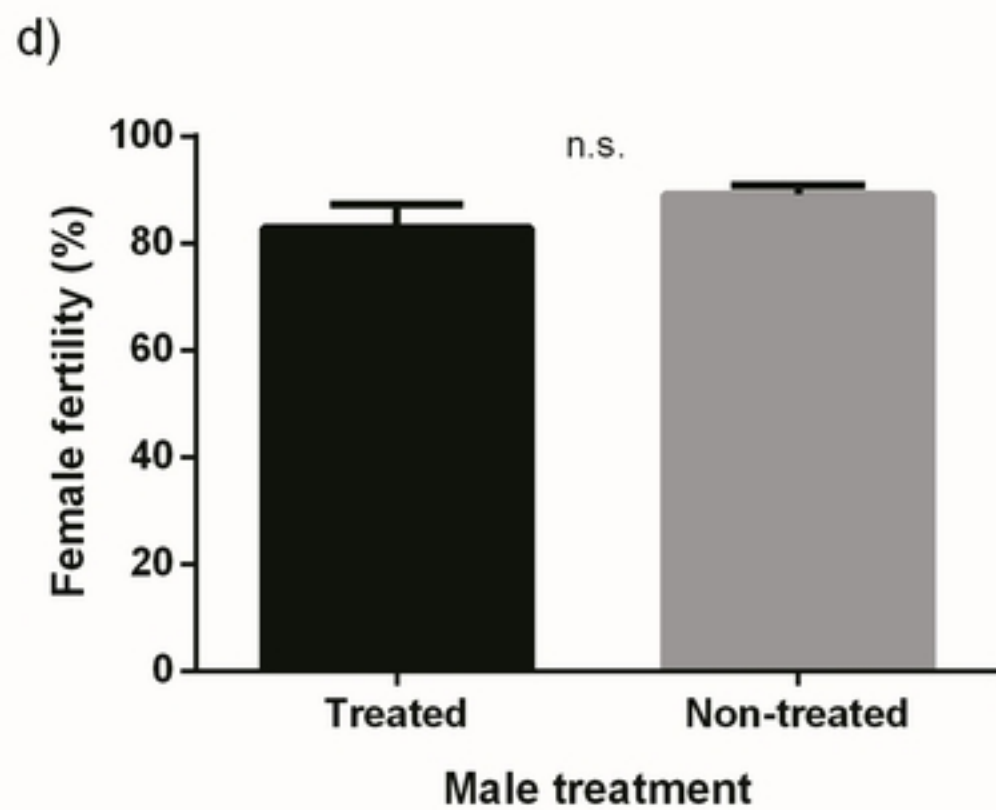
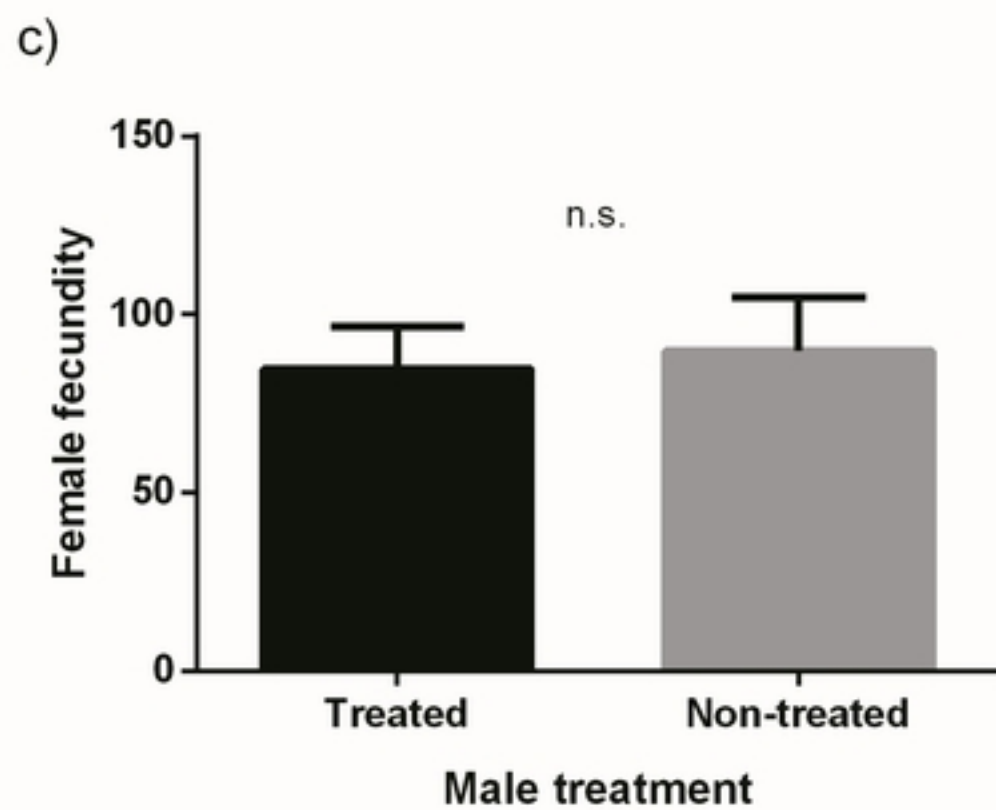
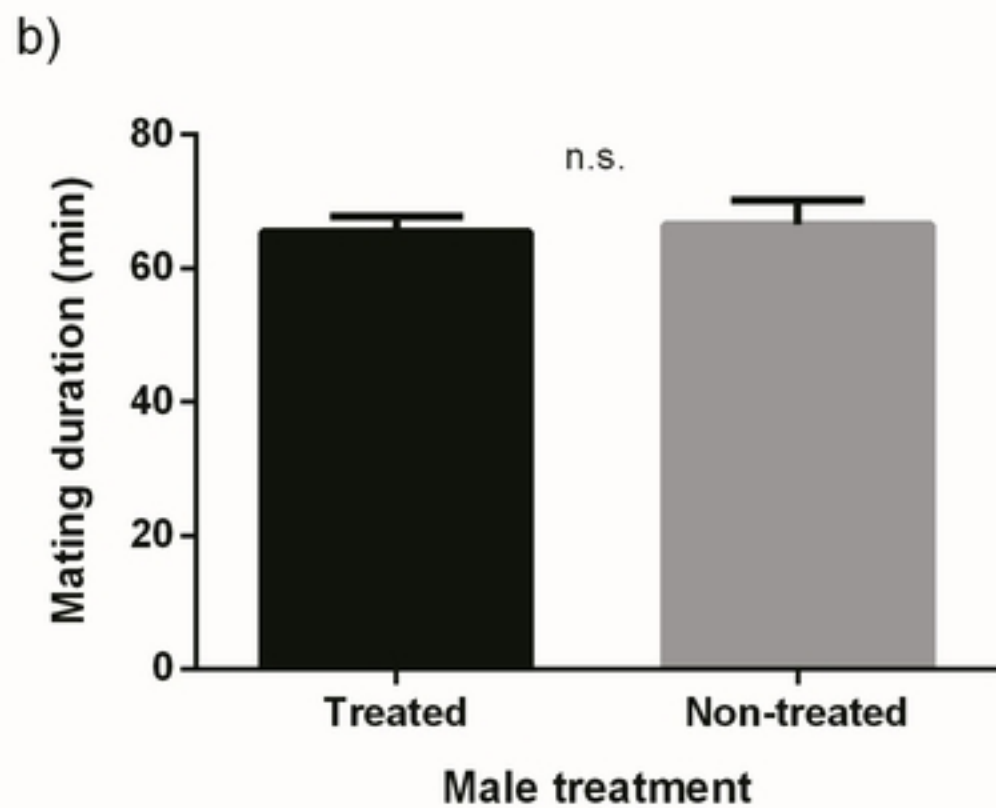
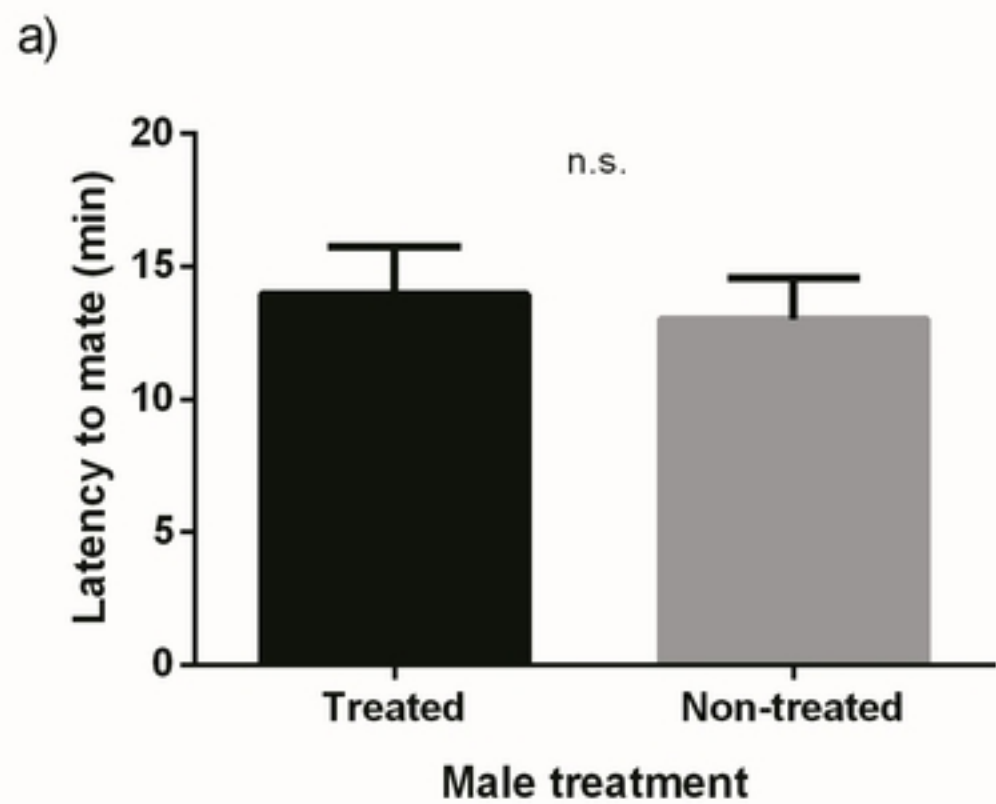


Figure 5

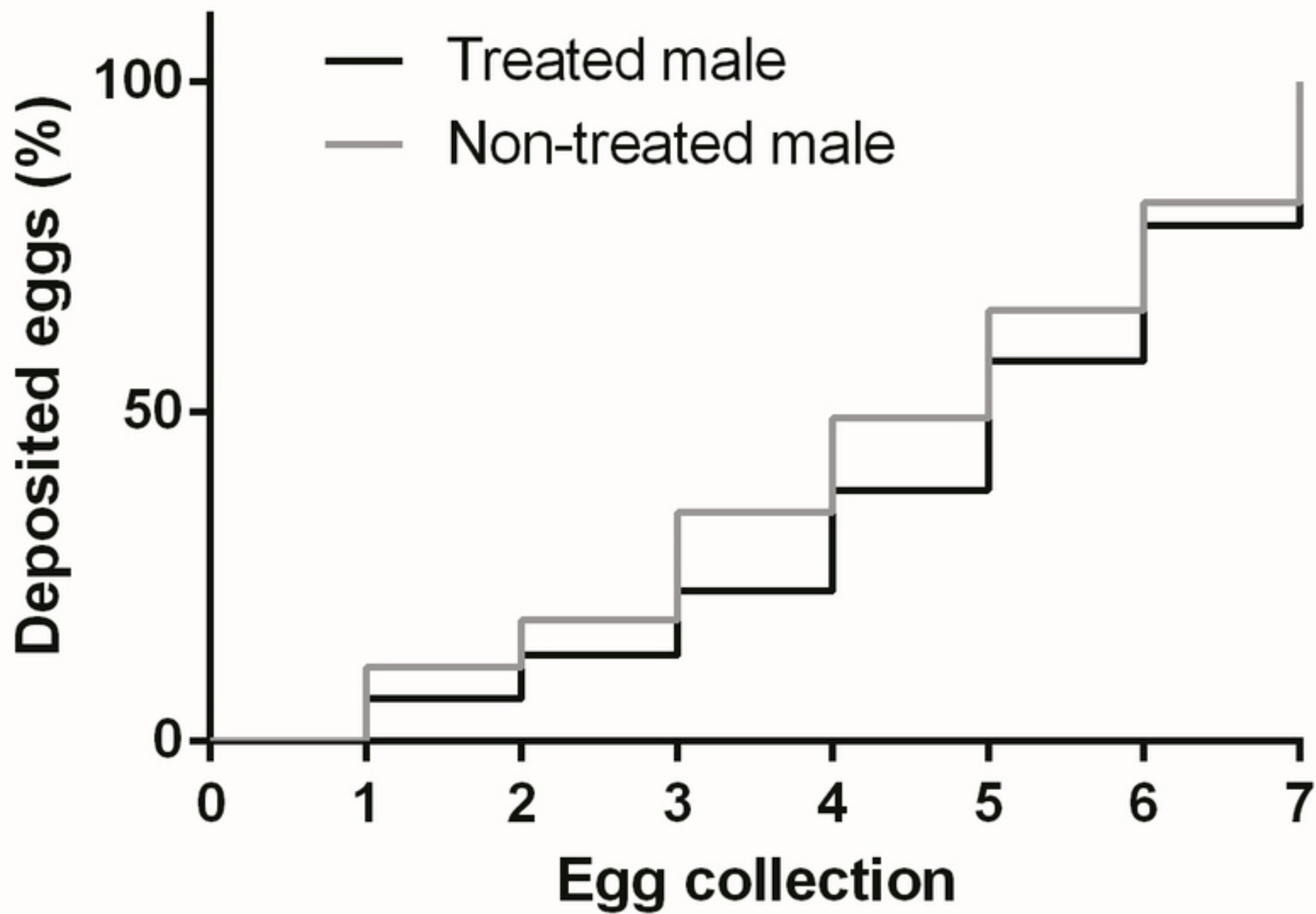


Figure 6

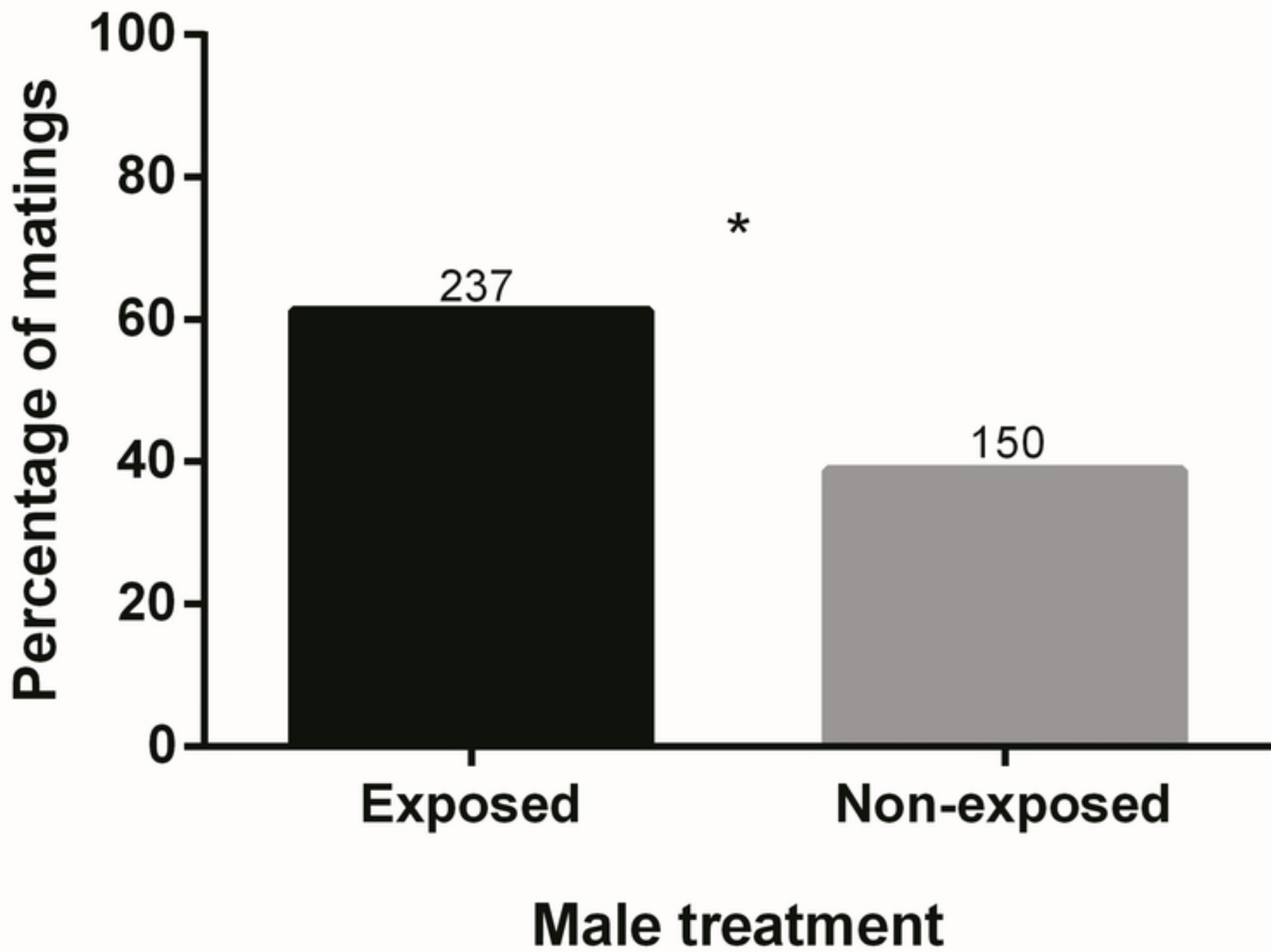


Figure 1

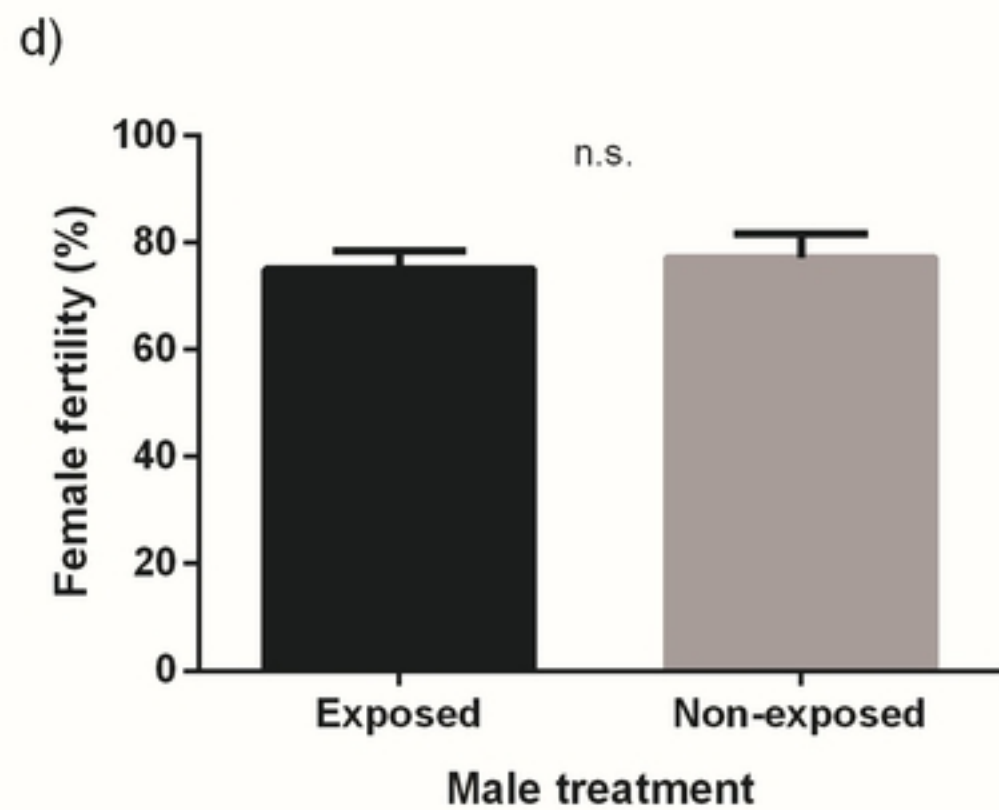
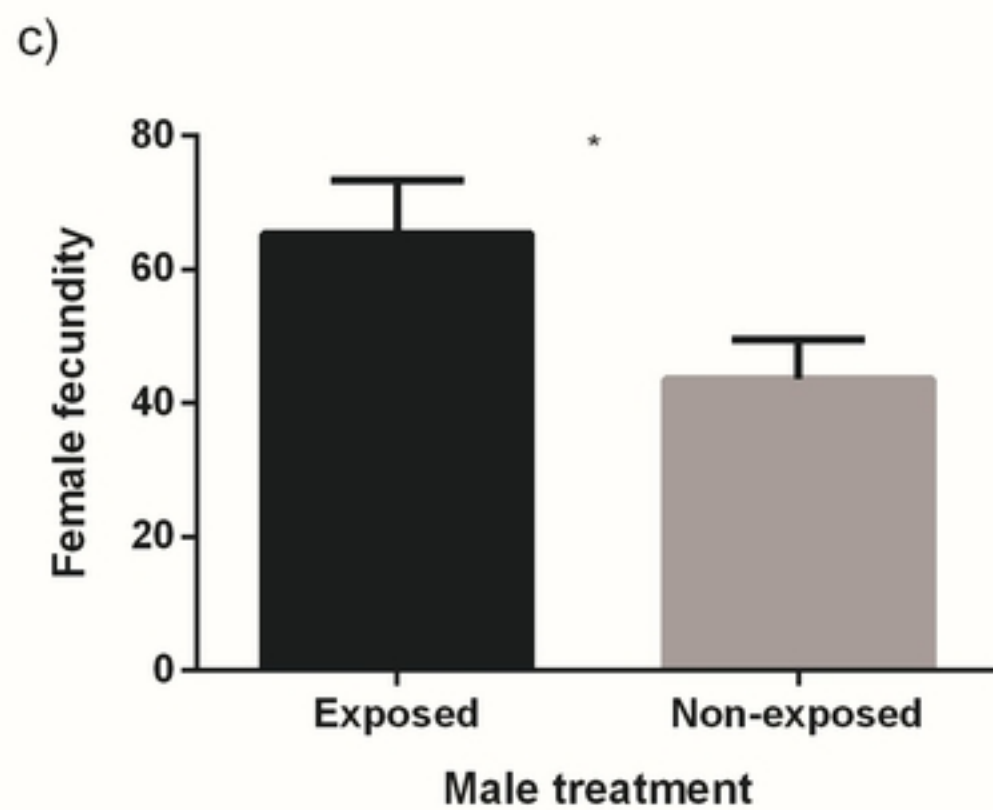
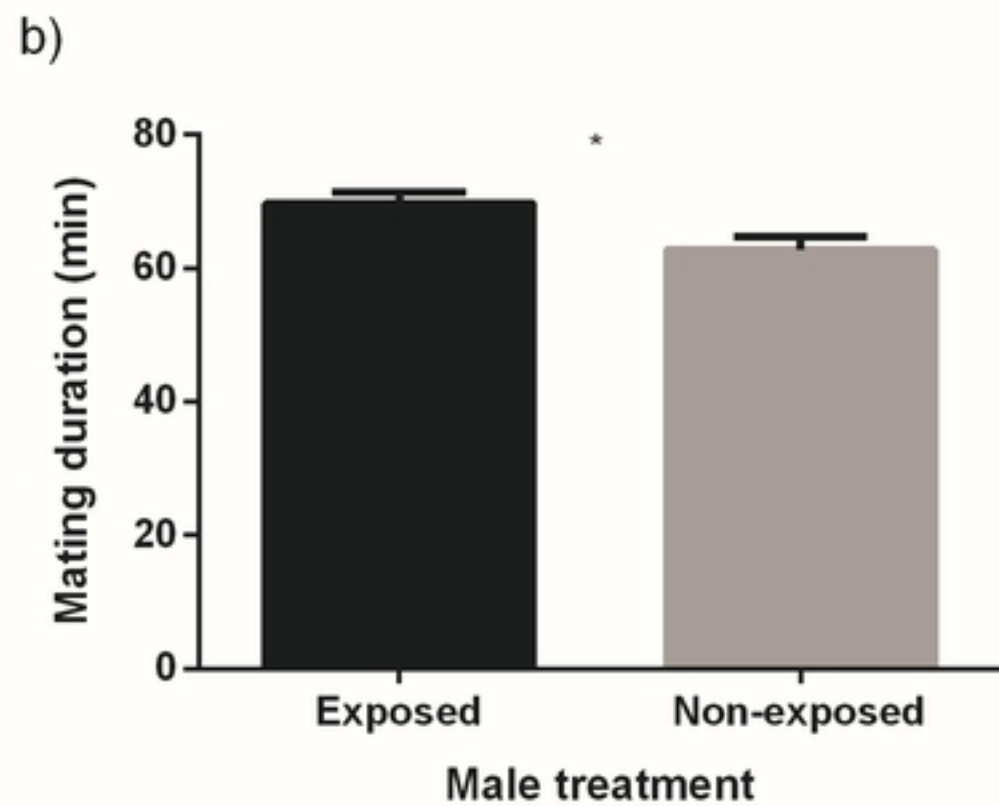
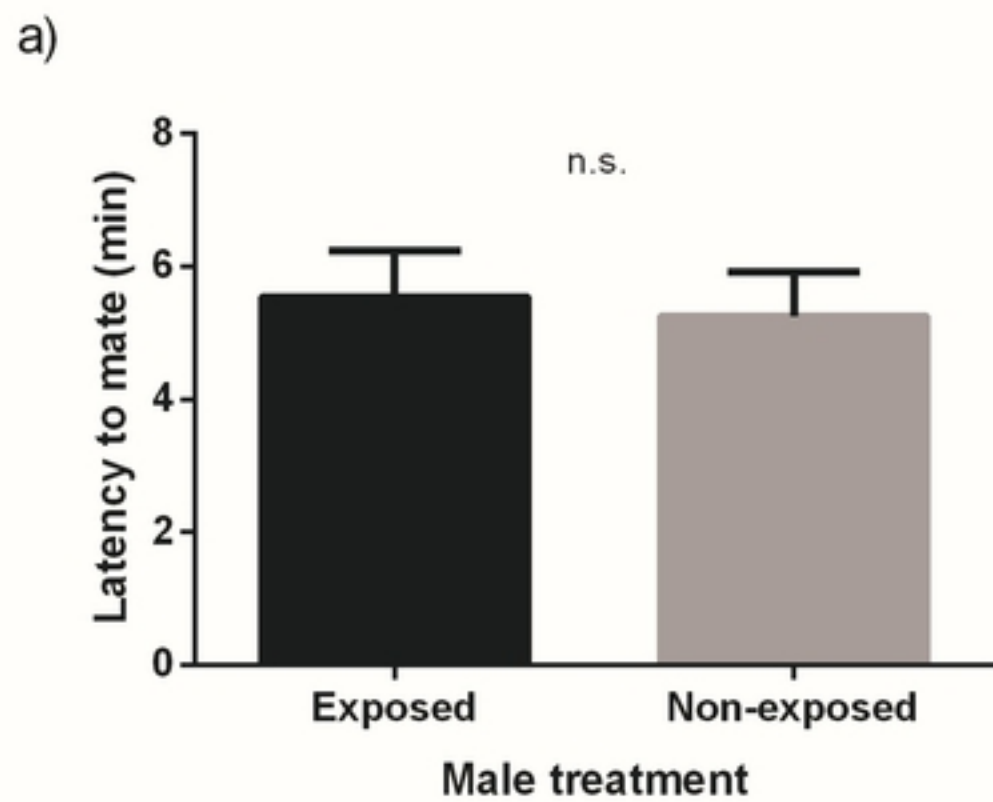


Figure 2

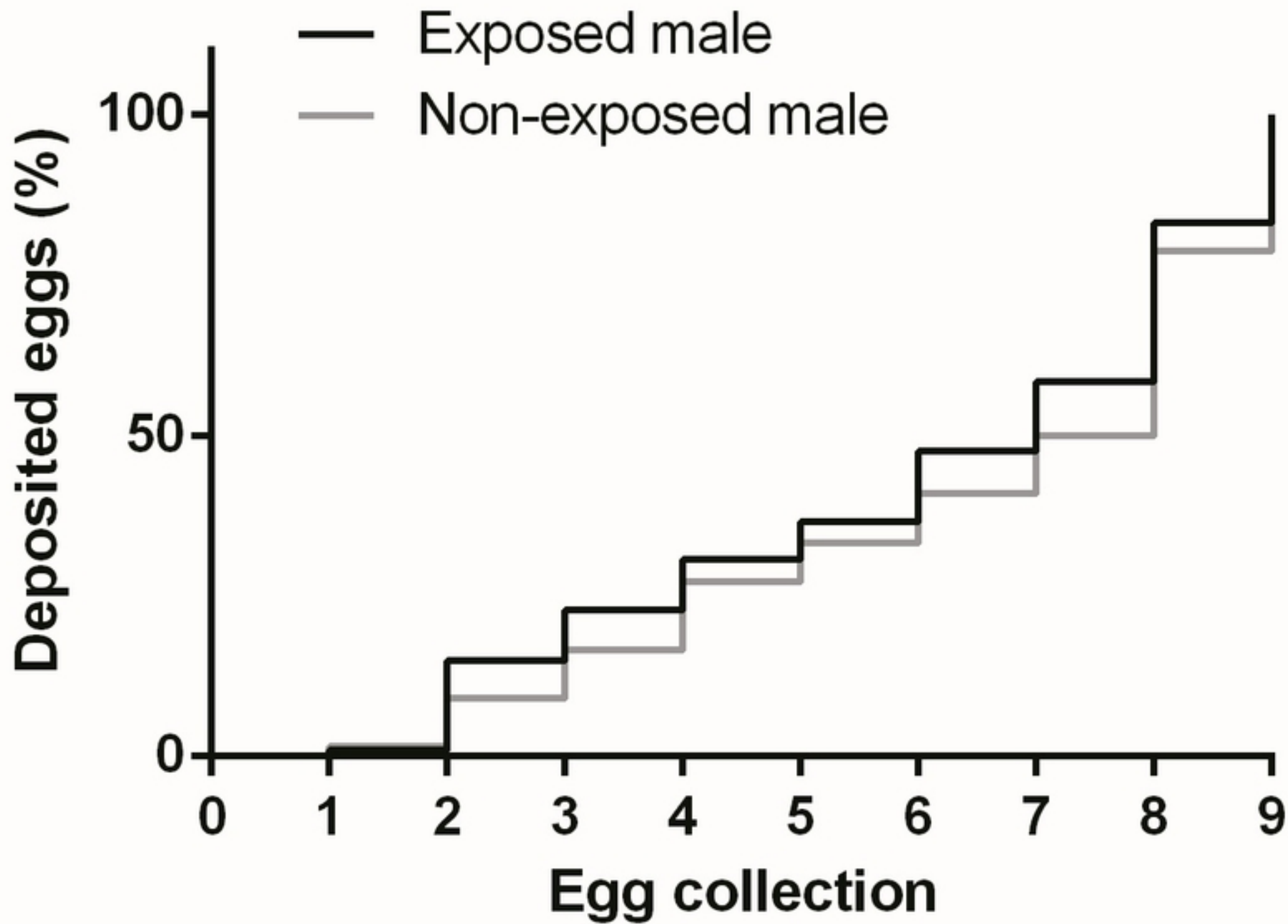


Figure 3

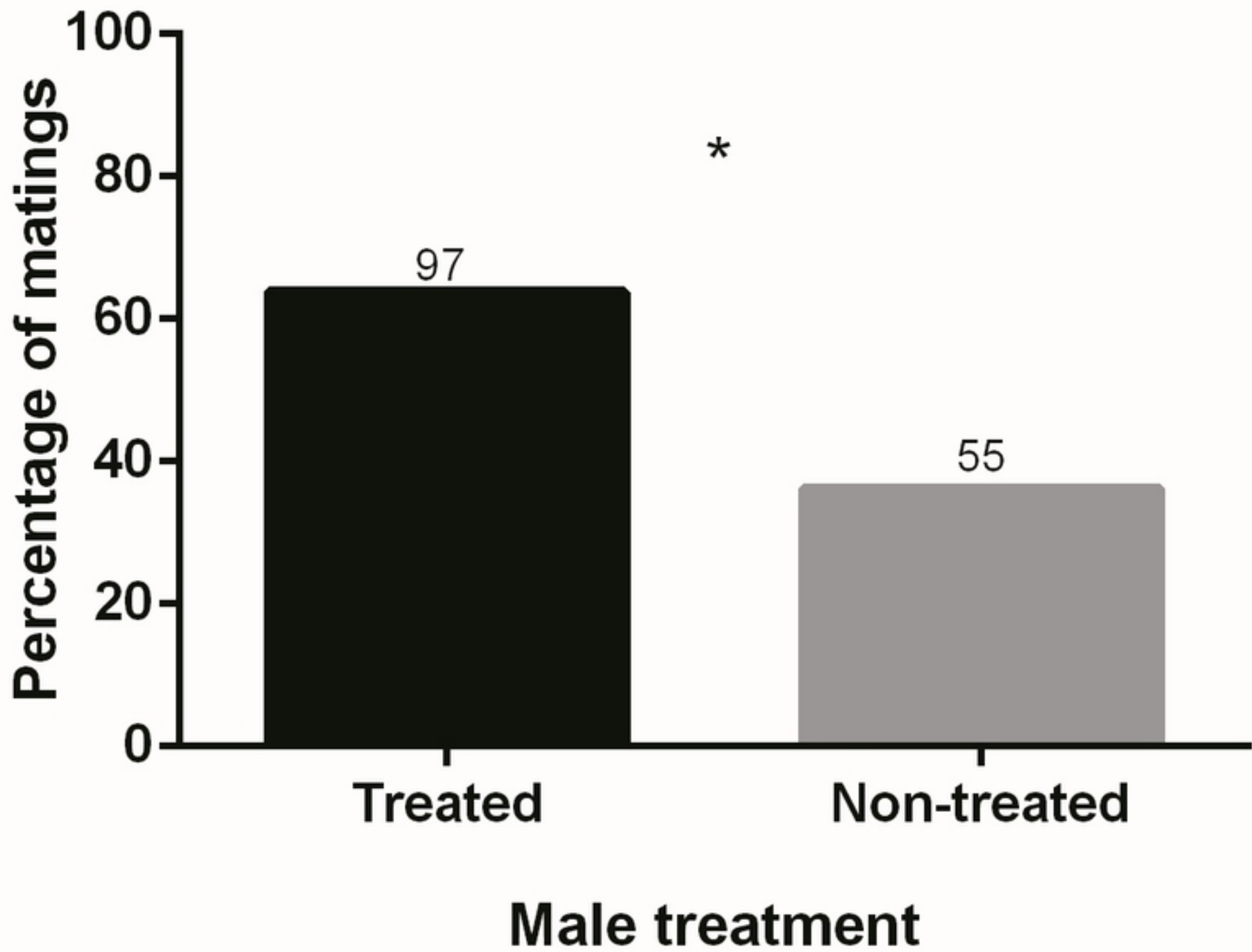


Figure 4