

1 **Epidemiology of *Staphylococcus aureus* in Neonates on Admission to a**
2 **Chinese Neonatal Intensive Care Unit**

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24 ABSTRACT

25 **Purpose:** Little is known about the molecular epidemiology of *Staphylococcus aureus* in
26 Chinese neonatal intensive care units (NICUs). We describe the molecular epidemiology of *S.*
27 *aureus* isolated from neonates on admission to Beijing Children's Hospital.

28
29 **Methods:** From May 2015-March 2016, nasal swabs were obtained on admission from 536
30 neonates. Cultures were also obtained from body sites with suspected infections. *S. aureus*
31 isolates were characterized by staphylococcal chromosomal cassette (SCCmec) type,
32 staphylococcal protein A (*spa*) type, multilocus sequence type (MLST), *sasX* gene, antimicrobial
33 susceptibility and cytotoxicity. Logistic regression assessed risk factors for colonization.

34
35 **Results:** Overall, 92 (18%) infants were colonized with *S. aureus* and 23 (4%) were diagnosed
36 with culture-positive *S. aureus* infection. Of the colonized infants, 72% harbored MSSA, while
37 74% of infected infants were culture-positive for MRSA. Risk factors for colonization included
38 female sex, age 7-28 days, birthweight and vaginal delivery. The most common MRSA and
39 MSSA clones were community-associated ST59-SCCmecIVa-t437 (60%) and ST188-t189
40 (15%), respectively. The *sasX* gene was not detected. Some MSSA isolates (16%) were
41 penicillin-susceptible and some MRSA isolates (18%) were oxacillin-susceptible. MRSA and
42 MSSA had similar cytotoxicity, but colonizing strains were less cytotoxic than strains associated
43 with infections.

44
45 **Conclusions:** *S. aureus* colonization was common in infants admitted to our NICU and two
46 community-associated clones predominated. Several non-modifiable risk factors for *S. aureus*

47 colonization were identified. These results suggest that screening infants for *S. aureus* upon
48 admission and targeting decolonization of high-risk infants and/or those colonized with high-risk
49 clones could be useful to prevent transmission.

50

51 **Key words:** *Staphylococcus aureus*, neonatal intensive care unit, colonization, MRSA, MSSA

52

53 INTRODUCTION

54 *Staphylococcus aureus* infections represent a significant clinical burden for infants worldwide
55 and were recently found to be the second most common cause of late-onset sepsis in very-low
56 birth weight (VLBW) infants admitted to neonatal intensive care units (NICU) in the United
57 States and United Kingdom.[1, 2] Preterm infants are also at high risk for *S. aureus*
58 colonization[3], a potential risk factor for subsequent infection. In a recent meta-analysis
59 involving patients admitted to NICUs and ICUs, methicillin-resistant *S. aureus* (MRSA)
60 colonization was associated with a 24.2 times increased MRSA infection risk.[4] Endemic
61 transmission and outbreaks due to MRSA and methicillin-susceptible *S. aureus* (MSSA) occur
62 frequently in NICUs.[5] Studying the molecular epidemiology and virulence factors of *S. aureus*
63 in the NICU population can promote an increased understanding of pathogenesis and ultimately
64 guide preventive strategies.

65
66 While the molecular characteristics of and risk factors for *S. aureus* colonization and infection
67 have been described for NICU populations across the globe and have increased our knowledge of
68 the global burden,[3, 6] no previous reports have described the molecular characteristics of *S.*
69 *aureus* strains isolated from neonates in NICUs in Mainland China. Many tertiary NICUs in
70 China are part of dedicated hospitals for children; neonates (less than 28 days old) served by
71 these units are mostly admitted from home after presenting as outpatients. Moreover, the
72 structural layout, patient population, and visiting policies for parents/guardians may differ
73 substantially from NICUs in other countries. For example, in the NICU of Beijing Children's
74 Hospital (BCH), parents are not allowed to visit and rooms contain four to eight neonates. In this
75 study, we aimed to determine the proportion of neonates colonized and/or infected with MSSA

76 and MRSA on admission to the NICU of BCH, as well as assess risk factors for *S. aureus*
77 colonization. We further aimed to describe the molecular epidemiology of both MSSA and
78 MRSA, including the most dominant clones, and their *in vitro* cytotoxicity. Ultimately, we will
79 use these data to inform future surveillance and *S. aureus* prevention efforts.

80

81 **METHODS**

82 **Study design, study population, and site**

83 From May 2015 to March 2016, we performed a prospective surveillance study on admission for
84 MSSA and MRSA among neonates ≤ 28 days of age hospitalized in the level 3, 50-bed NICU of
85 BCH. This hospital does not have an obstetrics unit; therefore, neonates (~700-750 annually) are
86 admitted to the NICU from home or other obstetric units. The most common admitting diagnoses
87 are infectious diseases (~60%), prematurity (~20%), and various congenital comorbidities
88 including cardiac, gastrointestinal, and neurologic disorders (~15%). The Ethics Committee of
89 BCH, affiliated with Capital Medical University, approved this study; parents and/or legal
90 guardians of infants provided written informed consent.

91

92 **Demographic and clinical data collection**

93 Selected demographic (e.g., sex, age, delivery type, birthweight) and clinical characteristics (e.g.,
94 congenital disease, respiratory support and previous antibiotic use) were abstracted from the
95 electronic medical records of enrolled neonates. Age was dichotomized as < 7 days and 7-28 days
96 of life for analysis, which was consistent with the way neonates have been previously defined[7]
97 [8]. Antibiotic exposure was defined as use of intravenous or oral antibiotics within the seven
98 days prior to admission. Respiratory support was defined as the use of nasal continuous positive
99 airway pressure (NCPAP) or mechanical ventilation within 24 hours of admission. Additionally,

100 diagnoses of suspected infections as described in the NICU admission notes, were also
101 abstracted.

102

103 **Surveillance and clinical specimen collection**

104 To detect *S. aureus* colonization, both anterior nares were swabbed within 24 hours of
105 admission, following a standard operational procedure. BBL™ Culture Swab™ Collection and
106 Transport System (Made by Copan for Becton, Dickinson, and Company, Sparks, USA) was
107 used. Specifically, only one swab was used for both nares. The swab should be inserted in the
108 nasal vestibule, introducing only the cotton part of the swab. The operator should rotate the swab
109 while circulating in the nasal vestibule for approximately 5 seconds. This procedure had to be
110 repeated in both nares.

111

112 Infants showing suggestive clinical symptoms were considered infected if *S. aureus* was isolated
113 from either a normally sterile site (eg, blood) or cultures obtained for clinical purposes (eg, skin
114 or eyes). The Clinical Microbiology Laboratory at BCH processed both surveillance and clinical
115 specimens. *S. aureus* was identified based on colony morphology and the coagulase test
116 (Saibaisheng, Beijing, China). PCR was used to detect the *mecA* gene.[9] Isolates with zone
117 sizes less than 21 mm for cefoxitin discs (Sigma, USA), according to the criteria of Clinical and
118 Laboratory Standards Institute (CLSI),[10] and which were also *mecA* gene positive were
119 considered MRSA. All *S. aureus* isolates were stored at -20° C.

120

121 **Molecular-typing and *sasX* detection**

122 To perform molecular studies, isolates were cultured by the research team on blood agar and
123 incubated overnight at 37°C. DNA was extracted and used to perform staphylococcal cassette

124 chromosome *mec* (*SCCmec*) typing,[11] multi-locus sequence typing (MLST)[12], and
125 staphylococcal protein A (*spa*) typing.[13] The *spa* types were assigned using the Ridom Staph
126 Database (Ridom, Germany).[13] Sequence types (STs) were assigned using the MLST database
127 (<http://saureus.mlst.net>).[14] Additionally, PCR was used to detect the presence of *sasX*, which
128 encodes for the cell wall-anchored protein-encoding gene.[15]

129

130 **Cytotoxicity assays**

131 Bacteriologically sterile culture filtrate preparations, obtained from early logarithmic-phase
132 growth (6 hours of incubation), were used for differentiating cytotoxic activity, as described
133 previously.[16] In brief, isolates were grown in 96-well, round bottomed plates in tryptic soy
134 broth for 16-18 hours with shaking at 37 °C. Cultures were diluted 1:75 with fresh Roswell Park
135 Memorial Institute plus casamino acids, and 150 µL of the diluted culture was regrown in 96-
136 well, round-bottomed plates for 6 hours at 37°C. Plates were centrifuged to pellet the bacteria,
137 and culture supernatants were collected and stored at –80°C until used. We then assayed the
138 cytotoxic activity of the culture supernatants using the human myeloid cell line HL-60,
139 differentiated into neutrophil-like cells (PMN-HL60), which have been shown to mimic the
140 sensitivity of human neutrophils to *S. aureus*. Twenty microliters of the *S. aureus* supernatant
141 were incubated with approximately 1.0×10^5 PMN-HL60 cells in a final volume of 100 µL (20%
142 v/v supernatants) for 2 hours at 37°C, followed by 2-hour incubation with the CellTiter reagent,
143 monitoring metabolic activity. Each sample was assayed in triplicate and independently repeated
144 at least twice. If >10% variation was observed between triplicate samples, the assay was
145 repeated.

146

147 **Antimicrobial susceptibility testing**

148 Antimicrobial susceptibility was determined by the agar dilution method, in accordance with the
149 CLSI.[10] MRSA and MSSA isolates were tested for susceptibility to penicillin, oxacillin,
150 gentamicin, ceftriaxone, rifampin, sulfamethoxazole-trimethoprim (TMP-SMX), erythromycin,
151 mupirocin, and levofloxacin. MRSA was also tested for susceptibility to vancomycin, linezolid,
152 fusidic acid, and tigecycline. *S. aureus* ATCC 29213 was used as the quality control.

153

154 **Statistical analysis**

155 The characteristics of colonized and uncolonized neonates were compared using Chi-squared and
156 Fisher's exact tests, as appropriate. Continuous variables, such as birthweight, were assessed
157 using Mann Whitney U test.

158

159 Categorical risk factors for *S. aureus* colonization were assessed by comparing the characteristics
160 of colonized vs. uncolonized neonates. For this analysis, birthweight, in grams, was categorized
161 into quartiles (e.g., 650-2500, 2501-3199, 3200-3500, 3501-5000 grams). Factors with $p < 0.10$ in
162 bivariate analysis were then assessed in a logistic regression model to determine risk factors for
163 overall *S. aureus* colonization. To determine independent risk factors for MRSA or MSSA
164 colonization, a multivariate multinomial logistic regression model was used. All statistical tests
165 were two-sided and performed in SAS 9.4 (Cary, NC); a p -value < 0.05 was considered
166 significant.

167

168 Cytotoxicity was expressed as the percentage of cells killed, and the median was compared
169 among MRSA versus MSSA isolates, isolates associated with infections versus colonization, and
170 among the most common STs. Cytotoxicity analyses were performed in GraphPad Prism 7.04

171 (GraphPad Software, La Jolla, CA) using the Mann Whitney U test.

172

173 **RESULTS**

174 **Demographic and clinical characteristics of study population**

175 From May 2015 to March 2016, 536 hospitalized neonates were admitted to the BCH NICU,
176 most of whom (520/536, 97%) were admitted from home after presenting as outpatients, another
177 16 neonates were transferred from other obstetric units within 24 hours of birth. All were
178 swabbed for *S. aureus* nasal colonization on admission. Neonates who were infected with *S.*
179 *aureus* on admission were excluded from analysis (n=23). Overall, 18% (n=92) of the 513
180 neonates had nasal colonization with *S. aureus*, 13% (n=66) were colonized with MSSA and
181 5.1% (n=26) were colonized with MRSA respectively. Colonized infants had a median
182 chronological age at admission of 14 (IQR [8-22.5]) days. Male and female infants had similar
183 ages at admission (mean 15 vs. 14 days, respectively, p=0.40). Colonized infants had
184 significantly higher birthweights (3270 IQR [2020-3655] grams) than uncolonized infants (3100
185 IQR [2500-3500] grams, p=0.001).

186

187 In this cohort, 255 infants were admitted with suspected infections; of these 23 (9.0%) were
188 culture-positive for *S. aureus*. Of the 23 *S. aureus* infections, 74% (n=17) were infected with
189 MRSA and 26% (n=6) were infected with MSSA. Conjunctivitis (n=8, 35%) and omphalitis
190 (n=8, 35%) were the most commonly diagnosed *S. aureus* infections, followed by pneumonia
191 (n=3, 13%), impetigo (n=2, 8.7%), cellulitis (n=1, 4.3%), and septicemia (n=1, 4.3%). Four
192 infected neonates were also colonized (2 with MSSA and 2 with MRSA).

193

194 **Risk factors for MSSA and MRSA colonization**

195 Risk factors for *S. aureus* colonization are shown in Table 1. In the multivariable adjusted
196 logistic regression model, female sex, age 7-28 days, birthweight and vaginal delivery were
197 associated with either *S. aureus* colonization, while antibiotic use in the week prior to admission
198 was protective. An additional adjusted logistic regression model assessing interaction between
199 birthweight quartiles and neonate age was assessed, but the interaction term was not significant
200 (p=0.09).

201
202 In multinomial logistic regression, female sex (p=0.02), vaginal delivery (p<0.0001), and age 7-
203 28 days (p<0.0001) remained significant risk factors for MSSA colonization, while female sex
204 (p=0.03) and age 7-28 days (p=0.001) remained significant risk factors for MRSA colonization.
205 Antibiotic use remained protective for both MSSA and MRSA, while birthweight was
206 unassociated with either MSSA or MRSA colonization.

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218 **Table 1 Risk Factors for *S. aureus* Colonization in Neonates Admitted to the NICU of Beijing**

219 **Children's Hospital**

Characteristics	Total ¹ (N=513) N (% of population or subpopulation)	Colonized ¹ (N=92)	Uncolonized (N=421)	Crude p-value	OR _{ADJ} (CI ₉₅) ²	Adj. p-value
Demographic						
Birthweight (g)³	3200 [2600-3500]	3270 [3030-3655]	3100 [2500-3500]	0.001	--	--
Sex				0.01	2.16 (1.29, 3.60)	0.0032
Female	198 (39%)	47 (24%)	151 (76%)			
Male	315 (61%)	45 (14%)	270 (86%)			
Age (days)				<0.0001	6.13 (3.34, 11.23)	<0.0001
<7	242 (53%)	19 (7.9%)	223 (92%)			
7-28	271 (47%)	73 (27%)	198 (73%)			
Delivery type				0.0005	2.36 (1.40, 3.98)	0.0012
Vaginal	248 (48%)	60 (24%)	188 (76%)			
Cesarean	265 (52%)	32 (12%)	233 (88%)			
Clinical						
Congenital Disease				0.36	--	--
Yes	86 (17%)	12 (14%)	74 (86%)			
No	427 (83%)	80 (19%)	347 (81%)			
Antibiotic Exposure, prior week				0.0058	0.25 (0.14, 0.45)	<0.0001
Yes	188 (37%)	22 (12%)	166 (88%)			
No	325 (63%)	70 (22%)	255 (78%)			
Respiratory Support				0.77	--	--
Yes	96 (19%)	16 (17%)	80 (83%)			
No	417 (81%)	76 (18%)	341 (82%)			

220

221 ¹ Excludes 4 neonates who were both colonized and infected on admission

222 ² Multivariable model adjusted for sex, age, delivery, birthweight (quartiles), and antibiotic use

223 ³ Median [IQR]

224 Abbreviations used: OR_{ADJ}=Adjusted Odds Ratio, CI₉₅=95% Confidence Interval, [IQR]=

225 Interquartile Range

226

227 **Molecular characteristics of *S. aureus* isolates**

228 MLST revealed 16 different sequence types (STs) among the 74 MSSA isolates (Figure 1A), the
229 most common of which were ST188 (n=12, 16%) and ST5 (n=12, 16%). Twenty-eight MSSA
230 *spa* types were identified, ST188-t189 (n=11, 15%) was the most common MSSA clone.

231
232 MLST revealed six different STs among the MRSA isolates (Figure 1B); ST59 was the most
233 common (n=35, 78%). Nine MRSA *spa* types were identified, SCCmecIVa was the most
234 common SCCmec type detected (n=32, 71%), followed by SCCmecV (n=6, 13%), SCCmecIVg
235 (n=5, 11%), and SCCmecIII (n=1, 2.2%); one isolate could not be typed. ST59-SCCmecIVa-
236 t437 (n=27, 60%) was the most common MRSA clone (Figure 2B).

237
238 Of 19 *S. aureus* isolates associated with skin and soft tissue infections (SSTIs), 3 (17%)
239 belonged to ST398. Three of the four neonates who were both colonized and diagnosed with
240 infection had the same *spa* type, ST, and SCCmec type identified in their isolate pairs; another
241 one infant had a point mutation in the *tpi* gene representing a different ST. No MSSA or MRSA
242 isolate harbored *sasX*.

243 244 **Antimicrobial susceptibilities**

245 All 74 MSSA isolates were susceptible to oxacillin, rifampin, TMP-SMX, and mupirocin; 99%
246 (73/74) were also susceptible to levofloxacin, 26% (19/74) to erythromycin and 16% (12/74) to
247 penicillin. All 45 MRSA isolates were susceptible to rifampin, TMP-SMX, mupirocin,
248 levofloxacin, vancomycin, linezolid and tigecycline; 98% (44/45) were also susceptible to
249 gentamicin and fusidic acid (44/45).

250 Eight MRSA isolates (18%) were oxacillin-susceptible (OS-MRSA), five (63%) of these
251 belonged to ST59-SCCmecIVa-t437.

252

253 **Cytotoxicity of MSSA and MRSA and of colonizing and infectious isolates**

254 The median cytotoxicity of the 119 isolates was 85% (IQR [76-88%]). The cytotoxicity of
255 MRSA (median: 84%, IQR [79-87%]) and of MSSA (median: 86%, IQR [75-88%]) were similar
256 ($p=0.85$) as shown in Figure 2A. The cytotoxicity of the 96 colonizing *S. aureus* isolates
257 (median: 85%, IQR [73-87%]) was less than that of the 23 infectious isolates (median: 88%, IQR
258 [82-88%], $p=0.0008$) (Figure 2B). Of the 3 main *S. aureus* clones, ST398-t571 had significantly
259 lower cytotoxicity (median: 72%, IQR [64-79%], $p=0.002$) compared to ST188-t159 (median:
260 85%, IQR [84-88%]) and ST59-t437 (median: 83%, IQR [74-87%]). Additionally, ST398-t571
261 had significantly lower cytotoxicity when compared to the cytotoxicity of all other clones
262 (median all other clones: 86% IQR [80-88%], $p=0.003$).

263

264 **DISCUSSION**

265 To our knowledge, this is the first study to assess the burden and molecular epidemiology of *S.*
266 *aureus* in a Chinese NICU. On admission, 18% of neonates ≤ 28 days of age had nasal
267 colonization with MSSA or MRSA. This rate was higher than previously reported for neonates
268 within 6 days of birth in Japan (10%)[17], neonates within 1 month of birth in the United States
269 (3.8%),[18] and neonates in a Taiwanese NICU (13%).[19] In the current study, 5.1% of
270 neonates were colonized with MRSA, which was also a higher rate than previously reported in
271 other NICUs in which rates ranged from 0.3-4.4%. [17]; [18]; [19] Our higher colonization rate
272 may reflect the admission patterns into our NICU, as 97% of neonates were admitted from home

273 and others were transferred from other obstetric units within 24 hours of birth. An *S. aureus* case
274 was considered community acquired if it was isolated from an outpatient or an inpatient within
275 48 h of hospitalization¹⁴, so all the *S. aureus* were more likely to have been transferred from the
276 community. Furthermore, 9% of neonates admitted with suspected infections were culture-
277 positive for *S. aureus*, 74% (17/23 infections) of which were due to MRSA. This contrasts with
278 previous reports in which MSSA represented a greater proportion of *S. aureus* infections than
279 MRSA.[20] Our findings demonstrated that admitted neonates continually imported *S. aureus*
280 into the NICU, and thus served as potential reservoirs of pathogens for other infants. This
281 suggests that routine surveillance followed by targeted intervention strategies could be useful in
282 reducing *S. aureus* infections. However, future studies should assess the effectiveness of
283 surveillance and decolonization in identifying high-risk infants and/ or targeting highly-cytotoxic
284 *S. aureus* clones in our NICU.

285
286 While others have assessed factors associated with *S. aureus* colonization in the NICU
287 population, including prematurity and intubation, few have examined the effect of age on
288 colonization risk.[4, 6] We found that neonates aged 7-28 days were at significantly increased
289 risk for both MRSA and MSSA colonization compared with younger neonates. Similarly,
290 Macnow et al found that infants transferred to the NICU at 7 days of age or older had
291 significantly increased odds of colonization with MRSA compared to younger infants,
292 presumably due to more MRSA exposure through interactions with staff, family members, and
293 the healthcare environment.[21] We also found that higher birthweight was associated with an
294 increased risk of *S. aureus* colonization, which contrasts with previous literature[3], we believe
295 this finding may also be related to the admission patterns of our NICU. In addition, we found
296 that female sex was a risk factor for both MSSA and MRSA colonization, which differed from

297 previous reports. A meta-analysis of risk factors for MRSA colonization in the NICU showed no
298 relationship between colonization and sex,[3] while another study reported male neonates were
299 at increased risk of both MRSA and MSSA colonization.[22] However, our admission patterns
300 did not elucidate an explanation for this finding nor were female neonates older than male
301 neonates on admission. We found vaginal delivery to be a risk factor for MSSA, but not for
302 MRSA colonization. In Shenzhen China, 7.3% vs. 1.7% of pre-partum women were colonized
303 with MSSA and MRSA, respectively, and vaginal delivery was associated with neonatal MSSA,
304 but not with MRSA colonization.[22] Similarly, Top et al reported MSSA and MRSA
305 anovaginal colonization rates of 11.8% and 0.6%, respectively, in pre-partum women, although
306 neonatal colonization was not assessed.[23] These findings suggest that vertical transmission of
307 *S. aureus* occurs, but is more relevant for MSSA than MRSA, presumably because fewer women
308 are colonized with MRSA.

309
310 In contrast, neonates who had received antibiotics within seven days of admission were at
311 decreased risk for both MSSA and MRSA colonization. Notably, 188 (37%) neonates had
312 received antibiotics within seven days of admission, as many infants had suspected infections
313 managed as outpatients. Oral antibiotics have not been shown to eradicate MRSA colonization in
314 hospitalized adults.[24] However, the relevance of these studies for neonates in whom the
315 organism burden may be lower or the duration of colonization is likely shorter is uncertain.
316 MSSA *spa* and STs were diverse, as noted in previous studies.[25] However, ST188 and ST5
317 were the most common MSSA clones, consistent with previous reports of community-associated
318 MSSA among adults and children in China.[26] ST188 virulence has been in part attributed to
319 epithelial cell adhesion and biofilm formation, properties which could facilitate nasal
320 colonization.[26] Additionally, an American study indicated that livestock-associated MRSA

321 ST5 isolates can adhere to human keratinocytes, which may facilitate colonization with this
322 strain.[27] Importation of common community-associated clones with enhanced adherence
323 properties could potentially facilitate MSSA transmission in the NICU.
324
325 ST59-SCC*mecIVa* was the most common MRSA clone, consistent with previous studies of
326 epidemic MRSA clones across Asia[14] and of community-associated MRSA identified in
327 Chinese children's hospitals.[28] In this study, several MSSA (n=10) and MRSA (n=2) isolates
328 belonged to ST398, which was originally reported to colonize livestock and their human
329 handlers,[29] but has recently been associated with colonization and infection in distinct human
330 populations in Europe, the Caribbean, and the northeastern United States.[30-32] In China,
331 ST398 is thought to account for as many as 20% of SSTIs caused by *S. aureus*.[29] Here, we
332 found that 16% (3/19) of SSTIs caused by *S. aureus* were ST398. As previous studies have
333 primarily focused on adult patients, the current study may help elucidate the role of ST398 in *S.*
334 *aureus* infections in the neonatal population.

335
336 We also explored the presence of the virulence gene *sasX*, which has been implicated in
337 epidemic spread of MRSA across China and has been demonstrated to play a key role in
338 colonization and pathogenesis, including promotion of immune evasion.[33] However, *sasX* was
339 not detected in the current study, potentially because no neonates harbored ST239, the dominant
340 global healthcare-associated MRSA clone which also predominantly carries *sasX*.[33]

341
342 Overall, 16% of MSSA strains were penicillin-susceptible. A recent study in Massachusetts
343 found a 3-fold (13% to 32%) increase in penicillin susceptibility among MSSA bloodstream
344 isolates over a decade, speculated to be the result of less selective pressure by β -lactam

345 agents.[34] Future studies should address this possibility in China. Additionally, 18% of MRSA
346 strains were oxacillin-susceptible (OS), which is the first time, to our knowledge, such isolates
347 have been reported in the neonatal population. OS-MRSA is increasingly associated with animal
348 and human infections worldwide;[35] 76% of MRSA isolates from bovine mastitis diagnosed in
349 four Chinese provinces were OS.[36] In our study, ST59-SCCmecIV-t437 was the most common
350 OS-MRSA clone (63%), which is consistent with a recent study from China which showed that
351 the most frequent OS-MRSA clones were ST338-t437-SCCmecV (32%) and ST59-t437-
352 SCCmecIV/V (21%).[37] Likewise, ST59-t437-SCCmecV_T was the most prevalent OS-MRSA
353 clone in a study from Taiwan.[38] However, this finding differs from other reports; ST88 and
354 ST8 were the most prevalent OS-MRSA clones in Africa[39] and OS-MRSA clones were highly
355 diverse in Brazil,[40] potentially due to geographic differences in genetic background. OS-
356 MRSA strains may be misidentified as MSSA by traditional susceptibility testing or as MRSA
357 by molecular detection of *mecA*, thereby complicating the diagnosis and appropriate treatment of
358 *S. aureus* infections. Surveillance for such emergent strains should thus be a public health
359 priority.

360
361 Furthermore we found that cytotoxicity of *S. aureus* was higher in infectious than in colonizing
362 isolates. This observation differs from a recent report by Maisem et al,[41] who found an
363 unexpected inverse correlation between *S. aureus* toxicity and disease severity when comparing
364 colonizing isolates and those isolated from SSTIs or bacteremia in adult patients. They suggested
365 that bacterial fitness in human serum could explain the unexpected association of low-toxicity
366 isolates with severe, invasive disease. Similarly, Rose et al[16] reported that low cytotoxic
367 activity and the CC8/239 clone, a weakly cytotoxic lineage, were independent predictors of

368 mortality in MRSA healthcare-associated pneumonia, suggesting that isolates with low
369 cytotoxicity may result in a depressed host response and ultimately worse patient outcomes.[16]
370 We also found that cytotoxicity was linked to the genetic background of *S. aureus*, as ST398-
371 t571 exhibited significantly lower cytotoxicity than the two most common clones, as well as all
372 other clones combined. The association between cytotoxicity, clinical presentations, and
373 outcomes should be studied to further elucidate *S. aureus* pathogenesis. Such studies could also
374 have important clinical implications and support targeted rather than universal decolonization of
375 neonates colonized with strains with specific molecular and virulence properties.

376
377 Several limitations to our study need to be considered. This was a single NICU cohort study with
378 unique admission patterns, which limits the generalizability of our findings. Since all *S. aureus*
379 isolates were collected from neonates upon admission, we could not explore hospital
380 transmission of MSSA or MRSA, nor ascertain whether colonized neonates subsequently
381 developed infections. Furthermore, given the study design, we could not assess the potential
382 relationship between cytotoxicity and subsequent infections or clinical outcomes. Because
383 surveillance cultures were only obtained from the nares of neonates and not from other body
384 sites, it is possible that we underestimated the true proportion of colonized neonates. We may
385 have also underestimated infections caused by *S. aureus*, as pre-admission antibiotics could have
386 resulted in negative clinical cultures.

387
388 In conclusion, the nasal colonization rate of *S. aureus* in neonates was high in the NICU of BCH.
389 Female sex, age 7-28 days, birthweight and vaginal delivery were risk factors for colonization.
390 While MSSA more frequently colonized neonates, MRSA more frequently infected neonates.
391 Most *S. aureus* strains were community-associated, reflective of NICU admission patterns.

392 Isolates associated with clinical infection exhibited higher cytotoxicity than colonizing isolates.
393 Our findings suggest that active surveillance of neonates for *S. aureus* should be considered as
394 part of strategies to detect importation and prevent transmission of both MRSA and MSSA
395 within the NICU.

396

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402

403 **Conflict of interest:**

404 The authors have no conflicts of interest to disclose.

405

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1 **Figure Legends**

2 **Figure 1A: Distribution of MSSA *spa* types among sequence types (ST) in neonates**
3 **admitted to the NICU of Beijing Children's Hospital, May 2015-March 2016.**

4 Overall, 16 STs and 29 *spa* types were identified in 74 MSSA isolates. The most common STs
5 were ST188 (n=12, 16%) and ST398 (n=10, 14%). Abbreviations used in figure: (MSSA,
6 methicillin-susceptible *Staphylococcus aureus*; MLST, multi-locus sequence typing; ST,
7 sequence type).

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9 **Figure 1B: Distribution of MRSA *spa* types among sequence types (ST) in neonates**
10 **admitted to the NICU of Beijing Children's Hospital, May 2015-March 2016.**

11 Overall, 6 STs and 9 *spa*-types were identified in 45 MRSA isolates. The most common ST was
12 ST59 (n=35, 78%). Abbreviations used in figure: (MSSA, methicillin-susceptible
13 *Staphylococcus aureus*; MLST, multi-locus sequence typing; ST, sequence type).

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1 **Figure Legends**

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3 **Figure 2A: Cytotoxicity of MSSA versus MRSA isolates obtained from neonates at Beijing**

4 **Children's Hospital.**

5 The cytotoxicity of MSSA isolates associated with colonization or infection (n=74) versus

6 MRSA isolates associated with colonization or infection (n=45) was similar (84 vs 86%,

7 p=0.85).

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9 **Figure 2B: Cytotoxicity of MSSA and MRSA isolates associated with colonization versus**

10 **infection obtained from neonates at Beijing Children's Hospital.**

11 The cytotoxicity of MRSA and MSSA isolates associated with colonization (n=96) was less

12 than the cytotoxicity of MSSA and MRSA isolates associated with infection (n=23) (p=0.008).

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