| 1 | Accounting for imperfect detection in species with sessile life cycle stages: a case study of |
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| 2 | bumble bee colonies |
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18 Abstract

| 19 | 1. | For bumble bees, colonies (not individual workers) are the functional unit of the population. |
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| 20 | | Estimates of colony density are thus critical for understanding population distribution and |
| 21 | | trends of this important pollinator group. Yet, surveys of bumble bee colonies and other taxa |
| 22 | | with sessile life cycle states rarely account for imperfect detection. |
| 23 | 2. | Here we demonstrate the use of mark-recapture methods to estimate the density of bumble |
| 24 | | bee colonies across the landscape using standardized survey protocols. |
| 25 | 3. | We found that the probability of detecting colonies in standardized surveys varied |
| 26 | | considerably across space, through time, and among colonies. |
| 27 | 4. | Using simulations, we also show that imperfect detection can obscure true variation in |
| 28 | | density among plots, or generate spurious variation in counts even when all plots have the |
| 29 | | same density. In both cases, we show that mark-recapture can be used to generate unbiased |
| 30 | | estimates of density, with relatively low search effort compared to conventional survey |
| 31 | | methods for bumble bee colonies. |
| 32 | 5. | Our study illustrates the advantages of mark-recapture for optimizing survey protocols for |
| 33 | | species with cryptic and sessile life cycle stages, which will be a valuable tool in ongoing |
| 34 | | studies of pollinator nesting ecology. |
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Key-words *Bombus*; closed population model; mark-recapture; monitoring; nesting habitat;
 occupancy

38 Introduction

| 39 | Measuring the size of natural populations is a primary goal of conservation monitoring |
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| 40 | and is often a prerequisite for management action or wildlife policy decisions (Williams et al. |
| 41 | 2002). Yet, estimating abundance and the factors that influence it can be challenging when |
| 42 | organisms are difficult to detect. Accordingly, study designs and analyses that account for |
| 43 | imperfect detection have a rich tradition in wildlife research (White and Burnham 1999; |
| 44 | Williams et al. 2002; Kéry and Schaub 2012). Failure to account for imperfect detection of |
| 45 | cryptic organisms can introduce both uncertainty and bias into population estimates, potentially |
| 46 | leading to erroneous conclusions about population status and habitat requirements (Gu and |
| 47 | Swihart 2004; Kéry and Schmidt 2008). |
| 48 | In contrast to studies of mobile vertebrates, imperfect detection is rarely accounted for in |
| 49 | population estimates for both invertebrates and taxa that are sessile or have a sessile life-cycle |
| 50 | stage, including plants and ground-nesting animals (Kellner and Swihart 2014; Berberich et al. |
| 51 | 2016). Kellner and Swihart (2014) found that only 9.0% of invertebrate and 1.4% of plant |
| 52 | population studies accounted for imperfect detection, compared to the mean of 23% across all |
| 53 | taxa. This discrepancy potentially stems from the incorrect assumption that detection probability |
| 54 | is inherently high in less mobile organisms. Yet, this assumption may be false if sessile |
| 55 | organisms are rare, inconspicuous, or logistically difficult to survey. For example, in a long-term |
| 56 | study of a 4.5 ha plot in Kansas, Slade et al. (2003) estimated that fewer than 4% of existing |
| 57 | Mead's milkweed plants were discovered in years without spring fires, and only 18% were |
| 58 | discovered in years with spring burning. In a study of Baltimore Checkerspot butterfly |
| 59 | demography, Brown et al. (2017) estimated the density of caterpillar aggregations on the |
| 60 | landscape using mark-resight surveys. The detection probability ranged from 0.32 to 0.56 on |

native *Chelone glabra* host plants, and from 0.17 to 0.25 on exotic *Plantago lanceolata* plants
(L.M. Browne and E.E. Crone, pers. comm, from analyses in Brown et al. 2017). Surprisingly,
Berberich et al. (2016) found that observers conducting one hour surveys for red wood ant nests
in 60m x 60m plots failed to detect up to 40% of ant nests larger than 50cm. Thus, even for
studies of sessile organisms, detection can be highly imperfect and systematically biased.

Bumble bees are an important group of pollinators with a sessile life-cycle stage. After a 66 queen bumble bee has established a colony and produced her first cohort or two of workers, she 67 remains in the nest until the colony expires (Goulson 2010). Since colonies are the functional 68 69 unit of the population for social insects such as bumble bees, studies of nesting habitat are particularly valuable for conservation planning. Yet, colonies are difficult to find, so relatively 70 few studies have investigated the correlates of nest density compared to the numerous studies of 71 72 habitat preference by foraging workers (but see examples in Harder 1986; Osborne et al. 2008; Waters et al. 2010; Lye et al. 2012; O'Connor et al. 2012, 2017). This disparity is problematic 73 74 because workers are highly mobile, e.g., workers often forage up to 1 km from their colony (Greenleaf et al. 2007) and may be attracted to areas rich in floral resources. This decoupling of 75 foraging and nesting sites potentially obscures the landscape drivers of population performance 76 (Heard et al. 2007). Indeed, Herrmann et al. (2007) reported that bumble bee colony densities 77 were uncorrelated with local worker densities across an agricultural landscape in Germany. 78 A variety of methods have been used to estimate the density of bumble bee nests across 79

particular habitat types for nests (Harder 1986; O'Connor et al. 2012; Rao and Skyrm 2013;

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82 O'Connor et al. 2017), canine-assisted searches (Waters et al. 2010; O'Connor et al. 2012), and

landscapes. These methods have included "free searches" in which observers haphazardly search

83 concentrated stationary observation of small plots by individual researchers or distributed citizen

84 science networks (Osborne et al. 2008; Lye et al. 2012). As an alternative to ground-based 85 searches, molecular (i.e., genetic) analysis of foraging workers has also been used to infer nest density at large spatial scales (Darvill et al. 2004; Goulson et al. 2010). Other studies have 86 87 inferred the relative density of nests across space based on the prevalence of spring prospecting behavior by newly emerged queens (Svensson et al. 2000; Kells and Goulson 2003; O'Connor et 88 89 al. 2017). Combined, these studies have reported considerable variation in nest densities (range: 0.1 to 50.1 nests ha^{-1} ; Table 2 in Appendix 1), potentially owing to ecologically relevant 90 differences across species, habitat types, and landscape configurations. However, O'Connor et al. 91 92 (2012) also reported a 20-fold difference in the number of nests detected between fixed and free searches, indicating an extreme degree of variation in nest detection among survey strategies. 93 Detection probability also often varies through time, across space, and between individuals 94 (Anderson 2001; Kéry and Schaub 2012). Therefore, unaccounted differences in detection 95 probability within and among studies could contribute to observed variation in nest density, 96 97 limiting the ability to generalize across studies and resolve the true environmental correlates of bumble bee population abundance. 98

Here, our objective was to generate unbiased estimates of bumble bee nest density using 99 mark-recapture methods while simultaneously examining the factors that influence imperfect 100 101 detection of nests. We first discuss the general application of closed-population modelling approaches to estimate the abundance of cryptic sessile organisms. We then review a classic 102 catalogue of model structures that can be used to correct for systematic bias in detection 103 104 probability, along with their specific relevance to our study of bumble bee nest density. We fit these models to empirical data and examine the consequences of imperfect nest detection in a 105 106 field setting. Finally, we conduct a simulation to illustrate the (spurious) variability induced into

count data by imperfect detection, and to demonstrate that this approach can be usefully applied
in cases where a large number of sites are only visited twice. Our study emphasizes the utility of
mark-recapture approaches for examining the ecological correlates of nest density for social
insects, and outlines a strategy for surveying bumble bee colonies with imperfect detection.

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1 Mark-recapture for estimating abundance with imperfect detection

Closed population models can be used to estimate abundance while accounting for 112 imperfect detection when the processes of birth, mortality, and movement do not alter the 113 number of individuals in plots over the course of a study (i.e., when the population is 114 115 "demographically closed" across the sampling period). For sessile organisms, this assumption is 116 satisfied when birth and death processes are unlikely to occur across the sampling period. Individuals can also be censored if they are known to have died during the sampling period. 117 118 Otis et al. (1978) outlined a classic catalogue of model structures that can be used to 119 examine drivers of variation in detection probability for closed populations. We adopt this 120 framework for our analysis of bumble bee nests to demonstrate how these models can be applied 121 to studies of sessile organisms and to link our specific study system to a well-defined body of mark-recapture research. Mathematical descriptions of each model are presented in Table 1. 122 123 The simplest model, M_0 , estimates a single detection probability that is common across all nests and sampling occasions. In reality, individuals may differ in their probability of being 124 detected, a phenomenon known as individual heterogeneity. Consequently, the model M_{H} 125 126 estimates both a mean and variance in detection probabilities across nests (i.e., individual 127 random effects). In our study of bumble bee nests, this could be due to unmeasured differences in 128 the worker activity level or the location of nests that increase the probability particular nests will be detected. Similarly, the model M_T estimates a mean detection probability across nests and a 129

130 variance in detection associated with visits (i.e., a temporal random effect). For bumble bees, this 131 is could be due to temporal variation in colony size, ambient temperature (which potentially affects activity level), or seasonal changes in vegetation within plots that alter the probability 132 nests will be detected on each visit. The model M_b described by Otis et al. (1978) accounts for a 133 discrete behavioral change in organisms that affects their individual detection in subsequent 134 135 marking occasions, commonly referred to as "trap-shyness" or "trap-happiness". In studies of sessile organisms, rather than behavioral changes of the study organisms themselves, this 136 137 response can plausibly occur if the vegetation surrounding focal organisms becomes trampled by researchers or if researchers remember the location of individuals (in this case, nests) they have 138 139 previously located. Either of these scenarios would result in different detection probabilities for initial and subsequent capture events in studies of sessile organisms. 140

Although not explicitly described by Otis et al. (1978), fixed effects of explicit covariates 141 142 can also be incorporated to examine the drivers of variation in detection probability (Kéry and 143 Schaub 2012). These can include age or size of the organism, habitat covariates, or explicit temporal covariates (e.g., to examine temporal trends in detection). In our study, four of the eight 144 models included explicit covariates affecting detection of bumble bee nests. Towards this goal, 145 146 we constructed a model that included different detection probabilities for each survey plot 147 $(M_{plot}; \text{ see Study Site in Methods section below})$, and three models that included effects of vegetation height above the nest (M_{veg}) , hour of the day at which plots were surveyed (M_{hour}) , 148 149 and ambient air temperature during the survey (M_{temp}) . Our objective was to demonstrate how 150 these effects could be incorporated to generate deeper insights into the processes influencing nest 151 abundance surveys, rather than to exhaustively examine the diverse suite of (potentially interacting) factors that influence nest detection. 152

153 Methods

154 Study Site

155 We conducted searches for bumble bee nests in three survey plots located at Appleton Farms (42.65°N, 70.86°W) in Ipswich, Massachusetts. Each plot was approximately 3000 m² in 156 area and not used for agriculture, although each plot is mowed annually to prevent succession. 157 158 Two plots were adjacent to one another, while the third plot was located approximately 1,000 159 meters away. Primary vegetation cover in each plot consisted of a variety of grasses, sedges 160 (*Carex* spp.), perennial forbs (e.g., *Plantago lanceolata*, *Linaria vulgaris*, *Lotus corniculatus*, 161 Solidago spp., Asclepias spp.), and ericaceous shrubs (e.g., Vaccinium angustifolium, Rubus spp.). Each plot was bordered by a hedgerow of trees or forest, and the surrounding landscape 162 was mixed agriculture (pasture and hay fields) and natural areas (forest and wet meadows). 163 Data Collection 164 Each plot was visited repeatedly in July and August of 2017, after bumble bee colonies 165 had produced several cohorts of workers, between 6:30 am and 7:30 pm, though the majority of 166 167 nest surveys took place in the morning. Searches were conducted independently by eight different investigators, several of whom surveyed each plot multiple times. Each plot was 168 surveyed for two hours between 12-16 times, for a total effort of 24-32 survey hours per plot. 169 170 During each survey, searchers moved slowly through the plot looking for bumble bee activity that might indicate the presence of nest (i.e., workers quickly descending to the ground, slowly 171 ascending from the ground, or conducting circular navigation flight behavior). Upon locating a 172 173 bumble bee nest, searchers placed an inconspicuous, numbered identifier next to the nest 174 entrance and recorded the nest location, identity, and whether the nest had been previously 175 located either by themselves or by other searchers. For each located nest, we recorded the height

of the tallest vegetation immediately above the nest entrance, which could affect searcher ability
to detect bumble bee movement near nest entrances. We extracted hourly ambient air
temperature measurements during each survey from the nearest weather station (approx. 9.5 km
away); ambient air temperature could affect bumble bee metabolism and nest activity, which
could influence our ability to detect nests. Finally, we also recorded the time of day of each
survey, as bumble bee activity varies throughout the day (Kwon and Saeed 2003).

182 Statistical Methods

The foundation of mark-recapture approaches is the encounter histories of individuals (in this 183 case bumble bee colonies) that are generated from repeated surveys of plots. Closed population 184 models assume the abundance of individuals within each plot does not change across sampling 185 periods; thus, variation in the number of individuals detected across repeated surveys is caused 186 entirely by observation error. The goal of closed population models is to estimate detection 187 probability (*p*) of individuals along with the spatial, temporal, and/or individual-level factors that 188 189 influence it. Once the factors that influence detection probability are estimated, the observed count of individuals can be corrected to generate an unbiased estimate of the true number of 190 191 individuals present at a site.

Successful detection of the individual *i* during each survey *t* occurs with some probability $p_{i,t}$ and the probability of not detecting the individual is $1 - p_{i,t}$. An encounter history of '0110' implies the individual was detected on the second and third survey of the plot and not detected on the first and fourth survey. The entire encounter history '0110' for individual *i* therefore occurs with probability $(1 - p_{i,1}) \times p_{i,2} \times p_{i,3} \times (1 - p_{i,4})$. The actual encounter history (i.e., the observed sequence of 0's and 1's) is assumed to arise from a series of Bernoulli trials. Additionally, a link function can be used to model the influence of covariates on detectionprobability, analogous to a logistic regression.

We fit the series of closed population models described in Table 1 using the empirical nest 200 201 encounter histories generated by our repeated surveys of plots. We fit models using Bayesian methods, outlined by Kéry and Schaub (2012, ch 6); though we note that such models can also 202 203 be fit in a frequentist framework using maximum likelihood approaches. Bayesian analysis 204 allows for random effects to be easily incorporated, for nests to be right-censored part way 205 through the study (e.g., if a nest was known to have failed, which violates an assumption of 206 closed population models), and for uncertainty in parameter estimates to be easily propagated to 207 the model output. The Bayesian implementation of closed population models uses an additional technique called "data augmentation" to estimate the true number of individuals in a plot, based 208 209 on the number of nests actually detected and their estimated detection probabilities (see Kéry and 210 Schaub 2012, ch. 6; Royle and Dorazio 2008, section 5.6 for further discussion of this technique; 211 also see our implementation of this approach in Supplementary Material 1).

212 Simulation Study

213 The methods described above demonstrate the application of a mark-recapture approach to the study of sessile insect life cycle stages when detection is highly imperfect. However, to 214 215 further illustrate the value of this approach, we conducted a simulation to illustrate a second (and 216 converse) problem of failing to account for imperfect detection: imperfect detection generates 217 spurious sampling variability in counts, even when no variation exists. This simulation was also 218 used to examine whether the models we used would converge with only two visits (the minimum 219 number of visits required for mark-recapture) if a larger number of plots had been surveyed. We 220 simulated 40 hypothetical plots, each containing exactly 5 bumble bee nests. We used a fixed

| 221 | detection probability of 0.30 for all nests (approximately equal to the mean detection probability |
|-----|--|
| 222 | we estimated; see Results section and Fig. 1a). We then simulated the number of nests observed |
| 223 | on a single visit to each plot by drawing from a binomial distribution with 5 trials for each plot |
| 224 | (i.e., one for each nest). We used this simulation to evaluate 1) the degree of spurious variability |
| 225 | introduced into counts simply by imperfect detection, and 2) whether the closed population |
| 226 | mark-recapture approach could correct for this spurious variability with only two visits to each |
| 227 | plot. |
| 228 | To facilitate the straightforward application of this analytical toolkit to other studies of |
| 229 | sessile insect life cycle stages, we include full encounter history data, covariate data, and |
| 230 | commented R code for Bayesian analysis and simulations as Supplementary Materials 1-9. Data |
| 231 | management and simulations were conducted in R version 3.4.4 and Bayesian analysis was |
| 232 | conducted in JAGS using the <i>jagsUI</i> library in R (Kellner 2018). |
| 233 | |
| 234 | Results |
| 235 | We located 18 bumble bee nests across the three survey plots (10, 5, and 3 nests in each |
| 236 | plot, respectively). All nests were constructed by Bombus impatiens, except for one that was |
| 237 | constructed by B. bimaculatus. We used all nests for subsequent analysis. The number of nests |
| 238 | located on single visits to each of the three plots ranged from 0 to 6, 0 to 3, and 0 to 3 for each |
| 239 | plot, respectively. The three plots were searched 11, 16, and 14 times by at least 6 different |
| 240 | observers. |
| 241 | Model results – patterns in nest detection |

The mean detection probability of nests based on the intercept-only model (M_0) was 0.30

(95% credible interval [CRI] = 0.24 - 0.36; Fig. 1a). A model that included heterogeneity in p

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| 244 | across nests (M_H) suggested that nests differed in their individual detection probabilities, with a |
|-----|--|
| 245 | median posterior estimate for p of 0.26 (95% CRI = 0.11 to 0.37) and standard deviation (on the |
| 246 | logit scale) of 0.79 (95% CRI = 0.14 to 1.84; Fig. 1b). Similarly, a model that included |
| 247 | heterogeneity in p across visits (M_T) indicated that detection probability varied through time with |
| 248 | a median p of 0.27 (95% CRI = 0.20 to 0.38) and standard deviation (on the logit scale) of 0.67 |
| 249 | (95% $CRI = 0.10$ to 1.29; Fig. 1b). We note that there was substantial uncertainty associated with |
| 250 | estimates of both individual and temporal random effects, as is common for random effect |
| 251 | models fit to relatively sparse data (Kéry and Schaub 2012). |
| 252 | There was weak evidence for different detection probabilities between the first and |
| 253 | subsequent capture occasions (Fig. 1c). Thus, nests were not more likely to be detected after their |
| 254 | initial discovery. |
| 255 | A model including different detection probabilities for nests within each survey plot |
| 256 | (M_{plot}) indicated nest detectability varied systematically across plots (Fig. 1d). Under this |
| 257 | model, median estimates of detection probabilities in each plot were 0.36 (95% $CRI = 0.27$ to |
| 258 | 0.45), 0.23 (95% CRI = 0.14 to 0.33), and 0.26 (95% CRI = 0.15 to 0.41). The probability that |
| 259 | detection probability was greater for nests in plot 1 than plot 2 was 0.98 (calculated directly from |
| 260 | posterior probability distributions). |
| 261 | We then constructed a series of models to examine effects of specific covariates on p. |

Height of vegetation above the nest entrance did not have a strong effect on *p* (standardized effect of vegetation height from $M_{veg} = 0.02$; 95% CRI = -0.02 to 0.06; Fig. 1e). However, *p* declined strongly throughout the day during sampling times (standardized effect of hour from $M_{hour} = -0.68$; 95% CRI = -1.25 to -0.25; Fig. 1e) and was negatively correlated with ambient air temperature during the survey (standardized effect of air temperature from $M_{temp} = -0.38$;

267 95% CRI = -0.69 to -0.09; Fig. 1e).

268 *Model results – nest abundance*

Across all eight models, median estimates of nest abundance were in close agreement.

All models estimated approximately 10, 5, and 3 nests in each of the three plots, respectively

(Fig. 2). The corresponding median estimate of nest density in each plot was therefore 33.3, 16.7,

and 10 nests \cdot ha⁻¹. Consequently, on single surveys of each plot, we located approximately 0-60%

of the nests in plots 1 and 2, and 0-100% of the nests in plot 3.

However, even with our large number of repeated searches (11-16 per survey plot), there

was a high probability that undiscovered nests remained in each plot at the end of our study (Fig.

276 2; note range of credible intervals). For example, while the median estimate of total nest

abundance from model M_0 was 18 (equal to the number of nests we located), the 95% credible

interval was 18 to 20, and the probability that the true abundance was greater than 18 was 0.25.

Notably, the credible intervals for estimates from model M_H were wide relative to other models.

280 This reflects two important features of individual heterogeneity: 1) a fraction of nests have

extremely low detection probabilities and it is difficult to estimate how many remained

undetected, and 2) the existing amount of heterogeneity is difficult to estimate, especially with

low sample sizes (see Fig. 1b).

Given that our counts of bumble bee nests were subject to substantial observation error, we performed an additional analysis to illustrate how imperfect detection can obscure large differences in the density of colonies among plots. We randomly selected nest counts from single surveys to each plot, calculated the resulting rank order of nest densities, and compared them to our estimates from mark-recapture models. We repeated this process 1000 times. This 289 analysis revealed that based on single visits to each field, the incorrect rank-order of density 290 between plots arises 70% of the time despite large differences in the relative density of nests in 291 each plot. Strikingly, the incorrect rank order between plots 1 and 3 arises 17% of the time, 292 despite a three-fold difference in estimated nest density between these two plots (33.3 vs 10 nests ha⁻¹). Next, to quantify the effort needed to reliably estimate differences in nest density 293 between plots, we sequentially re-fit model M₀ for different numbers of visits. With our small 294 295 number of plots and so few nests initially detected, the model would not converge with only 2 296 visits to each plot. This also occurred when models were fit with maximum likelihood in 297 program MARK instead of using Bayesian methods. The model converged with 3 visits to each 298 plot, but uncertainty associated with abundance estimates was extremely large (Fig. 3). After 5 visits to each plot, clear differences in abundance between plots 1 and 3 were apparent. As 299 300 expected, uncertainty in estimates continued to decline as the number of surveys increased.

301 Simulation results – correcting spurious variation in nest abundance

Our final simulation illustrates the sampling variability induced into count data even 302 303 when detection rates are fixed at 0.3. We then used this simulation to examine whether this spurious source of variability could be corrected by visiting each plot only twice if a larger 304 number of plots were visited. Despite the true presence of exactly 5 nests in each of the 40 305 306 simulated plots (Fig. 4; solid black line), the number of nests detected per plot ranged from 0 to 4 with a mean of 1.51 and standard deviation across plots of 0.95 (Fig. 4, dashed line and dots). 307 308 Thus, based on a single visit to each plot, there is considerable (but spurious) variation in nest 309 abundance across the 40 plots. We then simulated a second visit to each plot, fit mark-recapture model M_0 to the resulting encounter histories, and estimated the number of nests in each plot 310 311 while accounting for imperfect detection. The model successfully converged with only 2 visits to

each plot and the credible intervals (Fig. 4; gray ribbons) overlapped the true abundance in every plot. The model M_0 therefore correctly indicates weak evidence for variation in abundance among the 40 simulated plots, despite the high degree of variation in nest counts from a single survey. Thus, for larger sample sizes of plots, two to three visits to each may be sufficient in estimating nest densities.

317

318 Discussion

319 Our study is the first to apply mark-recapture methods to estimate the density of bumble bee 320 nests, which represent a critical and understudied life cycle stage for this important pollinator 321 group. In our study, single surveys of bumble bee nest abundance were subject to considerable bias and observation error, owing to imperfect detection. On average, we only detected 30% of 322 existing nests on each 2-hour survey of a 0.3 ha plot, and we show that the low detection 323 324 probability on single surveys can introduce substantial and spurious variation into counts (Fig. 325 4). Thus, in order to understand the nesting ecology and monitoring requirements of bumble bees, imperfect detection of nests must be properly accounted for. 326

Our estimate of detection probability is well within the range of reported rates for surveys of other sessile organisms: 0.01 to 1.0 in plants (Chen et al. 2013; Kellner and Swihart 2014), 0.17 to 0.60 for insect nests (Berberich et al. 2016; Brown et al. 2017), and 0.09 to 0.93 for patches of freshwater mussels (Reid 2016). These studies use mark-recapture approaches to improve estimates of density or occupancy on the landscape for sessile organisms that are not perfectly detectable, and our study adds bumble bees to this list of taxa. The range of bumble bee nest densities we detected are comparable to those reported in

Osborne et al. (2008), who used intensive fixed searches by citizen science volunteers to count

335 nests in UK gardens and countryside habitats. Of our three plots, the highest density we detected was 33.3 nests ha⁻¹, similar to hedgerow (29.5 nests ha⁻¹), garden (35.8 nests ha⁻¹), and fence line 336 $(37.2 \text{ nests} \cdot \text{ha}^{-1})$ habitats reported in Osborne et al. (2008). Notably, Cumber (1953) is the only 337 other study to report higher nest densities than these; his estimate of 48.6 nests ha⁻¹ was based on 338 intensive free searches of a refuse dump in England. Conversely, our lowest density plot 339 contained 10 nests ha⁻¹, which is similar to the density of 10.9 nests ha⁻¹ reported in Harder 340 341 (1986) who intensively surveyed a successional field in Ontario, Canada. This estimate is also similar to the lowest densities in Osborne (10.8 nests ha⁻¹ in woodland and 11.4 nests ha⁻¹ in 342 short grassland habitat). Therefore, our three plots seem to have captured the range of nest 343 densities observed in other studies, if we restrict these studies to those with intensive search 344 effort and extremely high detection probabilities. 345

346 Other studies have reported far lower nest densities than those in our study or those in 347 Osborne et al. (2008), Cumber (1953), and Harder (1986). However, comparisons among studies are ultimately hampered by differences in survey efforts, and thus, differences in detection error. 348 349 For example, low-intensity free searches by researchers or volunteers produced estimates of nest density ranging from 1.4 to 3.6 nests \cdot ha⁻¹ similar to the range of nest densities discovered by 350 351 bumble bee "sniffer dogs" (O'Connor et al. 2012, 2017). Both of these studies acknowledge that 352 detection error is likely substantial for these methods. Molecular studies also typically yield 353 estimates of nest density that are 1-2 orders of magnitude lower than intensive ground-based searches (range: 0.13 to 1.9 nests ha⁻¹; Supplement 2). Several molecular studies have used ad-354 355 hoc approaches to account for imperfect detection, but these approaches likely under-estimate 356 the true nest density (Goulson et al. 2010). Molecular methods also integrate habitat quality over 357 larger spatial extents than ground-based surveys, and likely incorporate areas that are unsuitable

358 for nesting (e.g., water bodies). Formal mark-recapture approaches are necessary to understand 359 the degree to which variation in nest density between studies is driven by ecologically relevant 360 factors (e.g., variation in habitat quality, differences in spatial scale at which studies occur) 361 versus unresolved differences in imperfect detection. 362 In addition to estimating overall probability of nest detection, we found that nest detection 363 declined when surveys were conducted later in the day and in warmer temperatures (Fig. 1e). Based on estimates from model M_{hour} , mean detection probability during 6 am surveys was 0.40 364 (95% CRI = 0.31 to 0.50), but was only 0.05 (95% CRI = 0.01 to 0.17) for surveys initiated at 6 365 366 pm. This result is consistent with Kwon and Saeed (2003) who found that colony traffic and 367 foraging activity of *Bombus terrestris* declined throughout the day and when temperatures were warmer. Similarly, Couvillon et al. (2010) reported that workers of all sizes conducted fewer 368 foraging trips in warmer temperatures. Thus, the results of our mark-recapture estimate of 369 370 detection probability are broadly consistent with prior knowledge of bumble bee foraging 371 ecology, and suggest that variation in forager behavior is a likely driver of differences in 372 detectability.

Although we did not measure them in our study, other factors could also influence colony 373 374 activity, and in turn, the probability that nests are detected on a given survey. For example, larger 375 colonies have higher traffic at nest entrances (Kwon and Saeed 2003) and are therefore likely to 376 be more detectable. Colony size, in turn, depends on floral resources available throughout the 377 season (Williams et al. 2012; Crone and Williams 2016). Thus, detection of nests may differ between high and low quality habitat owing to systematic differences in colony size. Here, we 378 379 found evidence for systematic differences in nest detection across our three survey plots. Plot 1 had the highest nest density $(33.3 \text{ nests} \cdot ha^{-1})$, and simultaneously, detection was highest for nests 380

in this plot (Fig. 1d). Mark-recapture approaches can correct for these spatio-temporal biases in
 detection probability, which in turn, improve comparisons of abundance within and between
 studies.

384 Population monitoring schemes will often have multiple objectives, in addition to generating reliable estimates of population size (e.g., sampling large numbers of individuals to assess body 385 condition, biochemistry, or disease status; monitoring behavior of individual animals in different 386 387 environmental contexts). For rare or highly cryptic species, intensive surveys over a small area may be required to generate reliable estimates of local abundance, but yield few individuals for 388 389 further detailed study. In the case of bumble bees, intensive fixed searches of small plots are considered sufficient to detect all existing nests and therefore reliably estimate nest density 390 (Osborne et al., 2008; O'Connor et al., 2012). However, intensive fixed searches are inefficient 391 392 when a large number of nests are desired (e.g., for examining microsite characteristics associated with nests, or for studying workers at nest entrances) and are logistically challenging to 393 394 implement over large areas without distributed citizen science networks (Osborne et al. 2008; 395 Lye et al. 2012). We were able to locate a high number of nests, generate precise estimates of abundance with only 5-6 surveys in each of our 3 plots (Fig. 3a), and produce relatively unbiased 396 estimates with an even smaller number of surveys at a larger number of sites (Fig 4). This 397 equates to much lower search effort than typical intensive fixed searches, while simultaneously 398 locating nests at a comparable rate to low-intensity free searches (O'Connor et al. 2012). Our 399 study illustrates the advantage of mark-recapture for optimizing survey protocols for cryptic and 400 401 sessile organisms. Further research in this area will be valuable in illuminating the ecological drivers of pollinator nesting ecology, a critical but understudied subject. 402

403

| 404 | Authors' Contributions |
|-----|--|
| 405 | All authors conceived of research ideas, designed methodology, and collected field data. |
| 406 | DI and EC led statistical analysis and writing of the manuscript. All authors contributed critically |
| 407 | to the drafts and gave final approval for publication. |
| 408 | |
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| 410 | All data and code associated with these analyses will be archived in Dryad Digital |
| 411 | Repository upon acceptance of this manuscript. |
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| 414 | The authors declare they have no conflict of interest. |
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| 419 | References |
| 420 | Anderson DR (2001) The need to get the basics right in wildlife field studies. Wildl Soc Bull |
| 421 | (1973-2006) 29:1294–1297 |
| 422 | Berberich GM, Dormann CF, Klimetzek D, Berberich MB, Sanders NJ, Ellison AM (2016) |
| 423 | Detection probabilities for sessile organisms. Ecosphere 7:e01546. |
| 424 | https://doi.org/10.1002/ecs2.1546 |

- 425 Brown LM, Breed GA, Severns PM, Crone EE (2017) Losing a battle but winning the war:
- 426 moving past preference–performance to understand native herbivore–novel host plant
- 427 interactions. Oecologia 183:441–453. <u>https://doi.org/10.1007/s00442-016-3787-y</u>
- 428 Chen G, Kéry M, Plattner M, Ma K, Gardner B (2013) Imperfect detection is the rule rather than
- the exception in plant distribution studies. J Ecol 101:183–191.
- 430 <u>https://doi.org/10.1111/1365-2745.12021</u>
- 431 Couvillon MJ, Fitzpatrick G, Dornhaus A (2010) Ambient air temperature does not predict
- 432 whether small or large workers forage in bumble bees (*Bombus impatiens*). Psyche: A
- 433 Journal of Entomology 2010:536430. <u>http://dx.doi.org/10.1155/2010/536430</u>
- 434 Crone EE, Williams NM (2016) Bumble bee colony dynamics: quantifying the importance of
- land use and floral resources for colony growth and queen production. Ecol Lett 19:460–
 468. http://dx.doi.org/10.1111/ele.12581
- 437 Cumber R (1953) Some aspects of the biology and ecology of humble-bees bearing upon the
- +57 Cumber R (1955) bonne aspects of the biology and ceology of number bees bearing upon an
- 438 yields of red-clover seed in New Zealand. New Zealand Journal of Science and
- 439 Technology 34:227–240
- 440 Darvill B, Knight ME, Goulson D (2004) Use of genetic markers to quantify bumblebee foraging
- 441 range and nest density. Oikos 107:471–478. <u>https://doi.org/10.1111/j.0030-</u>
- 442 <u>1299.2004.13510.x</u>
- Goulson D (2010) Bumblebees: behaviour, ecology, and conservation, 2nd edn. Oxford
 University Press, New York, pp 317
- 445 Goulson D, Lepais O, O'Connor S, Osborne JL, Sanderson RA, Cussans J, Goffe L, Darvill B
- 446 (2010) Effects of land use at a landscape scale on bumblebee nest density and survival. J
- 447 Appl Ecol 47:1207–1215. <u>https://doi.org/10.1111/j.1365-2664.2010.01872.x</u>

| 448 | Gu W, Swihart RK (2004) Absent or undetected? Effects of non-detection of species occurrence |
|-----|--|
| 449 | on wildlife-habitat models. Biol Conserv 116: 195-203. https://doi.org/10.1016/S0006- |
| 450 | <u>3207(03)00190-3</u> |
| 451 | Harder LD (1986) Influences on the density and dispersion of bumble bee nests (Hymenoptera: |
| 452 | Apidae). Ecography 9:99–103. <u>https://doi.org/10.1111/j.1600-0587.1986.tb01196.x</u> |
| 453 | Heard M, Carvell C, Carreck N, Rothery P, Osborne J, Bourke A (2007) Landscape context not |
| 454 | patch size determines bumble-bee density on flower mixtures sown for agri-environment |
| 455 | schemes. Biol Lett 3:638-641. https://doi.org/10.1098/rsbl.2007.0425 |
| 456 | Herrmann F, Westphal C, Moritz RF, Steffan Dewenter I (2007) Genetic diversity and mass |
| 457 | resources promote colony size and forager densities of a social bee (Bombus pascuorum) |
| 458 | in agricultural landscapes. Mol Ecol 16:1167–1178. https://doi.org/10.1111/j.1365- |
| 459 | <u>294X.2007.03226.x</u> |
| 460 | Kellner KF, Swihart RK (2014) Accounting for imperfect detection in ecology: a quantitative |
| 461 | review. PLoS ONE 9:e111436. https://doi.org/10.1371/journal.pone.0111436 |
| 462 | Kellner KF (2018) jagsUI: A Wrapper Around 'rjags' to Streamline 'JAGS' Analyses. R package |
| 463 | version 1.5.0. https://CRAN.R-project.org/package=jagsUI |
| 464 | Kells AR, Goulson D (2003) Preferred nesting sites of bumblebee queens (Hymenoptera: |
| 465 | Apidae) in agroecosystems in the UK. Biol Conserv 109:165–174. |
| 466 | https://doi.org/10.1016/S0006-3207(02)00131-3 |
| 467 | Kéry M, Schaub M (2012) Bayesian population analysis using WinBUGS: a hierarchical |
| 468 | perspective. Academic Press, Cambridge |
| 469 | Kéry M, Schmidt B (2008) Imperfect detection and its consequences for monitoring for |

470 conservation. Community Ecol 9:207–216. <u>https://doi.org/10.1556/ComEc.9.2008.2.10</u>

- 471 Kwon YJ, Saeed S (2003) Effect of temperature on the foraging activity of *Bombus terrestris*
- 472 L.(Hymenoptera: Apidae) on greenhouse hot pepper (*Capsicum annuum* L.). Appl
- 473 Entomol Zool 38:275–280. <u>https://doi.org/10.1303/aez.2003.275</u>
- 474 Lye GC, Osborne JL, Park KJ, Goulson D (2012) Using citizen science to monitor Bombus
- 475 populations in the UK: nesting ecology and relative abundance in the urban environment.
- 476 J Insect Conserv 16:697–707. <u>https://doi.org/10.1007/s10841-011-9450-3</u>
- 477 O'Connor S, Park KJ, Goulson D (2012) Humans versus dogs; a comparison of methods for the
- detection of bumble bee nests. J Apicult Res 51:204–211.
- 479 <u>https://doi.org/10.3896/IBRA.1.51.2.09</u>
- 480 O'Connor S, Park KJ, Goulson D (2017) Location of bumblebee nests is predicted by counts of
- 481 nest-searching queens. Ecol Entomol 42:731–736. <u>https://doi.org/10.1111/een.12440</u>
- 482 Osborne JL, Martin AP, Shortall CR, Todd AD, Goulson D, Knight ME, Hale RJ, Sanderson RA
- 483 (2008) Quantifying and comparing bumblebee nest densities in gardens and countryside
- 484 habitats. J Appl Ecol 45:784–792. <u>https://doi.org/10.1111/j.1365-2664.2007.01359.x</u>
- 485 Otis DL, Burnham KP, White GC, Anderson DR (1978) Statistical inference from capture data
- 486 on closed animal populations. Wildl Monogr 62: 3–135
- 487 Rao S, Skyrm KM (2013) Nest Density of the Native Bumble Bee, Bombus nevadensis Cresson
- 488 (Hymenoptera: Apoidea), in an Agricultural Landscape. J Kans Entomol Soc 86:93–97.
- 489 <u>https://doi.org/10.2317/JKES120708.1</u>
- 490 Reid S (2016) Search effort and imperfect detection: Influence on timed-search mussel (Bivalvia:
- 491 Unionidae) surveys in Canadian rivers. Knowl Manag Aquat Ecosyst 1–8.
- 492 <u>https://doi.org/10.1051/kmae/2016004</u>

- 493 Slade NA, Alexander HM, Dean Kettle W (2003) Estimation of population size and probabilities
- 494 of survival and detection in Mead's milkweed. Ecology 84:791–797.
- 495 https://doi.org/10.1890/0012-9658(2003)084[0791:EOPSAP]2.0.CO;2
- 496 Svensson B, Lagerlöf J, Svensson BG (2000) Habitat preferences of nest-seeking bumble bees
- 497 (Hymenoptera: Apidae) in an agricultural landscape. Agr Ecosyst Environ 77:247–255.
- 498 https://doi.org/10.1016/S0167-8809(99)00106-1
- 499 Waters J, O'Connor S, Park KJ, Goulson D (2010) Testing a detection dog to locate bumblebee
- colonies and estimate nest density. Apidologie 42:200–205.
- 501 <u>https://doi.org/10.1051/apido/2010056</u>
- 502 White GC, Burnham KP (1999) Program MARK: survival estimation from populations of
- 503 marked animals. Bird Study 46:S120–S139. <u>https://doi.org/10.1080/00063659909477239</u>
- 504 Williams BK, Nichols JD, Conroy MJ (2002) Analysis and management of animal populations.
- 505 Academic Press
- 506 Williams NM, Regetz J, Kremen C (2012) Landscape Scale resources promote colony growth
- 507 but not reproductive performance of bumble bees. Ecology 93:1049–1058.
- 508 <u>https://doi.org/10.1890/11-1006.1</u>
- 509
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512 Figures







518 Fig. 2 Estimated abundance of nests in each of the three survey plots (with associated 95%

519 Bayesian credible intervals) based on each of the eight closed population models and all survey

520 data



Fig. 3 Estimated nests in each plot and associated mean detection probability from closed
population models, based on model M₀ after different numbers of visits to each plot. Points
represent median estimates from Bayesian posterior distribution; lines denote associated 95%
credible intervals



Fig. 4 Results of a simulation with 40 survey plots, each with exactly 5 nests, and each nest with a 0.3 probability of detection on a single visit. Black dashed lines and black dots represent counts on a single visit to each plot. Gray lines and dots depict estimates based on a closed population model with 2 visits to each plot. Gray ribbon indicates 95% credible intervals for estimates in each plot. Thick solid line is true number of nests in each plot (n = 5)

532 Tables

533 Table 1 Classic closed population model structures used in our study to examine variation in the

probability of detecting bumble bee nests. $p_{i,t}$ refers to the detection probability of nest *i* on survey *t*,

535 and β_i refer to various fitted model coefficients

536 537

| # | Model Abbrev. | Model Name | Formula | Description |
|---|-----------------------|-----------------------------|--|---|
| 1 | <i>M</i> ₀ | Intercept-only | $logit(p_{i,t}) = \beta_0$ | Constant value of p across all i nests and t visits |
| | | | Models with Random Effects | |
| 2 | M_H | Individual heterogeneity | $logit(p_{i,t}) = \beta_0 + \alpha_i$ with $\alpha_i \sim Normal(0, \sigma_{nest}^2)$ | Random effect for <i>p</i> among <i>i</i> nests |
| 3 | M_T | Temporal heterogeneity | $logit(p_{i,t}) = \beta_0 + \alpha_t$ with $\alpha_t \sim Normal(0, \sigma_{time}^2)$ | Random effect for <i>p</i> among <i>t</i> visits |
| | | | Models with Categorical Covariate | <u>25</u> |
| 5 | M _b | Capture/recapture effect | $logit(p_{i,t}) = \beta_0 + \beta_1 * recap_{i,t}$ | <i>p</i> differs between initial capture and subsequent recaptures (where $recap_{i,t}$ is an indicator variable set to 0 until initial capture, 1 after initial capture) |
| 4 | M _{plot} | Plot effect | $logit(p_{i,t}) = \beta_0 + \beta_1 * Plot_i$ | <i>p</i> differs among survey plots (i.e., group effect) |
| | | | Models with Continuous Covariates | |
| 6 | M_{veg} | Veg. height | $logit(p_{i,t}) = \beta_0 + \beta_1 * veg_i$ | <i>p</i> depends on vegetation height above the nest entrance |
| 7 | M _{hour} | Hour of day | $logit(p_{i,t}) = \beta_0 + \beta_1 * hour_t$ | <i>p</i> depends on the time of day the survey was conducted |
| 8 | M _{temp} | Air temperature | $logit(p_{i,t}) = \beta_0 + \beta_1 * temp_t$ | <i>p</i> is affected by air temperature during the survey |

538

540 Appendix 1

541 **Table 2** Summary of previous studies that estimated bumble bee nest density, with description of

542 methods used, study region, species detected, habitat types associated with density estimates,

area searched, and estimated density. Note that search area does not apply to molecular methods

as nest density is inferred from genetic relatedness between workers and estimates of worker

545 foraging range

| Study | Method | Study Region | Species Detected | Habitat Type | Search area (ha) | Density (nests·ha ⁻¹) |
|----------------------|---------------|-----------------|---------------------|---------------------|------------------------|--------------------------------------|
| Osborne et al. 2008 | Fixed Search | UK | Multiple | Grassland <10 cm | 0.44 | 11.4 |
| Osborne et al. 2008 | Fixed Search | UK | Multiple | Grassland >10 cm | 0.75 | 14.6 |
| Osborne et al. 2008 | Fixed Search | UK | Multiple | Woodland | 0.19 | 10.8 |
| Osborne et al. 2008 | Fixed Search | UK | Multiple | Fence line | 0.16 | 37.2 |
| Osborne et al. 2008 | Fixed Search | UK | Multiple | Hedgerow | 0.41 | 29.5 |
| Osborne et al. 2008 | Fixed Search | UK | Multiple | Woodland edge | 0.25 | 19.9 |
| Osborne et al. 2008 | Fixed Search | UK | Multiple | Lrg Garden | 0.89 | 34.9 |
| Osborne et al. 2008 | Fixed Search | UK | Multiple | Med Garden | 1.13 | 31.9 |
| Osborne et al. 2008 | Fixed Search | UK | Multiple | Sm Garden | 0.40 | 50.4 |
| Osborne et al. 2008 | Fixed Search | UK | Multiple | Garden (all) | 2.43 | 35.8 |
| O'Connor et al. 2012 | Fixed Search | UK | Multiple | Woodland | 0.14 | 27.8 |
| O'Connor et al. 2012 | Detection Dog | UK | Multiple | Woodland | 6.94 | 1.41 |
| O'Connor et al. 2012 | Free Search | UK | Multiple | Woodland | 6.94 | 1.44 |
| O'Connor et al. 2017 | Free Search | UK | Multiple | Grassland | 5.00 | 3.6 |
| O'Connor et al. 2017 | Free Search | UK | Multiple | Woodland | 5.00 | 3 |
| Cumber 1953 | Free Search | UK | Multiple | Refuse dump | 0.80 | 48.6 |
| Darvill et al. 2004 | Molecular | UK | B. pascuorum | Forest/farmland | NA | 1.9 |
| Knight et al. 2005 | Molecular | UK | B. pascuorum | Farmland | NA | 0.3 |
| Knight et al. 2009 | Molecular | UK | B. pascuorum | Farmland | NA | 1.7 |
| Darvill et al. 2004 | Molecular | UK | B. terrestris | Forest/farmland | NA | 0.13 |
| Knight et al. 2005 | Molecular | UK | B. terrestris | Farmland | NA | 0.3 |
| Knight et al. 2005 | Molecular | UK | B. lapidarius | Farmland | NA | 1.2 |
| Knight et al. 2005 | Molecular | UK | B. pratorum | Farmland | NA | 0.3 |
| Waters et al. 2010 | Sniffer Dog | UK | B. muscorum | Upland Heath | NA | 0.5 |
| Waters et al. 2010 | Sniffer Dog | UK | Multiple | Lowland Heath | NA | 0.27 |

| Waters et al. 2010 | Sniffer Dog | UK | Multiple | Machair | NA | 2.13 |
|--------------------|-------------|-----|---------------|------------|------|-------|
| Waters et al. 2010 | Sniffer Dog | UK | Multiple | Dune | NA | 1.47 |
| Rao and Skyrm 2013 | Free Search | USA | B. nevadensis | Crop field | NA | 18.8 |
| Rao and Strange | | | В. | | | |
| 2012 | Molecular | USA | vosnesenskii | Crop field | NA | 0.76 |
| Harder 1986 | Free Search | CAN | Multiple | Old field | 3.20 | 10.93 |

546

Description of supplementary information 547

| 548 | Supplementary material 1: R code used to examine variation in the probability of detecting |
|-----|---|
| 549 | bumble bee nests using bayesian closed population models and generate Fig. 1 and Fig. 2 |
| 550 | |
| 551 | Supplementary material 2: R code used to demonstrate that imperfect detection can obscure |
| 552 | differences in colony density among plots and generate Fig. 3 |
| 553 | |
| 554 | Supplementary material 3: R code used to estimate bumblebee populations for simulated mark- |
| 555 | recapture data and generate Fig. 4 |
| 556 | |
| 557 | Supplementary material 4: Capture histories for 10 bumble bee nests for survey plot 1 |
| 558 | |
| 559 | Supplementary material 5: Capture histories for 5 bumble bee nests for survey plot 2 |
| 560 | |
| 561 | Supplementary material 6: Capture histories for 3 bumble bee nests for survey plot 3 |
| 562 | |
| 563 | Supplementary material 7: Covariate data for each nest, including the height of the tallest |
| 564 | vegetation immediately above the nest entrance |
| 565 | |

566 Supplementary material 8: Hourly ambient air temperature measurements during each survey

567 from the nearest weather station