

To block or not to block: the adaptive manipulation of plague transmission

S. Gandon¹, L. Heitzmann¹, F. Sebbane²

1. CEFE UMR 5175, CNRS - Université de Montpellier - Université Paul-Valéry Montpellier – EPHE,
1919 route de Mende, 34293 Montpellier, France.

2. Inserm, Univ. Lille, CNRS, CHU Lille, Institut Pasteur de Lille, U1019 – UMR8204 – CIIL– Center for
Infection and Immunity of Lille, F-59000 Lille, France.

The ability of the agent of plague, *Yersinia pestis*, to form a biofilm blocking the gut of the flea has been considered to be a key evolutionary step in maintaining flea-borne transmission. However, blockage decreases dramatically the life expectancy of fleas, challenging the adaptive nature of blockage. Here we develop an epidemiological model of plague that accounts for its different transmission routes, as well as the within-host competition taking place between bacteria within the flea vector. We use this theoretical framework to identify the environmental conditions promoting the evolution of blockage. We also show that blockage is favored at the onset of an epidemic, and that the frequencies of bacterial strains exhibiting different strategies of blockage can fluctuate in seasonal environments. This analysis quantifies the contribution of different transmission routes in plague and makes testable predictions on the adaptive nature of blockage.

Keywords: Plague, transmission, flea blockage, biofilm, parasite manipulation, epidemiology,
evolution, multi-host pathogens

24 **Significance statement:** Plague transmission relies on the ability of infected fleas to inoculate *Y. pestis*
25 bacteria to vertebrate hosts. The production of a biofilm by the bacteria blocks the foregut of the flea
26 and increases infectivity. But the adaptive nature of blockage remains controversial because it has a
27 massive survival cost on the infected fleas and reduces dramatically the length of the infection: an
28 extreme form of the classical virulence-transmission tradeoff. Here we develop a comprehensive
29 model of the multiple routes of plague transmission, we determine when blockage can be viewed as
30 an adaptive manipulation of its flea vector and we generate several testable predictions on the
31 evolution of plague in both endemic and epidemic situations.

32 *Yersinia pestis* is the bacterium that caused 3 plague pandemics and had a profound effect on human
33 history (1). A combination of comparative genomics analyses and experimental studies have unveiled
34 the different evolutionary steps leading to the emergence and the spread of one of the deadliest
35 human pathogen. *Y. pestis* recently emerged from *Yersinia pseudotuberculosis*, a food- and
36 waterborne enteric pathogen causing a benign disease of the digestive tract in humans (2–5). Only a
37 handful of genetic events, including acquisition of genes by horizontal transfer and loss of functional
38 genes, led to the production of flea-borne transmission of plague (3,4,6,7). Notably, the horizontal
39 acquisition of the *Yersinia* murin toxin gene (*ymt*) that protects from a bacteriolytic agent generated
40 during the digestion of the blood meal has been essential to colonize the flea’s midgut and foregut (8).
41 Truncation of the urease accessory protein UreD due to the insertion of a single nucleotide in the *ureD*
42 locus (pseudogenization) reduced the toxicity of the ancestral strain, thereby prolonging the duration
43 of infection in the vector (9,10). Lastly, a series of pseudogenization which led to the loss of the
44 functional accessory regulatory protein RcsA and of two phosphodiesterses (PDE) unlocked the pre-
45 existing capability of the ancestral strain to form a biofilm thanks to the *hmsHFRS* operon, enabling
46 persistent colonization of the proventriculus and ultimately blockage of flea’s gut (7,11).

47 When the proventriculus of the flea is blocked the biofilm prevents the incoming blood to enter
48 the midgut. The blood meal is contaminated upon contact with the bacterial mass, and is regurgitated
49 at the flea-bite site, leading to transmission of plague (12). Another consequence of the blockage is an
50 increase in the biting rate as the flea starves to death. Therefore, blockage is often viewed as a key
51 adaptation of *Y. pestis* because it boosts bacterial transmission by increasing both infectivity (the
52 number of bacteria inoculated in a new host) and the biting rate of infected fleas (7,11). Yet, the
53 adaptive nature of blockage is challenged by the fact that it drastically increases the mortality rate of
54 the flea (7,11). Besides, a combination of experimental observations and empirical studies suggest that
55 other routes of transmission may be involved in plague epidemics (13–16,6,17,18,4). In particular,
56 some flea transmission may also occur in an early-phase of the infection of unblocked flea. In other

57 words, blockage may be viewed as a by-product of the colonization of the foregut but not as an
58 adaptive manipulation of the biting rate of its insect vector.

59 To evaluate the relative importance of blockage on plague transmission we first develop a
60 theoretical framework that accounts for the multiple transmission routes of *Y. pestis* (**Figure 1**). In a
61 second step, we use this theoretical framework to study the evolution of the propensity to block the
62 flea. To analyse pathogen evolution we study the competition between bacterial strains with varying
63 blockage strategies. This competition takes place at a between-host level when bacteria are trying to
64 infect new hosts. But bacteria may also compete within-host when, for instance, a flea is coinfecting by
65 different strains after feeding on two infected hosts. We derive threshold conditions allowing the
66 invasion of a mutant strain with a specific blockage strategy in a stable environment. We also analyse
67 the evolution of the plague during epidemics and show how bacteria with different rates of blockage
68 can fluctuate in a seasonal environment.

69 **The model**

70 *Y. pestis* bacteria can live and/or persist in three different habitats: (i) a vertebrate host, (ii) a flea and
71 (iii) the soil. Our epidemiological model accounts for the complex life-cycle of *Y. pestis* through these
72 three different compartments of the environment (**Figure 1**). For the sake of simplicity we assume that
73 the densities of both the vertebrate host (the host) and the flea (the vector) are constant and equal to
74 N_H and N_F , respectively (we will relax this assumption later on in the analysis). The natural mortality
75 rates of hosts and vectors are m_H and m_F , respectively. Because we are interested in plague evolution
76 we assume that multiple bacterial strains can circulate. We note P_i the density of the free-living stage
77 of the strain i , and I_i the density of hosts infected with the strain i . After feeding on a host infected
78 with strain i , the infected flea is assumed to be “unblocked” state (state $F_{U,i}$). Infectious fleas can
79 become “blocked” (state $V_{B,i}$) and the transition between the “unblocked” and the “blocked” states
80 occurs at a rate ϵ_i (the rate of blockage), which is assumed to vary among different strains of *Y. pestis*.
81 We also assume that blocked fleas can become unblocked (return to the state $F_{U,i}$) at a constant rate

82 γ . Infection increases the mortality of the host (α_H), and the mortality of both the blocked and the
 83 unblocked fleas (α_B and α_U , respectively). It is important to note that blockage has a major impact on
 84 flea survival ($\alpha_B > \alpha_U$) (11,7). Hence, bacterial strains that promote blockage are associated with
 85 higher virulence in the flea because blockage decreases survival. Note, however, that once the infected
 86 flea is blocked (or unblocked) all the strains have the same mortality rates. The host can acquire the
 87 infection horizontally from other infected hosts at a rate $\beta_H I_i$, from the propagules in a contaminated
 88 environment at a rate $\beta_P P_i$ and from the infected vectors at rates $\beta_U F_{U,i}$ and $\beta_B F_{B,i}$. The parameters
 89 β_H , β_P , β_U and β_B modulate the relative importance of these four different routes of transmission.
 90 Crucially, experimental studies have demonstrated that blockage increases the infectiousness of fleas
 91 and thus $\beta_B > \beta_U$ (11,19,20,7). This life cycle can be summarized in the following system of equations
 92 (see **Table S1** for the definition of all the parameter of this model):

$$\begin{aligned}
 S &= N_H - \sum_i I_i \\
 F_S &= N_F - \sum_i (F_{U,i} + F_{B,i}) \\
 \dot{I}_i &= (\beta_H I_i + \beta_P P_i + \beta_U F_{U,i} + \beta_B F_{B,i}) S - (m_H + \alpha_H) I_i \\
 \dot{F}_{U,i} &= \sigma F_S I_i + \gamma F_{B,i} - (m_V + \alpha_V + \epsilon_i) F_{U,i} + \sum_{j \neq i} s[\epsilon_j, \epsilon_i] I_i F_{U,j} - \sum_{j \neq i} s[\epsilon_i, \epsilon_j] I_j F_{U,i} \\
 \dot{F}_{B,i} &= \epsilon_i F_{U,i} - (m_V + \alpha_B + \gamma) F_{B,i} \\
 \dot{P}_i &= \theta I_i - \delta P_i
 \end{aligned} \tag{1}$$

93

94 The above model accounts also for the competition taking place between bacterial strains in the early
 95 stage of the infection (i.e. in unblocked fleas). Indeed, when an unblocked flea infected with strain i
 96 feeds on a host infected by strain j the superinfection function $s[\epsilon_i, \epsilon_j]$ determines the probability that
 97 strain i is replaced by strain j . We assume that the competitiveness of the bacteria may be associated
 98 with the propensity to form biofilms and to block the flea. We used the following function to model
 99 superinfection:

$$s[\epsilon_i, \epsilon_j] = \frac{s_0}{s_0 + (1 - s_0)e^{\frac{-s_0'(\epsilon_j - \epsilon_i)}{s_0(1-s_0)}}} \quad (2)$$

100 where $s_0 = s[\epsilon_i, \epsilon_i]$ is the value of the probability of superinfection at the origin (when both strains
101 have the same value of ϵ) and $s_0' = ds[\epsilon_i, \epsilon_j]/d\epsilon_j|_{\epsilon_i=\epsilon_j}$ is the slope of the superinfection function at
102 the origin (**Figure S1**).

103 Note that we neglect the possibility that competition may occur in blocked fleas and in
104 vertebrate hosts because the bacterial density reached in blocked fleas and in infected hosts hampers
105 invasion by new strains. This is arguably a very simplified view of the way within-host competition
106 among bacterial strains may occur in this system. Yet, as we will see below, the simplicity of this model
107 shows the potential implications of within-host competition on plague evolution and leads to novel
108 adaptive hypothesis for the evolution of blockage.

109 **Epidemiology and evolution in a stable environment**

110 First, we focus on a scenario where the population of the bacteria is monomorphic and all the
111 parameters of the model are constant. The basic reproduction ratio R_0 of the pathogen is given by (see
112 **Appendix**):

$$R_0 = \frac{N_H}{m_H + \alpha_H} \left(\beta_H + \beta_P \frac{\theta}{\delta} + \beta_U \frac{\sigma N_F (m_F + \gamma + \alpha_B)}{A} + \beta_B \frac{\sigma \epsilon N_F}{A} \right) \quad (3)$$

113 with $A = m_F(m_F + \gamma + \epsilon) + \alpha_U(m_F + \gamma) + \alpha_B(m_F + \alpha_U + \epsilon)$. The above expression is useful to
114 identify the relative importance of the different routes of transmission on the epidemiology of plague.
115 Indeed, each term in the parenthesis are associated with the contribution of each of the 4 different
116 routes of transmission to R_0 : (i) direct horizontal transmission, (ii) transmission via propagules, (iii)
117 transmission via unblocked fleas, (iv) transmission via blocked fleas.

118 This expression is also particularly useful to identify the conditions promoting the ability of the
119 pathogen to trigger an epidemic in an uninfected host population where $S = N_H$ and $F_S = N_F$. When

120 $R_0 > 1$, the pathogen can invade the host population and the system reaches an endemic equilibrium
 121 where the host, the vector and the pathogen can coexist (the notation \bar{X} is used to refer to the
 122 equilibrium density of the variable X at this endemic equilibrium). Numerical exploration of the system
 123 (1) revealed that this endemic equilibrium was always locally stable.

124 In the following, we study the long-term evolutionary dynamics of plague using the classical
 125 formalism of Adaptive Dynamics, where mutation rate is assumed to be low which allows decoupling
 126 evolutionary and epidemiological dynamics (21–24). To study plague evolution we derive the invasion
 127 fitness per-generation of a *mutant* strain which has the strategy ϵ_m , at the endemic equilibrium set by
 128 a resident population of the pathogen which has the strategy ϵ (25) (**Appendix**):

$$R_m = \frac{\bar{S}_H}{m_H + \alpha_H} \left(\beta_H + \beta_P \frac{\theta}{\delta} + \frac{\sigma}{A_m} (\beta_U(m_F + \gamma + \alpha_B) + \beta_B \epsilon_m) (\bar{F}_S + s[\epsilon, \epsilon_m] \bar{F}_U) \right) \quad (4)$$

129 with: $A_m = m_F(m_F + \gamma + \epsilon_m) + \alpha_U(m_F + \gamma) + \alpha_B(m_F + \alpha_U + \epsilon_m) + \sigma s[\epsilon_m, \epsilon] \bar{I}(m_F + \gamma + \alpha_B)$.
 130 The mutant will invade the resident population if $R_m > 1$ and this invasion fitness can be used to derive
 131 the gradient of selection on blockage at the endemic equilibrium (i.e. \bar{F}_S , \bar{F}_U and \bar{S}_H) set by the resident
 132 strategy.

133 We used this invasion fitness to identify the conditions leading the evolution of higher rates of
 134 blockage (**Appendix**). In particular, under the assumption that the superinfection function is constant
 135 and equal to s_0 , we find that higher rates of blockage are selected for when:

$$\frac{\beta_B}{m_F + \alpha_B} > \frac{\beta_U}{m_F + \alpha_U + \sigma s_0 \bar{I}} \quad (5)$$

136 Hence, in spite of the complexity of the life cycle, the evolution of blockage boils down to a very simple
 137 condition that does not depend on the other routes of transmission. The left and the right hand sides
 138 of (5) measure of the relative quality of blocked and unblocked fleas, respectively. The quality of a
 139 vector depends on the instantaneous rate of transmission (β_B and β_U) but also the duration of the
 140 infection which is modulated by the mortality rates (m_F , α_U and α_B) as well as the rate of

141 superinfection in unblocked fleas ($\sigma s_0 \bar{I}$). Blockage evolves whenever the blocked fleas are better
142 vectors than unblocked fleas. When condition (5) is satisfied blockage evolves to maximal values. In
143 contrast, when condition (5) is not satisfied, blockage does not evolve and the evolutionary stable
144 strategy is $\epsilon^* = 0$.

145 The invasion condition can also be used to determine the conditions favoring the evolution of
146 blockage when the probability of superinfection depends on the investment in blockage of the
147 competing strains (i.e., $s_0' \neq 0$). For instance, under the simplifying assumption that the resident strain
148 does not block ($\epsilon = 0$) the condition for the invasion of a mutant strain that blocks the flea is:

$$\frac{\beta_B}{m_F + \alpha_B} > \frac{\beta_U}{m_F + \alpha_U + \sigma s_0 \bar{I}} - s_0' B \quad (6)$$

149 where $B = \frac{\beta_U(m_F + \alpha_B + \gamma)}{m_F + \alpha_B} \left(\frac{\bar{F}_U}{\bar{F}_S + s_0 \bar{F}_U} + \frac{\sigma \bar{I}}{m_F + \alpha_U + \sigma s_0 \bar{I}} \right)$.

150 The above condition shows that if the ability to block the flea is associated with a higher competitive
151 ability of the bacteria (i.e., $s_0' > 0$), blockage can evolve more readily. In contrast, if the production of
152 a biofilm is costly and induces a lower competitive ability (i.e., $s_0' < 0$), it is more difficult to evolve
153 blockage. Besides, adding a cost on biofilm production allows some intermediate blockage strategy to
154 be evolutionary stable (**Figure S2**).

155 Evolution in a fluctuating environment

156 Because plague dynamics is often characterized by dramatic temporal fluctuations (26,27) we
157 examined the evolution of blockage away from the endemic equilibrium. Numerical simulations show
158 that, at the onset of an epidemic a mutant strain with a higher ability to block the flea can increase in
159 frequency (**Figure 2**) even if this blockage strategy does not verify conditions (5) or (6). To understand
160 pathogen evolution during this transient phase of the epidemics it is important to track both the
161 *frequency* of the different strains and the *densities* of the pathogen in the different compartments of

162 the model (28–31). In the following, we derive the dynamics of the frequencies p_i^X , of the strain i in
 163 the compartment X :

$$\begin{aligned} \dot{p}_i^I &= \left(\beta_P \frac{P}{I} (p_i^P - p_i^I) + \beta_U \frac{F_U}{I} (p_i^{F_U} - p_i^I) + \beta_B \frac{F_B}{I} (p_i^{F_B} - p_i^I) \right) S \\ \dot{p}_i^{F_U} &= \sigma \frac{I}{F_U} F_S (p_i^I - p_i^{F_U}) + \gamma \frac{F_B}{F_U} (p_i^{F_B} - p_i^{F_U}) - (\epsilon_i - \bar{\epsilon}^{F_U}) p_i^{F_U} \\ &\quad + I \left(\sum_{j \neq i} s[\epsilon_j, \epsilon_i] p_j^I p_j^{F_U} - \sum_{j \neq i} s[\epsilon_i, \epsilon_j] p_j^I p_i^{F_U} \right) \end{aligned} \quad (7)$$

$$\dot{p}_i^{F_B} = \frac{F_U}{F_B} \left((\epsilon_i - \bar{\epsilon}^{F_U}) p_i^{F_U} - \bar{\epsilon}^{F_U} (p_i^{F_B} - p_i^{F_U}) \right)$$

$$\dot{p}_i^P = \frac{\theta I}{P} (p_i^I - p_i^P)$$

164 where $\bar{\epsilon}^{F_U} = \sum_i p_i^{F_U} \epsilon_i$ is the average value of blockage in unblocked fleas.

165 Focusing on the dynamics of mutant frequency is particularly useful to understand the
 166 interplay between epidemiology and evolution. For instance, let us focus on the scenario where two
 167 bacterial strains compete: a mutant strain that blocks the fleas at a rate ϵ_m and a resident strain that
 168 never blocks the fleas. In this case $p_m^{F_B} = 1$ because only the mutant can block the fleas. If we neglect
 169 superinfections and assume the initial frequency of the mutant is low, the above dynamical system
 170 reduces to:

$$\begin{aligned} \dot{p}_i^I &= \left(\beta_P \frac{P}{I} (p_m^P - p_m^I) + \beta_U \frac{F_U}{I} (p_m^{F_U} - p_m^I) + \beta_B \frac{F_B}{I} (1 - p_m^I) \right) S \\ \dot{p}_i^{F_U} &= \sigma \frac{I}{F_U} F_S (p_m^I - p_m^{F_U}) + \gamma \frac{F_B}{F_U} (1 - p_m^{F_U}) - \epsilon_m p_m^{F_U} \\ \dot{p}_i^P &= \frac{\theta I}{P} (p_m^I - p_m^P) \end{aligned} \quad (8)$$

171 Initially, the mutant frequency is expected to be low in all the 3 other compartments of the model (I ,
 172 F_U and P) which yields the following approximation for the change in mutant frequency in the infected
 173 host compartment: $\dot{p}_i^I \approx \beta_B \frac{F_B}{I} S$. This indicates that the mutant frequency is initially increasing in the

174 infected host compartment. This initial increase occurs even if the mutant is ultimately selected against
175 (**Figure 2**). This transient selection for the mutant is due to the fitness benefit associated with higher
176 transmission rates when there are a lot of susceptible hosts around (30,31).

177 If we impose periodic fluctuations in the densities of the host and the vector (e.g. induced by
178 the seasonality of the environment), we observe periodic fluctuations of the incidence of the disease
179 across time. These fluctuations maintain the pathogen away from the endemic equilibrium and can
180 favor different blockage strategies in different phases of these recurrent epidemics. More blockage is
181 selected for at the onset of the epidemics and it is selected against when the epidemics is fading away
182 (**Figure 3**).

183 Discussion

184 The emergence and the evolution of plague results from a series of adaptations that increased the
185 efficacy of flea-borne transmission of *Y. pestis* (3,4,7). But whether or not the blockage of the flea is an
186 adaptation remains a controversial issue (7,14,15,17). Our analysis is an attempt to clarify the
187 conditions that can promote the evolution of blockage. Here we consider a situation where a mutant
188 bacteria with a distinct blockage strategy is introduced in a population of *Y. pestis* and we determine
189 if such a mutant can invade or not. For instance, different genetic variants in the *hmsHFRS* operon are
190 known to affect dramatically the colonization of the proventriculus and the formation of a biofilm: the
191 *hmsFRS+* mutant is known to yield flea blockage while *hmsFRS-* never blocks the fleas and the mortality
192 of fleas blocked by the *hmsFRS+* mutant is considerably larger than unblocked fleas (7,11). Does the
193 gain in transmission due to blockage compensates this increased mortality? Our analysis allows us to
194 answer this question. More specifically, the condition (5) shows that blockage is adaptive, in the
195 absence of within-flea competition, if the ratio of mortality rates between blocked and unblocked fleas
196 is lower than the ratio of transmission rates between blocked and unblocked fleas:

$$\frac{m_F + \alpha_B}{m_F + \alpha_U} < \frac{\beta_B}{\beta_U} \quad (9)$$

197 Available data on blocked and unblocked rat flea *Xenopsylla cheopsis* (one of the main flea vector)
198 suggest that that the life expectancy of a blocked flea is around 2 days while the life expectancy of an
199 infected (but unblocked) flea is around 100 days (11,19,32,7). The ratio between mortality rates of
200 blocked and unblocked fleas is thus expected to be around 50. In other words, condition (9) indicates
201 that transmission rate of blocked fleas must be 50 times higher than transmission rate or unblocked
202 fleas for blockage to be adaptive. Available experimental data on *X. cheopsis* suggests that transmission
203 of blocked fleas is likely to be much higher than this threshold value. First, the ratio of the biting rates
204 of blocked and unblocked fleas is likely to be higher than 3 (19). Second, the number of *Y. pestis*
205 bacteria transmitted by blocked fleas is several order of magnitudes higher (19). Given that
206 regurgitation of a larger inoculum increases the chance of the bacteria to establish a successful
207 infection in the mammalian host, the ratio $\frac{\beta_B}{\beta_U}$ is likely to be higher than a few hundreds. Obviously,
208 obtaining more accurate estimates of transmission and mortality rates in *X. cheopsis* (but also in other
209 flea species) is particularly important to conclude on the adaptive nature of blockage.

210 Our analysis introduces also the possibility of within-flea competition between different
211 variants of *Y. pestis*. In particular, we contend that the production of a biofilm may be a way to
212 outcompete other bacteria in the foregut of the flea. Within-flea competition adds another dimension
213 in the adaptive value of blockage. In particular, conditions (5) and (6) indicate that this mechanism is
214 likely to promote the evolution of blockage. Recent experimental studies have explored the outcome
215 of competition between different strains of *Y. pestis* in fleas (33–36). Unfortunately, experiments
216 following the competition taking place between *hms* variants in the flea remain to be carried out.

217 Empirical evidence of plague dynamics reveal the highly epidemic nature of plague outbreaks
218 which is likely to be driven by seasonal variations of the environment (26,27). In such a fluctuating
219 environment, our analysis reveals that selection for blockage is likely to vary through time. Blockage

220 should be more strongly selected at the onset of epidemics, when many hosts are uninfected. In
221 contrast, blockage is expected to decrease when the epidemic is fading away because a smaller
222 number of susceptible hosts are available. It would be interesting to study the variability of the ability
223 to produce a biofilm and to block the fleas in natural populations. Analysis of bacteria sampled at
224 different points in space or in time would allow to test our prediction that temporal fluctuations in the
225 environment drives the maintenance of variability in *Y. pestis* populations.

226 Even though our model tries to capture multiple routes of transmission, it is important to
227 acknowledge that plague transmission involves a multitude of host species (37). Our model focuses on
228 a simple scenario with a single species of vertebrate host and a single species of flea. Yet, the
229 competence of fleas, their propensity to develop blockage and their mortality rates (after blockage)
230 are known to differ widely (7,32,38). Besides, the infectious blood source is also known to affect the
231 development of *Y. pestis* in the fleas (39). A full understanding of the ecology and evolution of the
232 plague thus requires a more comprehensive description of the network of host and vector species
233 involved in its transmission.

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335

336 **Appendix:**

337 **Derivation of R_0**

338 The ability of the pathogen to invade an uninfected host population is determined by, R_0 , its basic
339 reproduction ratio. To derive R_0 we need to consider the dynamics of equation (1) when $S = N_H$ and
340 $F_S = N_F$:

$$341 \quad \dot{X} = (F - M) \cdot X$$

342 where:

$$343 \quad X = \begin{pmatrix} I \\ F_U \\ F_B \\ P \end{pmatrix}$$

$$344 \quad F = \begin{pmatrix} \beta_H & \beta_U & \beta_B & \beta_P \\ 0 & 0 & 0 & 0 \\ 0 & 0 & 0 & 0 \\ 0 & 0 & 0 & 0 \end{pmatrix}$$

$$345 \quad M = \begin{pmatrix} m_H + \alpha_H & 0 & 0 & 0 \\ -\sigma N_F & m_F + \alpha_U + \epsilon & -\gamma & 0 \\ 0 & -\epsilon & m_F + \alpha_B + \gamma & 0 \\ -\theta & 0 & 0 & \delta \end{pmatrix}$$

346 The basic reproduction ratio is the dominant eigenvalue of $F \cdot M^{-1}$ which yields equation (3) in the
347 main text.

348 **Pathogen evolution**

349 To study pathogen evolution we first track the dynamics of a rare mutant invading the population of a
350 resident pathogen when the system has reached an endemic equilibrium. For the sake of simplicity,
351 we assume that coinfections with the resident and the mutant pathogens are not feasible but we do
352 allow for superinfections in the vector which yields the dynamical system (1). In matrix form this yields
353 the following dynamical system:

$$354 \quad \dot{\mathbf{X}}_m = (\mathbf{F}_m - \mathbf{M}_m) \cdot \mathbf{X}_m$$

355 where:

$$356 \quad \mathbf{X}_m = \begin{pmatrix} I_m \\ F_{U,m} \\ F_{B,m} \\ P_m \end{pmatrix}$$

$$357 \quad \mathbf{F}_m = \begin{pmatrix} \beta_H & \beta_U & \beta_B & \beta_P \\ 0 & 0 & 0 & 0 \\ 0 & 0 & 0 & 0 \\ 0 & 0 & 0 & 0 \end{pmatrix}$$

$$358 \quad \mathbf{M}_m = \begin{pmatrix} m_H + \alpha_H & 0 & 0 & 0 \\ -\sigma N_F - S_1 & m_F + \alpha_U + \epsilon_m + S_2 & -\gamma & 0 \\ 0 & -\epsilon_m & m_F + \alpha_B + \gamma & 0 \\ -\theta & 0 & 0 & \delta \end{pmatrix}$$

359 With $S_1 = s[\epsilon, \epsilon_m] \bar{I}$ and $S_2 = s[\epsilon_m, \epsilon] \bar{V}_I$

360 The basic reproduction ratio is the dominant eigenvalue of $\mathbf{F}_m \cdot \mathbf{M}_m^{-1}$ which yields equation (4) in the
361 main text.

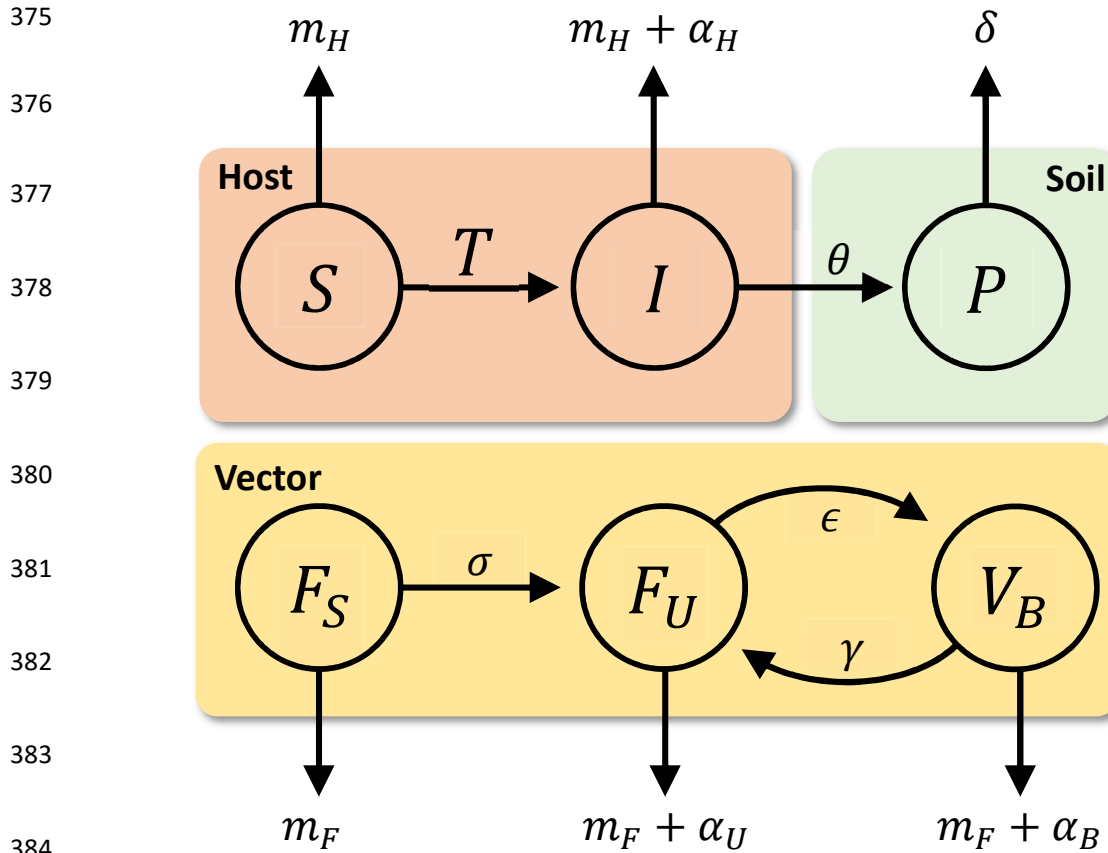
362 Simulations

363 In **Figure 2** we present a simulation of the dynamical system (1) with two strains: one strain never
364 blocks the flea ($\epsilon_1 = 0$) and another strain can block infected fleas ($\epsilon_2 = 1$). To illustrate the dynamics
365 occurring during an epidemic we assumed that none of the vectors are initially infected ($F_S = N_F$) and
366 we introduced a small density of infected hosts: $I_1 = 10^{-4}$ and $I_2 = 10^{-6}$. **Figure 2** shows the
367 epidemiological and the evolutionary dynamics when condition (5) is satisfied or not (panel (B) and
368 (A), respectively).

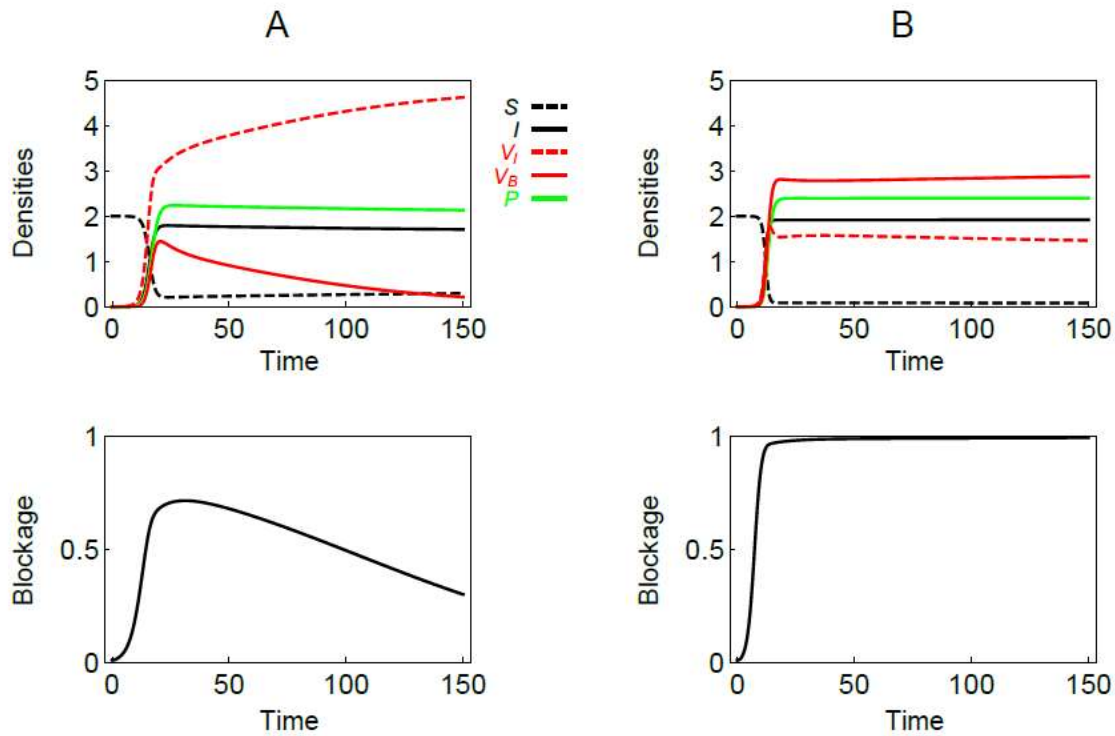
369 In **Figure 3** we present a simulation of the dynamical system (1) under the assumption that N_H and N_F
370 vary periodically because of seasonality with two strains: one strain never blocks the flea ($\epsilon_1 = 0$) and
371 another strain can block infected fleas ($\epsilon_2 = 0.8$). Under the parameter values we chose, the two

372 strains can coexist in the long-term. We show the epidemiological and evolutionary dynamics for 3
373 consecutive seasons, when the system has reached a stable limit cycle.

374



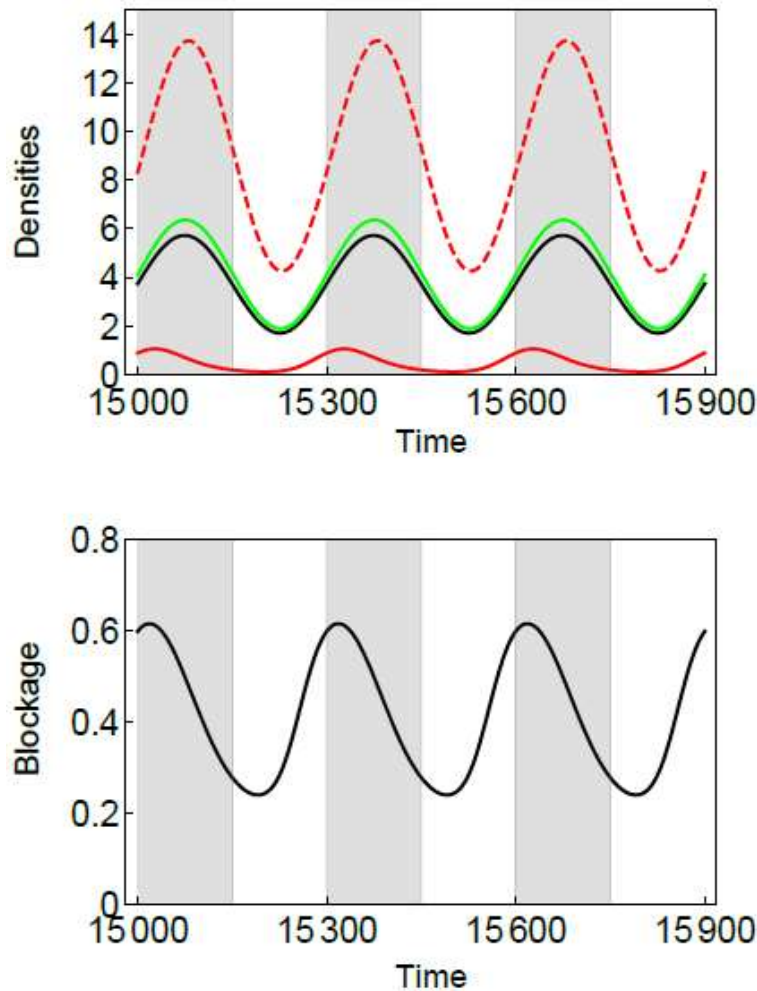
385 **Figure 1: The multiple routes of plague transmission.** Our model accounts for the circulation
 386 of *Y. pestis* in three different habitats: (1) a vertebrate host, (2) the vector and (3) the soil. The
 387 rate at which uninfected hosts become infected is determined by the sum of the force of
 388 infection from the different compartments of this system (see description of life cycle in the
 389 main text): $T = \beta_H I + \beta_P P + \beta_U F_U + \beta_B F_B$.



390

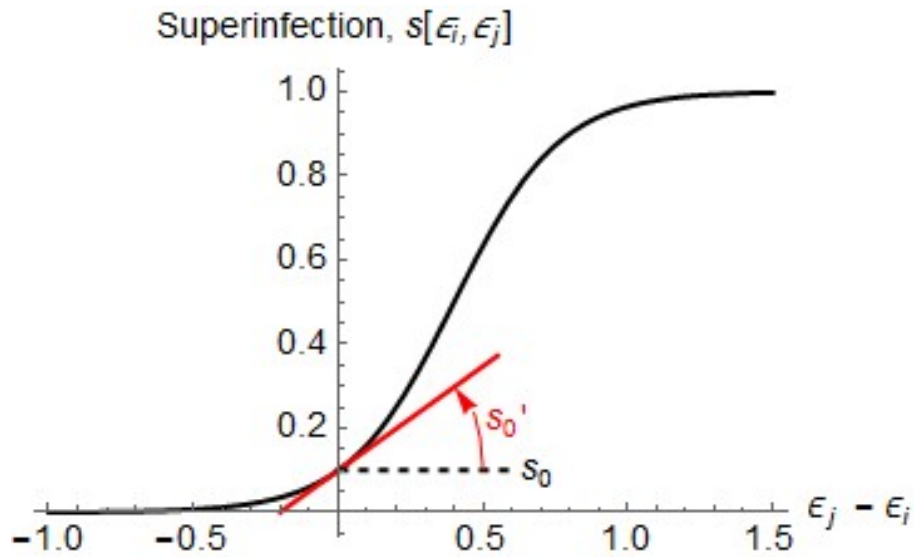
391 **Figure 2: Epidemiology and evolution of plague during an epidemic.** We present the
 392 epidemiological dynamics and the evolutionary dynamics in the absence of superinfection.
 393 The top figures show the dynamics of the densities of the different compartments of the
 394 model during an epidemic. The bottom figures show the dynamics of the mean value of the
 395 blockage strategy. We allow competition between two strains with very different blockage
 396 strategy (i.e. $\epsilon = 0$ or 1). In panel (A) $\beta_B = 0.3$ and blockage is maladaptive according to
 397 condition (5) (i.e., $\frac{\beta_B}{m_F + \alpha_B} < \frac{\beta_U}{m_F + \alpha_U + \sigma s_0 I}$) but blockage is selected for at the beginning of the
 398 epidemic. In panel (B) $\beta_B = 0.7$ and blockage is adaptive according to condition (5) (i.e.,
 399 $\frac{\beta_B}{m_F + \alpha_B} > \frac{\beta_U}{m_F + \alpha_U + \sigma s_0 I}$). Other parameter values (see **Appendix** for more details about the
 400 simulation procedure): $N_H = 2, N_F = 5, \gamma = 0.2, \theta = 1, \sigma = 0.5, \delta = 0.8, m_H = 0.002, m_F =$
 401 $0.01, \alpha_H = 0.1, \alpha_U = 0.01, \alpha_B = 0.2, \beta_H = 0.1, \beta_P = 0.06, \beta_U = 0.05$.

402



403

404 **Figure 3: Epidemiology and evolution of plague in a seasonal environment.** We allow the
 405 densities N_H and N_F to fluctuate periodically with the function $f(t) = 1 + \text{Sin}(2\pi t/T)$,
 406 where $T = 300$ (the shaded area indicates time when $f(t) > 1$). We allow competition
 407 between two strains with very different blockage strategy (i.e. $\epsilon = 0$ or 0.8). The two strains
 408 coexist but their relative frequencies fluctuate with the variations of the abundance of hosts
 409 and vectors. The strain that blocks the flea increases in frequency with the abundance of the
 410 host and the vector. Other parameter values (see **Appendix** for more details about the
 411 simulation procedure): $N_H = 2(1 + f[t])$, $N_F = 5(1 + f[t])$, $\gamma = 0.2$, $\theta = 1$, $\sigma = 0.25$, $\delta =$
 412 0.9 , $m_H = 0.002$, $m_F = 0.04$, $\alpha_H = 0.1$, $\alpha_U = 0.01$, $\alpha_B = 0.2$, $\beta_H = 0.1$, $\beta_P = 0.06$, $\beta_U =$
 413 0.05 , $s_0 = 0.5$, $s'_0 = -2$.

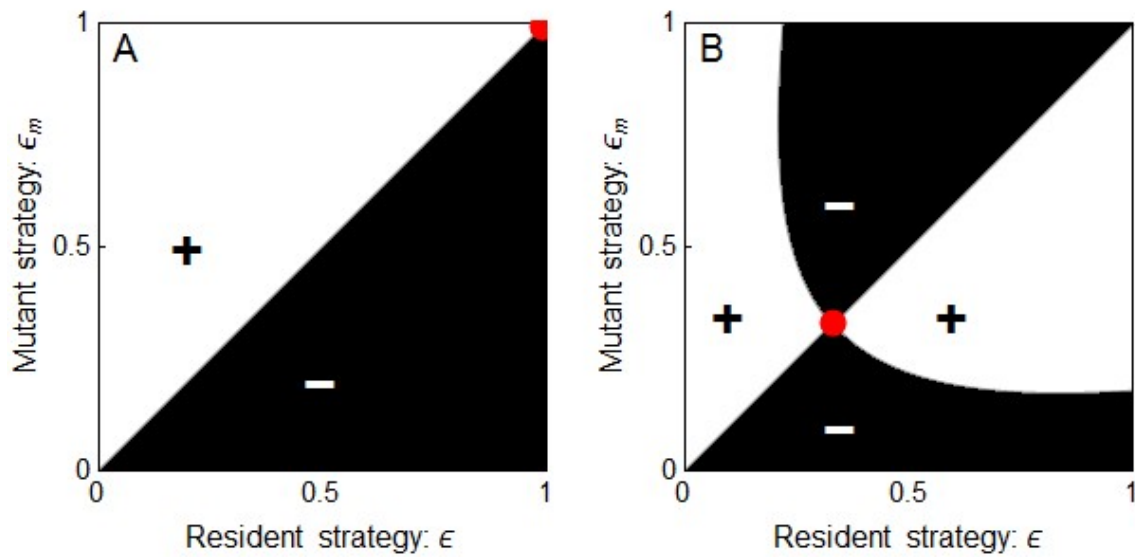


414

415

416 **Figure S1: The superinfection function.** This function measures the probability that a strain j
417 with the blockage strategy ϵ_j , outcompetes a resident strain i with the blockage strategy ϵ_i , in
418 the flea vector (see description of the parameters of the superinfection function in the main
419 text). In the scenario illustrated in the above figure, higher investment in the production of a
420 biofilm is associated with a within-host fitness advantage in the flea vector ($s'_0 > 0$).

421



422

423 **Figure S2: Pairwise invasibility plot on blockage.** We use equation (4) to plot the ability of the
 424 mutant strategy ϵ_m to invade a resident population with strategy ϵ . When $R_m > 1$ the mutant
 425 can invade (white) and when $R_m < 1$ the mutant fails to invade the resident population
 426 (black). In (A) $s'_0 = 0$ and in (B) $s'_0 = -1$. Pairwise invisibility plots can be used to find the
 427 ultimate evolutionary outcome (red dot) but also to identify pairs of strategies that can
 428 coexist. Panel (B) shows that an intermediate strategy can be evolutionary stable. Other
 429 parameter values: $N_H = 1, N_F = 6, \gamma = 0.23, \theta = 5, \sigma = 0.28, \delta = 0.75, m_H = 0.002, m_F =$
 430 $0.04, \alpha_H = 0.8, \alpha_U = 0.01, \alpha_B = 0.2, \beta_H = 0.1, \beta_P = 0.073, \beta_U = 0.02, \beta_B = 0.2, s_0 = 0.5$.

431 **Table S1:** Definitions of the main parameters of the model

Main parameters	Definitions
N_H	Density of the vertebrate host
N_F	Density of the flea vector
S, I	Densities of uninfected and infected hosts
F_S, F_U, F_B	Densities of uninfected, infected-unblocked and infected-blocked fleas
P	Density of free-living <i>Y. pestis</i> bacteria in the soil
β_P	Infectivity of free-living propagules for the host
β_H	Direct transmission rate among hosts
β_U	Infectivity of unblocked fleas
β_B	Infectivity of blocked fleas
m_H	Natural mortality rate of the host
m_F	Natural mortality rate of the flea
δ	Mortality rate of free-living propagules
θ	Rate of production of free-living propagules by infected hosts
α_H	Virulence (mortality induced by the pathogen) in the host
α_U	Virulence (mortality induced by the pathogen) in the unblocked fleas
α_B	Virulence (mortality induced by the pathogen) in the blocked fleas

432