1	Thresholding of cryo-EM density maps by false discovery rate control
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21	

22 Abstract

23 Cryo-EM now commonly generates close-to-atomic resolution as well as intermediate 24 resolution maps from macromolecules observed in isolation and in situ. Interpreting 25 these maps remains a challenging task due to poor signal in the highest resolution 26 shells and the necessity to select a threshold for density analysis. In order to facilitate 27 this process, we developed a statistical framework for the generation of confidence 28 maps by multiple hypothesis testing and false discovery rate (FDR) control. In this 29 way, 3D confidence maps contain separated signal from background noise in the form 30 of local detection rates of EM density values. We demonstrate that confidence maps 31 and FDR-based thresholding can be used for the interpretation of near-atomic 32 resolution single-particle structures as well as lower resolution maps determined by subtomogram averaging. Confidence maps represent a conservative way of 33 34 interpreting molecular structures due to minimized noise. At the same time they 35 provide a detection error with respect to background noise, which is associated with 36 the density and particularly beneficial for the interpretation of weaker cryo-EM 37 densities in cases of conformational flexibility and lower occupancy of bound 38 molecules and ions to the structure.

39 **1. Introduction**

40 Cryo-EM based structure determination has undergone remarkable technological 41 advances over the last several years leading to a sudden multiplication of near-atomic 42 resolution structures (Patwardhan, 2017). Before these transformative changes, only 43 highly regular specimens such as helical or icosahedral viruses were resolved at such 44 detail (Unwin, 2005; Sachse et al., 2007; Zhang et al., 2008; Yonekura et al., 2003; 45 Yu et al., 2008; Ge & Zhou, 2011). With the advent of direct electron detectors 46 (McMullan et al., 2016) and simultaneous improvements in image processing software 47 (Scheres, 2012b; Lyumkis et al., 2013; Punjani et al., 2017), smaller, less regular and 48 more heterogeneous single-particle specimens became amenable to be routinely 49 imaged below 4 Å resolution (Bai et al., 2013; Li et al., 2013; Liao et al., 2013). Recently, the highest resolution structures have become available at ~2 Å resolution 50 (Merk et al., 2016; Bartesaghi et al., 2018; 2015) and sub-4 Å structures below 100 51 kDa from images obtained with and without an optical phase plate have been resolved 52 53 (Merk et al., 2016; Khoshouei et al., 2017). These studies established the technical 54 routines for determining atomic models of structures that were previously thought to 55 be impossible to be resolved by cryo-EM or any other technique (Bai et al., 2015; Galej 56 et al., 2016; Fitzpatrick et al., 2017; Gremer et al., 2017). Electron tomography is the 57 visualization technique of choice for more complex samples including the cellular 58 environment. Due to the poor signal-to-noise ratio (SNR) individual tomograms suffer 59 from substantial noise artifacts. In case tomograms contain identical molecular units 60 they can be averaged by orientationally aligning particle volumes (Briggs, 2013). 61 Recently, with the increase of data quality and improved image processing routines, 62 this approach also yielded near-atomic resolution maps from HIV capsid (Schur et al., 63 2016).

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The resulting reconstructions regardless of whether they originate from single-particle and subtomogram averaging are inherently limited in resolution and suffer from contrast loss at high resolution (Rosenthal & Henderson, 2003). In the raw reconstructions, the high-resolution features are barely visible as the amplitudes follow an exponential decay described by the B-factor quantity that combines effects of radiation damage, imperfect detectors, computational inaccuracies and molecular

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71 flexibility. The Fourier shell correlation (FSC) is the accepted procedure to estimate 72 resolution (Saxton & Baumeister, 1982; van Heel et al., 1982; Rosenthal & Henderson, 73 2003) and can be compared with a given spectral signal-to-noise ratio (SSNR) 74 (Penczek, 2002). Consequently, B-factor compensation by sharpening is essential 75 and common practice. Sharpening is combined with signal-to-noise weighting to limit 76 the enhancement of noise features (Rosenthal & Henderson, 2003). Based on 77 sharpened maps, atomic models are built and further improved by real-space or 78 Fourier-space coordinate refinement (Adams et al., 2010; Murshudov, 2016). This process is particularly challenging at resolutions between 3 and 5 Å commonly 79 80 achieved in cryo-EM. Recently, we proposed a method to sharpen maps by using local 81 radial amplitude profiles computed from refined atomic models (Jakobi et al., 2017). 82 This method facilitates interpretation of densities with resolution variation, but also 83 requires the prior knowledge of a starting atomic model with correctly refined atomic 84 B-factors. Despite this advance, a more general approach is needed at the initial 85 stages of density interpretation in particular in the absence of prior model information. 86 Tracing of amino acids derived from the primary structure as well as placing non-87 protein components into density maps remains a laborious and time-consuming task. 88 In particular, the EM density contains a large dynamic range of gray values for which 89 only a small percentage of voxels is relevant for the interpretation using isosurface-90 rendered thresholded representations. In practice, the process of choosing a threshold 91 is helped by the empirical recognition of binary density features matching those of 92 expected protein features at the given resolution. Therefore, it would be desirable to 93 have more robust density thresholding methods at hand to reduce subjectivity and 94 provide statistical guidance in deciding which map features are considered significant 95 with respect to background noise.

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97 Extracting significant information from noisy data is a common problem in many fields 98 of science. The simplest approach is based on thresholding corresponding to multiples 99 of a standard deviation σ from an expected mean value. The experimental values are 100 only considered significant when above and rejected as noise when below this 101 threshold. In X-ray crystallography and cryo-EM, this σ -approach is commonly applied 102 to the determined maps and σ -thresholds are often reported when isosurface 103 renderings of the density are displayed. In EM maps in particular, the σ -levels reported 104 for interpretation are not universal and will be chosen by the interpreter as they vary 105 from structure to structure between 1 and 5 multiples of σ and often to a smaller extent 106 within the structure. The reason for the observed variation is that the high-resolution 107 amplitudes of density peaks are very weak and can be compromised by noise after 108 sharpening. In statistical theory, it has been recognized that the simple o-method 109 tends to increase the probability of declaring significance erroneously with larger 110 number of tests (Miller et al., 2001), which is referred to as the multiple testing problem. 111 To account for this effect, the probability of correct detection could be increased by 112 controlling the false discovery rate (FDR) (Benjamini & Hochberg, 1995). This 113 statistical procedure has been applied to noisy images in astronomy (Miller et al., 114 2001) and to time recordings of brain magnetic resonance images (Genovese et al., 115 2002) to better discriminate signal from noise.

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117 Due to the low SNRs of cryo-EM maps at high resolution, separating signal from noise 118 remains a daunting task. At present, the visualization and interpretation of the density 119 requires experience of the operator and thus relies on subjectively chosen isosurface 120 thresholds. As sharpening procedures also amplify noise alongside the high-resolution 121 signal, a more robust assessment of the statistical significance of those features by a 122 particular detection error is desirable. Here, we propose to apply the statistical 123 framework of multiple hypothesis testing by controlling the FDR to cryo-EM maps. The 124 resulting maps we refer to as confidence maps represent the FDR of a per-voxel basis 125 and allow the separation of signal from noise background. Confidence maps provide 126 complimentary information to EM densities from single-particle reconstructions and 127 subtomogram averaging as they allow detection of particularly weak features based 128 on statistical significance measures.

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130 2. Methods

131 2.1. Statistical framework

132 In order to overcome limitations in interpreting density features with respect to
133 significance, we apply multiple hypothesis testing using FDR control to cryo-EM maps.
134 In this workflow, we estimate the noise distribution from the background of a

135 sharpened cryo-EM map, apply subsequent statistical hypothesis testing for each 136 voxel and control the FDR (Fig. 1a). For the background noise, we assume a Gaussian 137 distribution or if required an empirical density distribution where the mean and variance 138 of the noise is estimated from four independent density cubes outside the particle 139 density along the central x, y and z axes (**Fig. 1b**). Subsequently, these estimates are 140 used to obtain upper bounds to assess signal from the particle with respect to 141 background noise (see Appendix). In addition, we assume that cryo-EM density to be 142 interpreted is of positive signal (see Results). Therefore, statistical hypothesis tests 143 are carried out by one-sided testing. To account for the total number of voxels and the 144 dependency between voxels, p-values are further corrected by means of a FDR 145 control procedure according to Benjamini and Yekutieli (Benjamini & Yekutieli, 2001). 146 The FDR-adjusted *p*-values (or *q*-values) of each voxel are directly interpretable as 147 the maximum fraction of voxels that have been mistakenly assigned to signal over the 148 background.

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150 As *q*-values of the respective voxels provide a well-established detection measure, we 151 further explored its use for density presentation and thresholding. Based on the FDR, 152 we inverted the map values to the positive predictive value (PPV) by PPV = 1 - FDR. 153 When the map is thresholded at PPV of 0.99, at least 99% of the binarized voxels are 154 truly positive density signal within the map, corresponding to a FDR of 1%. We term 155 these maps confidence maps, referring to the fact that PPVs serve as a measure of 156 the confidence by which we can discriminate signal from the noise. These confidence 157 maps can then be visualized like usual cryo-EM maps with common visualization 158 software, with the difference that the threshold for visualization is now given by 1-FDR 159 rather than the density potential.

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161 2.2. Simulations

The simulated images were 400x400 pixels in size. The scaled grid was generated by adding two orthogonal two-dimensional cosine waves with a period of 5 pixels, where all values smaller than zero were set to 0, and multiplying the sum by a factor of 0.5 in order to scale the maximum to 1. The scaled grid was 200x200 in size and embedded in the center of the 400x400 image. Gaussian distributed noise with a mean

of 0 and given variance of 0.01 (Fig. 1c), 0.1 (Fig. S1a) or 1.33 (Fig. S1b),
respectively, was added to the grid image. Mean and variance for the multiple testing
procedure were estimated outside the scaled grid and the FDR-procedure was carried
out as described. Simulations were implemented in MATLAB (Mathworks Inc.).

171 **2.3. Software**

172 The algorithm is implemented in Python, based on NumPy (Walt et al., 2011) and the 173 mrcfile I/O library (Burnley et al., 2017). Local resolutions were calculated using The 174 ResMap (Kucukelbir al., 2014). software available et is at 175 https://git.embl.de/mbeckers/FDRthresholding. Figures were prepared with UCSF 176 Chimera (Pettersen et al., 2004).

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178 3. Results

179 3.1. FDR-based hypothesis testing yields improved signal detection in 180 simulations

181 In order to evaluate the principal performance of the proposed method on simulated 182 data, we prepared a two-dimensional grid of continuous density waves (Fig. 1c, left). 183 We added white noise to a series of test images containing SNRs between 3.9 and 184 0.3 as they occur in high-resolution shells of 3D reconstructions when the FSC curve 185 drops from 0.67 up to 0.143 often reported as the resolution cutoff. First, we generated 186 a test image with a SNR of 1.2 and noted that in the power spectrum computed from the simulated noise images, signal from high-resolution features cannot be detected 187 188 although being present in the noise-free power spectrum (Fig. 1c, right). The 189 detection of these high-resolution features, however, can be recovered from the 190 corresponding confidence images that we generated as described above, even at 191 SNRs ranging between 3.9 and 0.3 (Fig. S1a-b). When comparing images 192 thresholded at conventional 3.0σ levels with confidence images thresholded at a PPV 193 of 0.99 or FDR of 0.01 (from here on referred to as 1% FDR), we note that FDR-194 controlled thresholding allows more faithful detection of weak density features closer 195 to noise levels. In this way, the density transformation to confidence images minimizes 196 false positive detection of pixels and improves the peak precision as adjacent noise 197 peaks are suppressed (Fig. S2).

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199 **3.2.** Choice of positive density model with Gaussian background noise

200 Although the model of Gaussian noise is often used to approximate background noise 201 in cryo-EM images and maps (Sigworth, 1998; Scheres, 2012a; Kucukelbir et al., 202 2014; Vilas et al., 2018), it is important to analyze actual maps to better understand 203 deviations from this assumption. For this purpose, we analyzed a total of 32 deposited 204 cryo-EM densities from 2 to 8 Å resolution and compared the empirical cumulative 205 density function (CDF) with the ideal Gaussian CDF (Fig. S3a). It is apparent that all 206 of them follow the ideal Gaussian CDF closely. For each map, we assessed normality 207 by Anderson-Darling hypothesis testing (Anderson & Darling, 1954) and found that 208 75% and 87.5% of the maps do not significantly deviate from normality when 209 conservative thresholds corresponding of 1% and 0.1% Family Wise Error Rates 210 (FWER) are chosen (Fig. S3b). One of the reasons for the observed deviations from 211 an idealized Gaussian distribution is a result of the 3D reconstruction procedure. In 212 principle, when truly aligned images containing white Gaussian noise are combined 213 by linear inversion, the obtained 3D volume will also have Gaussian distribution. In 214 practice, in cases when uncertainties reside on the 5 orientation parameters, 215 background noise is not necessarily Gaussian distributed. Moreover, resulting 3D 216 reconstructions will contain local correlations, i.e. "colored noise". Therefore, we 217 analyzed the resulting noise of 3D reconstructions generated from pure noise images 218 with even angular sampling. The resulting amplitude spectrum shows that it differs 219 from pure white noise due to correlations between adjacent pixels (Fig. S3c, left). 220 Furthermore, variances estimated for each voxel from 900 reconstructions show that 221 they can be approximated uniform over the central sphere (Fig. S3c, right).

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223 For the map of EMD-6287, which according to the Anderson-Darling test deviates 224 strongly from normality, we generated a confidence map using the Gaussian and the 225 empirical CDF. We inspected these confidence maps (Fig. S3d) and find that the 226 visual agreement between the two maps is very high. To highlight potential 227 differences, we computed a difference map between the two confidence maps created 228 by the two approaches and observe no systematic variation when deviation from 229 normality is assumed. Therefore, for interpreting confidence maps, small deviations 230 from normality do not appear to have practical limitations. In order to rule out any 231 potential unforeseen effects when maps deviate more strongly, we routinely 232 implemented the monitoring the degree of deviation from ideal Gaussian CDF. For 233 instance when the deviation of the empirical CDF from the Gaussian CDF exceeds 234 0.01, referring to the fact that *p*-values deviate by more than 1 %, can optionally use 235 the empirical CDF for the generation of confidence maps.

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237 The second assumption of the proposed confidence map assumes that protein gives 238 rise to positive density in cryo-EM maps. When inspecting EM density maps, it is 239 evident that not all signal present in the map is positive. Therefore, we analyzed 240 whether significant negative densities can be detected in confidence maps generated 241 from inverted densities. Indeed the confidence maps from negative densities reveal 242 significant signal in regions between protein density often in a spatially complementary 243 way (Fig. S4a left). Using the independently determined X-ray structure of the 20S 244 proteasome (PDB code 1PMA), we tested whether negative density coincides with the 245 atomic model. Overall, negative density has only a very small 2.5 % overlap with 246 atoms, which is close to the predicted false discovery rate of 1 % (Fig. S4b). When 247 using positive density, however, we find that a large fraction of 60 % of the PDB atoms 248 are found in the 1 % FDR contoured confidence map and 10% of that volume is 249 occupied by modelled atoms. In conclusion, we show that negative density presents 250 significant signal in cryo-EM maps, however, that only a very small fraction is occupied 251 by atoms. The largest fraction of negative densities are found next to positive protein 252 density most likely due to the fact that the molecular density is lower than in the particle 253 surrounding solvent area. Based on this analysis and our objective to identify those 254 voxels that arise from protein density, we include the restraint of testing for positive 255 signal into the generation of confidence maps and include an additional option to test 256 for negative signal.

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3.3. Confidence maps from near-atomic resolution maps separate signal from 259 background suited for molecular structure interpretation

260 In order to assess the potential of confidence maps for the interpretation of cryo-EM 261 densities, we applied the algorithm to the near-atomic resolution map of TMV determined at a resolution of 3.35 Å (EMD 2842) (Fromm et al., 2015). Variances 262

263 could be estimated reliably outside the helical rod from a range of different window 264 sizes from 10 to 30 voxels using the cryo-EM density (Fig. S5). To generate the 265 confidence map, we transformed the cryo-EM density to p-values and subsequently 266 to confidence maps in an equivalent manner to the simulated confidence images 267 above. Next, we inspected a longitudinal TMV section through the four helical bundle 268 of the coat protein and compared the confidence map with the cryo-EM density (Fig. 269 2a and b). The confidence map revealed backbone traces that contain values close 270 to 1 corresponding to the helical pitch of the LR helix. They clearly stand out with 271 respect to background noise that is suppressed towards values of 0. The associated 272 histogram of the confidence map revealed a strong peak beyond 0.99 PPV or below 273 1 % FDR separating signal over background and thresholding 5.7 % of voxels within 274 the density. In the case of the deposited cryo-EM map, the subjectively fine-tuned and 275 recommended 1.2 σ threshold also yielded a recognizable outline of helical pitch 276 contours while detecting only 3.7 % of voxels from the density. In analogy to 277 isosurface-rendered cryo-EM densities, confidence map exhibit recognizable 278 structural details, such as the α -helical pitch and many side chains of the central 279 helices (Fig. 2c). When applying a lower FDR of 0.01 %, polypeptide density becomes 280 discontinuous and smaller density features disappear. When going to higher FDR 281 thresholds such as 10 %, noise starts to be included in the density. At the 282 recommended 1 % FDR threshold, the appearance of noise is minimal and well 283 controlled in confidence maps. This is in contrast to cryo-EM densities, where the 284 appearance of noise is very sensitive to small changes in threshold level in particular 285 at lower σ . In fact, the recommended 1.2 σ contour includes only 52 % of the atoms of 286 the model whereas the 1 % FDR threshold contour already contains 73 % with 287 minimized noise. In order to include the same amount of atoms in a contour, a 288 threshold of 0.7 σ would be required, which at the same time will lead to a noticeable 289 increase of obstructing noise. Furthermore, we also examined two additional 290 confidence maps from EMDB model challenge targets determined at near-atomic 291 resolution: 20S proteasome (Campbell et al., 2015) and y-secretase (Bai et al., 2015) 292 (Fig. S6a and S6b). These confidence maps confirm the previous observation that 293 when displayed at FDR levels of 1 %, they provide structural details at near-atomic 294 resolution while effectively separating signal from noise.

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296 3.4. Confidence maps provide a map detection error with respect to 297 background noise

When confidence maps are generated from cryo-EM densities, the main aim of the 298 299 approach is to determine a voxel-based confidence measure of molecular density 300 signal with respect to background noise. In principle, the confidence measure could 301 also be interpreted as a broader error estimate of the EM map referring to the rate of 302 falsely discovered of voxels. The error, however, as it arises from a cryo-EM 303 experiment is a comprehensive quantity, which results from multiple contributions in 304 the form of the solvent scattering, detector noise as well as computational sources of 305 alignment and reconstruction algorithms in addition to variation of signal by multiple 306 molecular conformations and radiation damage effects (Frank & Liu, 1995; Penczek 307 et al., 2006). Estimating the complete series of error contributions including signal 308 variation is currently not possible in the context of common cryo-EM collection 309 schemes. Therefore, the most straightforward way of estimating noise is measuring 310 the variance of the map solvent area. This variance mainly captures errors as they 311 arise from detector noise and solvent scattering while neglecting contributions of 312 computation and local molecular variations. Detector noise can be considered to be 313 distributed uniformly over the 3D reconstruction whereas solvent scattering distribution 314 will not be uniform as pure solvent noise next to the particle is higher when compared 315 with solvent noise projected through the particle due to solvent displacement and 316 variations of water thickness in the particle view (Penczek, 2010). Consequently, 317 measuring noise in the solvent area of cryo-EM maps, will lead to an effective 318 overestimation of noise and therefore to an underestimation of confidence (see 319 Appendix Proposition 1). Although these deviations from a uniform Gaussian noise 320 model do not allow absolute error determination, in practice estimating solvent 321 variance can be used as conservative upper bounds for error rates without including 322 errors arising from computation and molecular variation. In conclusion, the error as it 323 arises from confidence maps should be considered a map detection error with respect 324 to background noise that can assist in the interpretation of cryo-EM densities.

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326 **3.5.** Robustness of FDR-controlled density transformation

327 In order to test the robustness of the approach, we systematically assessed the effects 328 of the required input on the resulting confidence map. First, we tested the influence of 329 severely underestimating noise for confidence map generation by using the 1/2 or 3/4 of the determined variance of the 20S proteasome densities (Fig. S6c). The resulting 330 331 confidence maps displayed at 1 % FDR revealed excessive declaration of background 332 as signal, which poses a principal risk for over-interpretation. This principal risk, 333 however, is less relevant, when the here proposed variance measurements outside 334 the particle is used as we tend to overestimate noise (see above and Appendix 335 Proposition 1). Therefore, we tested the effect of overestimating the variance by 1.25, 336 2 and 8 fold and generated confidence maps according to the defined procedure. The 337 resulting confidence maps show the disappearance of map features at the 1 % FDR 338 threshold only when the variance is severely overestimated by a factor of 8 but for 339 small overestimations is hardly noticeable in the map appearance. Another important 340 noise-related parameter prior to the proposed procedure is the applied sharpening level. Therefore, we tested a series of B-factors from 0 to -250 Å² applied to the 20S 341 342 proteasome maps and converted them into confidence maps. First, with increasing 343 negative B-factors the corresponding confidence maps displayed at 1 % FDR show 344 loss of features due to the drop in relative significance. This is in contrast to crvo-EM 345 densities that become severely over-sharpened and density features are dominated 346 by noise (Fig. S6d). Second, when under-sharpened maps are used for noise 347 estimation, maps will contain only low-resolution features lacking high-resolution detail 348 at the respective significance level in analogy to cryo-EM densities. Therefore, when 349 over-sharpened maps are used for noise estimation, confidence maps inherently avoid 350 enhancement of noise features that could be mistakenly interpreted as signal. 351 Although noise estimation is important for the procedure, tests show that smaller 352 variance overestimation does not have a noticeable effect on map interpretation of 1 353 % FDR confidence maps. In conclusion, confidence maps represent a conservative 354 way of displaying maps at defined significance while avoiding the problem of over-355 sharpening, which represents a principal benefit over visualization of σ -thresholded 356 sharpened EM densities.

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358 **3.6.** Confidence maps facilitate detection of weak density features

359 In order to evaluate further molecular details of the confidence map, we inspected 360 more ambiguous density features of the TMV map. Peripheral density at lower and 361 higher radius of the virus was notoriously difficult to interpret in previous works (Fromm 362 et al., 2015; Sachse et al., 2007; Namba & Stubbs, 1986). For these regions, we found 363 that there are well-defined features present in the 1% FDR confidence maps. Densities 364 of the coat protein for loops Q97 – T103 located at the inner radius and T153 – G155 365 at the outer radius are not present in the respective EM map, but clearly traceable in 366 the 1% FDR confidence map (Fig. 2d, center). In addition, side-chain density for K53 367 contacting the adjacent subunit was found to be clearly significant while being 368 discontinuous in the original map (Fig. 2d, bottom left). Based on confidence maps, 369 readjustment of side-chain rotamers was possible, illustrated for example by 370 significant density for R61, which suggests a realignment of R61 to form stabilizing 371 interactions with aromatic W152 (Fig. 2d, bottom right). The presented examples of 372 TMV illustrate that confidence maps represent an alternative for density display, which 373 can help in the process of molecular feature detection. Although threshold adjustments 374 in cryo-EM maps can also help model interpretation in ambiguous regions and 375 enhance weak density features, they also amplify noise features and increase the risk 376 of noise fitting.

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378 We also tested cases of more heterogeneous densities such as the V-ATPase SidK 379 complex (EMD-8724), which was determined at 6.8 resolution (Zhao et al., 2017). 380 First, the deposited EM map contains very weak density of the bacterial effector SidK 381 EM density due to low occupancy and flexible motion. The corresponding confidence 382 map of the V-ATPase SidK complex reveals that the SidK density is not significant as 383 continuous density when thresholded at 1% FDR as it is too noisy for further analysis 384 (Fig. S7a). Below in section 3.8, we will deal with cases of local resolution and SNR 385 variation that can be accommodated by a locally adjusted FDR procedure. Second, 386 we analyzed confidence maps from three conformational states generated by 3D 387 classification (EMD-8724, EMD-8725, EMD-8726). The generated confidence maps 388 thresholded at 1 % FDR of state 1, 2 and 3 confirm previous observations using EM 389 maps (Fig. S7b). Taken together, confidence maps provide an inherent significance

level associated with the density and minimize false positive noise detection. In this
way, confidence maps can guide atomic model interpretation of cryo-EM density maps
in particular in density regions of ambiguous quality.

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394 **3.7.** Confidence maps from subtomogram averages

395 We further explored whether structures determined at lower resolution may also 396 benefit from the approach. For this purpose, we examined the in situ determined sub-397 tomogram average of the HeLa nuclear pore complex computed from 8 pore particles 398 at 90 Å resolution (Mahamid et al., 2016). The deposited map clearly shows 399 continuous densities for the cytoplasmatic and inner ring molecules whereas density 400 below and above the pore is noisy when visualized at a threshold of 2.0 σ (Fig. 3a). 401 The corresponding the 1% FDR confidence map shows continuous features of the ring 402 structure with minimized noise, which makes interpretation straightforward. In order to 403 generate a confidence map for a subtomogram average structure, care must be taken 404 in identifying areas of noise devoid of any signal in order to estimate the noise variance 405 reliably (Fig. S8a). The same tomograms recorded from lamella of HeLa cells also 406 yielded a subtomogram average of ER-associated ribosomes. The ribosome structure 407 itself could be determined at 35 Å at the membrane with weak density below the 408 ascribed to a translocon-associated protein complex and membrane an 409 oligosaccharyltransferase (Mahamid et al., 2016). The corresponding densities can 410 only be visualized at low thresholds corresponding to 0.8 σ while increasing the 411 amount of background noise and hampering molecular interpretation (Fig. 3b). The 1 412 % FDR confidence maps, however, display the additional protein complexes in the 413 absence of noise. In this case, the confidence map discriminates between specific 414 association of the TRAP complex and the looser association of ribosomes within the 415 polysome assembly. Further, we examined deposited and confidence maps of the 23 416 Å resolution nuclear pore structure determined by subtomogram averaging (Appen et 417 al., 2015) (Fig. 3c). While the overall densities look very similar, we focused our 418 comparison on ambiguous density assignment of the linker region of Nup133. 419 Presence of density in the 1% FDR confidence maps confirms the continuity of this 420 density stretch and the author's interpretation of placing the Nup133 linker region 421 connecting the N-terminal β -propeller and C-terminal α -helical domain (**Fig. 3c, upper**)

right). In addition, we identified additional densities in the connecting densities
between the inner and nuclear as well as the inner and the cytoplasmic ring (Fig. 3c,
bottom). Both densities are not visible at the recommended σ-threshold of 2.1 but
they are reliably displayed in the 1% FDR confidence map. Taken together, confidence
maps generated from lower resolution subtomogram averages as well as from nearatomic resolution reconstructions assist in the density interpretation by separating
signal with respect to background noise.

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430 3.8. Confidence maps benefit from local SNR adjustment in cases of 431 resolution variation

432 After establishing the usefulness for maps covering a range of resolutions, we wanted 433 to further explore how FDR-controlled confidence maps cope with large resolution 434 differences within a single map. For this purpose, we analyzed the very high-resolution 435 2.2 Å map of β -galactosidase (β -gal) (EMD2984) (Bartesaghi *et al.*, 2015) in more 436 detail as it covers resolution ranges from 2.1 to 3.8 Å. In order to reveal high-resolution 437 details in the center of the map, high sharpening levels were required and 438 consequently less well resolved parts in the periphery of the map resulted in over-439 sharpened densities. When we applied our method to the cryo-EM density volume, we 440 found the 1% FDR confidence to be well defined in the center of the map but fading 441 out for large parts of the periphery in support of the B-factor test series on the 20S 442 proteasome (Fig. S8c). We reasoned when resolution differs across the map as a 443 consequence of molecular flexibility and computational errors, the SNR will vary in 444 correspondence. To compensate for these effects, noise levels can be adjusted in 445 cryo-EM maps by applying local low-pass filtrations in Fourier space according to local 446 resolutions (Cardone et al., 2013). Consequently, a local variance can be estimated 447 for each voxel by applying the same low-pass filter to the background noise windows 448 (Fig. S9a). Application of this procedure followed by the FDR control yield a more 449 evenly distributed 1% FDR confidence map including the β -gal periphery (**Fig. 4a, b** 450 top). At the same time, side chain details such as holes in aromatic rings can be 451 resolved at the same significance level as exemplified for W585 in analogy to the 452 appropriately filtered density (Fig. 4a, b bottom). Closer inspection of the cryo-EM 453 density shows that we did not observe density for the peripheral loops of the β -gal

454 complex at the 4.5 σ -threshold but clearly detected continuous loop density at a FDR 455 of 1% of the resolution-compensated confidence map (**Fig. 4c, left and right**). These 456 observations show that the statistical power of the procedure can be improved, i.e. the 457 amount of missed signal can be reduced in cases non-uniform noise levels, while still 458 controlling the FDR by incorporation of local resolution information (see Appendix for 459 detailed discussion).

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461 We recently introduced a local map sharpening tool for cryo-EM maps based on 462 refined atomic B-factors (Jakobi et al., 2017). When refined atomic coordinates are 463 available, the concept of resolution-compensated confidence maps based on adjusted 464 variances derived from local resolution filtering can be easily extended by scaling the 465 radial amplitude falloff of the noise window against the local reference model for estimating the resulting local noise levels (Fig. S9b). In order to directly compare 466 467 confidence maps generated by different filtering or scaling approaches, we focused 468 on the inspection of the peripheral regions of the β -gal enzyme as the densities are 469 weak in particular for loops extending from the particle. When we compared the 470 confidence map of this region generated using the local resolution filtering with the 471 original confidence map, we confirm the observation that adjustments according to 472 local resolutions improve the density connectivity (Fig. S10a, b). When we used the 473 local amplitude scaling approach, we obtained a confidence map with improved 474 density coverage when compared with the original confidence map but less coverage 475 when using local resolution filtering (Fig. S10b, c). In combination, when local variance 476 is estimated based on local amplitude scaling and filtering, we find optimal coverage 477 of density and the atomic model (Fig. S10d). Another example from the EMDB model 478 challenge is the TRPV1 channel determined at 3.4 Å resolution (EMD5578) (Liao et 479 al., 2013). The structure contains a well-defined transmembrane region and a more 480 flexible cytoplasmic domain that is less well resolved. The application of locally 481 adjusted SNRs to the confidence map yields a map with well interpretable density 482 including molecular details (Fig. 4d and 4e). In analogy to the examples above, the 483 cytoplasmic domain is only visible at lower thresholds than the core of the protein. The 484 1 % FDR confidence map captures all density occupied by the protein including the 485 more flexible regions at the cytoplasmic domain. The example of the TRPV1 channel confirms the observation of β-gal that local resolution differences need to be taken into
account for correct generation of confidence maps. When maps exhibit strong local
variation of noise due to molecular flexibility and computational errors, local variances
can be estimated based on local resolution measurements or on local sharpening
procedures and yield well-interpretable confidence maps at a single FDR threshold.

492 **3.9.** Confidence maps confirm detection of bound molecules

493 The majority of near-atomic resolution maps obtained by cryo-EM are in the resolution 494 range between 3 and 4.5 Å. Although main-chain and large side-chain density can 495 often be modeled reliably, smaller side chains and ordered non-protein components 496 such as water molecules and ions are inherently difficult to model at these resolutions 497 and pose the risk of noise fitting. Therefore, we investigated whether confidence maps can help to mitigate this problem and inspected a putative Mg²⁺ site coordinated by 498 499 E416, E461, H418 and three additional H₂O molecules inside of the β -gal enzyme. We rigidly placed of the Mg²⁺ ion and coordinated water molecules based on the 1.6 500 Å resolution X-ray crystal structure (Wheatley et al., 2015) (PDB 4ttg) and superposed 501 502 them onto the deposited EM density map. The map at lower 3.5 σ threshold shows 503 convincing density for only two out three water molecules. (Fig. 5a top left). In 504 contrast, the 1% FDR confidence map based on local variance estimation reveals 505 distinct density peaks for all three suspected H₂O molecules (Fig. 5a top right). 506 Furthermore, β-gal had been imaged in the presence of the small molecule inhibitor 507 PETG. Locating and conformational modeling of the ligand remains challenging due 508 to flexibility and lower occupancy (Fig. 5a bottom left). Ligand placement is facilitated 509 using confidence maps, with density well resolved for the complete small molecule 510 inhibitor (Fig. 5a bottom right). The confidence density confirms previous re-511 refinement of the inhibitor position and conformation (Jakobi et al., 2017). In addition, 512 we also tested whether detection of smaller ions can be facilitated by confidence 513 maps. For this purpose, we turned again to the TRPV1 channel and inspected the 514 density surrounding G643 known as the selectivity filter for the ions passing the 515 channel. The deposited map reveals a density peak in the symmetry center that is 516 compatible with a small ion. In support, the confidence map also shows a density peak 517 at the same position supporting the presence of an ion with a confidence of 1 % FDR 518 (Fig. 5b bottom right). In correspondence, there are multiple cryo-EM structures 519 reporting putative ion densities along an array of carbonyl forming an inner cavity of 520 the channel (Lee & MacKinnon, 2017; McGoldrick et al., 2018). Closer inspection of 521 the y-secretase complex reveals significant density for a membrane-embedded 522 phosphatidylcholine (PC) lipid molecule. In order to detect the two PC acyl chains, the 523 deposited EM map requires thresholding at two different σ -levels of 4 and 5 524 presumably due to differences in chain mobility (Fig. 5c). In contrast, the 525 corresponding 1% FDR confidence map encompasses most of the density of two acyl 526 chains without the need of threshold adjustments. In conclusion, confidence maps 527 from cryo-EM structures possess minimized noise and can be directly used to evaluate 528 the significance of density features to be present by providing a map detection error 529 that e.g. 1 % of the peaks are expected to be falsely discovered. Using complementary 530 information for the interpretation of cryo-EM structures will help to reduce subjectivity 531 involved in the process of density interpretation.

532

533 4. Discussion

534 In the current manuscript, we introduced FDR-based statistical thresholding of cryo-535 EM densities as a complementary tool for map interpretation. This approach is used 536 successfully in other fields of image processing sciences (Genovese et al., 2002). 537 Based on a total of five near-atomic resolution EM maps from the EMDB model 538 challenge (http://challenges.emdatabank.org), one intermediate resolution (6.8 Å) 539 structure and three subtomogram averages in the resolution range between 90 and 23 Å, we showed that using 1% FDR confidence maps are well suited for detailed 540 541 molecular feature detection and result in better confidence in particular for assignment 542 of weak structural features. Although for all maps different σ -levels ranging between 1 543 and 5 could be used for the interpretation of relevant cryo-EM map features, 544 confidence maps thresholded at a common 1 % FDR level show consistent 545 interpretability of molecular features for these maps. The advantage of confidence 546 maps is that they effectively separate signal from a background noise estimate by 547 assigning a confidence scale from 0 to 1 and at 1 % FDR. This way they show 548 consistent inclusion of signal while minimizing noise. In contrast, for cryo-EM densities 549 small changes of the isosurface σ -threshold can have severe consequences for the

interpretability of molecular features and bear the risk of mistakenly including noise.
Therefore, confidence maps and associated FDR thresholds provide a common and
conservative thresholding criterion for the interpretation of cryo-EM maps.

553

554 Included in the algorithm is a direct assessment of the signal significance with respect 555 to background noise associated with particular density features visible in cryo-EM 556 maps, which adds an additional objectivity to reporting of ambiguous density features. 557 Based on these properties, high-resolution confidence maps will be helpful in initial 558 atomic model building when no or little atomic reference structures are available and 559 for assessment of critical details such as side chain conformations and non-protein 560 molecules in the density. The use of these maps will improve the quality of initial 561 atomic models before launching real-space or reciprocal atomic coordinate refinement 562 (Murshudov, 2016; Adams et al., 2010), which should proceed with sharpened or 563 alternatively model-based sharpened maps as refinement targets (Jakobi et al., 2017). 564 The molecular interpretation based on confidence maps is not limited to maps of close-565 to-atomic resolution as we demonstrated its benefit for cases of intermediate 566 resolution single-particle and subtomogram averaging with three maps ranging in 567 resolution from 7 – 90 Å. In these cases, the interpretation of an unassigned density 568 using a confidence level is a beneficial property in particular in the absence of atomic 569 model information.

570

571 We also showed that the generation of confidence maps is a robust procedure. From 572 the sharpened cryo-EM density, we compute the CDF from the solvent background, 573 which in most cases can be approximated by a Gaussian distribution. In addition, we 574 assume protein density to be positive as the overwhelming majority of determined 575 atoms density resides in positive density. Moreover, we find that the region selected 576 for noise estimation is critical as it has to contain pure noise devoid of signal. We found 577 this particularly important for generating confidence maps from subtomogram 578 averages with particle boundaries less well defined. Generally, when estimating 579 background noise outside the particle, we tend to overestimate noise due to smaller 580 ice thickness in particle regions. Smaller deviations from noise estimation show little 581 effect on the conversion to confidence maps (Fig. S6b). We show that when

582 suboptimally sharpened input maps are used to generate confidence maps, the 583 operator avoids the common risk of mistakenly interpreting noise as signal in over-584 sharpened cryo-EM densities. In contrast, confidence maps generated from over-585 sharpened input maps will only result in insufficient declaration of density signal, which 586 is an important safety feature. Once noise is estimated, the procedure of generating 587 confidence maps is statistically clearly defined (Benjamini & Hochberg, 1995; Benjamini & Yekutieli, 2001) and does not contain any free parameters to optimize. 588 589 Only in cases of substantial resolution variation due to molecular flexibility and 590 computational errors, it may be required to locally adjust SNRs by including prior 591 information through local resolution filtering. More sophisticated approaches such as 592 amplitude scaling can also be used in cases where atomic reference structures are 593 available. Adjusting FDR control based on prior information is routinely implemented 594 in other applications of statistical hypothesis testing (Chong et al., 2015; Ploner et al., 595 2006). With the manuscript, we provide a program that requires a 3D volume as input 596 and allows specification of the location of density windows used for noise estimation. 597 The presented implementation including local resolution filtration is computationally 598 fast, taking from 30 s to 2 min on a Xeon Intel CPU for the maps produced in this 599 manuscript.

600

601 We presented several cases in our simulation and EMDB maps where confidence 602 maps displayed weak structural features more clearly while minimizing the occurrence 603 false positive pixels (Figures 1–5). This is a particularly useful property of confidence 604 maps. Weak densities close to inherent noise levels are present in most cryo-EM maps 605 and they result as a consequence of the molecular specimen as well as from the 606 applied computational procedures. For example, they can originate from side chain 607 mobility in the form of multiple rotamers or side-chain specific radiation damage 608 (Fromm et al., 2015; Allegretti et al., 2014; Bartesaghi et al., 2014). In addition, ligands 609 including small organic compounds or larger protein complex components may have 610 lower occupancy or partial flexibility {Zhao:2017hi}. In many complexes, peripheral 611 loops exposed to the solvent tend to have larger molecular flexibility than the core of 612 the protein (Hoffmann et al., 2015). We showed that thresholding confidence maps 613 yield higher voxel detection rates than thresholding in common cryo-EM densities. We

believe that is a result of the fact that the human operator prefers to recommend a more conservative *o*-threshold to avoid excessive inclusion of noise while as a consequence one misses out on signal. Using confidence maps, this type of noise can be suppressed and as a result more reliable signal can be interpreted.

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619 With the increasing number of near-atomic resolution cryo-EM structures, the process 620 of building atomic models has become increasingly important but remains time-621 consuming and labor-intense. Confidence maps can assist the user throughout this 622 process. In X-ray crystallography, multiple complementary maps are being used 623 routinely in the process of model building. Real-space model building and optimization is typically performed using maximum likelihood-weighted 2mFo-DFc, assisted by 624 mFo-DFc difference map used to highlight errors in the model. Various forms of omit 625 626 maps computed from phases of models in which a selection of atoms (e.g. a ligand) 627 has been omitted are used to confirm the presence of ligand and ambiguous density. 628 Similarly, confidence maps display a complementary aspect of cryo-EM maps in 629 helping to reduce ambiguity in density interpretation of e.g. weakly bound ligands, 630 alternative side-chain rotamers, conformationally heterogeneous structures including 631 incomplete or flexible parts of the complex. It is evident that confidence maps would 632 not be suitable for model refinement, as they do not discriminate the scattering mass 633 of different atoms or relative uncertainties of atomic positions. These properties are 634 usually modelled by atomic electron form factors and atomic displacement factors 635 (atomic B-factors). However, owing to the increased precision of density peaks and 636 noise suppression, it is perceivable that confidence maps could be used to guide 637 positional coordinate refinement if implemented as a peak searching procedure. In 638 addition, defined confidence values for density stretches should also be useful and 639 potentially beneficial for automated model building approaches. Interpreting cryo-EM 640 densities by means of an atomic model is often the final step of a cryo-EM experiment. 641 In practice, atomic models are even used as a validation tool to examine density 642 features for side chains at expected positions. One of the key advantages of the here 643 proposed confidence maps is that they can be generated without prior knowledge of 644 an atomic model. As the conversion of cryo-EM densities to FDR controlled maps is a 645 conceptually simple and computationally straightforward, confidence maps could be

routinely consulted for providing complementary information of statistical significance
during the intricate process of interpreting ambiguous densities in cryo-EM structures
resulting from molecular flexibility or partial occupancy.

649

650 Author contributions

M.B. and C.S. initiated the project. M.B. developed and implemented the code for the
algorithm. A.J.J. helped with structure comparison and implementation including
LocScale integration. C.S. supervised the project. M.B. and C.S. wrote the manuscript
with input from A.J.J.

655

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663

664 **Competing financial interests**

- 665 The authors declare that no competing financial interests exist.
- 666

667 **5. Appendix**

668 5.1. Statistical model

For each voxel in the reconstructed 3D volume, where the voxels are indexed with i,j,k, the intensity $X_{i,j,k}$ is modeled as

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 $X_{i,j,k} = \mu_{i,j,k} + \epsilon_{i,j,k} \qquad (1),$

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674 with $\epsilon_{i,j,k}$ a real valued random variable representing the background noise with mean 675 $\mu_{0,i,j,k} \in \mathbb{R}$ and variance $\sigma_{i,j,k}^2 \in \mathbb{R}_{>0}$ and where $\mu_{i,j,k} \in \mathbb{R}$ is the true intensity as 676 observed without background noise. 677 We developed an algorithm by means of multiple hypothesis testing, that controls the 678 maximum amount of false positive signal in the map, i.e. the FDR with respect to 679 background noise. First, we limit the tested voxels to the reconstruction sphere and 680 voxels located outside of a diameter larger than the box size are disregarded as they 681 arise from a smaller subset of averaged images than the voxels inside. Second, we 682 focus on the detection of voxels with positive deviations from background noise (see 683 section 3.2). In addition, voxels that contain significant signal are affected by further 684 sources of noise like flexibility, incomplete binding of ligands and structural 685 heterogeneity, leading to intensity variations of the signal. Consequently, these 686 sources lead to an increase of the variance for these voxels as part of incoherent 687 signal, which we do not consider here as it is going beyond the scope of detecting 688 signal beyond background. Background noise of experimental cryo-EM data, however, 689 poses principal challenges to the statistician, as it can result in non-uniform 690 distributions across the map: although background noise variances from images of 691 uniform noise over the pixels can be assumed uniform over the central sphere (Fig. 692 S3c right), background noise outside the particle is higher when compared with 693 background noise affecting the particle itself due to solvent displacement and 694 variations of relative ice thickness at the particle (Penczek et al., 2006). Therefore, 695 estimating noise in the solvent region outside the particle could lead to an 696 overestimation of the actual influence of the background noise on the particle (see 697 section 3.4). Although this may cause several problems for comprehensive 698 probabilistic modelling, these estimates can be interpreted as conservative bounds for 699 the signal significance of the particle over background noise. For this reason, we use 700 multiple hypothesis testing in order to calculate these upper bounds for detection 701 errors of false positive rates, as we prove in Proposition 1. In cases when alternative 702 noise estimates are available, they can be supplied as additional input to the 703 procedure in order to generate confidence maps.

704

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For each voxel a *z*-test is carried out, which identifies significant deviations from
background noise. The value of the test statistic *Z* at each voxel is then given as

$$z_{i,j,k} = \frac{x_{i,j,k} - \mu_{0,i,j,k}}{\sigma_{i,j,k}}$$
(2),

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709

where $x_{i,j,k} \in \mathbb{R}$ is the reconstructed mean intensity at the respective voxel. We are 710 711 testing for true intensity $\mu_{i,i,k}$ higher than 0, thus the null and alternative hypotheses 712 for each voxel become 713 714 $H_0: \mu_{i,i,k} = 0$ (3). *H*₁: $\mu_{i,i,k} > 0$ 715 716 717 The null hypothesis H_0 states that the true intensity $\mu_{i,i,k}$ at the respective voxel is 0, 718 i.e. no signal beyond background noise, while the second hypothesis H_1 states the deviation towards higher values. Testing for deviations towards negative values, i.e. 719 720 negative densities, is easily accomplished in this setting by multiplying the normalized 721 map intensities $z_{i,i,k}$ with -1, leading to a left-sided test procedure. Both options can

be chosen by the user.

Under the null hypothesis H_0 and by approximating the background noise with a Gaussian distribution (Kucukelbir *et al.*, 2014; Vilas *et al.*, 2018), the test statistic *Z* follows a standard Gaussian distribution. The *p*-values in our procedure are then calculated as

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$$p_{i,j,k} = \begin{cases} P(Z_{i,j,k} \ge z_{i,j,k} | H_0) = 1 - \Phi(z_{i,j,k}), & \text{if } x_{i,j,k} \ge \widetilde{\mu} \\ 1, & \text{if } x_{i,j,k} < \widetilde{\mu} \end{cases}$$
(4),

729

730 with $Z_{i,j,k}$ being the random variable representing the test statistic at voxel *i,j,k*, $z_{i,j,k}$ the particular realization, $\tilde{\mu}$ the background noise as estimated from the solvent area 731 732 and the cumulative distribution function $\Phi()$ of the standard Gaussian distribution. 733 Alternatively, *p*-values can also be calculated in a non-parametric way without any 734 assumptions about the underlying background noise distribution by simply replacing the cumulative distribution function $\Phi($) of the standard Gaussian distribution with the 735 empirical cumulative distribution function $\hat{F}()$ estimated from the sample of 736 737 background noise, given as

738

 $\hat{F}(t) = \frac{number of elements in the sample \le t}{total number of elements in the sample}, \quad t \in \mathbb{R}$ (5).

740

741 This allows the complete procedure to be carried out without any distribution 742 assumptions. However, comparisons show that the background noise can be well 743 approximated with a Gaussian distribution even in the tail areas, which are most 744 important for the calculation p-values (see section 3.2, Fig. 1b and S3a). The 745 respective method for *p*-value calculation, i.e. non-parametric or with Gaussian 746 assumption, can be chosen by the user. All presented cases in the manuscript, if not 747 stated differently, were calculated with the assumption of Gaussian distributed 748 background noise. Note, the here defined *p*-values differ only marginally from the *p*-749 values commonly used for one-sided testing in a way that for all voxels with intensities 750 smaller than the expected mean noise level μ_0 their value is here set to one. This 751 definition allows the control of the FDR in the more general setting of allowed 752 overestimated mean and variance (see Proposition 1).

753

754 **5.2.** Multiple testing correction

The respective hypothesis tests are applied to each voxel in the 3D volume. To account for the multiple testing problem with up to more than a million tests, we choose to control the FDR. Control in this context is meant in giving upper bounds for the occurring error. The FDR is defined as the expected amount of false rejections, i.e.

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$$FDR := \begin{cases} \mathbb{E}\left(\frac{V}{V+R}\right), & \text{if } V+R \neq 0\\ 0, & \text{if } V+R = 0 \end{cases}$$
(6),

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with $V \in \mathbb{N}_0$ the number of false rejections, $R \in \mathbb{N}_0$ the number of true rejections and E() denotes the expectation value. Due to dependencies between hypotheses at voxels close to each other, we choose the Benjamini-Yekutieli procedure (Benjamini & Yekutieli, 2001), giving an FDR-adjusted *p*-value for each voxel, which are often referred to as *q*-values. To describe the adjustment of *p*-values according to Benjamini and Yekutieli in more detail and for the ease of notation, we will now use a sequence of voxels from the map and denote the number of hypotheses, i.e. tested voxels, with 769 *m*. The *p*-values p_i , i = 1, ..., m are then sorted, from small to large, resulting in sorted 770 *p*-values $p_{(i)}$, i = 1, ..., m. *q*-values are then calculated as

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$$q_{(i)} = \min_{i \le k \le m} \left(p_{(k)} \frac{m}{k} \gamma \right) \tag{7},$$

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with *m* the number of hypotheses, *k* a running index and $\gamma = \sum_{l=1}^{m} \frac{1}{l}$. By recognizing the correct index in the sequence of voxels for each index (*i*), *i* = 1, ..., *m* in the sorted array and subsequent conversion into the 3D volume, we can assign each voxel position *i*, *j*, *k* its corresponding *q*-value. In order to interpret the resulting map, the *q*value for each voxel then gives the minimal FDR that has to be imposed at the thresholding in order to call the respective voxel a significant deviation from the background. The final value associated with voxel *i*, *j*, *k*, *q*'_{*i*, *j*, *k*, is then calculated as}

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- 782

$$q'_{i,j,k} = 1 - q_{i,j,k}$$
(8)

783

where $q_{i,j,k}$ is the *q*-value at the voxel indexed with *i,j,k*. Thus, visualization of the map at a value of 0.99 corresponds to a maximal FDR of 1%, or a minimal PPV of 99%, and therefore means that from all the visible voxels at this threshold, a maximum of 1% are expected to be background noise.

788

Next, we show that the presented procedure with *p*-values as defined above controls
the FDR even in the case of overestimated background noise, i.e. by using the
possibly overestimated background noise estimates from the solvent area in Equation
(2) for all voxels.

793

794 Proposition 1:

Consider Gaussian distributed random variables representing the background noise at all voxels *i*, *j*, *k* in the 3D map with true mean $\mu_{0,i,j,k} \in \mathbb{R}$ and variance $\sigma_{i,j,k}^2 \in \mathbb{R}_{>0}$. Moreover, let $\tilde{\mu} \ge \mu_{0,i,j,k}$ and $\tilde{\sigma}^2 \ge \sigma_{i,j,k}^2, \tilde{\mu} \in \mathbb{R}, \tilde{\sigma}^2 \in \mathbb{R}_{>0}$ for all *i*, *j*, *k*, the overestimated background noise parameters. Then $\tilde{q}_{i,j,k} \ge q_{i,j,k}$, where $\tilde{q}_{i,j,k}$ corresponds to the *q*- value as defined in Equation (7) and calculated with our procedure with parameters $\widetilde{\mu}, \widetilde{\sigma^2}$, and $q_{i,j,k}$ the q-value, as obtained with the true parameters $\mu_{0,i,j,k}$ and $\sigma_{i,j,k}^2$.

- 801
- 802 <u>Proof</u>:

In order to prove the statement, we will now recapitulate the algorithm and prove the inequality at all necessary steps. We start showing that the true *p*-value at voxel position *i*, *j*, *k*, $p_{i,j,k}$, is smaller when compared with the *p*-value $\widetilde{p_{i,j,k}}$ calculated from the overestimated background noise parameters using Equation (4). In other words, we want to show that $p_{i,j,k} \leq \widetilde{p_{i,j,k}}$ or equivalent to that, $\widetilde{p_{i,j,k}} - p_{i,j,k} \geq 0$. If $x_{i,j,k} <$ $\widetilde{\mu}$, then the statement is trivial, because $\widetilde{p_{i,j,k}} = 1$ and $p_{i,j,k} \leq 1$, which is a general property of *p*-values.

- 810 For $x_{i,j,k} \ge \tilde{\mu}$, considering Equations (2) and (4), it follows:
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812
$$\widetilde{p_{i,j,k}} - p_{i,j,k} = 1 - \frac{1}{2} (1 + \operatorname{erf}(\frac{x_{i,j,k} - \widetilde{\mu}}{\sqrt{2}\,\widetilde{\sigma}})) - 1 + \frac{1}{2} (1 + \operatorname{erf}(\frac{x_{i,j,k} - \mu_{0,i,j,k}}{\sqrt{2}\sigma_{i,j,k}})) =$$

813
$$-\frac{1}{2} \operatorname{erf}(\frac{x_{i,j,k} - \tilde{\mu}}{\sqrt{2}\,\tilde{\sigma}}) + \frac{1}{2} \operatorname{erf}(\frac{x_{i,j,k} - \mu_{0,i,j,k}}{\sqrt{2}\sigma_{i,j,k}}) \quad (9).$$

814

815 As the error function erf() is monotonically increasing, it is sufficient to show that 816 $\frac{x_{i,j,k} - \mu_{0,i,j,k}}{\sqrt{2}\sigma_{i,j,k}} \ge \frac{x_{i,j,k} - \widetilde{\mu}}{\sqrt{2}\widetilde{\sigma}}$. Because $x_{i,j,k} - \widetilde{\mu} \ge 0$ and thus also $x_{i,j,k} - \mu_{0,i,j,k} \ge 0$, as well as

- 817 $\widetilde{\sigma} \geq \sigma_{i,j,k}$, we have
- 818

819
$$\frac{x_{i,j,k}-\mu_{0,i,j,k}}{\sqrt{2}\sigma_{i,j,k}}-\frac{x_{i,j,k}-\widetilde{\mu}}{\sqrt{2}\widetilde{\sigma}}=\frac{(x_{i,j,k}-\mu_{0,i,j,k})\widetilde{\sigma}-(x_{i,j,k}-\widetilde{\mu})\sigma}{\sqrt{2}\widetilde{\sigma}\sigma_{i,j,k}}\geq\frac{(x_{i,j,k}-\mu_{0,i,j,k})\sigma-(x_{i,j,k}-\widetilde{\mu})\sigma}{\sqrt{2}\widetilde{\sigma}\sigma_{i,j,k}}=$$

820
$$\frac{\left(-\mu_{0,i,j,k}+\widetilde{\mu}\right)\sigma}{\sqrt{2}\widetilde{\sigma}\sigma_{i,j,k}} \ge 0 \quad (10),$$

821

where in the last inequality it was used that $\tilde{\mu} \ge \mu_{0,i,j,k}$ and $\tilde{\sigma} \ge \sigma_{i,j,k} > 0$. This gives the desired result of $\tilde{p}_{i,j,k} \ge p_{i,j,k}$.

824 Recapitulating the calculation of *q*-values in Equation (7) together with the conversion 825 of the 3D volume to a sequence, it follows:

826

827

$$q_{(a)} = \min_{a \le k \le m} \left(p_{(k)} \frac{m}{k} \gamma \right) \le \min_{a \le k \le m} \left(\widetilde{p_{(k)}} \frac{m}{k} \gamma \right) = \widetilde{q_{(a)}}, \ a = 1, \dots, m$$
(11),

828

829 with *m* the number of hypotheses, *k* a running index and $\gamma = \sum_{l=1}^{m} \frac{1}{l}$. This gives the 830 desired result:

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- 832

 $\widetilde{q_{i,j,k}} \ge q_{i,j,k} \tag{12}.$

833 834

835 As the Benjamini-Yekutieli procedure controls the FDR when using true parameters, 836 our procedure (i.e. Benjamini-Yekutieli applied to the modified *p*-values) will give a 837 more conservative estimate of the FDR (as shown in Proposition 1). Therefore, our 838 algorithm controls the FDR sufficiently well by giving an upper conservative bound for 839 the FDR. Thus, Propositon 1 states that even in the setting of non-uniform background 840 noise with higher noise levels in the region of background noise estimation, the FDR 841 is controlled and thus robust in the sense that the maximum FDR is still guaranteed. 842 Furthermore, it has to be mentioned that estimates of the background noise levels are 843 not the only factor contributing to FDR estimation. Both the number voxels as well as 844 their dependencies within the map have an important influence and are considered in 845 the FDR-adjustment. This makes the generation of confidence maps even with 846 severely overestimated background noise parameters a powerful procedure (Fig. S6), 847 where powerful is used here in its statistical sense of decreasing the error of missing 848 true signal. However, the power of the procedure can be even increased, i.e. the 849 amount of true missed signal reduced while controlling the FDR, by including 850 information about local resolutions, cutoffs in reciprocal space where no signal is 851 expected beyond, while, at the same time, controlling the FDR.

852

853 5.3. Testing with local filtering

In the presence of extreme resolution variation, using uniformly sharpened and filtered maps will lead to confidence maps of insufficient representation of features in both areas of either lower than the average B-factor or higher than the average B-factor. 857 Therefore, in the next two sections, we will show how noise levels can be locally 858 adjusted and subsequently estimated by inclusion of local resolution information as 859 well as atomic B-factors and how this can be used to increase the power to detect 860 weaker features while controlling the FDR. Local filtration of EM maps according to 861 the local resolution (Cardone *et al.*, 2013) has been shown to be a powerful approach 862 as it leads to local reductions of background noise. These variations of noise levels 863 between different voxels at different resolutions from local filtering, can be also 864 accounted for in the generation of confidence maps. For each voxel, a map duplicate 865 volume is filtered at the corresponding resolution and the noise distribution estimated 866 from the solvent area outside the particle. This procedure results in three 3D maps, 867 the estimates of local variances of the background noise at each voxel after local 868 filtration, the estimates of local means of the background noise at each voxel after 869 local filtration and the locally filtered map. These three maps are subsequently used 870 for the testing procedure. Thus, the value of the test statistic (2) is calculated by

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$$z_{i,j,k} = \frac{x_{i,j,k} - \tilde{\mu}_{i,j,k}}{\tilde{\sigma}_{i,j,k}}$$
(13),

873

where $x_{i,j,k} \in \mathbb{R}$ is the intensity of the locally filtered map at voxel i, j, k and $\tilde{\mu}_{i,j,k} \in \mathbb{R}$ and $\tilde{\sigma}_{i,j,k} \in \mathbb{R}_{>0}$ are the local mean and standard deviation estimate of the background noise at the respective voxel. All subsequent steps of the algorithm remain identical as well as the validity of Proposition 1.

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879 5.4. Testing with local amplitude scaling

As for the local filtration, local amplitude scaling gives rise to varying noise levels at different voxels. In order to obtain both mean and variance estimates for each voxel after local amplitude scaling, a duplicate window outside the particle containing pure noise is scaled according to the rolling window used in local amplitude scaling for each voxel, i.e. the amplitudes of the Fourier transform of the box containing pure noise at frequency *s*, denoted as $F_{noise}(s)$, are multiplied with a frequency dependent sharpening factor $k(s) \in \mathbb{R}_{\geq 0}$, which is consequently given as

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$$k(s) = \begin{cases} \frac{F_{sharpened}(s)}{F_{observed}(s)}, & \text{if } F_{observed}(s) \neq 0\\ 0, & \text{if } F_{observed}(s) = 0 \end{cases}$$
(14),

889

where $F_{sharpened}(s) \in \mathbb{R}_{\geq 0}$ and $F_{observed}(s) \in \mathbb{R}_{\geq 0}$ are rotationally averaged amplitudes of the Fourier transform at frequency *s* given at the respective rolling window for the sharpened and the observed experimental map, respectively. The noise distribution is then estimated from the scaled noise sample. In analogy to the case of locally filtered maps, this procedure results again in three 3D maps of estimated means, variances and intensities of the locally sharpened map for each voxel that can be incorporated with Equation (13) in the testing procedure. Proposition 1 remains valid.

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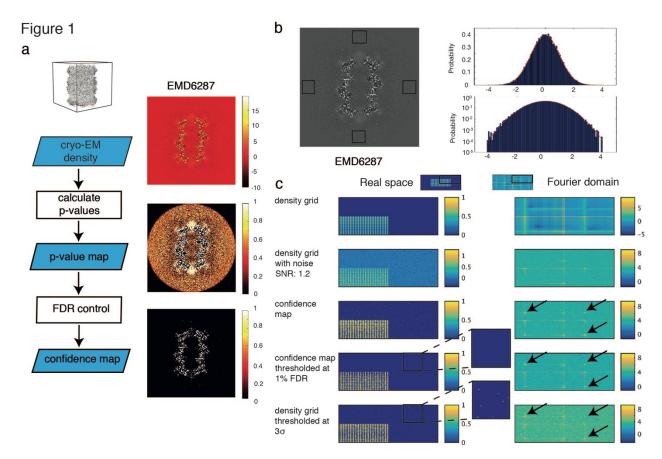
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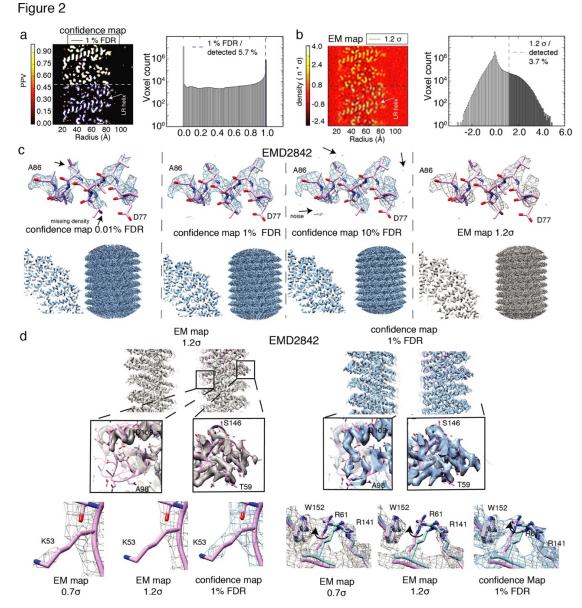
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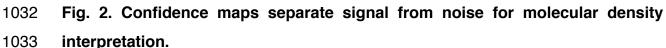
1013 Fig. 1. False discovery rate (FDR) analysis of cryo-EM maps.

1014 (a) Left. Flowchart of confidence map generation: the cryo-EM map is converted to pvalues and finally FDR controlled. Right. Slice views through a cryo-EM map of 20S 1015 1016 proteasome (EMD6287) depicted at the respective stages of the algorithm (blue 1017 boxes) on the left. Note, the strong increase in contrast when the sharpened map is 1018 converted to the confidence map. (b) Left. Estimation of the background noise from 1019 windows (red) outside the particle. Right. Histograms (top. probability in linear scale, 1020 bottom. probability in log-scale) of the background window together with the probability 1021 density function of the estimated Gaussian distribution. (c) Evaluation of the algorithm 1022 on a simulated 2D density grid. The upper right guadrant of images in real space (left 1023 column) together with the corresponding power spectrum in the Fourier domain (right 1024 column) are displayed. Density grid with added normally distributed noise at a signalto-noise ratio of 1.2 leads to loss of contrast at high resolution. Confidence maps 1025 1026 recapitulate these high-resolution features (arrows), showing that high-resolution 1027 signal is detected with high sensitivity. FDR thresholding at 1 % recovers a similar 1028 binary grid in comparison with 3σ -thresholding while minimizing noise contributions 1029 while minimizing detected noise (zoomed insets).

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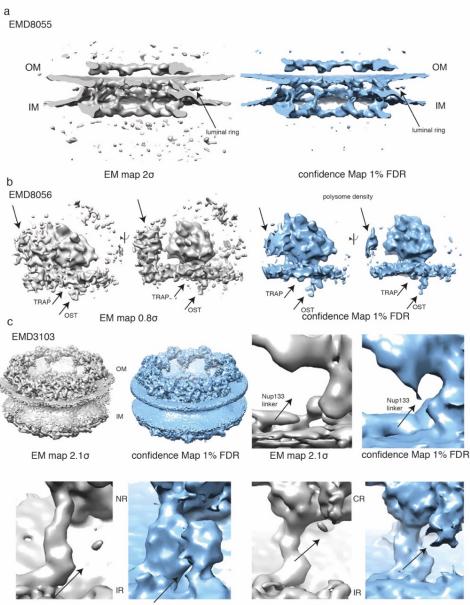
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1034 (a) Left. Confidence map with longitudinal section through TMV coat protein displayed 1035 indicating a-helical pitch of LR helix. Lower half shows the chosen contour at 1 % FDR in blue with 5.7 % of voxels detected. Right. Corresponding histogram of confidence 1036 map with signal separated above 0.99 PPV (1 % FDR). (b) Left. Same section as in 1037 (a) from cryo-EM density and the recommended threshold contoured at 1.2 σ in gray 1038 1039 with 3.7 % of voxels detected. Right. Corresponding histogram of cryo-EM density with 1040 thresholded values displayed in gray. (c) Isosurface rendered of thresholded confidence maps at 0.01 %, 1 % and 10 % FDR (left, center left, center right) shown 1041

1042 in blue and sharpened cryo-EM density with 1.2σ threshold (right) in gray from TMV 1043 (EMD2842). Shown are helix A86 – D77 (top), quarter cross section (bottom left) and 1044 side view (bottom right) of TMV map. (d) Detailed analysis of TMV density. Slice view 1045 through TMV rod with zoomed inset for inner and outer radii density (top). K53 side 1046 chain density (left) and molecular environment of R61 side chains (right) at 0.7, 1.2 σ 1047 threshold and at 1 % FDR confidence map. Figure 3



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EM map 2.1σ

confidence Map 1% FDR

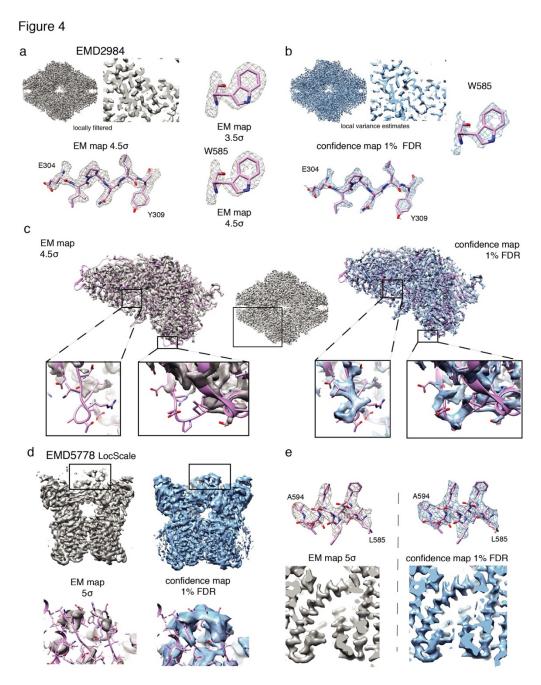
EM map 2.1σ

confidence Map 1% FDR

1049 Fig. 3. Confidence maps from subtomogram averages

1050 (a) Nuclear pore structure at 90 Å (EMD 8055) from 8 pore particles: cryo-EM map at 2.0σ threshold (left, gray) and confidence map at 1 % FDR threshold (right, blue). 1052 Note, the confidence map minimizes appearance of noise. (b) ER-associated

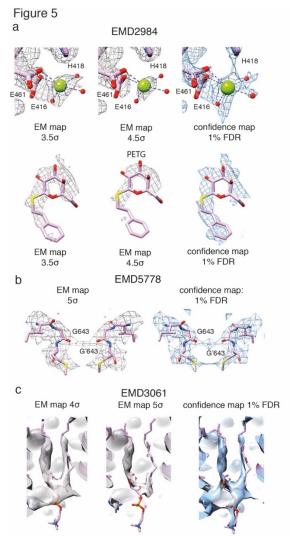
ribosome structure at 35 Å resolution (EMD 8056) in two side views at 0.8σ threshold 1053 1054 (left) and 1 % FDR confidence map (right). Note, in confidence maps weaker densities assigned to peripheral protein complexes TRAP and OST (arrows) can be easily 1055 visualized in the absence of noise. (c) Nuclear pore structure at 23 Å resolution (EMD 1056 1057 3103) comparing cryo-EM map at 2.1 σ threshold (left) and 1 % FDR confidence map 1058 (right). Comparison of map pairs for Nup133 linker density (top right), densities located between inner and nuclear ring (bottom left) and inner and cytoplasmic ring (bottom 1059 1060 right). In contrast to sharpened cryo-EM maps at 2.1σ threshold, confidence maps show consistently densities at the connections between the inner and outer rings at 1 1061 % FDR threshold (arrows). 1062



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1064 Fig. 4. Confidence maps benefit from local SNR adjustment based on local 1065 resolution.

1066 (a) β -galactosidase (EMD 2984) locally filtered cryo-EM map left (gray) displayed at 1067 4.5 σ threshold and (b) confidence map (blue) including signal-to-noise adjustment 1068 based on local resolution at 1% FDR threshold (right) in side view and cross section. 1069 High resolution features like E304 – E398 and holes in aromatic rings W585 in the 1070 3.5/4.5 σ -thresholded cryo-EM map (a) in comparison with the 1% FDR confidence 1071 map (b). (c) Comparison of density features from peripheral loop regions not covered 1072 by density in the locally filtered cryo-EM map (left) compared with the 1% FDR 1073 confidence map that shows densities for the respective loops. (d) TRPV1 (EMD5778) 1074 side view (top) with zoom-in to peripheral cytoplasmic domain density (bottom) 1075 comparing LocScale density displayed at 5 σ threshold (left) and 1 %FDR confidence 1076 map. (e) Detailed density stretch A594 – L585 (top) and transmembrane helix S5 1077 including S4-S5 linker (bottom) comparing LocScale density and 1 % FDR confidence 1078 map.

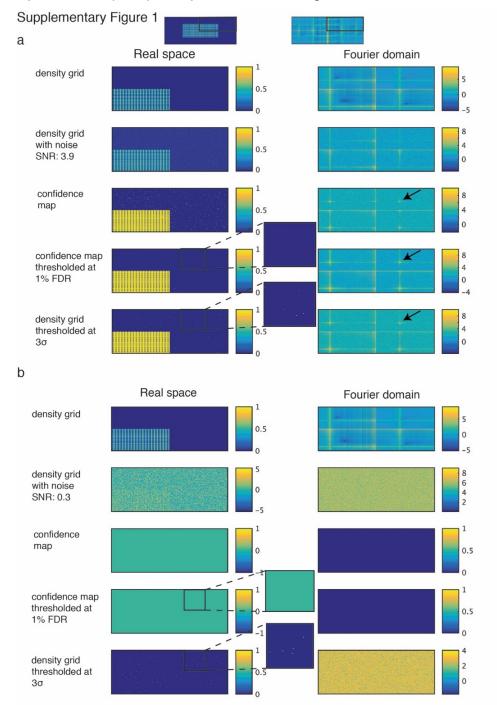


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1080 Fig. 5. Confidence maps confirm localization of non-protein components.

1081 (a) β-galactosidase (EMD 2984) with 3.5/4.5 *σ*-thresholded cryo-EM map (left, center, 1082 gray) and 1 % FDR thresholded confidence map (right, blue): Mg^{2+} ion coordinated by 1083 E461, E416, H418 and 3 H₂O molecules (top). Density of bound PETG ligand the 1084 3.5/4.5 *σ*-thresholded cryo-EM map and in the 1% FDR confidence map (bottom). (b) 1085 TRPV1 channel (EMD 5778) with 5 *σ*-thresholded cryo-EM map (left) and 1 % FDR 1086 thresholded confidence map (right): selectivity filter formed by carbonyls of symmetry-

- 1087 related G643 residues. The presence of a putative ion is supported by the confidence
- 1088 map. (c) γ -secretase (EMD 3061) with 4σ and 5σ -thresholded cryo-EM map (left) and
- 1089 1 % FDR thresholded confidence map (right): The confidence map reveals density for
- 1090 both acyl chains of phosphatidyl choline at a single threshold.



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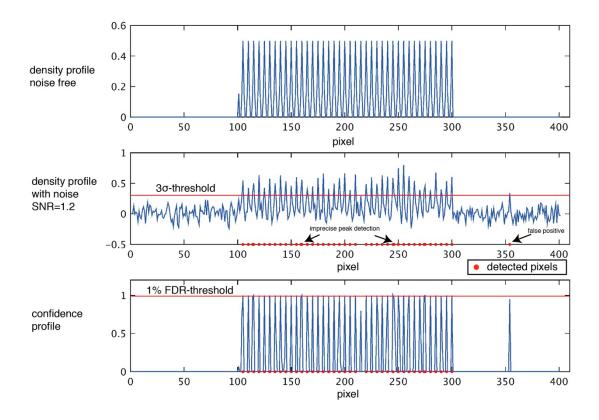
1092 Fig. S1. Comparison of σ and FDR thresholding of simulated density grids with

1093 varying signal-to-noise ratios.

1094 Thresholding with simulated density grids at signal-to-noise ratios and variance of (a)

1095 3.9 (0.01) and (b) 0.3 (1.33), respectively. The same simulations as in Fig. 1c are

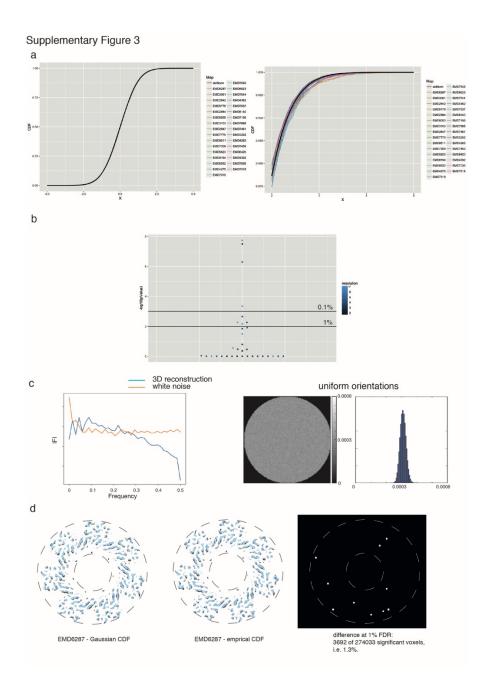
- 1096 repeated with lower and higher variances of the background noise. At low signal-to-
- noise ratio, the 1% FDR thresholding is devoid of false positives whereas conventional
- 1098 3*o*-thresholding approach yields many false positive pixels (zoomed inset).
 - Supplementary Figure 2



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1100 Fig. S2. Effect of σ and FDR thresholding on 1D density profiles.

1101 One-dimensional stacked plots of grid density with noise-free original (top), at signal-1102 to-noise ratio of 1.2 (center) and confidence map (bottom). The noisy density grid is 1103 thresholded at 3σ and the confidence map is thresholded at 1 % FDR. Conventional 1104 3σ -thresholding yields higher rates of false positives and some imprecise peak 1105 positions (arrows).

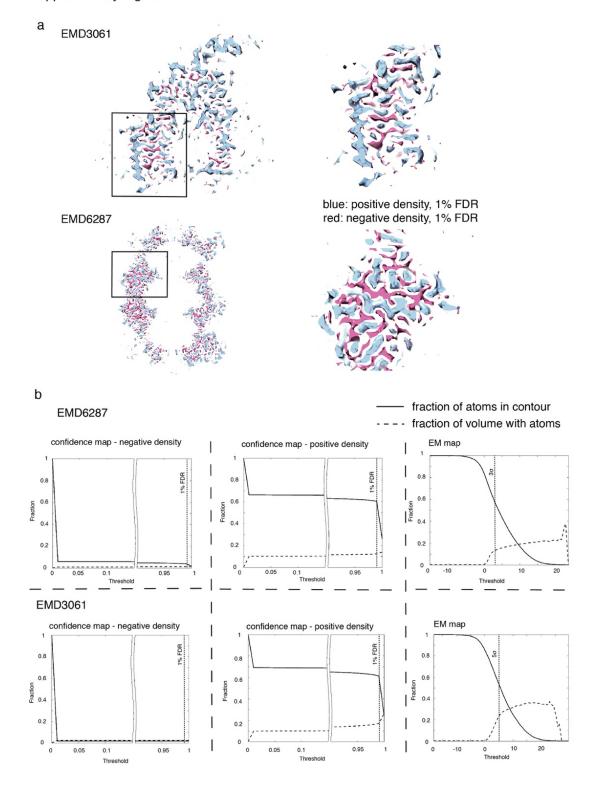


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1108 Fig. S3. Analysis of normality of cryo-EM densities.

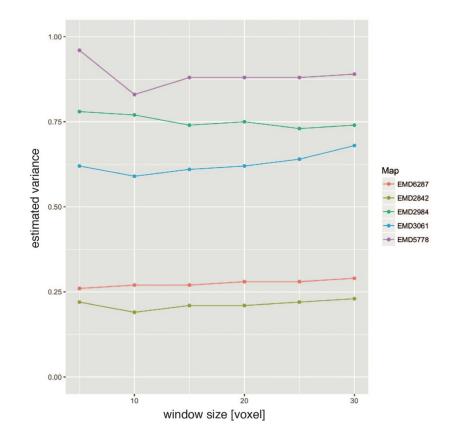
1109 (a) Left. Overlay of 32 cumulative density functions (CDF) derived from the above 1110 EMDB entries with ideal Gaussian CDF in black. Right. Zoomed inset to better 1111 highlight small differences. (b) 32 map entries are assessed with respect to normality 1112 according to the Anderson-Darling test, significance thresholds are displayed 1.0 and 1113 0.1 % respectively. (c) Left. Rotational power spectrum of a 3D reconstruction of white 1114 noise images in comparison with pure white noise spectrum. Right. Slice through 3D volume of variances estimated from 900 independent reconstructions from Gaussian 1115 white noise images with similar uniform orientations together with a histogram of the 1116

- 1117 estimated variances, showing that background noise can be assumed uniform over
- 1118 the central sphere in the reconstructed volume. (d) Cross-sectional view of confidence
- 1119 maps generated of EMD6287 using Gaussian and empirical CDF. Difference map
- 1120 between 1 % FDR binarized confidence maps in the respective image slice. Supplementary Figure 4



1122 Fig. S4. Analysis of positive and negative densities using confidence maps.

- 1123 (a) Overlay of 1% FDR positive (blue) and negative (red) confidence maps from
- 1124 original and inverted densities of EMD3061 (top) and EMD6287 (bottom) respectively.
- 1125 (b) Comparison of detected signal with corresponding atomic models by determining
- 1126 the fraction of overlap of atoms with volume and fraction of volume with atoms as a
- 1127 function of threshold for negative (left), positive (center) confidence maps and cryo-
- 1128 EM maps (right), respectively. Supplementary Figure 5

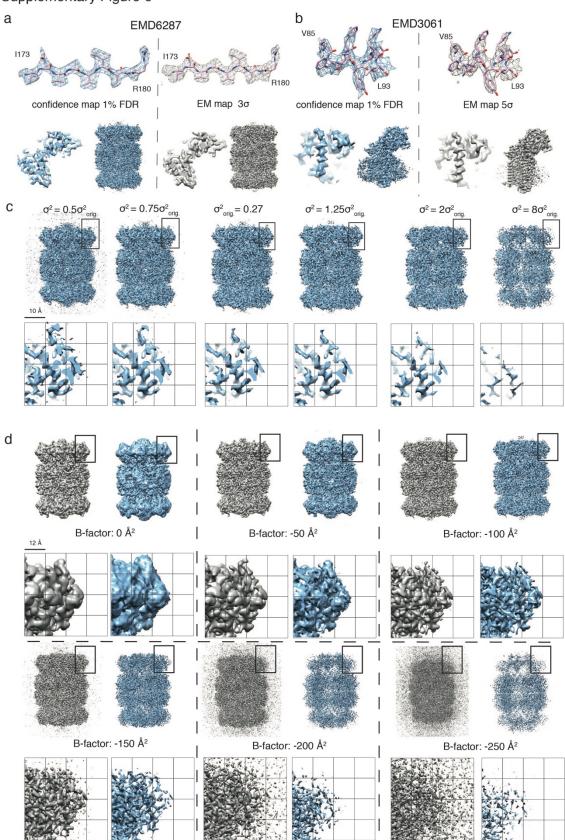


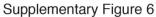
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1130 Fig. S5. Effect of window size on estimated variance.

1131 Estimated variance is stable with increasing window size from 5 to 30 voxels for a

- 1132 series of EMD entries.
- 1133







confidence maps shown at 1% FDR and EM maps shown at 3σ

1135 Fig. S6. Confidence maps and effect of incorrect noise estimation.

1136 (a) 20S proteasome map (EMD 6287) comparison of 1% FDR density (left) and 3*o*thresholded map (right). Shown are molecular details from 1173 - R180 (top), slice 1137 view (bottom left) and side view (bottom right) of density. (b) v-secretase map (EMD 1138 1139 3061) comparison of 1% FDR confidence map and 5σ -thresholded map. (c) Six 1140 confidence maps of 20S proteasome (EMD-8267) including magnified inset based on incorrect variance estimation: 1st and 2nd left noise is underestimated by 0.5 and 0.75 1141 times the variance (σ^2). In comparison with the correctly estimated noise (3rd), they 1142 show excessive noise features declared as signal at 1 % FDR. When noise is 1143 overestimated, which is more likely for cryo-EM maps, confidence maps are quite 1144 insensitive to changes in map appearance. For multiples like 1.25 σ^2 and $2\sigma^2$ no 1145 apparent density changes become visible (4th and 5th) unless strong overestimation 1146 like $8\sigma^2$ (6th) leads to disappearance of map features at a 1 % FDR threshold. (d) When 1147 applying a series of B-factors to the 3D reconstruction of the 20S proteasome map. 1148 1149 we see that with higher B-factors, sharpened EM densities become dominated by noise whereas corresponding confidence maps displayed at 1 % FDR show 1150 1151 disappearance of significant features thereby avoids over-interpreting noise features. 1152

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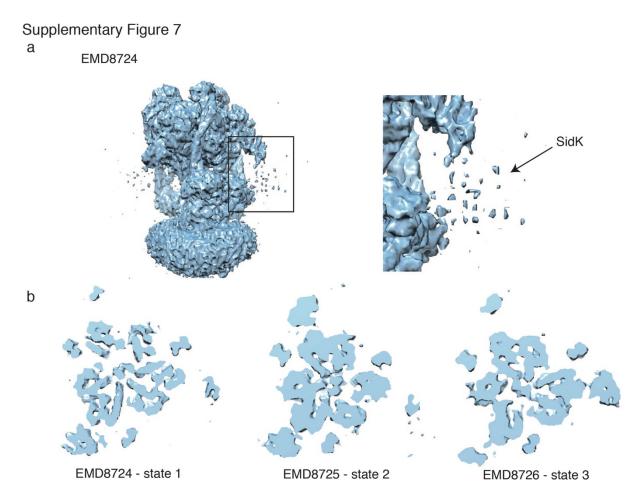
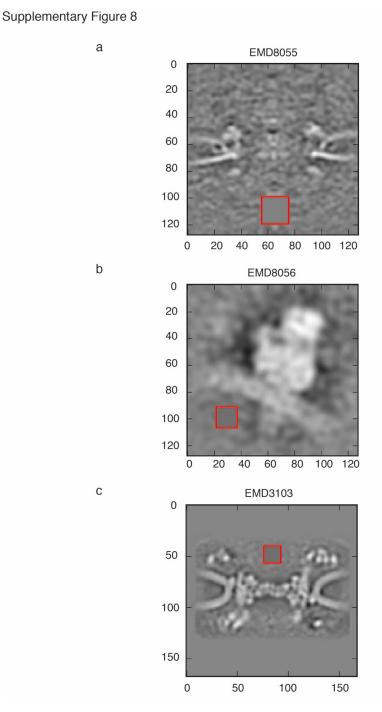


Fig. S7. Confidence maps of compositionally and conformationally heterogeneous complexes.

(a) Confidence map of yeast V-ATPase with Legionella pneumophila effector SidK
(EMD8724) at 1% FDR (left) together with a zoom on the flexible domains of SidK
(right). The confidence map shows significant density for the flexible domains,
however, not as continuous density. (b) Slices through confidence maps of 3D
classified cryo-EM maps. Different rotational states can be resolved in the confidence
maps.

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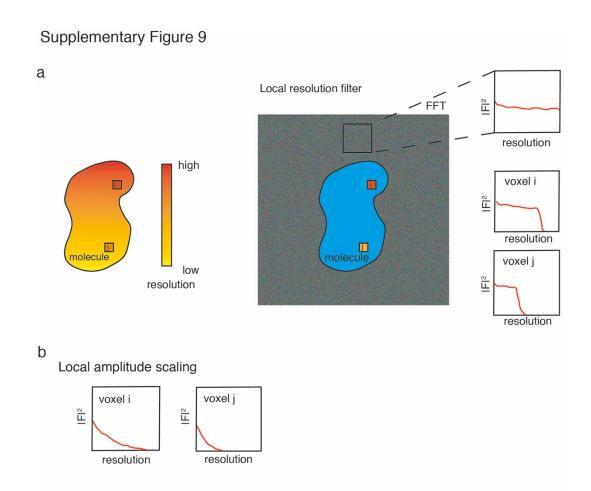
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1173 Fig. S8. Noise estimation in subtomogram averages.

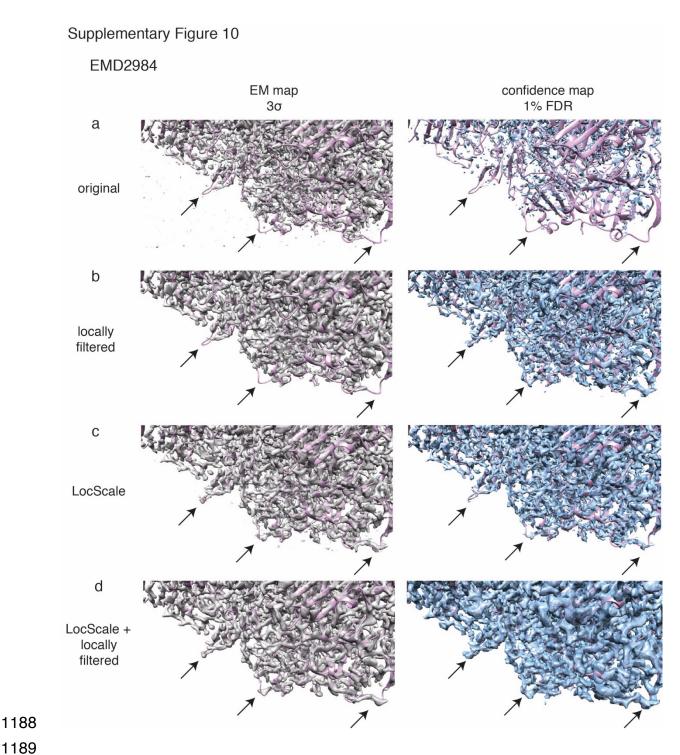
Gray-scale density slices with red windows for the voxel region used for variance
estimation: (a) EMD 8055: nuclear pore from HeLa cells by FIB-SEM, (b) EMD 8056:
ER-associated ribosomes, (c) EMD3103: 23 Å resolution nuclear pore subtomogram
average.



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1180 Fig. S9. Variance adjustment based on local resolution and local amplitude 1181 profile.

(a) Adjusting the local signal-to-noise ratio based on local resolution measurements:
for each voxel, the background windows are filtered according to the local resolution
at the respective voxels in order to estimate the noise levels of each voxel in the locally
filtered map. (b) In analogy, local sharpening is applied to background noise in order
to estimate resulting local noise distributions.



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1190 Fig. S10. Effect of local variance adjustments on confidence maps

1191 β -galactosidase (EMD 2984) cryo-EM map at 3.0 σ threshold (left, gray) and 1 % FDR 1192 confidence map based on different post-processing methods (right, blue). Global 1193 sharpening with uniform filtering, local filtering based on local resolution 1194 measurements, local sharpening and the combination of local sharpening with local 1195 filtering were compared. Confidence maps were generated with local noise estimate

- 1196 based on local resolution measurement, locally scaled window from a model reference
- 1197 structure and the combination of both, which in this case shows the best preservation
- 1198 of molecular density with respect to confidence.