# pymfinder: a tool for the motif analysis of binary and quantitative complex networks

Bernat Bramon Mora<sup>1,\*</sup>, Alyssa R. Cirtwill<sup>2</sup> and Daniel B. Stouffer<sup>1</sup>

<sup>1</sup>Centre for Integrative Ecology, School of Biological Sciences, University of Canterbury, Christchurch, New Zealand

 $^2$  Department of Physics, Chemistry, and Biology, Linköping University, Linköping, Sweden.  $^* bernat. bramon@gmail.com$ 

## $_{\scriptscriptstyle 1}$ Abstract

- <sup>2</sup> We developed *pymfinder*, a new software to analyze multiple aspects of the so-called network
- motifs—distinct n-node patterns of interaction—for any directed, undirected, unipartite or
- 4 bipartite network. Unlike existing software for the study of network motifs, pymfinder
- 5 allows the computation of node- and link-specific motif profiles as well as the analysis of
- 6 weighted motifs. Beyond the overall characterization of networks, the tools presented in this
- work therefore allow for the comparison of the "roles" of either nodes or links of a network.
- 8 Examples include the study of the roles of different species and/or their trophic/mutualistic
- 9 interactions in ecological networks or the roles of specific proteins and/or their activa-
- o tion/inhibition relationships in protein-protein interaction networks. Here, we show how
- 11 to apply the main tools from pymfinder using a predator-prey interaction network from a
- marine food web. pymfinder is open source software that can be freely and anonymously
- downloaded from https://github.com/stoufferlab/pymfinder, distributed under the
- 14 MIT License (2018).

## Introduction

The use of network theory has proven insightful in multiple fields, from the study of the spread of disease epidemics (1) to the characterization of neuronal networks (2). In ecology, this approach has been crucial to understanding the ways different species interact with each other, and the network perspective has justly become a central topic in community ecology (3). Over recent years, multiple methods for studying the topology of ecological networks have been successfully developed. Examples include models to generate realistic 21 ecological communities (4) or tools for studying different network metrics such as compartmentalization (5), nestedness (6) or intervality (7). Following these advances, one of the most versatile ways to understand the structure of complex ecological networks is via the so-called network motifs—i.e. the analysis of small subgraphs representing the distinct patterns of interaction involving any set of n species. These subgraphs have been referred to as the 'building blocks' of complex networks (8). 27 The study of network motifs has been applied to multiple ecological systems over the 28 recent years, including those composed of trophic (9) and mutualistic interactions (10). Non-ecological examples include in protein-protein interaction networks (11) and transcriptional regulation networks (12). There are typically two main approaches that are taken involving network motifs. First, counting the number of appearances of any given n-node pattern of interactions provides an overall perspective of the structure of a network. This has been done in different ecological studies, including the characterization of food webs (13), plant-pollination (14) and host-parasitoid networks (15). Second, other ecological studies have focused on the role of different species (16) and interactions (17), defining their position within the network based on which network motifs they form a part of. Following this work on network motifs, multiple tools for the counting of network motifs have been developed over the last decades (18, 19, 20). Most of the methodological work has focused on providing tools to efficiently quantify the overall structure of directed 40 and undirected unipartite networks—i.e. graphs consisting of one set of interacting nodes. 41 Unfortunately, to our knowledge, we are still lacking general-purpose software to also analyze bipartite networks—i.e. graphs consisting of two interacting sets of non-overlapping nodes—as well as to quantify the node- and link-specific motif profiles in both unipartite

and bipartite networks. In addition, there is no tool to date that allows the user to include information regarding the interaction strengths of a network within the analysis of motifs. 46 In response, we present pymfinder, software for motif analysis of network structure plus of the nodes and links of any type of network—i.e. directed/undirected, bipartite/unipartite, and weighted/binary networks. pymfinder is an open-source and versatile tool for the study of network motifs and the 50 result of long-standing research involving the study of ecological networks. For example, pymfinder was used to shed light on the ecological mechanisms underlying food-web structure (9), which, together with Bascompte 2005 (21) and Camacho et. al. (22), was one of the first studies to put network motifs into a purely ecological context. Building on these foundational studies, network motifs and pymfinder were shown to provide a useful way to characterize species' roles, showing them to be evolutionary conserved across communities (16). Similarly, the roles of links involving parasite species were characterized through the study of network motifs, generating an understanding of how different types of feeding links are distributed within a food web (17). The same software has also been used to study bipartite networks. For instance, a study on host-parasitoid networks showed how species' roles seem to be conserved over spatial scales as well as consistent over time 61 (15). Perhaps more importantly, the software presented here has also been a central piece of very recent research. For example, the tools in pymfinder were used to relate species' roles to multiple ecological traits in five marine food webs, showing that feeding environment is particularly strongly related to such roles (23). Likewise, the variability of species' roles in plant-pollinator communities in the Arctic has recently shown to be related to the 66 variability in community composition (24). Finally, the description of species' roles has 67 also been key to comparing entire networks by means of aligning species to each other, resulting in the identification of common backbones shared across food webs form different ecosystems (25). Overall, the tools included in pymfinder are and have been instrumental 70 to the development of a diverse set of projects over the years, and we believe that they 71 have the potential to be valuable for many others. This article describes the main structure of pymfinder and showcases some of its principal applications using a detailed ecological

dataset as the backdrop.

# 75 Design and implementation

## 76 General description

77 pymfinder is a Python library that combines Python methods for network-motif analysis.

<sup>78</sup> Some of the engine underneath is a modified version of *mfinder*—a software tool for network-

motif detection developed by Kashtan et. al. (8, 18). Originally, mfinder was written in C

and made available solely as an executable, and we use it within *pymfinder* for its underlying

81 efficiency. The mfinder code has been both included and modified here with the explicit

consent of Nadav Kashtan, the author of mfinder 1.2.

As input, pymfinder accepts any type of network. That is, the analyses can be performed 83 for both unipartite and bipartite networks. The format in which the networks are passed 84 to the different functions of the package is either as text files, Python arrays or pymfinderobjects. Text files must describe the set of links comprising the networks, where each link appears as a separate line in the files. For example, a given line "A B w" would describe 87 a single link  $A \to B$  between nodes A and B, where w represents the strength or weight 88 associated to such link (see Appendix). Similarly, Python arrays need to represent the list 89 of interactions forming the networks. Notice that the direction of the links is important. Therefore, in bipartite networks, nodes of each group need to consistently be placed on 91 the same side of the interactions—e.g. in a plant-pollinator networks the direction of the 92 interactions in the input must all go from a plant to a pollinator (or vice-versa). Importantly, 93 undirected networks can also be analyzed by pymfinder; however, any links between two 94 nodes A and B in such networks need to be characterized by the two parallel links A B and A. The output of pymfinder, is a high-level data type ('class') that contains different descriptors of the motif composition of the network under study (see Appendix).

# 98 Structure of the package

At their core, all of the analyses performed by *pymfinder* are based around the identification of all the different *n*-node patterns of interaction found within a given network. To do this, *pymfinder* will always start by enumerating the unique motifs/subgraphs that make up the overall structure of the network under study. This analysis can be performed for multiple motif sizes. This is especially important for bipartite networks, where three-node motifs are minimally informative and one needs to explore bigger motifs (15). Notice, however, that increasing the number of nodes can be computationally challenging for unipartite networks since the number of unique motifs quickly increases with their size—i.e. there are 13 unique three-node motifs, 199 unique four-node motifs and 9364 unique five-node motifs.

For the sake of simplicity, we will focus most of the description of the methods presented here on the analysis of three-node network motifs. For any given network, this analysis is a three-step process. First, pymfinder loops through all the rows i of the adjacency matrix A associated with the network. For each non-zero element  $a_{ij}$  found in row  $A_i$ , it then searches for any connected element  $a_{jk} = 1$ ,  $a_{kj} = 1$ ,  $a_{ik} = 1$ , and/or  $a_{ki} = 1$ , revealing the existence of any motif comprised of the nodes i, j, and k. If i, j and k define a motif and this motif has not already been identified, the corresponding motif and the position of each node within the motif is recorded.

Based on this initial motif enumeration, *pymfinder* can perform three subsequent analyses:

(i) the analysis of the overall network structure, (ii) the nodes and links' participation in

the different motifs, and (iii) the nodes and links' role in each of the motifs.

## 119 Motif structure

The most basic application of pymfinder is the analysis of the overall motif structure of a 120 given network. In particular, such analysis generates a description of the distribution of 121 distinct n-node patterns of interaction found within the network (up to 8-node motifs). The 122 application also includes the possibility of estimating the null motif composition expected 123 for such network (see Appendix). To generate this null composition, pymfinder uses an 124 MCMC algorithm to perform a randomization of the network while preserving the in-125 and out-degree of the nodes and each node's number of single and double links (26, 27). 126 Comparing the observed motif frequency to the random expectation, the application can 127 be used to determine which interaction patterns are over- or under-represented relative to 128 this null model (9). To do so, pymfinder calculates the mean and standard deviation of the 129 null expectation as well as the z-scores for its comparison with the actual observations. 130

An additional feature of pymfinder is the possibility of incorporating information regard-131 ing the link strength into the analysis of the motif structure. This is notable in particular 132 since there is no software available to explore the way the interaction strengths are dis-133 tributed within networks across motifs. To do so, pymfinder will account for each motif within a given weighted network as a function of the strength of the links forming them (Fig. 135 1). Note that the algorithm allows the user to choose how the weight of a motif is defined. 136 Specifically, given a motif formed by the set of links with strengths  $\{l\} = \{l_1, l_2, ..., l_L\},\$ 137 pymfinder will calculate the weight of such motif as  $f(\{l\})$ , where f is the function defined 138 by the user. By default, pymfinder uses the arithmetic mean as the function f. Similar 139 to unweighted networks, analysis of the motif structure of a weighted network returns the 140 average and standard deviation of the weight of each motif, as well as the median and the 141 first and third quartiles. 142

### 143 Motif participation

The study of network motifs can also be used as a way to classify nodes based on which 144 patterns of interactions they are part of. For any given network, this application deter-145 mines the frequency of appearance of every node across each of the different motifs (Fig. 146 1b), defining their participation across these distinct patterns of interactions. This a useful 147 perspective for motif analysis because it provides a node-based description of the networks 148 that can be used to understand the nature of specific nodes (e.g. different species in eco-149 logical networks or different proteins in protein-protein interaction networks) as well as 150 decomposes the overall structure of the network at a finer resolution (21). Similarly, the 151 same analysis can also be performed for the links forming the network. That is, pymfinder 152 can quantify the frequency with which every link forms part of each distinct motif. As for 153 the analysis of the overall structure of the networks, the motif participation of both nodes and links can also be calculated for any given motif size up to 8 nodes for weighted and 155 unweighted networks. Again, pymfinder will account for each motif within a given weighted 156 network as a function of the strength of the links forming it (Fig. 1), and the algorithm 157 allows the user to choose this function just as described above for motif structure. 158

## 9 Motif-role profiles

Within any given motif, nodes can play multiple roles. For example, in the two-node motif 160  $A \to B$ , there are two distinct positions A and B, which define two different roles—e.g. a 161 predator and a prey in a food web. In contrast, for the two-node motif  $A \leftrightarrow B$ , A and B 162 occupy indistinguishable positions; therefore, there is a single distinct role. The same idea 163 can be extended to all n-node motifs. For example, there are 30 distinct node positions and 164 24 distinct link positions across the 13 unique three-nodes motifs. These distinct positions 165 within the different motifs are important because the number of times that a node appears 166 in each of them can be used as a way to define its structural role in a community (16). 167 That is, we can characterize a node's structural role based on the number of times that it 168 occupies each distinct position of the n-node motifs. pymfinder provides a way to determine 160 such n-node motif-role profiles for both the nodes (Fig. 1c) and the links (Fig. 1d) of a 170 given network. Notice, however, that this function can only be run for two- and three-nodes 171 motifs in unipartite networks, and two- to six-nodes motifs in bipartite networks. 172

The analysis of node and link motif-role profiles can also incorporate information regarding the strengths of interactions between nodes. As before, consider a motif m formed by
the set of nodes  $\{i\}$  and the set of links with strengths  $\{l\}$ . For any node j in  $\{i\}$ , pymfindercalculates the contribution  $c_{jm}$  of motif m to any of the positions of j's motif-role profile
as:

$$c_{jm} = \frac{f(\{l_j\})}{\sum_i f(\{l_i\})} f(\{l\})$$
 (1)

where  $\{l_i\}$  is the set of strengths of all links in m involving node i, and f is a function defined by the user. By default, pymfinder again uses the arithmetic mean as f for weighted motifrole profiles. Notice that the contribution  $c_{im} = 1$  when ignoring the weights, or f is the arithmetic mean and all weights are equal to the motif size. When analyzing the motif-role profile of a link k forming such motif, the contribution  $c_{km}$  is assumed to be exactly equal to its link strength  $l_k$ .

#### 84 Basic tests

To ensure the reliable functioning of pymfinder, we included a set of basic tests in the 185 package. All these basic tests are based around the idea of analyzing the structure of 186 artificial networks containing only a single motif of each type for a given motif size—up 187 to five-node motifs for bipartite networks and three-node motifs for unipartite networks. 188 In addition, those networks are also set up so that any given node or link is only involved 180 in a single motif and role. Using these single-motif networks, we tested the functions of 190 pymfinder by ensuring that the analysis of such artificial networks does not result in the 191 misrepresentation of any motif, node, link or role. 192

# $_{\scriptscriptstyle{193}}$ Results/Discussion

The tools provided by *pymfinder* can be used in a large variety of systems and do not depend on the nature or providence of the networks. To illustrate the capabilities and potential of the software, we outline the study of a food web from a marine ecosystem as a representative study system (28). This specific network describes the predator-prey interactions between approximately 250 of the species found across an extensive area of the Caribbean Sea.

We first analyzed the overall three-species motif structure of the network and compared 200 it to the random expectation (Fig. 2b). For this example, we used the z-score values to 201 draw this comparison, which assume normality of the motif distribution. Notice, however, 202 that pymfinder also returns the mean number of motif counts in the randomized networks, 203 which allows for other types of statistical analyses. We found that the observed motif 204 distribution is generally significantly different from the random expectation, showing either 205 over- or under-representation relative to the results of the null model used here. This is evidence of a non-random organization of ecological communities (21, 29), which speaks to 207 the eco-evolutionary mechanisms shaping the ways in which different species interact with 208 each other. We then studied the distribution of link weights across motifs to test whether or 209 not different motifs are generally made of different interaction strengths. For this particular 210 example, we log-transformed the link weights to be approximately normally distributed as 211

well as scaled them so that the weakest and strongest links had a weight of zero and one,
respectively. In general, we found that interaction strengths are distributed in a similar
manner across the different motifs of the network under study (Fig. 2c). Notice that these
results are subject to the logarithmic transformation applied to the weight data, which is
generally very skewed (28).

Following the analysis of the overall motif structure, we examined the motif participation 217 of the different nodes and links that make up this food web. We found that some nodes 218 (e.g., sea cucumbers and algae) share almost identical motif-participation profiles while 219 others (e.g., filefish and sea cucumbers) have very distinct profiles (Fig. 3a). This shows 220 how motifs can be a valuable and insightful way to classify and compare the species across 221 communities. Perhaps more importantly, we observed how the information regarding the 222 interaction strengths forming the motifs changed those motif-participation profiles (Fig. 223 **3b**). Therefore, adding interaction strengths allowed us to distinguish between the roles of species with similar unweighted profiles. This is important because it suggests that, 225 from a node-specific perspective, interaction strengths are not equally distributed across 226 motifs. The uneven distribution of interaction strengths has important implications for the 227 relationship between network structure and species-interaction strengths and the stability 228 of food webs(30, 31). We also looked at the motif-participation profiles of the links (Fig. 229 3c). We found that those profiles could also be an indicator of the observed differences on 230 the way interaction strengths are distributed across motifs, as suggested by previous work 231 (17).232

Finally, we studied the motif-role profiles of the species of the marine network. This 233 analysis is similar to the motif participation analysis of nodes and links; however, it provides a finer resolution to the role that different species or links might play in the community. 235 Using the proportion of times that the different species are in each of the 30 unique positions 236 of the three-species network motifs, we performed an analysis of multivariate homogeneity of 237 238 group dispersions to compare the roles of the species in the network (32). To do this, we first calculated the euclidean distance between the roles of every pair of species in the network, 239 generating a dissimilarity matrix of all species. We then performed a basic clustering 240 analysis of the species-role dissimilarity matrix to find the most distinct groups of roles (Fig. 241

4). Finally, we used the function betadisper from the R package vegan (33) to perform the Principal Coordinates Analysis (PCoA) of the data. We found four characteristic groups of species presenting very distinct motif-role profiles. Notice that the same analysis can also be done for the motif-role profile of every link in the network. This is useful because it shows the diversity of structural roles in this community and underlines how those profiles could be used to compare species, links or networks within and across ecosystems, environments and biomes (34, 25).

# Availability and Future Directions

pymfinder is open source software that can be freely and anonymously downloaded from 250 https://github.com/stoufferlab/pymfinder. The documentation of the package is at-251 tached as supplementary material and the data used to test the software can be found 252 within the github repository. pymfinder has been tested to run on any platform that 253 supports Python. pymfinder will require you to have the Python modules Numpy and Se-254 tuptools installed in your machine. Data used to present the software has been previously 255 published by Bascompte et. al. (28). We are currently working on additional software that 256 uses the weighted motif-role profiles of nodes to efficiently align bipartite networks multiple 257 times.

## 259 References

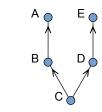
- $^{260}$  [1] Mark EJ Newman. Spread of epidemic disease on networks. *Physical review E*,  $^{261}$   $^{66}$  (1):016128, 2002.
- [2] Olaf Sporns. Network analysis, complexity, and brain function. *Complexity*, 8(1):56–60, 2002.
- [3] Jordi Bascompte and Pedro Jordano. Mutualistic networks. Princeton University Press,
   2013.
- [4] Richard J Williams and Neo D Martinez. Simple rules yield complex food webs. Nature,
   404(6774):180–183, 2000.

- [5] Roger Guimera, Marta Sales-Pardo, and Luís A Nunes Amaral. Module identification in bipartite and directed networks. *Physical Review E*, 76(3):036102, 2007.
- [6] Jordi Bascompte, Pedro Jordano, Carlos J Melián, and Jens M Olesen. The nested
   assembly of plant-animal mutualistic networks. Proceedings of the National Academy
   of Sciences, 100(16):9383-9387, 2003.
- [7] Daniel B Stouffer, Juan Camacho, and Luís A Nunes Amaral. A robust measure of
   food web intervality. Proceedings of the National Academy of Sciences, 103(50):19015–
   19020, 2006.
- [8] Ron Milo, Shai Shen-Orr, Shalev Itzkovitz, Nadav Kashtan, Dmitri Chklovskii, and Uri Alon. Network motifs: simple building blocks of complex networks. *Science*, 298(5594):824–827, 2002.
- [9] Daniel B Stouffer, Juan Camacho, Wenxin Jiang, and Luís A Nunes Amaral. Evidence
   for the existence of a robust pattern of prey selection in food webs. *Proc R Soc Biol Sci*, 274(1621):1931–1940, 2007.
- [10] Carsten F Dormann, Jochen Fründ, Nico Blüthgen, and Bernd Gruber. Indices, graphs and null models: analyzing bipartite ecological networks. *The Open Ecology Journal*, 284 2(1), 2009.
- [11] Esti Yeger-Lotem, Shmuel Sattath, Nadav Kashtan, Shalev Itzkovitz, Ron Milo, Ron Y
   Pinter, Uri Alon, and Hanah Margalit. Network motifs in integrated cellular networks
   of transcription-regulation and protein-protein interaction. *Proceedings of the National* Academy of Sciences of the United States of America, 101(16):5934-5939, 2004.
- <sup>289</sup> [12] Shai S Shen-Orr, Ron Milo, Shmoolik Mangan, and Uri Alon. Network motifs in the transcriptional regulation network of escherichia coli. *Nature genetics*, 31(1):64, 2002.
- <sup>291</sup> [13] Janis Klaise and Samuel Johnson. The origin of motif families in food webs. *Scientific*<sup>292</sup> Reports, 7(1):16197, 2017.
- [14] María C Rodríguez-Rodríguez, Pedro Jordano, and Alfredo Valido. Functional consequences of plant-animal interactions along the mutualism-antagonism gradient. *Ecology*, 98(5):1266–1276, 2017.

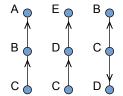
- [15] Nick J Baker, Riikka Kaartinen, Tomas Roslin, and Daniel B Stouffer. Species' roles in
   food webs show fidelity across a highly variable oak forest. *Ecography*, 38(2):130–139,
   2015.
- [16] Daniel B Stouffer, Marta Sales-Pardo, M Irmak Sirer, and Jordi Bascompte. Evolutionary conservation of species' roles in food webs. Science, 335(6075):1489–1492,
   2012.
- Journal of Animal Ecology, 84(3):734—744, 2015.
- [18] Nadav Kashtan, Shalev Itzkovitz, Ron Milo, and Uri Alon. Efficient sampling algorithm
   for estimating subgraph concentrations and detecting network motifs. *Bioinformatics*,
   20(11):1746–1758, 2004.
- [19] Sebastian Wernicke and Florian Rasche. Fanmod: a tool for fast network motif detection. *Bioinformatics*, 22(9):1152–1153, 2006.
- <sup>310</sup> [20] Gabor Csardi and Tamas Nepusz. The igraph software package for complex network research. *InterJournal, Complex Systems*, 1695(5):1–9, 2006.
- [21] Jordi Bascompte and Carlos J Melián. Simple trophic modules for complex food webs.
   Ecology, 86(11):2868–2873, 2005.
- <sup>314</sup> [22] Juan Camacho, Daniel B Stouffer, and Luís A Nunes Amaral. Quantitative analysis of the local structure of food webs. *Journal of Theoretical Biology*, 246(2):260–268, 2007.
- <sup>316</sup> [23] Alyssa R Cirtwill and Anna Eklöf. Feeding environment and other traits shape species' roles in marine food webs. *Ecology letters*, 21(6):875–884, 2018.
- [24] Alyssa R Cirtwill, Tomas Roslin, Claus Rasmussen, Jens Mogens Olesen, and Daniel B
   Stouffer. Between-year changes in community composition shape species' roles in an
   arctic plant-pollinator network. Oikos, 2018.
- <sup>321</sup> [25] Bernat Bramon Mora, Dominique Gravel, Gilarranz J. Luis, Poisot Thimothee, and
  <sup>322</sup> Daniel B. Stouffer. Identifying a common backbone of interactions underlying food
  <sup>323</sup> webs from different ecosystems. *Nature Communications*, 2018. (accepted).

- <sup>324</sup> [26] Ron Milo, Nadav Kashtan, Shalev Itzkovitz, Mark EJ Newman, and Uri Alon. On the uniform generation of random graphs with prescribed degree sequences. arXiv preprint cond-mat/0312028, 2003.
- <sup>327</sup> [27] James G Sanderson, Michael P Moulton, and Ralph G Selfridge. Null matrices and the analysis of species co-occurrences. *Oecologia*, 116(1-2):275–283, 1998.
- <sup>329</sup> [28] Jordi Bascompte, Carlos J Melián, and Enric Sala. Interaction strength combinations <sup>330</sup> and the overfishing of a marine food web. *Proceedings of the National Academy of* <sup>331</sup> Sciences of the United States of America, 102(15):5443–5447, 2005.
- <sup>332</sup> [29] Jonathan J Borrelli. Selection against instability: stable subgraphs are most frequent <sup>333</sup> in empirical food webs. *Oikos*, 124(12):1583–1588, 2015.
- 334 [30] Anje-Margriet Neutel, Johan AP Heesterbeek, and Peter C de Ruiter. Stability in real 335 food webs: weak links in long loops. *Science*, 296(5570):1120–1123, 2002.
- [31] Mark Emmerson and Jon M Yearsley. Weak interactions, omnivory and emergent
   food-web properties. P Roy Soc Lond B Bio, 271(1537):397–405, 2004.
- [32] Marti J Anderson. Distance-based tests for homogeneity of multivariate dispersions.
   Biometrics, 62(1):245-253, 2006.
- [33] Jari Oksanen, F. Guillaume Blanchet, Michael Friendly, Roeland Kindt, Pierre Legendre, Dan McGlinn, Peter R. Minchin, R. B. O'Hara, Gavin L. Simpson, Peter Solymos,
   M. Henry H. Stevens, Eduard Szoecs, and Helene Wagner. vegan: Community Ecology
   Package, 2017. R package version 2.4-4.
- [34] Timothée Poisot, Elsa Canard, David Mouillot, Nicolas Mouquet, and Dominique
   Gravel. The dissimilarity of species interaction networks. *Ecol Lett*, 15(12):1353–1361,
   2012.





#### b) Network motifs



#### c) Node roles

				~	<b>7</b> 0
	0_		<b>&gt;</b> ○	•	<b>→</b>
Α	0	0	1	0	0
В	0	1	0	0	1
С	2	0	0	1	0
D	0	1	0	0	1
Е	0	0	1	0	0

#### d) Link roles

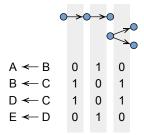


Figure 1: Main components of network-motif analysis. (a) A simple network that could represent a simple ecological community—where nodes would characterize species and the arrows would indicate the interactions between them—or a protein-protein interaction network—where nodes would represent different proteins and the arrows indicate either activation or inhibition. (b) All three-node motifs found in the network from (a); from this classification, we can compute the overall network structure and the number appearances of every node in each motif. (c) The characterization of every node's motif-role profile. This characterization is based on the number of appearances of every link's motif-role profile, which is based on the number of appearances of every link in each of the unique motif link-positions. Notice that we excluded any motif or role that was not represented in the network.

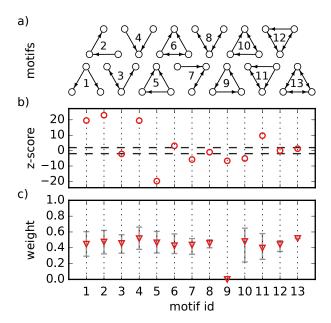


Figure 2: Analysis of the overall motif structure of the marine food web under study. The first panel (a) shows all the possible three-species motif structures. In this case, any arrow indicates the direction of energy flow from a prey to its predator. The second panel (b) presents the z-scores obtained from comparison between the empirical motif frequency and the random expectation. The dotted lines indicate the thresholds for significant over- and under-representation (z = 1.97 and z = -1.97, respectively). The third panel (c) shows the median weight found for each motif. The error bars represent the first and third quartiles. Note that the motif id given on the x-axis corresponds to the indexing in (a), and that the interaction strengths have been transformed to approximately be normally distributed and strictly positive.

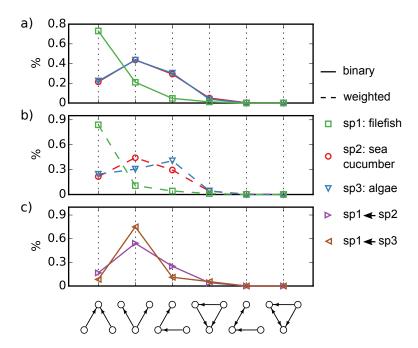
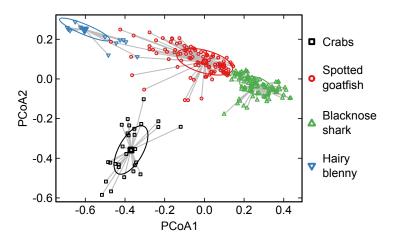


Figure 3: Analysis of the species' motif participation in the marine food web under study. The first panel (a) shows the motif-participation profiles of three representative species from the web; here, every point describes the proportion of times that these species are found in any of the possible motifs. For simplicity, we excluded the seven motifs in which these species never appear. The second panel (b) presents the motif-role profiles for the same three species when adding information regarding the interaction strengths. In this case, every point represents the relative weight associated with the motifs in which each species participates. The third panel (c) shows the motif-participation profiles for the links involving the same three species.



**Figure 4**: Principal coordinate analysis of the dissimilarity matrix containing the pairwise distances between all nodes' motif-role profiles in the marine food web under study. Every point represents a different species and each color corresponds to a group characterizing a distinct role. The species in the legend are those corresponding to the medoids of each group. The ellipses are the one-standard-deviation ellipses about the group medians.