1 Complex patterns of sex-biased demography in canines

2 Tanya N. Phung¹, Robert K. Wayne², Melissa A. Wilson Sayres^{3,5*}, Kirk E. Lohmueller^{1,2,4,5*}

3 Affiliations:

- ⁴ ¹Interdepartmental Program in Bioinformatics, University of California, Los Angeles, CA 90095, USA.
- ⁵ ²Department of Ecology and Evolutionary Biology, University of California, Los Angeles, CA 90095,
- 6 USA.
- 7 ³School of Life Sciences and Center for Evolution and Medicine, The Biodesign Institute, Arizona State
- 8 University, Tempe, AZ 85281
- 9 ⁴Department of Human Genetics, David Geffen School of Medicine, University of California, Los
- 10 Angeles, CA 90095, USA.
- ⁵These authors contributed equally to this work.
- 12
- 13 *To whom correspondence should be addressed:
- 14 Melissa A. Wilson Sayres
- 15 School of Life Sciences
- 16 Arizona State University
- 17 427 E Tyler Mall
- 18 Tempe, AZ, 85281
- 19 (480) 727-6366
- 20 melissa.wilsonsayres@asu.edu
- 21
- 22 Kirk E. Lohmueller
- 23 Department of Ecology and Evolutionary Biology
- 24 University of California, Los Angeles
- 25 621 Charles E. Young Drive South
- 26 Los Angeles, CA 90095-1606

- 27 (310)-825-7636
- 28 klohmueller@ucla.edu
- 29

30 INTRODUCTION

31 Studies of genetic variation have shown that the demographic history of dogs has been extremely 32 complex, involving multiple bottleneck and admixture events. However, existing studies have not explored the variance in the number of reproducing males and females, and whether it has changed across 33 evolutionary time. While male-biased mating practices, such as male-biased migration and multiple 34 35 paternity, have been observed in wolves, recent breeding practices could have led to female-biased mating 36 patterns in breed dogs. In addition, breed dogs are thought to have experienced the popular sire effect, 37 where a small number of males father many offspring with a large number of females. Here we use 38 genetic variation data to test how widespread sex-biased mating practices in canines are during different 39 time points throughout their evolutionary history. Using whole genome sequence data from 33 dogs and 40 wolves, we show that patterns of diversity on the X chromosome and autosomes are consistent with a higher number of reproducing males than females over ancient evolutionary history in both dogs and 41 42 wolves, suggesting that mating practices did not change during early dog domestication. In contrast, since 43 breed formation, we found evidence for a larger number of reproducing females than males in breed dogs, 44 consistent with the popular sire effect. Our results confirm that the demographic history of canines has been complex, with unique and opposite sex-biased processes occurring at different times. The signatures 45 observed in the genetic data are consistent with documented sex-biased mating practices in both the wild 46 47 and domesticated populations, suggesting that these mating practices are pervasive.

48

49 Dogs were the first animals known to be domesticated and have lived alongside humans and shared our 50 environment ever since¹. There is tremendous interest in understanding their genetics and evolutionary 51 history²⁻⁵. Many studies have shown that dogs have a complex evolutionary history; they experienced a 52 population size reduction (i.e. bottleneck) associated with domestication and additional breed-specific

bottlenecks associated with breed formation during the Victorian era⁶. In addition to bottleneck events. 53 dogs experienced admixture with wolves during the domestication process⁷. Studies have disagreed about 54 the process of domestication, including when, where, and how many times dogs were domesticated^{2,8-12}. 55 56 However, despite the extensive work on understanding dog demographic history, existing studies have 57 not explored the population history of males and females across dog domestication. Departures from an equal number of reproducing males and females are called sex-biased demographic processes, and leave 58 signatures in the genome (reviewed in Wilson Sayres 2018¹³). Previous ecological and field studies 59 suggested that mating practices have been sex-biased in canines. In the wild populations, vonHoldt et al. 60 61 (2008) observed that in some cases, Yellowstone male wolves would migrate to an existing wolf pack to mate with the alpha female when the alpha male $dies^{14}$. The migration into an existing wolf pack is 62 63 therefore male-biased. An additional source of male biased migration may come from male wolves called "Casanova wolves". These wolves leave their natal packs and visit a nearby wolf pack around mating 64 season to mate with the subordinate females¹⁵. Lastly, there has also been evidence of multiple paternity 65 in Ethiopian wolves and foxes^{16,17}. In the domesticated populations, it is thought that more females 66 contributed to breed formation than males, indicating female-biased processes¹⁸. In addition, recent 67 reproductive practices, such as the popular sire effect, which involves a small number of males 68 reproducing with a large number of females can lead to female-biased demography¹⁹. Despite these 69 observations of mating practices suggesting the numbers of reproducing males and females has been 70 unequal during canid evolution, it is unclear how pervasive these processes are, and which have had the 71 72 dominant effect on shaping patterns of diversity.

73

To test how widespread sex biased demography has been throughout canid evolution, we calculated and compared measures of genetic diversity on the X chromosome to those on the autosomes. This ratio has been termed Q in Emery et al. (2010) and we will use this notation throughout²⁰. In male-heterogametic sex-determining systems (XX/XY) with equal numbers of reproducing males and females, there are three copies of the X chromosome for every four copies of the autosomal genome. Therefore, in a constant size population without any natural selection or sex-biased processes, Q is expected to be 0.75 (reviewed in Webster and Wilson Sayres 2016²¹). Specifically, $Q = N_X/N_A \cong 0.75$. Deviations from this expected ratio could be indicative of sex-biased processes. If Q < 0.75, there are fewer copies of the X chromosome than expected, suggesting a larger number of reproducing males than reproducing females, indicative of male-biased processes. If Q > 0.75, there are more copies of the X chromosome than expected, suggesting a larger number of reproducing males, indicative of female-biased processes.

86

Studies comparing measures of genetic diversity between the X chromosome and autosomes have 87 resulted in many insights into the evolutionary history of humans. Hammer et al. (2008) computed Q by 88 fitting a model of demographic history to the ratio in the mean of genetic diversity within the X 89 chromosome and autosomes: $Q_{\pi} = \pi_X / \pi_A^{22}$. They found that Q_{π} is greater than 0.75 in all human 90 populations examined, suggesting female-biased processes that have led to more reproducing females 91 than males during human evolutionary history²². Later, Keinan et al. $(2009)^{23}$ computed Q by calculating 92 the ratio in fixation index, F_{ST} , between the X chromosome and the autosomes: $Q_{FST} = \frac{\ln (1-2F_{ST}^A)}{\ln (1-2F_{ST}^A)}$. They 93 found that Q_{FST} is less than 0.75 only when comparing a non-African population to an African 94 population²³. This result suggests that there was a male-biased migration out of Africa, where there were 95 more reproducing males than females. Even though these two studies came to different conclusions 96 regarding the sex ratio in human history, a later study reconciled these seemingly disparate findings by 97 98 demonstrating that Q can detect bias in sex ratios at different timescales, depending on whether it is calculated from genetic diversity (Q_{π}) or the fixation index $(Q_{FST})^{20}$. Specifically, Q_{π} can detect sex bias 99 in ancient timescales, which is before or immediately after the split between populations, whereas Q_{FST} 100 detects sex-biased demography on recent timescales, after the populations split from each other²⁰. Emery 101 et al. (2010) reconciled results from Hammer et al. (2008) and Keinan et al. (2009) by showing that 102 103 evolutionary processes within human history are consistent with an earlier female bias followed by a male

bias during the migration of some humans out of Africa²⁰. Additionally, direct comparisons of the two
 studies were complicated by linked selection on the X chromosome^{24,25}. In addition to humans, comparing
 the genetic diversity between the X chromosome and autosomes has also been used to study sex-biased
 processes in many other species¹³.

108

109 Given how examining patterns of genetic diversity on the X chromosome and the autosomes has facilitated our understanding of sex-biased demography in other species and what has been observed 110 regarding sex-biased mating practices in canines, we wanted to test how widespread these mating 111 112 practices are throughout different time points during canine evolutionary history. We utilized whole-113 genome sequences of 21 dogs and 12 wolves. Using the estimator of the effective sex ratio based on nucleotide diversity, we found that Q_{π} is less than 0.75 in both dogs and wolves, indicative of an ancient 114 115 male bias either in the shared ancestral population, or immediately after their split. We then inferred the effective sex ratio in a population genetic model, demonstrating that a population size reduction by itself 116 117 cannot generate the empirical patterns. Rather, a male-biased sex ratio was needed in conjunction with a 118 population size reduction to recapitulate empirical patterns. Finally, using the estimator of the effective 119 sex ratio based on the fixation index, we showed that while the demographic history in wolves has 120 remained male-biased in recent history, the demographic history in dogs has changed from male-biased in 121 the ancient timescale to female-biased in recent times. These results add to our current understanding 122 about the canine demographic history and suggest the need to incorporate sex-biased demography in 123 future studies.

124

125 **RESULTS**

126 Description of the data

127 We collected a dataset of 33 female canid whole genomes that include 4 German Shepherds, 5 Tibetan

- 128 Mastiffs, 12 dog individuals from a variety of breeds, 6 Arctic Wolves, and 6 Grey Wolves
- 129 (Supplementary Table 1). The German Shepherd and Tibetan Mastiff data were sequenced by Gou et al.

 $(2014)^{26}$ and the *fasta* files were downloaded from NCBI SRA. We combined 12 high coverage (>15X) 130 131 whole genome sequences of female dogs from multiple breeds that were included in Marsden et al. $(2016)^{27}$ because we were interested in how results differ between using a group of one breed versus using 132 a group consisting of multiple breeds. We named this pooled group the "Pooled Breed Dogs". The Arctic 133 134 Wolf data were sequenced by Robinson et al (Submitted). These Arctic Wolves were located in Northern Canada (north of the Arctic circle). The longitudinal and latitudinal locations for these Arctic Wolves are 135 included in Supplementary Table 1. We also used high coverage (>15X) whole genome sequences of 136 female Grey Wolves from Marsden et al. (2016)²⁷. Since these Grey Wolves originated from Europe, 137 138 Asia, and Yellowstone, we named this population the "Pooled Grey Wolves". Details about coverage and 139 accession numbers for the individuals in this study are summarized in Supplementary Table 1. 140 Estimating the effective sex ratio based on genetic diversity 141 142 Previous work has shown that dogs experience male mutation bias, where the mutation rate is higher in males compared to females due to more germline cell divisions in males at reproduction^{28–30}. Male 143 mutation bias has a significant impact on measurements of genetic diversity because it can inflate raw 144 metrics of genetic diversity on the autosomes compared to on the X chromosome (reviewed in Webster 145 and Wilson Sayres 2016^{21}). To confirm that male mutation bias exists in our data, we computed male 146 mutation bias for each population using dog-cat divergence (see Methods). We observed that the level of 147 male mutation bias is around 2, which is consistent with previous reports^{29,30} (Supplementary Table 2). 148 149 Therefore, we controlled for male mutation bias in all estimates of genetic variation by normalizing 150 autosomal and X chromosome diversity by dog-cat divergence in the corresponding regions. 151 Natural selection is thought to be more efficient at reducing genetic diversity on the X chromosome than 152 on the autosomes because males have only one X chromosome which is exposed directly to selection 153 (reviewed in Webster and Wilson Sayres 2016^{21}). To control for natural selection affecting the X 154 155 chromosome more than the autosomes, we used regions of the genome in which mutations would be

putatively neutral by removing sites that are functional. Specifically, we removed genic and conservedsites (see Methods).

158

To understand whether any evolutionary process has been sex-biased over ancient timescales, we computed Q_{π} . We found that in both dog and wolf populations, Q_{π} is significantly less than 0.75 (Figure 1, No cM cutoff), suggesting a male-biased sex ratio, with more males reproducing relative to females.

162

 Q_{π} of less than 0.75 could occur due to the effect of natural selection on linked neutral sites. Specifically, 163 164 natural selection could have reduced diversity in linked neutral regions on the X chromosome more than on the autosomes, as seen in humans $^{23-25}$. Further, it is possible that there is more constraint on noncoding 165 regions near genes on the X chromosome than on the autosomes³¹. To measure how neutral diversity is 166 affected by linked selection, we compared diversity on the X chromosome and autosomes in regions near 167 genes versus putatively unconstrained regions 0.4 cM away from the nearest gene. Diversity increased 168 169 more with increasing distance from genes on the X chromosome than on the autosomes, consistent with 170 natural selection reducing diversity more on the X chromosome than on the autosomes near genes 171 (Supplementary Table 3).

172

173 To test whether stronger linked selection acting on the X chromosome relative to the autosomes could cause Q_{π} to be less than 0.75, we expanded our filtering criteria to remove sites that are near genes, 174 175 defined by genetic distance (see Methods). Since we did not know a priori what the minimum genetic distance would be required to obtain sites that are not affected by selection, we included several 176 177 thresholds. We removed sites whose genetic distance to the nearest genes is less than 0.2 cM, 0.4 cM, 0.6 178 cM, 0.8 cM, and 1 cM. We observed that even after removing sites whose genetic distance to the nearest genes are less than 1 cM, Q_{π} is still less than the expected 0.75 in both dog and wolf populations, except 179 180 for the German Shepherd (Figure 1). In the German Shepherd, when using the thresholds of 0.8 cM and 1 181 cM, Q_{π} approaches 0.75. However, since there are significantly fewer sites and variants left after 182 removing sites whose genetic distance to the nearest genes is less than 0.8 cM or 1 cM, we could not 183 exclude the possibility that we are underpowered to detect any signal in the data (Supplementary Table 4). 184 Nonetheless, these results suggest that while linked selection may partially account for Q_{π} of less than 185 0.75, especially in the German Shepherd, linked selection by itself cannot explain why Q_{π} is less than 186 0.75 across all dog and wolf populations. In sum, our results suggest that there has been male-biased sex 187 ratios in both dogs and wolves over ancient evolutionary timescales.

188

189 Inference of sex-biased demographic processes under population genetic models

Pool and Nielsen (2007) demonstrated that a Q_{π} of less than 0.75 could be explained by a reduction in 190 population size even with an equal number of breeding males and females³². To test whether population 191 bottlenecks can explain the reduction in diversity on the X chromosome, we fitted a demographic model 192 193 that includes a bottleneck using the autosomal site frequency spectrum (SFS) (Supplementary Figure 1) and asked whether the best fitting demographic model on the autosomes could also account for the level 194 195 of diversity on the X chromosome when using an N_X/N_A ratio of 0.75. If a demographic model including 196 a bottleneck by itself can generate a Q_{π} of less than 0.75, we would expect that scaling the population size of the X chromosome to be three-quarters that of the autosomes should result in a Q_{π} comparable to the 197 198 empirical data. Additionally, we then employed a composite likelihood framework to directly infer the 199 N_X/N_A ratio from the SFS while accounting for the complex non-equilibrium demography.

200

First, we fitted a demographic model that includes a bottleneck using the SFS on the autosomes using *fastsimcoal2*³³ for each population considering regions of greater than 0.4 cM, 0.6 cM, 0.8 cM, and 1 cM from genes. We reasoned that we would not be able to exclude the role of selection when not removing sites near genes or using too small of a threshold (i.e. 0.2 cM). We also corrected for male mutation bias using mutation rates that we inferred from dog-cat divergence in the same windows (see Methods;

206 Supplementary Table 2). The inferred demographic parameters that resulted in the best likelihood of the 207 data are presented in Supplementary Table 5. To test whether the inferred demographic parameters can recapitulate the autosomal data, we used *fastsincoal2* to generate the expected SFSs. In all populations 208 209 except the German Shepherds, across all thresholds examined, we observed that the SFSs generated using 210 the inferred demographic parameters visually match with the empirical autosomal SFSs (Supplementary 211 Figure 2). The differences in log-likelihood between the simulated SFSs and the empirical SFSs are also 212 small (Supplementary Table 6), confirming our visual inspection of the fit of the demographic models. In 213 addition, autosomal genetic diversity (π) computed from the demographic model is comparable to the empirical estimates of π (Supplementary Figure 3). Thus, these lines of evidence demonstrate that the 214 215 inferred demographic parameters can recapitulate the empirical data on the autosomes, except for the 216 more stringent filtering on the German Shepherd (See Supplementary Note 1).

217

To understand whether the demographic model including a bottleneck that was fitted to the autosomal 218 219 data could account for the level of diversity on the X chromosome, we used the inferred demographic 220 parameters to simulate the SFSs for the X chromosome. To account for the differences in population size between the X chromosome and the autosomes, we adjusted the population size on the X chromosome by 221 a constant value which we called C, where $N_X = CN_A$. If a bottleneck by itself without any sex biased 222 223 demography can generate a Q_{π} of less than 0.75, we expected that using a C value of 0.75 would recapitulate the empirical data. If a bottleneck model by itself is not sufficient to generate a Q_{π} of less 224 than 0.75, and sex-biased processes need to be invoked, we expected that rescaling the population size on 225 226 the X chromosome to be three-quarters of the population size on the autosomes would not fit well. Rather, 227 a different value of C would yield a better fit.

To assess whether a null *C* value of 0.75 or a different *C* value yielded a better fit to the empirical SFSs
on the X chromosome, we searched over a grid of *C* values. We found the maximum likelihood value of

231 C for each population and filtering threshold. To do this, for each C on a grid of C values, we first 232 calculated the population size on the X chromosome, which is $N_X = CN_A$. We then used *fastsimcoal2* to 233 simulate an SFS and assess the fit by comparing the Poisson log-likelihood to the SFS on the X chromosome (see Methods). For each population and for each threshold, we found a set of C values that 234 235 maximizes the likelihood of the data (Figure 2, Table 1, Supplementary Table 7). 236 237 With the exception of the German Shepherd at the most stringent filtering thresholds (>0.8 cM and >1 238 cM), we inferred that C is less than 0.75 for all population and filtering thresholds. When using a filtering threshold of 0.4 cM from genes, we found that C ranges from 0.61 to 0.68. The full model, where we 239 240 inferred C for each comparison, fits the observed X chromosome SFS significantly better than a model where C is constrained to be 0.75 (Likelihood Ratio Tests > 30, p-value < 10^{-8} ; Table 1). Further, the null 241 C value of 0.75 does not visually fit the SFSs on the X chromosome (Supplementary Figure 4, blue bars), 242 243 suggesting that we can reject an equal number of reproducing males and females. Third, we observed that diversity on the X chromosome from simulating with a null C value of 0.75 overestimated the empirical X 244 chromosome diversity (Supplementary Figure 5, blue bars). These results suggest that a model including 245 246 both a bottleneck and a male-bias sex ratio can generate Q_{π} of less than 0.75 and recapitulate the observed 247 SFSs and genetic diversity. Only in the German Shepherd population when using the most stringent

threshold (>0.8 cM and >1 cM), can a demographic history including a bottleneck by itself generate a

249 Q_{π} of less than 0.75.

250

251 Female-biased sex ratio within dogs in recent history

Since estimates of sex ratios from levels of genetic diversity are sensitive to ancient sex-biased processes (prior to or immediately after the split between two species), we wanted to determine whether the pattern of male-biased contributions remained constant throughout the evolutionary history of canines²⁰. To study sex-biased demography on recent timescales, we computed Q_{FST} for each pair of populations (see 256 Methods). In the dog to dog comparison, we computed Q_{FST} between German Shepherds and Tibetan 257 Mastiffs, between German Shepherds and Pooled Breed Dogs, and between Tibetan Mastiffs and Pooled 258 Breed Dogs. We observed that Q_{FST} is greater than 0.75 for all three pairs and across all thresholds, 259 suggesting a female-biased sex ratio within the dog populations in recent history (Figure 3 and 260 Supplementary Figure 6). This is consistent with fewer reproducing males than females in the population since the formation of different dog breeds. In the wolf to wolf comparison, we computed Q_{FST} between 261 262 Arctic Wolves and Pooled Grey Wolves. In contrast to the breed dogs, we found that Q_{FST} is less than 263 0.75 when using the thresholds of >0.4 cM and >0.6 cM, suggesting that a male-biased sex ratio has been maintained within the wolf populations in recent history (Figure 3 and Supplementary Figure 6). 264 However, we noted that when using a more stringent threshold (>0.8 cM or >1 cM), Q_{FST} within wolves 265 approaches 0.75 or greater than 0.75 (Supplementary Figure 6). We could not exclude the possibility that 266 267 we are unable to detect a true signal in the data due to significantly fewer sites and variants left after the 268 more stringent filtering (Supplementary Table 4). Overall, these results indicate that while the process within wolves has probably maintained a male-bias from ancient to recent history, the process within dogs 269 270 has changed to female-bias, potentially because of breeding practices that have led to female-biased 271 processes such as the popular sire effect.

272

273 DISCUSSION

In this study, we used two different statistics to estimate the ratio of reproducing males to females in canines and found that the demographic history of dogs and wolves has been sex-biased, but not always in the same direction. Estimating the sex ratio based on the levels of genetic diversity (Q_{π}) from the X chromosome and autosomes showed a male-biased sex ratio in both dogs and wolves on an ancient timescale, which cannot be explained by linked selection or a population size reduction on its own (Figure 1 and Figure 2). Instead, in both dogs and wolves, there has been a larger number of reproducing males than females. In wolf packs, the alpha male and female are the dominant reproducers, but subdominant

reproduction is common and may involve multiple fathers for a single litter¹⁴. Multiple paternity is a 281 282 unique aspect of canid reproduction and may help drive a male bias in reproduction, as offspring of a single litter can only have a one mother, but may have multiple fathers and litter size may be as large as 283 16 individuals³⁴. In addition, wolves migrating to existing wolf packs are predominantly male-biased¹⁴. 284 285 Further, "Casanova wolves" who stay near a wolf pack during mating season to mate with the non-alpha females could also cause male-biased mating patterns¹⁵. Multiple paternity and male-biased migration 286 likely occurred in early dogs, but under more recent controlled breeding, valuable sires would be the only 287 father of a litter. Hence the controlled nature of breeding in modern dog breeds, and the focus on a subset 288 289 of "popular" sires could drive the female bias in reproduction. The population sire effect also reduces the 290 effective size of breeds and effects such as inbreeding further skew evolution in modern breeds.

291

In addition, we observed that determining the amount of bias based on the absolute value of Q_{π} by itself 292 can lead to overestimation, because the reduction of diversity on the X chromosome due to a population 293 294 size reduction is not accounted for. For example, in Tibetan Mastiff, when using a threshold of 0.6 cM to remove linked neutral sites, a Q_{π} of 0.52 suggests an N_X/N_A ratio of 0.52. However, we inferred a C 295 296 value of 0.57 (confidence interval: 0.56-0.6) using our modelling framework, indicating that the sex ratio is higher than when just examining the absolute value of Q_{π} . This difference exists because the estimate 297 of Q_{π} could be affected by a population size reduction differentially influencing diversity on the X and 298 autosomes³², but our inference framework accounts for this effect. Our findings suggest that inferring the 299 sex ratio in a model-based framework should yield a more accurate estimate than the absolute Q_{π}^{22} . 300

301

302 Our results add to the growing literature on the complex demographic history of dogs (reviewed in 303 Freedman et al. 2016³ and Ostrander et al. 2017⁴). In addition to multiple episodes of bottleneck and 304 admixture events, we now present evidence for sex-biased demographic processes. Furthermore, we 305 provide evidence that sex-biased processes within dogs have changed throughout evolution, switching 306 from a male-bias in ancient timescales to a female-bias in recent timescales, reflecting how modern 307 breeding practices influence the sex ratio. To the best of our knowledge, this is the first genomic study of 308 sex-biased demography in dogs. Some limitations in this study provide avenues for future work. First, our study was limited by the availability of high coverage (>15X coverage) whole-genome sequences of 309 310 female individuals at the time of analysis. Future studies could utilize more female individuals and a 311 variety of populations to understand whether there are differences in sex-biased processes between breeds. Second, future work could extend our modelling framework by including more complex 312 313 demographic scenarios such as migration events to better capture the autosomal data, especially the German Shepherds, Finally, future studies could examine whether processes such as admixture with 314 315 wolves or introgression has been sex-biased. 316 317 **METHODS** Whole-genome sequence processing 318 We followed Genome Analysis Toolkit's (GATK) documentation for variant discovery best practices^{35–37}. 319 320 Scripts used for processing whole-genome sequencing for each of the following steps can be found at https://github.com/tnphung/NGS pipeline. 321 322 323 Data pre-processing for variant calling First, we converted all fastq files to raw unmapped reads using Picard FastqToSam³⁸. Second, we marked 324 Illumina adapters using Picard MarkIlluminaAdapters³⁸. Third, we mapped to the reference dog genome 325 (canFam3) using bwa-mem³⁹. Fourth, we marked duplicates using Picard MarkDuplicates³⁸. We then 326 327 recalibrated base quality scores using GATK where we performed three rounds of recalibration to obtain 328 analysis-ready reads in BAM file format. 329 Variant calling with GATK 330 We used GATK Haplotype caller for variant calling^{35–37}. We first generated a gVCF file for each 331 332 individual. We then performed joint-genotyping for all 33 individuals in our study.

2	2	2
-≺	-≺	-≺
-	-	-

334 Filtering to obtain high quality sites

- 335 To obtain sites that are high confidence, we retained sites whose depth (annotated as DP in VCF file
- format) is between 50% and 150% of the mean depth across all sites. In addition, we only kept sites that
- 337 were genotyped in all 33 individuals (i.e. the total number of alleles in called genotypes, AN, is equal to
- **338** 66).
- 339

340 Variant filtering

- 341 We obtained variant sites from the VCF files by using GATK SelectVariants^{35–37}. We then filtered these
- variants by applying GATK Hard Filter (QD < 2.0, FS > 60.0, MQ < 40.0, MQRankSum < -12.5,
- ReadPosRankSum < -8.0). In addition, we only selected biallelic SNPs and removed any clustered SNPs
- defined by having 3 SNPs within 10bp.
- 345

346 Filtering nucleotide sites

347 Filtering out the pseudoautosomal regions (PARs) of the X chromosome

348 Previous work showed that the PARs in canines span the first 6.59Mb of the X chromosome⁴⁰. Therefore,

349 we filtered out the PARs by removing any site that overlaps with the first 6.59Mb of the X chromosome.

- In humans, it was shown that genetic diversity does not drop abruptly at the PAR boundary⁴¹. Rather,
- 351 genetic diversity decreases gradually over the PAR boundary and reaches nonPAR diversity past the PAR

boundary⁴¹. One concern is that filtering out the PARs is not sufficient to avoid any inflation of X-linked

- variation. However, if this is the case, we would expect Q_{π} we calculated to be higher than the actual Q_{π} .
- Therefore, Q_{π} less than 0.75 is not caused by not sufficiently filtering sites on the nonPARs.

- 356 *Filtering sites that could be under the direct effect of selection*
- 357 To control for the effects of direct selection, we removed sites that are potentially functional and therefore
- are more likely to be affected by purifying or positive selection. Specifically, we removed sites that

overlap with a gene transcript as defined by Ensembl (gene transcripts include both exons and introns).
We also removed sites that are conserved across species. To obtain conserved sites, we downloaded
phastConsElements100way for hg19 from the UCSC Genome Browser and used liftOver command line
tool to convert hg19 coordinates to canFam3 coordinates.

363

364 *Filtering out sites that could be affected by linked selection*

365 To control for the effect of natural selection on linked neutral sites, we employed a filtering criterion to remove sites near genes as defined by genetic distance to the nearest genes. We used the genetic distance 366 367 map based on patterns of linkage disequilibrium from Auton et al. (2013) because this genetic map 368 includes information for the X chromosome whereas the pedigree map from Campbell et al. (2016) does not have information on the X chromosome 42,43 . For each site that is outside of genes and conserved 369 370 regions, we found its nearest gene in terms of physical distance. We then converted physical distance to genetic distance using the genetic map from Auton et al. $(2013)^{42}$. Since we did not know *a priori* what 371 372 the minimum genetic distance is required to remove sites near genes to control for linked selection, we used multiple thresholds. Specifically, we removed sites whose genetic distance to the nearest gene is less 373 than 0.2 cM, less than 0.4 cM, less than 0.6 cM, less than 0.8 cM, and less than 1 cM. 374

375

376 Identifying sites that are alignable between dog and cat

377 Since we controlled for mutation rate variation by normalizing the uncorrected genetic diversity by dog-

378 cat divergence, we identified regions of the genome that are alignable between dog and cat. We

downloaded the pairwise alignment between dog and cat from the UCSC Genome browser⁴⁴. We then

380 generated BED files whose coordinates represent regions of the genome that are alignable between dog

and cat.

- In summary, for our empirical analyses, we used regions of the genome that are (1) not affected directly
- by selection, (2) not affected by linked selection using multiple thresholds, (3) high in quality (see the
- section on filtering to obtain high quality sites above), and (4) alignable between dog and cat.
- 386

387 Computing Q_{π}

- 388 Computing uncorrected average pairwise differences between sequences (π)
- 389 We computed genetic diversity, π , defined as the average number of differences between pairs of 390 sequences⁴⁵:

391
$$\pi = \frac{n}{n-1} \sum_{i}^{\text{all sites}} p_i (1-p_i)$$
 where p_i is the allele frequency and n is the number of alleles. For each

region of the genome that satisfies the filtering criteria above, we computed π for the X chromosome and

autosomes. To obtain the mean in diversity, π/site , we calculated: $\pi/\text{site} = \frac{\sum_{i}^{\text{regions}} \pi}{\sum_{i}^{\text{regions}} \text{total sites}}$.

394

395 *Computing dog-cat divergence*

396 For each region of the genome that satisfies the filtering criteria above, we tabulated the number of DNA

397 differences between dog and cat. To obtain the mean in divergence, we calculated $\frac{\text{divergence}}{\text{site}} =$

398
$$\frac{\sum_{i}^{\text{regions}} \text{number of divergent sites}}{\sum_{i}^{\text{regions}} \text{total sites}}$$

399

400 *Computing male mutation bias*

401 We computed male mutation bias (α) using divergence on the X chromosome and on the autosomes as

402 follows⁴⁶:
$$\alpha = \frac{4-3\frac{X}{A}}{3\frac{X}{A}-2}$$
.

403

404 *Computing corrected diversity*

- 405 To control for variation in mutation rates across chromosomes, we normalized diversity by dog-cat
- 406 divergence by dividing π /site by divergence/site.
- 407
- 408 Constructing 95% confidence interval by bootstrapping
- 409 We generated bootstrap replicates of the BED file that we used to compute genetic diversity and
- 410 divergence by randomly selecting a fragment from the BED file with replacement. For each bootstrap
- 411 replicate, the number of fragments chosen was equal to the number of fragments in the original BED file.
- 412 We generated 1000 bootstrap replicates. For each of the 1000 bootstraps on the X chromosome, we
- 413 computed uncorrected π , dog-cat divergence, and corrected π . We did the same calculations for each of
- 414 the 1000 bootstraps on the autosomes. We then divided corrected π on the X chromosome by corrected π
- 415 on the autosomes to obtain Q_{π} . We calculated 95% confidence interval using 1000 bootstrapped values of
- 416 corrected π_X , 1000 bootstrapped values of corrected π_A , and 1000 bootstrapped values of corrected Q_{π} by
- 417 selecting the values at the 2.5 and 97.5 percentiles.
- 418

419 Computing Q_{FST}

420 *Computing* F_{ST}

We computed Weir and Cockerham's F_{ST} for each pair of populations using the *SNPRelate* package implemented in R^{47} . For dog-to-dog comparison, we computed F_{ST} for German Shepherds and Tibetan Mastiffs, German Shepherds and Pooled Breed Dogs, and Tibetan Mastiff and Pooled Breed Dogs. For wolf-to-wolf comparison, we computed F_{ST} for Arctic Wolves and Grey Wolves. Since the number of individuals differs between populations, we subsampled such that there were four individuals in each population (Supplementary Table 8). We computed F_{ST} for the X chromosome and for the autosomes.

428 Computing Q_{FST}

429 We computed
$$Q_{FST}$$
 using: $Q_{FST} = \frac{\ln(1-2F_{ST}^A)}{\ln(1-2F_{ST}^X)} \frac{20,23}{20,23}$.

43	0
----	---

- 431 *Constructing 95% confidence interval by bootstrapping*
- 432 Since the input to *SNPRelate* to calculate F_{ST} is a VCF file format, we generated 1000 bootstrapped VCF
- 433 files by randomly selecting variants from the VCF file with replacement. The number of variants selected
- 434 for each bootstrapped VCF is equal to the number of variants in the empirical VCF file. For each
- 435 bootstrapped VCF, we computed F_{ST} and Q_{FST} as explained above. From the 1000 values of bootstrapped
- 436 Q_{FST} , we then calculated 95% confidence interval by selecting the values at the 2.5 and 97.5 percentiles.
- 437

438 Modeling framework to estimate the N_X/N_A ratio (C)

439 *Obtaining the site frequency spectrum (SFS)*

440 We computed the folded SFSs using Equation 1.2 of Wakely's An Introduction to Coalescent Theory⁴⁸,

441 reproduced as follows:

442
$$\eta_i = \frac{\xi_i + \xi_{n-i}}{1 + \delta_{i,n-i}} \quad 1 \le i \le [n/2]$$

where ξ_i is the number of sites where the alternate allele is present at *i* copies, $\delta_{i,n-i}$ is equal to 0 when $i \neq n-i$ and is equal to 1 when i = n - i. For each population and for each threshold to remove linked neutral sites (>0.4 cM, >0.6 cM, >0.8 cM, and >1 cM), we computed the folded SFSs for the X chromosome and autosomes.

447

448 *Computing mutation rates*

449 We utilized dog-cat divergence to infer the mutation rates for the X chromosome and autosomes.

450 Specifically, $\mu = \frac{D}{2t_{enlit}}$, where D is the divergence/site between dog and cat (see Computing dog-cat

- 451 divergence section above) and t_{split} is the split time between dog and cat in unit of generation. We used
- 452 54 million years as the split time between dog and cat and a generation time of 3 years per generation 49,50 .

453

The estimates of mutation rates are in the same order of magnitude as estimate from ancient DNA

454 (Supplementary Table 4)⁸.

455

456 Inferring demographic parameters

457 We inferred demographic parameters from the autosomal data (SFSs on the autosomes) using a maximum likelihood framework as implemented in *fastsimcoal2*³³. We specified a bottleneck demographic model 458 and inferred four parameters: NANC which is the population size in the ancestral population, NBOT which is 459 the population size during the bottleneck, N_{CUR} which is the population size in the current day, and T_{BOT} 460 461 which is the duration between the end of the bottleneck and current day (Supplementary Figure 1). Further, we repeated the inference of the previous four parameters for values BOT_{DUR} (the duration of the 462 bottleneck) ranging from 75 to 100 generations (Supplementary Figure 1) and chose the value that yielded 463 the highest likelihood. We implemented this procedure for each population and for thresholds of >0.4 cM, 464 >0.6 cM, >0.8 cM, and >1 cM to remove linked sites. The demographic parameters that maximized the 465

466 likelihood are summarized in Supplementary Table 5.

467

468 Inferring N_X/N_A ratio (C)

To account for differences in population size between the X chromosome and autosomes, we scaled the population size on the X chromosome to that on the autosomes by a constant factor we called *C*, where $N_X = CN_A$. To find the maximum likelihood estimate of *C*, we searched over a grid for values of *C*, including 0.75, to find a value that resulted in the highest likelihood. Because the number of SNPs at particular frequencies contains substantial information about demography, we used a Poisson likelihood for the number of SNPs in each entry of the SFS to compute the Poisson log-likelihood as in Beichman et al. $(2017)^{51}$.

476

477 Accessing fit of MLEs of C to π

- 478 We computed diversity from the simulated SFSs under the demographic models fit to the autosomes
- using the MLEs of C (Table 1) and compared that to the empirical uncorrected diversity.
- 480

481 Data availability

- 482 All scripts can be found at https://github.com/tnphung/SexBiased. SRA numbers for *fastq* files are listed
- 483 in Supplementary Table 1. Post base quality score calibration (BQSR) BAM files and VCF files are

484 available upon request.

485

486 ACKNOWLEDGEMENTS

- 487 We thank Jacqueline Robinson for providing the sequencing data for the Arctic Wolves and Christian
- 488 Huber for helpful discussions. This work was supported by the National Institute of General Medical
- 489 Sciences (NIGMS) of the National Institutes of Health (NIH) grant R35GM119856 to K.E.L and NIGMS
- 490 grant R35GM124827 to M.A.W.S. T.N.P. was supported by NIH-NCI National Cancer Institute
- 491 T32CA201160.
- 492

493 FIGURES

- 494 Figure 1. X-linked and autosomal genetic diversity across canids. Genetic diversity measured as the
- 495 average pairwise differences between sequences (π) corrected for mutation rate variation using divergence
- 496 (see Methods) on the X chromosome and autosomes in multiple canid populations. *Q* denotes the ratio of
- 497 π on the X chromosome to that of the autosomes. The horizontal red line denotes the null expectation of
- 498 0.75. Bins along the x-axis denote different filtering based on genetic distances from genes. Error bars
- denote 95% confidence intervals obtained through bootstrapping (see Methods).

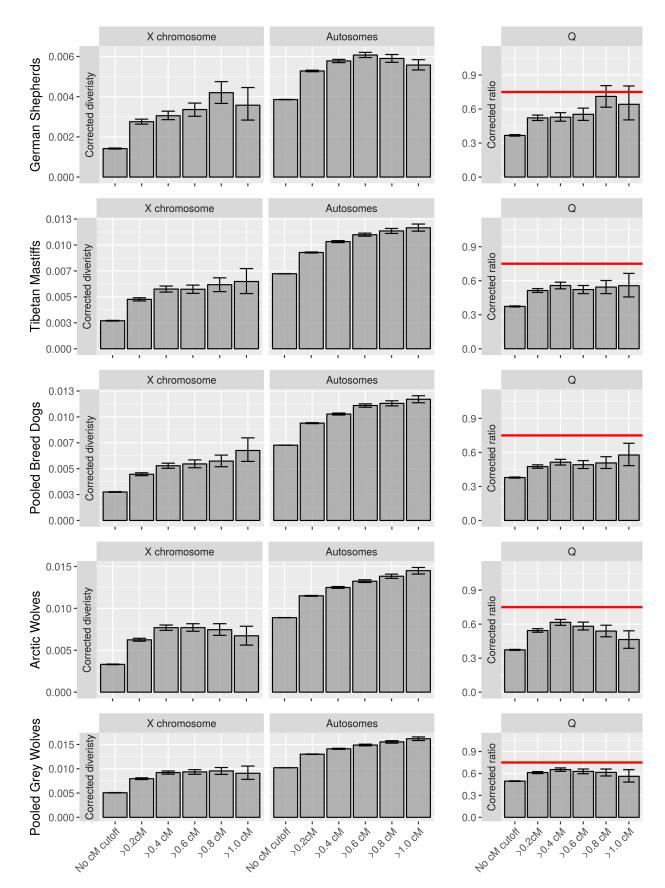


Figure 2. Effective population size estimates for multiple canid populations. Maximum likelihood 501

- 502 estimates (MLEs) of the effective population size on the X chromosome relative to that of the autosomes
- $(C = N_X/N_A)$ are shown for German Shepherds, Tibetan Mastiffs, Pooled Breed Dogs, Arctic Wolves and 503
- Pooled Grey Wolves with increasing distance from genes. Error bars denote approximate asymptotic 95% 504
- 505 confidence intervals obtained as the parameter values within 2 log-likelihood unites of the MLE.

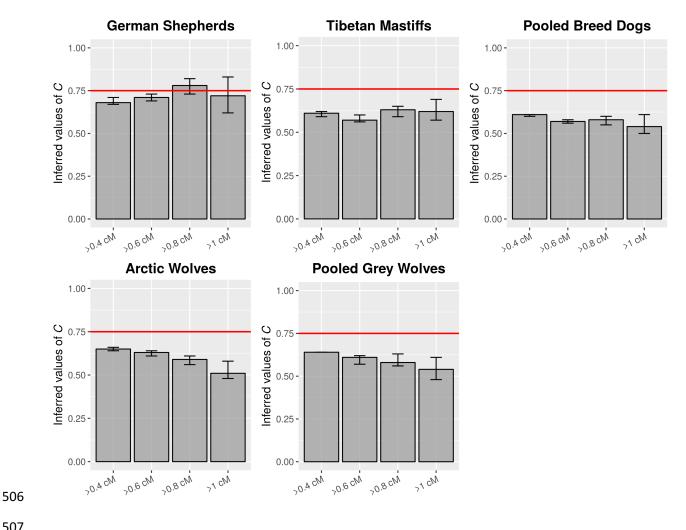
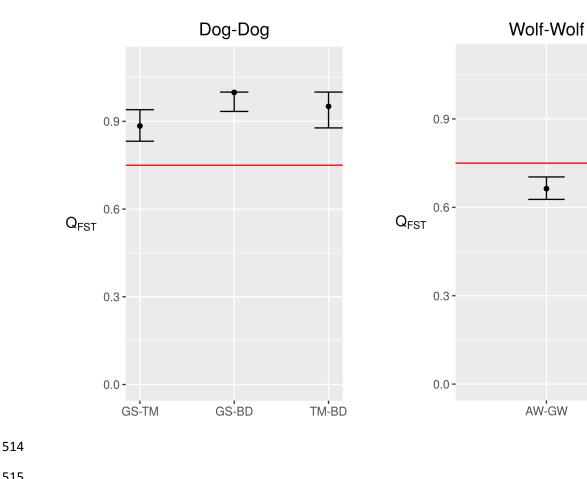


Figure 3. Sex biased demography on recent time scales. Estimates of the sex ratio for a pair of 508

- 509 populations computed using F_{ST} (see Methods) using a threshold of >0.6 cM to remove linked neutral
- 510 sites are shown. The horizontal red line denotes the null expectation of 0.75. Error bars denote 95%
- 511 confidence intervals obtained through bootstrapping (see Methods). Abbreviations: GS (German
- 512 Shepherds), TM (Tibetan Mastiffs), BD (Pooled Breed Dogs), AW (Arctic Wolves), GW (Pooled Grey
- 513 Wolf).



516 TABLES

517 Table 1. Likelihood ratio tests comparing models of sex-biased demography in multiple canid

518 **populations.** Likelihood ratio tests of the amount of sex-biased demography are shown when removing

- any sites whose genetic distance to the nearest genes is less than 0.4 cM. For the other thresholds, see
- 520 Supplementary Table 7.

Population	С	Log-likelihood	Likelihood ratio test	p-value
German Shepherds	Null ($C = 0.75$)	7460.957	34.779	3.69 X 10 ⁻⁹
	Best ($C = 0.68$)	7478.347		
Tibetan Mastiffs	Null ($C = 0.75$)	16050.84	208.972	2.30 X 10 ⁻⁴⁷
	Best ($C = 0.61$)	16155.32		
Pooled Breed Dogs	Null ($C = 0.75$)	16875.26	249.627	3.13 X 10 ⁻⁵⁶
	Best ($C = 0.61$)	17000.07		
Arctic Wolves	Null (<i>C</i> = 0.75)	23035.46	133.341	7.62 X 10 ⁻³¹
	Best ($C = 0.65$)	23102.13		
Pooled Grey Wolves	Null ($C = 0.75$)	30767.69	188.719	6.05 X 10 ⁻⁴³
	Best ($C = 0.64$)	30862.05		

521

523 **REFERENCES**

- Hemmer, H. Domestication: The Decline of Environmental Appreciation. (Cambridge University Press, 1990).
- 526 2. Freedman, A. H. *et al.* Genome sequencing highlights the dynamic early history of dogs. *PLoS*527 *Genet.* 10, e1004016 (2014).
- 528 3. Freedman, A. H., Lohmueller, K. E. & Wayne, R. K. Evolutionary history, selective sweeps, and
 529 deleterious variation in the dog. *Annu. Rev. Ecol. Evol. Syst.* 47, 73–96 (2016).
- 530 4. Ostrander, E. A., Wayne, R. K., Freedman, A. H. & Davis, B. W. Demographic history, selection
 531 and functional diversity of the canine genome. *Nat. Rev. Genet.* 18, 705–720 (2017).
- 5. Freedman, A. H. & Wayne, R. K. Deciphering the origin of dogs: from fossils to genomes. *Annu. Rev. Anim. Biosci.* 5, 281–307 (2017).
- 534 6. Boyko, A. R. The domestic dog: man's best friend in the genomic era. *Genome Biol.* 12, 216
 535 (2011).
- vonHoldt, B. M. *et al.* A genome-wide perspective on the evolutionary history of enigmatic wolflike canids. *Genome Res.* 21, 1294–1305 (2011).
- 538 8. Larson, G. *et al.* Rethinking dog domestication by integrating genetics, archeology, and
 539 biogeography. *Proc. Natl. Acad. Sci.* 109, 8878–8883 (2012).
- 540 9. Thalmann, O. *et al.* Complete mitochondrial genomes of ancient canids suggest a European origin
 541 of domestic dogs. *Science* 342, 871–874 (2013).
- 542 10. Botigué, L. R. *et al.* Ancient European dog genomes reveal continuity since the Early Neolithic.
 543 *Nat. Commun.* 8, 16082 (2017).
- Frantz, L. A. F. *et al.* Genomic and archaeological evidence suggest a dual origin of domestic
 dogs. *Science* 352, 1228–1231 (2016).
- 546 12. Drake, A. G., Coquerelle, M. & Colombeau, G. 3D morphometric analysis of fossil canid skulls
 547 contradicts the suggested domestication of dogs during the late Paleolithic. *Sci. Rep.* 5, 8299 (2015).
- 548 13. Wilson Sayres, M.A. Genetic diversity on the sex chromosomes. *Genome Biol. Evol.* 10, 1064–
 549 1078 (2018).
- Vonholdt, B. M. *et al.* The genealogy and genetic viability of reintroduced Yellowstone grey
 wolves. *Mol. Ecol.* 17, 252–274 (2008).
- 15. Casanova wolves | Natural History. Available at: https://retrieverman.net/2010/12/17/casanova wolves/. (Accessed: 26th June 2018)
- Baker, P. J., Funk, S. M., Bruford, M. W. & Harris, S. Polygynandry in a red fox population:
 implications for the evolution of group living in canids? *Behav. Ecol.* 15, 766–778 (2004).

556 17. Sillero-Zubiri, C., Gottelli, D. & Macdonald, D. W. Male philopatry, extra-pack copulations and 557 inbreeding avoidance in Ethiopian wolves (Canis simensis). Behav. Ecol. Sociobiol. 38, 331-340 (1996). 558 Sundqvist, A.-K. et al. Unequal contribution of sexes in the origin of dog breeds. Genetics 172, 18. 559 1121-1128 (2006). 560 19. Ostrander, E. A. & Kruglyak, L. Unleashing the canine genome. Genome Res. 10, 1271-1274 561 (2000).Emery, L. S., Felsenstein, J. & Akey, J. M. Estimators of the human effective sex ratio detect sex 562 20. 563 biases on different timescales. Am. J. Hum. Genet. 87, 848-856 (2010). 564 21. Webster, T. H. & Wilson Sayres, M. A. Genomic signatures of sex-biased demography: progress 565 and prospects. Curr. Opin. Genet. Dev. 41, 62-71 (2016). 566 22. Hammer, M. F., Mendez, F. L., Cox, M. P., Woerner, A. E. & Wall, J. D. Sex-biased 567 evolutionary forces shape genomic patterns of human diversity. PLoS Genet. 4, e1000202 (2008). 23. Keinan, A., Mullikin, J. C., Patterson, N. & Reich, D. Accelerated genetic drift on chromosome X 568 during the human dispersal out of Africa. Nat. Genet. 41, 66-70 (2009). 569 570 24. Hammer, M. F. et al. The ratio of human X chromosome to autosome diversity is positively 571 correlated with genetic distance from genes. Nat. Genet. 42, 830-831 (2010). 572 Arbiza, L., Gottipati, S., Siepel, A. & Keinan, A. Contrasting X-linked and autosomal diversity 25. 573 across 14 human populations. Am. J. Hum. Genet. 94, 827-844 (2014). 574 26. Gou, X. et al. Whole-genome sequencing of six dog breeds from continuous altitudes reveals 575 adaptation to high-altitude hypoxia. Genome Res. 24, 1308-1315 (2014). 27. 576 Marsden, C. D. et al. Bottlenecks and selective sweeps during domestication have increased 577 deleterious genetic variation in dogs. Proc. Natl. Acad. Sci. 113, 152-157 (2016). 578 28. Li, W. H., Yi, S. & Makova, K. Male-driven evolution. Curr Opin Genet Dev 12, (2002). 579 29. Wilson Sayres, M. A. & Makova, K. D. Genome analyses substantiate male mutation bias in many species. BioEssays News Rev. Mol. Cell. Dev. Biol. 33, 938-945 (2011). 580 581 30. Lindblad-Toh, K. et al. Genome sequence, comparative analysis and haplotype structure of the 582 domestic dog. Nature 438, 803-819 (2005). 583 31. Narang, P., Wilson Savres, M. A. Variable autosomal and X divergence near and far from genes 584 affects estimates of male mutation bias in great apes. Genome Biol. Evol. 8, 3393-3405 (2016). 585 32. Pool, J. E. & Nielsen, R. Population size changes reshape genomic patterns of diversity. Evolution 61, 3001–3006 (2007). 586 587 33. Excoffier, L., Dupanloup, I., Huerta-Sánchez, E., Sousa, V. C. & Foll, M. Robust demographic inference from genomic and SNP data. PLoS Genet 9, e1003905 (2013). 588

Stahler, D. R., MacNulty, D. R., Wayne, R. K., vonHoldt, B. & Smith, D. W. The adaptive value
of morphological, behavioural and life-history traits in reproductive female wolves. *J. Anim. Ecol.* 82,
222–234

592 35. McKenna, A. *et al.* The Genome Analysis Toolkit: A MapReduce framework for analyzing next-593 generation DNA sequencing data. *Genome Res.* **20**, 1297–1303 (2010).

594 36. DePristo, M. A. *et al.* A framework for variation discovery and genotyping using next-generation
595 DNA sequencing data. *Nat. Genet.* 43, 491–498 (2011).

596 37. Van der Auwera, G. A. *et al.* From FastQ data to high confidence variant calls: the Genome
597 Analysis Toolkit best practices pipeline. *Curr. Protoc. Bioinforma.* 43, 11.10.1-33 (2013).

598 38. Picard Tools - By Broad Institute. Available at: http://broadinstitute.github.io/picard/. (Accessed:
599 9th March 2018)

600 39. Li, H. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM.
 601 *ArXiv13033997 Q-Bio* (2013).

40. Young, A. C., Kirkness, E. F. & Breen, M. Tackling the characterization of canine chromosomal
breakpoints with an integrated in-situ/in-silico approach: The canine PAR and PAB. *Chromosome Res.*16, 1193–1202 (2008).

605 41. Cotter, D. J., Brotman, S. M. & Wilson Sayres, M. A. Genetic diversity on the human X
606 chromosome does not support a strict pseudoautosomal boundary. *Genetics* 203, 485-492 (2016).

42. Auton, A. *et al.* Genetic recombination is targeted towards gene promoter regions in dogs. *PLoS Genet* 9, e1003984 (2013).

Campbell, C. L., Bhérer, C., Morrow, B. E., Boyko, A. R. & Auton, A. A pedigree-based map of
recombination in the domestic dog genome. *G3 GenesGenomesGenetics* 6, 3517–3524 (2016).

611 44. Kent, W. J. *et al.* The Human Genome Browser at UCSC. *Genome Res.* **12**, 996–1006 (2002).

45. Tajima, F. Evolutionary relationship of DNA sequences in finite populations. *Genetics* 105, 437–
460 (1983).

46. Link, V., Aguilar-Gómez, D., Ramírez-Suástegui, C., Hurst, L. D. & Cortez, D. Male mutation
bias is the main force shaping chromosomal substitution rates in monotreme mammals. *Genome Biol. Evol.* 9, 2198–2210 (2017).

47. Zheng, X. *et al.* A high-performance computing toolset for relatedness and principal component
analysis of SNP data. *Bioinforma. Oxf. Engl.* 28, 3326–3328 (2012).

619 48. Wakely, J. Coalescent Theory: An Introduction. (Macmillan Learning, 2016).

49. Hedges, S. B., Marin, J., Suleski, M., Paymer, M. & Kumar, S. Tree of life reveals clock-like
speciation and diversification. *Mol. Biol. Evol.* 32, 835–845 (2015).

- 50. Hedges, S. B., Dudley, J. & Kumar, S. TimeTree: a public knowledge-base of divergence times
 among organisms. *Bioinforma. Oxf. Engl.* 22, 2971–2972 (2006).
- 51. Beichman, A. C., Phung, T. N. & Lohmueller, K. E. Comparison of single genome and allele
 frequency data reveals discordant demographic histories. *G3 Genes Genomes Genet.* 7, 3605–3620

^{626 (2017).}