1	The effect of nutrient storage on courtship behavior and copulation frequency in the
2	fruit fly, Drosophila melanogaster.
3	
4 5 6 7	Hannah Ananda Bougleux Gomes ^{a1&,} , Justin R. DiAngelo ^{b&} , and Nicholas Santangelo ^{a*&}
, 8 9	^a Department of Biology, 114 Hofstra University, Hempstead, NY, 11549
10 11	^b Division of Science, Penn State Berks, Reading, PA 19610
12 13 14	¹ Present address: Harvard University, Department of Stem Cell and Regenerative Biology, 7 Divinity Ave, Cambridge, MA 02138
15	
16	*Corresponding author:
17 18 19 20	E-mail: <u>Nicholas.santangelo@hofstra.edu</u>
21	^{&} These authors contributed equally to the work
22	
23	
24	
25	
26	
27	
28	
29	
30	

31 Abstract

32 Nutrient storage and metabolism effects on reproductive behavior are well studied in 33 higher vertebrates like mammals, but are less understood in simpler systems. Drosophila 34 *melanogaster* is well suited to study the ramifications of diet and metabolic energy 35 storage on reproductive behaviors as they are commonly used to explore energy 36 mobilization pathways. We tested, for the first time, courtship of the naturally occurring 37 *adipose (adp⁶⁰)* mutant which over-accumulates triglycerides and glycogen on a normal 38 diet. We also fed wild type (WT) flies either a normal diet, high fat diet or food deprived 39 them before measuring courtship, copulations, and glycogen and triglyceride levels. 40 Adipose mutants decreased both courtship and copulation frequency, yet showed the 41 highest glycogen and triglyceride levels. We suggest the adp^{60} physique and/or an altered 42 ability to utilize mobilize energy explains these effects. Food deprived WT flies had the 43 lowest glycogen and triglycerides but exhibited shortened courtship latencies with 44 increased courtship behaviors. This may be due to a decreased lifespan of food deprived 45 flies leading to a greater reproductive drive. However, high fat fed flies copulated more 46 frequently and had the highest triglycerides among WT groups, yet equal glycogen levels 47 to the normal fed WT group. Thus, a high fat diet either increases male attractivity or 48 male courtship persistence. Taken together, available diet and nutrient storage affects 49 male fly reproductive behavior in a unique manner, which may be explained by their 50 natural history, and provides a paradigm for understanding energetics based on 51 reproductive potential.

52

53

54 Introduction

55 Caloric intake is known to play an important role in different organisms' behaviors. For 56 example, consumption of a high-calorie diet alters the function of the mammalian 57 circadian clock, thereby affecting behavioral processes, such as locomotor activity, sleep 58 and energy homeostasis [1]. In the fruit fly Drosophila melanogaster, the presence of 59 food promotes aggressive behaviors in males mediated by sweet-sensing gustatory 60 receptor neurons [2]. Conversely, caloric restriction can also affect behaviors. A long-61 term experiment with humans in free-living conditions showed that caloric restriction 62 resulted in decreased physical activity levels [3]. Studies such as these show that altering 63 caloric intake not only impacts behavior via activity level decisions, but social 64 interactions as well.

65 In addition to caloric intake influencing behavior, the storage of energy as 66 glycogen and fat and the ability to mobilize these stores also influences reproductive 67 behaviors. For example, available energy resources during mating of tetrapods are well 68 known to be related to the physiological ability of these animals to carry out reproduction 69 (mammals: reviewed in [4], [5], [6], [7]; birds: reviewed [8], [9]; reptiles: [10], [11]). The 70 clear link between reproductive physiology and energy reserves suggests that appetitive 71 reproductive behaviors should also be tightly linked to these energy reserves. Such a 72 mechanism would enable organisms to adequately assess whether the proper energetic 73 resources are available to complete the process of reproduction once started. This is well 74 known to happen in mammals, where the hormonal pathways that control energy balance 75 are directly linked to the ones that control sexual reproduction (reviewed in [6]). 76 However outside of mammals, data on the link between reproductive behavior and

77 caloric intake and mobilization is lacking (but see [12] and review within on birds). In 78 addition, males are rarely the subject of such studies, yet one would expect that, despite 79 reproduction requiring less energetic output for males, the same selective pressure of 80 monitoring metabolic energy resources in making reproductive decisions should remain. 81 Cheng et al. [13] showed that male oriental fruit flies (*Bactrocera dorsalis*) reared on 82 food with high levels of D-glucose exhibit higher success in mating. Also, Mediterranean 83 fruit flies (Ceratitis capitata) caught while undergoing reproductive behaviors had higher 84 lipid levels than flies at rest [14] with males having much more variable levels of lipids 85 than females. Yet the effect of energy availability on appetitive reproductive behaviors 86 such as courtship is not as well known. Such data could provide the beginnings of a 87 framework to understand how metabolic energy resources shape appetitive behaviors. 88 In order to explore any correlation that may exist between reproductive behavior 89 and metabolism, energy storage can be manipulated genetically and/or through diet. The 90 fruit fly *Drosophila melanogaster* has been a useful model system for these types of 91 studies because of its high genetic conservation to humans, its ease of obtaining large 92 sample sizes, the ease of genetic and dietary manipulations, and well-characterized and 93 robust reproductive behaviors that can be easily quantitated. In D. melanogaster there 94 exist mutants in metabolic genes that result in lean or obese phenotypes similar to that 95 seen when wild type flies are food deprived or fed a high-fat diet, respectively [15]. 96 These mutants can be utilized to test whether altering metabolic pathways affects 97 behavior. One such gene that can be useful in this regard is *adipose (adp)*. The most well-98 characterized mutation of this gene is adp^{60} , a 23 nucleotide deletion resulting in 99 increased storage of triglycerides in the fly fat body, providing an obesity phenotype,

100 similar to the triglyceride accumulation observed when flies are fed a high fat diet [16], [17]. While both the adp^{60} mutation and feeding flies a high fat diet leads to an obesity 101 102 phenotype, the adp^{60} mutation results in chronic obesity as the fat accumulation is 103 observed in both the larval and adult stages of development [16], [18]. This is different 104 than the obesity observed after feeding flies a high fat diet as this obesity is acute and 105 only appears in adult flies after 4-5 days of being exposed to the altered diet [17]. While 106 much is known about the metabolic consequences due to the loss of the *adipose* gene, little is known about how the obesity phenotype resulting from this *adp*⁶⁰ mutation affects 107 108 reproductive behaviors. 109 Metabolic mutants can potentially have similar changes in metabolism as those 110 fed different diets without manipulating the feeding regimen, thus strengthening the link 111 between energy storage and reproductive behaviors. Both approaches, and reproductive 112 behavior monitoring, can be easily carried out in *D. melanogaster* making them an ideal 113 model for which to explore this paradigm. For example, drive to reproduce can be 114 assessed through courtship behaviors such as wing vibrations, wing scissoring, tapping, 115 thrusting, and copulation attempts and copulations [19]. In this study, we compare these 116 reproductive behaviors from *adp*⁶⁰ male flies, wild type males fed a high-fat diet, or wild 117 type males food deprived for 24 hours to wild type male flies fed a normal diet in order to 118 determine the effects of changes in nutrient storage and metabolism on reproductive 119 behaviors. For the purposes of this study, female flies are kept constant (i.e. are only of 120 WT genotype and not diet manipulated) in order to isolate any potential effects of 121 nutrient storage on male courtship decisions.

122 Therefore, in addressing the effects of obese mutants and high fat feeding and 123 starvation on reproduction, our study is an opportunity to increase the base of knowledge 124 on the behavioral effects of obesity by utilizing the adp^{60} mutant in addition to the diet-125 induced obesity model. By comparing the courtship behaviors and frequency of 126 copulation of male flies fed a high-fat diet, males food deprived for 24 hours, and male *adp*⁶⁰ mutants, with wild type *D. melanogaster* males fed a normal diet, we aim to 127 128 understand how energy resources and nutrient availability affect reproductive behaviors. 129 The use of both diet and *adp*⁶⁰ mutants will help establish whether any resulting changes 130 in male courtship due to diet regime is related to changes in nutrient storage (i.e. presence 131 or absence of adp^{60} gene), or is the simply the cue of caloric intake (i.e. high or low 132 caloric diet).

Relative to normal fed wild type flies, we predict adp^{60} mutant flies, and flies fed 133 134 a high-fat diet, to show an increase in copulation frequency and an increase in courtship 135 behaviors because of the enhanced energy reserves that could be invested in courtship. 136 This prediction is consistent with data from Kauffmann and Rissman [20] where the 137 hormone GnRH-II mediates sexual behavior in mice when there are enough energy 138 resources present. This helps ensure enough energy is available to mediate successful 139 reproduction. Though invertebrates such as insects may utilize a different hormonal 140 mechanism, we predict the relationship between energetic requirements and successful 141 reproduction to be a relatively conserved relationship. Conversely, we expect food 142 deprived flies to show a decrease in copulation and courtship for similar reasons. We 143 expect that the *adp*⁶⁰ mutants show a similar duration of courtship behaviors and courtship latency to the wild type flies fed a high-fat diet and thus that adp^{60} mutants are 144

145 in fact a good model for understanding the effects of obesity on outward male sexual

146 behavior.

147

148 Materials and Methods

149 Fly husbandry

150 Flies used in this study were: OreR (BL#2376) and *adp*⁶⁰ outcrossed into the OreR

151 genetic background (a gift from Ronald Kuhnlein). For the courtship assays, 1-2 day old

152 male flies were separated from females and aged 4-5 days before being put into the

153 courtship apparatus with 3-5-day old wild type OreR virgin females. For high fat diet

154 studies, OreR males were fed cornmeal-sucrose medium (9g Drosophila agar, 40g

sucrose, 65g cornmeal, 25g Red Star whole yeast, 100 mL Karo lite corn syrup in 1200

156 mL dH₂O) supplemented with 30% coconut oil as described previously [17] during the 4-

157 5-day aging period described above. For starvation experiments, male flies were food

deprived on 1% agar for the last 24 hours of the aging period described above.

159

160 **Courtship analysis**

161 Males and females were transferred to different sides of the divider in Aktogen ®

162 courtship chambers by cooling them on ice and transferring with forceps. Once flies

163 recovered from cooling and were walking normally in the chamber, the divider was

- 164 removed allowing flies to interact. A video camera was placed over the chamber and all
- 165 pairs were video recorded for three hours. Four chambers were always run together; two

166 normal fed wild type male trials were always paired with two trials of one of the other 167 three groups (food deprived, high fat diet, and adp^{60}).

168 Videos were uploaded to a behavioral event recorder program, Observer [®]. The 169 person scoring the behaviors did so blind to which group was being scored. The male 170 behaviors analyzed were courtship latency, orientation, wing vibration, wing scissor, 171 tapping, thrust, copulation attempt and copulation [19]. Orientation was defined as when 172 the male oriented at any direction close to the female's body and typically done for the 173 entirety for courtship. The start of orientation signaled the start of courtship, and 174 therefore the start of when the divider was removed allowing a male to see a female to 175 when orientation began was considered the latency to courtship. Wing vibration was 176 when the male's wings vibrated horizontally and vertically at different angles. This 177 behavior is known to create the "courtship song" which is important in the courtship 178 process of Drosophila [21]. Wing scissor was when a fly's wings were extended, crossed 179 and re-crossed. Tapping was when the male touched any female body part, usually 180 sideways, with its tarsus. Thrusting, which occurs during copulation and copulation 181 attempts, was when the male grasped the female's abdomen with his foretarsi, curled his 182 abdomen downward and forward, making a fast contact with the female's genitalia. 183 Copulation occurred when a male stabilized on top of the female for a considerable 184 amount of time, typically 10 -15 minutes. A copulation attempt was counted when the 185 male grasped the female's abdomen and maintained his abdomen in contact with the 186 female's genitalia for more than one second (i.e. longer than a thrust), but did not 187 complete the copulation (i.e. did not stabilize for an extended copulation period of time). 188

189 Triglyceride, Glycogen and Protein Measurements

190 Triglyceride, glycogen and protein levels were measured as previously described [22]. 191 Briefly, 2 male flies were homogenized in lysis buffer (140 mM NaCl, 50 mM Tris-HCl, 192 pH 7.4, 0.1% Triton-X 100, and 1X protease inhibitor cocktail (Roche Life Sciences) by 193 sonication. Thus, one "sample" consisted of the macromolecule levels measured in two 194 flies with WT N = 16, adp^{60} mutant N = 17, high fat diet N = 21, food deprived N = 17. 195 Triglycerides and protein were measured using the Triglycerides Liquicolor Test 196 (Stanbio) and the BCA Protein Assav Kit (ThermoScientific), respectively, according to 197 manufacturer's instructions. Total glucose levels were measured using the Glucose 198 Oxidase Reagent (Pointe Scientific) after treating samples with 8 mg/mL 199 amyloglucosidase (Sigma) in 0.2M citrate buffer, pH 5.0 for 2 hours. Free glucose was 200 measured in samples not treated with amyloglucosidase and glycogen levels were 201 determined by subtracting free glucose from total glucose. Triglycerides and glycogen 202 were normalized by dividing each by the total protein concentration of each sample. 203

204 Statistical Analysis

All behaviors (except orientation latency) were corrected for the length of an individual's courtship period (i.e. orientation period) and then compared statistically. The number of wing scissors, number of wing vibrations, and duration of copulation conformed to parametric assumptions and were analyzed via ANOVA. Total courtship time, courtship latency, wing scissor duration rate, wing vibration duration rate, and total number of thrusts were ln transformed to fit parametric assumptions and analyzed via ANOVA. An overall significant ANOVA test was followed by Tukey tests to identify which groups in

212	the overall test were significantly different. Number of tapping behaviors, rate of tapping
213	behavior, and rate of thrusting could not be transformed to fit parametric assumptions and
214	so were analyzed via Kruskal Wallis. No Kruskal Wallis test was significant thus no
215	multiple comparison testing was necessary for non-parametric analyses. Copulation
216	frequency between groups was tested via a Pearson Chi-Square test followed up with a
217	post hoc test using the residuals if the overall was found to be significant using a
218	Bonferroni adjustment for multiple comparisons and thus an alpha level of 0.01. Energy
219	data (glycogen, triglyceride and protein content) were analyzed via ANOVA as these data
220	met all parametric assumptions. Significant overall effects were followed by Tukey
221	multiple comparisons.

222

223 **Results**

224 Copulatory and Courtship Behavior

225 There was a difference in copulation frequency across groups $(X^2_{3, N=252} =$

42.395, P < 0.001; Fig 1). Post hoc tests revealed that the adp^{60} mutants had a

significantly lower frequency of copulations (13 / 75, 17.3%) relative to the other three

groups (P = 0.001), while the high fat fed flies had a significantly greater proportion of

copulations (24/33, 72.7%) than the other three groups (P < 0.001). Food deprived and

wild type flies showed a similar proportion of copulations (food deprived: 20/30, 66.7%,

231 P = 0.02; wild type: 62/114, 54.4%, P = 0.03).

232

Fig 1. Percentage of successful copulations. This figure shows the percentage of

successful copulations of food deprived, high fat fed, adp^{60} , and normal fed flies.

Numbers in parentheses represent number of trials run in each group. Different letters indicate frequencies that were significantly different at P < .01 (based on Bonferroni adjustment; see text for details).

238

239 Total courtship time among all four groups did not significantly differ (F $_{3,115}$ =

240 0.789, P = 0.50). However, latency to court did significantly differ (F $_{3,115}$ = 13.307, P <

241 0.001; Fig 2) with food deprived flies showing a significantly shorter latency time than

either high fat fed flies (P = 0.036) or wild type flies (P < 0.001), but not relative to adp^{60}

243 mutants (P = 0.378). There were no significant differences in latency to court among high

fat fed, wild type, and adp^{60} mutant groups (P > 0.1 in all instances).

245

Fig 2. Courtship latencies. This figure shows the number of seconds required for flies in each group to engage in orientation behavior of food deprived, high fat fed, adp^{60} , and normal fed flies. Boxes indicate interquartile ranges (IQR); whiskers indicate 1.5x IQR; "X" indicates points outside 1.5 x IQR; dark circles indicate mean. Different letters indicate groups that were significantly different at P < 0.05.

251

The total number of courtship behaviors did not significantly differ among all groups for any behavior (wing scissors: F $_{3,115} = 0.522$, P = 0.66; wing vibrations: F $_{3,115} =$ 0.945, P = 0.42; thrusts: F $_{3,115} = 0.212$, P = 0.88; tapping: χ^2 (3) = 0.25, P = 0.96), yet the rate these behaviors were conducted during courtship did significantly differ for some behaviors. Rate of wing scissoring significantly differed across groups (F $_{3,115} = 10.711$, P < 0.001; Fig 3) with the *adp*⁶⁰ mutant exhibiting a significantly lower rate than all other

258	groups (vs. wild type: $P = 0.002$; vs. high fat flies: $P < 0.001$; and vs. food deprived flies:
259	P < 0.001). Food deprived flies also showed a significantly higher rate of wing scissoring
260	relative to wild type flies ($P = 0.013$), but was not different relative to high fat fed flies (P
261	= 0.551). The rate of wing scissoring for wild types did not significantly differ from high
262	fat fed flies ($P = 0.357$). Wing vibrations showed the same differences across groups as
263	did wing scissoring. Specifically, the rate of wing vibrations significantly differed across
264	groups (F $_{3,115}$ = 12.343, P < 0.001) with the <i>adp</i> ⁶⁰ mutant exhibiting a significantly lower
265	rate than all other groups ($P < 0.001$ for all comparisons). Food deprived flies showed a
266	significantly higher rate of wing vibration relative to wild type flies ($P = 0.006$), but was
267	not different relative to high fat fed flies ($P = 0.587$). Wild type and high fat fed flies
268	were again not different ($P = 0.199$). There was no significant effect on the rate of
269	tapping ($\chi^2(3) = 0.45$, P = 0.92) or rate of thrusting ($\chi^2(3) = 4.503$, P = 0.21) across all
270	groups.

271

272 Fig 3. Courtship behavior rate. The rate at which courtship behavior (wing scissor and 273 wing vibration) occurred for each group (vertical hatching = food deprived WT flies; light gray = high fat fed WT flies; dark gray = adp^{60} mutant flies; stipple = normal WT 274 275 fed). Rates are duration of behavior divided by duration of courtship (start of orientation 276 to start of copulation). Boxes indicate IQR; whiskers indicate data range; dark circles 277 indicate means. Different capital letters indicate groups that are significantly different for 278 wing scissor behavior at P < 0.05. Different lower case letters indicate groups that are 279 significantly different for wing vibration behavior at P < 0.05.

280

281 Triglyceride and Glycogen levels

282 To confirm the efficacy of the high fat diet and the 24-hour food deprivation as well as 283 the adp^{60} mutation, triglyceride and glycogen levels were measured in these groups and 284 compared to wild type flies fed normal food. Consistent with previous reports, there was 285 a significant overall effect of treatment on the triglyceride per protein ratio (F $_{3.67}$ = 286 173.293, P < 0.001; Fig 4A) with wild type flies showing a lower ratio than adp^{60} 287 mutants (P < 0.001) and high fat fed flies (P = 0.023), but a higher ratio compared to food 288 deprived flies (P < 0.001) [16], [17], [23], [24]. Food deprived flies also showed a lower 289 ratio compared to adp^{60} mutants (P < 0.001) and high fat fed flies (P < 0.001). High fat 290 fed flies showed a lower ratio compared to adp^{60} mutants (P < 0.001). There was also a significant effect of fly/diet type on the glycogen per protein ratio (F $_{3,67}$ = 38.524, P = 291 292 0.01; Fig 4B) with wild type flies showing a lower glycogen to protein ratio than adp^{60} 293 mutants (P < 0.001), a higher level than food deprived flies (P = 0.002) similar to that 294 shown in [24] and no difference than the high fat diet fed group (P = 0.874). Food deprived flies also showed a lower glycogen to protein ratio compared to *adp*⁶⁰ mutants 295 296 (P < 0.001) and high fat fed flies (P < 0.001). High fat fed flies showed a lower ratio 297 compared to adp^{60} flies (P < 0.001).

298

Fig 4. Fat and sugar per protein of flies A) Triglyceride content per total body protein for food deprived, high fat fed, adp^{60} , and normal fed flies. Different capital letters indicate significant differences at P < 0.05. B) Glycogen content per total body protein for food deprived, high fat fed, adp^{60} , and normal fed flies. See Figure 2 for box and whisker details. Different lower case letters indicate significant differences at P < 0.05.

304

305 **Discussion**

306 Availability of energy, both in terms of availability of nutrients and lipid and glycogen 307 storage affected reproductive behaviors in *D. melanogaster* similarly. Contrary to our 308 prediction, food deprived wild type flies had a decreased latency to court when compared 309 to all other fly groups indicating that limiting caloric intake actually increases appetitive 310 reproductive behavior. In addition, once courtship started, food deprived flies showed the 311 greatest rate of courtship in terms of wing scissoring and wing vibrations which further 312 supports this notion. However, this increased drive to reproduce did not result in more 313 copulations. In fact, the high fat fed group enjoyed a significantly greater copulation 314 frequency than all other groups, with the food deprived and normal fed flies showing 315 similar copulation frequencies and adp^{60} showing the least. Thus, while food deprived 316 flies showed a greater drive and effort to reproduce, the higher copulation rate by high fat 317 fed flies could indicate that flies fed diets high in fat are more attractive. This suggests 318 that while energy availability influences male reproductive drive, other aspects of male 319 appearance or courtship quality (two aspects known to influence female choice [25]) 320 interact to impact reproductive success.

We suspect that the higher drive to reproduce shown by food deprived flies is due to the limited availability of metabolic resources, which could indicate a shortened time to live and thus a greater immediate investment in reproduction. Previous work has shown that food deprived flies and flies reared on highly nutrient restricted diets show decreased longevity compared to those raised on normal diets [26]. Flies who live shorter periods of time are also known to show increased drive to reproduce. For example, in *D*.

327 *nigrospiracula*, male flies infected with parasites lived shorter lives but dedicated 328 significantly more time to courting females than uninfected males [27]. Curiously, the 329 courtship latency among normal fed wild type flies, high fat fed wild type flies and adp⁶⁰ 330 mutants were all similar despite adp⁶⁰ mutants storing excess nutrients compared to wild 331 type flies. However, courtship latency is indicative of courtship drive, and it is known 332 that the lifespan of adp^{60} homozygous males is similar to that of wild type males [28]. In 333 addition, adp^{60} mutants are starvation resistant [23]. Thus, adp^{60} mutants do not appear to 334 have as compromised reproductive potential based on survival as that of food deprived 335 wild type flies.

336 Though longevity can be related to reproductive drive in species such as 337 Drosophila, it remains unclear what is the direct cue to this increased drive – the actual 338 intake of energy-rich molecules and/or the physiological availability of these metabolic 339 reserves. Food deprived flies, who showed the shortest latency to court along with the 340 highest rates of courtship behaviors (i.e. wing scissoring and wing vibrations), had the 341 lowest food availability, and the lowest storage of both glycogen and triglycerides per body protein content. However, the adp^{60} mutants, who had a constant availability of 342 343 food but do not metabolize their triglycerides and glycogen in the same way as wild type 344 flies (thus leading to an increased amount of each in our analysis) showed significantly 345 less wing scissoring and wing vibrations. When considering these mutants show a similar 346 reproductive drive to normal and high fat fed flies, potentially the availability of nutrients 347 is involved with reproductive drive and courtship latency, while the appropriate storage 348 and usage of these sources of energy once ingested is involved in carrying out courtship 349 song caused by the wing vibration behavior. Perhaps combining diet alteration and the

 adp^{60} mutation may allow the determination of whether the availability of nutrients or normal storage and usage of these nutrients is important for regulating courtship behaviors. However adp^{60} mutants are known to have a "lethargic" phenotype where their overall mobility is less than that of wild type flies [23]. This phenotype may be due to this altered ability to metabolize nutrients and/or the gross obesity of these animals and could be related to the decreased wing scissoring and wing vibrations, but it did not affect the latency to court.

357 An important point to note is that this process could work differently in species 358 with a different natural history. For example, it is well known that when physiological 359 fuels are low in mammals, the reproduction cycle stops until such time as there is enough 360 energy to put into reproduction [7]. This again is likely a function of longevity where 361 such species can afford to wait for the proper resources. In mammals, it is well 362 established that there are related, but independent mechanisms that link reproduction to 363 food intake and physiological energy availability [6]. For example, kissipeptin, which is 364 known to regulate GnRH and the reproductive axis, is influenced by ghrelin and PYY 365 secretion (gastrointestinal hormones released upon ingestion) as well as by nutritional 366 status [29]. Less is known about such pathways in *D. melanogaster*; however, there is 367 some evidence to support that reproductive behavior is related to food availability (i.e. 368 intake) and energy storage. For example, altering the amounts of protein and 369 carbohydrates in the female diet will affect the rate of egg production in wild type D. 370 *melanogaster* [30]. Schultzhaus [31] analyzed the effect of high fat diets on courtship in 371 D. melanogaster and found that while feeding female flies a high fat diet had more 372 negative effects on reproductive behavior than the same diet did on males, male judgment

373 of female attractiveness was influenced by the high fat feeding. Considering only the wild 374 type groups in the current study (food deprived, high fat fed, and normal fed), the nutrient 375 storage data suggest that the reserves of glycogen, and not triglycerides, may serve as a 376 cue for the change in sexual behavior we see in our study. Food deprived flies had lower 377 glycogen and triglycerides than all other groups while normal fed and high fat fed flies 378 courted similarly and had similar glycogen levels but different triglycerides. It follows 379 that animals assess short term energy stores, such as glycogen, to establish levels of 380 activity and not long term energy stores like triglycerides. We suspect that these limited 381 metabolic resources culminated in a greater immediate investment in reproduction. 382 It is unclear why the adp^{60} mutant contained the highest levels of glycogen and 383 triglycerides yet courted the least, but many possibilities exist, each of which could 384 contribute to and help explain the overall reduced mobility phenotype of the *adp*⁶⁰ mutant 385 addressed above. One possibility is the physical structure of increased fat deposits 386 decreases wing dexterity therefore affecting courtship song. Another is that the adp^{60} 387 mutation has additional unrealized effects on muscle activity. The adp^{60} mutation results 388 in chronic obesity with excess fat accumulation in both the larval and adult stages of 389 development [16], [18], [23]; Figure 4A). Potentially these mutants do not mobilize these 390 fuels in the same way as wild type flies as lipid metabolism in these mutants is suspected 391 to be different throughout development (see [32] for further discussion). Any negative 392 effect these metabolic pathways have on muscle function, in turn, could limit their ability 393 to function properly and affect courtship. Lastly, when these male courtship songs are 394 altered in this way, males might simply be choosing to decrease their song either because 395 of a female's lack of interest, or they are responding to their own song. While we cannot

address which of these mechanisms is at play, they would all lead to reduced song and/or low quality song that could lead to an increased rejection by the female fly. In fact, the lack of courtship song from the adp^{60} mutants could help explain the low copulation rates of this group via females "allowing" a copulatory grasp as various *Drosophila* species are known to reject males on the basis of song [33], [34].

401 While the lack of courtship song via wing vibrations could explain the

402 significantly lower percentage of successful copulations for *adp*⁶⁰ mutants, food deprived

403 flies did show the most wing vibrations, yet it was high fat fed flies who had the most

404 copulations. Possibly females enable copulations based on multiple signals with courtship

405 song being only one. Size of the male could be another (see [25] for further discussion],

406 as well as number or quality of the cuticular hydrocarbons, which are known chemical

407 communicators between males and females and can be affected by nutritional regime

408 (reviewed in [35]). While high fat diets are known to not alter male cuticular

409 hydrocarbons [31], the effect of *adp*⁶⁰ and food deprivation is not known. Larger males

410 may generally indicate better quality sperm based on nutrition; however, Partridge et al.

411 [25] also argued that larger males are more active and generally more persistent, so the

412 increased copulation seen in the current study from the high fat fed group may be due to a

413 form of male coercion.

414

415 1. Conclusions

416 Overall, we find that altering nutrient storage through diet or genetic manipulation has

417 effects on courtship in the fruit fly, *D. melanogaster*. The increased drive to reproduce by

418 food deprived flies was unexpected, but is reasonable based on species' reproductive

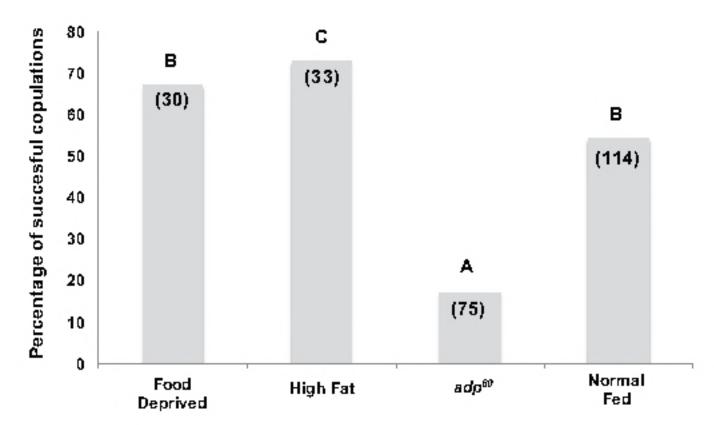
419	investment and longevity. We suspect that such findings would be different in species
420	with a different natural history, such as those with a greater longevity where one can
421	afford to wait for the proper resources to become available. More research is needed to
422	identify the exact physiological cue that activates reproductive behavior across species
423	with such varied life histories to better understand reproductive motivation in the context
424	of natural history. We believe utilizing a combination of manipulation of animal diets and
425	the use of mutants to manipulate physiological pathways can help illuminate the resource
426	mechanisms that drive the reproduction axis. Species such as D. melanogaster are
427	particularly powerful in this regard as such mutants exist both naturally, such as the adp^{60}
428	mutants used here, and can be artificially created to probe the effects of specific genes
429	and metabolic processes on organismal reproduction.
430	
431	2. Acknowledgements
432	We would like to thank Grzegorz Polak and Justin Palermo for help with fly husbandry.
433	This research did not receive any specific grant from funding agencies in the public,
434	commercial, or not-for-profit sectors.
435	
436	References:
437 438 439 440 441 442	 Kohsaka A, Laposky AD, Ramsey KM, Estrada C, Joshu C, Kobayashi Y, et al. High- fat diet disrupts behavioral and molecular circadian rhythms in mice. Cell Metab. 2007;6:414–421. doi: 10.1016/j.cmet.2007.09.006 Lim RS, Eyjólfsdóttir E, Shin E, Perona P, Anderson DJ. How food controls aggression in Drosophila. PloS One 2014;9:e105626. doi:10.1371/journal.pone.0105626
443	3. Redman LM, Heilbronn LK, Martin CK, De Jonge L, Williamson DA, Delany JP, et

3. Redman LM, Heilbronn LK, Martin CK, De Jonge L, Williamson DA, Delany JP, et
al. Metabolic and behavioral compensations in response to caloric restriction:
implications for the maintenance of weight loss. PloS One 2009;4:e4377. doi:
10.1371/journal.pone.0004377

447	4. Bronson F. Puberty and energy reserves: a walk on the wild side. In: Wallen, K.,
448	Schneider JE, editors. Reproduction in Context: Social and Environmental
449	Influences on Reproduction. Cambridge, Massachusetts; London, England: MIT
450	Press; 2000. p.15–33.
451	5. Gittleman JL, Thompson SD. Energy allocation in mammalian reproduction. Am.
452	Zool. 1988;28:863-875. doi: 10.1093/icb/28.3.863
453	6. Schneider JE. Energy balance and reproduction. Physiol. Behav. 2004; 81:289–317.
454	doi: 10.1016/j.physbeh.2004.02.007
455	7. Schneider JE, Wade GN. Inhibition of reproduction in service of energy budget. In:
456	Wallen, K., Schneider JE, editors. Reproduction in Context: Social and
457	Environmental Influences on Reproduction. Cambridge, Massachusetts; London,
458	England: MIT Press; 2000 p.15–33.
459	8. Martin TE. Food as a limit on breeding birds: a life-history perspective. Annu. Rev.
460	Ecol. Syst. 1987;18:453–487.
461	9. Sibly RM, Witt CC, Wright NA, Venditti C, Jetz W, Brown JH. Energetics, lifestyle,
462	and reproduction in birds. Proc. Natl. Acad. Sci. 2012;109:10937-10941. doi:
463	10.1073/pnas.1206512109
464	10. Lind CM, Beaupre SJ. Male Snakes Allocate Time and Energy according to
465	Individual Energetic Status: Body Condition, Steroid Hormones, and
466	Reproductive Behavior in Timber Rattlesnakes, Crotalus horridus. Physiol.
467	Biochem. Zool. 2015;88:624–633. doi: 10.1086/683058
468	11. Van Dyke JU, Griffith OW, Thompson MB. High Food Abundance Permits the
469	Evolution of Placentotrophy: Evidence from a Placental Lizard, Pseudemoia
470	entrecasteauxii. Am. Nat. 2014;184:198-210. doi: 10.1086/677138
471	12. Ottinger MA, Mobarak M, Abdelnabi M, Roth G, Proudman J, Ingram DK. Effects of
472	calorie restriction on reproductive and adrenal systems in Japanese quail: are
473	responses similar to mammals, particularly primates? Mech. Ageing Dev. 2005;
474	126:967–975. doi: 10.1016/j.mad.2005.03.017
475	13. Cheng D, Chen L, Yi C, Liang G, Xu Y. Association between changes in
476	reproductive activity and D-glucose metabolism in the tephritid fruit fly,
477	Bactrocera dorsalis (Hendel). Sci. Rep. 2014; 4(7489):1-9.14. doi:
478	10.1038/srep07489
479	14. Warburg MS, Yuval B. Effects of energetic reserves on behavioral patterns of
480	Mediterranean fruit flies (Diptera: Tephritidae). Oecologia 1997;112:314-319.
481	doi: 10.1007/s004420050314
482	15. Baker KD, Thummel CS. Diabetic larvae and obese flies—emerging studies of
483	metabolism in Drosophila. Cell Metab. 2007;6:257–266.
484	16. Häder T, Müller S, Aguilera M, Eulenberg KG, Steuernagel A, Ciossek T, et al.
485	Control of triglyceride storage by a WD40/TPR-domain protein. EMBO Rep.
486	2003;4: 511–516. doi: 10.1038/sj.embor.embor837
487	17. Birse RT, Choi J, Reardon K, Rodriguez J, Graham S, Diop S, et al. High-fat-diet-
488	induced obesity and heart dysfunction are regulated by the TOR pathway in
489	Drosophila. Cell Metab. 2010;12:533–544. doi: 10.1016/j.cmet.2010.09.014
490	18. Reis T, Van Glist MR, Hariharan IK. A buoyancy-based screen of Drosophila
491	larvae for fat-storage mutants reveals a role for Sir2 in coupling fat storage to

492	nutrient availability. PloS Genet. 2010; 6(11): e1001206. doi:
493	10.1371/journal.pgen.1001206.
494	19. Cobb M, Connolly K, Burnet B. Courtship behaviour in the melanogaster species
495	sub-group of Drosophila. Behaviour. 1985; 95:203–230. doi:
496	10.1163/156853985X00136
497	20. Kauffman, A.S., Rissman, E.F., 2004. A critical role for the evolutionarily conserved
498 499	gonadotropin-releasing hormone II: mediation of energy status and female sexual behavior. Endocrinology 145, 3639–3646. doi: 10.1210/en.2004-0148
500	21. Ewing AW, Bennet-Clark H. The courtship songs of Drosophila. Behaviour 1968; 31:
501	288–301. doi: 10.1163/156853968X00298
502	22. Gingras RM, Warren ME, Nagengast AA, DiAngelo JR. The control of lipid
503 504	metabolism by mRNA splicing in Drosophila. Biochem. Biophys. Res. Commun. 2014;443: 672–676. doi: 10.1016/j.bbrc.2013.12.027
505	23. Suh JM, Zeve D, McKay R, Seo J, Salo Z, Li R, et al. Adipose is a conserved dosage-
506	sensitive antiobesity gene. Cell Metab. 2007;6:195–207.
507	24. Schwasinger-Schmidt TE, Kachman SD, Harshman LG. Evolution of
508	starvation resistance in Drosophila melanogaster: measurement of direct and
509	correlated responses to artificial selection. J. Evol. Biol. 2012;25:378-387. doi:
510	10.1111/j.1420-9101.2011.02428.x
511	25. Partridge L, Ewing A, Chandler A. Male size and mating success in Drosophila
512	melanogaster: the roles of male and female behaviour. Anim. Behav. 1987;35:
513	555–562. doi: 10.1016/S0003-3472(87)80281-6
514	26. Magwere T, Chapman T, Partridge L. Sex differences in the effect of dietary
515	restriction on life span and mortality rates in female and male Drosophila
516	melanogaster. J. Gerontol. A. Biol. Sci. Med. Sci. 2004;59:B3-B9. doi:
517	10.1093/gerona/59.1.B3
518	27. Polak M, Starmer WT. Parasite-induced risk of mortality elevates reproductive effort
519	in male Drosophila. Proc. R. Soc. Lond. B Biol. Sci. 1998;265:2197-2201. doi:
520	10.1098/rspb.1998.0559
521	28. Doane WW. Developmental physiology of the mutant female sterile (2) adipose of
522	Drosophila melanogaster. I. Adult morphology, longevity, egg production, and
523	egg lethality. J. Exp. Zool. Part Ecol. Genet. Physiol. 1960;145:1–21.doi:
524	10.1002/jez.1401450102
525	29. Fernandez-Fernandez R, Martini A, Navarro V, Castellano J, Dieguez C, Aguilar E,
526	et al. Novel signals for the integration of energy balance and reproduction. Mol.
527	Cell. Endocrinol. 2006; 254: 127–132.
528	30. Lee KP, Simpson SJ, Clissold FJ, Brooks R, Ballard JWO, Taylor PW, et al. Lifespan
529	and reproduction in Drosophila: new insights from nutritional geometry. Proc.
530	Natl. Acad. Sci. 2008;105:2498–2503. doi: 10.1073/pnas.0710787105
531	31. Schultzhaus JN, Bennett CJ, Iftikhar H, Yew JY, Mallett J, Carney GE. High fat diet
532	alters Drosophilia melanogaster sexual behavior and traits: decreased
533	attractiveness and changes in pheromone profiles. Sci. Rep. 2018; 8:5387.
534	doi:10.1038/s41598-018-23662-2
535	32. Teague BD, Clark AG, Doane WW. Developmental analysis of lipids from wild-type
536	and adipose60 mutants of Drosophila melanogaster. J. Exp. Zool. Part Ecol.
537	Genet. Physiol. 1986;240:95–104.

- 33. Hoikkala A, Aspi J, Suvanto L. Male courtship song frequency as an indicator of male genetic quality in an insect species, Drosophila montana. Proc. R. Soc.
 - Lond. B Biol. Sci. 1998;265:503-508. Doi: 10.1098/rspb.1998.0323
- 34. Ritchie MG, Saarikettu M, Livingstone S, Hoikkala A. Characterization of female
- preference functions for Drosophila montana courtship song and a test of the temperature coupling hypothesis. Evolution 2001;55:721-727. doi: 10.1554/0014-
- 3820(2001)055[0721:COFPFF]2.0.CO;2
- 35. Ferveur JF. Cuticular hydrocarbons: their evolution and roles in Drosophila pheromonal communication. Behav. Genet. 2005;35:279-295. doi: 10.1007/s10519-005-3220-5



bioRxiv preprint doi: https://doi.org/10.1101/331660; this version posted May 25, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under a CC-BY 4.0 International license.

