

1 **Genome sequence and description of *Blautia brookingsii* str SG772 nov., a new species of**
2 **anaerobic bacterium isolated from healthy human gut.**

3

4 Sudeep Ghimire^{1,2}, Roshan Kumar^{1,2}, Eric Nelson^{1,2}, Jane Christopher-Hennings^{1,2}, and Joy
5 Scaria^{1,2*}

6

7

8 *Corresponding Author

9 Email: joy.scaria@sdstate.edu

10

11

12

13

14

15

16

17 ¹Department of Veterinary and Biomedical Sciences, South Dakota State University, Brookings,
18 SD, USA.

19 ²South Dakota Center for Biologics Research and Commercialization, SD, USA.

20

21 **Abstract**

22 Strain SG-772 is a Gram positive, strictly anoxic bacterium isolated from the feces of a healthy
23 human fecal donor. Based on 16S rRNA gene sequence, the strain showed maximum similarity
24 (94.39%) with *Blautia stercoris* GAM6-1 in EZ-Taxon server and thus assigned the genus
25 *Blautia*. The scanning electron micrograph of the bacterium revealed the characteristic
26 coccobacillus shape as well as the complete absence of flagellum, suggesting its non-motile
27 phenotype. This strain was found to utilize 27 substrates based on Biolog AN plates assay, with
28 maximum preference for D-mannitol. Additionally, the strain was found to be resistant to
29 tetracycline and streptomycin. Genome sequencing and analysis revealed an overall genome size
30 of 3.49 Mbp and GC content of 43.97%. Based on RAST annotation server, the closest neighbor
31 was *Blautia hansenii* DSM20583. Average Nucleotide Identity (ANI) of these strains were
32 81.69%, suggesting a high level of genomic variation. The comparative genome analysis of
33 strain SG772 with *B. hansenii* DSM20583 revealed a total of 411 orthologous genes coding for
34 basic metabolic functions. Furthermore, the genomes were functionally distinct based on COG
35 categories. Thus, based on all these differences, we propose a novel species of genus *Blautia*
36 named as *Blautia brookingsii* SG772.

37

38

39

40

41 **Keywords:** *Blautia*, *brookingsii* SG772, Human gut, Genome, Anaerobe

42

43 **Introduction**

44 The members of family *Lachnospiraceae* are commensal bacteria inhabiting the human
45 gastrointestinal tract (1). These group of bacteria degrade complex polysaccharides to produce
46 short chain fatty acids. Acetate, butyrate and propionate produced as the result of digestion of
47 polysaccharides can be used as an energy source for the host (2). *Lachnospiraceae* in human gut
48 microbiota plays important role in maintaining health and disease balance (3-7). Recently, a new
49 genus *Blautia* has been formed under *Lachnospiraceae* family by reclassifying few members of
50 genus *Clostridium* and *Ruminococcus* (8). *Blautia* has been reported to provide colonization
51 resistance against *Enterococcus faecium* (9) and *Clostridium difficile* (5).

52 The members of genus *Blautia* are gram- positive, non-motile, coccoid shaped; obligate
53 anaerobes which can produce a wide variety of compounds as the by-product of fermentation.
54 Most of the members of this genus were either isolated from the human or mammalian fecal
55 samples (8, 10). During the culturomics of human feces, a novel *Blautia*-like strain was isolated
56 from Brain Heart Infusion (BHI) based medium and named as strain SG772. Based on 16S
57 rRNA gene sequence phylogeny, it was taxonomically clustered with the genus *Blautia*.
58 Additionally, using taxo-genomics approach, the bacterium SG772 showed distinct
59 morphological, physiological and genomic attribute as compared to its closest neighbor. Based
60 on all these differences, we propose a novel species of genus *Blautia* named as *Blautia*
61 *brookingsii* SG772.

62 **Organism information**

63 **Growth conditions and biochemical properties**

64 During culturomics of healthy human fecal sample, strain SG772 was isolated from a healthy
65 human fecal sample using Brain Heart Infusion (BHI) agar medium anaerobically at 37⁰C.
66 Morphologically, colonies were round, whitish, convex and smooth after 48 hours incubation on
67 BHI agar. Further, cellular morphology of strain SG772 was examined using a scanning electron
68 microscope. Microscopic study revealed the complete absence of flagellum, indicating that strain
69 SG772 is non-motile. The size of the bacterium varied between 0.5-0.8 x 1.8-2.5 µm (Figure 1).
70 The Gram-stain test was performed using a Gram staining kit (HiMedia) and the strain was
71 found to be Gram-positive. Optimum temperature and pH for growth of the bacteria was found to
72 be 37⁰C and 6.8 respectively.

73 As the strain SG772 exhibited no significant scores on Matrix-assisted desorption-ionization
74 time-of-flight (MALDI-ToF) (Bruker Daltonics, Germany) (11), the genes encoding for 16S
75 rRNA gene was amplified, sequenced using Applied Biosystems 3500xL, Genetic analyser
76 (Applied Biosystems, MA, USA) at Department of Veterinary and Biomedical Sciences, South
77 Dakota State University. The sequence was assembled using Sequencing Analysis version 5.1.1
78 Genious 10.2.3 (NJ, US). A continuous stretch of 1440 bp of the 16S rRNA gene of strain
79 SG772 was obtained and this sequence was subjected to similarity search against EzTaxon- e-
80 server (12). Analysis of 16S rRNA gene revealed highest sequence similarity (94.39%) to
81 *Blautia stercoris* GAM6-1. Thereafter, the phylogenetic tree was constructed based on 16S
82 rRNA gene sequence, together with top 30 taxonomically characterized strains. The
83 evolutionary distance matrix was calculated using the distance model of Jukes & Cantor (1969)
84 and an evolutionary tree was reconstructed using the neighbor joining method. The sequence of
85 *Atobobium minutum* NCFB2751 was used as an outgroup (Figure 2). The resultant tree topology
86 was evaluated by bootstrap analysis based on 1000 replicates (Felesenstein J, 1985) in Mega

87 version 5.2.2. Strain SG772 falls in the clade containing members belonging to the genus
88 *Blautia*. (Figure. 2)

89 The growth pattern for the strain SG772 was determined using growth curve assay (Figure 3A).
90 The biphasic growth pattern indicate its ability to use simple sugars in media initially with
91 utilization of complex carbohydrates when simple sugars are depleted. Additionally, in order to
92 elucidate the pattern of substrate oxidation, Biolog assay was performed using Biolog AN
93 MicroPlate™ (Biolog Catalog # 70007) anaerobically. For this, 100 µl of overnight grown
94 culture was plated onto 150 mm BHI agar plate and incubated for 48 hours anaerobically at 37⁰
95 C. Grown colonies were picked by a sterile cotton swab and inoculated to AN inoculating fluid
96 (Biolog Catalog # 72007) until OD₆₅₀ reached 0.3. From this suspension, 100µl was pipetted into
97 each well of 96 well biolog AN microplate in triplicate and incubated at 37⁰C anaerobically.
98 OD₆₅₀ readings were taken at 0 hour and 24 hours post inoculation and results were analyzed and
99 compared against *Blautia stercoris* GAM6-1 (Table 1). The strain was found to utilize 27
100 different substrates with maximum preference for D-mannitol (Figure 3B).

101 Antimicrobial susceptibility of the strain was tested using agar diffusion method in BHI agar
102 medium. One hundred microliters of overnight grown culture of the strain was plated in BHI
103 agar medium and antibiotic discs were placed. Zone of inhibition by antibiotics was measured
104 after 24 hours of anaerobic incubation to determine the antibiotic sensitivity of the strain. The
105 strain was found to be resistant to tetracycline and streptomycin. In contrast, the strain was
106 susceptible to chloramphenicol, ampicillin, erythromycin and novobiocin.

107 **Genome sequencing information**

108 **DNA isolation**

109 Strain SG772 was sub-cultured on BHI agar and incubated at 37⁰C anaerobically for 48 hours.
110 Further, it was cultured in 5 ml of BHI broth for 24 hours. Isolation of DNA was performed
111 following E.Z.N.A DNA isolation kit (Omega, Biotek) protocol. Initially, 500µl of bacterial
112 broth was pelleted by centrifuging at 10,000×g for 1 min. The pellet obtained was lysed in tris
113 EDTA buffer using lysozyme for 1 hour and protein was digested using proteinase K overnight
114 at 55⁰C. Subsequently, the mixture was pelleted by centrifuging at 8,000×g for 1 min and
115 supernatant was spun in spin column followed by two washes of wash buffer. Finally, DNA was
116 eluted using 50 µl of nuclease free water and kept at -20⁰C until use.

117 **Genome sequencing, assembly and annotation**

118 The genome was sequenced using Illumina MiSeq (Illumina Inc, CA) using 2x 250 paired end
119 chemistry. Further, it was assembled using SPAdes 3.9.0 (13) and validated using Quast (14).
120 Coding sequences were predicted using Glimmer 3.0 (15) and annotated with RAST 2.0 server
121 (16) and the NCBI Prokaryotic Genomes Annotation Pipeline (PGAP) version 4.4. For core
122 genome analysis, GenBank files were retrieved for strains SG772 along with its RAST neighbor
123 *Blautia hansenii* DSM 20583 and were used for core genome analysis using OrthoMCL
124 clustering algorithm (17) at 75% query coverage and sequence similarity (18). For functional
125 annotation, the amino acid sequences were searched against the COG database to predict the
126 abundance of COG gene families and heat map was constructed using Pearson correlation
127 method and hierarchical clustering.

128 **Genomic attributes**

129 Assembly of the *B. brookingsii* SG772 genome produced 55 contigs with the genome size of
130 3.46 Mbp (N50 = 179,176) and 43.97% GC content (Table 2). The largest contig was of 401,827

131 bp while the smallest was of 624 bps. The genome contains 57 tRNA genes, 3343 coding
132 sequences, a phage element and a CRISPR element (Table 2).

133 **Insights from genome sequence**

134 **Insight into the genetic repertoire**

135 In order to compare the newly sequenced genome of strain SG772 to its closest RAST neighbor
136 *B. hansenii* DSM 20583 (8), we used BLAST Ring Image Generator (BRIG) (19) as a tool to
137 show genome wide sequence similarity using *B. hansenii* DSM 20583 as a reference genome
138 (Figure 4). Based on average nucleotide identity, these two genomes were 81.69% identical (19).
139 Furthermore, this comparison revealed several differences in genome size, GC content and RNA
140 copies (Table 2). For functional annotation, the amino acid sequences were searched against the
141 COG database to predict the abundance of COG gene families and were compared against *B.*
142 *hansenii* DSM 20583 (21). The gene families for energy production and conversion (C), amino
143 acid transport and metabolism (E), carbohydrate transport and metabolism (G), transcription,
144 ribosomal structure and biogenesis (J), transcription (K), replication, recombination and repair
145 (L), cell wall/membrane/envelope biogenesis (M), inorganic ion transport and metabolism (P),
146 defense mechanism (V) and unknown functions (S) were differentially enriched among these two
147 strains (Figure 3C). Core genome analysis revealed a total of 411 core genes coding for basic
148 metabolic functions. A total of 4,880 COG functions were annotated out of which only 70%
149 were assigned to known categories. Additionally, resistance to cadmium, tetracycline,
150 vancomycin, beta lactamase and fluoroquinolones along with dormancy and sporulation genes
151 were relatively abundant in strain SG772. Furthermore, the genome was devoid of genes for
152 motility and chemotaxis suggesting its non-motile phenotype.

153 **Conclusions**

154 This study presents the genome sequence for the strain SG772, with marked physiological and
155 genomic differences from its neighbors *i.e.*, *B. stercoris* GAM6-1 and *B. hansenii* DSM20583.
156 Based on the differences, we propose a novel species of genus *Blautia* named as *Blautia*
157 *brookingsii* SG772.

158 **Taxonomic and nomenclatural proposals**

159 *Blautia brookingsii* (*brookingsii* referring to the isolation site of the type strain from Brookings,
160 SD, USA). The cells are Gram stain positive, non-motile, coccobacillus, and 1.8-2.5 μm in
161 length and 0.5-0.8 μm in diameter. Colonies were whitish, circular, smooth, and convex after 48
162 h of incubation in BHI agar. Optimal growth was observed at 37⁰C and pH of 6.8. It assimilates
163 D-Mannitol, D-Melezitose, Adonitol, D-Glucosaminic acid, D-Arabitol, D- Sorbitol, D-
164 Trehalose etc. Furthermore, the strain was resistant to tetracycline and streptomycin antibiotics
165 whereas, it was susceptible to chloramphenicol, erythromycin, novobiocin and ampicillin.
166 Genome sequence revealed the genome size of the strain SG772 to be 3.46Mbp with G+C
167 content of 43.97 %. The type strain SG772 isolated from hlthy human fecal sample collected at
168 Brookings, SD, USA and has been deposited in the Microbial Culture Collection at the National
169 Centre for Cell Science, India.

170 **Declarations**

171 Authors declare no competing interests.

172 **Acknowledgements**

173 This work was supported in part by the USDA National Institute of Food and Agriculture, Hatch
174 projects SD00H532-14 and SD00R540-15 to JS, and a grant from the South Dakota Governor's
175 Office of Economic Development awarded to JS, EN and JH.

176 **References**

- 177 1. McLellan SL, Newton RJ, Vandewalle JL, Shanks OC, Huse SM, Eren AM, Sogin ML. 2013. Sewage
178 reflects the distribution of human faecal Lachnospiraceae. *Environ Microbiol* 15:2213-27.
- 179 2. Biddle A, Stewart L, Blanchard J, Leschine S. 2013. Untangling the Genetic Basis of Fibrolytic
180 Specialization by Lachnospiraceae and Ruminococcaceae in Diverse Gut Communities. *Diversity*
181 5:627-640.
- 182 3. Young VB. 2017. The role of the microbiome in human health and disease: an introduction for
183 clinicians. *BMJ* 356:j831.
- 184 4. Million M, Diallo A, Raoult D. 2017. Gut microbiota and malnutrition. *Microb Pathog* 106:127-
185 138.
- 186 5. Takahashi K, Nishida A, Fujimoto T, Fujii M, Shioya M, Imaeda H, Inatomi O, Bamba S, Sugimoto
187 M, Andoh A. 2016. Reduced Abundance of Butyrate-Producing Bacteria Species in the Fecal
188 Microbial Community in Crohn's Disease. *Digestion* 93:59-65.
- 189 6. Reeves AE, Koenigsnecht MJ, Bergin IL, Young VB. 2012. Suppression of *Clostridium difficile* in
190 the gastrointestinal tracts of germfree mice inoculated with a murine isolate from the family
191 Lachnospiraceae. *Infect Immun* 80:3786-94.
- 192 7. Kameyama K, Itoh K. 2014. Intestinal colonization by a Lachnospiraceae bacterium contributes
193 to the development of diabetes in obese mice. *Microbes Environ* 29:427-30.
- 194 8. Liu C, Finegold SM, Song Y, Lawson PA. 2008. Reclassification of *Clostridium coccoides*,
195 *Ruminococcus hansenii*, *Ruminococcus hydrogenotrophicus*, *Ruminococcus luti*, *Ruminococcus*
196 *productus* and *Ruminococcus schinkii* as *Blautia coccoides* gen. nov., comb. nov., *Blautia*
197 *hansenii* comb. nov., *Blautia hydrogenotrophica* comb. nov., *Blautia luti* comb. nov., *Blautia*
198 *producta* comb. nov., *Blautia schinkii* comb. nov. and description of *Blautia wexlerae* sp. nov.,
199 isolated from human faeces. *Int J Syst Evol Microbiol* 58:1896-902.
- 200 9. Caballero S, Kim S, Carter RA, Leiner IM, Susac B, Miller L, Kim GJ, Ling L, Pamer EG. 2017.
201 Cooperating Commensals Restore Colonization Resistance to Vancomycin-Resistant
202 *Enterococcus faecium*. *Cell Host Microbe* 21:592-602 e4.
- 203 10. Eren AM, Sogin ML, Morrison HG, Vineis JH, Fisher JC, Newton RJ, McLellan SL. 2015. A single
204 genus in the gut microbiome reflects host preference and specificity. *ISME J* 9:90-100.
- 205 11. Seng P, Abat C, Rolain JM, Colson P, Lagier JC, Gouriet F, Fournier PE, Drancourt M, La Scola B,
206 Raoult D. 2013. Identification of rare pathogenic bacteria in a clinical microbiology laboratory:
207 impact of matrix-assisted laser desorption ionization-time of flight mass spectrometry. *J Clin*
208 *Microbiol* 51:2182-94.
- 209 12. Chun J, Lee JH, Jung Y, Kim M, Kim S, Kim BK, Lim YW. 2007. EzTaxon: a web-based tool for the
210 identification of prokaryotes based on 16S ribosomal RNA gene sequences. *Int J Syst Evol*
211 *Microbiol* 57:2259-61.
- 212 13. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI,
213 Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA.

- 214 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J*
 215 *Comput Biol* 19:455-77.
- 216 14. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome
 217 assemblies. *Bioinformatics* 29:1072-5.
- 218 15. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and
 219 endosymbiont DNA with Glimmer. *Bioinformatics* 23:673-9.
- 220 16. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM,
 221 Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D,
 222 Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O.
 223 2008. The RAST Server: rapid annotations using subsystems technology. *BMC genomics* 9:75.
- 224 17. Li L, Stoeckert CJ, Jr., Roos DS. 2003. OrthoMCL: identification of ortholog groups for eukaryotic
 225 genomes. *Genome Res* 13:2178-89.
- 226 18. Contreras-Moreira B, Vinuesa P. 2013. GET_HOMOLOGUES, a versatile software package for
 227 scalable and robust microbial pangenome analysis. *Appl Environ Microbiol* 79:7696-701.
- 228 19. Konstantinidis KT, Tiedje JM. 2005. Genomic insights that advance the species definition for
 229 prokaryotes. *Proc Natl Acad Sci U S A* 102:2567-72.

230

231 **Table 1:** Physiological features of strain SG-772 compared to phylogenetic neighbor *Blautia*
 232 *stercoris* GAM6-1.

Characteristics	SG772	<i>Blautia stercoris</i> GAM6-1*
Habitat	Human gut	Human gut
Classification	Domain: Bacteria	Domain: Bacteria
	Phylum: Firmicutes	Phylum: Firmicutes
	Class: Clostridia	Class: Clostridia
	Order: Clostridiales	Order: Clostridiales
	Family: Lachnospiraceae	Family: Lachnospiraceae
	Genus: <i>Blautia</i>	Genus: <i>Blautia</i>
	Species: <i>brookingsii</i>	Species: <i>stercoris</i>
Optimum temperature	37 ⁰ C	37 ⁰ C
Shape	Coccobacilli	Cocci
Optimum pH	6.8	6.2
α-Galactosidase activity	-	+
β-Galactosidase activity	-	+
α-Glucosidase	-	+
β-Glucosidase	-	+
N-Acetyl-D-glucosamine	-	+
Turanose	-	-
Erythritol	-	+
Salicin	-	+
L-Glutamine	-	+
α-methyl-D-glucoside	-	NA
Glycyl-L- Proline	-	NA

Glycyl-L- Glutamine	-	-
Dextrin	+	NA
Arbutin	+	NA
D-Saccharic Acid	+	NA
a-D-Glucose	+	NA
L-Valine	+	NA
D-Cellobiose	+	NA
L-Rhamnose	+	NA
Succinic Acid	+	NA
D-Galactose	+	-
D-Mannose	+	NA
Palatinose	+	NA
Uridine-5'- Mono- phosphate	+	-
L-Fucose	+	NA
D-Glucose- 6-Phosphate	+	NA
Lactulose	+	+
D-Trehalose	+	NA
Succinic Acid Mono-Methyl Ester	+	NA
b-Cyclodextrin	+	NA
D-Sorbitol	+	NA
3-Methyl-D- Glucose	+	NA
D-Arabitol	+	NA
D-Glucosaminic Acid	+	NA
Adonitol	+	+
D-Melezitose	+	NA
D-Mannitol	+	NA

233 (*Data adopted from Park et al., 2012)

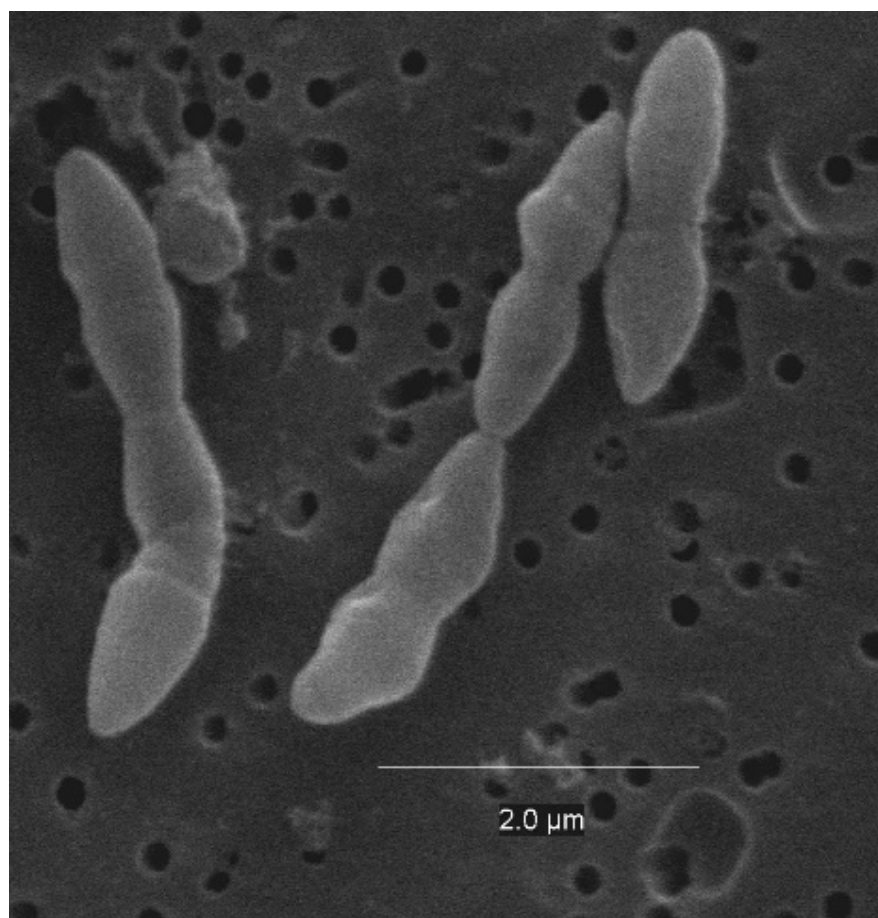
234

235 **Table 2:** General attributes of *Blautia brookingsii* SG772 with *Blautia hansenii* DSM20583

Characteristics	<i>Blautia brookingsii</i> SG772	<i>Blautia hansenii</i> DSM20583
Genome size (bp)	3,459,071	3,065,946
Number of contigs	55	1
Coding sequences	3343	3073
%GC content	43.97	39.04
tRNA	57	65
rRNAs (5S, 16S, 23S)	1, 8, 7	5, 5, 5

ncRNAs	4	4
Pseudogenes	152	68
CRISPR elements	1	2
Phage	1	0

236



237

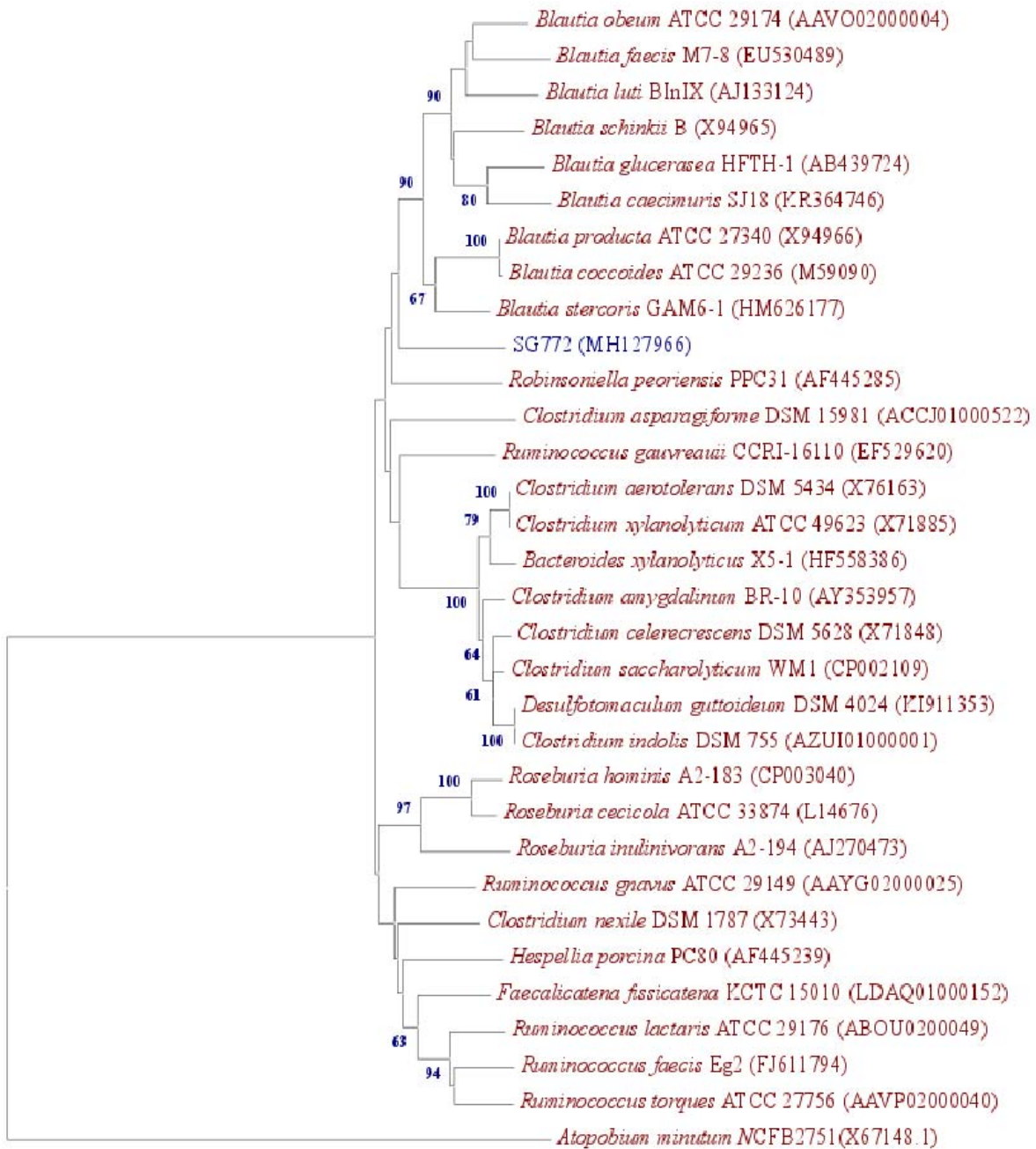
238

239 **Figure 1:** Scanning electron micrograph of strain SG772 grown on BHI agar at 37⁰C for 24
240 hours. Bar, 2.0 μm

241

242

243

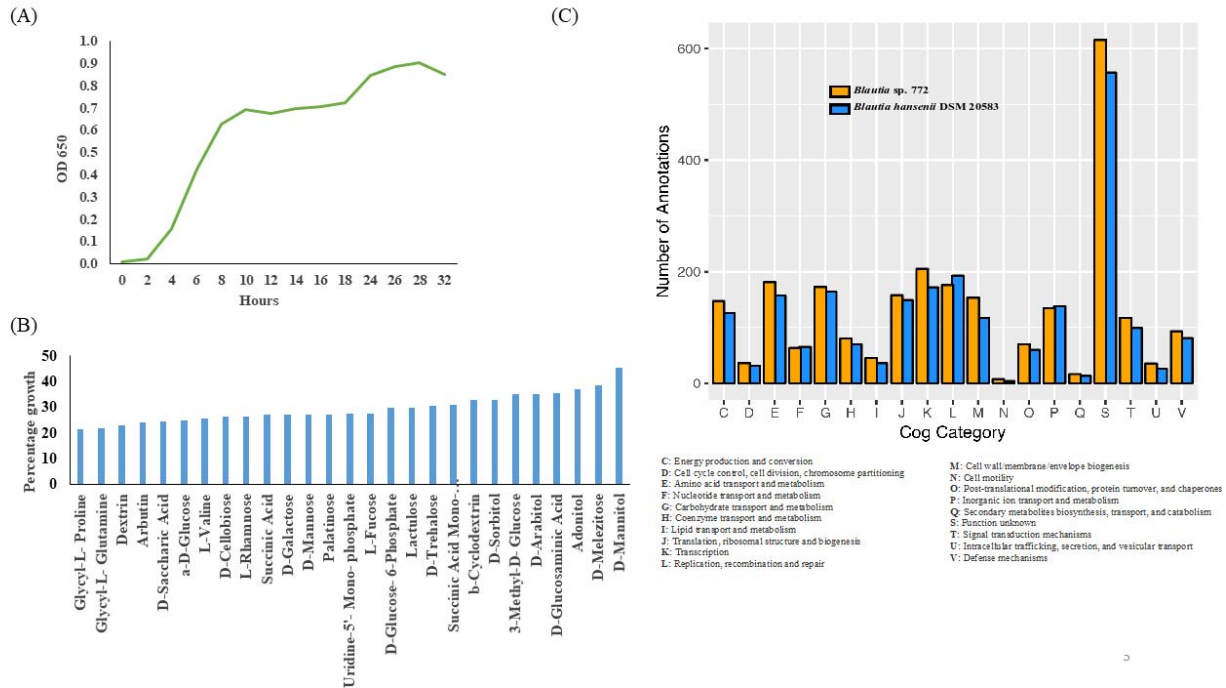


244

245 **Figure 2:** 16s rRNA sequence based phylogenetic tree constructed using neighbor-joining

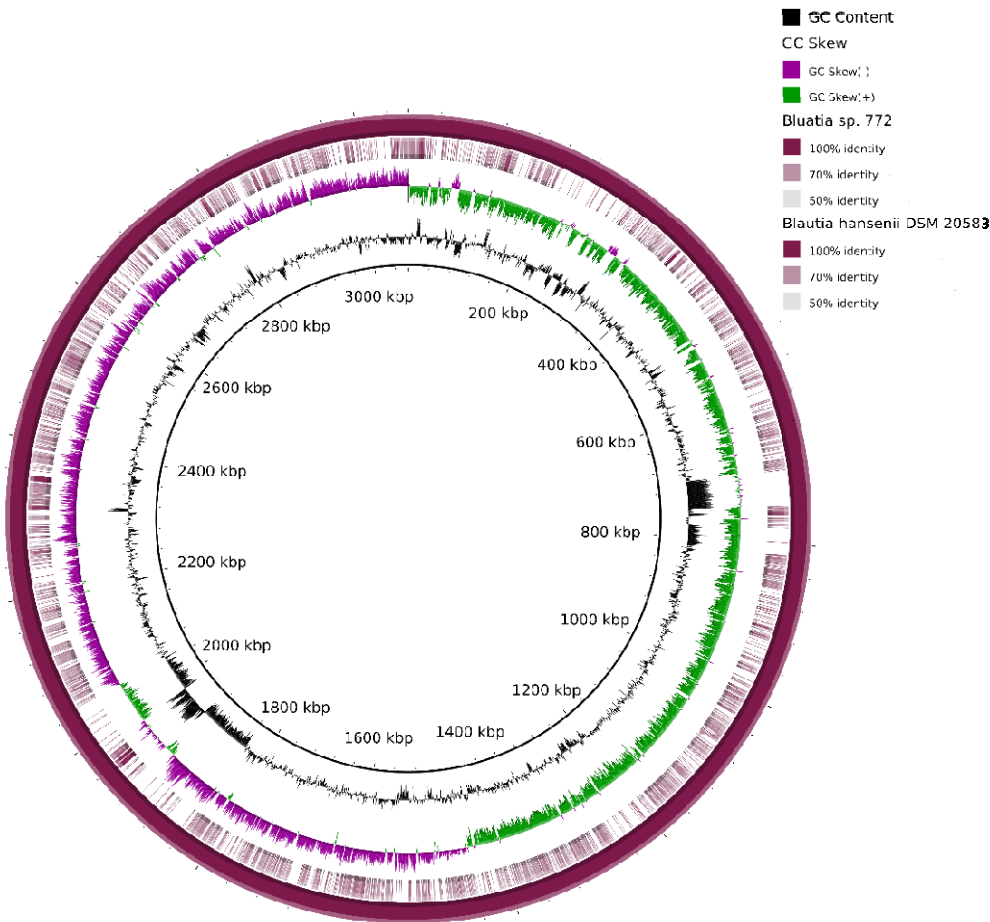
246 method. The 16S rRNA gene sequence was compared against the EZ-taxon, which showed the

247 highest similarity (94.39%) with *Blautia stercoris* GAM-61. The sequence of *Atobobium*
 248 *minutum* NCFB2751 was used as an outgroup. Numbers at nodes indicate bootstrap values
 249 expressed as percentages of 1000 replications. Bar, 0.02 accumulated changes per nucleotide.
 250 GenBank accession numbers are shown in parentheses.



251
 252 **Figure 3: Phenotypic and genotypic profiling of *Blautia sp. SG772*-** (A) Growth curve of the
 253 strain *Blautia sp. SG772*, (B) Bar plot depicting different patterns of substrate utilization, (C) Bar
 254 graph depicting comparative abundance of COG gene families between the two genomes.

255



256

257 **Figure 4: Comparative genome map of *Blautia* sp. SG772 using *B. hansenii* DSM 20583 as a**
258 **reference.** From the inside out, circle 1 represents the mean centered G+C content; circle 2
259 shows GC skew calculated as $(G - C) / (G + C)$; circle 3 represents the genome sequence
260 similarity of *Blautia* sp. SG772 based on color gradient against the reference genome of *B.*
261 *hansenii* DSM 20583; while circle 4 represents the genome of *B. hansenii* DSM 20583.

262