1 2	Combined use of procalcitonin and C-reactive protein levels can help clinically diagnose bacterial co-infections in children infected with H1N1 influenza
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15	
16	Abstract
17	Objective
18	This study evaluated the diagnostic value of measuring the levels of procalcitonin (PCT) and C-reactive protein
19	(CRP) to differentiate children co-infected with H1N1 influenza and bacteria from children infected with H1N1
20	influenza alone and to provide a reliable clinical diagnostic support system with improved accuracy and precision
21	control.
22	Methods
23	Consecutive patients (children aged <5 years) with laboratory-confirmed H1N1 influenza who were hospitalized or
24	received outpatient care from a tertiary-care hospital in Canton, China between 1 January 2012 and 1 September
25	2017 were included in the present study. Laboratory results, including serum PCT and CRP levels, white blood cell
26	(WBC) counts, and blood and sputum cultures, were analyzed. The predictive value of the combination of
27	biomarkers versus either biomarker alone for diagnosing bacterial co-infections was evaluated using logistic
28	regression analyses.
29	Results
30	Of 3180 children infected with H1N1 influenza, 226 (7.1%) met the bacterial co-infection criteria, with
31	Staphylococcus pneumoniae being the most commonly identified bacteria (36.28%). Significantly higher PCT
32	(1.46 vs 0.21 ng/ml, $p$ <0.001) and CRP (19.20 vs 5.10 mg/dl, $p$ <0.001) levels were detected in the bacterial
33	co-infection group than in the H1N1 infection only group. Multivariate logistic regression analysis showed

- 34 independent associations between PCT (odds ratio [OR]: 1.73, 95% confidence interval [CI], 1.34-2.42, p<0.001)
- 35 and CRP levels (OR:1.09, 95% CI, 1.06-1.13, p<0.001) with bacterial co-infections. Using PCT or CRP levels
- alone, the areas under the curves (AUCs) for predicting bacterial co-infections were 0.801 (95%CI, 0.772-0.855)
- and 0.762 (95% CI, 0.722-0.803), respectively. Using a combination of PCT and CRP, the logistic regression-based
- 38 model, Logit(P)=-1.912+0.546 PCT+0.087 CRP, showed significantly greater accuracy (AUC: 0.893, 95%CI:
- **39** 0.842-0.934) than did the other three biomarkers.

40 Conclusions

- 41 The combination of PCT and CRP levels could provide a useful method of distinguishing bacterial co-infections
- 42 from an H1N1 influenza infection alone in children during the early disease phase. After further validation, the
- 43 flexible model derived here could assist clinicians in decision-making processes.
- 44 Keywords
- 45 H1N1 influenza; Bacterial co-infection; Procalcitonin; C-reactive protein
- 46

## 47 Introduction

48 Co-infections with bacterial pathogens are a major cause of morbidity and mortality in children with H1N1 49 influenza infections worldwide [1]. Most deaths that occurred during several H1N1 influenza pandemics in 50 1918-1919 were due to bacterial co-infections rather than direct effects of the virus[2]. A recently study estimated 51 that bacterial co-infection was approximately 33% in patients hospitalized with H1N1infection[3], resulting in 52 more than 74% of patients receive antibiotic therapy after admission for initial H1N1 influenza infection [4], 53 despite adverse effects, the costs and increasing antibiotic resistance. Therefore, an early and rapid diagnosis was 54 recognized as a priority in managing bacterial co-infections, which may assist clinicians in initiating appropriate 55 antibiotic treatments to improve patient outcomes[5].

However, an early diagnosis of bacterial co-infections among patients with H1N1 influenza is challenging, because of the many overlapping symptoms and the lack of specific clinical manifestations of bacterial co-infections compared with H1N1 infection alone[6]; furthermore, young children cannot accurately describe their own disease symptoms, making the diagnosis even more difficult. Microbiological culture is the gold standard for diagnosing bacterial co-infections; however, current microbiological culture was time-consuming cultivation of bacteria before identification via colony and biochemical profiling, and the routine testing procedure may take several days and can also result in false-negative results.

63 Consequently, the availability of an efficient biomarkers system would be crucial in helping to quickly

64 differentiate bacterial co-infections from H1N1 infections alone. Recently, several inflammatory biomarkers have 65 been evaluated for their abilities to distinguish co-infections with H1N1 and bacteria from H1N1 infections alone. 66 Among these biomarkers, traditional biomarkers such as a white blood cell (WBC) count [7] and C-reactive 67 protein (CRP) levels[8] are commonly used to differentiate between bacterial and viral etiologies. Although 68 previous studies have focused on using CRP levels to detect bacterial co-infections in patients with H1N1 69 infections, the evidence from these studies is inconsistent. Studies suggested serum CRP as a potential diagnostic 70 biomarker [9-11], whereas Piacentini et al.[12] found that CRP levels were unable to distinguish bacterial 71 co-infections from H1N1 infections. Another interesting biomarker is procalcitonin (PCT), the prohormone of 72 calcitonin produced by C cells in the thyroid. Plasma PCT concentrations are low in healthy individuals and 73 increase during bacterial, parasitic, or fungal infections, whereas they remain at normal levels during viral 74 infections or noninfectious inflammatory reactions[13]. Studies have attempted to assess PCT levels in patients 75 with H1N1 infection and found that PCT helped to distinguish bacterial co-infections from H1N1 infections[14, 76 15]. Nevertheless, to the best of our knowledge, previous studies published to date have focused on adults[14, 15] 77 and patients with severe disease[16], but have included few patients with H1N1 infections.

78 Thus, in the present study, we conducted a retrospective analysis of 3180 children with H1N1 infection, to 79 evaluated the diagnostic levels of serum PCT, CRP and WBC alone and in combination in differentiating bacterial 80 co-infections from H1N1 influenza infections alone in children, to provide a reliable clinical diagnostic support 81 system for improving diagnostic accuracy and for enabling early treatment of bacterial co-infections during H1N1 82 influenza infections.

83

84

85 Methods

## 86 Settings and participants

We performed a retrospective cohort study of consecutive patients with laboratory-confirmed H1N1 influenza infections, all of whom were children <5 years old who were hospitalized or received outpatient care from a tertiary-care hospital in Canton, China between 1 January 2012 and 1 September 2017. Demographic and clinical characteristics, including age, gender, weight, diagnoses, total length of hospital stay, intensive care unit (ICU) admission, total length of ICU stay, total cost and in-hospital mortality were recorded. Data from initial laboratory exams, including serum PCT and CRP levels, WBC counts, and blood and sputum cultures were collected. The

- 93 ethics committee of Guangzhou Woman and Children's Medical Center approved our study, and written informed
- 94 consent was obtained from all the participants' parents or designated guardians.
- 95

## 96 Definitions

97 Patients diagnosed with H1N1 influenza infection confirmed by real-time reverse transcriptase polymerase chain 98 reaction (RT-PCR) [17] of nasopharyngeal secretions or bronchoalveolar lavage fluid samples within the first 48 99 hours hospitalization were included in the study. Bacterial co-infection was defined as a positive H1N1 influenza 100 viral PCR result with one or more bacterial pathogens detected. Bacterial cultures were obtained from blood, valid 101 sputum, lower respiratory tract samples or samples of other normally sterile fluids within the first 48 hours 102 hospitalization. We selected patients for this study who did not receive antibiotics prior to hospitalization to better 103 differentiate patients co-infected with H1N1 and bacteria from patients infected with H1N1 alone.

104

#### 105 Inflammatory biomarkers (PCT, CRP and WBC) measurements

Venous blood samples were collected from the patients infected with H1N1 upon admission. Serum PCT levels
were determined using an enzyme-linked fluorescence analysis (ELFA, VIDAS BRAHMS PCT kit, bioMerieux
SA, France). Serum CRP levels were determined using BNPProSpec automatic protein analyzer (Dade Behring
BN Prospec, USA)[18], and WBC counts were analyzed by using an Sysmex XE-2100 haematology analyser
(Sysmex, Kobe, Japan).

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## 113 Statistical analysis

114 Categorical variables are summarized using absolute values and percentages, and continuous variables are 115 presented as medians and interquartile ranges (IQR). The Chi-square tests (for nominal variables) or the Wilcoxon 116 rank sum test (for continuous variables) was employed for between-group comparisons. Univariate logistic 117 regression analysis was used to assess the ability of each biomarker (PCT, CRP and WBC) to diagnose bacterial 118 co-infections. Furthermore, iterative biomarker(s) were selected (including biomarker with p < 0.10) using 119 automatic forwards stepwise regression, and the multivariate logistic regression model was built. The performance 120 of the models was then assessed by calculating the area under the receiver-operating characteristic (ROC) 121 curve(AUC). The AUC values were compared for each biomarkers individually and in conjunction with 122 biomarkers model by Hanley and McNeil method[19]. The sensitivity, specificity, positive predictive value (PPV)

123	and negative predictive value (NPV) were also reported. The Youden index (sensitivity + specificy-1) was used to
124	determine the optimal ROC cutoff value. Moreover, 10-fold cross-validation to evaluate the robustness of the
125	estimates obtained from the constructed model, as previously described [20], was performed. Then, we averaged
126	the AUC, sensitivity and specificity values obtained from the 10-fold cross-validations to generate summary
127	performance estimates.
128	All statistical analyses were performed using R Software, version 3.4.2 (www.r-project.org). A two-tailed p value

- 129 <0.05 was considered significant.
- 130
- 131 Results

#### 132 Study population and bacterial pathogen characteristics

During the study period, 3180 children with laboratory-confirmed H1N1 influenza infection were included, with a median age of 3.6 years (IQR, 1.8-7.5 years); 1784 (52.3%) were males. Among these patients, 226 (7.1%) had a proven bacterial co-infection. *Streptococcus pneumoniae* was the most frequent pathogens causing the bacterial co-infection in 82(36.2%) cases, followed by *Staphylococcus aureus* in 55 (24.3%) cases and *Pseudomonas aeruginosa* in 34 (15.0%) cases (Table S1). Eight children (3.5%) displayed two positive respiratory tract bacterial cultures.

139

140 When the baseline characteristics and clinical outcomes of the H1N1 plus bacterial co-infection group were 141 compared, children in the H1N1-alone group were older, but this result was not significant (median age, 2.5 vs 2.4 142 years, p=0.197). Differences in gender or weight were not observed between the two groups; however, the 143 bacterial co-infection group showed significantly higher inpatient admission (14.3% vs 50.4%, p<0.001) and ICU 144 admission rates (2.6% vs 36.3%, p < 0.001) than patients in the H1N1-alone group. The bacterial co-infection group 145 also required longer hospital stays (5 vs 10 days, p=0.003) than H1N1-alone group and thus had much higher 146 hospital costs (median hospital cost, 1213.2 vs 3467.3 RMB, p<0.001). Moreover, a higher in-hospital mortality 147 rate was noted for the bacterial co-infection group than the H1N1 alone group (0.1% vs 4.8%, p < 0.001) (Table 1). 148

# Comparison of serum PCT, CRP and WBC levels between H1N1-alone and H1N1 with bacterialco-infection groups

Serum PCT, CRP and WBC levels were analyzed to identify potential biomarkers that distinguished between
H1N1 infections and H1N1 and bacterial co-infections. The median serum PCT, CRP and WBC levels were all

- 153 significantly higher in the H1N1 with bacterial co-infection group than in the H1N1-alone group (median PCT
- 154 level, 1.46 vs 0.21 ng/ml, *p*<0.001; median CRP level, 19.20 vs 5.10 mg/dl, *p*<0.001, median WBC count, 8.50 vs
- 155  $6.90 \times 10^9$  cells/l, *p*=0.019) (Figure 1).
- 156

## 157 Univariate and multivariate logistic regression analyses

Univariate analysis revealed significant associations of serum PCT, CRP and WBC levels with co-infections with H1N1 and bacteria (odds ratio [OR]:1.65, 95 % confidence interval [CI] 1.34-2.06, p<0.001; OR: 1.08, 95 % CI 1.06-1.09, p < 0.001; OR:1.06, 95% CI 1.04-1.09, p=0.02, respectively). The associations with PCT and CRP levels remained statistically significant(p<0.05) after the application of the forwards regression model, whereas WBC counts were excluded from the model (p<0.05). Then, multivariate logistic regression analysis showed that CRP (OR:1.09, 95% CI 1.06-1.13, p<0.001) and PCT levels (OR:1.73, 95%CI 1.34-2.42, p<0.001) were significant independent diagnostic biomarkers. (Table 2).

165

## 166 Comparison and validation of the model's diagnostic ability

167 Because the serum PCT and CRP levels were independent predictors that differentiated patients with bacterial 168 co-infections from patients infected with H1N1 alone, we constructed a new model, PCT&CRP [Logit (P) = -1.912 169 + 0.546 PCT+ 0.087 CRP], that combined the PCT and CRP levels. The performance of the ROC curves of the 170 constructed model, PCT, CRP, and WBC levels for differentiating children with H1N1 and bacterial co-infections 171 from children infected with H1N1 alone were compared. The AUC, sensitivity, specificity, PPV, and NPV are 172 shown in Table 3. The constructed model exhibited the largest AUC (0.893, 95%CI 0.852-0.934). The p values of 173 the ROC curve comparison between the constructed model and CRP and PCT levels were all less than 0.01. The 174 AUCs for PCT, CRP, and WBC levels were 0.801(95%CI, 0.772-0.855), 0.762(95%CI, 0.722-0.803), and 175 0.551(95%CI, 0.502-0.592), respectively. The optimum cutoff values for PCT, CRP, and WBC were 0.52 ng/ml, 176 13.55 mg/l and  $11.56 \times 10^9$  cells/l, respectively. Significant differences were observed among the ROC curves of 177 the PCT, CRP, and WBC (p < 0.05). The diagnostic ability of each model followed the order of PCT&CRP > PCT > 178 CRP > WBC (Figure 2). The PCT&CRP was superior to use of the PCT, CRP and WBC alone in differentiating 179 patients with bacterial co-infections from those infected with H1N1 alone. The robustness of PCT&CRP was 180 internally evaluated through 10-fold cross-validation. On average, the constructed model presented an AUC of 181 0.872, a sensitivity of 0.754 and a specificity of 0.896.

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183

#### 184 Discussion

185 Bacterial co-infection is especially known to excess the mortality and morbidity of H1N1 influenza.

186 Unfortunately, it is difficult to correctly diagnose bacterial coinfection based only on clinical criteria, as well as 187 bacterial culturing is time-consuming. There is a crucial need to differentiate H1N1 patients with bacterial 188 co-infection from H1N1 infection alone. The diagnostic and predictive value of serum PCT and CRP levels as 189 biomarkers has been discussed in several studies[10, 14, 15, 21]. Shin et al. [10]found that serum PCT was a good 190 indicator of in discriminating bacterial co-infections infection from H1N1 infections alone in 60 adult patients in 191 ICU. Guervilly et al. [21] report that PCT values were statistically higher in patients with bacterial co-infections. In 192 addition, PCT has been suggested to exclude bacterial co-infections in patients with an H1N1 infection and to 193 reliably and accurately reduce inappropriate antibiotic exposure [14]. Our results showed that serum PCT levels 194 was significantly higher in patients with bacterial co-infection compared to those infected H1N1 infection alone, 195 reminding us that PCT was association with bacterial co-infection. Furthermore, the results of ROC curve analysis 196 indicated that an AUC value of 0.801 (95% CI, 0.772-0.855), with cutoff value was 0.52ng/ml, supported the 197 prognostic value of PCT in children with or with bacterial co-infection.

198 The diagnostic utility of CRP to differentiate bacterial co-infection from H1N1 infection is disputed [9-11, 15]. 199 Haran et al. [11]found that CRP as predictor of bacterial infection among patients with an H1N1 infection. 200 Similarly, Shin et al. [10] reported that serum CRP levels was significantly higher in patients with bacterial 201 co-infection compared to those infected with H1N1 alone. But other study suggested that CRP levels were unable 202 to distinguish bacterial co-infections from H1N1 infections [9]. Our study showed that serum CRP levels was 203 significantly higher in patients with bacterial co-infection compared to those infected H1N1 infection alone, it 204 indicating that those biomarkers could be aid in discriminating between these conditions. Furthermore, our study 205 showed that the diagnostic efficacy of PCT for bacterial co-infection in H1N1 infection was better than that of 206 CRP (AUC 0.801 and 0.783, respectively; p<0.05), consistent with the results of a previous study[11]. However, 207 the AUC of WBC counts in diagnosing bacterial co-infections was 0.551 (95%CI, 0.502-0.592), indicating that 208 WBC may not be a valuable biomarker for our cohort of children.

Previous study use of a combination of CRP and PCT levels for evaluating for bacterial co-infections increased the accuracy of differentiating children with bacterial co-infections from those infected with H1N1 alone[10]. Similar observations were reported in the present study, we used a multivariate logistic regression analysis to construct a new model using the PCT and CRP levels: [Logit(P)=-1.912+0.546 PCT+0.087 CRP]. The ROC curve analysis

yielded an AUC value for the model of up to 0.893, which was clearly superior to PCT or CRP levels alone
(p<0.05). Furthermore, the constructed model [Logit(P) =-1.912+0.546PCT +0.087CRP] was internally validated</li>
through 10-fold cross-validation, resulting in high diagnostic accuracy. Therefore, the joint detection of PCT and
CRP levels clearly improves the prognosis of children with H1N1 bacterial co-infection. Based on the results from
our study, the combination of serum PCT and CRP levels will help clinicians determine the appropriate antibiotic
therapy[22], thus potentially improving patient outcomes and reducing antibiotic overuse [5].

This study involved 3180 children with H1N1, 7.11% of whom presented a confirmed bacterial co-infection, after including both outpatients and inpatients. The proportion of bacterial co-infection while similar to that previously reported for H1N1. Nevertheless, previous studies of children with H1N1 influenza infection reported a bacterial co-infection rate ranging from 18% to 60%[23, 24]. These rates may be overestimated because the previous studies were limited to pediatric patients in the ICU, which represent a population with moderate to severe H1N1 influenza infection. Moreover, children with bacterial co-infections exhibited a higher percentage of ICU admission rates in the current study.

226 Our study shows that Streptococcus pneumoniae was the leading cause of bacterial co-infection with H1N1, 227 followed by Staphylococcus aureus and Streptococcus pyogenes, consistent with the results from previous 228 studies[25, 26]. Additionally, children with H1N1 influenza infection and bacterial co-infection have been reported 229 to exhibit a higher risk of severe outcomes [26-28]. In our study, patients co-infected with bacteria and H1N1 230 exhibited increased percentages of inpatient and ICU admissions, higher costs and longer hospital stays. 231 Furthermore, a significantly higher hospital mortality rate was observed in children with H1N1 and bacterial 232 co-infections because bacterial co-infections represent an important mortality risk factor, possibly suggesting early 233 empiric antibiotics treatment in severe patients may improve outcomes.

The potential limitations of our study should be mentioned. First, the levels of selected biomarkers (PCT, CRP and WBC) were evaluated only once. Second, our diagnostic model was derived and validated at a single hospital center, and it should be validated in a multicenter trial center before its broad application. Finally, we also acknowledge that, we may have created a bias, due to bacterial organisms cannot be confirmed solely with blood, sputum, lower respiratory tract samples.

239

#### 240 Conclusion

In conclusion, we detected serum PCT and CRP levels and revealed that they represent promising biomarkers and
 useful clinical tools for differentiating pediatric patients with bacterial co-infections from those infected with

- 243 H1N1 alone. Furthermore, the combination of PCT and CRP levels could represent a useful method for screening
- 244 bacterial co-infections from H1N1 influenza infections alone in children during the early disease phase. After
- further validation, the flexible model reported here may assist clinicians with decision-making processes.

246

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- 250

# 251 Reference

- 252 [1] H. Nair, W.A. Brooks, M. Katz, A. Roca, J.A. Berkley, S.A. Madhi, J.M. Simmerman, A. Gordon, M.
- Sato, S. Howie, Global burden of respiratory infections due to seasonal influenza in young children: a
   systematic review and meta-analysis, The Lancet, 378 (2011) 1917-1930.
- [2] J.A. McCullers, The co-pathogenesis of influenza viruses with bacteria in the lung, Nature Reviews
   Microbiology, 12 (2014) nrmicro3231.
- [3] C. Cillóniz, S. Ewig, R. Menéndez, M. Ferrer, E. Polverino, S. Reyes, A. Gabarrús, M.A. Marcos, J.
  Cordoba, J. Mensa, Bacterial co-infection with H1N1 infection in patients admitted with community
  acquired pneumonia, Journal of Infection, 65 (2012) 223-230.
- [4] S. Jain, L. Kamimoto, A.M. Bramley, A.M. Schmitz, S.R. Benoit, J. Louie, D.E. Sugerman, J.K.
  Druckenmiller, K.A. Ritger, R. Chugh, Hospitalized patients with 2009 H1N1 influenza in the United
  States, April–June 2009, New England journal of medicine, 361 (2009) 1935-1944.
- [5] D.S. Chertow, M.J. Memoli, Bacterial coinfection in influenza: a grand rounds review, Jama, 309(2013) 275-282.
- [6] R. Libster, J. Bugna, S. Coviello, D.R. Hijano, M. Dunaiewsky, N. Reynoso, M.L. Cavalieri, M.C.
  Guglielmo, M.S. Areso, T. Gilligan, Pediatric hospitalizations associated with 2009 pandemic influenza
  A (H1N1) in Argentina, New England Journal of Medicine, 362 (2010) 45-55.
- [7] S. Lindbäck, U. Hellgren, I. Julander, L. Hansson, The value of C-reactive protein as a marker of
  bacterial infection in patients with septicaemia/endocarditis and influenza, Scand. J. Infect. Dis., 21
  (1989) 543-549.
- [8] S. Sanders, A. Barnett, I. Correa-Velez, M. Coulthard, J. Doust, Systematic review of the diagnostic
  accuracy of C-reactive protein to detect bacterial infection in nonhospitalized infants and children with
  fever, The Journal of pediatrics, 153 (2008) 570-574. e573.
- [9] P.R. Ingram, T. Inglis, D. Moxon, D. Speers, Procalcitonin and C-reactive protein in severe 2009
  H1N1 influenza infection, Intensive care medicine, 36 (2010) 528-532.
- 276 [10] S. Ahn, W.Y. Kim, S.H. Kim, S. Hong, C.M. Lim, Y. Koh, K.S. Lim, W. Kim, Role of procalcitonin and
- C reactive protein in differentiation of mixed bacterial infection from 2009 H1N1 viral pneumonia,
   Influenza and other respiratory viruses, 5 (2011) 398-403.
- 279 [11] J.P. Haran, F.L. Beaudoin, S. Suner, S. Lu, C-reactive protein as predictor of bacterial infection
- among patients with an influenza-like illness, The American journal of emergency medicine, 31 (2013)
- 281 137-144.

[12] E. Piacentini, B. Sánchez, V. Arauzo, E. Calbo, E. Cuchi, J.M. Nava, Procalcitonin levels are lower in
 intensive care unit patients with H1N1 influenza A virus pneumonia than in those with
 community-acquired bacterial pneumonia. A pilot study, Journal of critical care, 26 (2011) 201-205.

[13] D. Gendrel, J. Raymond, M. Assicot, F. Moulin, J.-L. Iniguez, P. Lebon, C. Bohuon, Measurement of
 procalcitonin levels in children with bacterial or viral meningitis, Clinical infectious diseases, 24 (1997)
 1240-1242.

[14] A.H. Rodríguez, F.X. Avilés-Jurado, E. Díaz, P. Schuetz, S.I. Trefler, J. Solé-Violán, L. Cordero, L.
Vidaur, Á. Estella, J.C.P. Laderas, Procalcitonin (PCT) levels for ruling-out bacterial coinfection in ICU
patients with influenza: a CHAID decision-tree analysis, Journal of infection, 72 (2016) 143-151.

291 [15] E. Cuquemelle, F. Soulis, D. Villers, F. Roche-Campo, C.A. Somohano, M. Fartoukh, A. Kouatchet, B.

Mourvillier, J. Dellamonica, W. Picard, Can procalcitonin help identify associated bacterial infection in
patients with severe influenza pneumonia? A multicentre study, Intensive care medicine, 37 (2011)
796-800.

[16] R. Pfister, M. Kochanek, T. Leygeber, C. Brun-Buisson, E. Cuquemelle, M.P. Machado, E. Piacentini,

296 N.E. Hammond, P.R. Ingram, G. Michels, Procalcitonin for diagnosis of bacterial pneumonia in critically

ill patients during 2009 H1N1 influenza pandemic: a prospective cohort study, systematic review and
individual patient data meta-analysis, Critical care, 18 (2014) R44.

- [17] C. Duncan, J.L. Guthrie, N. Tijet, N. Elgngihy, C. Turenne, C. Seah, R. Lau, L. McTaggart, G. Mallo, S.
   Perusini, Analytical and clinical validation of novel real-time reverse transcriptase–polymerase chain
   reaction assays for the clinical detection of swine-origin H1N1 influenza viruses, Diagnostic
   microbiology and infectious disease, 69 (2011) 167-171.
- 303 [18] A. Esfahani, N. Makhdami, E. Faramarzi, M. Asghari Jafarabadi, A. Ostadrahimi, M. Ghayour
  304 Nahand, Z. Ghoreishi, Prealbumin/CRP based prognostic score, a new tool for predicting metastasis in
  305 patients with inoperable gastric cancer, Gastroenterology research and practice, 2016 (2016).

[19] B.J. McNeil, J.A. Hanley, Statistical approaches to the analysis of receiver operating characteristic
 (ROC) curves, Medical decision making, 4 (1984) 137-150.

308 [20] R. Tibshirani, T. Hastie, B. Narasimhan, G. Chu, Diagnosis of multiple cancer types by shrunken
 309 centroids of gene expression, Proceedings of the National Academy of Sciences, 99 (2002) 6567-6572.

310 [21] C. Guervilly, Y. Coisel, E. Botelho-Nevers, S. Dizier, M. Castanier, R. Lepaul-Ercole, O. Brissy, A. Roch,

J.-M. Forel, L. Papazian, Significance of high levels of procalcitonin in patients with influenza A (H1N1)
 pneumonia, Journal of Infection, 61 (2010) 355-358.

313 [22] W.C. Albrich, S. Harbarth, Pros and cons of using biomarkers versus clinical decisions in start and
314 stop decisions for antibiotics in the critical care setting, Intensive care medicine, 41 (2015) 1739-1751.

315 [23] S.B. Carr, E.E. Adderson, H. Hakim, X. Xiong, X. Yan, M. Caniza, Clinical and demographic

characteristics of seasonal influenza in pediatric patients with cancer, The Pediatric infectious diseasejournal, 31 (2012) e202.

318 [24] E. Cordero, P. Pérez - Romero, A. Moreno, O. Len, M. Montejo, E. Vidal, P. Martín - Dávila, M.
319 Fariñas, N. Fernández - Sabé, M. Giannella, Pandemic influenza A (H1N1) virus infection in solid organ

- transplant recipients: impact of viral and non viral co infection, Clinical Microbiology and Infection,
  18 (2012) 67-73.
- 322 [25] Q. Zhang, W. Ji, Z. Guo, Z. Bai, N.E. MacDonald, Risk factors and outcomes for pandemic H1N1
- 323 influenza compared with seasonal influenza in hospitalized children in China, Canadian Journal of
- 324 Infectious Diseases and Medical Microbiology, 23 (2012) 199-203.

[26] C. Reed, A.J. Kallen, M. Patton, K.E. Arnold, M.M. Farley, J. Hageman, L. Finelli, Infection with
 community-onset Staphylococcus aureus and influenza virus in hospitalized children, The Pediatric

327 infectious disease journal, 28 (2009) 572-576.

328 [27] T. Nguyen, U.G. Kyle, N. Jaimon, M.H. Tcharmtchi, J.A. Coss-Bu, F. Lam, J. Teruya, L. Loftis,

- 329 Coinfection with Staphylococcus aureus increases risk of severe coagulopathy in critically ill children
- 330 with influenza A (H1N1) virus infection, Critical care medicine, 40 (2012) 3246.
- 331 [28] N. Bhat, J.G. Wright, K.R. Broder, E.L. Murray, M.E. Greenberg, M.J. Glover, A.M. Likos, D.L. Posey,
- A. Klimov, S.E. Lindstrom, Influenza-associated deaths among children in the United States, 2003–2004,
- 333 New England Journal of Medicine, 353 (2005) 2559-2567.

- 354 Table

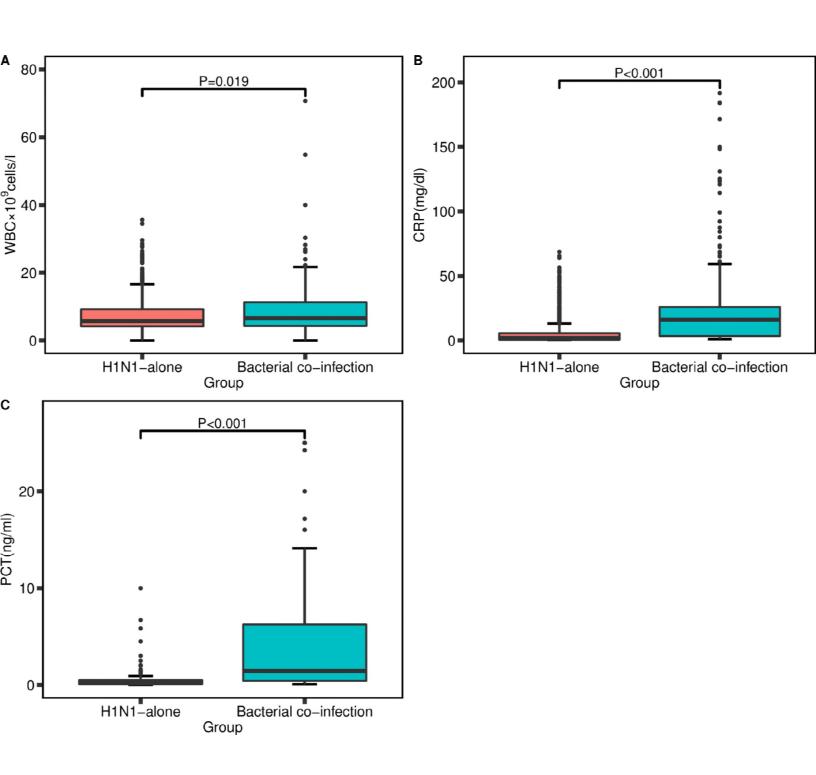
**Table 1** Demographic and clinical characteristics of patients with H1N1 influenza who presented with and without

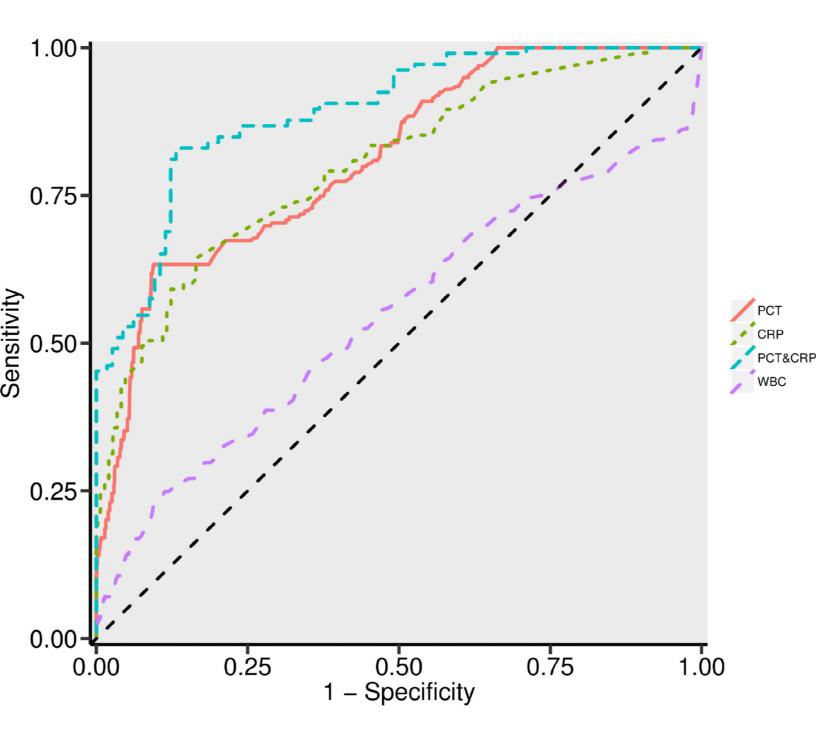
bacterial co-infections.

357 Abbreviations: IQR: interquartile range; ICU: intensive care unit; \* The differences between the H1N1-alone

358 359	group and bacterial co-infection group were examined using the Wilcoxon rank sum test or Chi-square tests.
360	Table 2 Univariate and multivariate logistic regression analyses of biomarkers for of bacterial co-infection in
361	H1N1 patients infected with H1N1.
362	Abbreviations: $\beta$ : regression coefficient; OR: odds ratio; CI: confidence interval; * In the multivariate logistic
363	regression analysis, WBC counts (p>0.05) were excluded from the final model based on the results of the forward
364	stepwise analysis.
365	
366	Table 3 Discriminatory performance of WBC, CRP, PCT and the constructed model for detecting patients with
367	H1N1 influenza and a bacterial co-infection.
368	Abbreviations: AUC: area under the receiver-operating-characteristic curve; CI: confidence interval; PPV: positive
369	predictive value; NPV: negative predictive value; PCT&CRP: Logit (P) = $-1.912 + 0.546$ PCT+ 0.087 CRP.
370	
371	Figure
372	Figure 1 Serum PCT (A), CRP (B) and WBC (C) levels in patients with H1N1 influenza who presented with and
373	without bacterial co-infections. The differences between the H1N1-alone group and bacterial co-infection group
374	were examined using the Wilcoxon rank sum test.
375	
376	Figure 2 ROC curves of PCT, CRP, WBC and PCT&CRP (Logit (P) = $-1.912 + 0.546$ PCT+ 0.087 CRP) for
377	differentiating patients with bacterial co-infections from those with infected H1N1 alone.
378	
379	Supplementary material

380 Table S1 Pathogens isolated in patients with H1N1 influenza and a bacterial co-infection





	H1N1 alone	Bacterial co-infection	
Characteristic	( <i>n</i> = 2954)	(n = 226)	<i>p</i> *
Age (years), median (IQR)	2.5 (1.2, 4.0)	2.4 (1.0, 4.1)	0.197
Male, <i>n</i> (%)	1794 (49.5)	105 (47.6)	0.240
Weight (kg), median (IQR)	9.4 (5.2, 20.8)	9.6 (4.8, 21.5)	0.368
Patients, n (%)			<0.001
Inpatient	432 (14.3)	114 (50.4)	
Outpatient	2522 (85.4)	112 (49.6)	
Total length of hospital stays (days), median (IQR)	5.0 (2.0, 9.0)	10.00 (6.0, 18.3)	<0.001
ICU admission, <i>n</i> (%)	77 (2.6)	82 (36.3)	<0.001
Total length of ICU stays (days), median (IQR)	6.0 (3.8, 10.0)	11.0 (6.3, 19.8)	<0.001
Total cost (RMB), median (IQR)	1213.2 (205.5, 3041.7)	3467.3 (1302.3, 41321.6)	<0.001
In-hospital mortality, n (%)	3 (0.1)	11 (4.8)	<0.001

 Table 1 Demographic and clinical characteristics of patients with H1N1 influenza who presented

 with and without bacterial co-infections.

Abbreviations: IQR: interquartile range; ICU: intensive care unit; \* The differences between the H1N1-alone group and bacterial co-infection group were examined using the Wilcoxon rank sum test or Chi-square tests.

	Univariate				Multivariate				
Variable	β	OR	95% CI	р		β	OR	95% CI	р
WBC	0.060	1.06	1.04-1.09	< 0.001		_*	-	-	-
PCT	0.498	1.65	1.34-2.06	< 0.001		0.546	1.73	1.34-2.42	< 0.001
CRP	0.073	1.08	1.06-1.09	< 0.001		0.087	1.09	1.06-1.13	< 0.001

 Table 2 Univariate and multivariate logistic regression analyses of biomarkers for of bacterial

 co-infection in H1N1 patients infected with H1N1.

Abbreviations:  $\beta$ : regression coefficient; OR: odds ratio; CI: confidence interval; \* In the multivariate logistic regression analysis, WBC counts (p>0.05) were excluded from the final model based on the results of the forward stepwise analysis.

Variables	AUC (95% CI)	cutoff level	Sensitivity	Specificity	PPV	NPV
WBC	0.551 (0.502-0.592)	11.56	0.267	0.887	0.144	0.910
CRP	0.762 (0.722-0.803)	13.55	0.633	0.856	0.330	0.971
PCT	0.801 (0.772-0.855)	0.52	0.643	0.886	0.773	0.852
PCT&CRP *	0.893 (0.852-0.934)	-	0.830	0.868	0.854	0.846

 Table 3 Discriminatory performance of WBC, CRP, PCT and the constructed model for detecting

 patients with H1N1 influenza and a bacterial co-infection.

Abbreviations: AUC: area under the receiver-operating-characteristic curve; CI: confidence interval; PPV: positive predictive value; NPV: negative predictive value; PCT&CRP: Logit (P) = -1.912 + 0.546 PCT+ 0.087 CRP.