Experimental increases in glucocorticoids alter function of the HPA axis in wild red squirrels without negatively impacting survival and reproduction

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Abstract

Hormones such as glucocorticoids (colloquially referred to as "stress hormones") have important effects on animal behavior and life history traits, yet most of this understanding has come through correlative studies. While experimental studies offer the ability to assign causality, there are important methodological concerns that are often not considered when manipulating hormones, including glucocorticoids, in wild animals. In this study, we examined how experimental elevations of cortisol concentrations in wild North American red squirrels (*Tamiasciurus hudsonicus*) affected their hypothalamic–pituitary–adrenal (HPA) axis reactivity, and life history traits including body mass, litter survival, and adult survival. The effects of exogenous cortisol on plasma cortisol concentrations depended on the time between treatment consumption and blood sampling. In the first nine hours after consumption of exogenous cortisol, individuals had significantly higher true baseline plasma cortisol concentrations, but adrenal gland function was impaired as indicated by their dampened response to capture and handling and to injections of adrenocorticotropic hormone compared to controls. Approximately 24 hours after consumption of exogenous cortisol, individuals had much lower plasma cortisol concentrations than controls, but adrenal function was restored. Corticosteroid binding globulin (CBG) concentrations were also significantly reduced in squirrels treated with cortisol. Despite these profound shifts in the functionality of the HPA axis, squirrel body mass, offspring survival, and adult survival were unaffected by experimental increases in cortisol concentrations. Our results highlight that even short-term experimental increases in glucocorticoids can affect adrenal gland functioning and CBG concentrations but without other side-effects.

Introduction

Associations between glucocorticoids (GCs) and life history or behavioral traits are increasingly studied, due to their role as a mechanistic link between the genome and the environment, and to uncover general relationships between hormones and fitness (Breuner et al., 2008; Dantzer et al., 2016). GCs are released by the hypothalamic-pituitary-adrenal (HPA) axis in response to environmental challenges and have widespread effects on physiology and behavior (Sapolsky et al., 2000; Romero, 2004). Correlational studies have helped advance our understanding of the relationships between GCs and phenotypic traits in wild animals, but establishing the causality of such relationships requires experimental manipulation of GCs. In laboratory settings, hormone manipulations are logistically feasible (e.g. Karatsoreos et al., 2010; Lussier et al., 2009), but experimental studies conducted in wild populations are likely to provide better insights into the ecologically relevant effects of GCs on life history variation.

Hormone manipulations in wild animals are more challenging than in the laboratory, but several methods have been developed (see Sopinka et al., 2015). That said, exogenous GCs may have unintended physiological side effects, which may influence or skew interpretation of the results obtained from manipulative studies. One potential problem with hormone manipulations is related to the fact that the endocrine system is a homeostatic system that is controlled by negative feedback mechanisms and tends to compensate for disruption. Therefore, if animals are treated with a hormone, the endogenous production of the hormone may be reduced after a few days, and longer treatment duration may lead to the regression of the endocrine gland, and have important consequences for endocrine homeostasis (Fusani, 2008). Such effects are well documented in humans, as both cortisol and synthetic GCs (which may be more potent, see Meikle and Tyler, 1977), are used to treat a range of ailments (Arabi et al., 2010; Kirwan et al., 2007). For example, cortisol administration may lead to side-effects, including

suppression of the HPA axis and reduced adrenal function (Broide et al., 1995; Feiwel et al., 1969; Jacobs et al., 1983). Although such side-effects are usually temporary (Morris and Jorgensen, 1971; Streck and Lockwood, 1979), in extreme cases, patients may develop more severe and long term conditions such as Cushing's syndrome (see Axelrod, 1976) or secondary adrenal insufficiency that can lead to Addison's disease (Arlt and Allolio, 2003).

If GC manipulations affect the adrenal glands, endogenous production of GCs, and endocrine homeostasis, this may lead to unintended consequences in wild animals. This could jeopardize the value of performing such studies, as they could adversely influence survival and reproduction. Indeed, some studies indicate that elevations in GCs reduce estimates of fitness (Bonier et al., 2009; Breuner et al., 2008; Wingfield et al., 1998), but it is unclear if this is due to an unintended complication from the manipulation rather than a natural consequence of increased GCs. Although these issues were highlighted 10-15 years ago (Romero, 2004; Fusani, 2008; see also Sopinka et al. 2015; Crossin et al., 2016), detailed studies about the potential complications of manipulating hormones in wild animals have not been widely performed except in birds. Torres-Medina et al. (2018) reviewed the consequences of experimentally elevated GCs on baseline and stress-induced corticosterone levels from previous studies on multiple bird species that were published 2005-2015. Many but not all of these studies were conducted in free-living birds. They showed that most studies that experimentally elevated GCs (using silastic implants, time release pellets, or osmotic pumps) examined how corticosterone treatment affected baseline corticosterone levels but very few investigated treatment effects on stress-induced corticosterone levels. Their results documented that birds treated with exogenous GCs exhibited lower stress-induced corticosterone levels, suggesting that experimental elevation of GCs can suppress the activity of the HPA axis in wild birds just as in studies in humans or laboratory rodents.

Unlike birds, studies that experimentally elevated GCs in free-living mammals are extremely rare and we are not aware of any study that has investigated both how treatment with GCs affects the HPA axis in wild mammals or how changes in the HPA axis induced by treatment with exogenous GCs affects fitness proxies. We examined how exogenous cortisol affected the HPA axis and life history traits of North American red squirrels (*Tamiasciurius hudsonicus*) by treating squirrels with exogenous cortisol or control vehicle. We expected that exogenous cortisol would increase fecal glucocorticoid metabolite (FGM) and plasma cortisol concentrations, but that, as may be the case in humans receiving GC therapy or birds with corticosterone implants, HPA axis responsiveness would decrease. We examined how administration of exogenous cortisol affected the responsiveness of the HPA axis by measuring the change in plasma cortisol concentrations following 1) capture and handling and 2) pharmaceutical suppression with dexamethasone (Dex) and pharmaceutical stimulation with adrenocorticotropin hormone (ACTH; hereafter "Dex/ACTH challenges"). Because we expected that administration of cortisol would suppress HPA axis responsiveness, we also examined how quickly the HPA axis recovered after administration of exogenous cortisol by measuring HPA axis responsiveness (using the Dex/ACTH challenges) in squirrels that received exogenous cortisol on the same day of sampling or the day after their last treatment. We expected that exogenous cortisol might lead to increased body mass in squirrels (Axelrod, 1976), but did not expect our treatment dosages to be sufficiently high to cause anorexia through sustained adrenal impairment (Arlt and Allolio, 2003). As we aimed to keep GCs within a physiologically-relevant ('normal') range for this species, we did not expect to see negative effects of our treatments on body mass or adult or litter survival.

Methods

Study population

All of our research was approved by the *Animal Care and Use Committee* at the University of Michigan (PRO00005866). We studied a natural population of red squirrels in the Yukon, Canada (61°N, 138°W) that has been monitored since 1987 (Boutin et al., 2006; McAdam et al., 2007). All squirrels in this population are individually identified by a unique ear tag in each ear, as well as a unique color combination of colored wires attached to each ear tag which allow researchers to identify individuals from a distance. Squirrels were live-trapped (Tomahawk Live Trap Co., WI, USA), during which they were weighed using a Pesola spring balance, and fecal samples were collected from underneath traps, placed on ice, and stored at -20 °C upon return to the field station (Dantzer et al., 2010). Female and male reproductive condition was assessed through palpation or by expressing milk from the teats in females (see McAdam et al., 2007).

Experimental manipulations of GCs

In 2015 and 2016, squirrels were randomly allocated to either control (8 g all natural peanut butter, 2 g wheat germ, no cortisol) or cortisol treatments (8 g peanut butter, 2 g wheat germ with 6, 8, or 12 mg of cortisol [H4001, Sigma Aldrich, USA]). Dosages of 0, 6, 8, and 12 mg cortisol were selected following previous studies in red squirrels (Dantzer et al., 2013) and laboratory rodents (Casolinia et al., 1997; Catalani et al., 2002; Mateo, 2008) that used similar dosages to induce a moderate increase in GCs. Treatments were provided directly to squirrels by putting the treatment in a bucket that was hung from trees on their territories (Dantzer et al., 2013). To ensure that target squirrels (identifiable through ear tags/radio-collars) were consuming the treatments, camera traps (Reconyx PC900 HyperFire Professional Covert IR) were placed by the buckets of 31 squirrels for five continuous days. Out of 155

d of camera trapping, conspecific pilferage was only observed ten times, and there was one case of heterospecific pilferage by a grey jay (*Perisoreus canadensis*). Consumption of each treatment was estimated daily by checking buckets for any leftovers and estimating these as a percentage. Squirrels consumed on average 91.9% of their total peanut butter treatments (median = 100%, SD = 12.13%, range = 43-100%).

Effects of exogenous cortisol on fecal glucocorticoid metabolites (FGM)

To evaluate the effects of cortisol treatments on FGM, fecal samples were collected in 2015 and 2016 from male and female squirrels fed with 0, 6, 8, and 12 mg cortisol/day (Table 1). Glucocorticoid metabolites from fecal samples were extracted and assayed as previously validated and described (Dantzer et al., 2011, 2010) using a 5α-pregnane-3β,11β,21-triol-20-one enzyme immunoassay (Touma et al., 2003). Intra- and inter-assay CVs for pools diluted 1:250 (n = 13 plates) were 7.4% and 15.4%. For pools diluted 1:500 (n = 13 plates) this was 7.5% and 17.9%. Pools diluted 1:100 (n = 9 plates) had intra and inter-assay CVs of 10.0% and 17.9%, and for pools diluted at 1:700 (n = 9 plates) this was 6.4% and 18.9%. Samples from control (n = 135) or cortisol (n = 237) treated squirrels included those collected before (range: 0-21 days before treatment started, mean ± SE: 7 ± 0.7 days), during, and after treatment (range:1-21 days after treatment, mean ± SE: 9.4 ± 0.8 days, Table 1).

Effects of exogenous cortisol on plasma cortisol concentrations and corticosteroid binding capacity

Non-breeding male squirrels were fed cortisol (8 mg/day) or control treatments for one (n = 40 squirrels) or two weeks (n = 26). The time that squirrels consumed their treatments was estimated by checking buckets at regular intervals between 25 minutes and a few hours (shown as hours:min, mean = 1:53 hrs, SD = 1:12 hrs). Squirrels were either blood sampled the same day they consumed their last treatment (n = 36 squirrels, mean = 3:30 hrs after treatment consumption, range = 0:57-8:55 hrs) or the

154 day after they consumed their last treatment (n = 30, mean = 22:46 hrs after treatment consumption,
155 range = 14:57-30:55 hrs). Note that for 19 next day bleed squirrels, the time of treatment consumption
156 was not recorded. Blood samples obtained within 3 min of squirrels entering a trap (n = 54 samples) are
157 referred to as *true baseline* samples (Romero et al. 2005). If the first blood sample was obtained >3 min
158 after squirrels went into traps (n = 12), this is referred to as *stress-induced* samples. Stress-induced
159 samples are considered to reflect the effects of capture and handling on plasma cortisol concentrations or
160 were taken to see if dexamethasone administration (see below) would reduce plasma cortisol
161 concentrations. Although the time of day we obtained blood samples varied, there was no daytime
162 sampling bias between cortisol treated (between 9:41 and 18:07, mean = 13:23) and control squirrels
163 (between 9:33 and 17:31, mean = 13:50, t-test, $t_{42.15}$= 1.14, p = 0.26). We also conducted a general linear
164 model containing time of day of sampling (b = -0.02, SE = 0.18, t_{14} = -0.12, P = 0.91) and a quadratic
165 term for time of day (b = -0.41, SE = 0.24, t_{14} = -1.68, P = 0.11) and found no significant effect of
166 sampling time on plasma cortisol.

167 Blood samples were obtained from the nailbed and collected into heparinized capillary tubes, and
168 plasma was separated via centrifugation and frozen at -20° C. Total plasma cortisol concentrations were
169 assayed using an ImmuChem coated tube cortisol radioimmunoassay (MP Biomedicals, New York,
170 USA) following the manufacturer's instructions, with the exception that, due to small sample volumes,
171 plasma and tracer volumes of 12.5 µL and 500 µL were used. This assay has already been validated and
172 used to measure plasma cortisol in other rodent species (Karatsoreos et al. 2010; Brooks and Mateo,
173 2013). We validated this kit by first showing linearity and then demonstrating that the assay reliably
174 responds to changes in HPA axis activity through the decreases in plasma cortisol we observed in
175 response to Dex and the increases in plasma cortisol in response to ACTH (Fig. 3). Linearity was tested
176 by pooling several samples and serially diluting these from 1 (neat) to 1:64. Results were plotted,

177 visually inspected, and evaluated with linear regression (R^2 adj = 0.991, $p < 0.001$). According to the
178 manufacturer, the assay detection limit is 1.7 ng/ml, and samples that read below this value (n = 8) were
179 set at 1.7 ng/ml. Most samples were run in duplicate, but because of small plasma volumes only one
180 estimate was obtained for 33.9% of samples. Average standard and sample intra-assay CVs were 7.9%
181 (n = 4 assays). Inter-assay CVs for the five standards provided (10, 30, 100, 300 and 1000 ng/ml
182 cortisol) were 11.1%, 15.4%, 8.8%, 4.0% and 7.7%.

183 Corticosteroid binding capacity (CBG) was measured in plasma stripped of endogenous steroids
184 using dextran-coated charcoal (DCC) and diluted to a final dilution of 1/50 in phosphate buffered saline
185 with 0.1% gelatin (PBS). Three tubes (final volume of 150 µL) were prepared for each sample: two
186 containing 160 nM cortisol (10% 1,2,6,7-^{3}H-cortisol, Perkin Elmer, Waltham, MA, and 90% non-
187 labeled cortisol, C-106, Sigma-Aldrich) to measure total binding, and one containing an additional 4 µM
188 non-labeled cortisol to measure nonspecific binding (primarily by albumin). After incubating tubes
189 overnight, 300 µL of ice-cold DCC was added and left for 15 minutes to strip free cortisol from the
190 plasma mixture. The tubes were then centrifuged at 2000 x *g* at 4 °C for 12 minutes. The supernatant
191 (containing bound cortisol) was decanted into scintillation vials, to which 4 mL of scintillation fluid
192 (Emulsifier-Safe cocktail, Cat. No. 6013389, Perkin Elmer, Groningen, Netherlands) was added. Vials
193 were counted in a scintillation counter. Specific binding by CBG was calculated by subtracting
194 nonspecific binding counts from total binding counts. Specific binding scintillation counts were
195 converted to nM binding by measuring the total counts in the 150 µL of the 160 nM solution and
196 adjusting for the plasma dilution. Some CBG-bound hormone is lost to the DCC during the 15 minute
197 DCC exposure. Using pooled plasma exposed to DCC for 5-20 minutes, we calculated the rate of loss of
198 CBG-bound cortisol (data not shown). From this, we calculated that the 15 minute DCC exposure
199 resulted in the loss of 28.5% of CBG-bound hormone, and all our specific binding measurements were

corrected accordingly. To calculate the percent free cortisol, we estimated free cortisol concentrations (i.e. not bound by CBG) using the total cortisol concentration, the equilibrium dissociation constant for red squirrels of 61.1 nM (Delehanty et al., 2015), and the equation in Barsano and Baumann (1989). As plasma volumes were limited, only 58 samples could be assayed for both CBG/percent free cortisol and total cortisol (see below for details).

Effects of exogenous cortisol on HPA axis reactivity

To determine how our cortisol treatments affected the responsiveness of the HPA axis, we used two different methods. First, we assessed the response to capture and handling ("handling stress") in cortisol-treated and control squirrels by acquiring a series of blood samples starting immediately after they entered the live-trap. Squirrels were bled at intervals of 0-3, 3-6, 6-12 and 18-22 min after trap doors closed either the same day as confirming they ate their last treatment (mean time elapsed = 3:36 hrs, range = 0:57-8:55 hrs, cortisol n = 10, control n = 13) or the next day (mean time elapsed = 22:46, hrs range = 14:57-30:55 hrs, cortisol n = 5, control n = 6).

Second, in a separate set of squirrels, we assessed how cortisol-treated and control squirrels (n = 32) responded to intra-muscular injections of dexamethasone (Dex, a glucocorticoid receptor antagonist) and adrenocorticotropic hormone (i.e., Dex/ACTH challenges). We used previously described protocols (Boonstra and McColl, 2000), with modified concentrations of Dex (3.2 mg/kg) and ACTH (4 IU/kg). Briefly, squirrels were captured and a true baseline blood sample (0-3 min after entering trap) was obtained followed by the acquisition of a second blood sample (stress-induced sample) an average of 12:37 min (min:sec, n = 13, range = 5:00-28:00 min) after the squirrel entered the trap (time of the stress-induced sample was not recorded for 13 different squirrels as traps were checked ~60 min after they were set). Squirrels were then injected with Dex and a third blood sample was acquired 60 min

after injection. Following the acquisition of the third blood sample, squirrels were injected with ACTH and then blood sampled 30 and 60 min after ACTH injection.

Effects of exogenous cortisol on adult squirrel body mass

We assessed how treatment with exogenous cortisol affected body mass of non-breeding females (n = 21 females, 64 body mass measures before treatment and 85 records during treatment) and non-breeding males (n = 47 males, 37 body mass measures before treatment and 28 records during treatment) by live-trapping them approximately once per week and weighing them to the nearest 5g with a spring scale (McAdam et al., 2007). We compared non-breeding squirrel body mass in cortisol treated and control squirrels sampled in 2015 and 2016 before and during the treatments.

Effects of exogenous cortisol on litter survival

As a part of our long-term data collection, we track the reproduction of females during pregnancy and lactation by capturing them, palpating their abdomens to identify pregnancy stage, and expressing milk from their teats to identify if they are lactating (McAdam et al., 2007). We also retrieve pups from their natal nest soon after parturition ("first nest entry") and approximately 25 d after parturition ("second nest entry") to collect a range of data described elsewhere (McAdam et al., 2007). We used these data to identify how treatment with exogenous cortisol affected litter survival in females treated only during pregnancy (n = 71) and in a separate group of females treated only during lactation (n=17) compared to controls. In these comparisons, we also included data on litter fate collected in 2012 from squirrels fed the same dosages (0, 6, 12 mg cortisol/day) for similar periods of time (see Dantzer et al., 2013). When females gave birth, their nests were located an average of 1.6 d after parturition (SD = 1.6 d, range = 0-6 days) and then again 25.5 d after parturition (SD = 1.5 d, range = 21-29 d).

We examined how our treatments affected whether females treated during pregnancy (control n = 24, 6 mg cortisol/day n = 9, 8 mg cortisol/day n = 16, 12 mg cortisol/day n = 22) or lactation (control n = 8, 12 mg cortisol/day n = 9) lost their litters before the first nest entry (via abdominal palpation) or between the first and second nest entry (indicated by cessation of lactation). Females treated during pregnancy were treated from the estimated last third of pregnancy (based on abdominal palpation), until five days post parturition (treatment duration range = 8 – 25 days, mean = 18, SD = 4). Females treated during lactation were treated for 10 continuous days, from days 5 to 15 post parturition (mean = 9.9 d, SD = 0.6, range = 8 -11 days).

Effects of exogenous cortisol on adult squirrel survival

The effects of treatment with exogenous cortisol on survival of adult squirrels was monitored through regular live trapping and behavioral observations (McAdam et al., 2007). Survival data were only available from squirrels studied in 2015 (n = 50, including 41 females and nine males). These squirrels were fed either control treatments (n = 25, 10-26 days, mean = 19, SD = 7) or 12 mg cortisol/day (n = 25, 8-35 days, mean = 20, SD = 7). Although we did not know the ages of all squirrels, there was no age bias between squirrels fed control (eight known ages, mean = 4.05, SD= 1.05 years) and those fed cortisol (nine known ages, mean = 3.97, SD = 0.87 years, $t_{13.8}$= 0.18, p = 0.86). We estimated survival until exactly 1 year after the treatments were stopped.

Statistical analyses

Analyses were conducted using R statistical software (v 3.3.3, R Core Team, 2017). When there were multiple measures for individual squirrels, linear mixed-effects models (LMMs) were conducted using packages ‘lme4’ (v 1.1.10, Bates et al., 2015) and all such models contained ‘squirrel ID’ as a random intercept term. If there were no repeated measures, general linear models (GLM) were used. To

265 make comparisons between groups, we used the 'glht' function in R package 'multcomp' (Hothorn et
266 al., 2017). Model residuals were plotted to check for conformity with homogeneity of variance and
267 normality (Zuur et al., 2010). Where necessary, data were ln transformed. Regression lines were
268 visualized using R package 'visreg' (v 2.2.2, Breheny and Burchett, 2016).

269 We tested effects of treatments on FGM concentrations using LMMs, analyzing female and male
270 data separately due to differences in reproductive states. Models for females included dose (0, 6, 8, 12
271 mg of cortisol/day), reproductive state (non-breeding, pregnant, lactating), Julian date, and whether the
272 squirrel was treated on the sampling day (yes/no) as fixed effects, with an interaction term for dose and
273 treatment (yes/no). Models for males included the same variables (but only doses of 0 and 8 mg) except
274 reproductive state (all were non-breeding).

275 To assess how our treatments affected the responsiveness of the HPA axis, we used two separate
276 LMMs to assess if cortisol treated and control squirrels differed in their plasma cortisol concentrations
277 following 1) our capture and handling stress experiments where we obtained a series of blood samples 2
278 to 28 min after the trap doors closed and 2) our Dex/ACTH challenges. For the LMM to assess the
279 effects of capture and handling on plasma cortisol concentrations, the model included a fixed-effect for
280 treatment (control or cortisol) and the time taken to acquire the blood sample expressed in minutes since
281 the squirrel was trapped (standardized following Schielzeth, 2010). For the LMM to assess plasma
282 cortisol concentrations following the Dex/ACTH challenges, the model included the fixed effect (control
283 or cortisol) and a categorical variable for when the blood sample was obtained ('true baseline', 'stress-
284 induced, '60 minutes after Dex injection' [hereafter; DEX], '30 minutes after ACTH injection'
285 [hereafter; ACTH30], and '60 minutes after ACTH injection' [hereafter; ACTH60]). Two plasma
286 samples with very low binding (<10%) were excluded from the analysis. Some models included
287 squirrels treated for either 1 or 2 weeks, and some included squirrels that were treated in both spring and

autumn (n = 18 squirrels, with treatments switched between periods, with the exception of two squirrels fed GCs twice and one squirrel fed control treatments twice). Where this was the case, treatment duration (1/2 weeks) and whether or not squirrels had been treated before (yes/no) were included in our initial models. Because these two variables (treated for one/two weeks, and whether or not squirrels had been treated previously) were not significant in any of the models, we do not discuss them below.

To assess effects of treatments on CBG concentrations and percent free cortisol, we subset samples collected at different intervals after squirrels entered the traps (effects of handling stress samples) and those from Dex/ACTH challenges. For our handling stress samples, data from samples collected on the same day (n = 12) and the day after (n = 3) the last treatment was consumed were pooled. This model included an interaction between the sampling day (same/next) and treatment (control or 8 mg cortisol/day). Due to limited data (only 58 samples were analyzed for CBG, across all categories), only the effects of treatment (control or 8 mg cortisol/day) on CBG and percent free cortisol were tested for squirrels Dex/ACTH challenged on the same day as consuming their last treatments (n = 16 squirrels). Models for squirrels ACTH challenged the day after consuming their last treatments (n = 27 squirrels) included interactions between sample time (stress-induced, Dex, ACTH30, ACTH60) and treatment (control or 8 mg cortisol/day).

To estimate the total plasma cortisol in a 24 h period, true baseline cortisol was plotted against the time since treatment was consumed. Regression line equations were used to calculate the area under these lines for both control and cortisol treated squirrels, using the 'trapzfun' command in package 'pracma' (Borchers, 2018), and areas under the curve were compared with χ^2 tests.

Data on body mass were subset into those collected in spring (non-breeding females fed 0 or 12 mg cortisol/day) and autumn (non-breeding males fed 0 or 8 mg cortisol/day). Body masses were compared using LMMs including a two-way interaction between treatment, and time (before/during

311 treatment). To assess differences between litter survival (lost/not lost), and adult survival (yes/no) GLMs
312 were applied using binomial errors. Models included a binary fixed effect for treatment (12 mg
313 cortisol/day or control) and sex (only for adult survival). Because the duration of the treatments varied
314 among different squirrels, we also included total days of treatment and an interaction between treatment
315 and treatment duration in all these models to assess how our treatments affected body mass and adult or
316 litter survival. Dispersion parameters (using R package blemco, Korner-Nievergelt et al., 2015) between
317 0.75 and 1.4 were taken to accept overdispersion was not problematic.

318

319

320 **Results**

321 *Effects of treatments on fecal glucocorticoid metabolite concentrations*

322 Overall, squirrels fed cortisol treatments (6, 8, 12 mg/day) had significantly higher FGM
323 concentrations than when they were not being fed, but the magnitude of increase depended on the
324 dosage ($F_{3,263.8}$= 11.5, $p < 0.001$, Fig. 1). Both female and male control squirrels fed plain peanut butter
325 had similar FGM concentrations when they were being fed their treatments compared to when they were
326 not being fed their treatments (females: b = -0.03, SE = 0.15, z = 0.22, p = 1.0; males: b =0.14, SE=
327 0.36, z = 0.41, p = 0.96). FGM concentrations in both females and males fed 6, 8, or 12 mg cortisol/day
328 were significantly higher compared when they were being fed compared to when they were not being
329 fed (6 mg: b = 0.78, SE = 0.28, z = 2.8, p = 0.032; 8 mg: females: b = 0.79, SE = 0.21, z = 3.9, p <
330 0.001; males: b =1.37, SE= 0.49, z = 2.82, p = 0.013; 12 mg: b = 1.39, SE = 0.17, z = 8.0, p < 0.001).
331 Concentrations of FGM during treatment in female squirrels treated with 12 mg vs 8 mg (b = 0.39, SE =
332 0.28, z = 1.4, p = 0.63), 6 mg vs 8 mg (b = 0.36, SE = 0.36, z = 1.0, p = 0.88), and 6 mg vs 12 mg
333 cortisol/day (b = 0.74, SE = 0.35, z = 2.1, p = 0.18) were not significantly different. Julian date did not

affect FGM concentrations in females ($F_{1,284.9} = 3.44$, $p = 0.06$) or males ($F_{1,15.2} = 0.73$, $p = 0.41$). Reproductive condition did not affect FGM in this dataset, possibly because of limited sample numbers on some reproductive states (see Table 1, $F_{2,105.0} = 1.37$, $p = 0.26$).

Effects of treatments on total plasma cortisol concentrations over 24 hr period

We plotted true baseline plasma cortisol concentrations against the time since treatment was consumed to estimate the area under these lines for both control and cortisol treated squirrels. The area under these lines was used to estimate the total plasma cortisol concentrations over a 24 hr period. Overall, we estimated that cortisol treated squirrels experienced significantly higher plasma cortisol (total area = 7907.4 units) than controls (total area = 4110.8 units) over a 24 h period ($\chi^2 = 614.4$, DF = 1, $p < 0.001$, Fig. 2).

Responsiveness of HPA axis to capture and handling in squirrels sampled same day or day after last treatment

The effects of capture and handling on plasma cortisol concentrations were significantly different between control and cortisol treated squirrels, in addition to whether the squirrels were sampled on the same day or day after their last treatment. In squirrels sampled the same day as receiving their last treatment, plasma cortisol concentrations were generally higher in cortisol treated squirrels compared to controls, but their responsiveness to capture and handling differed (Fig. 3A). In control squirrels sampled the same day as receiving their last treatment, plasma cortisol concentrations significantly increased as handling time increased ($b = 0.31$, $SE = 0.11$, $t = 2.8$, DF = 61.4, $p = 0.007$, Fig. 3A) whereas they declined as handling time increased in cortisol treated squirrels ($b = -0.56$, $SE = 0.17$, $t = -3.3$, DF = 61.4 $p = 0.002$, Fig. 3A).

In squirrels sampled the day after receiving their last treatment, plasma cortisol concentrations were generally lower in cortisol treated squirrels than in control squirrels ($b = -1.8$, $SE = 0.81$, $t = -2.2$,

357 DF = 8.4, p = 0.057, Fig. 3B), though this difference was not significant. Handling time increased
358 plasma cortisol concentrations in both control and cortisol treated squirrels (b = 0.42, SE = 0.20, t = 2.1,
359 DF = 19.2, p = 0.048) and this was not affected by treatment (b = 0.40, SE = 0.1.4, DF= 18.7, p = 0.16,
360 Fig. 3B).

361 *Responsiveness of HPA axis to Dex/ACTH challenges in squirrels sampled on the same day as*
362 *consuming last treatment*

363 In squirrels sampled on the same day as consuming their last treatment, HPA axis responsiveness
364 to our Dex/ACTH challenges differed between control and cortisol treated squirrels ($F_{9,35.5} = 6.4$, p <
365 0.001, Fig. 3A). Squirrels treated with cortisol (8 mg/day) had significantly higher true baseline cortisol
366 concentrations (611.4±104.4 ng/ml) than control squirrels (214.5±41.3 ng/ml, b = 394.7, SE = 88.7, z =
367 4.4, p < 0.001, Fig. 3A) but cortisol treated and control squirrels had similar stress-induced plasma
368 cortisol concentrations (b = 15.9, SE = 114.5, z = 0.14, p = 1, Fig. 3A). Both cortisol treated (205.8±50.2
369 ng/ml) and control (292.2±44.5 ng/ml) squirrels responded to Dex, as indicated by the reductions in their
370 plasma cortisol concentrations 60 min after the Dex injection compared to stress-induced plasma cortisol
371 concentrations, although these reductions in were not significant (control: b = -33.9, SE = 81.2, z = -
372 0.44, p = 1.0; cortisol treated: b = -139.6, SE = 88.7, z = -1.57, p = 0.62). Control squirrels had
373 significantly higher plasma cortisol concentrations in samples taken 30 minutes after ACTH injection
374 compared to those obtained 60 min after the Dex injection (604.9±93.6 ng/ml, b = 321.1, SE = 81.2, z =
375 3.97, p < 0.001) but not in samples taken 60 min after ACTH injection (404.6±76.3 ng/ml, b = -213.4,
376 SE = 85.1, z = -2.51, p = 0.10). In cortisol treated squirrels, plasma cortisol concentrations were
377 unaffected by ACTH as plasma cortisol concentrations in samples taken 30 minutes after ACTH
378 injection (153.5±22.1 ng/ml, b = -51.8, SE = 88.7, z = -0.58, p = 1.0) and 60 minutes after ACTH

injection (163.1±36.7 ng/ml, b = 9.5, SE = 83.5, z = 0.11, p =1.0) were no different from those obtained 60 min after the Dex injection.

Responsiveness of HPA axis to Dex/ACTH challenges in squirrels sampled the day after consuming their last treatment

In squirrels sampled the day after consuming their last treatment, HPA axis responsiveness to our Dex/ACTH challenges differed between control and cortisol treated squirrels ($F_{4,46.5} = 9.2$, $p < 0.001$, Fig. 3B). Cortisol treated squirrels that were sampled the day after consuming their last treatment had lower plasma cortisol concentrations than controls at all sampling times (Fig. 3B). Although true baseline plasma cortisol concentrations did not differ between cortisol treated and control squirrels (b = 48, SE = 82.7, z = 0.6, p = 0.99, Fig. 3B), stress-induced plasma cortisol concentrations (35.9±21.2 ng/ml) were on average 92.3% lower in cortisol treated squirrels than in control squirrels (468.5±34.8 ng/ml, b = -404.8, SE = 67.4, z = -6.0, p < 0.001, Fig. 3B). Plasma cortisol concentrations after the Dex injection were an average of 76.4% lower in cortisol treated squirrels (56.5±34.6 ng/ml) than controls (239.8±24.8 ng/ml, b = -183.3, SE = 53.7, z = -3.4, p = 0.006). Cortisol treated squirrels had plasma cortisol concentrations (mean = 97.9±19.9 ng/ml) that were on average 80.4% lower than in control squirrels (mean = 498.8±34.6 ng/ml, b = 398.1, SE = 57.5, z = -6.9, p < 0.001) 30 min after the ACTH injection. This difference remained 60 min after the ACTH injection (cortisol mean = 101.7±11.4 ng/ml, control mean = 469.0±89.2 ng/ml, a difference of 78.3%, b = -368.2, SE = 56.8, z = -6.5, p < 0.001, Fig. 3B).

In squirrels sampled the day after receiving their last treatment, control squirrels (b = -211.9, SE = 53.7, z = -3.9, p < 0.001), but not cortisol treated squirrels (b = 9.6, SE = 49.4, z = 0.2, p = 1.0), had significantly lower plasma cortisol concentrations 60 minutes after Dex injections compared to stress-induced concentrations. Thirty minutes after the ACTH injection (ACTH30), control squirrels (b

= 257.3, SE = 45.6, z = 5.6, $p < 0.001$), had significantly higher plasma cortisol concentrations than 60 minutes after Dex injections. In cortisol treated squirrels, plasma cortisol concentrations were higher at 30 minutes after ACTH injection (ACTH30) than 60 minutes after Dex injection, but this was not significant (b = 42.6, SE = 44.7, z = 0.95, p = 0.95, Fig. 3B).

Effects of cortisol treatment on plasma CBG and free cortisol concentrations

Squirrels treated with cortisol had significantly lower CBG concentrations in plasma samples obtained during our Dex/ACTH challenges compared to controls regardless of whether the samples were obtained on the same day as consuming their last treatments ($F_{1,5.0} = 51.0$, $p < 0.001$) or the day after consuming their last treatment ($F_{1,12.9} = 29.3$, $p < 0.001$, Fig. 6A). Consequently, squirrels treated with cortisol had significantly higher free cortisol concentrations in plasma samples obtained during the Dex/ACTH challenges regardless of whether they were sampled on the same day ($F_{1,5.0} = 7.6$, $p = 0.04$) or the day after consuming their last treatments ($F_{1,12.1} = 15.3$, $p = 0.002$, Fig. 6B). CBG concentrations in plasma samples obtained during the Dex/ACTH challenges from squirrels sampled the day after consuming their last treatment did not vary among the different sample types (stress-induced, Dex, ACTH30, ACTH60: $F_{3,6.2} = 1.0$, $p = 0.46$), nor did percent free cortisol ($F_{3,12.3} = 0.5$, $p = 0.71$).

In plasma samples obtained from squirrels at regular intervals after they entered our live traps, CBG concentrations were significantly lower in plasma samples acquired from cortisol treated squirrels compared to controls ($F_{1,7.0} = 24.5$, $p = 0.002$, Fig. 6A), but percent free cortisol did not differ between cortisol treated and control squirrels ($F_{7.0} = 2.3$, $p = 0.17$). The effect of treatment on plasma CBG concentrations ($F_{7.0} = 1.3$, $p = 0.30$) and percent free cortisol ($F_{1,7.0} = 1.9$, $p = 0.21$) was not different between squirrels sampled the same day or the day after last treatment.

Effects of treatments on body mass

Both control and cortisol treated females (effect of treatment period, b = 11.9, SE = 3.7, $t_{87.4}$ = 3.26, p = 0.001) but not males (b = 1.33, SE = 3.2, $t_{32.4}$ = 0.41, p = 0.68) were heavier when they were being treated (n = 23 females, n = 11 males) compared to before they were being treated (n = 24 females, n = 17 males). However, control and cortisol treated females and males did not differ in body mass while they were being treated, as indicated by the lack of significant interactions between treatment and treatment period in both females (b = -3.83, SE = 4.9, $t_{87.2}$ = -0.79, p = 0.43) and males (b = 6.5, SE = 4.84, $t_{29.5}$ = 1.34, p = 0.19). Although females and males in both treatment groups (control or cortisol) were treated for varying lengths of time, treatment duration did not affect body mass as indicated by the lack of significant interactions between treatment and treatment duration for both females (b = -1.36, SE = 12.1, $t_{17.8}$ = -0.11, p = 0.91) and males (b = 1.73, SE = 3.8, $t_{49.7}$ = 0.45, p = 0.65).

Effects of treatments on litter survival

For females that were treated during pregnancy, there was no significant difference in litter survival rates before the first nest entry between females treated with cortisol (6, 8, 12 mg/day) during pregnancy (14/47 litters lost) and controls (7/24 litters lost; z = 0.28, p = 0.78). Similarly, there were also no significant difference in litter survival between the first and second nest entry for females treated with cortisol during pregnancy (17/47 litters lost) and controls (4/24 litters lost: z = 0.01, p = 0.99). Even though control (range = 11-23 d) and cortisol treated (range = 8-25 d) pregnant females were treated for different lengths of time, there was no effect of treatment duration for both control and cortisol treated females on litter survival prior to the first nest entry (treatment x treatment duration, z = -0.39, p = 0.69) and between the first and second nest entry (z = -0.16, p = 0.87).

For females treated during lactation (n = 17), there was no significant difference in litter survival between the first and second nest entry, with 1/8 control females losing their litter and 3/9

447 cortisol fed (12 mg/day) females losing their litter ($z = 0.001$, $p = 1$). Although treatment duration varied
448 for control (range = 10-11 d) and cortisol treated (range = 8-10 d) lactating females, there was once
449 again no effect of treatment duration for both control and cortisol treated females (treatment x treatment
450 duration, $z = -0.005$, $p = 0.97$).

451 *Effects of treatments and treatment duration on adult survival*

452 There was no difference in survival to one year following cessation of the treatments between
453 controls (18/25 survived to one year) and those fed cortisol (14/25 survived to one year; $z = -1.09$, $p =$
454 0.27). Although the duration of the cortisol or control treatments slightly varied among cortisol treated
455 (range = 8-35 d) or control (range = 10-26 d) squirrels, treatment duration did not affect adult survival as
456 indicated by the lack of significant interaction between treatment and treatment duration ($z = 0.63$, $p =$
457 0.53). There was no difference in survival between males (7/9 survived to one year), and females (25/41
458 survived to one year, $z = 1.19$, $p = 0.24$).

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460

461 **Discussion**

462 Our results on cortisol manipulations in wild red squirrels, spanning a range of dosages, life
463 history stages, and including both sexes, provide important information regarding the response of wild
464 animals to such hormone manipulation including the possible fitness consequences of hormone
465 manipulation. Squirrels treated with cortisol had higher FGM concentrations and true baseline plasma
466 cortisol concentrations over a 24 h period. However, similar to studies in humans, laboratory rodents, or
467 wild birds (see Introduction), our results highlight that exogenous GCs can cause the adrenals to stop
468 responding to handling stress or pharmaceutical (Dex/ACTH) challenges. Although we documented that

469 treatment with exogenous GCs affected the responsiveness of the HPA axis, these effects were short-
470 lived and did not affect fitness proxies, including body mass, and offspring or adult survival.

471 Our results indicate that concentrations of CBG were significantly reduced in squirrels treated
472 with cortisol for one or two weeks, suggesting that chronically elevated GCs reduce CBG
473 concentrations. This reduction in CBG may result in a higher bioavailability of plasma cortisol (Breuner
474 et al., 2013). Previous studies in rats have shown that administration of exogenous GCs can inhibit the
475 rate of CBG production and secretion in the liver (Feldman et al., 1979), and one study found that, 24
476 hours after acute stress, CBG concentrations were reduced in rats (Fleshner et al., 1995). Chronic
477 elevations in GCs has also been shown to lead to reduced CBG concentrations in most species studied to
478 date (Armario et al., 1994; Breuner et al., 2013). A previous study found that CBG concentrations in red
479 squirrel plasma started to decrease as quickly as four hours after the start of Dex/ACTH challenges,
480 suggesting that although high concentrations of CBG may buffer squirrels from the effects of high
481 concentrations of free cortisol caused by acute stressors, these concentrations decline rapidly when the
482 duration of the stressor is longer than a few hours (Boonstra and McColl, 2000). However, this does not
483 seem to carry any noticeable cost as there were no changes in body mass or litter and adult survival in
484 response to our treatments.

485 Previous reviews have emphasized the importance of maintaining hormone concentrations
486 within a physiological range when performing hormone manipulations (Crossin et al., 2016; Fusani,
487 2008; Zera, 2007). Studies in mammals have shown that acute experimental challenges can cause
488 increases in plasma cortisol that are comparable to those achieved by our treatments. For example, in
489 both laboratory rats and wild animals, physical restraint, open field trials, and maze tests may cause >10
490 fold increases in plasma GCs (Cockrem, 2013). In our study, the highest recorded true baseline plasma
491 cortisol concentration in cortisol treated squirrels was approximately seven times higher than the

492 average control true baseline plasma cortisol concentration. This indicates that the increase in plasma
493 cortisol caused by our 8 mg cortisol/day treatment is within the physiological range for a squirrel.
494 However, it is possible that the duration of elevated plasma cortisol caused by our treatments is longer
495 than that caused by a natural ecological factor that elicits an increase in plasma cortisol. Studies in rats
496 show that plasma GCs increase quickly in response to acute stress, but returns to baseline concentrations
497 within 2-5 hours after the stressor is removed (Marin et al., 2007; Mizoguchi et al., 2001), but in cortisol
498 treated squirrels, plasma cortisol remained elevated, compared to control squirrels, for an estimated 17
499 hours post-treatment (Fig. 2).

500 Our results highlight the importance of regularly provisioning individuals with treatments to
501 sustain increases in hormone concentrations. Although we found that squirrels fed cortisol had
502 significantly higher concentrations of plasma cortisol over a 24 hr period than the controls, it was
503 important to provision individuals with the treatments every 24 hrs. This was because plasma cortisol in
504 cortisol treated squirrels did decrease to concentrations well below those of control squirrels >20 hrs
505 after consuming their treatments. When it is feasible, daily supplementation may be effective in
506 maintaining sustained elevations in hormone concentrations and provide an alternative to other methods
507 like implants that carry some disadvantages (Sopinka et al., 2015). For example, Torres-Medina et al.
508 (2018) showed that silastic implants, time release pellets, or osmotic pumps that contain corticosterone
509 can also suppress the responsiveness of the HPA axis in birds, which could decrease overall exposure to
510 circulating corticosterone. Our study shows that even regular provisioning of exogenous GCs rather than
511 implants, such as through food in our study (see also Dantzer et al., 2017) or through other methods
512 (Vitousek et al. 2018), may also decrease the activity of the HPA axis and cause a reduced ability to
513 mount an increase in circulating GCs in response to an environmental challenge.

514 Our results also highlight the potentially adverse consequences that may occur when ending
515 hormone manipulations in wild animals. When sampled less than a day after the end of cortisol
516 treatment, squirrels did not respond to handling stress or ACTH injection, and appeared to have
517 impaired adrenal function and lower CBG concentrations. Data collected from cortisol treated squirrels
518 the day after consuming their last treatment showed that plasma cortisol was very low compared to
519 control squirrels, suggesting exogenous GCs have been excreted but endogenous GCs were being
520 produced at a lower rate than in control animals. However, plasma cortisol did increase with handling
521 stress, suggesting some recovery of adrenal function within 24 hrs of stopping the treatments. Our
522 results suggest that the adrenal gland may need time to recover from treatment, and endogenous cortisol
523 production may not return to pre-treatment levels for several days.
524 Hormone manipulations can provide powerful tools to study relationships between hormones
525 and life history traits, and in recent years methods have been developed to achieve this (Sopinka et al.,
526 2015). Many studies aim to experimentally elevate GCs to test the "cort-fitness hypothesis", which
527 proposes that elevations in baseline GCs decreases survival or reproduction (Bonier et al., 2009), or to
528 document the effects of elevated GCs on behavior or life history traits (Crossin et al., 2016). We show
529 that elevation of plasma cortisol concentrations within the physiological range for 1-2 weeks had
530 profound effects on measures of HPA axis reactivity and CBG concentrations. This is similar to the
531 results of Torres-Medina et al. (2018) that showed that treatment with corticosterone implants can also
532 cause reduced corticosterone levels in response to capture and handling. However, our study took the
533 result from Torres-Medina et al. (2018) one step further as we showed how treatment with exogenous
534 GCs suppresses CBG concentrations and we also investigated the possibility of fitness consequences of
535 reduced HPA axis reactivity. Despite these observed shifts in the functionality of the neuroendocrine
536 stress axis and the sustained elevations in GCs, we found no change in body mass or offspring and adult

537 survival. This indicates that some species can tolerate bouts of increased GCs and rapid reorganization
538 of the stress axis without negatively impacting survival and reproduction.
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542 **References**

543 Arabi, Y.M., Aljumah, A., Dabbagh, O., Tamim, H.M., Rishu, A.H., Al-Abdulkareem, A., Al Knawy,
544 B., Hajeer, A.H., Tamimi, W., Cherfan, A., 2010. Low-dose hydrocortisone in patients with
545 cirrhosis and septic shock: A randomized controlled trial. Can. Med. Assoc. J. 182, 1971–1977.
546 doi:10.1503/cmaj.090707

547 Arlt, W., Allolio, B., 2003. Adrenal insufficiency. Lancet 361, 1881–1893. doi:10.1016/S0140-
548 6736(03)13492-7

549 Armario, A., Giralt, M., Martí, O., Gavaldà, A., Hidalgo, J., Hsu, B.R.., Kuhn, R.W., 1994. The effect of
550 acute and chronic ACTH administration on pituitary-adrenal response to acute immobilization
551 stress. Relationship to changes in corticosteroid-binding globulin. Endocr. Res. 20, 139–149.

552 Axelrod, L., 1976. Glucocorticoid therapy. Medicine (Baltimore). 55, 39–65.

553 Barsano, C.P., Baumann, G., 1989. Simple algebraic and graphic methods for the apportionment of
554 hormone (and receptor) into bound and free fractions in binding equilibria; or how to calculate
555 bound and free hormone? Endocrinology 124, 1101–1106.

556 Bates, D., Maechler, M., Bolker, B., Walker, S., Christensen, R.H.., Singmann, H., Dai, B.,
557 Grothendieck, G., Green, P., 2015. Package "lme4."

558 Bonier, F., Martin, P.R., Moore, I.T., Wingfield, J.C., 2009. Do baseline glucocorticoids predict fitness?
559 Trends Ecol. Evol. 24, 634–642. doi:10.1016/j.tree.2009.04.013

560 Boonstra, R., McColl, C.J., 2000. Contrasting stress response of male Arctic ground squirrels and red
561 squirrels. J. Exp. Zool. Part A Ecol. Genet. Physiol. 286, 390–404.

562 Borchers, H.., 2018. Package "pracma." doi:https://cran.r-project.org/package=pracma

563 Boutin, S., Wauters, L.A., McAdam, A.G., Humphries, M.M., Tosi, G., Dhondt, A.A., 2006.
564 Anticipatory reproduction and population growth in seed predators. Science (80-.). 314, 1928–
565 1930. doi:10.1126/science.1135520

566 Breheny, P., Burchett, W., 2016. Package "visreg." doi:https://cran.r-
567 project.org/web/packages/visreg/vis

568 Breuner, C.W., Delehanty, B., Boonstra, R., 2013. Evaluating stress in natural populations of
569 vertebrates: total CORT is not good enough. Funct. Ecol. 27, 24–36. doi:10.1111/1365-2435.12016

570 Breuner, C.W., Patterson, S.., Hahn, T.., 2008. In search of relationships between the acute
571 adrenocortical response and fitness. Gen. Comp. Endocrinol. 157, 288–95.

572 Broide, J., Soferman, R., Kivity, S., Golander, A., Dickstein, G., Spirer, Z., Weisman, Y., 1995. Low-
573 dose adrenocorticotropin test reveals impaired adrenal function in patients taking inhaled
574 corticosteroids. J. Clin. Endocrinol. Metab. 80, 1243–1246. doi:10.1210/jcem.80.4.7714095

575 Casolinia, P., Cigliana, G., Alemaa, G.., Ruggieria, V., Angeluccia, L., Catalania, A., 1997. Effect of
576 increased maternal corticosterone during lactation on hippocampal corticosteroid receptors, stress
577 response and learning in offspring in the early stages of life. Neuroscience 79, 1005–1012.

578 Catalani, A., Casolini, P., Ciglianab, G., Scaccianoce, S., Consoli, C., Cincque, C., Zuena, A..,
579 Angelucci, L., 2002. Maternal corticosterone influences behavior, stress response and
580 corticosteroid receptors in the female rat. Pharmacol. Biochem. Behav. 73, 105–114.

581 Cockrem, J.F., 2013. Individual variation in glucocorticoid stress responses in animals. Gen. Comp.
582 Endocrinol. 181, 45–58. doi:10.1016/j.ygcen.2012.11.025

583 Crossin, G.T., Love, O.P., Cooke, S.J., Williams, T.D., 2016. Glucocorticoid manipulations in free-
584 living animals: Considerations of dose delivery, life-history context and reproductive state. Funct.
585 Ecol. 30, 116–125. doi:10.1111/1365-2435.12482

586 Dantzer, B., Goncalves, I.B., Spence-Jones, H.C., Bennett, N.C., Heistermann, M., Ganswindt, A.,
587 Dubuc, C., Gaynor, D., Manser, M.B., Clutton-Brock, T.H. 2017. The influence of stress hormones
588 and aggression on cooperative behaviour in subordinate meerkats. Proc. Roy. Soc. B. 284,
589 20171248. doi: 10.1098/rspb.2017.1248

590 Dantzer, B., McAdam, A.G., Palme, R., Boutin, S., Boonstra, R., 2011. How does diet affect fecal
591 steroid hormone metabolite concentrations? An experimental examination in red squirrels. Gen.
592 Comp. Endocrinol. 174, 124–131. doi:10.1016/j.ygcen.2011.08.010

593 Dantzer, B., Mcadam, A.G., Palme, R., Fletcher, Q.E., Boutin, S., Humphries, M.M., Boonstra, R.,
594 2010. Fecal cortisol metabolite levels in free-ranging North American red squirrels: Assay
595 validation and the effects of reproductive condition. Gen. Comp. Endocrinol. 167, 279–286.
596 doi:10.1016/j.ygcen.2010.03.024

597 Dantzer, B., Newman, A.E.M., Boonstra, R., Palme, R., Boutin, S., Humphries, M.M., Mcadam, A.G.,
598 2013. Density triggers maternal hormones that increase adaptive offspring growth in a wild
599 mammal. Science 340, 1215–1217. doi:10.1126/science.1235765

600 Dantzer, B., Westrick, S.E., van Kesteren, F., 2016. Relationships between endocrine traits and life
601 histories in wild animals: insights, problems, and potential pitfalls. Integr. Comp. Biol. 56, 185–
602 197. doi:10.1093/icb/icw051

603 De Nardo, D.F., Sinervo, B., 1994. Effects of corticosterone on activity and home-range size of free-

604 ranging male lizards. Horm. Behav. 28, 53–65. doi:10.1006/hbeh.1994.1005

605 Delehanty, B., Hossain, S., Jen, C.C., Crawshaw, G.J., Boonstra, R., 2015. Measurement of free
606 glucocorticoids: quantifying corticosteroid-binding globulin binding affinity and its variation
607 within and among mammalian species. Conserv. Physiol. 3, 1–13. doi: 10.1093/conphys/cov020

608 Ebensperger, L.A., Ramírez-Estrada, J., León, C., Castro, R.A., Tolhuysen, L.O., Sobrero, R., Quirici,
609 V., Burger, J.M., Soto-Gamboa, M., Hayes, L.D., 2011. Sociality, glucocorticoids and direct fitness
610 in the communally rearing rodent, Octodon degus. Horm. Behav. 60, 346–352.
611 doi:10.1016/j.yhbeh.2011.07.002

612 Feiwel, M., James, V.H.., Barnett, E.., 1969. Effect of potent topical steroids on plasma cortisol levels of
613 infants and children with eczema. Lancet 293, 485–487. doi:10.1016/S0140-6736(69)91588-8

614 Feldman, D., Mondon, C.., Horner, J.., Weiser, J.N., 1979. Glucocorticoid and estrogen regulation of
615 corticosteroid-binding globulin production by rat liver. Am. J. Physiol. - Endocrinol. Metab. 237,
616 493–499.

617 Fleshner, M., Deak, T., Spencer, R.L., Laudenslager, M.L., Watkins, L.R., Maier, S.., 1995. A long-term
618 increase in basal levels of corticosterone and a decrease in corticosteroid-binding globulin after
619 acute stressor exposure. Endocrinology 136, 5336–5342.

620 Fusani, L., 2008. Endocrinology in field studies: Problems and solutions for the experimental design.
621 Gen. Comp. Endocrinol. 157, 249–253. doi:10.1016/j.ygcen.2008.04.016

622 Hothorn, T., Bretz, F., Westfall, P., Heiberger, R.., Schuetzenmeister, A., Scheibe, S., 2017. Package
623 "multcomp." doi:https://cran.r-project.org/package=multcomp

624 Jacobs, S., Pullan, P.., Potter, J.., Shenfield, G.., 1983. Adrenal suppression following extradural

625 steroids. Anaesthesia 38, 953–956. doi:10.1111/j.1365-2044.1983.tb12025.x

626 Karatsoreos, I.N., Bhagat, S.M., Bowles, N.P., Weil, Z.M., Pfaff, D.W., McEwen, B.S., 2010. Endocrine
627 and physiological changes in response to chronic corticosterone: A potential model of the
628 metabolic syndrome in mouse. Endocrinology 151, 2117–2127. doi:10.1210/en.2009-1436

629 Kirwan, J.R., Bijlsma, J.W., Boers, M., Shea, B., 2007. Effects of glucocorticoids on radiological
630 progression in rheumatoid arthritis (Review). Cochrane Database Syst. Rev. 1, 1–86.
631 doi:10.1002/14651858.CD006356

632 Korner-Nievergelt, F., Roth, T., von Felten, S., Guelat, J., Almasi, B., Korner-Nievergelt, P., 2015.
633 Package "blemeco." doi:https://cran.r-project.org/web/packages/blmeco/blme

634 Lussier, A.L., Caruncho, H.J., Kalynchuk, L.E., 2009. Repeated exposure to corticosterone, but not
635 restraint, decreases the number of reelin-positive cells in the adult rat hippocampus. Neurosci. Lett.
636 460, 170–174. doi:10.1016/j.neulet.2009.05.050

637 Marin, M.T., Cruz, F.C., Planeta, C.S., 2007. Chronic restraint or variable stresses differently affect the
638 behavior, corticosterone secretion and body weight in rats. Physiol. Behav. 90, 29–35.
639 doi:10.1016/j.physbeh.2006.08.021

640 Mateo, J.., 2008. Inverted-U shape relationship between cortisol and learning in ground squirrels.
641 Neurobiol. Learn. Mem. 89, 582–590.

642 McAdam, A.G., Boutin, S., Sykes, A.K., Humphries, M.M., 2007. Life histories of female red squirrels
643 and their contributions to population growth and lifetime fitness. Ecoscience 14, 362–369.
644 doi:10.2980/1195-6860(2007)14[362:LHOFRS]2.0.CO;2

645 Meikle, A.W., Tyler, F.H., 1977. Potency and duration of action of glucocorticoids. Am. J. Med. 63,

200–207. doi:10.1016/0002-9343(77)90233-9

Mizoguchi, K., Yuzurihara, M., Ishige, A., Sasaki, H., Chui, D.., Tabira, T., 2001. Chronic stress differentially regulates glucocorticoid negative feedback response in rats. Psychoneuroendocrinology 26, 443–459. doi:10.1016/S0306-4530(01)00004-X

Morris, H.., Jorgensen, J.., 1971. Recovery of endogenous pituitary-adrenal function in corticosteroid-treated children. J. Pediatr. 79, 480–488. doi:10.1016/S0022-3476(71)80163-4

R Core Team, 2017. R version 3.3.3. doi:https://www.r-project.org/

Sapolsky, R.M., Romero, L.M., Munck, A.U., 2000. How do glucocorticoids influence stress responses? Integrating permissive, suppressive, stimulatory, and preparative actions. Endocr. Rev. 21, 55–89. doi:10.1210/edrv.21.1.0389

Schielzeth, H., 2010. Simple means to improve the interpretability of regression coefficients. Methods Ecol. Evol. 1, 103–113. doi:10.1111/j.2041-210X.2010.00012.x

Sopinka, N.M., Patterson, L.D., Redfern, J.C., Pleizier, N.K., Belanger, C.B., Midwood, J.D., Crossin, G.T., Cooke, S.J., 2015. Manipulating glucocorticoids in wild animals: Basic and applied perspectives. Conserv. Physiol. 3, 1–16. doi:10.1093/conphys/cov031

Streck, W., Lockwood, M.., 1979. Pituitary adrenal recovery following short-term suppression with corticosteroids. Am. J. Med. 66, 910–914. doi:10.1016/0002-9343(79)90444-3

Torres-Medina, F., Cabezas, S., Marchant, T.A., Wikelski, M., Romero, L.M., Hau, M., Carrete, M., Tella, J.L., Blas, J. 2018. Corticosterone implants produce stress-hyporesponsive birds. J. Exp. Biol. *in press* doi: 10.1242/jeb.173864

666 Touma, C., Sachser, N., Mostl, E., Palme, R., 2003. Effects of sex and time of day on metabolism and
667 excretion of corticosterone in urine and feces of mice. Gen. Comp. Endocrinol. 130, 267–278.
668 doi:10.1016/S0016-6480(02)00620-2

669 Vitousek, M.N., Taff, C.C., Ardia, D.R., Stedman, J.M., Zimmer, C., Salzman, T.C., Winkler, D.W.
670 2018. The lingering impact of stress: brief acute glucocorticoid exposure has sustained, dose-
671 dependent effects on reproduction. Proc. Roy. Soc. B 285, 20180722. doi: 10.1098/rspb.2018.0722

672 Wingfield, J.C., Maney, D.L., Breuner, C.W., Jacobs, J.D., Lynn, S., Ramenofsky, M., Richardson,
673 R.D., 1998. Ecological bases of hormone - behavior interactions : The "emergency life history
674 stage." Avian Endocrinol. F. Investig. Methods 38, 191–206. doi:10.1093/icb/38.1.191

675 Zera, A.J., 2007. Endocrine analysis in evolutionary-developmental studies of insect polymorphism:
676 Hormone manipulation versus direct measurement of hormonal regulators. Evol. Dev. 9, 499–513.
677 doi:10.1111/j.1525-142X.2007.00181.xfun

678 Zuur, A.F., Ieno, E.N., Elphick, C.S., 2010. A protocol for data exploration to avoid common statistical
679 problems. Methods Ecol. Evol. 1, 3–14. doi:10.1111/j.2041-210X.2009.00001.x

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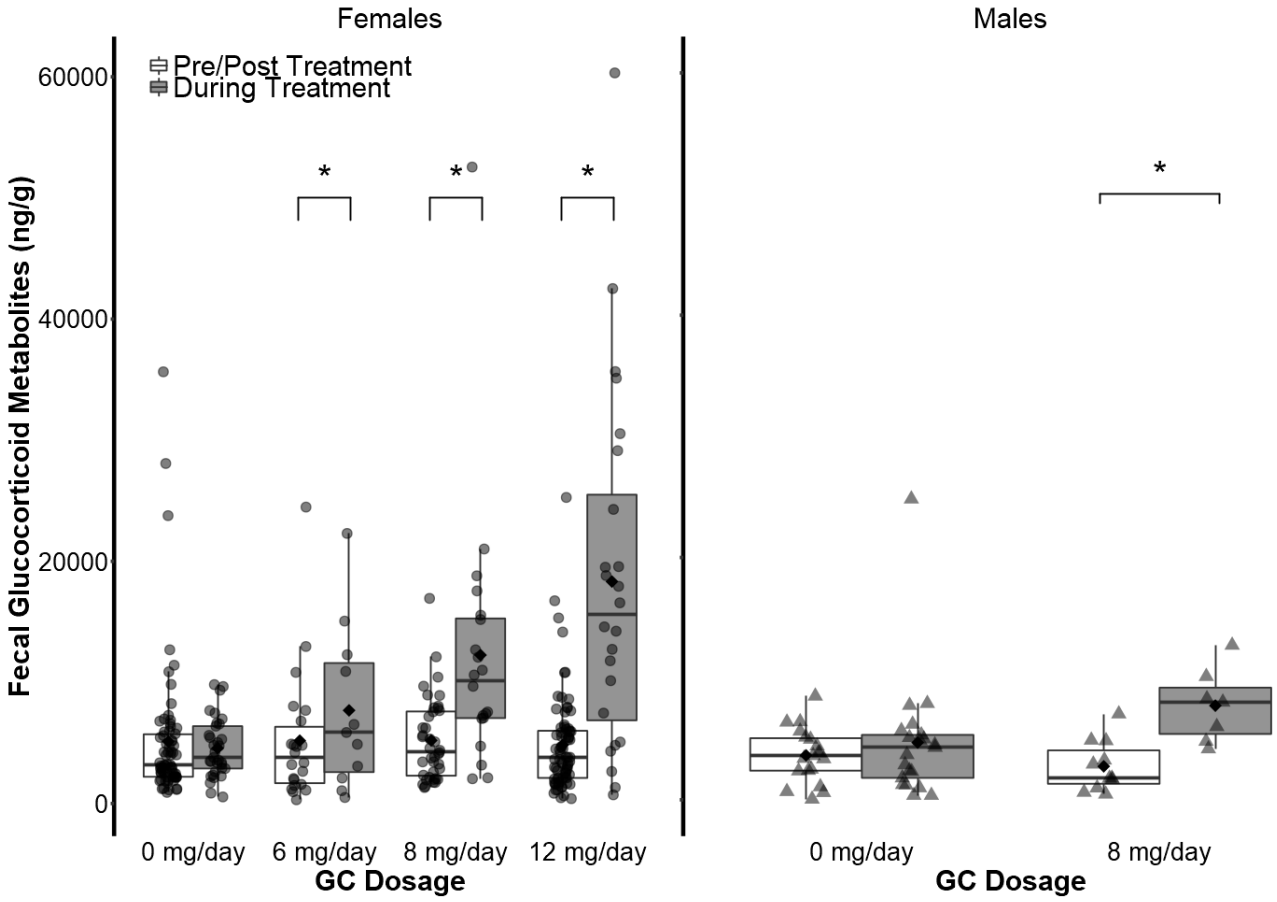
682 **Figures**

683 **Fig 1**. FGM concentrations in squirrels treated with control (0 mg/day) or with cortisol (6, 8, or 12
684 mg/day) treatments. Asterisks (*) indicate significant differences ($p < 0.05$). Upper and lower hinges
685 correspond to the first and third quartiles. Upper/lower whiskers extend from the hinge to the
686 highest/lowest value that is within 1.5x the interquartile range. White diamonds indicate means.



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691 **Fig. 2.** True baseline plasma cortisol concentrations in squirrels fed cortisol (GC, 8 mg/day) and controls
692 (0 mg/day) sampled between 1 to 31 hours after confirming they consumed their last treatments.
693 Different points correspond to different individual squirrels.

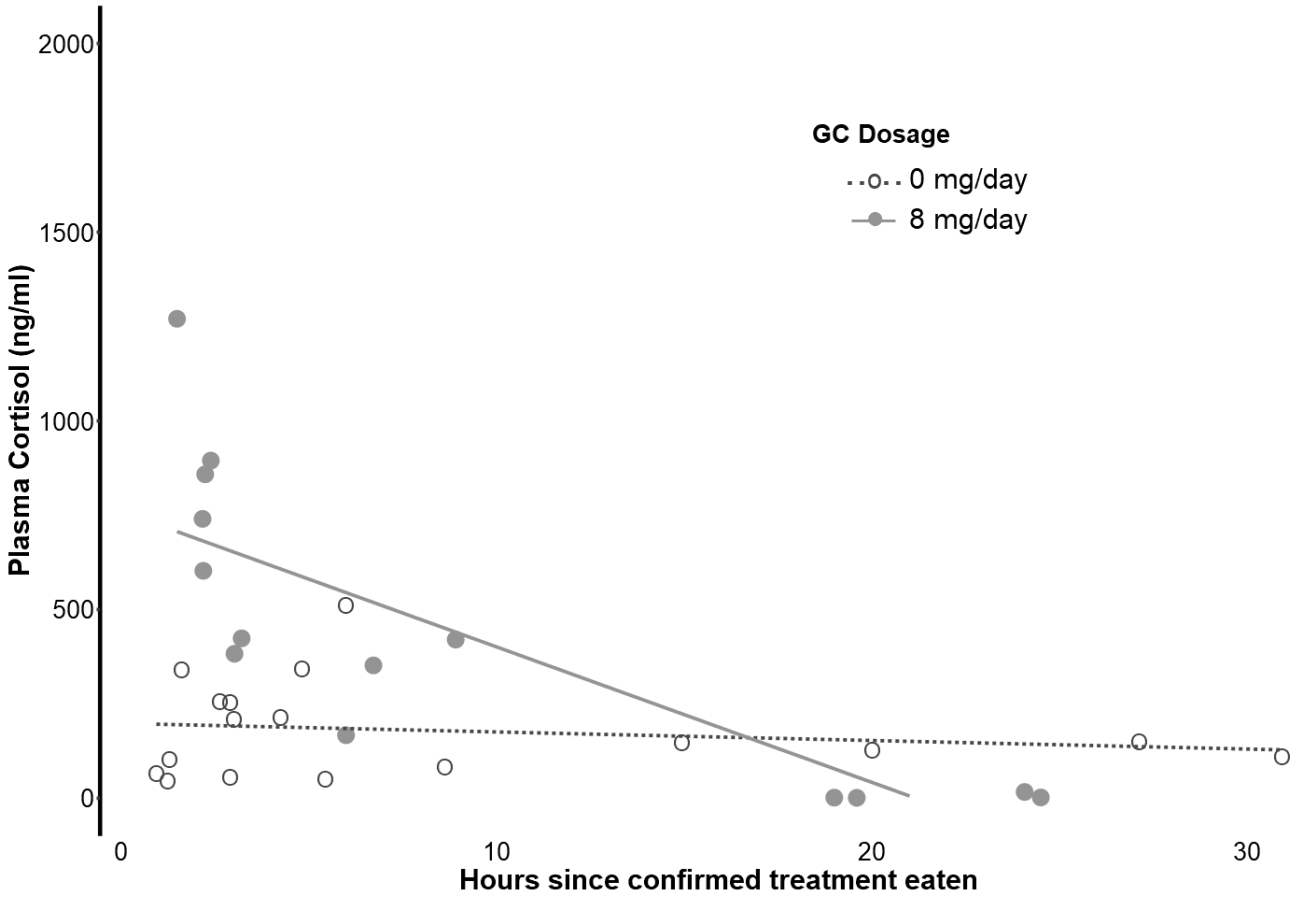

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Fig. 3. Effect of time elapsed since squirrels entered traps ("handling stress") on plasma cortisol concentrations from control squirrels (0 mg/day), or those treated with GCs (8 mg/day) and (A) sampled the same day as consuming their last treatment or (B) sampled the day after consuming their last treatment. Mean and SE are shown.

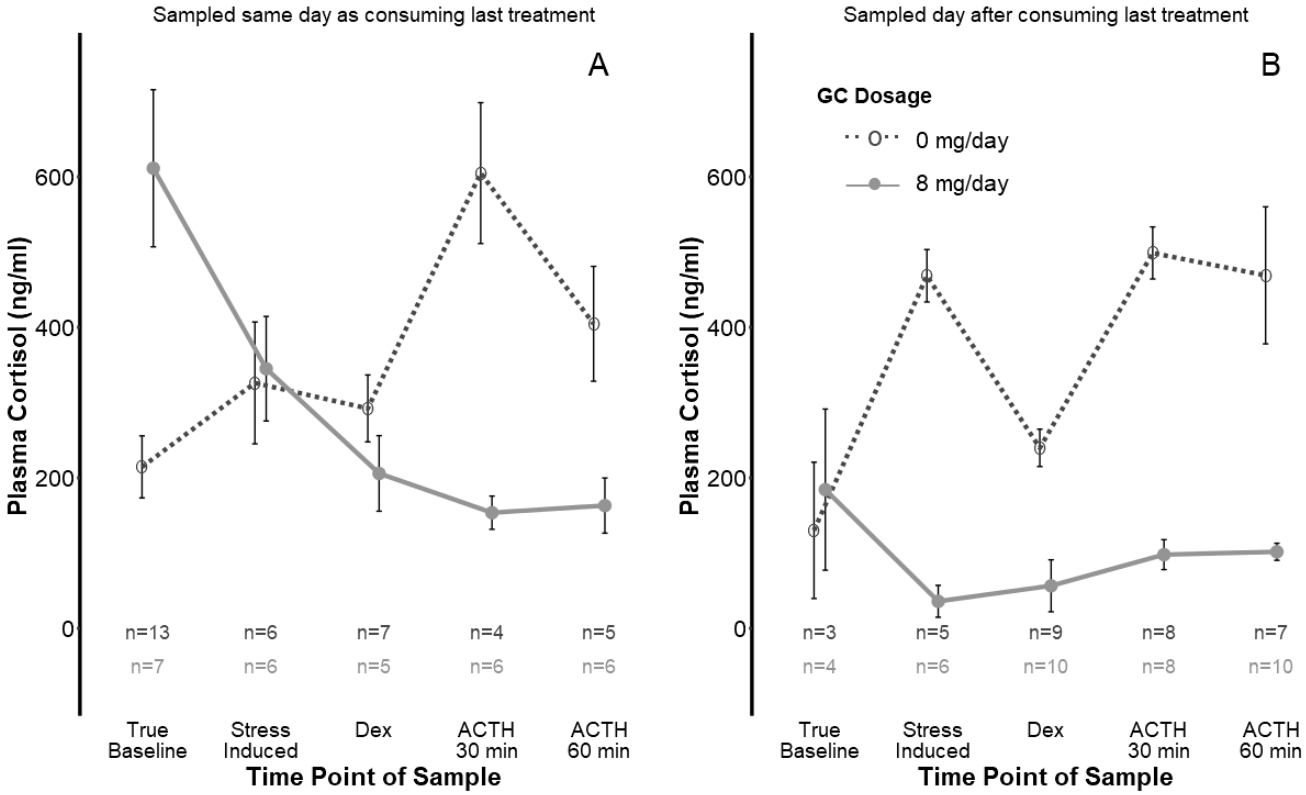

Fig 4. Plasma cortisol concentrations following Dex/ACTH challenges conducted on male squirrels treated with 0 mg (Control) or 8 mg cortisol/day for 7-14 days (GCs). A) Males were trapped the same day as confirming they consumed their last treatment. B) Males were trapped the day after being fed their last treatment, but note that the time they consumed their treatments was not recorded. Note that true baseline samples for cortisol treated squirrels were highly variable (two samples of 321.2 and 412.7 ng/ml, and two of 1.7 ng/ml). Means and standard errors are shown.

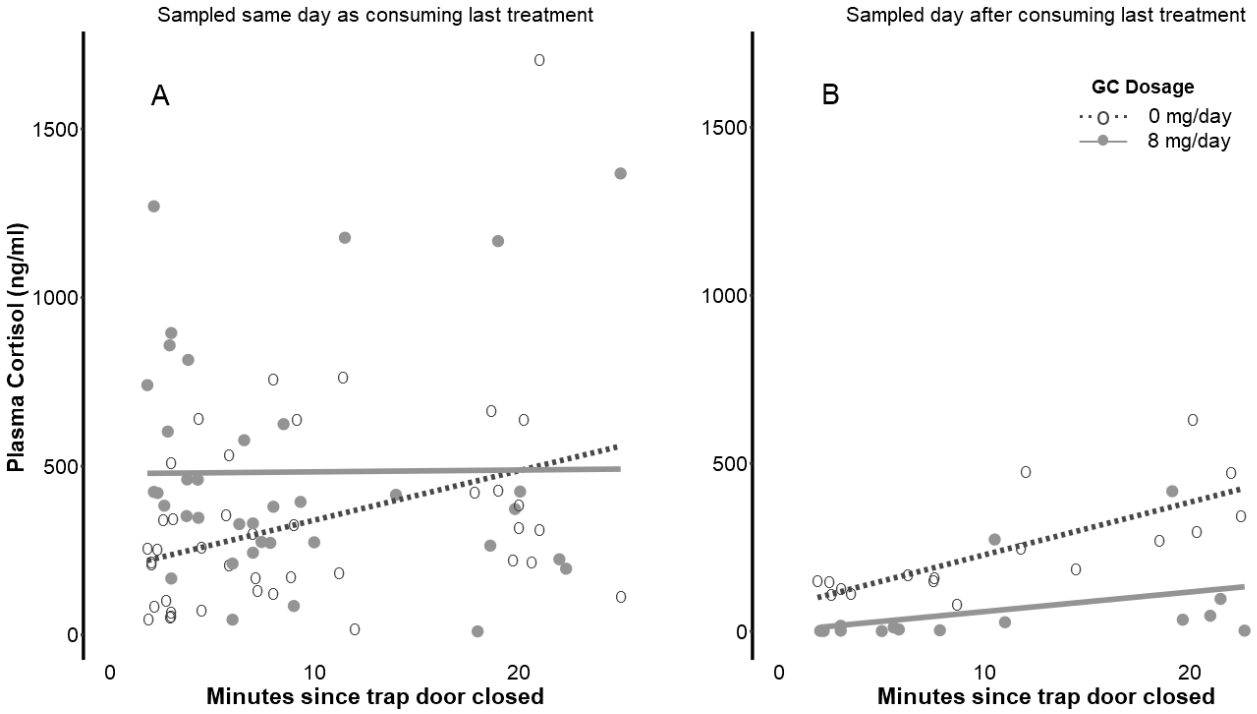

Fig. 5. A) Plasma corticosteroid binding globulin (CBG) and B) percent free cortisol in control and cortisol treated squirrels subjected to both Dex/ACTH challenges (Dex/ACTH Challenge) and response to handling stress (blood samples obtained 0-3, 3-6, 6-12 and 18-22 minutes after entering trap were lumped together). The figure includes squirrels that were sampled both on the same day as consuming their last treatment and the day after consuming their last treatment. Upper and lower hinges correspond to the first and third quartiles. Upper/lower whiskers extend from the hinge to the highest/lowest value that is within 1.5x the interquartile range. White diamonds indicate means.