1 On the relationship between high-order linkage disequilibrium and

- 2 epistasis
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- 4 Yanjun Zan^{1*}, Simon K. G. Forsberg^{1*+} and Örjan Carlborg^{1§}
- ¹Department of Medical Biochemistry and Microbiology, Uppsala University, SE-751
- 6 23 Uppsala, Sweden
- 7
- 8 *Authors contributed equally
- 9 ⁺Present address: Ecology and Evolutionary Biology Department; Lewis Sigler
- 10 Institute for Integrative Genomics, Princeton University, Princeton, New Jersey,
- 11 08540 and Department of Neuroscience, Functional Pharmacology, Uppsala
- 12 University, BMC, Box 593, 751 24 Uppsala, Sweden
- 13 [§]Corresponding author: <u>orjan.carlborg@imbim.uu.se</u>
- 14 Örjan Carlborg
- 15 Uppsala University
- 16 Medical Biochemistry and Microbiology
- 17 BMC Box 582, SE-751 23, Uppsala, Sweden.
- 18 Phone: +46 18 4714592
- 19
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- 22
- 23 Data availability:
- i) Genome wide re-sequencing data are available as part of Arabidopsis thaliana 1001
- 25 genomes project http://1001genomes.org/data-center.html, ii) 250 K SNP chip data
- 26 were available in the *Arabidopsis thaliana* Regmap panel
- 27 (<u>http://bergelson.uchicago.edu/?page_id=790</u>), iii) The Molybdenum level for 340
- 28 Arabidopsis thaliana are available in
- 29 https://doi.org/10.1371/journal.pgen.1005648.s005. iv) Corresponding genotypes are
- 30 extracted from a subset of the Regmap panel
- 31 (http://bergelson.uchicago.edu/?page_id=790).
- 32
- 33 Running title: High-order LD and statistical epistasis
 - 1

34

ABSTRACT

35 A plausible explanation for statistical epistasis revealed in genome wide association 36 analyses is the presence of high order linkage disequilibrium (LD) between the 37 genotyped markers tested for interactions and unobserved functional polymorphisms. 38 Based on findings in experimental data, it has been suggested that high order LD 39 might be a common explanation for statistical epistasis inferred between local 40 polymorphisms in the same genomic region. Here, we empirically evaluate how prevalent high order LD is between local, as well as distal, polymorphisms in the 41 42 genome. This could provide insights into whether we should account for this when 43 interpreting results from genome wide scans for statistical epistasis. An extensive and 44 strong genome wide high order LD was revealed between pairs of markers on the high 45 density 250k SNP-chip and individual markers revealed by whole genome sequencing 46 in the A. thaliana 1001-genomes collection. The high order LD was found to be more 47 prevalent in smaller populations, but present also in samples including several 48 hundred individuals. An empirical example illustrates that high order LD might be an even greater challenge in cases when the genetic architecture is more complex than 49 50 the common assumption of bi-allelic loci. The example shows how significant 51 statistical epistasis is detected for a pair of markers in high order LD with a complex 52 multi allelic locus. Overall, our study illustrates the importance of considering also 53 other explanations than functional genetic interactions when genome wide statistical 54 epistasis is detected, in particular when the results are obtained in small populations of 55 inbred individuals.

57

INTRODUCTION

58 The genetic architecture of most biological traits is complex and involves multiple 59 genes, whose effects are often influenced by interactions with other genes and 60 environmental factors. To study the relative contributions by genes, environmental 61 factors and their interactions in segregating populations, statistical genetic approaches 62 are commonly used to partition the genetic variance to additive and dominance 63 variance of individual loci and epistatic interaction variance between them (Lynch and Walsh 1998). In principle, the variance partitioning is performed by associating 64 65 the phenotypic variation for a trait in a population with linear combinations of the 66 genotypes within and/or across loci. How the genotypes are combined (parameterized) 67 in the model is determined by the genetic model used in the analysis. The classic 68 quantitative genetics models are parameterized to capture the genetic variance in a 69 hierarchical manner. First, a main additive allele-substitution is defined. Then, if 70 accounted for, dominance is modeled as a single-locus deviation from additivity and 71 genetic interactions as multi-locus deviations from single locus additivity and 72 dominance (Nelson et al. 2013). As a consequence of this, the genetic contributions of 73 individual and combinations of loci described as additive, dominance and epistatic 74 variances are unlikely to reflect the underlying biological mechanisms (Carlborg et al. 75 2006; Phillips 2008; Huang et al. 2012; Sackton and Hartl 2016; Forsberg et al. 2017).

76

77 Although the ultimate aim of a genetic association study is generally to detect 78 functional polymorphisms, most often genotypes are only scored for a reduced set of 79 polymorphisms (genetic markers). These reduced marker sets are selected with the 80 aim to tag as many of the unobserved functional polymorphisms as possible. The 81 statistical inferences of the underlying genetic architecture made from such reduced 82 sets of markers can, however, be problematic in some cases. For example, multiple 83 unobserved functional polymorphisms can lead to associations to individual markers 84 that do not properly represent the causal variants (Platt et al. 2010), and high order 85 linkage disequilibrium (LD) to single functional polymorphism can lead to indirect 86 statistical epistatic associations to pairs of markers (Wood et al. 2014). Here, we focus 87 on high order linkage disequilibrium defined as when two genotyped markers tag an 88 un-genotyped polymorphism (see Materials and Methods section). It is still unknown 89 how prevalent and strong such high order LD is in the genome, making it difficult to

90 estimate how many reported pairwise statistical epistatic interactions are due to such 91 LD. However, the study by Wood et al (Wood et al. 2014) presented results 92 suggesting that many of the significant statistical epistatic interactions detected 93 between pairs of local markers by Hemani et al. (Hemani et al. 2014) might be due to 94 high-order LD to unobserved, linked sequence polymorphisms in the same genomic 95 region. Many past and current studies of genetic interactions in, for example, 96 Drosophila, plant, animal and human populations (Shimomura et al. 2001; Anholt et 97 al. 2003; Caicedo et al. 2004; Segrè et al. 2004; Carlborg et al. 2006; Hemani et al. 98 2014) rely on genome-wide statistical analyses of pairwise interactions between 99 selected sets of markers as in (Hemani et al. 2014). With the increasing interest in, 100 and availability of, sufficiently large datasets for epistatic association analyses it is 101 therefore important to also evaluate the risk of making false inferences about loci 102 being involved in functional genetic interactions from findings of statistical epistasis, 103 when they instead are due to high order LD.

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105 Here, we empirically explore the prevalence and strength of high order LD within and 106 between chromosomes in publically available high-density SNP and whole-genome 107 re-sequencing data from the model plant Arabidopsis thaliana. Two locus LDs are 108 calculated between the markers selected for the 250k A. thaliana SNP chip that have 109 been the basis for many GWAS analyses in the past, and the additional SNPs revealed 110 by whole genome sequencing using data from the 1001 genomes project (Atwell et al. 111 2010; Cao et al. 2011; Horton et al. 2012; Schmitz et al. 2013; Alonso-Blanco et al. 112 2016). Strong high order LD was found to be common both within and across 113 chromosomes between pairs of markers from the SNP-chip and the sequencing 114 polymorphisms and often the combined genotype of the marker pair tagged the 115 genotype of the sequencing markers better than any single marker on the SNP chip. 116 The risk of falsely inferring genetic interactions between markers on different 117 chromosomes in a two-locus interaction analysis might increase in situations when the 118 underlying genetic architecture is more complex, for example when a single locus 119 contains multiple functional alleles. This is illustrated using an empirical example 120 from a second public A. thaliana dataset (Forsberg et al. 2015). Overall, this study 121 provides new insights that deepen our understanding about the link between high 122 order LD and statistical epistasis to guide researchers when interpreting results

123 obtained from epistatic genetic association analyses.

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- 125 126

MATERIALS AND METHODS

127 Methods

When an individual marker is in complete linkage disequilibrium $(r^2 = 1)$ with a 128 129 functional polymorphism affecting a studied trait, a single-locus association test between the marker and the trait will capture all the phenotypic variance contributed 130 131 by the functional polymorphism. A basic assumption in genetic association studies is 132 that at least one genotyped marker will be in sufficiently high LD with each functional 133 polymorphism to detect it in this way. In reality, however, not all functional 134 polymorphism will be in such perfect LD with a genotyped marker, and then there is a 135 risk that the joint genotype of two (or more) markers tags the genotype of the 136 functional polymorphism better than any single marker (high-order LD > single-137 marker LD). This will, as discussed below, influence the significances of the trait-138 marker associations detected in a genetic association analysis and the inferences made 139 about the genetic architecture of the trait.

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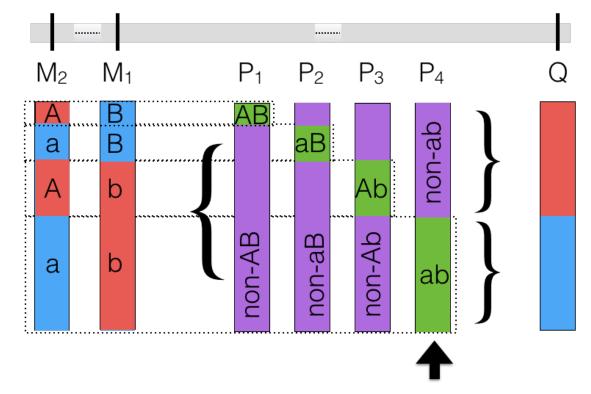
141 Quantifying high order linkage disequilibrium

We calculate the high order LD between pairs of predictors (here genotyped SNP
markers) and single targets (here un-genotyped SNP polymorphisms) following (Hao *et al.* 2007).

145

146 Consider a pair of bi-allelic predictor SNPs (M_1 and M_2 ; Figure 1). These markers can 147 together form four two-locus genotypes: AB, Ab, aB and ab (Figure 1). We now want 148 to know whether any two-locus predictor could tag the single locus target genotype 149 better than any of the individual predictor genotypes (i.e. evaluate whether 150 $\max(\text{second-order } LD) > \max(\text{single order } LD))$. To calculate the high order LD 151 between the two predictors $(M_1 \text{ and } M_2)$ and the single target (Q), the two-locus 152 M_1M_2 genotype is used to create a multi-allelic pseudo marker (P) with four alleles (Figure 1). In this way, a second-order LD (r^2) can be calculated for each of the 153 154 possible ways that M_1 and M_2 together can tag the genotype at Q (Figure 1).

156 The calculation of the second order LD therefore first involves creating the four 157 possible bi-allelic pseudomarkers (P₁, P₂, P₃ & P₄; Figure 1) from the two locus M_1/M_2 genotypes. These are assigned the genotypes P_1 {AB, non-AB}, P_2 {Ab, non-158 Ab}, P_3 {aB, non-aB} and P_4 {ab, non-ab}, respectively. The LD-r² is then computed 159 between the target (Q) and the four bi-allelic pseudomarkers (P₁, P₂, P₃ & P₄). For 160 each pair of predictors, the second order LD is then defined as the $LD-r^2$ for the 161 pseudomarker with the highest $LD-r^2$ to the target. Pseudomarkers with higher $LD-r^2$ 162 to the target (Q) than 0.3 are kept for further analyses. The $LD-r^2$ values were 163 computed using the software LdCompare (Hao et al. 2007). 164



165

166 *Figure 1.* Illustration of how the pseudomarkers (P_1, P_2, P_3, P_4) used in the estimation of the second **167** order linkage disequilibrium between a pair of linked or unlinked markers (predictors; M_1 and M_2 ,)

168 and a third linked or unlinked functional polymorphism (target; *Q*) are created. The pseudomarkers

169 together represent the possible bi-allelic formulations of the two-locus M_1M_2 genotypes. The maximum

170 pairwise $LD-r^2$ between the target and the four pseudomarkers (P_4) defines the second order LD

- 171 between the predictors (M_1, M_2) and the target (Q).
- 172
- 173

174 Statistical epistasis emerging from high order linkage disequilibrium

175 In a genetic association study in an inbred or haploid population, two-locus epistasis

is typically modelled as:

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178
$$Y = a_1\beta_1 + a_2\beta_2 + a_1a_2\beta_{12} + e$$
 [1]
179
180 Here, a_1 and a_2 are indicator variables for the genotypes at two genotyped markers, M_1
181 and M_2 , taking values 1/-1 for the two alternative homozygous genotypes AA vs aa
182 and BB vs bb, respectively. a_1a_2 is an indicator variable for the interaction between a_1
183 and a_2 taking value 1 for the two-locus genotypes AABB and aabb and -1 for AAbb
184 and aaBB. β_1 , β_2 and β_{12} are the corresponding estimates for the marginal (additive)

effects and the additive-by-additive interaction between the loci.

185 186

187 The aim of a statistical epistatic analysis is to include an interaction term in the model [1] to estimate the deviations of the two-locus genotype-values (AABB, AAbb, aaBB 188 189 and aabb) from the predictions obtained by the marginal (additive) effects (Alvarez-190 Castro and Carlborg 2007). However, a non-zero estimate of the interaction term in 191 model [1] does not, as noted e.g. by Wood et al. (Wood et al. 2014) necessarily have 192 to result from a genetic interaction. It could, for example, instead emerge from a 193 second-order LD between two markers and a single functional polymorphism. Here, 194 refer back to Figure 1. Now assume that a trait is determined by a single functional 195 locus (Q). Two markers, M_1 and M_2 , are genotyped but neither of these markers 196 individually tag the causal genotype (blue) at Q well. However, the causal (blue) 197 allele at Q is, tagged perfectly by one of the two-locus M_1M_2 genotypes (ab; Figure 1), while the other three M₁M₂ two-locus genotypes (aB, Ab and AA; Figure 1) are only 198 199 present together with the no-effect (red) allele at locus Q. When fitting model [1] to 200 the genotypes of marker M_1 and M_2 , the estimate for the interaction term (β_{12}) will be 201 non-zero, illustrating how statistical epistasis can emerge from the second-order LD 202 between M₁ and M₂ and Q. This example illustrates a scenario similar to what was 203 empirically observed in (Wood et al. 2014), where physically linked markers in low 204 haplotypes that were in high order LD with a LD with each other tagged 205 polymorphism that was unobserved in the original study.

206

207 Classifying identified high order linkage disequilibrium triplets depending on the
208 distance between the loci

209 Here, we evaluate the prevalence and strength of high order LD between pairs of 210 markers selected for genotyping on a 250k SNP chip (predictors) and a third locus 211 revealed by whole genome sequencing (targets) using publicly available datasets in A. 212 thaliana (Cao et al. 2011; Alonso-Blanco et al. 2016). Three types of high order LD 213 are defined based on the locations of the predictors relative to the target. If both 214 predictors are located within 1Mb of the target it is classified as cis-cis. If only one 215 predictor is closer than 1Mb it is classified as cis-trans. If none is closer than 1Mb it is 216 classified as trans-trans. The choice of a 1Mb threshold to define cis vs trans 217 predictors is arbitrary, but we consider it useful for evaluating how common high 218 order LD is between predictors near (local/cis) and far (global/trans) from the target.

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220 Material

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The genome wide prevalence of high order linkage disequilibrium in publically
available Arabidopsis thaliana datasets

224 The A. thaliana 1001-genomes project has released complete genome sequences for 225 hundreds of wild collected accessions (http://www.1001genomes.org). Here, we used 226 whole-genome SNP data on 728 accessions scored by whole genome re-sequencing 227 (Cao et al. 2011; Alonso-Blanco et al. 2016). The predictors used in our analysis was 228 a subset of the SNPs selected for the 250k A. thaliana SNP chip (Horton et al. 2012) 229 (n = 200,352 in total; MAF > 0.05) and the targets a subset of the SNPs revealed 230 using whole-genome re-sequencing (n = 1,641,240 in total; MAF > 0.05) (Table 1). Although the results from the analyses of this data will be specific to this species and 231 232 dataset, it is assumed that the relationships between targets and predictors will be a 233 realistic representation of what to be expected also in other populations. This is 234 because the selection of markers for the high-density 250k SNP chip, was done for the 235 purpose of genetic association studies following similar procedures as used also in 236 other species and populations.

237

The reason for only studying a subset of the possible targets and predictors is that it was not computationally feasible to exhaustively evaluate the high order LD between all possible pairs of predictors selected for the 250k SNP chip and all the targets revealed by genome sequencing. Instead, the second order LD was exhaustively

calculated for all targets and predictors i) within a randomly selected 6 Mb window on chromosome 2 as well as ii) between three randomly selected windows from different chromosomes (Table 1). Computations were performed for the entire population (n =728 individuals) and two smaller random samples of n = 100 and n = 50 individuals. The results for the populations with n = 100 and n = 728 are reported in the main manuscript and the results for n = 50 is reported in the Supplementary material.

249 *Table 1. Regions and SNPs selected for evaluation of second order LD.*

		-	-		
	Window 1	Window 2	Targets ¹	Predictors ²	Filtered targets ³
Region 1	Chr2: 8-14Mb	-	70,712	6,053	-
Regionpair 1	Chr1: 10-12Mb	Chr3: 10-12Mb	29,133	6,245	20,239
Regionpair 2	Chr2: 10-12Mb	Chr4: 10-12Mb	23,751	5,302	15,887
Regionpair 3	Chr2: 10-12Mb	Chr3: 10-12Mb	23,751	5,212	15,884
Genome			1,486,942	154,298	1,229,012

¹Total number of polymorphic SNPs in the evaluated windows/genome in the population revealed via whole-genome re-sequencing (Alonso-Blanco et al. 2016). ²Total number of polymorphic SNPs in the two windows/genome included on the 250k AT SNP-chip (Horton et al. 2012); ³Number of target SNPs in the two windows/genome with $LD-r^2 < 0.6$ to any individual predictor.

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The predictor pairs in the evaluated windows in the genome with high order $LD-r^2 >$ 0.6 to a target were classified as cis-cis/cis-trans/trans-trans. To extrapolate these findings to the genome level, the proportions of all evaluated predictor pairs that displayed these patterns were calculated and then multiplied with the total number of possible cis-cis/cis-trans/trans-trans pairs in the genome (Table S1).

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261 Analyzing a public A. thaliana dataset for two locus statistical epistasis

262 A publicly available dataset including 340 Arabidopsis thaliana accessions were used 263 for a genome wide association analysis. In short, the plants were grown in a controlled 264 environment with 6 biological replicate plants per accession. Analyses by Inductively 265 Coupled Mass Spectroscopy (ICP-MS) provided estimates of leaf molybdenum 266 concentration as described in (Baxter et al. 2010; Forsberg et al. 2015). The 267 accessions were genotyped for 141.385 SNP markers with MAF > 0.15 (Atwell *et al.* 268 2010; Baxter et al. 2010; Shen et al. 2012; Forsberg et al. 2015). A more thorough 269 description of the dataset can be found in (Baxter et al. 2010; Forsberg et al. 2015). In 270 an earlier study of this dataset (Forsberg et al. 2015), it was revealed that a large

271 fraction of the genetic variance for this trait was explained by a single linkage block 272 containing several low-frequency, large effect structural variants that were poorly 273 tagged by the genotyped SNPs. This linkage block was originally identified due to its 274 large marginal, variance heterogeneity effect in the population (Shen et al. 2012). It is 275 known that statistical epistasis and genetic variance heterogeneity can emerge from 276 similar genetic architectures (Forsberg et al. 2015), and this population was therefore 277 selected for further evaluations of whether high order LD between the genotyped 278 SNPs and these hidden polymorphisms could lead to statistical epistasis in a two locus 279 association analysis. We performed an exhaustive, two-dimensional genome scan for 280 pairwise statistical epistasis between the genotyped markers and the level of 281 molybdenum in the leaf using the software *plink* (Purcell *et al.* 2007) without control 282 for population structure. Thereafter, each pair of loci that passed the genome wide 283 significance threshold in the initial scan was fitted in a two-locus epistatic genetic 284 model [1] using hglm function in hglm package (Rönnegård et al. 2010) to correct for 285 the possible effects of population structure via the genomic kinship matrix as in (Forsberg et al. 2015). The significance threshold used to infer significant interacting 286 pairs ($p < 3.2 \times 10^{-10}$) was defined as a Bonferroni corrected nominal 5% significance 287 288 threshold. The correction was done for an estimated number of independent 289 association tests assumed to equal the number of independent LD blocks in the A. 290 thaliana genome as described in (Lachowiec et al. 2015).

291

292 Data availability

293 Genome wide re-sequencing data are available as part of the *Arabidopsis thaliana*

294 1001 genomes project <u>http://1001genomes.org/data-center.html</u>. The 250 K SNP chip

295 data are available as part of the genotype data for the *Arabidopsis thaliana* Regmap

296 panel (<u>http://bergelson.uchicago.edu/?page_id=790</u>). The Molybdenum levels for the

297 340 Arabidopsis thaliana accessions are available in

298 https://doi.org/10.1371/journal.pgen.1005648.s005

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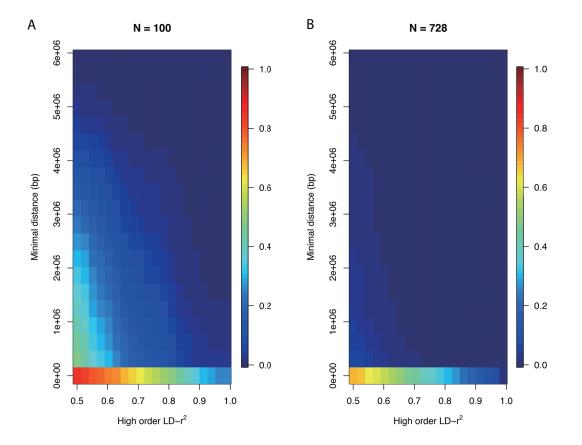
RESULTS

This study aims to answer the following questions by analyzing two public *A*. *thaliana* datasets: How common can we expect high order LD to be between pairs of

304 SNPs selected for genotyping and hidden sequence variants in the genome? Is high 305 order LD primarily observed between predictors tightly linked to a target functional 306 polymorphism (in cis) as in (Wood et al. 2014), or is it also observed for predictors 307 unlinked to the target (in trans)? How dependent is the prevalence of high order LD 308 and cis vs trans predictors on the population size? We also present an empirical 309 example where high order LD exists between a cis-trans predictor pair with 310 significant statistical epistasis and a locus displaying a strong genetic variance 311 heterogeneity due to independent contributions by multiple linked polymorphisms 312 (Forsberg *et al.* 2015). This illustrates how complex inheritance patterns of individual loci, something usually not explored in GWAS data, further complicates the 313 314 interpretation of detected statistical epistatic signals.

315

316 The population size affects the prevalence and location of predictors in high order LD The high order $LD-r^2$ values for all pairs of predictors and individual targets in a 6Mb 317 window on Chromosome 2 (Table 1) is shown for populations with n = 100 and n =318 728 individuals in Figure 2. The strongest second order $LD-r^2$ was observed where at 319 320 least one predictor is located near the target (y-axis). When the sample size was smaller (n = 100; Figure 2A), strong second order $LD-r^2$ was rather common also 321 when both predictors were located far from the target. For example, 20% of the 322 targets had a high order $LD-r^2 > 0.65$ with a predictor pair where at least one of the 323 324 predictors was located more than 1Mb away from it. Even though the prevalence of strong high order LD- r^2 decreases when the sample size increases, it is still common 325 326 in the large population (n = 728; Figure 2B), with the highest prevalence when at least 327 one of the predictors is located close to the target.



328

Figure 2. Illustration of how the prevalence of high order $LD-r^2$ to the targets in a 6Mb window on A. thaliana chromosome 2 (8 – 14Mb) depends on distance of the predictors from the target. The color gradient illustrates the proportion of predictor pairs that reach a particular $LD-r^2$ (x-axis) depending on the distance between the nearest predictor and the target (y-axis). Results are presented for populations with n = 100 (A) and n = 728 (B) individuals.

334

Strong high-order $LD-r^2$ between a predictor pair and a target is mostly observed when at least one of the predictors is in strong individual $LD-r^2$ with the target. However, as illustrated in Figure 3, many cases also exist where the high order $LD-r^2$ is strong while the $LD-r^2$ to the individual predictors is weak.

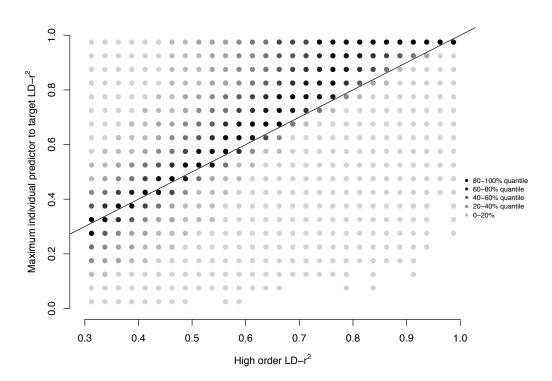


Figure 3. Strong second order $LD-r^2$ exists also when the individual predictor to target $LD-r^2$ is weak. The intensity of each dot illustrates the number of cases with a particular high order $LD-r^2$ / maximum individual predictor to target $LD-r^2$ combination. Dots below the line are cases where the high order $LD-r^2$ stronger than any individual predictor to target $LD-r^2$ (n=728).

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345 *Estimating the genome wide prevalence of strong high order linkage disequilibrium*

Figure 2 illustrates that high-order $LD-r^2$ exists where one or both predictors are 346 347 located close to the target as well as when one or both predictors are located further away in the evaluated 6Mb window. The genome-wide prevalence of high order LD-348 r^2 for the three different classes of predictor pairs, cis-cis/cis-trans/trans-trans (as 349 350 defined above) were next explored in three pairs of distant 2Mb windows in the 351 genome (Table 1) to provide data to estimate their genome-wide prevalence. Here, only cases when individual predictors in the windows had lower individual $LD-r^2$ than 352 353 0.6 to the targets were considered.

354

Overall, the fraction of predictor pairs that display higher second-order LD (LD- r^2 > 0.6) is low. In the smaller population (n = 100), less than 1 out of 10⁶ evaluated predictor pairs and in the larger population (n = 768) less than 1 out of 10⁷ (Table S1). However, since the total number of evaluated pairs was very large (around 10¹¹).

359 many cases were still detected. Regardless of population size, cis-cis and cis-trans 360 pairs dominated (42/44% for n = 100, and 56/58\% for n = 728; Figure 4A-C; Table 361 S1). Trans-trans pairs existed, but were much less common (~1% for n = 100, <0.01% 362 for n = 728, respectively, Figure 4A-C; Table S1). When extrapolating these results to 363 a genome wide scale, this picture, however, changes dramatically (Figure 4D). Trans-364 trans and cis-trans predictor pairs are now much more common than cis-cis pairs due 365 to their much higher genome-wide prevalence (35/18-fold for n = 100 and 35/0.3 for n = 728 more common; Figure 4D, Table S1). This result illustrates that it is a 366 367 considerable risk to disregard high-order LD as a possible explanation for statistical 368 epistatic interactions even at larger sample-sizes.



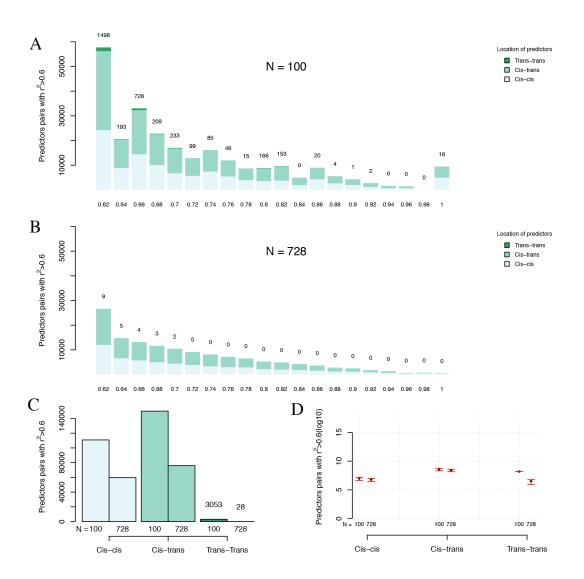




Figure 4. Number of predictor pairs of different classes in strong high order $LD-r^2$ (>0.6) to targets detected in the evaluated windows and estimated genome wide. The distribution of $LD-r^2$ values > 0.6

373 for the cis-cis, cis-trans, trans-trans predictor pairs for (A; n = 100) and (B; n = 728) The total number

374 of predictor pairs with high order $LD-r^2$ above 0.6 in the three classes are summarized in (C) and used

- 375 to estimate the total expected number of predictor pairs in the entire genome (D; error bars show the
- 376 *estimation error estimated from the results obtained for the three window (Materials and Methods).*
- 377

378 Linking high order LD and statistical epistasis in a two locus epistatic association
379 analysis in A. thaliana

380 A publicly available dataset including 340 Arabidopsis thaliana accessions were used 381 for a genome wide association analysis for leaf molybdenum concentration This 382 dataset was earlier used by (Forsberg et al. 2015) to dissect a locus with a highly 383 significant variance heterogeneity association for leaf molybdenum concentration 384 (Shen et al. 2012) to the contributions of four independent associations in an extended 385 LD block on chromosome 2. Several of these associations were found to structural 386 variants that were poorly tagged by the SNP markers (Forsberg et al. 2015). Our 387 pairwise genome wide scans for pairs of epistatic loci identified 396 significant SNP 388 pairs. For 290 pairs both markers were located in the narrow region on chromosome 2 389 that was earlier dissected in detail (Forsberg et al. 2015). All these are examples of 390 cis-cis predictor pairs. The remaining 106 pairs contained one predictor in the 391 chromosome 2 region and another one elsewhere in the genome, being examples of 392 cis-trans predictor pairs.

393

394 The strongest pairwise epistasis was detected for a cis-trans predictor pair (Figure 5A). 395 The accessions with the AA genotype at the predictor located in trans to the 396 chromosome 2 region (chromosome 1:5,315,502 bp) all have an intermediate 397 molybdenum level in the leaf (Figure 5A). The accessions with the GG allele at the 398 trans predictor have different levels of molybdenum in their leaves depending on 399 whether they carry the CC or TT genotype at the cis predictor in on chromosome 2 400 (10,928,720 bp). These differences explain the significant statistical epistasis detected 401 when fitting the two-locus epistatic model [1] to this data.

402

This statistical interaction could be due to a true genetic interaction. An alternative explanation is however presented in Figure 5 There, the overlap between the two locus genotypes for the cis-trans predictor pair (Figure 5A) and the alleles at the four

406 loci earlier reported to be associated with leaf molybdenum levels in this region 407 (Forsberg *et al.* 2015) are illustrated. The multi-locus genotypes of the predictor pair 408 tags different combinations of minor alleles at the four loci that were found to either 409 increase (mGWA1, mGWA2, 326ins) or decrease (53del) leaf molybdenum levels in 410 the accessions (Forsberg et al. 2015). The statistical epistatic interaction was detected due to the difference in molybdenum levels between accessions carrying the GGCC 411 412 genotype (low molybdenum) and GGTT (high molybdenum). Figure 5B shows that the accessions with the GGCC genotype have the lowest frequency of the 413 414 molybdenum increasing allele mGWA2 and the highest frequency of the molybdenum 415 decreasing allele 53del. The accessions with the GGTT genotype instead have the highest frequencies of the molybdenum increasing alleles at mGWA2, mGWA1 and 416 417 326ins. The genotypes AACC and AATT, with intermediate molybdenum levels, both have intermediate frequencies of the mGWA1 and mGWA1 increasing alleles and 418 419 lack the 53del and 326ins alleles. A more parsimonious interpretation of these results 420 is thus that the statistical epistasis at the predictor pair is due to the high order LD 421 between them and the genotypes at the four loci located in the region on chromosome 422 2.

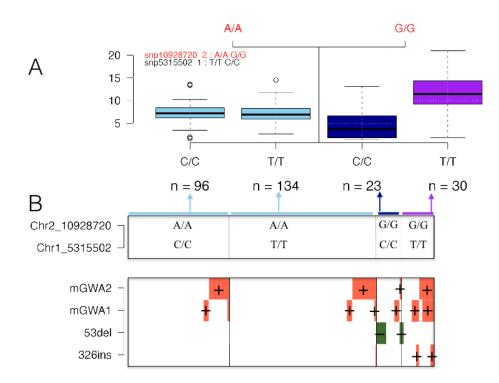


Figure 5. An illustration of how the high order LD between four polymorphisms affecting the level of
molybdenum in the A. thaliana leaf (Forsberg et al. 2015), likely explains the significant statistical

426 epistasis detected for a cis-trans predictor pair. (A) Boxplots illustrating the phenotypic distribution in 427 the four genotype classes defined by the cis-trans predictor pair with the strongest significant epistatic 428 interaction to the level of molybdenum in the A. thaliana leaf. (B) Illustration of the connection 429 between the two-locus genotypes of the predictor pair and the minor alleles at the four linked loci 430 associated with this trait on chromosome 2 (Forsberg et al. 2015). The top box in (B) illustrates the 431 two-locus genotype for the predictor pair, with the width of each sub-box indicating the number of 432 individuals in each genotype class in the population. In the bottom box in (B), each individual is 433 represented as a column, where green (molybdenum decreasing) and orange (molybdenum increasing) 434 colors indicates that the individual carry the minor alleles at the four loci identified in (Forsberg et al. 435 2015). mGWA1 and mGWA2 are SNP markers associated with the trait and 53del and 326 are 436 structural polymorphisms (Forsberg et al. 2015).

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DISCUSSION

439 High order linkage disequilibrium between combinations of genotyped markers, and 440 unobserved functional polymorphisms, can result in significant statistical epistasis in 441 genome wide association analyses. This was earlier illustrated empirically for linked 442 pairs of genotyped predictor SNPs and ungenotyped target polymorphisms in humans 443 by Wood et al. (Wood et al. 2014). Here, we present a new example from A. thaliana 444 where significant statistical epistasis between pairs of predictors is due to the effects 445 at a single loci and that only one of the statistically interacting loci was located near 446 the target. By exploring the prevalence of second order LD in the genome of the 447 1001-genomes A. thaliana collection, we find that although the total amount of high 448 order linkage disequilibrium decreases with increasing population sizes, it is still 449 highly prevalent both within and across chromosomes even in relatively large 450 populations (n = 728). It is was found to be most common when one predictor is in 451 high LD to (and located physically near) the target, but many cases exist where the 452 LD to the individual predictors is very weak but the high order LD is strong. The 453 choice of target and predictor SNPs in this study is arbitrary and therefore it it is 454 difficult to assess how representative they are for the prevalence of high order LD in 455 other populations. However, they do suggest that strong high order LD can be 456 prevalent also in larger populations, indicating that statistical epistasis observed in 457 studies based on reduced representation genotyping (such as SNP-chips) need to be 458 interpreted with caution.

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460 The most prevalent type of high order LD on a genome wide basis is that of cis-trans 461 predictor pairs, but also cis-cis pairs are common regardless of population size. The 462 prevalence of trans-trans pairs is high in smaller populations but decreases rapidly as 463 the population size increases. A possible biological explanation for the observation 464 that cis-cis and cis-trans high order LD pairs is relatively prevalent also at larger 465 population sizes would be that the number of, and variation in, the trans located predictors is sufficiently large on a genome-wide basis to complement any 466 467 imperfection in the tagging of the functional polymorphism by the cis located 468 predictor. Whereas trans-trans high order LD will always result in falsely associated 469 loci, cis-trans and cis-cis high order LD presents an opportunity to identify true 470 functional loci for the trait. The problem in a real data analysis is that statistical 471 epistasis between a pair of predictors can emerge from true interactions or high order 472 LD within and across chromosomes. However, as the sample sizes increase the risk of 473 detecting pairs of predictors where none is located close to the true functional 474 polymorphism decreases. Before concluding that the detected association is due to 475 two interacting loci, further analyses of the associated pair are however recommended.

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477 Whole-genome sequencing provides unprecedented opportunities to genotype most 478 segregating single nucleotide polymorphisms in the genome. Despite this, it is 479 unlikely that these will be able to tag all functional polymorphisms, such as larger 480 structural variants or multi-allelic functional loci due to tandem repeats. Hence, even 481 though the scenario of reduced representation genotyping with SNP-chips or similar 482 will soon be a technology of the past, association analyses will still be challenged by 483 the need to tag hidden polymorphisms with imperfect markers as illustrated in our 484 analyses of the complex locus affecting molybdenum levels in the A. thaliana leaf. In 485 fact, it is not unlikely that the problem with high order LD between SNP predictors 486 and hidden, complex functional loci will remain a major challenge in the future as the 487 increased number of markers generated by sequencing also increases the chance of 488 finding combinations of cis-cis or cis-trans predictors that tag these functional 489 polymorphisms better than any single marker. To evaluate the extent of this problem 490 one will, however, need a more comprehensive dataset than the one studied here 491 including a more complete scoring of all types of non-SNP polymorphisms in the

492 genome with potential effect on traits of interest.

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494 The prevalence of high order LD is likely to be more of a concern in populations of 495 inbred or haploid individuals. These include, for example, inbred lines derived from 496 bi- and multi-parental crosses of plants and animals, as well as populations of wild 497 collected inbred plants (Churchill et al. 2004; Valdar et al. 2006; Kover et al. 2009; 498 Cao et al. 2011; Mackay et al. 2012). As heterozygotes are not present in these 499 populations, the number of multi locus genotype classes is smaller than in outbred 500 populations, making them attractive for studies of genetic interactions. As a common 501 approach to detect interactions in such populations is to identify pairs of loci 502 displaying significant statistical epistasis, such results need to be interpreted with 503 caution, as the analyzed populations are generally small. If one, or more, of the 504 functional polymorphisms in the genome are unknown and poorly tagged by the 505 genotyped markers, there is a risk that statistical interactions arise from high-order LD 506 between the genotyped markers and the hidden functional polymorphisms. Hence, 507 even though these populations increase the power to map loci displaying statistical 508 epistasis, there is also a risk of falsely concluding that the underlying genetic 509 architecture involves genetic interactions.

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CONCLUSIONS

Statistical epistasis detected in genome wide association analyses can result from high 512 513 order LD between genotyped markers and unobserved functional polymorphisms. 514 This study revealed extensive and strong genome wide high order LD between pairs 515 of markers on a high density 250k SNP-chip and individual markers revealed by whole genome sequencing in the A. thaliana 1001-genomes collection. The high 516 517 prevalence of strong high order LD in this dataset suggests that epistatic variance 518 detected between pairs of markers in association analyses, especially in small inbred 519 populations genotyped for reduced representation sets of markers, need to be 520 interpreted with caution. An empirical example is presented where a pair of markers 521 with significant statistical epistasis in a genome wide association analysis is in high 522 order LD with a complex multi allelic locus with large effects on the analyzed trait. 523 As complex functional loci such as this are unlikely to be captured by individual bi-524 allelic SNP markers, even if millions of them are scored by whole genome sequencing,

525 it is important to evaluate also other explanations of statistical epistasis than 526 underlying genetic interactions in particular when small populations of inbred 527 individuals are studied. 528 529 **ACKNOWLEDGEMENTS** 530 This study was funded by grants from the Swedish Research Council (2012-4634) and 531 the Swedish Research Council Formas (2013-450) to ÖC. 532 533 **AUTHOR CONTRIBUTIONS** 534 ÖC and SF initiated the study. ÖC, SF and YZ designed the project and the statistical 535 analyses; SF and YZ wrote analysis scripts and performed the data analyses. ÖC and 536 YZ summarized the results and wrote the initial version of the manuscript. All authors 537 contributed to the writing of the final version of the manuscript. 538 539 **DISCLOSURE DECLARATION** 540 The authors declare no competing interest. 541 542 SUPPLEMENTARY MATERIAL 543 Supplementary material is provided in Supplementary Figure 1 and Supplementary 544 Table 1. 545 546 **REFERENCES:** 547 1001 Genomes Consortium., 2016 1,135 Genomes Reveal the Global Pattern of 548 Polymorphism in Arabidopsis thaliana. Cell 166: 481-491. 549 Anholt R. R. H., Dilda C. L., Chang S., Fanara J.-J., Kulkarni N. H., et al., 2003 The 550 genetic architecture of odor-guided behavior in Drosophila: epistasis and the 551 transcriptome. Nat Genet 35: 180-184. 552 Atwell S., Huang Y. S., Vilhjálmsson B. J., Willems G., Horton M., et al., 2010 553 Genome-wide association study of 107 phenotypes in Arabidopsis thaliana inbred 554 lines. Nature 465: 627-631. 555 Aulchenko Y. S., Ripke S., Isaacs A., van Duijn C. M., 2007 GenABEL: an R

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