# 1 **RefSeq database growth influences the accuracy of** *k***-mer-based**

# 2 species identification

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9

### 10 ABSTRACT

11 Accurate species-level taxonomic classification and profiling of complex microbial communities 12 remains a challenge due to homologous regions shared among closely related species and a 13 sparse representation of non-human associated microbes in the database. Although the database 14 undoubtedly has a strong influence on the sensitivity of taxonomic classifiers and profilers, to 15 date, no study has carefully explored this topic on historical RefSeq releases and explored its 16 impact on accuracy. In this study, we examined the influence of the database, over time, on k-17 mer based sequence classification and profiling. We present three major findings: (i) database 18 growth over time resulted in more classified reads, but fewer species-level classifications and 19 more species-level misclassifications; (*ii*) Bayesian re-estimation of abundance helped to recover 20 species-level classifications when the exact target strain was present; and (iii) Bayesian re-21 estimation struggled when the database lacked the target strain, resulting in a notable decrease in 22 accuracy. In summary, our findings suggest that the growth of RefSeq over time has strongly 23 influenced the accuracy of k-mer based classification and profiling methods, resulting in

24	different classification results depending on the particular database used. These results suggest a
25	need for new algorithms specially adapted for large genome collections and better measures of
26	classification uncertainty.

27

28 Keywords: Taxonomic classification, Reference database, Metagenomics, Microbiome,

29 Comparative analysis

30

## 31 INTRODUCTION

Fundamental questions of a metagenomic survey are: (*i*) what microbes are present in each sample, (*ii*) how abundant is each organism identified in a sample, (*iii*) what role might each microbe play (i.e. what gene functions are present) and (*iv*) how do the previous observations change across samples and time? Specifically, there have been numerous studies highlighting the utility of metagenomic datasets for pathogen detection, disease indicators, and health <sup>1,2</sup>. Addressing each of these fundamental questions is predicated on the ability to assign taxonomy and gene function to unknown sequences.

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40 Several new tools and approaches for taxonomic identification of DNA sequences have emerged 41  $^{3-5}$ , in addition to community-driven 'bake-offs' and benchmarks <sup>6</sup>. *k*-mer based classification 42 methods such as Kraken or CLARK <sup>3,7</sup> are notable for their exceptional speed and specificity, as 43 both are capable of analyzing hundreds of millions of short reads (ca. 100 base pairs) in a CPU 44 minute. These *k*-mer based algorithms use heuristics to identify unique, informative, *k*-length 45 subsequences (*k*-mers) within a database to help improve both speed and accuracy. A challenge 46 for *k*-mer based classification approaches is that closely related species and strains often contain 47 many identical sequences within their genomes. This challenge is typically addressed by 48 assigning the query sequence with the lowest common ancestor (LCA) of all species that share 49 the sequence. A comprehensive benchmarking survey indicated that Kraken offered the best  $F_1$ 50 score (a measure considering both precision and recall) among the k-mer based taxonomic classifiers evaluated at the species level<sup>8</sup>. Bracken, a Bayesian method that refines Kraken 51 52 results, is capable of estimating how much of each species is present among a set of ambiguous species classifications by probabilistically re-distributing reads in a taxonomic tree<sup>9</sup>. We thus 53 54 selected Kraken and Bracken as representative tools from the genre of k-mer based classification 55 methods. The focus on this study was not to examine a specific software tool, but rather to 56 decouple the performance of k-mer based methods from the underlying database. 57 58 Available k-mer based methods for taxonomic identification and microbiome profiling rely on 59 existing reference databases. While several investigations have examined the influence of 60 contamination in specific database releases, and identified idiosyncrasies specific to a release <sup>10,11</sup>, no study has examined the specific influence of perhaps the most popular database from 61 62 which to build classification databases, the repository of sequenced and assembled microbes 63 (RefSeq), across all releases of the database. Additionally, metagenomic classification and 64 profiling tools are commonly compared to each other using simulated datasets on a fixed 65 database, with leave-one-out analysis, but never compared to each other across recent trajectories 66 in database growth. The aim of this study was to elucidate the influence of RefSeq database 67 growth over time on the performance of k-mer based taxonomic identification tools. 68

69 **RESULTS** 

### 70 RefSeq database growth

71 Since its release in June 2003 bacterial RefSeq, on average, has doubled in size (giga base pairs, 72 Gbp) every 1.5 years (Fig. 1A), with the number of unique 31-mers in the database growing at a 73 similar rate (Fig. 1B). A more recent release, bacterial RefSeq version 84 (released 9/11/2017), 74 totaled over 700 Gbp of sequence data. The Simpson's index of diversity is a metric with values 75 between zero and one that reports the probability that two individuals randomly selected from a 76 sample will not belong to the same species. Samples with a high Simpson's index of diversity 77 (i.e. closer to one) may be considered more diverse than those with low values (i.e. closer to 78 zero). The diversity for each version of the bacterial RefSeq database increased until April 2013 79 where the Simpson's index of diversity for each subsequent bacterial RefSeq release has trended 80 downward (Fig. 1C). A slower growth is also seen in the number of new bacterial species in 81 each RefSeq version, indicating many of the same species are being sequenced repeatedly (Fig. 82 1D).

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#### 84 Taxonomic classification over time with a simulated metagenome

85 Kraken's own simulated validation set of ten known genomes was searched against nine versions 86 of bacterial RefSeq (1, 10, 20, 30, 40, 50, 60, 70, 80) and the MiniKraken database (4GB 87 version) (Fig. 2). The accuracy of each Kraken run depends on the RefSeq version used in the 88 search (Fig. 2; Table 1). Correct genus-level classifications increased as RefSeq grew, but 89 correct species-level classifications peaked at version 30 and tended to decline thereafter (Fig. 2). 90 The decrease in correct species classifications is due to more closely-related genomes appearing 91 over time in RefSeq, making it difficult for the classifier to distinguish them and forcing a move 92 up to the genus level. Overall, misclassified species-level calls were consistently rare, as reads

93 were misclassified at the species level an average of 7% of the time (Table 1; Fig. 2). The 94 fraction of reads classified at any taxonomic level, regardless of accuracy, increases as RefSeq 95 grows over time (Fig. 3). However, the fraction of species-level assignments (again, regardless 96 of accuracy) peaks at RefSeq version 30 and begins to decline thereafter, while the fraction of 97 genus-level classifications begins to increase.

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99 Bracken was used to re-estimate the abundances of classifications made by Kraken when 100 searching the simulated reads against eight bacterial RefSeq database versions (1, 10, 20, 30, 40, 101 50, 60, 70). Bracken first derives probabilities that describe how much sequence from each 102 genome is identical to other genomes in the database. This step requires searching a Kraken 103 database against itself with Kraken, which could not be performed for the MiniKraken DB (as 104 there is no FASTA file for this database) or bacterial RefSeq version 80 (as it would require 105 extensive computation for a database that size). Bracken was able to re-estimate species 106 abundances for 95% of the input data using RefSeq version 70, while Kraken only classified 107 51% of reads at the species level. Because Bracken may probabilistically distribute a single 108 read's classification across multiple taxonomy nodes, its performance must be measured in terms 109 of the predicted abundances. Bracken typically included the correct species in its re-estimation, 110 but sometimes included incorrect species in the abundance estimation (on average 15% of reads 111 were associated with a genome outside of the ten knowns).

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#### 113 Taxonomic classification of difficult to classify genomes over time

114 The challenging nature of classifying sequences belonging to the *Bacillus cereus* sensu latu

group has been previously documented  $^{12,13}$ . The *B. anthracis* species within this group is a well-

116 defined monophyletic subclade of the larger *B. cereus* group, and the base of the *B. anthracis* 117 clade is commonly denoted by a single nonsense mutation in the plcR gene <sup>14</sup> which is conserved 118 in all known *B. anthracis* genomes and has been shown to confer a regulatory mutation essential 119 for maintaining the pXO1 and pXO2 plasmids that carry the virulence factors characteristic of 120 anthrax<sup>15</sup>. However, not all *B. anthracis* cause disease in humans, such as *B. anthracis* Sterne (missing the pXO2 plasmid) and some *B. cereus* strains do cause anthrax-like disease  $^{16}$ , 121 122 complicating a precise species definition. Thus, it is not a surprise that accurate species-level 123 classification within this group has proven challenging for k-mer based methods, especially those 124 methods not based on phylogenetic evidence. To demonstrate how difficult sequences from this 125 group have been to classify over time, simulated reads were created for two Bacillus cereus 126 strains. The first, B. cereus VD118, is a strain available in RefSeq version 60 and beyond, and the second, *B. cereus* ISSFR-23F<sup>17</sup>, was recently isolated from the International Space Station 127 128 and is not present in any of the RefSeq releases tested. It is phylogenetically close to B. 129 anthracis, but lacks the phylogenetic and species characteristics of B. anthracis. Again, as 130 bacterial RefSeq grows over time, the number of genus-level classifications made by Kraken 131 increases (Fig. 4). While the number of genus-level calls made by Kraken increases over time 132 the number of unclassified and misclassified species calls decreases (most commonly B. 133 anthracis, B. thuringensis, and B. weihenstephanensis).

134

Bracken made species-level predictions for all reads no matter which version of bacterial RefSeq was used (Fig. 4). However, the increased rate of species-level predictions came at the cost of accuracy, as Bracken correctly identified *B. cereus* VD118 and *B. cereus* ISSFR-23F an average of 72% and 29% of the time, respectively, across RefSeq versions 1 through 70. The fraction of

139 reads assigned to each *Bacillus* species varied substantially from each database tested. The range

140 of *Bacillus* species predictions for *B. cereus* VD118 were: *B. cereus* 81% (max=100%,

141 min=18%), B. anthracis 48% (max=48%, min=0%), B. thuringiensis 23% (max=23%, min=0%),

142 and *B. weihenstephanensis* 76% (max=76%, min=0%). While the range of *Bacillus* species

143 predictions for *B. cereus* ISSFR-23F were: *B. cereus* 45% (max=50%, min=5%), *B. anthracis* 

144 90% (max=95%, min=5%), and *B. thuringiensis* 54% (max=54%, min=0%).

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#### 146 CPU/Memory performance over time

147 Historical bacterial RefSeq versions were recreated and used to build Kraken databases with

148 default settings. While most databases were constructed with ease and in less than a day, version

149 70 required 500 GB of RAM and 2 days (single compute node using on 64 cores), while version

150 80 required ca. 2.5 TB of RAM and ca. 11 days (single compute node using on 64 cores). Given

151 this trend, future releases will likely require over 4 TB of RAM and weeks of computation to

build, putting into question the feasibility of building and profiling *k*-mer databases on future

153 RefSeq versions. Recent studies <sup>18</sup> have suggested alternative approaches for database

154 construction that would help to circumvent future computational bottlenecks.

155

#### 156 **DISCUSSION**

The results of our study support three conclusions: (*i*) the RefSeq bacterial database composition and diversity is dynamic, varying from release to release; (*ii*) the database composition strongly influences the performance of *k*-mer based taxonomic identification methods, and (*iii*) Bayesian based methods can help mitigate some of the effect, but struggle with novel genomes that have close relatives in the database.

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### 163 Database influences on k-mer based taxonomic classification

164 Using Bracken, the majority of *Bacillus cereus* ISSFR-23F simulated reads were not correctly

- assigned to *B. cereus* but were more frequently mis-assigned as *Bacillus anthracis* or *Bacillus*
- 166 *thuringiensis* (Fig. 4B). This, in part, is not surprising as two of the three species in this group, B.
- 167 *cereus* and *B. thuringiensis*, have no clear phylogenetically defined boundary, though *B.*
- 168 *anthracis* is phylogenetically distinct from *B. cereus* and *B. thuringiensis*. Furthermore, any two
- 169 genomes within the *Bacillus cereus* sensu lato group are likely to be over 98% identical <sup>9</sup>. Given
- 170 that *k*-mer based methods are not phylogenetically-grounded, but rather based on sequence

171 composition, they are susceptible to misidentification in clades where the taxonomy is in partial

172 conflict with phylogeny, such as the *Bacillus cereus* sensu lato group. One clear example of

173 misidentification within this group was the false identification Anthrax in public transit systems

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Another observation worth highlighting is that the fraction of simulated reads classified as one of the three *B. cereus* sensu lato species varied across database versions (Fig. 4), with the exception of *B. cereus* VD118, which was present in RefSeq releases 60 and 70 (Fig. 4A). The variation in species classifications across database versions indicates that even when using the same tools to analyze the same dataset, the conclusions derived from this analysis can vary substantially depending on which version of a database you are searching against, especially for genomes belonging to difficult to classify species (i.e. require phylogenetic-based approaches).

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184 Imperfect data

185 The genomic data deluge has helped to expand public repositories with a broader and deeper 186 view of the tree of life, but has also brought with it contamination and misclassification. Contamination in public databases is well-documented<sup>21</sup> and represents an additional 187 188 confounding factor for k-mer based methods. While several custom tools have been built to deal 189 with imperfect data<sup>22</sup>, there is a need for database 'cleaning' tools that can preprocess a database 190 and evaluate it for both contamination (genome assemblies that contain a mixture of species) and 191 misclassified species and strains (genomes that are assigned a taxonomic ID that is inconsistent 192 with its similarity to other genomes in the database). The misclassification issue often is in the 193 eye of the beholder; species have been named based on morphology, ecological niche, toxin 194 presence/absence, isolation location, 16S phylogenetic placement, and average nucleotide 195 identity across the genome. This, coupled with an often ambiguous species concept in microbial genomes due to horizontal gene transfer and mobile elements<sup>23</sup>, brings into question the reliance 196 197 on the current taxonomic structure for assigning names to microbes sequenced in metagenomic 198 samples. A more robust approach would be for the classification databases to derive their own 199 hierarchical structure directly from the data, rather than taxonomy, and then map back the 200 internally derived hierarchy to widely-used taxonomic names.

201

#### 202 CONCLUSION

Our findings demonstrate that changes in RefSeq over time have influenced the accuracy of *k*mer based classification and profiling methods. Bayesian re-estimation approaches are helpful for species or strain level prediction but can result in false positives and are computationally prohibitive with larger databases. Despite recent progress in *k*-mer based methods for metagenome profiling and classification these tools should likely be used as step one in a multistep process, which also includes read mapping, assembly, feature prediction, and annotation.

- Additionally, priority should be given to the breadth, not depth, of species added to reference
- 210 databases over time.
- 211

### 212 METHODS

#### 213 Acquisition of bacterial RefSeq databases versions 1 through 80

- 214 FASTA files of previous versions of bacterial RefSeq are not publically available for download.
- 215 Therefore, sequences from previous versions of bacterial RefSeq were acquired using custom
- 216 scripts (https://github.com/dnasko/refseq\_rollback). Briefly, the process involved downloading
- 217 the current bacterial RefSeq release (ver. 84 as of the date of the analysis) FASTA files
- 218 (ftp.ncbi.nlm.nih.gov/refseq/release/bacteria) and concatenating them into one file. Then, the
- 219 catalog file associated with the desired version is downloaded
- 220 (ftp.ncbi.nlm.nih.gov/refseq/release/release-catalog/archive), which contains the identifiers for
- sequences present in that version of bacterial RefSeq. Sequence identifiers in that version's
- 222 catalog file are pulled from the current RefSeq FASTA file and written to a new file. Using the
- 223 refseq\_rollback.pl script any version of bacterial RefSeq can be created. For this study only

versions 1, 10, 20, 30, 40, 50, 60, 70, and 80 were recreated.

225

#### 226 Taxonomic classification on simulated datasets

Two simulated read datasets were used to test Kraken and Bracken performance with different versions of the bacterial RefSeq database. The first simulated dataset was downloaded from the Kraken website (ccb.jhu.edu/software/kraken) and was previously used in the Kraken manuscript as a validation set <sup>3</sup>. Briefly, this simulated dataset was composed of 10 known bacterial species: 231 Aeromonas hydrophila SSU, Bacillus cereus VD118, Bacteroides fragilis HMW 615,

232 Mycobacterium abscessus 6G-0125-R, Pelosinus fermentans A11, Rhodobacter sphaeroides

233 2.4.1, Staphylococcus aureus M0927, Streptococcus pneumoniae TIGR4, Vibrio cholerae

234 CP1032(5), and *Xanthomonas axonopodis* pv. Manihotis UA323. Each genome had 1,000

single-end reads (101 bp in size) for a total of 10,000 reads. We selected this dataset as it has

been widely used as a benchmark for other k-mer based classification methods  $^{3,7}$  and represents

a breadth of species. This simulated read dataset was classified against each of the recreated

238 bacterial RefSeq databases using Kraken (ver 1.0) with default settings.

239

240 To test the ability to classify reads from genomes not in the bacterial RefSeq database 10,000

simulated single-end Illumina reads (101 bp) were created using Grinder <sup>24</sup> with default settings

from: (i) a Bacillus cereus genome, B. cereus VD118, not present in RefSeq until verion 60 and

243 beyond; and (*ii*) a novel *B. cereus* genome, *B. cereus* ISSFR-23F<sup>17</sup>, never present in any of the

244 RefSeq versions tested. We decided to use these genomes as they are members of the *B. cereus* 

sensu lato group, containing a collection of species that are known to be challenging for *k*-mer

246 methods to distinguish between  $^{19,20}$ . These datasets were classified with Kraken (ver. 1.0) and

247 Bracken (ver. 1.0.0)<sup>9</sup> both with default settings (Bracken "read-length" set to 101).

248

#### 249 Running Bracken on Kraken output

Bracken (ver. 1.0.0) was run on the output of each Kraken search (except for release 80 and
KrakenMiniDB). Default parameters were used except for "read-length", which was set to 101.

### 253 Bacterial RefSeq diversity metric calculations

- 254 Diversity metrics were calculated for every version of bacterial RefSeq (1-84) by parsing the
- 255 catalog files for each version. An operational taxonomic unit (OTU) table was constructed using
- the NCBI taxonomy identifiers as taxonomic units (see create\_otu\_table.pl in the refseq\_rollback
- repository). The OTU table was imported to QIIME (ver. MacQIIME 1.9.1-20150604)<sup>25</sup>.
- 258 Diversity metrics (Simpson, Shannon, Richness) were calculated using the "alpha\_diversity.py"
- script and plotted using the R base package.
- 260

### 261 ABBREVIATIONS

262 OTU: Operational taxonomic unit; LCA: Lowest common ancestor

#### 263 **DECLARATIONS**

#### 264 Acknowledgements

- 265 This work utilized the computational resources of the NIH HPC Biowulf cluster
- 266 (https://hpc.nih.gov). The authors would like to thank Mihai Pop for his feedback and

267 discussion of this project in its early development.

268

## 269 Funding

- 270 S.K. and A.M.P. were supported by the Intramural Research Program of the National
- 271 Human Genome Research Institute, National Institutes of Health. D.J.N. and T.J.T
- 272 were supported by the FunGCAT program from the Office of the Director of National
- 273 Intelligence (ODNI), Intelligence Advanced Research Projects Activity (IARPA), via

274	the Army Research Office (ARO) under Federal Award No. W911NF-17-2-0089. The
275	views and conclusions contained herein are those of the authors and should not be
276	interpreted as necessarily representing the official policies or endorsements, either
277	expressed or implied, of the ODNI, IARPA, ARO, or the US Government.
278	
279	Availability of Data and Materials
280	Scripts used in this analysis are available on GitHub
281	(github.com/dnasko/refseq_rollback). Datasets and genomes used in this analysis are
282	available online and referenced in the text.
283	
284	Authors' contributions
285	T.J.T. and D.J.N. designed the experiments. D.J.N. wrote the analysis scripts. D.J.N.,
286	S.K, and T.J.T. performed the experiments. D.J.N., S.K., A.M.P. and T.J.T. wrote the
287	paper.
288	
289	Ethics approval and consent to participate
290	NA
291	Consent for publication
292	NA
293	Competing interests
294	NA

## 295 Additional files

296

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358

## 360 Table 1. Fractions of unclassified (Unclass.), correctly classified (Correct), and misclassified

## 361 (Misclass.) simulated reads from ten genomes using Kraken against different versions of

### 362 bacterial RefSeq.

			Genus		Species	
Release	Date	Unclass.	Correct	Misclass.	Correct	Misclass.
1	2003-06-30	0.62	0.38	0.00	0.29	0.08
10	2005-03-06	0.53	0.46	0.01	0.38	0.07
20	2006-11-05	0.49	0.49	0.01	0.40	0.08
30	2008-07-07	0.25	0.74	0.00	0.60	0.07
40	2010-05-07	0.22	0.77	0.00	0.54	0.08
50	2011-11-08	0.21	0.78	0.01	0.52	0.07
60	2013-07-19	0.03	0.96	0.00	0.57	0.09
70	2016-03-03	0.03	0.95	0.01	0.42	0.09
80	2017-01-09	0.03	0.94	0.01	0.28	0.08

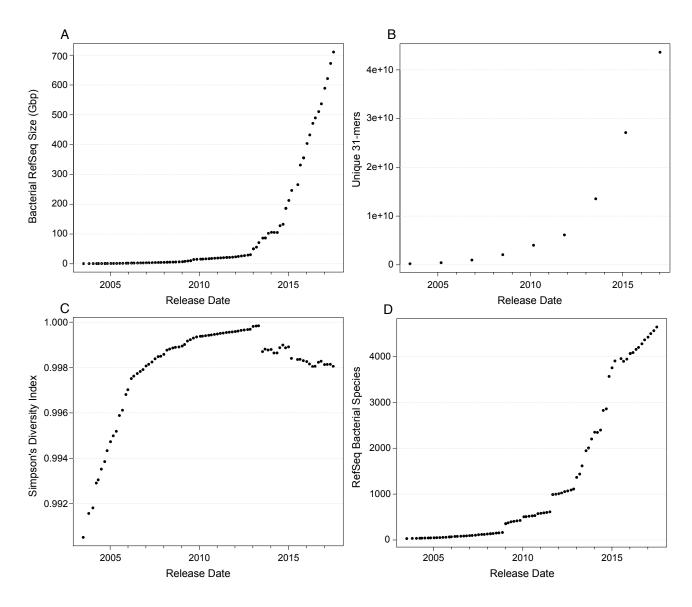




Figure 1: Simpson diversity index of bacterial RefSeq has decreased every release since April 2013. (A) The number of base pairs in bacterial RefSeq continues to grow exponentially, but (B) the number of unique 31-mers and (D) the number of bacterial species added increases slower. (C) The Simpson's diversity index grew every release up to April 2013 where it has declined every release since.

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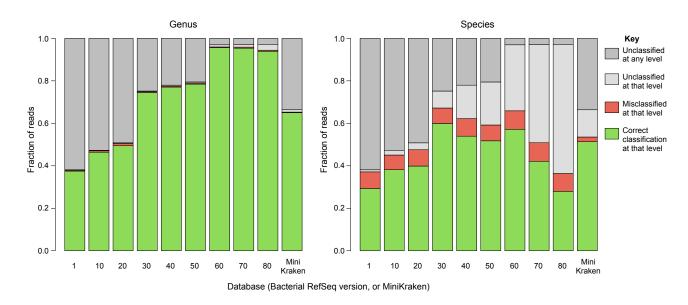
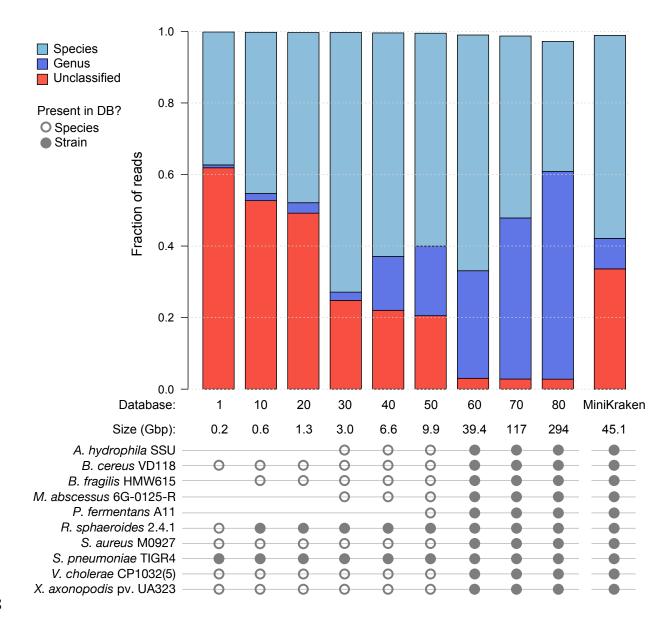


Figure 2: The fraction of correct species classifications (right) decreases in later RefSeq database versions because they are only being classified at the genus level (left). Kraken classification results of simulated reads from known genomes against nine versions the bacterial RefSeq database and the MiniKraken database. Misclassifications at the genus and species levels remain consistently low across database versions. 



<sup>388</sup> 

390 **bacterial RefSeq grows.** Fraction of simulated reads classified at different taxonomic levels,

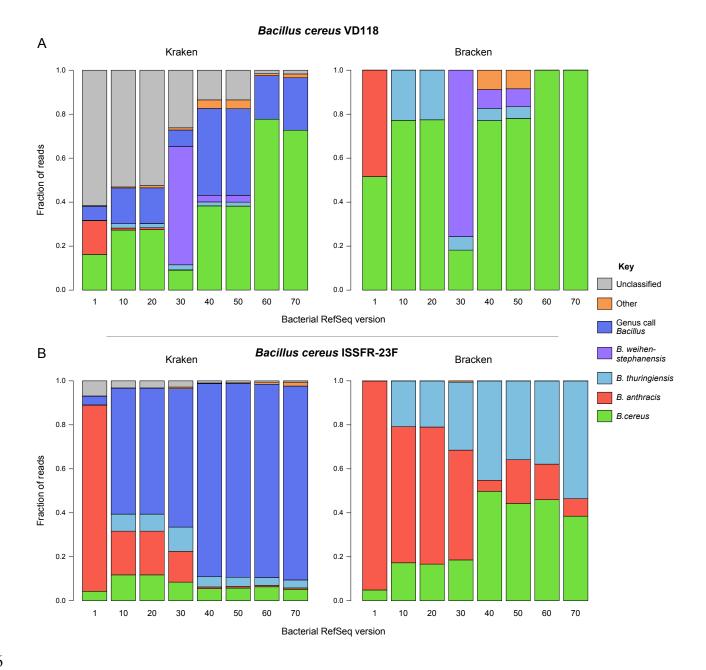
391 regardless of accuracy, using Kraken against ten databases. The circles below indicate when

ach genome's species/strain is in a database. Although the MiniKraken database contains all 10

393 genomes it yields results comparable to bacterial RefSeq version 40.

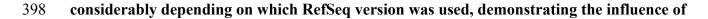
394

**<sup>389</sup>** Figure 3: Species-level classifications decrease, and genus-level classifications increase as









- 399 the database on a *k*-mer based taxonomic classification. (A) Classifying simulated *B. cereus*
- 400 VD118 reads with Kraken (left) and Bracken (right) against different version of RefSeq.
- 401 Species-level classifications varied, and the fraction of unclassified reads decreased with Kraken,

- 402 as the database grew. Once *B. cereus* VD118 appeared in the database (ver. 60) Bracken
- 403 correctly classified every read. (B) Species-level classifications decrease with Kraken as RefSeq
- 404 grows using simulated reads from an environmental *Bacillus cereus* not in RefSeq. Fraction of
- 405 simulated *B. cereus* ISSFR-23F reads classified using Kraken ver. 1.0 (left) and Bracken ver.
- 406 1.0.0 (right) against different versions of bacterial RefSeq. Bracken classification pushed all
- 407 reads to a species-level call, though these classifications were often for other *Bacillus* species.