

1 **Chromosomally Encoded *mcr-5* in Colistin Non-susceptible *Pseudomonas aeruginosa***

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9 **Running Title:** *mcr-5* in colistin-resistant *P. aeruginosa*

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15 **Abstract**

16 Whole genome sequencing (WGS) of historical *Pseudomonas aeruginosa* clinical isolates
17 identified a chromosomal copy of *mcr-5* within a Tn3-like transposon in *P. aeruginosa* MRSN
18 12280. The isolate was non-susceptible to colistin by broth microdilution and genome analysis
19 revealed no mutations known to confer colistin resistance. To the best of our knowledge this is
20 the first report of *mcr* in colistin non-susceptible *P. aeruginosa*.

21 **Manuscript**

22 *Pseudomonas aeruginosa* is a leading cause of infection among immunocompromised
23 patients and those receiving treatment in Intensive Care Units (ICU) (1). Antimicrobial treatment
24 of *P. aeruginosa* is challenging due to the intrinsic resistance of this species to many antibiotics
25 and a proclivity to develop resistance to other antibiotics via point mutations in intrinsic genes
26 (2, 3). Furthermore, *P. aeruginosa* can readily acquire transmissible antibiotic resistance (AbR)
27 genes resulting in the emergence of successful multi- or extensively-drug resistant strains (4).
28 The emergence of these resistant strains has resulted in a greater reliance on colistin (polymixin
29 E) as a key antipseudomonal agent (5). Unfortunately, colistin resistance in *P. aeruginosa* has
30 been extensively reported, primarily due to mutations in regulatory two-component systems (
31 reviewed in (6)). However, colistin resistance mediated by the transferable colistin resistance
32 gene *mcr* has not been described in this species to date. In this report we describe colistin non-
33 susceptible *P. aeruginosa* MRSN 12280 carrying a chromosomal copy of *mcr-5*.
34 *P. aeruginosa* MRSN 12280 was sequenced as part of a larger effort to sequence all *P.*
35 *aeruginosa* isolates in the Multi-drug resistant organism Repository and Surveillance Network
36 (MRSN) repository (n=2,440; manuscript in preparation). The isolate was cultured from a sacral
37 wound of an elderly male patient treated in the USA in 2012. Minimum inhibitory concentrations
38 (MICs) of colistin were determined using broth microdilution (BMD) with cation-adjusted
39 Mueller Hinton broth (CA-MHB) according to the Clinical & Laboratory Standards Institute
40 (CLSI) guidelines, and also with calcium-enhanced Mueller-Hinton (CE-MH) as recommended
41 by Gwozdziński and colleagues for *Enterobacteriaceae* carrying *mcr* (7). *Escherichia coli*
42 MRSN 388734 carrying *mcr-1* (8) and *P. aeruginosa* ATCC 27298 were used as positive and
43 negative controls, respectively. Colistin MICs were 4µg/ml (intermediate) and 8µg/ml (resistant)

44 for *P. aeruginosa* MRSN 12280 in CA-MH and CE-MH medium, respectively (**Table 1**).

45 Notably, the MIC of colistin in the control strains *E. coli* MRSN 388734 and *P. aeruginosa*

46 ATCC 27298 also increased in CE-MH, but interpretations did not change (**Table 1**).

47 Short and long-read whole genome sequencing (WGS) was performed on a NextSeq 550

48 (Illumina, San Diego, CA) and PacBio RS II (Pacific Biosciences, Menlo Park, CA),

49 respectively, as previously described (9). *In silico* multi-locus sequence typing (MLST) assigned

50 *P. aeruginosa* MRSN 12280 to a novel sequence type (ST) that is a single-loci variant of ST-

51 2613. An analysis of the WGS data detected five AbR genes that are commonly found in *P.*

52 *aeruginosa* (*aph(3')-IIB*, *bla_{OXA-50}*, *bla_{PAO}*, *catB7*, and *fosA*) and the recently described colistin

53 resistance gene, *mcr-5* (10). *Mcr-5* was first reported in 2017 in a cluster of colistin non-

54 susceptible *Salmonella enterica* and shares a protein sequence identity of just 36.11% with *Mcr-*

55 1 (10). Borowiak and colleagues reported that the gene was part of a Tn3-family transposon that

56 was found primarily on small, multi-copy ColE-type plasmids. However, in one isolate (*S.*

57 *enterica* 12-02546-2) the gene was present in a single copy on the chromosome and had a

58 colistin MIC of 4mg/L (10). In *P. aeruginosa* MRSN 12280, a single copy of *mcr-5* was also

59 present on the chromosome and was part of an 8,522bp Tn3-family transposon. This transposon

60 was identical to the one described by Borowiak and colleagues except for the insertion of IS5

61 into a gene encoding a putative major facilitator superfamily (MFS) directly downstream of *mcr-*

62 5 (**Figure 1**). The transposon was flanked by 38bp inverted repeats (IR) and generated a 5bp

63 target site duplication (TCCAT) upon insertion.

64 Colistin resistance in *P. aeruginosa* has primarily been attributed to mutations in up to

65 five different two-component regulatory systems (PhoPQ, PmrAB, ParR/S, ColR/S, and CprR/S)

66 (Reviewed in (6)) but to the best of our knowledge, *Mcr*-mediated colistin resistance has not

67 been described in *P. aeruginosa* to date. As mutations in the two-component regulatory systems
68 could potentially contribute to colistin resistance in *P. aeruginosa* MRSN 12280, we examined
69 the amino acid sequences of PmrA, PmrB, PmrE, PhoQ, ParR, ParS, ColR, ColS, MigA, LpxC,
70 and CprS for non-synonymous mutations (6, 11-13). When compared to *P. aeruginosa* PA01, *P.*
71 *aeruginosa* MRSN 12280 had non-synonymous mutations in PhoQ (Y85F), PmrA (L71R),
72 PmrB (S2P, A4T, G68S, Y345H, G362S), ParR (L153R, S170N), and ParS (H398R). However,
73 all of the mutations in PhoQ, PmrA, and PmrB have previously been reported in colistin
74 sensitive strains (12, 13). An analysis of 1,135 *parS* genes from the National Center for
75 Biotechnology Information (NCBI) revealed that 1,117 sequences have an arginine at position
76 398, indicating that the ParS protein from *P. aeruginosa* PA01 is a poor representative of ParS in
77 *P. aeruginosa*. Finally, a search of NCBI for the L153R and S170N mutations in ParR revealed
78 that these mutations are present in a cluster of *P. aeruginosa* belonging to ST-235 from East
79 Asia. *P. aeruginosa* VRFP04 from this cluster has been analyzed in detail and it is colistin
80 sensitive (14). Though additional experiments are underway to confirm these findings, the data
81 strongly suggests that colistin non-susceptibility in *P. aeruginosa* MRSN 12280 is due to Mcr-5.

82 We report the first identification of *mcr-5* in a colistin non-susceptible strain of *P.*
83 *aeruginosa*. The gene was chromosomally encoded and embedded within a Tn3-family
84 transposon that was related to the original transposon carrying *mcr-5* (10). Of note, while part of
85 Mcr-5 and the transposon were identified in two *P. aeruginosa* assemblies using BLASTp (10),
86 no other *P. aeruginosa* isolate in the MRSN repository contained the gene. This suggests that
87 *mcr-5* is not widely distributed in this species but has the potential to disseminate via the Tn3-
88 family transposon.

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144 **Table 1. Colistin susceptibility testing**

Strain	MIC of colistin in ¹	
	CA-MH	CE-MH
<i>P. aeruginosa</i> MRSN 12280	4	8
<i>E. coli</i> MRSN 388634	8	16
<i>P. aeruginosa</i> ATCC 27298	0.25	1

145 Abbreviations used: MIC, minimum inhibitory concentration; CA-MH, cation-adjusted Mueller

146 Hinton; CE-MH, Calcium-enhanced Mueller Hinton.

147 ¹ MICs were performed in BMD according to CLSI guidelines. Results represent the average

148 value from three independent tests.

149 **Figure Legends**

150 **Figure 1. Alignment of Tn3-family transposons carrying *mcr-5*.** Comparison of the Tn3-like
151 transposon carrying *mcr-5* in plasmid pSE13-SA01718 from *Salmonella enterica* (10) with the
152 Tn3-like transposon carrying *mcr-5* in *P. aeruginosa* MRSN 12280. Transposons are encased in
153 a rectangle with the inverted repeats (IR) depicted as shaded, rotated triangles. Open arrows
154 represent coding sequences (red arrows, *mcr-5*; white arrows, genes associated with DNA
155 mobility; grey arrows, other genes) and indicate direction of transcription.

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