

Eco-evolutionary community turnover following environmental change

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Abstract

Co-occurring species often differ in intraspecific genetic diversity, which in turn can affect adaptation in response to environmental change. Specifically, the simultaneous evolutionary responses of co-occurring species to temporal environmental change may influence community dynamics. Local adaptation along environmental gradients combined with gene flow can enhance genetic diversity of traits within populations. Here I build off existing quantitative genetic theory to study community dynamics of locally adapted species in response to temporal environmental change. I show that species with greater gene flow have lower equilibrium population size due to maladaptive immigrant genotypes (migration load). However, following abrupt environmental change that leaves all species initially maladapted, high gene flow species adapt faster due to greater standing genetic diversity. As a result, species can transiently reverse their relative abundances, but sometimes only after long lag periods. If constant temporal environmental change is applied, the community exhibits a shift toward stable dominance by species with intermediate gene flow. Notably, populations of fast-adapting high gene flow species can increase under environmental change because the change suppresses superior competitors with lower gene flow. This eco-evolutionary competitive release stabilizes ecosystem function. The community dynamics observed here parallel the purely ecological successional dynamics following disturbances. My results demonstrate how interspecific variation in life history can have far-reaching impacts on eco-evolutionary community response to environmental change.

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28 **1 Introduction**

29 Genetic diversity in quantitative traits serves as the raw material for selection (Lush
30 1937). Understanding how rapid changes in selection impact populations is a question
31 with tremendous importance in biodiversity conservation, agriculture, and medicine
32 (Gomulkiewicz and Holt 1995; Bell and Gonzalez 2009; Read et al. 2011; Alexander et
33 al. 2014; Lasky et al. 2015; Bay et al. 2017). A substantial portion of genetic diversity
34 in phenotypes within species is maintained due to population adaptation to local
35 environments (Turesson 1922; Clausen et al. 1940; Leimu and Fischer 2008; Hereford
36 2009). Local adaptation is defined as a genotype-by-environment interaction favoring
37 home genotypes (Kawecki and Ebert 2004). When populations are locally adapted,
38 greater gene flow can increase within-population diversity due to immigration from
39 populations adapted to other environments (Barton 2001; Lenormand 2002; Garant et
40 al. 2007). Given that local adaptation is common (Leimu and Fischer 2008; Hereford
41 2009; Sanford and Kelly 2010) and multiple co-occurring species can be simultaneously
42 adapted to local environments, these processes could impact genetic diversity of co-
43 occurring species and community responses to environmental change. Here I build
44 on previous theory to study the complex role gene flow plays in communities due to
45 its effect on genetic diversity, which induces migration load on populations but also
46 speeds up adaptation (Pease et al. 1989; Polechová et al. 2009; Kremer et al. 2012).

47 A major body of theory explores the conditions under which selective gradients lead
48 to stable polymorphism and local adaptation (Haldane 1930; Slatkin 1973; Felsenstein
49 1977; Kirkpatrick and Barton 1997; Behrman and Kirkpatrick 2011; Yeaman and
50 Whitlock 2011; Le Corre and Kremer 2012). When populations are locally adapted,
51 immigrant alleles to a given location may be poorly suited to the local environment,
52 as these immigrants originate from populations adapted to different environments
53 (Haldane 1956; Mayr 1963; Kirkpatrick and Barton 1997; Lenormand 2002; Polechová
54 and Barton 2015). These alleles can impose a “migration load” on populations,
55 reducing population size due to lower average fitness of individuals in a population
56 (Barton 2001; Lenormand 2002; Farkas et al. 2013; Polechová and Barton 2015).
57 Assuming organisms have a limited ability to disperse into appropriate environments
58 (e.g. passive dispersers), migration load increases with increasing rate and spatial scale
59 of gene flow (among other factors discussed below, Slatkin 1973; Kirkpatrick and
60 Barton 1997; Polechová and Barton 2015).

61 The observation that humans are rapidly changing global environments has motivated
62 studies of temporal changes in selection (Bay et al. 2017; Siepielski et al. 2017).
63 Environmental change can cause population decline, extinction, or persistence via
64 plasticity or evolution (Aitken et al. 2008). Theoretical and experimental studies have
65 largely focused on two scenarios of environmental change: 1) a rapid, abrupt shift
66 from a historical selection regime to a new one (Gomulkiewicz and Holt 1995; Orr and
67 Unckless 2008) or 2) sustained change in selection through time (Pease et al. 1989;

68 Lynch and Lande 1993; Polechová et al. 2009; Gonzalez et al. 2013). Most theoretical
69 studies have focused on the binary outcome of whether species survive or go extinct
70 following environmental change. For example, a number of authors have investigated
71 factors influencing the probability of evolutionary rescue (Gomulkiewicz and Holt 1995;
72 Orr and Unckless 2008; Bell and Gonzalez 2009; Uecker et al. 2014), which is defined
73 as adaptation that prevents extinction following environmental change (Gonzalez et
74 al. 2013). Pease et al. (1989) and Lynch and Lande (1993) studied the maximal
75 rates of environmental change that allow populations to persist. Still, little is known
76 about how evolutionary response to rapid environmental change impacts abundance
77 patterns, apart from equilibrium abundance of individual populations (Polechová et al.
78 2009). Despite this gap, community and ecosystem processes are strongly influenced
79 by abundance dynamics of component species, such that understanding abundance
80 responses to environmental change is a key goal of community and ecosystem ecology
81 (Loreau 2010; Clark et al. 2014*b*). An emerging area of inquiry has investigated
82 community evolutionary rescue, roughly defined as evolutionary rescue of multiple
83 co-occurring species (Fussmann and Gonzalez 2013; Kovach-Orr and Fussmann 2013;
84 Low-Décarie et al. 2015).

85 Among the factors that determine population response to environmental change are
86 initial population size and genetic diversity in the trait(s) under selection. When
87 populations are initially small before environmental change, a species faces a greater
88 risk of stochastic extinction following environmental change (Gomulkiewicz and Holt
89 1995). Additionally, if genetic variants do not exist within a population that are
90 beneficial after environmental change then a population will wait for new mutations
91 or immigrant alleles (e.g. Orr and Unckless 2008), a scenario most relevant when
92 adaptation is oligogenic. Alternatively, standing variation within populations may
93 allow more rapid adaptation, if adaptive variants are already present at the time
94 of environmental change (Bonhoeffer and Nowak 1997). Such standing variation
95 can be caused by gene flow along spatial selective gradients (Barton 2001). In
96 particular, quantitative genetic models of local adaptation are relevant to adaptation
97 to anthropogenic change because phenotypes involved in climate adaptation are often
98 complex with polygenic architecture (Bay et al. 2017).

99 The effects of rapid environmental change on biodiversity are partly influenced by how
100 multiple co-occurring species simultaneously respond to environment (Bradshaw 1984;
101 Jackson and Overpeck 2000; Gilman et al. 2010; Urban et al. 2012). Typically studies
102 of community and ecosystem responses to environmental change focus on ecological
103 mechanisms, e.g. interspecific variation in demographic and physiological response
104 to environment (Deutsch et al. 2008; Clark et al. 2014*a*; Lasky et al. 2014). For
105 example, interspecific variation in dispersal ability is expected to have major effects on
106 community response to environmental change, as some species are better able to track
107 spatial shifts in environmental niches (Ackerly 2003; Gilman et al. 2010; Urban et
108 al. 2013). However, most approaches ignore another level of complexity: intraspecific

109 variation and evolutionary response within members of a community. Authors have
110 studied how multiple species simultaneously evolve following environmental change
111 using simulation (De Mazancourt et al. 2008; Moran and Ormond 2015; Vanoverbeke
112 et al. 2015). However, many multi-species models typically focus on species that begin
113 having niche differentiation along climate gradients (e.g. De Mazancourt et al. 2008;
114 Price and Kirkpatrick 2009; Norberg et al. 2012; Moran and Ormond 2015), but what
115 happens for species occupying similar climatic niches remains to be explored (but see
116 Fussmann and Gonzalez 2013; Osmond and Mazancourt 2013). To date there have
117 been few analytical results for how evolutionary responses of multiple species impact
118 community responses to environmental change.

119 Here I build on an existing quantitative genetic theory of local adaptation (Barton
120 2001) and adaptation to a shifting optimum (Pease et al. 1989; Lynch and Lande
121 1993; Polechová et al. 2009). I reframe this theory to demonstrate the complex
122 role interspecific variation in gene flow plays in communities due to its effect on
123 genetic diversity, which induces migration load on populations but also causes faster
124 adaptation (Pease et al. 1989; Polechová et al. 2009; Kremer et al. 2012). I then ask
125 how interspecific variation in gene flow and other traits impact community dynamics
126 following environmental change due to ecological and evolutionary processes.

127 2 Model and Results

128 I start with a model of locally-adapted populations following Pease et al. (1989),
129 Barton (2001) and Polechová et al. (2009), a stochastic version of which was studied
130 by Polechová and Barton (2015). The model I use is a deterministic model of a
131 population with logistic growth and a quantitative trait z subject to hard selection
132 with a spatially-varying selective gradient. The mean per capita reproductive rate is
133 given by

$$\bar{r} = r_m \left(1 - \frac{N}{K}\right) - \frac{(\bar{z} - \theta)^2}{2V_S} - \frac{V_P}{2V_S} \quad (1)$$

134 where r_m is population growth rate of optimal phenotype individuals at low density,
135 N is census population size, K is carrying capacity, and V_P is variance of phenotype z .
136 The first term on the right-hand side of equation 1 determines a reduction in fitness
137 due to negative density dependence. The second term gives reduction in fitness due
138 to the mismatch between the population mean phenotype \bar{z} and the local optimum
139 θ , and V_S gives the inverse strength of stabilizing selection. Even if the population
140 is adapted to the local optimum (i.e. $\bar{z} = \theta$) there still may be many maladapted
141 individuals (i.e. $V_P > 0$), whose contribution to population mean fitness is determined
142 by the last term in equation 1.

143 The optimal trait value θ changes in space (x) at rate b such that $\theta(x) = bx$ (Kirkpatrick
144 and Barton 1997). The mean trait \bar{z} at a given location x changes through time due

145 to curvature of the cline in space, asymmetric gene flow (modeled as a Gaussian with
146 standard deviation σ) across the cline due to spatial trends in abundance, and selection,
147 given by the first three terms on the right hand side of equation 2, respectively

$$\frac{\partial \bar{z}}{\partial t} = \frac{\sigma^2}{2} \frac{\partial^2 \bar{z}}{\partial x^2} + \sigma^2 \frac{\partial \ln(N)}{\partial x} \frac{\partial \bar{z}}{\partial x} + V_G \frac{\partial \bar{r}}{\partial \bar{z}}. \quad (2)$$

148 The final term in equation 2 is the classic quantitative genetic result where change
149 in the mean trait \bar{z} is a function of genetic variance (V_G) multiplied by the selection
150 function ($\frac{\partial \bar{r}}{\partial \bar{z}}$). Population dynamics at x are given by

$$\frac{\partial N}{\partial t} = \frac{\sigma^2}{2} \frac{\partial^2 N}{\partial x^2} + \bar{r}N \quad (3)$$

151 where the first term on the right-hand side gives change to due spatial trends in
152 abundance, and the second term gives change due to average individual fitness.
153 Note that here there is no frequency or density-dependent selection, i.e. intraspecific
154 competition (or apparent competition) is not dependent on z in any way, beyond the
155 effects of z on N . This assumption may be well-justified for traits involved in abiotic
156 stress-tolerance (e.g. cold or heat tolerance) where selection does not result in any
157 diversity in z .

158 Assuming constant K through space, a stable equilibrium exists where all populations
159 are locally adapted along the linear environmental gradient b , i.e. $\bar{z} = \theta$ at all x
160 (Barton 2001). An additional consequence of local adaptation and a linear cline in
161 \bar{z} is that $\frac{\partial^2 \bar{z}}{\partial x^2} = 0$ and constant population size in space, $\frac{\partial \ln(N)}{\partial x} = 0$. I ignore spatial
162 boundary conditions that would result in asymmetric gene flow.

163 Barton (2001) allowed genetic variance within a population (V_G) to change as a
164 function of gene flow. As gene flow increases, so does immigration of maladaptive
165 genotypes into any given population such that $V_G = b\sigma\sqrt{V_S}$ and $V_P = V_G + V_E$ where
166 V_E is stochastic environmental variation in z (Barton 2001).

167 2.1 Impacts on community structure

168 Two traits that ecologists commonly study are important in this model: the rate
169 and scale of dispersal/gene flow (determined by σ) and reproductive rate at low
170 density (r_m). Maladapted immigrants depress mean fitness (known as migration load,
171 equation 1). The equilibrium census population size (Polechová and Barton 2015) as
172 a proportion of carrying capacity K , \hat{N} , is given by

$$\hat{N} = 1 - \frac{b\sigma}{2\sqrt{V_S}r_m} + \frac{V_E}{2V_Sr_m} \quad (4)$$

173 where the second term on the right gives migration load. Migration load can thus
174 introduce uneven community structure when species differ in σ or r_m . To identify the
175 maximum σ capable of persistence I set \hat{N} to zero and solve the inequality to obtain

$$\sigma < \frac{2V_S r_m - V_E}{b\sqrt{V_S}} \quad (5)$$

176 Here I am interested in complex effects of species traits that might yield unexpected
177 results under environmental change. While greater r_m decreases migration load
178 (equation 4) it does not impact the rate of adaptation $\frac{\partial \bar{z}}{\partial t}$ (equation 2). However, gene
179 flow, σ , plays a more complex role.

180 To study how interspecific variation in σ could structure communities along spatiotem-
181 poral environmental gradients, I now consider a community of species that vary only
182 in σ (but not other parameters e.g. K , V_S , V_E). For mathematical convenience I start
183 with communities lacking species interactions. I follow with simulations that introduce
184 competition among species.

185 In the Barton (2001) model, greater σ increases V_G and migration load and thus
186 decreases equilibrium population size. From equation 4, the proportional reduction in
187 \hat{N} due to migration load is equal to $\frac{b\sigma}{2\sqrt{V_S r_m}}$. I differentiate with respect to σ to obtain

$$\frac{d\hat{N}}{d\sigma} = -\frac{b}{2\sqrt{V_S r_m}} \quad (6)$$

188 which gives the slope of species equilibrium abundance versus gene flow. Thus the
189 species abundance distribution for a community (McGill et al. 2007) could be obtained
190 using the distribution of σ and applying equation 6. The parameters on the right of
191 equation 6 are each constrained to be positive so that when holding these constant
192 across species of varying σ there is a negative relationship between σ and \hat{N} . The effect
193 of migration load is stronger and abundance distribution is steeper as the selective
194 gradient b is steeper.

195 Note that in the equation (4) for \hat{N} , species that differ in b (the slope of selective
196 gradients) will have similar differences in \hat{N} as species differing in σ . The product $b\sigma$
197 gives change in optimal phenotype z over one dispersal standard deviation (Kirkpatrick
198 and Barton 1997). Here I focus on variation in σ among species, given that interspecific
199 variation in dispersal ability of propagules and gametes is a major interest in community
200 ecology.

201 **2.2 Abrupt environmental change and transient community** 202 **turnover**

203 The interesting effects of gene flow in a community context arise from the dual role
204 of σ following environmental change. Greater σ can have a fitness benefit when

205 population mean traits differ from the optimum, $\bar{z} \neq \theta$, such as in populations that
206 have experienced recent environmental change (Polechová et al. 2009; Kremer et al.
207 2012) or populations colonizing new environments. Greater σ proportionally increases
208 V_G , which proportionally increases the speed of adaptation $\frac{\partial \bar{z}}{\partial t}$ for a given selection
209 regime $\frac{\partial \bar{r}}{\partial \bar{z}}$ (third term on right-hand side of equation 2).

210 I studied the effect of σ on the speed of adaptation using numerical simulations. I
211 simulated non-overlapping generations whose dynamics were governed by discretized
212 versions the above equations. Simulations were initialized with a locally-adapted
213 population at equilibrium population size, $N = \hat{N}$ and $\bar{z} = \theta_x$ where the subscript
214 x on θ indicates the optimal trait is for location x . I chose biologically plausible
215 parameter values (although below I study other values): $b = 0.05$, $V_S = 1$, $V_E = 0.05$,
216 $r_m = 0.5$, $x = 0$ and thus $\theta_x = 0$ (Polechová and Barton 2015). I then imposed an
217 instantaneous change in θ_x such that a new phenotype, $\theta_{x1} = 1$, was favored, and the
218 change in selection was the same at all locations, i.e. the slope b of the spatial gradient
219 did not change, $\theta_1(x) = bx + 1$ (Figure 1). This scenario is mathematically convenient
220 because all populations experience the same relative change and dynamics and thus
221 no spatial trend in abundance emerges ($\frac{\partial \ln(N)}{\partial x} = 0$) nor does the cline in \bar{z} change
222 ($\frac{\partial^2 \bar{z}}{\partial x^2} = 0$). As a result, V_G is unchanged.

223 I first compare evolution of z for two species differing only in σ ($\sigma_1 = 0.326$ and
224 $\sigma_2 = 3.069$). Both species were subject to the same selective gradient $b = 0.05$ and the
225 clines in the mean phenotype \bar{z} of the two species were equal before environmental
226 change, but with the second species having greater variance within any local population
227 (i.e. greater V_G , Figure 1). I found rapid adaptation to θ_{x1} in the high gene flow species
228 with the low σ species lagging far behind (Figure 1). I then simulated communities
229 with a log uniform distribution of σ values across 100 species under the same conditions
230 as the two example species to illustrate the differences in adaptation due to σ .

231 Faster adaptation following a shift in environment (Figure 1) will lead to more rapid
232 recovery of population mean fitness because the difference between \bar{z} and θ decreases
233 more rapidly (eqn. 1). Although species with high σ are less abundant than low
234 σ species in communities in a stable environment (eqn. 4), the faster adaptation
235 of high σ species may allow them to increase their relative abundance following an
236 environmental change. These two example species differ only in σ ($\sigma = 0.326$ and
237 $\sigma = 3.069$, respectively) and exhibit a transient reversal in relative abundance as the
238 high σ species is more abundant for an interval following the environmental change
239 (Figure 2). The reversal is transient because the stable environment after change
240 favors low σ .

241 In a diverse community with species having a range of gene flow one can ask how
242 composition might shift due to different evolutionary responses. From the aforemen-
243 tioned simulation of species with a range of σ values (Figure 1E), I calculated which
244 species was most abundant at each time point. Under equilibrium, the species with
245 lowest σ has highest N (eqn. 4 and Figure 2). Following an instantaneous shift in θ ,

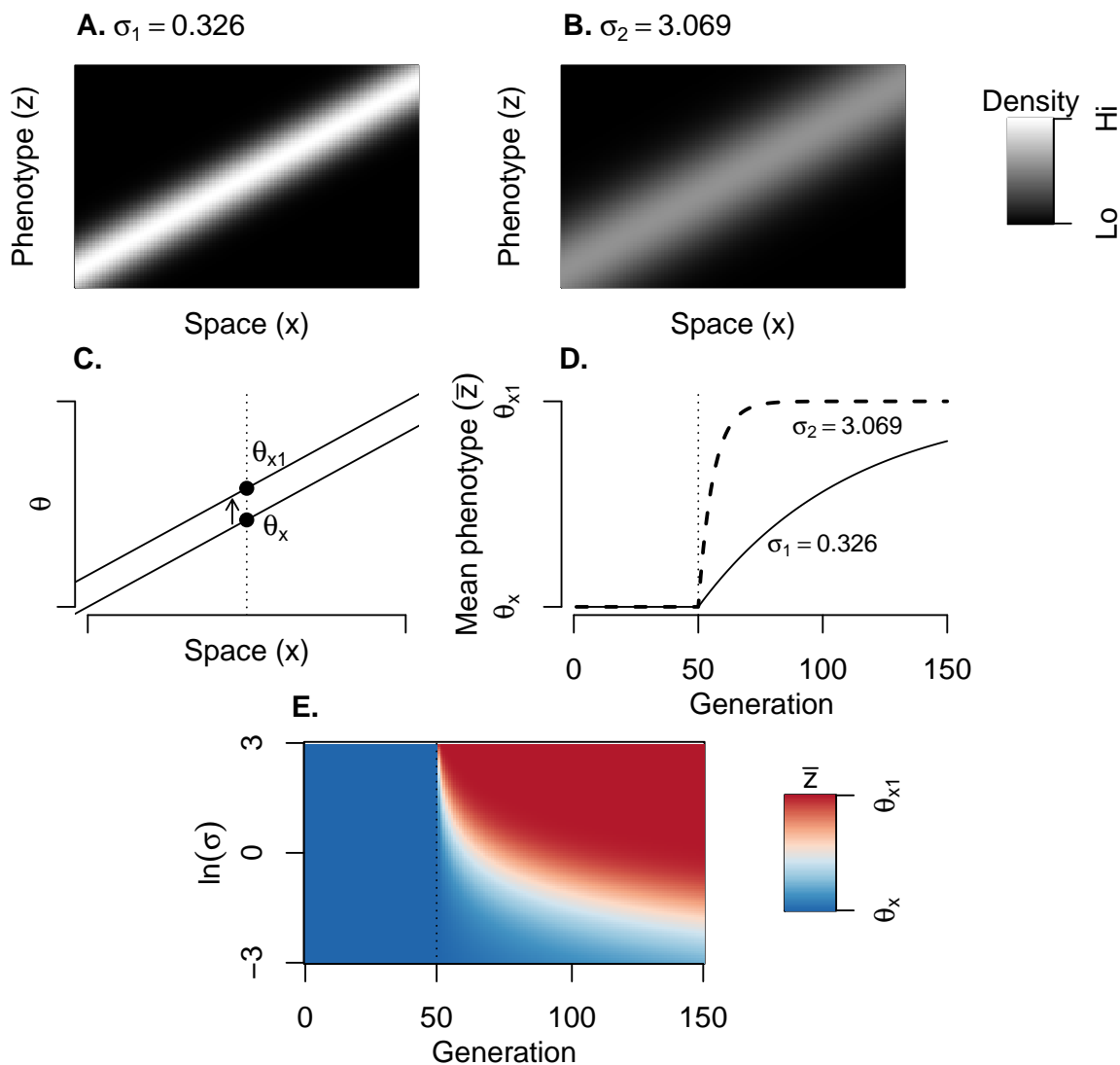


Figure 1: In a locally adapted system, interspecific variation in σ , determining the rate and scale of gene flow, determines differences in genetic diversity and rate of adaptation. Species with low (A) and high (B) σ (gene flow) are subject to the same selective gradient b (favoring an increase in phenotype value through space from left to right) and all populations are locally adapted. However, the high σ species (B) has higher diversity of the trait under selection within populations (V_G) at a given location in space (evident via thicker gray region for any given location along the x-axis) due to maladaptive immigration. I simulated an instantaneous change in optimal phenotype (C) at generation 50. Higher σ species adapt to the new optimum faster, (D) comparing low and high σ species, (E) comparing trait evolution for species with a range of σ values. (E) Blue is the optimal trait prior to the change, and all species begin locally adapted despite differences in V_G . Red is the optimal trait following the change, and high σ quickly adapt while low σ species lag. Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, $r_m = 0.5$, and $\theta_{x1} - \theta_x = 1$.

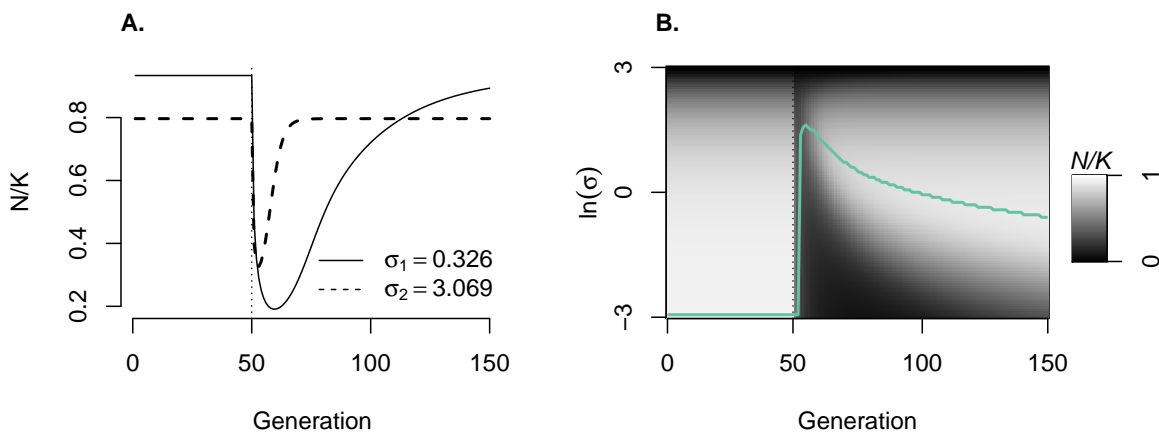


Figure 2: Changes in population size following an abrupt environmental change for species differing only in σ . (A) Time series of population size of two species (the same species as in Fig. 1A-B,1D), one with high σ and high V_G (dashed line) and one with low σ and low V_G (solid line). Populations are at equilibrium for the first 50 generations, after which an instantaneous environmental change occurs. Following this change, the species with high σ adapts faster and reaches equilibrium N before the species with low σ . (B) Comparing N trajectories for species with a range of σ values. High σ quickly adapt while low σ species lag. The most abundant species at any generation is indicated by the green line. Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, $r_m = 0.5$, and $\theta_{x1} - \theta_x = 1$.

246 higher σ species dominate but gradually give way to lower σ species because all species
 247 \hat{N} are unchanged. However, the lag before poor dispersers adapt can be quite long
 248 given the very low rate of adaptation for the lowest σ (Figure 2). This interspecific
 249 variation in adaptation following environmental change will likely have impacts on
 250 the distribution of traits in a community, which is often of interest to community and
 251 ecosystem ecologists (Muscarella and Uriarte 2016; Šímová et al. 2018). For example,
 252 ecosystem function may be influenced by the mass-averaged functional traits in a
 253 community (Grime 1998). I study dynamics in community-weighted mean z in the
 254 Appendix.

255 I next studied how factors that mediate the tradeoffs associated with σ (migration
 256 load versus speed of adaptation) impact community dynamics. Because the transient
 257 advantage of higher σ species comes from the faster approach of \bar{z} to new θ (Equation
 258 2), the magnitude of environmental change might influence the degree of community
 259 turnover. Under a weak shift in θ , the benefit to adapting faster for high σ species is
 260 lower. Figure 3 illustrates these effects. When the magnitude of the environmental
 261 shift is large, community turnover (as determined by which species dominate following
 262 the environmental shift) is also large. Notably, subtle shifts in environment lead to
 263 subtle, though delayed changes in the most dominant species (blue lines in Figure 3).
 264 This lag emerges because when a species starts with greater N at equilibrium the

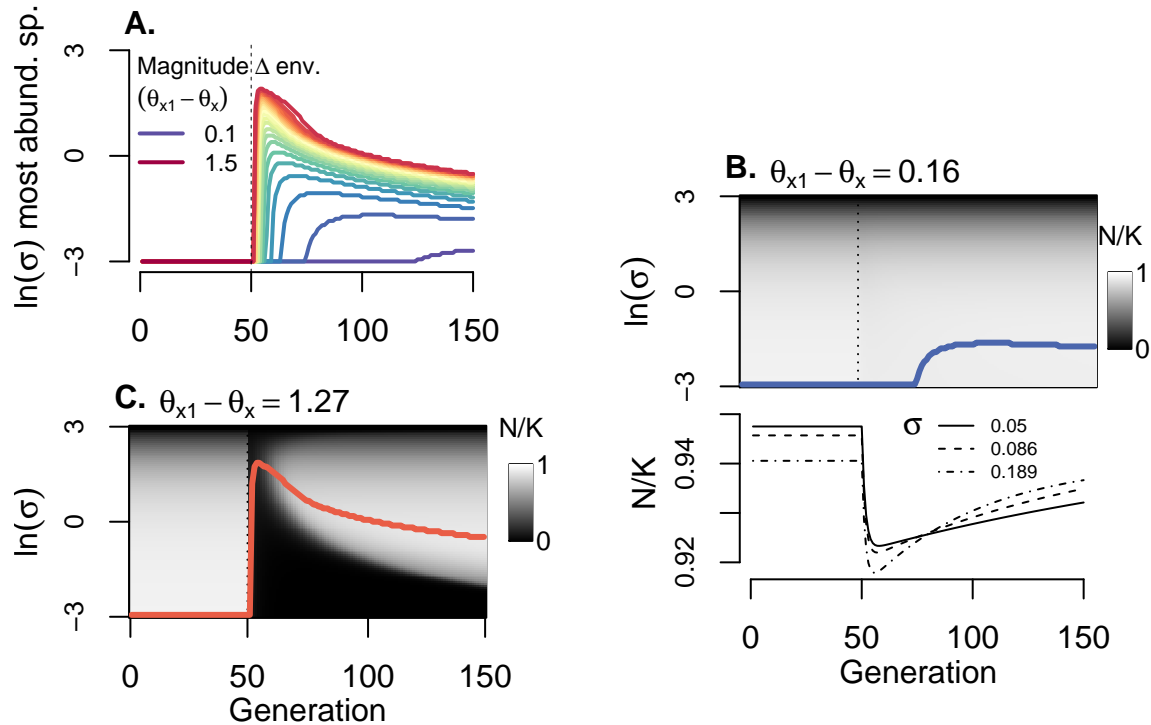


Figure 3: How the magnitude of environmental shift affects the magnitude of community turnover. (A) In general, the greater the environmental change, the higher the σ of the most abundant species soon after the environmental change (B). When the environmental change is too extreme, all species go extinct, as for a change in θ of 1.5 in this example. (C) When environmental change is smaller, a lag between environmental change and change in species relative abundances can occur. Populations are at equilibrium and adapted to θ_x for the first 50 generations, when an instantaneous environmental change to θ_{x1} occurs. Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, and $r_m = 0.5$.

265 differences between species in maladaptation take time to erode the initial advantage
 266 (Figure 3). Despite the lag in reversal of species relative abundances, the differences
 267 among species in \bar{r} are quickly evident in the form of differences in $\frac{\partial N}{\partial t}$ (i.e. there is
 268 rapid emergence of differences among species in slope of N trajectories, Figure 3B).

269 Migration load is ameliorated by high r_m (equation 4), thus r_m may impact eco-
 270 evolutionary community dynamics. Greater r_m reduces the effects of maladaptive
 271 immigration on \hat{N} and allows for persistence (i.e. $\hat{N} > 0$) of species with higher σ
 272 (inequality 5). My simulations showed opposing effects of r_m on community dynamics.
 273 When r_m is low, high σ species cannot persist and thus the magnitude of community
 274 turnover is lower. However, because r_m is low, the recovery of species from low density
 275 is slow, and the community is dominated by relatively higher σ species for a long
 276 period of time (Figure 4). By contrast, high r_m allows for high σ species and the rapid

277 environmental change causes strong, but shorter lived, community turnover.

278 Interspecific trait variation is often correlated across multiple trait axes, corresponding
279 to ecological strategies and life histories. Thus it is unlikely that empirical variation
280 in σ would be independent of other traits. To explore potential impacts of trait
281 covariation, I studied the situation where σ and r_m positively covary such that higher
282 gene flow species also exhibit higher per capita population growth when rare. For
283 example, plants with high reproductive rates tend to have greater dispersal distances
284 (Beckman et al. 2018). To test how this trait covariation would influence eco-
285 evolutionary community turnover, I simulated a positive relationship similar to the
286 observed empirical relationship (Beckman et al. 2018), $r_m = a + c \ln(\sigma)$, where a is an
287 intercept and c determines the rate at which r_m increases for species of higher σ . This
288 correlation has opposing effects on migration load and \hat{N} : r_m decreases load but σ
289 increases load (equation 4). Thus intermediate σ species have greatest abundance at
290 equilibrium (Figure 4). Notably, this correlation between r_m and σ leads to weaker
291 eco-evolutionary community turnover because intermediate σ species were already
292 dominant before environmental change so their dominance after environmental change
293 means the community is relatively consistent.

294 Migration load is also ameliorated under shallower environmental gradients (lower
295 b), though low b also reduces V_G and hence adaptation. In nature, the slope of
296 environmental gradients varies in space and are thought to be important drivers of
297 biodiversity patterns (Yeaman and Jarvis 2006). An important observation is that in
298 a system with low b , there will be predominantly gene flow between like environments.
299 The slope of the curve relating species abundance to gene flow ($\frac{d\hat{N}}{d\sigma}$) is proportional to
300 b thus lower b will result in a abundance curve, i.e. a more even community. That is,
301 migration load is reduced and species differing in σ have similar abundances.

302 I investigated the impacts of varying the slope of spatial gradients on turnover in
303 communities following rapid environmental change. When I varied b , the most obvious
304 impact is on the magnitude of community turnover following environmental change.
305 Immediately after the environmental change, high σ species dominate when b is low.
306 Note that when b is low, differences in abundance of species differing in σ are subtle
307 due to low migration load, though there is relatively high turnover in which species are
308 most abundant following the environmental change. When b is high, the environmental
309 change results in turnover favoring species of intermediate σ . Surprisingly, the change
310 in relative species abundances following the environmental change happens at a similar
311 rate regardless of b (lines in Figure 5 have similar trajectories following environmental
312 change), although higher b results in faster return to equilibrium because the initial
313 community turnover was less. The consistency of the rate of community turnover
314 is likely due to species proportional differences in V_G and rate of adaptation being
315 constant despite differences in b (equation 2).

316 Barton (2001) and Polechová and Barton (2015) investigated how faster change in
317 environments at range margins, i.e. increasing magnitude of b , impacts local adaptation.

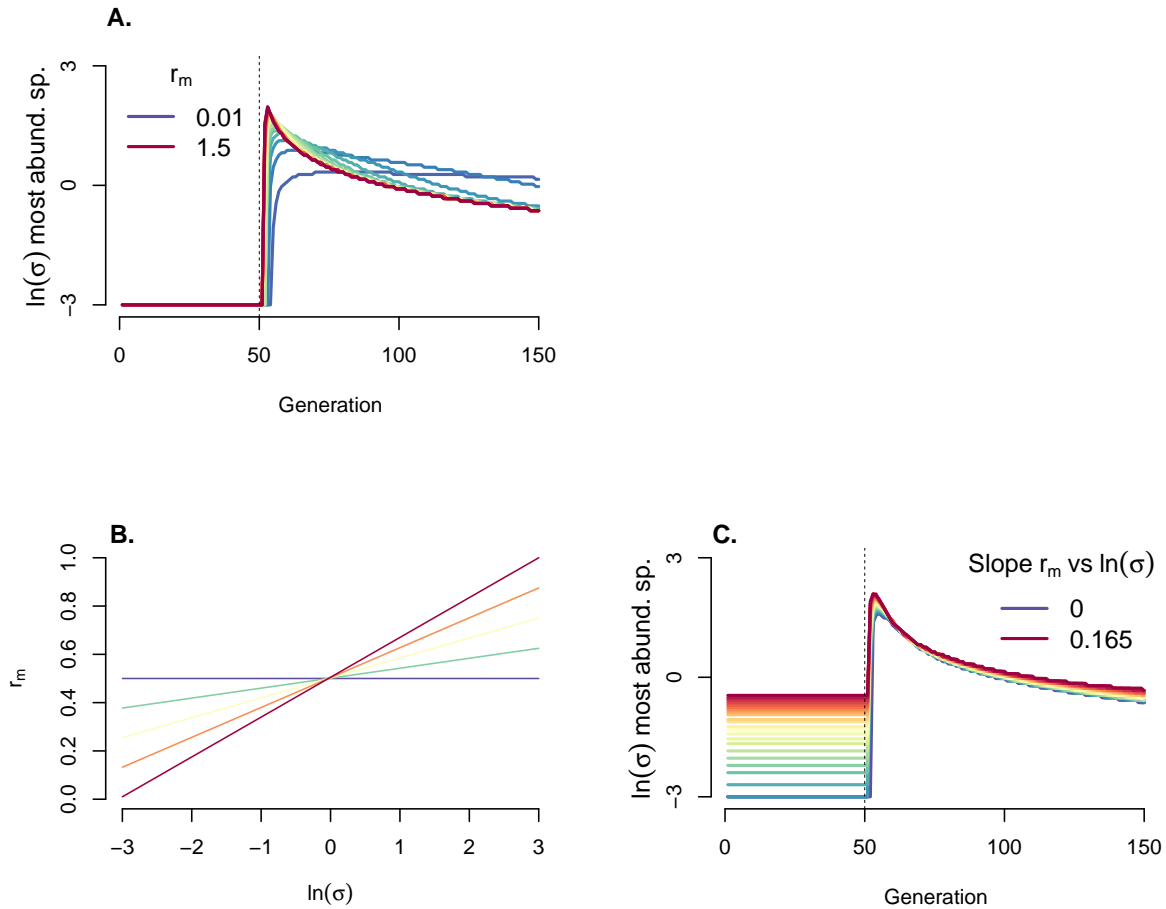


Figure 4: (A) r_m affects the magnitude of community turnover following an abrupt environmental change. Greater r_m results in an initially greater community turnover because r_m reduces migration load and allows high σ species to leverage their faster adaptation following environmental change. Lower r_m increases migration load, limits the ability of high σ to take advantage of their faster adaptation, but also slows the rebound of eventually dominant low σ species. (B) Correlation between reproductive rate at low density (r_m) and gene flow (σ) affects the magnitude of community turnover following an abrupt environmental change (C). Greater correlation results in dominance by intermediate (as opposed to low) σ species at equilibrium under constant environments. After 50 generations the instantaneous environmental change occurs and higher σ species briefly dominate. Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, and $\theta_{x1} - \theta_x = 1$.

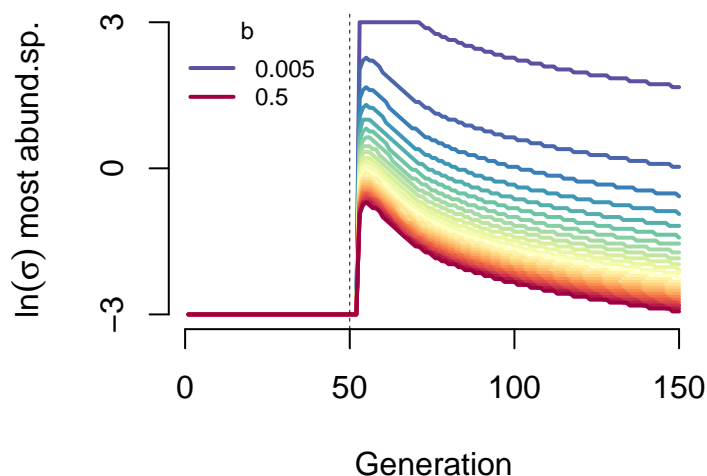


Figure 5: The slope of the spatial selective gradient (b) affects the magnitude of community turnover following an abrupt environmental change. Greater b results in dominance by intermediate σ species following abrupt environmental change (imposed after 50 generations). Lower b allows higher σ species to briefly dominate, although in these scenarios migration load is low and relative abundance at equilibrium (\hat{N}) under stable environments (generations 1-50) is only weakly related to σ . Parameter values (unless otherwise noted) were $V_S = 1$, $V_E = 0.05$, $r_m = 0.5$, and $\theta_{x1} - \theta_x = 1$.

318 My results on how b influences community turnover due to differential evolutionary
319 response to environmental change may apply to such changes in b in space. The
320 present model can be applied assuming that the rate of change in b is subtle, such
321 that $\partial\bar{z}/\partial x$ remains approximately linear. If b is sharper at range margins (for an
322 assemblage of species, this would correspond to ecotones at the margin of ecoregions,
323 for example along very steep altitudinal gradients), migration load would be stronger at
324 margins and would have a stronger influence on community composition at equilibrium
325 (i.e. steeper $\frac{d\hat{N}}{d\sigma}$). However, following environmental change, the change in species rank
326 abundance will be greater in the range core (low b) while there would be lesser change
327 in species rank at range margins (high b).

328 **2.3 Community turnover under sustained environmental** 329 **change**

330 Temporal environmental change can take any functional form. In the previous section
331 I simulated an instantaneous shift in environment that then stabilized (Gomulkiewicz
332 and Holt 1995; Orr and Unckless 2008). Alternatively, environments may undergo
333 more gradual sustained directional shifts. This scenario has been analyzed previously
334 by Pease et al. (1989), Lynch and Lande (1993), and Polechová et al. (2009). Here, I
335 build on this framework by explicitly considering the role of gene flow on population

336 dynamics in this scenario. In the Lynch and Lande (1993) model, the optimum θ
 337 changes at a rate k per unit time t , so that $\theta(x, t) = bx + kt$ (Polechová et al. 2009).
 338 After a enough time has passed to allow for a balance between adaptation versus the
 339 shifting optimum, the mean trait (\bar{z}) at location x lags behind the optimum a stable
 340 distance, which Lynch and Lande (1993) calculated as equal to $-k\frac{V_S}{V_G}$. In the present
 341 model, greater σ increases V_G and causes lower lag in \bar{z} behind the shifting optimum.
 342 Substituting the Barton (2001) equation for V_G in a locally adapted system into the
 343 previous expression results in a lag in \bar{z} equal to

$$(\theta + kt) - \bar{z} = -\frac{k\sqrt{V_S}}{b\sigma} \quad (7)$$

344 i.e. lag in \bar{z} for a given species was proportional to σ^{-1} (Polechová et al. 2009 identified
 345 this expression in a population genetic model of this scenario). Thus stronger stabilizing
 346 selection reduces the lag, though to a lesser degree than identified by Lynch and Lande
 347 (1993; $\sqrt{V_S}$ versus V_S , Kremer et al. 2012). This is because when stabilizing selection
 348 is stronger (low V_S) the fitness advantage of adapted genotypes is higher but stronger
 349 stabilizing selection also reduces V_G from immigration, slowing adaptation.

350 Lynch and Lande (1993) also derived the critical rate of environmental change above
 351 which populations go extinct (assuming large N_e) as $k_c = V_G\sqrt{2\frac{r_m - \frac{V_P}{2V_S}}{V_S}}$ (see also
 352 Polechová et al. 2009). I substitute the Barton (2001) equation for V_G in a locally
 353 adapted system into the previous equation to obtain

$$k_c = b\sigma\sqrt{2r_m - \frac{b\sigma}{\sqrt{V_S}} - \frac{V_E}{V_S}}. \quad (8)$$

354 This equation shows how k_c is non-monotonically related to σ , i.e. k_c is greatest for
 355 intermediate values of σ (Polechová et al. 2009). Low σ species have low V_G , and
 356 hence slower $\frac{\partial\bar{z}}{\partial t}$ but high σ species suffer from high migration load (high V_P). The
 357 difference in this sustained environmental change scenario (compared to the abrupt
 358 change scenario above) is that slower $\frac{\partial\bar{z}}{\partial t}$ continues to be a drag on \bar{r} (known as lag
 359 load), whereas after the abrupt change that then stops, slower $\frac{\partial\bar{z}}{\partial t}$ becomes unimportant
 360 as \bar{z} approaches θ .

361 To determine how the shifting optimum impacts community structure as t becomes
 362 large, I substituted the lag in \bar{z} to the previous equation for \hat{N} (equation 4). Thus at
 363 equilibrium trait lag under an environment shifting so that the optimal trait changes
 364 at rate k ,

$$\hat{N} = 1 - \frac{b\sigma\sqrt{V_S} + V_E}{2V_S r_m} - \frac{k^2}{2b^2\sigma^2 r_m} \quad (9)$$

365 where the first subtracted term includes migration load, which worsens with σ , while
 366 the second subtracted term gives the lag load, which is ameliorated by σ . These
 367 opposing effects result in species with intermediate values of σ and hence V_G being

368 most abundant (Figure 6, Polechová et al. 2009). Differentiating with respect to σ gives

369

$$\frac{d\hat{N}}{d\sigma} = \frac{k^2}{b^2\sigma^3 r_m} - \frac{b}{2\sqrt{V_S} r_m}. \quad (10)$$

370 The maximum \hat{N} is attained by species with σ that cause the right hand side of
371 equation 10 to equal zero, i.e. the σ with maximum \hat{N} is equal to $(2k^2\sqrt{V_S})^{1/3}/b$.
372 Note this expression equals zero when k is zero, thus consistent with results on locally
373 adapted systems in constant environments where $\sigma = 0$ is favored due to lack of
374 migration load (equation 6). Thus greater rates of environmental change through time
375 (k) favor higher σ species, but at a decreasing rate ($k^{\frac{2}{3}}$, equation 10).

376 In this scenario of sustained environmental change, steepening selective gradients
377 (higher b) results in a lower σ having maximum \hat{N} . Thus these results are similar to
378 those following an abrupt change in environment: at range margins or ecotones where
379 b may be steeper, the magnitude of change in the most abundant species will be less,
380 compared to where b is shallower.

381 2.4 Effects of species interactions

382 Species interactions could change the relative importance of some of the processes
383 studied above. For example, interspecific competition could depress the mean fitness
384 of species, pushing them closer to extinction, and also exacerbate relative population
385 differences. I simulated both scenarios of environmental change with non-zero species
386 interactions. Here I present simulation results for species within a community compet-
387 ing with each other, using the Lotka-Volterra form. Instead of equation 1, I used a
388 discrete time version of the following

$$\bar{r} = r_m \left(1 - \frac{N_i - \sum_{i \neq j}^J N_j \alpha_{ij}}{K}\right) - \frac{(\bar{z} - \theta)^2}{2V_S} - \frac{V_P}{2V_S} \quad (11)$$

389 where N_i is the population size of the focal species i and there are J total competitor
390 species each with population sizes of N_j . α_{ij} determines the strength of interspecific
391 competition. Interactions were symmetric among species such that all $\alpha_{ij} = \alpha_{ji}$.

392 I here repeat analyses from above, but with $\alpha_{ij} > 0$. Note that per equation 2, I
393 assume adaptation is not influenced by such competitive interactions (i.e. competition
394 does not influence $\frac{\partial \bar{r}}{\partial \bar{z}}$; α_{ij} is unrelated to z_i and z_j) (Fussmann and Gonzalez 2013; cf.
395 Osmond and Mazancourt 2013). In simulations, I initiated species at a low abundance
396 ($N = 10^{-5}$), but then allowed 500 generations for population growth with interspecific
397 competition before imposing change in θ .

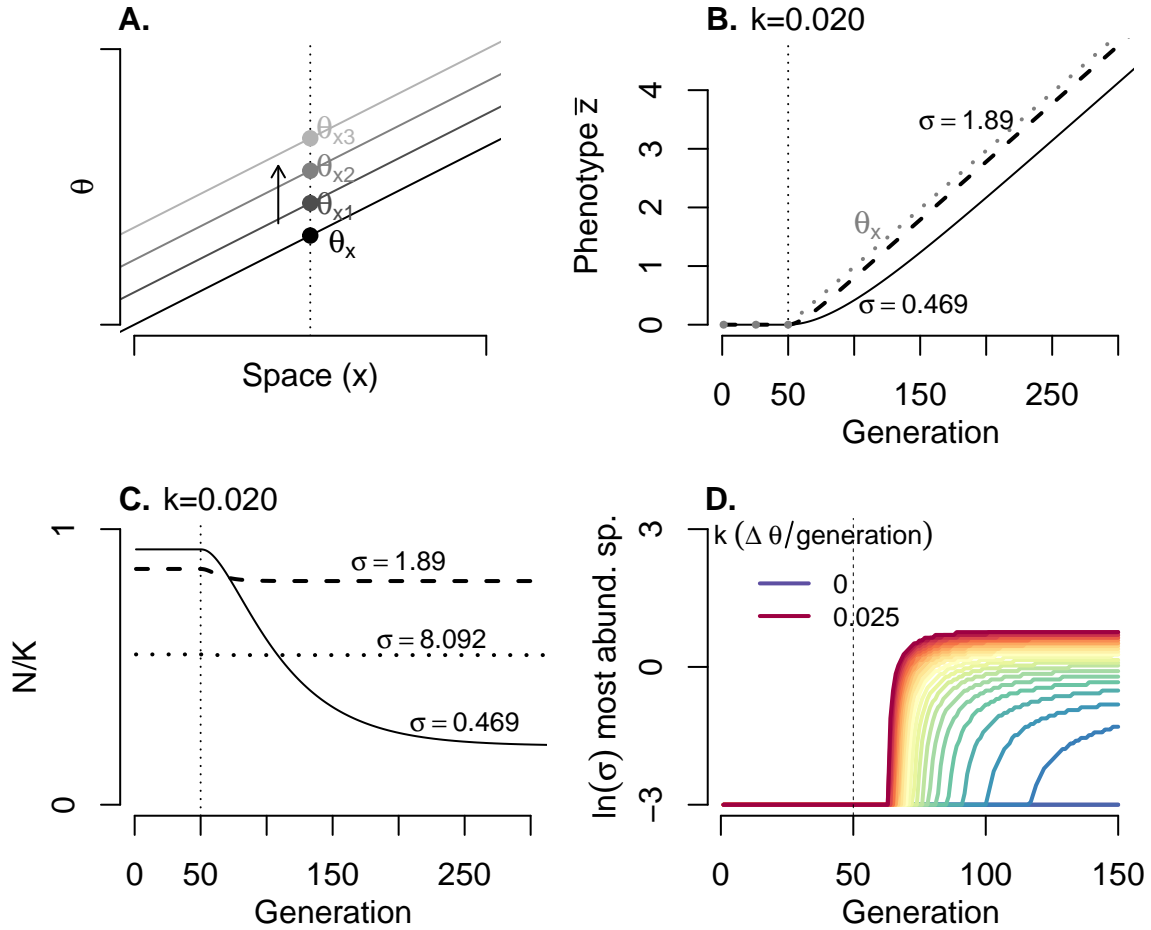


Figure 6: Effects of a sustained environmental change (i.e. a change in θ_x through time). (A) Illustration of the scenario of shifting θ across all locations, from a historical θ_x to which species were locally adapted, to new θ . (B-D) Environment is constant (constant θ) until the vertical dashed line at which point θ_x changes at a constant rate k . (B-C) Illustration with $k = 0.020$ for example species. (B) Evolution of \bar{z} for two example species differing in σ relative to the shifting optimum (θ_x). (C) Population size trajectories for the same two species in addition to one higher σ species. (D) Effects of differing rate of environmental change (k) on community turnover (i.e. the most abundant species under environmental change). Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, and $r_m = 0.5$

398 2.4.1 Species interactions and response to abrupt environmental change

399 The previous simulations involved communities of $J = 100$ species differing in σ
400 but with all $\alpha_{ij} = 0$. I now study a community having weak pairwise interactions
401 between all species, $\alpha_{ij} = 0.01$. Because all species experience approximately equal
402 effects of interspecific competition, the relative differences among species in \hat{N} remain
403 approximately the same, albeit with a decrease in the maximum σ capable of persisting
404 (Figure 12). Adding symmetrical and weak species interactions had only weak effect
405 on turnover in the most abundant community member, compared to the scenario with
406 no interactions (Figure 12). Reductions in population size were nearly equivalent for
407 all species in a diverse community with weak competition. Thus the main effect of
408 adding weak species interactions in a diverse community was to reduce the maximal σ
409 capable of persisting. Similarly, variation in the magnitude of abrupt environmental
410 change had similar impact on community dynamics, as measured as σ of the most
411 dominant species, regardless of the simulated weak competition.

412 I also simulated ten strongly competing species ($\alpha_{ij} = 0.75$) and found substantial
413 differences in community dynamics. Here, competition again had little effect on how
414 the σ of the most abundant species changed with time (Figure 12). However, this
415 similarity obscured underlying community changes caused by the combination of
416 eco-evolutionary response to changing θ and competition. In the presence of strong
417 competition, species that have relatively lower abundance following environmental
418 change remained suppressed for longer periods of time and at very low densities (Figure
419 12). Note that my deterministic simulations lack stochastic extinction, which is
420 likely a major problem for populations at very low density. In my simulations, the
421 low σ species that dominated under stable environments but reached low density
422 following environmental change are on a steady upward population trend at the end of
423 simulations. Thus the dominance of higher σ species is still transient, though with a
424 much slower return to the pre-environmental change equilibrium \hat{N} . Higher σ species
425 that dominate communities can actually see increased absolute abundance following
426 environmental change, despite going from being locally-adapted to being maladapted.
427 This surprising change results from the release from competitive suppression by low σ
428 species.

429 2.4.2 Species interactions and sustained environmental change

430 I also simulated how interspecific competition impacts the eco-evolutionary community
431 response to a sustained environmental change. I used the same model of species
432 interactions as described above (equation 11) under the scenario of shifting θ at rate k
433 through time. I again began by simulating a diverse community of weakly interacting
434 species ($J = 100$ and $\alpha_{ij} = 0.01$). I found that the σ of the dominant species
435 under environmental change was largely the same regardless of whether $\alpha_{ij} = 0$ or

436 $\alpha_{i,j} = 0.01$ (Figures 6 & 7). When increasing interaction strength ($\alpha_{i,j} = 0.75$)
437 in less diverse communities ($J = 10$), I again found similar patterns comparing
438 $\alpha_{i,j} = 0.75$ to $\alpha_{i,j} = 0$ in terms of which species were most abundant through
439 time (both $J = 10$, Figure 7). However, this similarity obscured differences in relative
440 abundance patterns among communitiy members. Under interspecific competition,
441 the most abundant species had greater relative abundance advantages. Interestingly,
442 in scenarios with interspecific competition, higher gene flow species often showed
443 dramatic increases in absolute abundance following the initiation of environmental
444 change (Figure 7). In these simulations, low gene flow species with low migration
445 load were supressed by environmental change and this allowed increased abundance of
446 higher gene flow species better able to adapt to shifting environments.

447 **2.4.3 Ecosystem resilience and interspecific interactions**

448 The increased absolute abundance exhibited by many intermediate to high σ species
449 under environmental change may have important community and ecosystem-level
450 implications. For example, biodiversity can impact ecosystem function when species
451 exhibit compensatory population dynamics through time, stabilizing ecosystem-level
452 processes (Micheli et al. 1999; Loreau 2010). The increased abundance of high σ species
453 under environmental change due to competitive release might stabilize ecosystem
454 function despite declining low σ species. I tested this hypothesis using the assumption
455 that total number of individuals of all species in the community corresponded to
456 ecosystem function.

457 I quantified biomass resilience using approaches specific to each scenario of environ-
458 mental change. For abrupt change, I calculated the time (number of generations)
459 until the community regained 75% of the biomass seen at equilibrium before the
460 environmental change. For sustained change, I calculated the biomass in the final
461 generation of simulations (500 generations following the initiation of change - when
462 populations had stabilized) as a proportion of the biomass under stable environments.

463 In both cases, simulations showed that communities with stronger interspecific compe-
464 tition also showed greater resilience under strong environmental change. In diverse
465 communities with weak interspecific competition, biomass either returned faster or
466 was maintained at higher relative levels, compared to similar communities without
467 interspecific competition (Figure 8). Communities with fewer species (10 species)
468 but stronger interspecific competition exhibited even greater resilience relative to
469 comparable communities without interspecific competition, under both scenarios of
470 environmental change. This resilience is clearly due to increases in abundance of high
471 σ species, which were released from competitive supression by previously dominant but
472 slow adapting low σ species, and which themselves adapted to changing environments
473 rapidly (Figures 7 & 8).

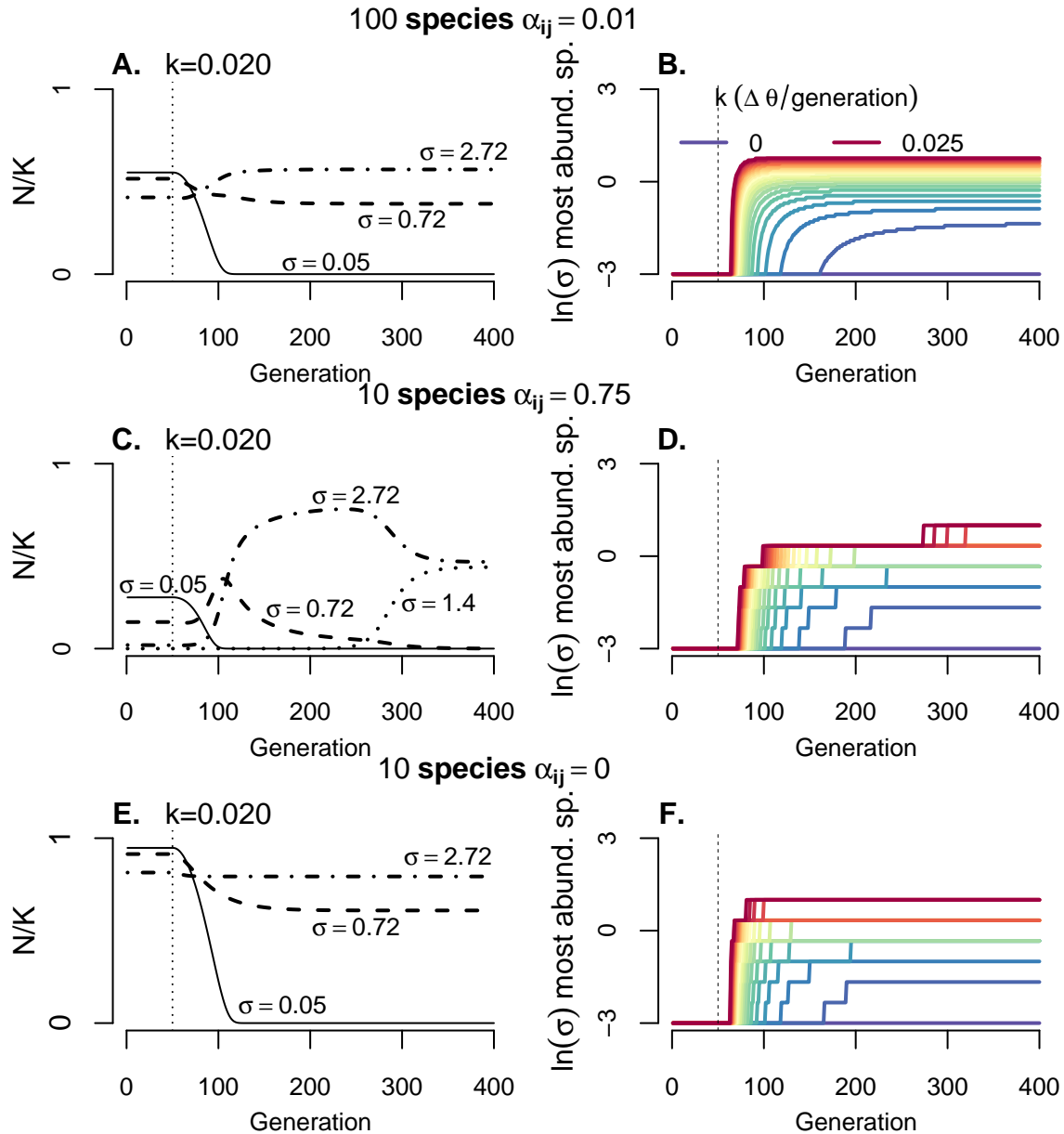


Figure 7: Effects of a sustained environmental change with interspecific competition, with example species highlighted in each scenario. Left panels demonstrate how absolute abundance of higher σ species can increase following environmental change, despite. (A-B) Diverse community with weak interspecific competition (C-D) Community with fewer species and strong interspecific competition. (E-F) A community composed of the same species as (C-D) but with no interspecific competition. Vertical dashed line indicates beginning of environmental change at generation 50 (450 generations were run under a stable environment before those shown). Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, and $r_m = 0.5$

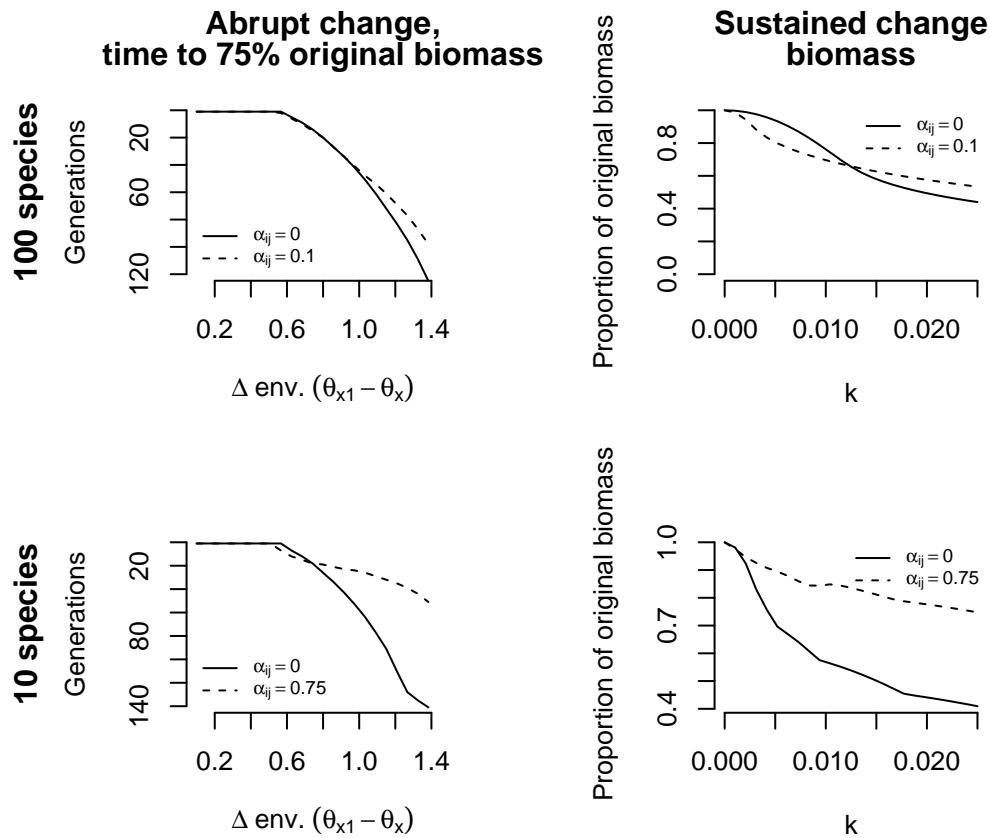


Figure 8: Communities with interspecific competition are more resilient to environmental change, measured in terms of (left panels) time to return to 0.75 of pre-environmental change biomass or (right panels) biomass in 500th generation under sustained linear temporal change. Note that in left panels the y-axis is reversed for comparability with right panels. Biomass is measured as the total number of individuals of all species. Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, and $r_m = 0.5$.

474 2.5 Relationship of eco-evolutionary community turnover to 475 ecological succession

476 The transient dominance of species with higher gene flow following an abrupt environ-
477 mental change is qualitatively similar to classic hypotheses explaining successional
478 turnover in communities. Specifically, early successional species may have better
479 dispersed propagules but lower fitness compared to later successional species. In the
480 present model, gene flow and propagule dispersal are one in the same (σ), i.e. there is
481 no mechanism of gene flow apart from propagule movement (no gamete movement). To
482 more formally investigate the similarity with succession, I studied how species differing
483 in σ in the present model respond to ecological disturbance, with no change in θ . In the
484 absence of any environmental change, consider an ecological disturbance that reduces
485 locally-adapted populations of different species by the same large proportion. For
486 simplicity, I assumed a localized disturbance that introduced non-zero $\frac{\partial^2 N}{\partial x^2}$ (equation
487 3) but did so orthogonally to b such that asymmetric migration had no effect on trait
488 evolution (i.e. $\frac{\partial \ln(N)}{\partial x}$ set equal to zero in equation 2).

489 Successional community turnover arises as species differ in the rate of population
490 growth (eqn 3) due to interspecific variation in immigration (favoring high σ species)
491 and fitness (favoring low σ species). However, note that the fitness advantage of low
492 σ species is dependent on reproduction by individuals already present, which are few
493 after disturbance. My simulations showed that the more intense the disturbance,
494 the slower the return to community equilibrium (Figure 9), analogous to the slower
495 return following greater abrupt changes in θ (Figure 3). Under a sustained ecological
496 disturbance (constant proportion of individuals lost each generation) ecological commu-
497 nity turnover exhibits qualitatively similar patterns to the eco-evolutionary response
498 to sustained change in θ (Figure 9). Specifically, sustained disturbance resulted in
499 consistent dominance by species with intermediate σ , similar to these species being
500 most abundant under sustained change in θ (Figure 6).

501 3 Discussion

502 Evolutionary genetic theory is a rich source of hypotheses for how life history impacts
503 evolution. On this rapidly changing planet, understanding and predicting evolutionary
504 responses environmental change will be particularly valuable (Bay et al. 2017; Gienapp
505 et al. 2017). Molecular data are providing a deeper view of the differences among
506 species in population genomic patterns (e.g. Romiguier et al. 2014). The present is
507 ripe for studying how interspecific trait differences impact evolutionary response to
508 environmental change and the consequences for communities and ecosystems. Here,
509 I took existing quantitative genetic models of adaptation (Lynch and Lande 1993;
510 Barton 2001; Polechová et al. 2009) and showed how interspecific trait variation

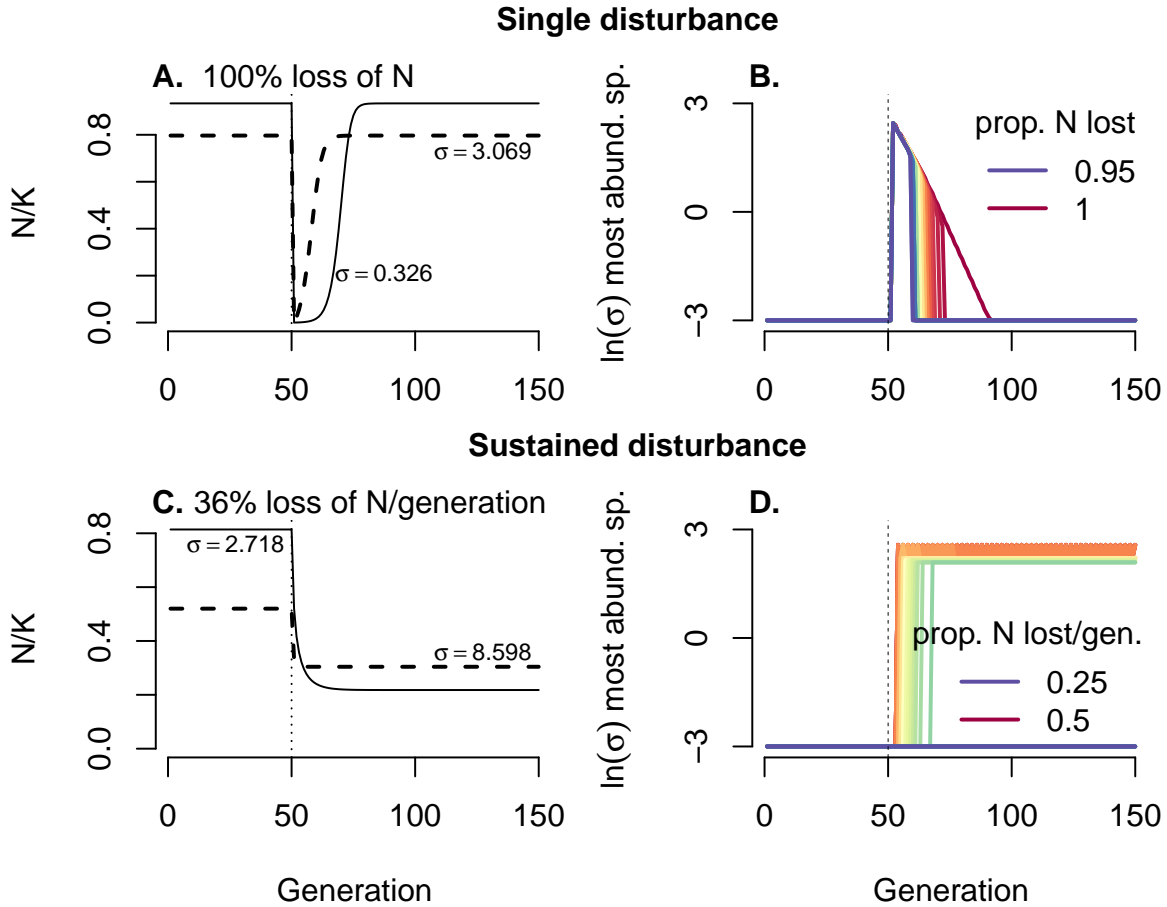


Figure 9: Variation in dispersal (σ) among species determines how communities of locally-adapted populations respond to ecological disturbance. (A-B) A single disturbance removes a large portion of each species' N after generation 50. (C-D) recurring disturbances are imposed in each generation, starting after generation 50. Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, $r_m = 0.5$, and $\frac{\partial^2 N}{\partial x^2} = 5$.

511 gives rise to differences in genetic diversity with non-monotonic effects on community
512 structure and dynamics. Many previous studies of what is referred to as evolutionary
513 rescue have largely focused on thresholds beyond which populations go extinct under
514 environmental change (Lynch and Lande 1993; Gomulkiewicz and Holt 1995; Bell and
515 Gonzalez 2009; Uecker et al. 2014). Even if populations of most species in a community
516 are able to avoid extinction under environmental change, my results highlight how
517 communities may change drastically in composition and function. Dominant species
518 can become rare and rare species can become dominant (Figure 7). This turnover has
519 important consequences for community diversity and ecosystem function.

520 In general, eco-evolutionary community inversions (i.e. reversals in relative abundances)
521 may arise in any system where there is a negative or complex relationship between
522 census population size and adaptability to environmental change. In my model,
523 these changes are driven by the fact that initially numerically abundant species are
524 more maladapted for longer periods of time following environmental change. Genetic
525 variance has a major influence on the rate of adaptation, but other traits, such as
526 generation time, vary among species in communities and may also result in eco-
527 evolutionary community turnover. For example, parasites may have shorter generation
528 time than hosts, allowing parasites to adapt faster to abiotic environmental change.
529 Both vertebrate hosts (Fraser 2013) and their parasites (Sternberg and Thomas 2014)
530 can be locally-adapted along temperature gradients, though parasites might adapt
531 to climate change faster than hosts. Alternatively, when census population size is
532 positively related to genetic variance in a trait under selection (Frankham 1996),
533 evolutionary responses to environmental change may reinforce the ecological responses,
534 reducing community diversity.

535 I identified a transient benefit to high gene flow following an abrupt environmental
536 change, due to faster adaptation. In their experimental microcosm study, Low-Décarie
537 et al. (2015) demonstrated how gene flow was key to the eco-evolutionary recovery
538 of soil microbial communities responding to a novel herbicide. Studies of genetic
539 variation (Lande and Shannon 1996) from dispersal (Polechová et al. 2009; Blanquart
540 and Gandon 2011) or mutation (Taddei et al. 1997) have yielded similar results. When
541 environment is constant, low mutation rates are favored, though mutator lineages
542 have transient benefits when they find adaptive mutations (Taddei et al. 1997).
543 Additionally, fluctuating environments can favor higher mutation rates (Travis and
544 Travis 2002). Indeed, co-occurring species can exhibit a range of mutation rates (Baer
545 et al. 2007), which may also play a role in species differences in the degree of local
546 adaptation and subsequent responses to environmental change (Orr and Unckless
547 2008). Here, I did not allow explicit evolution of dispersal distance (σ), though
548 the comparison of population sizes for my species of differing σ provide insight into
549 how dispersal would evolve in this system. In a temporally constant environment
550 (prior to generation 50 in simulations), dispersal is maladaptive due to the spatial
551 selective gradient (Balkau and Feldman 1973). However, once temporal change

552 in environment is introduced, greater dispersal can be favored with the functional
553 form of temporal environmental change determining the optimal σ (see Blanquart
554 and Gandon 2011 for more detailed analysis). I did not investigate interspecific
555 variation in phenotypic plasticity, which may supplant local adaptation as a response
556 to environmental gradients. As with migration load, if census population size is related
557 to the degree of local adaptation versus plasticity (i.e. habitat specialization versus
558 generalization) then changing environments may cause complex community change.
559 Under some models, greater dispersal across environmental gradients can favor plastic
560 responses to environment (Sultan and Spencer 2002; reviewed by Hendry 2016).

561 The form of environmental change may have dramatic effects on how eco-evolutionary
562 responses influence communities. Previous theory has shown how the benefits of genetic
563 variation (Lande and Shannon 1996) and dispersal (Blanquart and Gandon 2011) can
564 depend on the functional form of environmental change. I found that communities can
565 exhibit distinct dynamics depending on a scenario of abrupt environmental change
566 (Gomulkiewicz and Holt 1995; Orr and Unckless 2008) versus sustained change (Pease
567 et al. 1989; Lynch and Lande 1993; Polechová et al. 2009). Specifically, sustained
568 change favors intermediate gene flow species and results in their stable dominance
569 (highest N) in communities, whereas abrupt environmental change results in only
570 transient community change favoring high to intermediate σ species. In nature any
571 form is possible and thus my results demonstrate how diverse forms of environmental
572 change may cause complex dynamics in nature.

573 Though I modeled community turnover in a single local population, all communities
574 in my model are equivalent and the processes I described would occur across species
575 ranges. This suggests that there is a large potential spatial extent of eco-evolutionary
576 responses to rapid environmental change, resulting in community change across large
577 regions. In nature b is non-linear and rugged, a feature worthy of study in future
578 simulation of response to temporal environmental change. Furthermore, multiple traits
579 may be under simultaneous spatially-varying selection (Guillaume 2011; Duputié et
580 al. 2012; MacPherson et al. 2015) and selective regimes on these traits may change
581 simultaneously. Given that environmental change can be complex, with different forms
582 of change in different environmental dimensions, it is possible that in nature changes
583 in selective gradients may take multiple functional forms simultaneously leading to
584 complex changes in relative abundance for species differing in σ .

585 The model studied here was simple and thus it is challenging to determine how
586 important my results are in natural systems. However, gene flow across spatial
587 selective gradients is likely a major source of within-population genetic variation in
588 traits under selection (Yeaman and Jarvis 2006; Paul et al. 2011; Farkas et al. 2013).
589 Findings on ponderosa pine suggest that greater b can cause greater V_G (Yeaman
590 and Jarvis 2006). Less is known, however, of how adaptability or V_G are related to
591 interspecific variation in census population size (abundance). The negative relationship
592 between these two quantities is the key to community turnover following environmental

593 change in my results. One problem with empirically studying the processes I described
594 there is often a substantial lag before better dispersing species dominate communities
595 (Figures 3 & 6). Thus researchers may overlook empirical population changes caused
596 by environmental change.

597 It may be a common feature of strongly interacting species that they experience
598 selective gradients driven by the same environmental variable (e.g. temperature).
599 Differences among these species in local adaptation to the same environmental variable
600 might lead to different eco-evolutionary responses to environmental change, causing
601 indirect effects on interacting species (Fussmann and Gonzalez 2013). For exam-
602 ple, multiple competing tree species may simultaneously be locally-adapted along
603 environmental gradients (Ikeda et al. 2014). Recent work by Brans et al. (2017)
604 has shown similar intraspecific trait clines in multiple co-occurring cladocerans along
605 urbanization gradients drives community patterns. Here I simulated competing species,
606 but interactions of different types (e.g. trophic) may yield additional eco-evolutionary
607 community responses to changing environments.

608 My work demonstrates how interspecific variation in gene flow alters communities
609 experiencing environmental change. Some authors have suggested assisted gene flow
610 as a technique to mitigate climate change impacts on wild populations, with gene
611 flow facilitating local adaptation of populations suddenly experiencing novel climates
612 (Aitken and Whitlock 2013). My results highlight how such approaches could have
613 important effects on community structure. Aitken and Whitlock (2013) suggested
614 that assisted gene flow efforts should be focused on ecologically dominant species (due
615 to importance for ecosystem functioning) and rare species (to prevent extinction).
616 My results show how such a strategy would likely change community structure, as
617 species not included (historically intermediate abundance species) in assisted gene
618 flow would be expected to decline in abundance due to slower adaptation to climate
619 change. Others have suggested breeding of wild species to promote adaptation to
620 future environments (Oppen et al. 2015). These management efforts would have to
621 be balanced across species of different abundances if they are to limit impacts on
622 community composition.

623 **3.1 Conclusion**

624 Community composition is defined by the population sizes of component species, but
625 greater population size might not correspond to greater adaptability to environmental
626 change. This discrepancy can result in complex community turnover as selection
627 regimes shift. The simple model studied here demonstrates some of the complexity
628 in eco-evolutionary community change. Future research could improve our ability
629 to predict responses to environmental change in nature by learning more about the
630 genetics and ecology of adaptation in addition to theoretical investigation of more
631 complex scenarios.

632 **3.2 Acknowledgments**

633 This manuscript benefited from comments by Hidetoshi Inamine, Martin Turcotte,
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635 Read, Katriona Shea, and Timothy Reluga.

636 4 Appendix

637 4.1 Impacts on community-mean traits

638 Interspecific variation in adaptation following environmental change will likely have
639 impacts on the distribution of traits in a community, which is often of interest to
640 community and ecosystem ecologists (Muscarella and Uriarte 2016; Šímová et al. 2018).
641 For example, ecosystem function may be influenced by the mass-averaged functional
642 traits in a community (Grime 1998). Under the scenario of abrupt environmental
643 change, the slow adaptation and return to equilibrium abundance of species that
644 dominate communities may have interesting effects on changes in community-weighted
645 mean (CWM) traits. Indeed, following abrupt environmental change, initially there is
646 a very rapid phase of change in CWM driven by fast-adapting high σ species (Figure
647 10). However, there is an abrupt slow-down in change in CWM as most high σ species
648 have adapted but low σ species remain maladapted. Nevertheless, the low migration
649 load of these low σ species contributes to their fitness and abundance and hence
650 influence over CWM traits. By contrast, when there is sustained change in θ over
651 time, species exhibit marked variation in their ability to adapt to the moving optimum.
652 Although the highest σ species are able to maintain \bar{z} close to the optimum, they are
653 less abundant than intermediate σ species due to migration load (equation 10, Figure
654 6). Thus the CWM exhibits a substantial and stable lag behind the optimum.

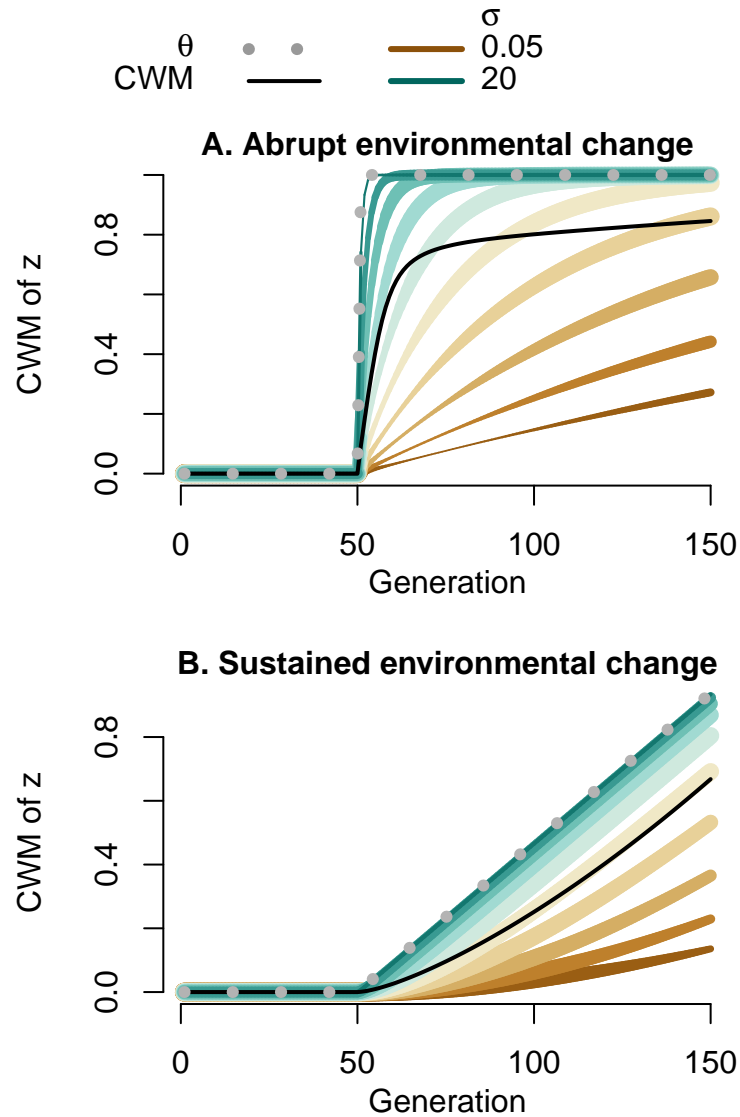


Figure 10: Effects of environmental change on community-weighted mean (CWM) traits under selection due to eco-evolutionary responses. Example species with a range of σ values are shown (colors), with line thickness indicating relative abundance. As in earlier presented simulations, communities were composed of species with a log uniform distribution of σ values. The CWM (black line) at each timepoint is an abundance-weighted average of z . Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, and $r_m = 0.5$. For (A), $\theta_{x1} - \theta_x = 1$. For (B), $k = 0.009$.

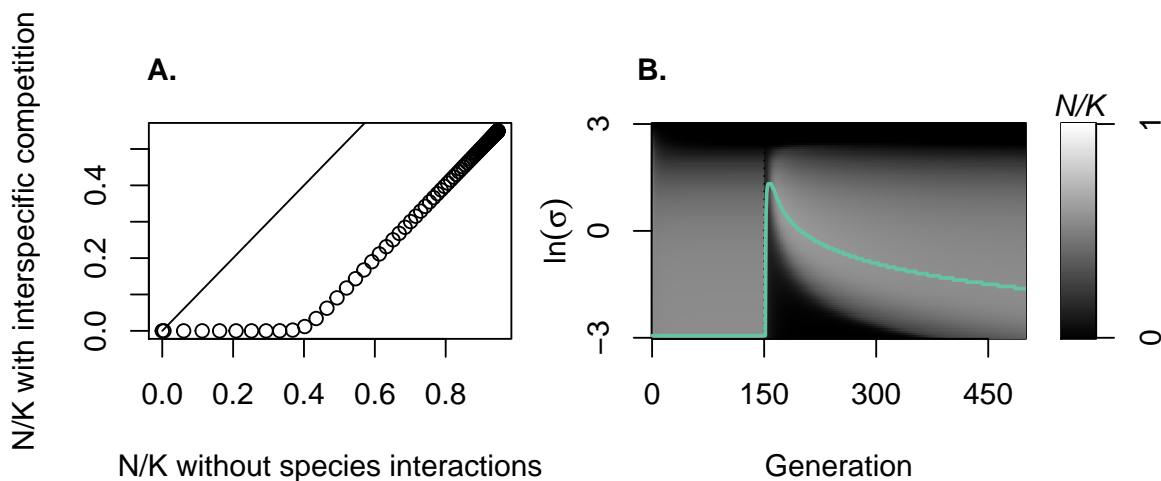


Figure 11: Community turnover under abrupt environmental change, with interspecific competition. (A) Comparing approximate equilibrium species' N for scenarios differing in presence of interspecific competition. Black circles show species in a diverse (100 species) community with weak interactions ($\alpha_{ij} = 0.01$). Straight line shows one-to-one relationship. (B) Here the trend in abundance for a diverse (100 species) community is shown, where species weakly compete ($\alpha_{ij} = 0.01$) according to Lotka-Volterra models. Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, $r_m = 0.5$, and $\theta_{x1} - \theta_x = 1$.

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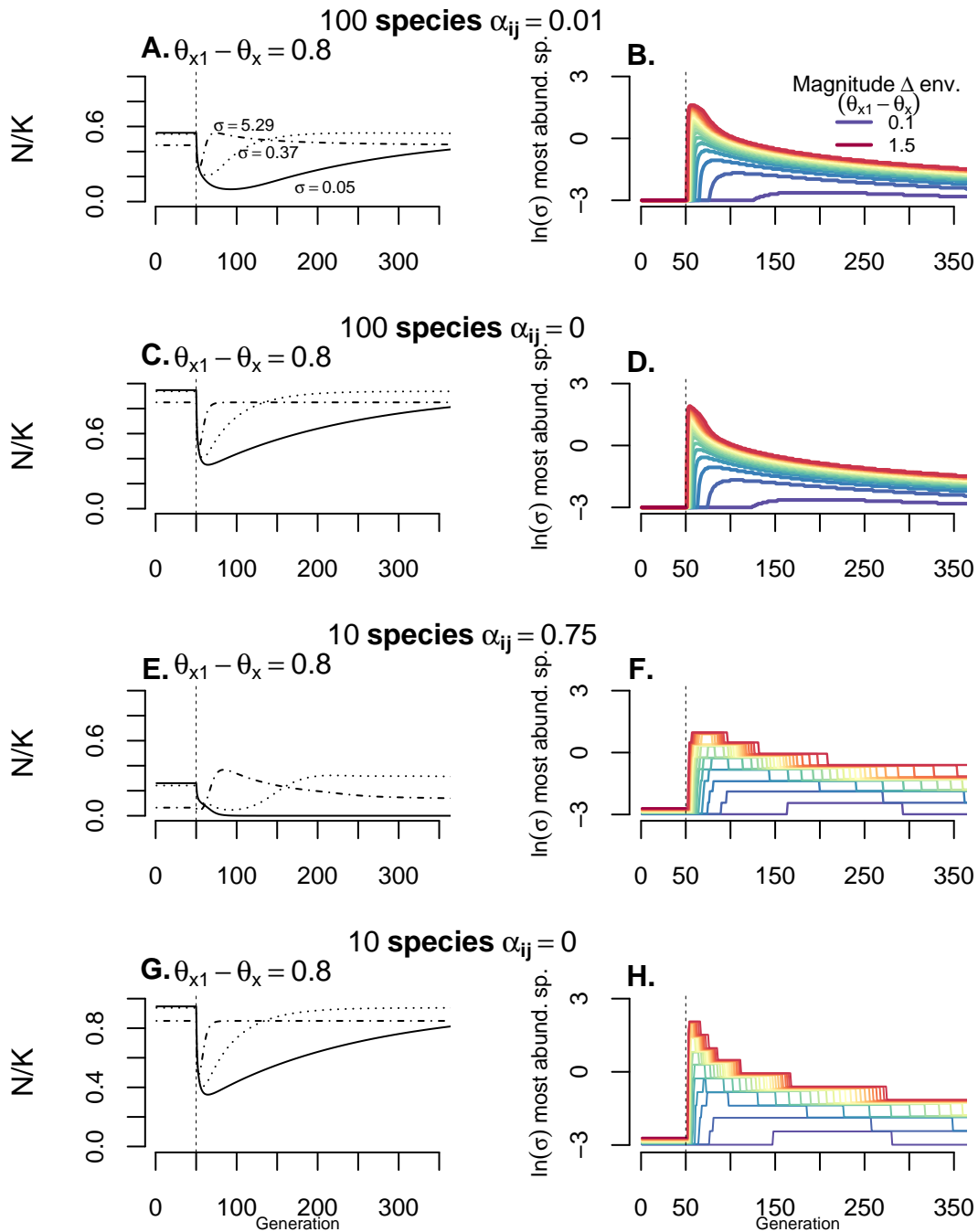


Figure 12: How the magnitude of environmental shift and interspecific competition affects community turnover. Left panels show the same three representative species of varying σ under different scenarios of interspecific competition. Right panels show which species are most abundant at any point in time, under different levels of abrupt environmental change. Populations are at approximate equilibrium and adapted to θ_x for the first 50 generations, when an instantaneous environmental change to θ_{x1} occurs. Parameter values (unless otherwise noted) were $b = 0.05$, $V_S = 1$, $V_E = 0.05$, and $r_m = 0.5$.

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