

1 Membrane proteins with high N-glycosylation, high expression, and multiple
2 interaction partners were preferred by mammalian viruses as receptors

3

4 Zheng Zhang^{1, #}, Zhaozhong Zhu^{1, #}, Wenjun Chen¹, Zena Cai¹, Beibei Xu², Zhiying
5 Tan², Aiping Wu^{3, 4}, Xingyi Ge¹, Xinhong Guo¹, Zhongyang Tan¹, Zanzian Xia⁵,
6 Haizhen Zhu^{1, 6, *}, Taijiao Jiang^{3, 4, *}, Yousong Peng^{1, *}

7

8 ¹ College of Biology, Hunan University, Changsha, China

9 ² College of Computer Science and Electronic Engineering, Hunan University,
10 Changsha, China

11 ³ Center of System Medicine, Institute of Basic Medical Sciences, Chinese Academy
12 of Medical Sciences & Peking Union Medical College, Beijing, China

13 ⁴ Suzhou Institute of Systems Medicine, Suzhou, China

14 ⁵ School of Life Sciences, Central South University, Changsha, China

15 ⁶ State Key Laboratory of Chemo/Biosensing and Chemometrics, Hunan University,
16 Changsha, China

17 # These authors contributed equally to this work

18 * Correspondence: zhuhaizhen69@yahoo.com (HZ), taijiao@ibms.pumc.edu.cn (TJ),

19 pys2013@hnu.edu.cn (YP)

20 **Abstract**

21 Receptor mediated entry is the first step for viral infection. However, the relationship
 22 between viruses and receptors is still obscure. Here, by manually curating a
 23 high-quality database of 268 pairs of mammalian virus-host receptor interaction,
 24 which included 128 unique viral species or sub-species and 119 virus receptors, we
 25 found the viral receptors were structurally and functionally diverse, yet they had
 26 several common features when compared to other cell membrane proteins: more
 27 protein domains, higher level of N-glycosylation, higher ratio of self-interaction and
 28 more interaction partners, and higher expression in most tissues of the host.
 29 Additionally, the receptors used by the same virus tended to co-evolve. Further
 30 correlation analysis between viral receptors and the tissue and host specificity of the
 31 virus shows that the virus receptor similarity was a significant predictor for
 32 mammalian virus cross-species. This work could deepen our understanding towards
 33 the viral receptor selection and help evaluate the risk of viral zoonotic diseases.

34 **Introduction**

35 In the new century, much progress has been made in prevention and control of
 36 infectious diseases, but the recent serial outbreaks of Zika virus ^[1], Ebola virus
 37 (EBOV) ^[2] and Middle East Respiratory Syndrome Coronavirus (MERS-CoV) ^[3]
 38 indicate that the viral infectious diseases still pose a serious threat to human health
 39 and global security. The virus is the most abundant biological entity on Earth and
 40 exists in all habitats of the world ^[4]. Nearly all cellular organisms are prey to viral

41 attack. Humans were reported to be infected by hundreds of viruses ^[5, 6]. Most of the
 42 human emerging infectious diseases are zoonotic, with viruses that originate in
 43 mammals of particular concern ^[7], such as the Human Immunodeficiency Virus (HIV)
 44 ^[8] and Severe Acute Respiratory Syndrome Coronavirus (SARS-CoV) ^[9]. Mammals
 45 are not only the most closely related animal to humans in phylogeny, but also contact
 46 with humans most frequently ^[7], especially for the livestock and pet. For effective
 47 control of human viral diseases, much attention should be paid to the mammalian
 48 virus.

49 Receptor-binding is the first step for viral infection of host cells ^[10-13]. Multiple types
 50 of molecules could be used as viral receptors ^[12, 14], including protein ^[15-17],
 51 carbohydrate ^[18, 19] and lipid ^[20]. How to select receptors by the virus is an important
 52 unsolved question ^[13, 14, 16, 21]. Specificity and affinity are two most important factors
 53 for viral receptor selection ^[14]. Carbohydrates and lipids are widely distributed on host
 54 cell surfaces and easy targets for viruses to grab ^[10, 11]. Compared to these molecules,
 55 proteins were reported to be more suitable receptors because of stronger affinity and
 56 higher specificity for viral attachment, which could increase the efficiency of viral
 57 entry and facilitate viruses to expand their host ranges and alter their tropisms ^{[10-12, 14,}
 58 ^{15]}. Previous studies have shown that proteins that were abundant in the host cell
 59 surface or had relatively low affinity for their natural ligands, were preferred by
 60 viruses as receptors, such as proteins involved in cell adhesion and recognition by
 61 reversible, multivalent avidity-determined interactions ^[10, 15]. This suggests that the
 62 selection of proteins by viruses as receptors should not be a random process. A

63 systematic analysis of the characteristics of the viral receptor could help understand
64 the mechanisms under the receptor selection by viruses.

65 The virus-receptor interaction was reported to be a principal determinant of viral host
66 range, tissue tropism and cross-species infection ^[11, 16, 22]. The existence and
67 expression of the virus receptor in a host (or tissue) should be a prerequisite for viral
68 infection of the host (or tissue) ^[21]. Usually, a virus mainly infects some particular
69 type of hosts or tissues. For example, the influenza virus mostly infects cells of the
70 respiratory tract ^[23]. However, the virus-receptor interaction is a highly dynamic
71 process. Some viruses can recognize one or more receptors ^[13, 14, 24], which can also
72 differ among virus variants or during the course of infections ^[14, 25, 26]. In some cases,
73 a few amino acid mutations in the viral protein or the receptor could abolish or
74 enhance viral infection ^[27-29]. Besides, the virus-receptor interaction is under
75 continuous evolutionary pressure to increase the viral infection efficiency, which may
76 result in the emergence of virus variants with altered host or tissue tropism. For
77 example, the SARS-CoV and MERS-CoV, which belong to the same genus,
78 *betacoronavirus*, have evolved to use different receptors (angiotensin I converting
79 enzyme 2 (ACE2) and dipeptidyl peptidase 4 (DPP4) respectively) and also infect
80 different hosts ^[11, 16, 28]. Despite of numerous studies about the tissue and host
81 specificity of the virus and the viral receptor, the systematical correlation
82 characteristics between them are still obscure.

83 Here, by manually curating a high-quality database of 268 pairs of mammalian
84 virus-receptor interaction, which included 128 unique viral species or sub-species and

119 virus receptors, we systematically analyzed the structural, functional,
 evolutionary and tissue-specific expression characteristics of mammalian virus
 receptors, which could not only deepen our understanding towards the mechanism
 behind the viral receptor selection, but also help to predict and identify viral receptors.
 Besides, we also investigated the associations between the tissue and host specificity
 of the virus and the viral receptor, and further evaluated the risk of viral cross-species
 based on viral receptors. It would help for early warning and prediction of viral
 zoonotic diseases.

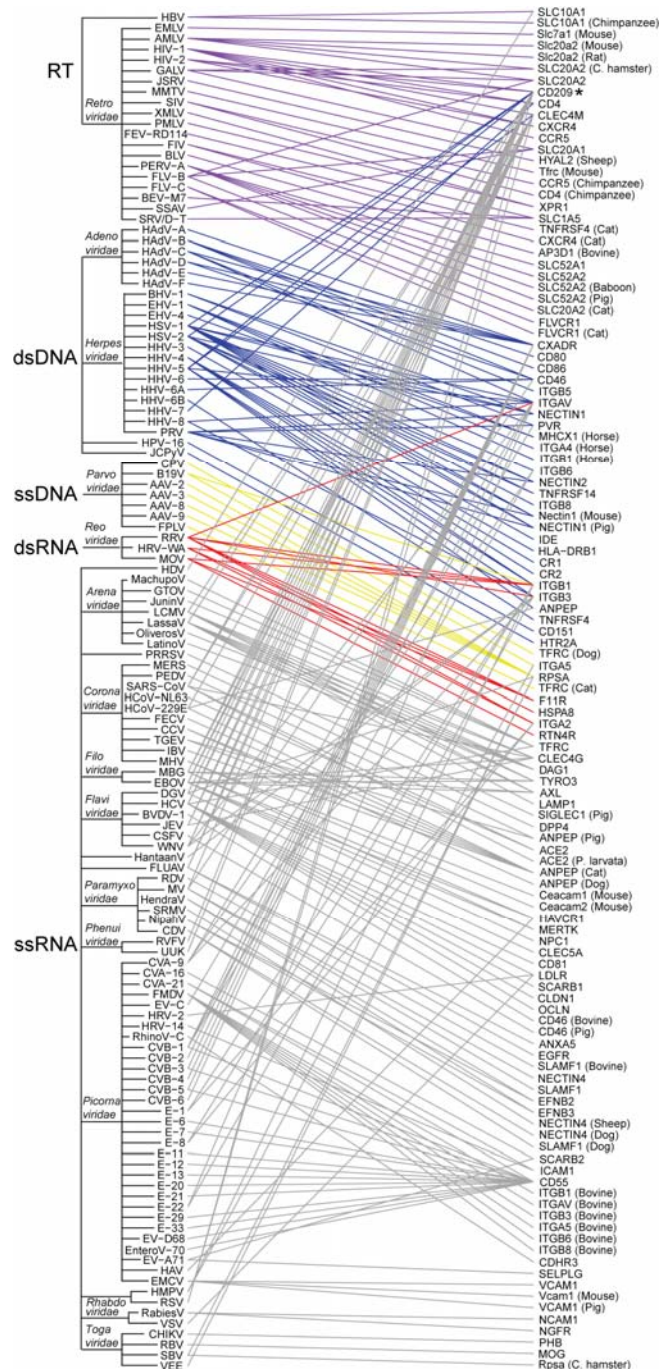
Results

Database of mammalian virus-host receptor interaction

To understand how the virus selects receptors, we manually curated a high-quality
 database of 268 pairs of mammalian virus-host receptor interactions (Figure 1 and
 Table S1), which included 128 unique viral species or sub-species from 21 viral
 families and 119 virus receptors from 13 mammal species. The viral receptor
 collected here belonged to 13 mammal species (Figure S1A), among which the human
 accounted for the most (74/119). The viruses included in the database covered all
 groups of viruses in the Baltimore classification (Figure 1). Among them, the
 single-stranded RNA (ssRNA) virus accounted for over half of all viruses (76/128),
 while the double-stranded RNA (dsRNA) virus accounted for the least (3/128). On the
 level of family, the family of *Picornaviridae* of ssRNA virus, *Retroviridae* of

106 Retro-transcribing viruses (RT) and *Herpesviridae* of double-stranded DNA (dsDNA)

107 viruses were the most abundant ones in the database (Figure 1 and Table S1).



109 **Figure 1.** The mammalian viruses and their related receptors in our database. The

110 lines between the virus and their related receptors were colored according to the group

111 of the virus in the Baltimore classification. The names of some viral families were
112 presented in italic. Viral names were displayed in abbreviation (see Table S1 for the
113 full name). The host names were given for the receptor of non-human mammal
114 species. The receptor CD209 was marked with an asterisk. For more details about the
115 mammalian virus and their receptors, please see the website
116 <http://www.computationalbiology.cn:5000/viralReceptor>.

117

118 **Association between mammalian viruses and their receptors**

119 Analysis of the association between the virus and their receptors showed that 60% of
120 viruses (77/128) used only one receptor (Figure 1 and Figure S1B), while the
121 remaining viruses used two or more receptors. Surprisingly, some viruses, such as the
122 Human alphaherpesvirus 1 (HSV-1) and Hepacivirus C (HCV), used more than five
123 receptors. We next analyzed the receptor usage on the level of viral family. For fifteen
124 viral families including two or more viruses in the database, all of them used two or
125 more sets of receptors, suggesting that different viruses of the same family tend to use
126 different receptors. For example, in the family of *Togaviridae*, the Chikungunya virus
127 (CHIKV), the Rubella virus (RBV) and the Sindbis virus (SBV) used the receptor of
128 prohibitin (PHB), myelin oligodendrocyte glycoprotein (MOG) and ribosomal protein
129 SA (RPSA), respectively. On the other hand, some viruses of different families or
130 even different groups used the same receptor (Figure 1). For example, HIV-2 and
131 EBOV, from the family of *Retroviridae* (RT group) and *Filoviridae* (ssRNA group)

132 respectively, both took CD209 molecule (CD209) (marked with an asterisk in Figure
133 1) as the receptor. On average, each receptor was used by more than two viruses.
134 More specifically, among 119 virus receptors, forty-four of them were used by more
135 than one virus (Figure 1 and Figure S1C); twenty-one of them were used by viruses of
136 more than one family and fifteen of them were used by viruses of more than one
137 group (Figure 1).

138 **Structural, functional, evolutionary and tissue-specific expression characteristics** 139 **of mammalian virus receptors**

140 To understand how the virus selects receptors, we systematically analyzed the
141 structural, functional, evolutionary and tissue-specific expression characteristics of the
142 mammalian virus receptor.

143 ***1) The mammalian virus receptor were structurally diverse***

144 We firstly investigated the structural characteristics of mammalian virus receptor
145 proteins. As expected, all the mammalian virus receptor protein belonged to the
146 membrane protein which had at least one transmembrane alpha helix (Figure S2A).
147 Twenty-four of them had more than five helices, such as 5-hydroxytryptamine
148 receptor 2A (HTR2A) and NPC intracellular cholesterol transporter 1 (NPC1). The
149 receptor protein was mainly located in the cell membrane. Besides, more than one
150 third (43/119) of them were also located in the cytoplasm, and thirteen of them were
151 located in the nucleus.

152 Then, the protein domain composition of the mammalian virus receptor protein was

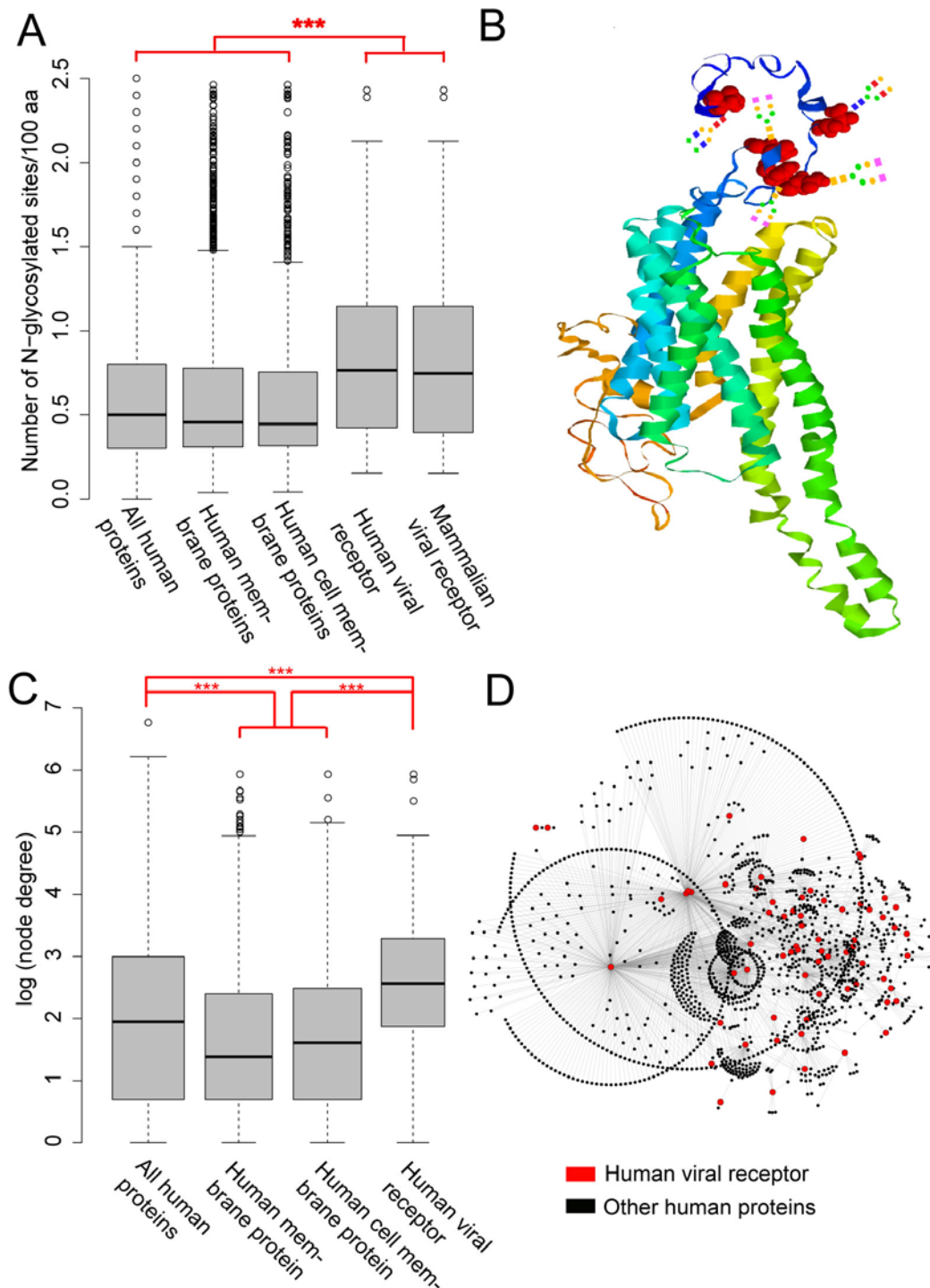
153 analyzed. The mammalian virus receptor proteins contained a total of 336 domains
154 based on the Pfam database, with each viral receptor protein containing more than two
155 domains on average (Figure S2B). This was significantly more than that of human
156 proteins or human membrane proteins (p-values < 0.001 in the Wilcoxon rank-sum
157 test). Some viral receptor proteins may contain more than 10 domains, such as
158 complement C3d receptor 2 (CR2) and low density lipoprotein receptor (LDLR). The
159 protein domains of the mammalian virus receptor protein could be grouped into 77
160 families in the Pfam database, suggesting the structure diversity of the mammalian
161 virus receptor protein. The most commonly observed Pfam families were
162 Immunoglobulin V-set domain, Immunoglobulin C2-set domain, Integrin beta chain
163 VWA domain, Integrin plexin domain, and so on (Figure S2C).

164 ***2) The mammalian virus receptor had high level of N-glycosylation***

165 Glycosylation of protein is widespread in the eukaryote cell. We next characterized
166 the glycosylation level of the mammalian virus receptor. N-glycosylation is the most
167 common type of glycosylation. We found that 93 of 119 mammalian virus receptors
168 were N-glycosylated with an average of 0.94 glycosylation sites per 100 amino acids
169 (Figure 2A). It increased to 0.97 glycosylation sites per 100 amino acids for the
170 human viral receptor (Figure 2A), among which 62 were N-glycosylated. Twelve
171 human viral receptors were observed to have ten or more N-glycosylation sites, such
172 as complement C3b/C4b receptor 1 (CR1) and lysosomal associated membrane
173 protein 1 (LAMP1). Figure 2B displayed the modeled 3D-structure of HTR2A, the
174 receptor for JC polyomavirus (JCPyV). Five N-glycosylation sites were highlighted in

175 red on the structure, which were reported to be important for viral infection ^[30]. For
176 comparison, we also characterized the N-glycosylation level for the human cell
177 membrane protein, human membrane proteins and all human proteins (Figure 2A). It
178 was found they had a significantly lower level of N-glycosylation than that of human
179 and mammalian virus receptors (p-values < 0.001 in the Wilcoxon rank-sum test),
180 which suggests the importance of N-glycosylation for the viral receptor.

181 O-glycosylation is also a common type of glycosylation. We found there was only a
182 small fraction of mammalian virus receptors (14/119) with O-glycosylation. Besides,
183 no significant difference was observed between the O-glycosylation level of
184 mammalian virus receptor proteins and that of human proteins (Figure S2D).



185

186 **Figure 2.** Analysis of N-glycosylation and protein-protein interactions of mammalian

187 virus receptors. (A) Comparison of the N-glycosylation level between mammalian

188 viral receptors, human viral receptors, human cell membrane proteins, human

189 membrane proteins and all human proteins. For clarity, the outliers greater than 2.5
190 were removed. “***”, p-value < 0.001. (B) The modeled 3D-structure of HTR2A.
191 Five N-glycosylation sites were highlighted in red. Artificial glycans were manually
192 added onto the site. (C) Comparison of the degree of proteins between human viral
193 receptors, human cell membrane proteins, human membrane proteins and all human
194 proteins in the human PPIN. For clarity, the node degree was logarithmically
195 transformed. “***”, p-value < 0.001. (D) Partial human PPI network composing of
196 the PPIs which involved at least one viral receptor protein (colored in red).

197

198 **3) Functional enrichment analysis of the human virus receptor**

199 We next attempted to identify the gene functions and pathways enriched in the
200 mammalian virus receptor. As was mentioned above, 74 of 119 mammalian virus
201 receptors belonged to the human. Besides, analysis showed that 36 of the remaining
202 non-human mammalian virus receptors were homologs of the human virus receptor
203 (Table S2). Therefore, we conducted the function enrichment analysis only for the
204 human virus receptor based on the databases of Gene Ontology (GO) and KEGG. For
205 the GO Cellular Component (Table S3), the human virus receptor was mainly
206 enriched in the membranes and junctions, the latter of which included the adherens
207 junction, cell-substrate junction, focal adhesion, and so on. For the GO Biological
208 Process (Table S3), the human virus receptor was mainly enriched in the process of
209 entry into the host. Besides, some terms related to the immune response were also

210 enriched, such as “Regulation of leukocyte activation” and “Lymphocyte activation”.

211 For the GO Molecular Function (Table S3), besides for the enrichment of terms

212 related to the virus receptor activity, the human virus receptor was also enriched in

213 terms of binding to integrin, glycoprotein, cytokine, and so on.

214 Consistent with the enrichment analysis of GO Cellular Component, the KEGG

215 pathways of “Cell adhesion molecules”, “Focal adhesion” and “ECM-receptor

216 interaction” were also enriched. Besides, the pathway of “Phagosome” was enriched

217 (Table S3), which may be associated with viral entry into the host cell. Interestingly,

218 some pathways associated with heart diseases were enriched, including “Dilated

219 cardiomyopathy”, “Hypertrophic cardiomyopathy”, “Arrhythmogenic right

220 ventricular cardiomyopathy” and “Viral myocarditis”.

221 ***4) Human virus receptors had more interaction partners than other proteins***

222 We next analyzed the protein-protein interactions (PPIs) which the mammalian virus

223 receptor protein took part in. As the reason mentioned above, we only used the human

224 virus receptor for PPI analysis. A human PPI network (PPIN) was constructed based

225 on the work of Menche et al ^[31]. It included a total of 13,460 human proteins that are

226 interconnected by 141,296 interactions. The degree and betweenness of each protein

227 in the PPIN were calculated, which could measure the importance of a protein in the

228 PPIN. It was found that the degrees for human membrane proteins and cell membrane

229 proteins were significantly smaller than those of other human proteins (Figure 2C &

230 Figure S3A, p-value < 0.001 in Wilcox rank-sum test) in the PPIN. Similar

231 observations could be found for the node betweenness in the PPIN (Figure S3B&C).

232 However, the human virus receptor protein, a subset of the human cell membrane

233 protein, was found to have significantly larger degrees and higher betweenness than

234 other human proteins in the PPIN (Figure 2C and Figure S3, p-value < 0.001 in

235 Wilcoxon rank-sum test). The median degrees for human virus receptors was 13 (Figure

236 2C), which was nearly twice as much as that of all human proteins. Six viral receptors

237 were observed to have degrees larger than 100 (Figure 2D), including epidermal

238 growth factor receptor (EGFR), heat shock protein family A member 8 (HSPA8), PHB,

239 RPSA, CD4 molecule (CD4) and integrin subunit beta 1 (ITGB1). Since the viral

240 receptor (colored in red in Figure 2D) interacted with lots of other human proteins

241 (colored in black in Figure 2D) in PPIN, we further investigated the functional

242 enrichment of these proteins by GO enrichment analysis. Interestingly, six of top ten

243 enriched terms in the domain of Biological Process were related to protein targeting

244 or localization (Table S3).

245 When looking at the interactions between viral receptors, we found that 38 of 74 viral

246 receptors interacted with themselves. This ratio ($38/74 = 51\%$) was much higher than

247 that of human proteins (22%), membrane proteins (11%) and human cell membrane

248 proteins (14%). However, we found the viral receptor tended not to interact with each

249 other (Figure S3D). Among 74 human virus receptor proteins, 36 of them had no

250 interactions with any other human virus receptor. There were only 50 PPIs between

251 different human virus receptor proteins, with each viral receptor protein interacting

252 with an average of only one other viral receptor protein.

253 **5) *The mammalian virus receptor was not more conserved than other genes***

254 Large degree of the human viral receptor in the human PPIN suggests the importance
255 of them in cellular activity. Analysis showed that 11 human viral receptors belonged
256 to the housekeeping gene. This ratio ($0.15 = 11/74$) was a little lower than that of
257 housekeeping genes in all human genes ($0.19 = 3804/20243$), suggesting that the
258 human viral receptor was not enriched in the housekeeping gene.

259 Then, we investigated the evolutionary conservation of mammalian virus receptors in
260 108 mammal species which were richly annotated in the NCBI Reference Sequences
261 (RefSeq) database (see Methods). Over half of mammalian virus receptors had
262 homologs (see Methods) in more than 100 mammal species (Figure S4A). We further
263 calculated the pairwise sequence identities between the viral receptor and their
264 homologs in mammal species. For nearly half of viral receptors, the average of
265 pairwise sequence identities was higher than 0.8 (Figure S4B). For comparison, we
266 also analyzed the evolutionary conservation of human proteins by randomly selecting
267 1000 human proteins from the NCBI RefSeq database. They were observed to have
268 similar conservation level with that of human viral receptors (Figure S4C&D and
269 Table S4).

270 **6) *Viral receptors expressed higher than other proteins in 32 major human tissues***

271 Since the virus has to compete with other proteins for binding to the receptor, proteins
272 with high expression should be preferred by viruses as receptors. Thus we measured
273 the average expression level of human viral receptors in 32 major human tissues

(Figure 3). For comparison, those of human membrane genes, human cell membrane genes and all human genes were also displayed in Figure 3. As was shown clearly, the expression level of human cell membrane genes (in cyan) was similar to that of all human genes (in black) in these tissues. Both of them were lower than that of human membrane genes (in blue). However, the human viral receptor (in red), which was part of the human cell membrane gene, expressed much higher than other sets of genes in nearly all these tissues. On average, they had an expression level of 24 transcripts per million (TPM) in these tissues, while this was 8, 4 and 4 TPM for the human membrane gene, the human cell membrane gene and all human genes (see the black arrow in Figure 3).

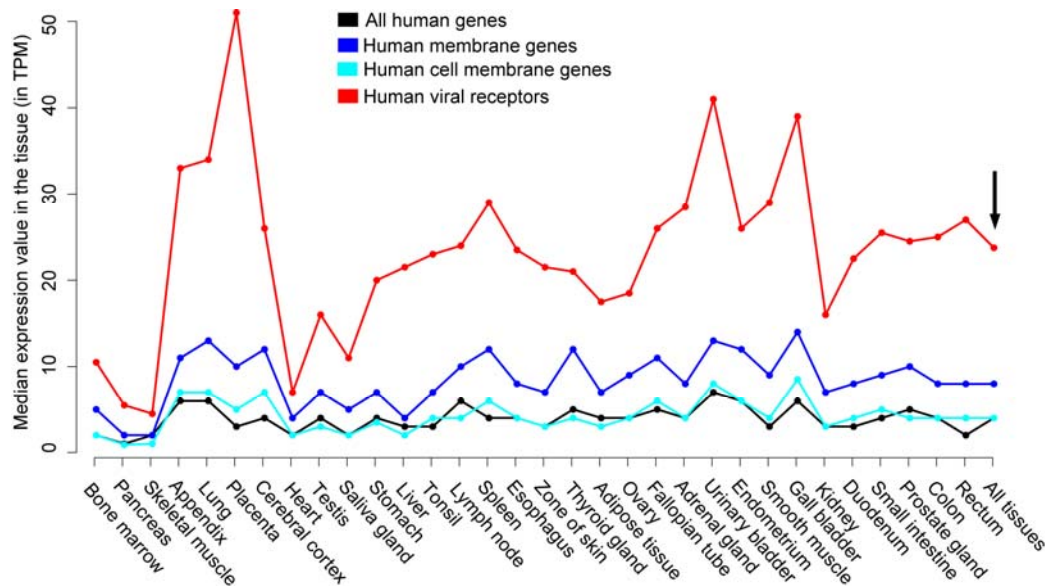


Figure 3. The average expression level of human viral receptors (red), human cell membrane genes (cyan), human membrane genes (blue) and all human genes (black) in 32 major human tissues. The expression level was measured with transcripts per million (TPM). The black arrow refers to the average expression level of genes in all

289 32 tissues.

290

291 **Viral receptors used by the same virus tended to co-evolve**

292 The results mentioned above shows that a total of 51 viruses used more than one viral
 293 receptor. These viral receptors may work together or independently. We then analyzed
 294 the relationship between them. Structural analysis shows that except for integrins
 295 which generally work in heterodimer, few of viral receptors used by the same virus
 296 shared the same protein domain (data not shown). This suggests that when the virus
 297 expands the use of receptors, it tends to select structurally diverse proteins. We
 298 continued to analyze the co-evolution between mammalian viral receptors in 108
 299 mammal species. We found that the average of Spearman Correlation Coefficients
 300 (SCCs, a measure of the extent of co-evolution) between viral receptors employed by
 301 the same virus was 0.54, which was significantly larger than that between other viral
 302 receptors (Figure 4A, p-value < 0.001 in the Wilcoxon rank-sum test). For example,
 303 SARS-CoV used four receptors, i.e., ACE2, CD209, C-type lectin domain family 4
 304 member G (CLEC4G) and member M (CLEC4M). The average of SCCs between
 305 these four receptors was as high as 0.86. In addition, we analyzed the extent of
 306 co-expression between human viral receptors in 32 tissues. It was found that the
 307 extent of co-expression between viral receptors employed by the same virus was a
 308 little larger than that between other viral receptors, yet this difference was not
 309 statistically significant (Figure 4B, p-value > 0.1 in the Wilcoxon rank-sum test).

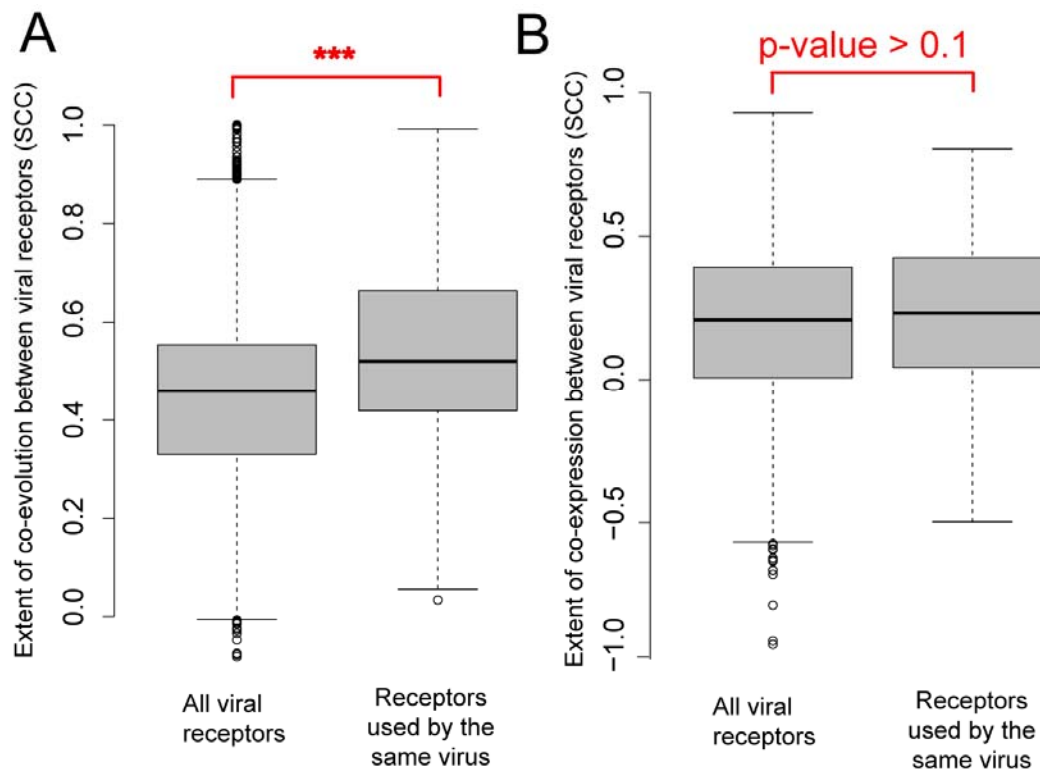


Figure 4. The co-evolution and co-expression of viral receptors. (A) Comparing the extent of co-evolution between mammalian virus receptors in 108 mammal species in the set of receptors used by the same virus and all mammalian virus receptors. “***”, p-value < 0.001 in the Wilcoxon rank-sum test. (B) Comparing the extent of co-expression between human viral receptors in 32 human tissues in the set of receptors used by the same virus and all human virus receptors.

Analysis of the association between the tissue and host specificity of the virus and the viral receptor

Although there were plenty of studies about the tissue and host specificity of the virus and the viral receptor, there was still a lack of systematic analysis towards the

association between them. Besides, few studies quantify such associations. Therefore, we further investigated systematically the association between the tissue and host specificity of the virus and the viral receptor.

1) Viral receptor expressed higher in tissues infected by viruses than in those not infected

To investigate the association between the tissue specificity of the virus and tissue-specific expression of viral receptors, we manually compiled the tissue tropism of viruses from the literature or Wikipedia and obtained that in 32 human tissues for a total of 52 viruses (Table S5). Some viral receptors had high expression levels in most tissues, most of which were housekeeping genes, such as CD81 molecule (CD81) and ITGB1. While for most viral receptors, their expression levels varied much in different tissues. Analysis of the association between the tissue-specific expression of viral receptors and viral tissue tropism showed that the viral receptor expressed higher in the tissues infected by viruses (marked with asterisks in Table S5) than in those not infected, yet this difference was not statistically significant (p-value > 0.1 in the Wilcoxon rank-sum test) (Figure S5). For example, the neural cell adhesion molecule 1 (NCAM1), which was employed by the Rabies lyssavirus (RabiesV) as the receptor, expressed much higher in the tissue of Cerebral cortex (infected by RabiesV) than in other tissues not infected by the virus (Table S5).

2) The viral receptor was a significant predictor in predicting viral cross-species in mammal species

343 Since the viral receptor determines the host specificity of the virus to a large extent, it
 344 is expected that the closer between the viral receptor and its homolog in a species, the
 345 more likely the virus which used the receptor would infect the species. To validate this
 346 hypothesis, we firstly calculated the sequence identities between the viral receptor and
 347 their homologs in 108 mammal species (Figure 5 and Table S6). For clarity, only 26
 348 mammal species, which were frequently observed, were presented in Figure 5. Then,
 349 we compared the sequence identities between viral receptor proteins and their
 350 homologs in the species infected by the virus which used the receptor (marked with
 351 asterisks and triangles), and in those not infected by the virus. As expected, the former
 352 was significantly higher than the latter (Figure 6A, $p\text{-value} < 0.001$ in the Wilcoxon
 353 rank-sum test).

354 Previous work by Olival et al. showed that phylogenetic relatedness of host was a
 355 major factor which influenced the cross-species of mammalian viruses^[7]. We
 356 compared the ability of the host relatedness and the viral receptor similarity in
 357 predicting viral cross-species in mammal species. The models based on the latter
 358 achieved an Area Under the ROC Curve (AUC) of 0.65 (Figure 6B), a little higher
 359 than that of the model based on the former (0.62), although this difference was not
 360 statistically significant ($p\text{-value} = 0.13$). This suggests that the viral receptor similarity
 361 should be a significant predictor in predicting viral cross-species in mammal species
 362 as well as the host relatedness.

Figure 5. Analysis of the association between the host specificity of the virus and the viral receptor. It listed the sequence identities (colored according to the legend) between the viral receptor and its homologs in 26 mammal species (at the bottom). The white referred to no homologs in the species. The viral receptor and the virus which used them were displayed in the left and right side of the figure respectively. For the non-human mammalian virus receptor, the species it belongs to was presented in brackets. The asterisk referred to the viral infection of the mammal species based on Olival's work, while the triangle referred to that based on our database. For more details, please see Table S6.

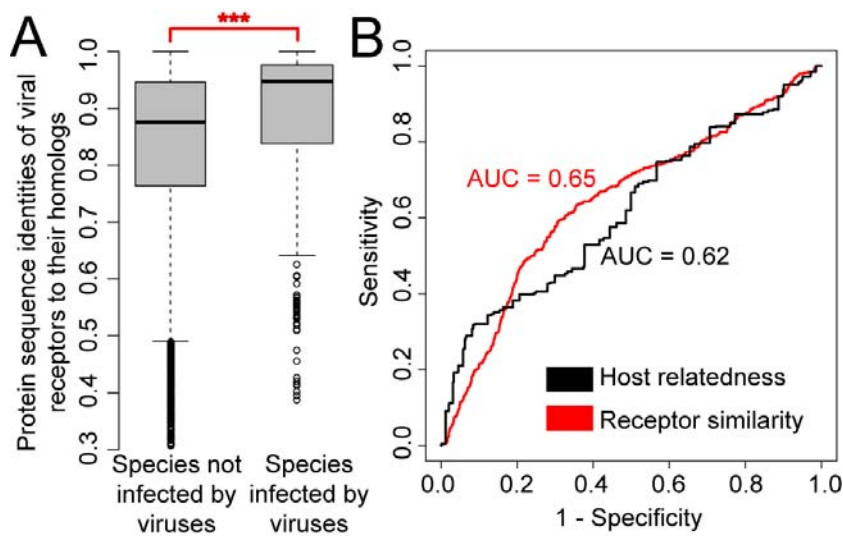


Figure 6. Quantify to what extent the viral receptor determine the host specificity of the virus. (A) Sequence identities between viral receptors and their homologs in mammal species infected by viruses and those not infected. “***”, p-value < 0.001. (B) The Receiver Operating Characteristic (ROC) curve for models of predicting viral cross-species in mammal species based on host relatedness (in black) and receptor sequence identity (in red). AUC, Area Under the ROC Curve.

380 Based on the results mentioned above, we continued to evaluate the risk of viral
381 cross-species transmission in 108 mammal species based on viral receptors. Figure 5
382 shows that more than 40% of viral receptors, such as insulin degrading enzyme (IDE)
383 and PHB, had high sequence identities with their homologs in most mammal species,
384 suggesting that the virus using them may have a high probability to infect these
385 species. On the other hand, some mammal species had homologs which were highly
386 similar to most viral receptors, such as the primates (Chimpanzee, *Macaca mulatta*
387 and Lowland gorilla). They may have a high risk of infection by the virus which used
388 these receptors.

389

390 **Discussion**

391 The viral receptor is essential for viral infection. By collecting the largest dataset ever
392 reported about the mammalian virus-host receptor interactions, we systematically
393 analyzed the structural, functional, evolutionary and tissue-specific expression
394 characteristics of mammalian virus receptors. We found that the viral receptors were a
395 subset of structurally and functionally diverse cell membrane proteins. They were
396 enriched in GO terms and KEGG pathways related to junctions, adhesion and binding,
397 which were typical features of viral receptors reported by previous studies ^{[10, 12, 14, 15,}
398 ^{21]}. Besides, our analysis identified several novel features of the viral receptor. Firstly,
399 the viral receptor had a higher level of N-glycosylation than other proteins. Then,
400 what's the relationship between glycosylation and viral receptor selection? As we

401 know, glycosylation of proteins is widely observed in eukaryote cells ^[32]. It plays an
 402 important role in multiple cellular activities, such as folding and stability of
 403 glycoprotein, immune response, cell-cell adhesion, and so on. Glycans are abundant
 404 on host cell surfaces. They were probably the primordial and fallback receptors for the
 405 virus ^[11]. To use glycans as their receptors, a large number of viruses have stolen a
 406 host galectin and employed it as a viral lectin ^[11, 33]. For example, the SJR fold, which
 407 was mainly responsible for glycan recognition and binding in cellular proteins, was
 408 observed in viral capsid proteins of over one fourth of viruses ^[33]. Thus, during the
 409 process of searching for protein receptors, the protein with high level of glycosylation
 410 could provide a basal attachment ability for the virus, and should be the preferred
 411 receptor for the virus.

412 Secondly, our analysis showed that the viral receptor protein had a tendency to
 413 interact with itself and had far more interaction partners than other membrane proteins.
 414 Besides the function of viral receptor, the receptor protein functions in the host cell by
 415 interacting with other proteins of the host, such as signal molecules and ligands.
 416 Therefore, the virus has to compete with these proteins for binding to the receptor ^[15].
 417 The protein with less interaction partners are expected to be preferred by the virus.
 418 Why did the virus select the proteins with multiple interaction partners as receptors?
 419 One possible reason is that the receptor proteins are closely related to the “door” of
 420 the cell, so that many proteins have to interact with them for in-and-out of the cell.
 421 This could be partly validated by the observation that for the interaction partners of
 422 human viral receptors, six of top ten enriched terms in the domain of GO Biological

423 Process were related to protein targeting or localization (Table S3). For entry into the
424 cell, the virus also selects these proteins as receptors. Another possible reason is that
425 viral entry into the cell needs cooperation of multiple proteins which were not
426 identified as viral receptors yet. Besides, previous studies show that the virus could
427 structurally mimic native host ligands ^[34], which help them bind to the host receptor.
428 Thus, membrane proteins with multiple interaction partners have a larger probability
429 to be used by viruses as receptors than other proteins.

430 Thirdly, the viral receptor was observed to have a much higher level of expression
431 than other genes in each of the 32 human tissues. This may be directly related to the
432 above finding that the viral receptor generally had multiple interaction partners: on the
433 one hand, the viral receptor needs multiple copies to interact with multiple proteins;
434 on the other hand, since the virus has to compete with other proteins for binding to the
435 receptor, high expression of the receptor will facilitate the virus's binding to the viral
436 receptor.

437 The virus-receptor interaction is a major determinant of viral host range and tissue
438 tropism. Previous case studies showed that the viral receptor expressed highly in the
439 tissues infected by the virus ^[35, 36]. Consistent with these studies, our systematic
440 analysis found that the tissues with high expression of the viral receptor, and the
441 mammal species with homologs highly similar to the viral receptor, were more
442 possibly to be infected by the virus. However, the opposites were also observed. Some
443 mammal species (or tissues) which had no receptor homolog (or low expression of the
444 viral receptor) were also infected by the virus. These viruses may use other receptors

not identified yet. Some mammal species (or tissues) with homologs highly similar to viral receptors (or high expression of the viral receptor) were observed to be not infected by the virus. This may be partly explained by the missed virus-host interactions in our data. Besides, it may also suggest that the host or tissue susceptibility to the virus is not solely determined by the viral receptor. More factors such as the host or tissue accessibility ^[7, 23], the cell defense system ^[37, 38] and the complex interaction between viral and host proteins ^[39, 40] may also influence viral infections.

There were some limitations within the study. Firstly, the viral receptor was biased towards the human, due to the bias of studies towards human viruses. Fortunately, the viral receptor was conserved in mammal species to a large extent, which may reduce the influence of this bias on the diversity of viral receptors. Secondly, the virus-host interactions were not complete due to limited surveys ^[7]. According to the risk analysis of viral cross-species based on viral receptors, much more mammal species may be infected by the mammalian virus analyzed in this study. High attention should be paid to this risk. Thirdly, due to the difficulties of identifying viral receptors ^[17, 41, 42], the database of mammalian virus-host receptor interaction was still limited in its size, which hindered us from a more comprehensive survey of the correlation characteristics between viruses and viral receptors. More effective methods, either experimental or computational ^[34], should be developed for identifying viral receptors, while the characteristics identified in this study may help such endeavors.

Overall, the structural, functional, evolutionary and tissue-specific expression

characteristics identified here should not only deepen our understanding of the viral receptor selection, but also help for development of more effective methods for identifying viral receptors. Besides, evaluating the risk of viral cross-species infection based on the viral receptor could also help for early warning and prediction of viral zoonotic diseases.

Materials and Methods

Database of mammalian virus-host receptor interaction

The data of mammalian virus-host receptor interaction were compiled from three sources: firstly, the literature related to viral receptors (a total of 1303 papers) were downloaded from NCBI Pubmed database^[43] by searching “virus receptor” [TIAB] or “viral receptor”[TIAB] on August 14th, 2017. The mammalian virus and their related host receptors were manually extracted from the literature; secondly, part of viral receptors were directly obtained from the database of ViralZone^[44] on September 9th, 2017; thirdly, proteins annotated with one of GO terms “virus receptor activity”, “viral entry into host cell” and “receptor activity” in UniprotKB database^[45] were collected on August 14th, 2017, and manually checked later. In combination, a database was created with 268 pairs of mammalian virus-host receptor interaction, which included 128 unique viral species or sub-species and 119 viral receptors (Table S1).

Analysis of structural features of viral receptors

488 The number of transmembrane alpha helix of the mammalian virus receptor was
 489 derived from the database of UniprotKB and the web server TMPred ^[46]. The location
 490 for the viral receptor was inferred from the description of “Subcellular location” for
 491 the receptor protein provided by UniProtKB, or from the GO annotations for them:
 492 the viral receptors annotated with GO terms which included the words of “cell surface”
 493 or “plasma membrane” were considered to be located in the cell membrane; those
 494 annotated with GO terms which included the words of “cytoplasm”, “cytosol” or
 495 “cytoplasmic vesicle”, or shown to be in the cytoplasm in UniProtKB, were
 496 considered to be located in the cytoplasm; those annotated with GO terms “nucleus”
 497 (GO:0005634) or “nucleoplasm” (GO:0005654) were considered to be located in the
 498 nuclear. The Pfam family, the N-glycosylation and O-glycosylation sites for the
 499 protein were obtained from the database of UniprotKB.

500 For comparison, the human proteins and their related structural characteristics were
 501 obtained from the database of UniProtKB/SwissProt on November 24th, 2017. The
 502 proteins which had at least one transmembrane alpha helix were considered as
 503 membrane proteins. The membrane proteins which were shown to be located in the
 504 cell membrane were considered as cell membrane proteins. In total, we obtained
 505 20243 human proteins, 5187 human membrane proteins and 2208 human cell
 506 membrane proteins.

507 The 3D structure for the viral receptor HTR2A were modeled with the help of
 508 I-TASSER ^[47] based on the protein sequence of HTR2A (accession number in the
 509 database of UniProt: P28223). The best model was selected, and visualized in RasMol

510 (version 2.7.5) ^[48].

511 **Functional enrichment analysis**

512 The GO function and KEGG pathway enrichment analysis for the human viral
513 receptor were conducted with functions of *enrichGO()* and *enrichKEGG()* in the
514 package “clusterProfiler” (version 3.4.4) ^[49] in R (version 3.4.2) ^[50].

515 **Protein-protein interaction (PPI) network analysis**

516 The human PPI network (PPIN) was constructed based on the work of Menche et al
517 ^[31]. The degree and betweenness for each protein in the PPIN were calculated with
518 functions of *degree()* and *betweenness()* in the package “igraph” (version 1.0.0) ^[51] in
519 R (version 3.2.5). The network was displayed with Cytoscape (version 2.6.2) ^[52].

520 For robustness of the results, we also conducted PPI analysis based on the human
521 PPIs derived from the database of STRING ^[53] on November 7, 2017. The human
522 PPIN was built based on the PPIs with median confidence (combined score equal to
523 or greater than 0.4). It included 710,188 PPIs and 17,487 proteins which could be
524 mapped to NCBI gene ids. Similar to those mentioned above, the viral receptor
525 protein was observed to have far more interaction partners and higher betweenness
526 than other proteins in the human PPIN (Figure S3A&C).

527 **Evolutionary analysis**

528 To identify the homolog of the mammalian virus receptor in other mammal species,
529 the protein sequence of each viral receptor was searched against the database of

mammalian protein sequences, which were downloaded from NCBI RefSeq database
^[54] on October 10th, 2017, with the help of BLAST (version 2.6.0) ^[55]. Analysis
showed that in the database of mammalian protein sequences, there were 108
mammal species which were richly annotated and had far more protein sequences
than other mammal species (Table S7). Therefore, only these 108 mammal species
were considered in the evolutionary analysis. Based on the results of BLAST, the
homolog for the viral receptor was defined as the hit with E-value small than 1E-10,
coverage equal to or greater than 80% and sequence identity equal to or greater than
30%. Only the closest homolog, i.e., the best hit, in each mammal species was used
for further analysis. For measuring the conservation level of viral receptors, two
indicators were used. The first indicator was the number of mammal species with
homolog of the viral receptor in 108 mammal species. The other indicator was the
average of the pairwise sequence identities between the viral receptor and its
homologs in 108 mammal species. For comparison, 1000 human protein sequences
were randomly selected from the NCBI RefSeq database (Table S4). Similar methods
as above were utilized to calculate the indicators of conservation level for these
proteins.

For analysis of co-evolution between viral receptors, firstly for each viral receptor, a
phylogenetic tree was built based on the protein sequences of the receptor and its
homologs in 108 mammal species with the help of phylip (version 3.68) ^[56]. The
neighbor-joining method was used with the default parameter. Then, the genetic
distances between the viral receptor and their homologs were extracted from the

552 phylogenetic tree with a perl script. Finally, for a pair of viral receptors, the spearman
553 correlation coefficient (SCC) was calculated between the pairwise genetic distances of
554 viral receptors and their homologs, which was used to measure the extent of
555 co-evolution between this pair of viral receptors.

556 The set of housekeeping gene in human was adapted from the work of Eisenberg et al
557 ^[57]. A total of 3804 genes were identified as the housekeeping gene.

558 **Analysis of the tissue-specific expression of human viral receptors**

559 The expression level for human viral receptors and other human genes in 32 human
560 tissues were derived from the database of Expression Atlas ^[58]. For analysis of the
561 association between viral infection and tissue-specific expression of viral receptors,
562 we manually compiled the tissue tropism of viruses from the literature or Wikipedia
563 and obtained that in 32 human tissues for a total of 52 viruses which used a total of 46
564 receptors (Table S5). When comparing the expression level of human viral receptors
565 and other set of human genes in 32 human tissues, to reduce the influence of extreme
566 values, the median instead of the mean of the expression values was used to measure
567 the average expression value of a gene set in a tissue.

568 The SCCs between the expression values of viral receptors in 32 tissues were
569 calculated to measure the extent of co-expression between viral receptors.

570 **Analysis of the host specificity of the virus and the viral receptor**

571 The mammalian virus-host interactions were primarily adapted from Olival's work ^[7].
572 One hundred and fifteen viruses in our database and 61 of 108 richly annotated

mammal species could be mapped to those in Olival's work (Table S8). These 115
viruses used a total of 116 viral receptors. The sequence identities of these viral
receptor proteins to their related homologs in the corresponding mammal species were
presented in Table S6.

For comparison, we also extracted genetic distances (host relatedness) between the
mammal species and the viral host with reported receptors based on Olival's work
(Table S9). Then, the genetic distance of the mammal species to the viral host with
reported receptors, and the sequence identity of the receptor homolog in the mammal
species to the viral receptor protein, was respectively used to predict whether a
mammal species could be infected by the virus which infected the host with reported
receptors. The method of Receiver Operating Characteristic (ROC) curve was used to
evaluate and compare their performance with the functions of *roc()*, *auc()*, *roc.test()*
and *plot.roc()* in the package of "pROC"^[59] in R (version 3.2.5).

Statistical analysis

All the statistical analysis was conducted in R (version 3.2.5)^[50]. The wilcoxon
rank-sum test was conducted with the function of *wilcox.test()*.

Acknowledgements

This work was supported by the National Key Plan for Scientific Research and
Development of China (2016YFD0500300 and 2016YFC1200200), the National
Natural Science Foundation of China (31500126, 31671371, 81730064 and

81571985), National Science and Technology Major Project (2017ZX10202201) and the Chinese Academy of Medical Sciences (2016-I2M-1-005). The authors would like to thank Pro. Xiangjun Du in Sun Yat-sen University for helpful suggestions.

The authors have declared that no competing interests exist.

Author contributions

HZ, TJ and YP conceived and designed the study. ZZ and ZZZ did the computational analysis. WC, ZC, ZT and BX compiled the database of mammalian viruses and their related receptors, and the tissue tropism of viruses. AW and XYG directed the computational analysis about the tissue and host specificity of viruses. YP and ZZ wrote the paper. AW, XYG, XHG, ZT, ZX, TJ and HZ reviewed and edited the manuscript. All authors read and approved the manuscript.

References

- [1] Mlakar J, Korva M, Tul N, et al. Zika virus associated with microcephaly. *New Engl J Med*, 2016, 374: 951-958
- [2] Maganga GD, Kapetshi J, Berthet N, et al. Ebola virus disease in the democratic republic of congo. *New Engl J Med*, 2014, 371: 2083-2091
- [3] Breban R, Riou J, Fontanet A. Interhuman transmissibility of middle east respiratory syndrome coronavirus: Estimation of pandemic risk. *Lancet*, 2013, 382: 694-699
- [4] Paez-Espino D, Eloie-Fadrosch EA, Pavlopoulos GA, et al. Uncovering earth's virome. *Nature*, 2016, 536: 425-+
- [5] Mihara T, Nishimura Y, Shimizu Y, et al. Linking virus genomes with host taxonomy. *Viruses-Basel*, 2016, 8:
- [6] Geoghegan JL, Senior AM, Di Giallonardo F, et al. Virological factors that increase the transmissibility of emerging human viruses. *Proceedings of the National Academy of Sciences of the United States of America*, 2016, 113: 4170-4175
- [7] Olival KJ, Hosseini PR, Zambrana-Torrel C, et al. Host and viral traits predict zoonotic

624 spillover from mammals. *Nature*, 2017, 546: 646-+

625 [8] Sharp PM, Hahn BH. Origins of hiv and the aids pandemic. *Csh Perspect Med*, 2011, 1:

626 [9] Ge XY, Li JL, Yang XL, et al. Isolation and characterization of a bat sars-like coronavirus

627 that uses the ace2 receptor. *Nature*, 2013, 503: 535-+

628 [10] Dimitrov DS. Virus entry: Molecular mechanisms and biomedical applications. *Nat Rev*

629 *Microbiol*, 2004, 2: 109-122

630 [11] Li F. Receptor recognition mechanisms of coronaviruses: A decade of structural studies.

631 *Journal of virology*, 2015, 89: 1954-1964

632 [12] Baranowski E, Ruiz-Jarabo CM, Domingo E. Evolution of cell recognition by viruses.

633 *Science*, 2001, 292: 1102-1105

634 [13] Grove J, Marsh M. The cell biology of receptor-mediated virus entry. *The Journal of cell*

635 *biology*, 2011, 195: 1071-1082

636 [14] Casasnovas JM. Virus-receptor interactions and receptor-mediated virus entry into host

637 cells. 2013[

638 [15] Wang JH. Protein recognition by cell surface receptors: Physiological receptors versus virus

639 interactions. *Trends in biochemical sciences*, 2002, 27: 122-126

640 [16] Li F. Structure, function, and evolution of coronavirus spike proteins. *Annu Rev Virol*, 2016,

641 3: 237-261

642 [17] Yan H, Zhong G, Xu G, et al. Sodium taurocholate cotransporting polypeptide is a

643 functional receptor for human hepatitis b and d virus. *eLife*, 2012, 1: e00049

644 [18] Peng WJ, de Vries RP, Grant OC, et al. Recent h3n2 viruses have evolved specificity for

645 extended, branched human-type receptors, conferring potential for increased avidity. *Cell host &*

646 *microbe*, 2017, 21: 23-34

647 [19] Isa P, Arias CF, Lopez S. Role of sialic acids in rotavirus infection. *Glycoconjugate journal*,

648 2006, 23: 27-37

649 [20] Mazzon M, Mercer J. Lipid interactions during virus entry and infection. *Cellular*

650 *microbiology*, 2014, 16: 1493-1502

651 [21] Marija Backovic FAR. Virus entry: Old viruses, new receptors. *Current opinion in virology*,

652 2012, 2: 10

653 [22] Coffin JM. Virions at the gates: Receptors and the host-virus arms race. *PLoS Biology*, 2013,

654 11:

655 [23] Kumlin U, Olofsson S, Dimock K, et al. Sialic acid tissue distribution and influenza virus

656 tropism. *Influenza and other respiratory viruses*, 2008, 2: 147-154

657 [24] Harris HJ, Davis C, Mullins JG, et al. Claudin association with cd81 defines hepatitis c virus

658 entry. *The Journal of biological chemistry*, 2010, 285: 21092-21102

659 [25] Ribeiro RM, Hazenberg MD, Perelson AS, et al. Naive and memory cell turnover as drivers

660 of ccr5-to-cxcr4 tropism switch in human immunodeficiency virus type 1: Implications for

661 therapy. *Journal of virology*, 2006, 80: 802-809

662 [26] Carter CC, McNamara LA, Onafuwa-Nuga A, et al. Hiv-1 utilizes the cxcr4 chemokine

663 receptor to infect multipotent hematopoietic stem and progenitor cells. *Cell host & microbe*, 2011,

664 9: 223-234

665 [27] Taubenberger JK, Kash JC. Influenza virus evolution, host adaptation, and pandemic

666 formation. *Cell host & microbe*, 2010, 7: 440-451

667 [28] Lu G, Wang Q, Gao GF. Bat-to-human: Spike features determining 'host jump' of

668 coronaviruses sars-cov, mers-cov, and beyond. *Trends in microbiology*, 2015, 23: 468-478

669 [29] Li F. Receptor recognition and cross-species infections of sars coronavirus. *Antiviral*

670 *research*, 2013, 100: 246-254

671 [30] Maginnis MS, Haley SA, Gee GV, et al. Role of n-linked glycosylation of the 5-ht2a receptor

672 in jc virus infection. *Journal of virology*, 2010, 84: 9677-9684

673 [31] Menche J, Sharma A, Kitsak M, et al. Uncovering disease-disease relationships through the

674 incomplete interactome. *Science*, 2015, 347:

675 [32] Corfield A. Eukaryotic protein glycosylation: A primer for histochemists and cell biologists.

676 *Histochemistry and cell biology*, 2017, 147: 119-147

677 [33] Krupovic M, Koonin EV. Multiple origins of viral capsid proteins from cellular ancestors.

678 *Proceedings of the National Academy of Sciences of the United States of America*, 2017, 114:

679 E2401-E2410

680 [34] Drayman N, Glick Y, Ben-nun-shaul O, et al. Pathogens use structural mimicry of native

681 host ligands as a mechanism for host receptor engagement. *Cell host & microbe*, 2013, 14: 63-73

682 [35] Boonarkart C, Champunot R, Uiprasertkul M, et al. Case report: Increased viral receptor

683 expression associated with high viral load and severe pneumonia in a young patient infected with

684 2009 h1n1 influenza a with no pre-existing conditions. *Journal of medical virology*, 2012, 84:

685 380-385

686 [36] Nowakowski TJ, Pollen AA, Di Lullo E, et al. Expression analysis highlights axl as a

687 candidate zika virus entry receptor in neural stem cells. *Cell stem cell*, 2016, 18: 591-596

688 [37] McNab F, Mayer-Barber K, Sher A, et al. Type i interferons in infectious disease. *Nature*

689 *reviews Immunology*, 2015, 15: 87-103

690 [38] Jost S, Altfeld M. Control of human viral infections by natural killer cells. *Annual review of*

691 *immunology*, 2013, 31: 163-194

692 [39] Randall G, Panis M, Cooper JD, et al. Cellular cofactors affecting hepatitis c virus infection

693 and replication. *Proceedings of the National Academy of Sciences of the United States of America*,

694 2007, 104: 12884-12889

695 [40] Konig R, Zhou Y, Elleder D, et al. Global analysis of host-pathogen interactions that

696 regulate early-stage hiv-1 replication. *Cell*, 2008, 135: 49-60

697 [41] Li W. The hepatitis b virus receptor. *Annual review of cell and developmental biology*, 2015,

698 31: 125-147

699 [42] Pillay S, Meyer NL, Puschnik AS, et al. An essential receptor for adeno-associated virus

700 infection. *Nature*, 2016, 530: 108-112

701 [43] Agarwala R, Barrett T, Beck J, et al. Database resources of the national center for

702 biotechnology information. *Nucleic Acids Res*, 2016, 44: D7-D19

703 [44] Masson P, Hulo C, De Castro E, et al. Viralzone: Recent updates to the virus knowledge

704 resource. *Nucleic Acids Res*, 2013, 41: D579-D583

705 [45] Bateman A, Martin MJ, O'Donovan C, et al. Uniprot: The universal protein knowledgebase.

706 *Nucleic Acids Res*, 2017, 45: D158-D169

707 [46] Hofmann K, Stoffel W. Tmpred: Prediction of transmembrane regions and orientation. 2017,

708 https://embnet.vital-it.ch/software/TMPRED_form.html

709 [47] Roy A, Kucukural A, Zhang Y. I-tasser: A unified platform for automated protein structure

710 and function prediction. *Nat Protoc*, 2010, 5: 725-738

711 [48] Bernstein HJ. Rasmol 2.7.5. 2017, <http://www.openrasmol.org/>

712 [49] Yu GC, Wang LG, Han YY, et al. Clusterprofiler: An r package for comparing biological
713 themes among gene clusters. *Omics*, 2012, 16: 284-287

714 [50] Team RC. R: A language and environment for statistical computing. R foundation for
715 statistical computing, vienna, austria. 2016, <https://www.R-project.org/>

716 [51] G C, T N. The igraph software package for complex network research, *interjournal*,
717 *complex systems* 1695. 2006, <http://igraph.org>

718 [52] Shannon P, Markiel A, Ozier O, et al. Cytoscape: A software environment for integrated
719 models of biomolecular interaction networks. *Genome research*, 2003, 13: 2498-2504

720 [53] Szklarczyk D, Franceschini A, Wyder S, et al. String v10: Protein-protein interaction
721 networks, integrated over the tree of life. *Nucleic Acids Res*, 2015, 43: D447-D452

722 [54] Pruitt KD, Tatusova T, Brown GR, et al. Ncbi reference sequences (refseq): Current status,
723 new features and genome annotation policy. *Nucleic Acids Res*, 2012, 40: D130-D135

724 [55] Altschul SF, Gish W, Miller W, et al. Basic local alignment search tool. *Journal of molecular*
725 *biology*, 1990, 215: 403-410

726 [56] Felsenstein J. Phylip - phylogeny inference package (version 3.2). *Cladistics*, 1989, 5: 3

727 [57] Eisenberg E, Levanon EY. Human housekeeping genes, revisited. *Trends Genet*, 2013, 29:
728 569-574

729 [58] Petryszak R, Keays M, Tang YA, et al. Expression atlas update—an integrated database of
730 gene and protein expression in humans, animals and plants. *Nucleic Acids Res*, 2016, 44:
731 D746-D752

732 [59] Robin X, Turck N, Hainard A, et al. Proc: An open-source package for r and s plus to
733 analyze and compare roc curves. *Bmc Bioinformatics*, 2011, 12: