

1 **Mapping diversification metrics in macroecological studies: Prospects and**
2 **challenges**

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16

17 **Abstract**

18 The intersection of macroecology and macroevolution is one of the most active
19 research areas today. Macroecological studies are increasingly using phylogenetic
20 diversification metrics to explore the role of evolutionary processes in shaping
21 present-day patterns of biodiversity. Evolutionary explanations of species richness
22 gradients are key for our understanding of how diversity accumulated in a region.
23 For instance, the present-day diversity in a region can be a result of *in situ*
24 diversification, extinction, or colonization from other regions, or a combination of
25 all of these processes. However, it is unknown whether these metrics capture well
26 these diversification and dispersal processes across geography. Some metrics
27 (e.g., mean root distance -MRD-; lineage diversification-rate -DR-; evolutionary
28 distinctiveness -ED-) seem to provide very similar geographical patterns
29 regardless of how they were calculated (e.g., using branch lengths or not). The
30 lack of appropriate estimates of extinction and dispersal rates in phylogenetic
31 trees can limit our conclusions about how species richness gradients emerged.
32 With a review of the literature and complemented by an empirical comparison, we
33 show that phylogenetic metrics by itself are not capturing well the speciation,
34 extinction and dispersal processes across the geographical gradients.
35 Furthermore, we show how new biogeographic methods can improve our
36 inference of past events and therefore our conclusions about the evolutionary
37 mechanisms driving regional species richness. Finally, we recommend that future
38 studies include several approaches (e.g., spatial diversification modelling,
39 parametric biogeographic methods) to disentangle the relative the role of

40 speciation, extinction and dispersal in the generation and maintenance of species

41 richness gradients.

42

43 Keywords: Phylogeny, Geography, Spatial diversification, Macroevolution, Species

44 richness, Regional assemblages

45 **Introduction**

46 The causes of spatial variation of biodiversity are one of the most
47 fundamental questions in ecology, biogeography and macroecology (Brown
48 1995, 2014, Brown and Lomolino 1998, Hawkins et al. 2012, Fine 2015, Jablonski
49 et al. 2017). Current studies are integrating in a single framework the ecological
50 and evolutionary mechanisms driving regional species diversity (McGaughan
51 2015, Pärtel et al. 2016, Cabral et al. 2017, Leidinger and Cabral 2017). However,
52 only three macroevolutionary processes ultimately can modify the number of
53 species in a region: speciation, extinction and dispersal (Wiens 2011, Fine 2015,
54 Jablonski et al. 2017) (Figure 1). These processes can be modulated by species'
55 traits varying within clades (Paper et al. 2016, 2017, Jezkova and Wiens 2017,
56 Moen and Wiens 2017), age of region (e.g., time-for-speciation effect; Stephens &
57 Wiens, 2003), geographical area (Losos and Schluter 2000), or climatic conditions
58 (Condamine et al. 2013, Lewitus and Morlon 2017).

59 The integration of different disciplines such as molecular phylogenetics,
60 palaeontology, and historical biogeography have allow to infer a series of
61 macroevolutionary processes across geography (Diniz-Filho et al. 2013, Fritz et al.
62 2013). It is well-known that fossil information is key to estimate with high
63 confidence rates of speciation and extinction (Sepkoski 1998, Foote 2000,
64 Quental and Marshall 2010, Rabosky 2010b). New methods correcting for
65 sampling bias are able to generate improvements in the estimates of the
66 speciation, extinction and net diversification rates (Silvestro et al. 2014, 2016).

67 However, the causal mechanisms that underlying the geographical diversity
68 gradients only can be established with greater confidence for a few taxonomic
69 groups with adequate fossil record, such as marine bivalves (Jablonski et al. 2006,
70 2017), mammals (Silvestro et al. 2014) or plants (Antonelli et al. 2015).

71 As fossil data is not available or incomplete for most extant groups, model-
72 based approaches used to estimate speciation and extinction rates in
73 palaeontology were adapted to study the macroevolutionary dynamics using
74 phylogenetic information (Nee et al. 1994, Morlon et al. 2010, Stadler 2013).
75 Molecular phylogenies are becoming essential to the study of diversification
76 dynamics across temporal and spatial scales for extant taxa (Wiens and
77 Donoghue 2004, Rabosky and Lovette 2008, Stadler 2013, Morlon 2014, Schluter
78 and Pennell 2017). Therefore, it is possible to reconstruct past diversification
79 process based on the branching events of a phylogeny using a set of birth-death
80 models (Nee et al. 1994, Nee 2006, Morlon et al. 2010, Stadler 2013, Morlon
81 2014). These birth-death models allow infer either a homogeneous process for an
82 entire clade (Nee et al. 1994, Magallón and Sanderson 2001) or a heterogeneous
83 process varying in time or in specific subclades of a tree (Paper et al. 2006,
84 Rabosky and Lovette 2008, Alfaro et al. 2009). However, these birth-death models
85 only account for temporal variation of the macroevolutionary processes and how
86 translate these processes to the geography is still a matter of debate.

87 Macroecological studies use two main approaches to link the estimates of
88 diversification with the geographical ranges of species (Hawkins et al. 2007, Algar
89 et al. 2009, Qian et al. 2014, Pinto-Ledezma et al. 2017, Velasco et al. 2018). The

90 first one uses a set of phylogenetic metrics as a proxy to capture the geographical
91 signature of lineage diversification dynamics (Diniz-Filho *et al.*, 2013; Fritz *et al.*,
92 2013; Table 1; Figure 2). These metrics provide either an estimate of a per-species
93 rate of diversification (e.g., mean root distance –MRD-, residual phylogenetic
94 diversity –rPD-, mean diversification rate –MDR-), the phylogenetic structure of
95 regional assemblages (e.g., phylogenetic species variability –PSV-) or the average
96 age of co-occurring lineages in a given area (e.g., mean age; see Table 1). Each
97 metric is calculated for each species in the phylogeny; therefore, we can associate
98 the species' values to its corresponding geographical range and generate a map
99 with average values for cells or regions. Although these phylogenetic metrics only
100 account for speciation events, macroecologists have used these maps as a proxy
101 to test some evolutionary-based hypothesis in macroecological research (Diniz-
102 Filho *et al.*, 2013; Fritz *et al.*, 2013; see Table 2 for a compendium of these
103 hypotheses).

104 The second approach used by macroecologists consists in the explicit
105 estimation of diversification parameters across geography (Goldberg *et al.* 2005,
106 2011, Ramiadantsoa *et al.* 2017). For instance, the geographic state speciation
107 and extinction model –GeoSSE (Goldberg *et al.* 2011; Table 1) allows estimating
108 speciation, extinction and dispersal parameters across two regions. It is possible
109 to disentangle the relative role of each one of these processes on the generation
110 and maintenance of the geographical diversity gradients (Rolland *et al.* 2014,
111 Pulido-Santacruz and Weir 2016, Pinto-Ledezma *et al.* 2017). In addition, a
112 recently developed Bayesian approach (BAMM; Rabosky 2014, Rabosky *et al.*

113 2014) allows to infer the balance of speciation and extinction in the generation of
114 these biodiversity gradients (Rabosky et al. 2015, Sánchez-Ramírez et al. 2015,
115 Morinière et al. 2016, Pinto-Ledezma et al. 2017). The BAMM approach allows
116 both the inference of macroevolutionary dynamics for an entire clade (i.e., a
117 macroevolutionary regime; Rabosky 2014) and also get estimates of per-species
118 diversification rates (i.e., as a phylogenetic metric; Rabosky 2016) that can be
119 mapped in a geographical domain. Although all these methods aim to obtain a
120 geographical picture of the diversification processes, it remains unexplored if they
121 can effectively capture these dynamics across regions.

122 In this paper, we conducted a review on macroecological literature to
123 evaluate how evolutionary and biogeographic processes contribute to shape
124 geographical species richness gradients. We review only those papers that make
125 explicit use of phylogenetic metrics and/or explicit diversification approaches
126 (Table 1). We divided our review in three main sections. In the first one, we
127 discuss how studies use phylogenetic metric to test some evolutionary-based
128 hypotheses underlying geographical diversity gradients and we explore some
129 limitations of these metrics (see also Table 2). Also, we discuss to what extent
130 these metrics are able to capture macroevolutionary dynamics in a spatial explicit
131 context. We illustrate these using three case studies (Furnariides birds, Hylid
132 frogs, and *Anolis* lizards; Figure 1) and explore how another approaches (e.g.,
133 diversification modelling and biogeographical approaches) can complement our
134 inferences about diversification process across geography. In the second section,
135 we discuss how dispersal and extinction processes are limiting these

136 diversification inferences and we propose some research avenues to attempt to
137 solve these problems. Using an explicit biogeographical approach, we test the
138 role of dispersal on the geographical species richness patterns of the three case
139 studies. Finally, in the third section, we call for the adoption of complementary
140 approaches (e.g., extensive simulations, parametric biogeographical methods) in
141 macroecological research with the aim to evaluate the relative role of speciation,
142 extinction and dispersal process driving geographical biodiversity gradients.

143

144 **LITERATURE REVIEW**

145 We conducted a literature search in Web of Science for studies that
146 explicitly have addressed questions on how speciation, extinction and dispersal
147 have shaped geographical species richness gradients. We selected those papers
148 that used either phylogenetic metrics (e.g., mean root distance -MRD-,
149 phylogenetic diversity -PD-, phylogenetic species variability -PSV-; diversification
150 rate -DR-; mean Ages; Table 1) or explicit macroevolutionary approaches (e.g.,
151 GeoSSE, BAMM; Goldberg *et al.*, 2011; Rabosky 2014). We compiled a list of 44
152 papers (Table A1), but we are aware that this likely is not an exhaustive search.
153 The majority of papers reviewed are testing historical process shaping latitudinal
154 diversity gradients (LDG) in various taxa.

155

156 **TESTING EVOLUTIONARY HYPOTHESIS USING PHYLOGENETIC METRICS** 157 **AND EXPLICIT DIVERSIFICATION APPROACHES**

158 Here, we discuss how phylogenetic metrics are used to test evolutionary
159 hypotheses related to the generation and maintenance of geographical diversity.
160 Several studies used the mean root distance -MRD- metric to evaluate whether
161 regional assemblages are composed of “basal” or “derivate” linages. First, this
162 terminology should be avoided because it provides an incorrect interpretation of
163 the phylogenetic trees (Baum et al. 2005, Crisp and Cook 2005, Omland et al.
164 2008). Although this metric does not incorporate information from branch lengths
165 (Algar et al. 2009, Qian et al. 2015), it does provide the average number of nodes
166 separating each species in a given region from the root of the phylogeny (Kerr and
167 Currie 1999). MRD therefore provides information about the number of
168 cladogenetic events (splits) that have occurred through the history of co-occurring
169 lineages in each region (Pinto-Ledezma et al. 2017, Velasco et al. 2018). Under
170 this view, MRD should be interpreted as a metric of total diversification (Rabosky
171 2009), where high MRD values indicating regional assemblages dominated by
172 extensive cladogenesis and low MRD values indicating assemblages with few
173 cladogenetic events. A main concern with this metric concern with the fact that it
174 does not provide any information about what macroevolutionary dynamics have
175 taken place in a region. For example, it is very hard to establish whether MRD
176 allows to distinguish between diversity-dependent (Rabosky 2009, Rabosky and
177 Hurlbert 2015) or time-dependent (Wiens 2011, Harmon and Harrison 2015)
178 processes dominating regional diversity. Although the distinction between these
179 two dynamics, and its relationship with the origin and maintenance of regional
180 diversity, is an intense topic in the macroevolutionary literature (Rabosky 2009,

181 2013, Wiens 2011, Cornell 2013, Harmon and Harrison 2015, Rabosky and
182 Hurlbert 2015). However, more empirical and theoretical work is necessary to
183 establish what scenario plays a significant role in regional species richness
184 assembly (Rabosky 2012, Etienne et al. 2012, Valente et al. 2015, Graham et al.
185 2018). It might worth to establish whether local ecological process scaling up to
186 regional scales or emergent effects (i.e., the existence of a strong equilibrium
187 process) governed the build-up of regional diversity (Cornell 2013, Harmon and
188 Harrison 2015, Rabosky and Hurlbert 2015, Marshall and Quental 2016).

189 The time-for-species effect hypothesis state that the regional build-up of
190 species richness is directly proportional to the colonization time of its constituent
191 clades (Stephens and Wiens 2003b; Table 2). However, many phylogenetic
192 metrics used to test this hypothesis, did not incorporate any age information (Fritz
193 and Rahbek 2012, Qian et al. 2015). Qian et al. (2015) did some additional
194 predictions for the time effect hypothesis regarding the phylogenetic structure of
195 regional assemblages. We suggest that these predictions are not easily deduced
196 from the original statement of the time-for-speciation effect hypothesis (Stephens
197 & Wiens 2003). For instance, Qian et al. (2015, p. 7) predicted that regions with
198 low species richness (e.g., extra-tropical regions) should be composed of more
199 closely related species than regions with high species richness (e.g., tropical
200 regions). This assumes that regions with low species richness were colonized
201 recently and therefore these lineages had little time for speciation. However, it is
202 also plausible consider that high extinction occurred in these poor species
203 richness regions by marginal climatic niche conditions preventing adaptive

204 diversification (Wellborn and Langerhans 2015). By contrast, regions with high
205 species richness might also be assembled by multiple dispersals from nearby
206 regions becoming to be a macroevolutionary sink (Goldberg et al. 2005). In this
207 latter case, the species richness was not build-up by *in situ* speciation mainly but
208 by continued dispersal through time. To evaluate which of these scenarios is more
209 plausible it is necessary to adopt an approach that explicitly infer the number of
210 the dispersal and cladogenetic events across areas (Roy and Goldberg 2007,
211 Dupin et al. 2017).

212 Differences in species diversification are also considered as a main driver of
213 the geographical diversity gradient for many groups (Kennedy et al. 2014, Pinto-
214 Ledezma et al. 2017). This hypothesis states (Table 2) that differences in net
215 diversification rates between areas are the main driver of differences in regional
216 species richness between areas. Davies and Buckley (2011) used the phylogenetic
217 diversity controlled by species richness (i.e., residual PD –rPD-) to distinguish
218 areas with different evolutionary processes. These authors predicted that areas
219 where rapid speciation and low immigration events from other areas occurred, are
220 dominated by large adaptive radiations (e.g., large islands; Losos and Schluter
221 2000). By contrast, areas with slow speciation and colonized by multiple lineages
222 through time should have high values of residual PD.

223 The “out of the tropics” -OTT- hypothesis (Jablonski et al. 2006; Table 2)
224 states that latitudinal diversity gradient is due to that the majority of lineages
225 originated in the tropics and then migrated to extratropical regions. Under this

226 hypothesis, tropics harbour higher net diversification rates (higher speciation and
227 lower extinction) than extratropical regions and dispersal rates are higher from the
228 tropics to extratropical regions than the reverse (Jablonski et al. 2006; Table 1).
229 For instance, Rolland et al. (2014) used the GeoSSE model to test this hypothesis
230 in the generation of the latitudinal mammal diversity gradient. They found that net
231 diversification rates (i.e., the balance of speciation minus extinction) was higher in
232 tropical than in temperate regions and dispersal rates were higher from the tropics
233 to temperate regions than the reverse. Also, Pinto-Ledezma et al. (2017) used the
234 GeoSSE model to test an analogue hypothesis to OTT, as form of Out of the
235 Forest hypothesis (OTF), using Furnariides birds as a clade model. Their favoured
236 a model where open areas have higher speciation, extinction and dispersal rates
237 than forest habitats. All these results suggest that it is reasonable to use either
238 phylogenetic metrics or explicit diversification approaches (e.g., the GeoSSE
239 model) to evaluate a set of evolutionary-based hypotheses as a main driver of
240 geographical diversity gradients. However, we show here (see below) that these
241 approaches fail to capture the evolutionary and biogeographic processes at
242 spatial scales.

243

244 ***Are phylogenetic metrics capturing well the diversification process across***
245 ***geography?***

246 A deep understanding of evolutionary processes affecting regional species
247 assemblages is coming from the integration of molecular phylogenies and fossil

248 record (Quental and Marshall 2010, Marshall 2017). From this integration of
249 neontological and paleontological perspectives, it is clear that both approaches
250 are necessary to test evolutionary-based hypothesis in macroecological research.
251 Several hypotheses were proposed to explain geographical diversity patterns,
252 particularly the latitudinal diversity gradient –LDG- (see Table 2 for a summary and
253 compilation of the main hypotheses reported in the literature). Although the ideal
254 approach is to generate robust conclusions from multiple lines of evidence (e.g.,
255 fossil record, molecular phylogenies, biogeographical inference) it is clear that this
256 information is scarce for many taxonomic groups. Many macroecological studies
257 have adopted either phylogenetic metrics or explicit diversification approaches
258 (e.g., the GeoSSE model) to evaluate the relative contribution of speciation,
259 extinction and dispersal on the resulting geographical diversity gradients (Table
260 S1).

261 Phylogenetic metrics can be easily visualized in a geographical context and
262 several inferences about ecological (e.g., dispersal) and evolutionary (e.g.,
263 speciation) process can be done. As these metrics provide a *per-species level*
264 *diversification* metric for each species in a phylogeny, it is possible to associate
265 these values with the corresponding species' geographical range and obtain a
266 mean value for cells or regions in a given geographical domain (Table 1; Figure 2).
267 By contrast, explicit diversification approaches (e.g., GeoSSE; BAMM; fitting
268 models) provide a *per-lineage level diversification* metric for a given clade or a
269 regional assemblage (Rabosky 2016a). However, in some cases, it is possible to

270 generate a *per-species level diversification* metric with these approaches. For
271 instance, Pérez-Escobar et al. (2017) used the function GetTipsRates in
272 BAMMtools (Rabosky et al. 2014) to map speciation rates for Neotropical orchids.

273 These two approaches (phylogenetic metrics and lineage diversification)
274 potentially can provide complementary pictures about how macroevolutionary
275 dynamics have taken place in the geography. On the one hand, it is possible to
276 estimate diversification rates for a given clade using the number of species, its
277 age and a birth-death models (Magallón and Sanderson 2001, Nee 2006,
278 Sánchez-Reyes et al. 2017). These model-fitting approaches allow to whether
279 diversity- or time-dependent diversification process has taken place in a regional
280 assemblage (Etienne et al. 2012, Rabosky 2014, Valente et al. 2015). On the other
281 hand, per-species diversification rate metrics allow establishing the potential of
282 each individual species to generate more species (Jetz et al. 2012, Rabosky 2014,
283 2016a). However, these approaches imply at least a different process, which left a
284 different signature on the geography. Phylogenetic metrics captures a total
285 diversification process (Rabosky 2009), whereas lineage diversification
286 approaches (e.g., BAMM) can potentially provide information about an individual
287 diversification process (Rabosky 2013). In addition, still is not clear whether
288 phylogenetic metrics can provide an accurate description of the diversification
289 dynamics across geography.

290 The first step to clarify how well these phylogenetic metrics behave is to
291 establish a comparison within and between different taxonomic groups. To

292 evaluate how different phylogenetic metrics vary across geography and their
293 relationship with species richness, we used two empirical data sets from our own
294 empirical work (furnariid birds and anole lizards; (Pinto-Ledezma et al. 2017,
295 Velasco et al. 2018) and a data set compiled from several sources (hylid frogs;
296 (Wiens et al. 2006, Algar et al. 2009, Pyron 2014a). We mapped across geography
297 five phylogenetic metrics (Table 1, Figure 2). We selected these three data sets
298 because previous work analysed how evolutionary-based hypotheses affected the
299 present-day species richness gradient (Wiens et al. 2006, Algar et al. 2009, Pinto-
300 Ledezma et al. 2017, Velasco et al. 2018).

301 Figure 2 shows the geographical pattern of species richness and the five
302 phylogenetic metrics for *Anolis* lizards, hylid frogs and Furnariides birds. For all
303 clades, there is a higher species concentration near to the Ecuador. Higher
304 species concentration for hylids and Furnariides can be found in the Amazon and
305 the Atlantic forest and for *Anolis* lizards in Central America and the Caribe (Figure
306 2A-C; see also Algar et al. 2009, Pinto-Ledezma et al. 2017, and Velasco et al.
307 2018, for a detailed description of the geographical species richness pattern for
308 these clades, respectively). In terms of the geographical pattern of each
309 phylogenetic metric (Figure 2D-R), in most of the cases cells with higher metric
310 values are related to cells that contain high species richness and vice versa
311 (Figure 2D-R; Figure A1). However, the degree and the direction of this
312 relationship changes according to the phylogenetic metric used. For example,
313 MRD, a metric of species derivedness, shows a negative correlation with species
314 richness (Figure 2J-L; Figure A1). Importantly, the spatial relationships between

315 species richness and phylogenetic metrics found in our analysis could simply be
316 the result of aggregated species-level attributes within cells or assemblages
317 (Hawkins et al. 2017). Hence, any conclusion derived from these relationships
318 needs to be considered carefully. In addition, there are different levels of
319 correlation between phylogenetic metrics (Figure A1). For example, MDR - MA
320 present a high but negative correlation, and rPD - PSV and MRD - MDR present a
321 mid-high positive correlation (Figure A1). Although there are few studies
322 comparing correlations between metrics (Vellend et al. 2010, Miller et al. 2017), to
323 our knowledge, none previous study compares the similarity of these
324 diversification metrics (Table 1). However, some of these metrics sharing
325 mathematical assumptions, which increase the likelihood of correlation between
326 them. For example, for ultrametric trees, metrics as MDR could be approximated
327 by considering the mean root distance (i.e. MRD metric) from the tips to the root
328 (Freckleton et al. 2008), so further studies exploring the mathematical relation
329 between metrics are needed.

330 In order to assess if the cells/assemblages on average do not represent a
331 random sampling from the species pool, we applied a simple permutation test to
332 explore the non-randomness in each of the phylogenetic metrics. We applied a
333 null model where the presence-absence matrix (i.e., PAM) was randomly shuffled
334 1000 times, but maintaining the frequency of species occurrence and observed
335 richness in the cells/assemblages (Gotelli 2000). This kind of null model is
336 standard in studies at the community/assemblage level that use phylogenetic
337 information (Cavender-Bares et al. 2004, 2006). Interestingly, none of the

338 phylogenetic metrics deviates from the null expectation for the three clades
339 (Figure 3). Also, very few cells/assemblages present p-values below the 0.05
340 threshold, thus indicating that the cells/assemblages present random association
341 among species (Figure 3). These results should be supported by repeating
342 analyses with more clades at different spatial extents, but again, we stress that
343 any result obtained with the use of phylogenetic metrics need to be interpreted
344 carefully.

345

346 ***A brief comparison between phylogenetic metrics and explicit diversification***
347 ***and biogeographic approaches***

348 We compared the phylogenetic metrics enunciated in Table 1, which have
349 been the most used in macroecological research. We explored whether the
350 geographical patterns of these phylogenetic metrics in three empirical examples
351 coincide with the macroevolutionary dynamics inferred using explicit modelling
352 diversification approaches. In particular, we implemented the GeoSSE model to
353 estimate the three parameters (speciation, extinction, and dispersal) between two
354 areas in each taxonomic group (Table 3 and 4). In addition, we used the BAMM
355 approach to generate the per-species level diversification metric implemented in
356 the software BAMM 2.5.0 (Rabosky 2014). In the following section, we discuss
357 each metric and we compare them with the explicit diversification approaches.

358

359 *Residual Phylogenetic Diversity (rPD)*

360 In the case of furnariid birds, we show that forest areas tend to exhibit
361 slightly higher values of rPD in contrast with open areas (Figure 4; see also Figure
362 2). According to Davies and Buckley's logic, these areas exhibit slow
363 diversification and frequent dispersal from open areas. Pinto-Ledezma et al.
364 (2017) using GeoSSE and BAMM approaches indicated that open areas exhibit
365 higher net diversification rates than open areas (Table 3). For hylid frogs, we found
366 that tropical areas tend to exhibit higher rPD values than extratropical regions
367 (Figure 5). However, by adopting an explicit diversification approach (GeoSSE and
368 BAMM), we found that net diversification rates were similar in both regions (Table
369 3). In the case of *Anolis* lizards, the rPD values were higher in the continent than in
370 the island areas (Figure 6). However, using GeoSSE and BAMM, we found that
371 both rates were similar (Table 3). In a recent paper, Poe et al. (in press) also found
372 that macroevolutionary rates are similar between insular and mainland clades. All
373 these results suggest that rPD likely does not provide an accurate signature of the
374 macroevolutionary dynamic at spatial scales. In fact, it seems that rPD tends to
375 overestimate differences between regions when a stationary diversification
376 process is occurring across geography. A potential solution might be rethinking
377 the way in which we visualize rPD across geography in contrast with the original
378 meaning by Davies & Buckley (2011; see also Forest et al. 2007).

379

380 *Mean root distance (MRD)*

381 As we discussed, *MRD* captures a total diversification value portraying the
382 number of cladogenetic events co-occurring in a given region. In the case of

383 furnariid birds, we found that MRD values tend to be higher in open than forest
384 areas (Figure 4). Accordingly, this metric suggests that more cladogenetic events
385 were accumulated in open areas (i.e., more total diversification; Rabosky 2009).
386 Therefore, this metric, for this bird clade, is consistent with results from explicit
387 diversification approaches (Table 3). For hyliid frogs, it seems that there are no
388 differences in MRD values between extratropics and tropics areas (Figure 5).
389 However, tropical areas have some cells with very high values. Again, MRD
390 provide an accurate description of the total diversification pattern in this clade
391 across the latitudinal gradient. In *Anolis* lizards, we found that MDR values tend to
392 be lower in islands in comparison with mainland areas (Figure 6). In this case,
393 MRD did not provide an accurate description of the evolutionary processes
394 occurring between the mainland and insular anole assemblages. However, there is
395 also a high probability that the high MRD values in the mainland are a direct reflect
396 of an idiosyncratic evolutionary trajectory of each one of the two clades that
397 radiated there (i.e., *Draconura* and *Dactyloa* clade; see Poe et al. 2017, Velasco et
398 al. 2018). These two clades seems to exhibit differential diversification dynamics
399 across geography (Velasco et al. 2018) but further research might be necessary to
400 evaluate these differences.

401

402 *Phylogenetic species variability (PSV)*

403 The PSV metric provides information about how related are the species in a
404 given regional assemblage. In hyliid frogs, we found that tropical assemblages
405 tend to be composed of more related species than extratropical assemblages

406 (Figure 5). Hylid assemblages in extratropical areas are composed of multiple
407 lineages that dispersed from tropical areas and then diversified there. We found
408 higher dispersal rates from tropical to temperate regions than vice versa (Table 3
409 and 4). The same tendency is present in the case of furnariid birds where open
410 areas exhibit higher PSV values than forest areas (Figure 4) and dispersal rates
411 were higher from open to forest areas than the reverse (Table 3). By contrast, we
412 did not find any evidence for differences in PSV values between island and
413 mainland *Anolis* assemblages (Figure 6). In addition, the dispersal rates were very
414 low between these two regions (Table 3; Poe et al. 2017). All these results confirm
415 that the PSV metric can provide some insights about how dispersal process have
416 shaped regional assemblages. We find evidence that low PSV values (i.e.,
417 phylogenetically over-dispersed faunas) are influenced by multiple dispersals
418 along its evolutionary history.

419

420 *Mean diversification rate (MDR)*

421 Jetz et al. (2012) proposed MDR metric as a species-level speciation rate
422 metric based in the branch length along the path from the root of a tree to each
423 individual species. In furnariid birds, we noted that MDR was slightly higher in
424 open versus forest areas and the same pattern is present using the BAMM
425 approach (Pinto-Ledezma et al. 2017; Figure 5; Table 3). For hylid frogs,
426 extratropical regions tend to exhibit higher values than tropical regions (Figure 5).
427 MDR seems to capture well the differences in macroevolutionary diversification for
428 these taxa along the latitudinal diversity gradient. A similar pattern is present when

429 the BAMM approach is used (Table 3). We consider that both metrics (MDR vs
430 per-species diversification rate from BAMM) leave the same signature in the
431 geography. In *Anolis*, we found that insular assemblages tend to exhibit higher
432 MDR values than continental assemblages (Figure 6). However, there is no
433 difference in the macroevolutionary dynamic between these two areas for the
434 *Anolis* lizards clade (Poe et al. in press, Velasco et al. 2018).

435

436 *Mean ages (MA)*.

437 The average of ages of co-occurring lineages are used to test evolutionary
438 hypothesis about whether a region maintains older lineages than others (e.g., a
439 museum) or a combination of old and recent lineages (e.g., OTT hypothesis, Table
440 1). Although this metric does not provide any inference of the ancestral area of the
441 clade, it is possible to implement an explicit biogeographic approach to test this
442 (see below). For example, in hylid frogs, we found that extratropical areas are
443 composed of older lineages than tropical regions (Figure 5). The biogeographic
444 parametric approach infers this same area as ancestral for the entire lineage
445 (Figure A2). In furnariid birds, mean ages metric revealed that older lineages have
446 accumulated more in forest than open areas (Figure 4). In accordance, the
447 ancestral area inferred with a parametric biogeographic method was the forest
448 area (Figure A3). In the case of the anole lizards, insular settings tend to be
449 composed of older lineages than continents. However, the ancestral area for the
450 entire anole clade is the mainland, particularly South America (Poe et al. 2017).
451 The mainland *Anolis* radiation is composed of two clades, one clade that

452 originated in South America (the *Dactyloa* clade; Poe et al. 2017) and colonized
453 Caribbean islands, and the other clade (the *Norops* clade; Poe et al. 2017) that
454 originated in the Caribbean islands and then colonized back the mainland in
455 Middle America and then dispersed to South America. Therefore, the
456 biogeographical history of the *Anolis* radiation is complex and involves multiple
457 dispersals between islands and mainland areas (Poe et al. 2017; Figure A4). In
458 general, mean ages does not provide enough information about the biogeographic
459 origin and maintenance of a clade. This happens because multiple dispersals and
460 *in situ* cladogenesis might erase any simplistic pattern elucidated for this metric,
461 as found in the case of the *Anolis* lizards.

462

463 **HOW DISPERSAL AND EXTINCTION AFFECT INFERENCES OF**

464 **GEOGRAPHICAL DIVERSIFICATION GRADIENTS?**

465 Dispersal is another key macroevolutionary process that ultimately determines the
466 number of a species in a region (Roy and Goldberg 2007, Eiserhardt et al. 2013,
467 Rolland et al. 2014, Chazot et al. 2016). However, few studies evaluated explicitly
468 how the direction of dispersals between region contributes to the generation of
469 regional differences between areas (Chown and Gaston 2000, Goldberg et al.
470 2005, 2011, Jablonski et al. 2006). Roy and Goldberg (2007) showed with
471 simulations that dispersal asymmetry between areas had a strong impact in the
472 regional species richness and the average age of these lineages. Accordingly,
473 phylogenetic metrics can be sensitive to dispersal between areas because it is
474 impossible to distinguish which lineages originated by *in situ* speciation or simply

475 due to dispersal from nearby areas. Goldberg et al. (2011) developed the GeoSSE
476 model to evaluate how range evolution affected diversification rates in a
477 phylogenetic comparative approach. The GeoSSE model only considers three
478 states (A: endemic species to a region; B: endemic species to another region; and
479 AB for widespread species) and makes a series of assumptions that can be
480 problematic. The first assumption of the GeoSSE model is that a time-dependent
481 process dominates the diversification dynamic in each region (Stephens and
482 Wiens 2003, Wiens 2011). This assumption conflicts with a diversity-dependent
483 process assumption and this debate is far from being resolved (Cornell 2013,
484 Harmon and Harrison 2015, Rabosky and Hurlbert 2015). The second problematic
485 assumption has to do with the fact that the GeoSSE model consider dispersal
486 rates as stable through time and lineages. In other words, the dispersal ability and
487 therefore the frequency of transitions between areas are constant across the
488 evolutionary history of a clade. There are many empirical evidence showing that
489 dispersal rates vary across time and space among lineages (McPeck and Holt
490 1992, Sanmartín et al. 2008, Robledo-Arnuncio et al. 2014).

491 Regardless of these major assumptions, the GeoSSE model has been
492 adopted to evaluate relative contributions of speciation, extinction and dispersal
493 to the generation of species richness gradients (e.g., Rolland et al. 2014, Pyron
494 2014b, Staggemeier et al. 2015, Looney et al. 2016, Morinière et al. 2016, Pulido-
495 Santacruz and Weir 2016, Alves et al. 2017, Hutter et al. 2017, Pinto-Ledezma et
496 al. 2017). In a recent study, Rabosky and Goldberg (2015) found that state-
497 dependent diversification models tend to inflate excessively the false discovery

498 rates (i.e., type I error rates). In particular, Rabosky and Goldberg (2015) found
499 that these models tend to find false associations between trait shifts and shifts in
500 macroevolutionary dynamics. Although the Rabosky and Goldberg's study was
501 not based on the GeoSSE model, it is clear that transitions between areas (i.e.,
502 dispersal events) can be falsely associated with shifts in speciation and extinction
503 rates across the phylogeny. Alves et al. (2017) also found that geographical
504 uncertainties in the assignment of species to a given area affect the parameter
505 estimates (i.e., speciation, extinction and dispersal rates) in the GeoSSE model.
506 Same authors also evaluated how incorrect assignments of bat species to tropical
507 or extra-tropical regions can generate erroneous conclusions about the relative
508 role of speciation, extinction and dispersal on a latitudinal diversity gradient. From
509 these studies, it is clear that dispersal is a major issue that needs to be evaluated
510 explicitly in macroecological studies.

511 Pulido-Santacruz and Weir (2016) also used the GeoSSE model to disentangle
512 the relative effect of speciation, extinction and dispersal on the latitudinal bird
513 diversity gradient. They found that extinction was prevalent across all bird clades
514 and therefore they suggest this as a main driver of the geographical bird diversity
515 gradient. Pyron (2014c), also using the GeoSSE model, found that temperate
516 diversity in reptiles is due to higher extinction in these areas. We consider that
517 extinction inferences from the GeoSSE model should be treated with caution. For
518 the few clades where fossil record is abundant (e.g., marine bivalves; Jablonski et
519 al. 2006), studies point out to conclude that extinction differences between
520 regions should be treated with caution due to the potential sampling bias

521 (Jablonski et al. 2006, 2017). In addition, studies based on extensive simulations
522 found that extinction inferences based only in molecular phylogenies are not
523 reliable (Rabosky 2010a, 2016b, Quental and Marshall 2010), although extinction
524 rates can be estimated relatively well using medium to large phylogenies (Beaulieu
525 & O'Meara 2015).

526 In a recent review, Sanmartín and Meseguer (2016) proposed that it is possible
527 to detect the extinction signature in molecular phylogenies using extensive
528 simulations and lineage-through-time –LTT- plots (see also Antonelli and
529 Sanmartín 2011). These authors also found that many birth-death models leave a
530 similar phylogenetic imprint, which make indistinguishable some scenarios. In
531 addition, extinction events can affect substantially the ancestral range estimates,
532 and therefore dispersal and extinction parameters in several parametric
533 biogeographic methods (e.g., Dispersal-Vicariance –DIVA- and Dispersal-
534 Extinction-Cladogenesis –DEC- models; Ronquist 1997, Ree et al. 2005).
535 Sanmartín and Meseguer (2016) finally proposed that the adoption of a
536 hierarchical Bayesian approach using continuous-time Markov Chain models will
537 allow a better estimation of extinction both in geography and in the phylogeny
538 (Sanmartín et al. 2008, Sanmartin et al. 2010).

539 Recently, Rabosky and Goldberg (2017) developed a semi-parametric
540 method (FiSSE) to correct the statistical problems found in BiSSE models by
541 themselves in a previous paper (Rabosky and Goldberg 2015). However, the
542 FiSSE method does not allow the evaluation of the contribution of dispersal on
543 regional species richness. In any case, the best suitable framework to estimate

544 relative contributions of speciation, extinction and dispersal might be the GeoSSE
545 model (or parametric biogeographic models; e.g., Matzke 2014; see below),
546 although it requires the simulation of a series of null scenarios to evaluate the
547 statistical power in each case (see Alves et al. 2017, Pinto-Ledezma et al. 2017 for
548 a few examples). For instance, Pinto-Ledezma et al. (2017) developed a
549 parametric bootstrapping approach simulating traits to evaluate whether empirical
550 inferences are different from the simulated. They simulated 100 datasets of neutral
551 characters along a set of empirical phylogenies and using this new information
552 repeated the same procedure with empirical data (see Appendix S1 in Pinto-
553 Ledezma et al. 2017 for details of the bootstrapping approach). This
554 bootstrapping procedure assumes no direct effect of the geographic character
555 states on the parameter estimations (Feldman et al. 2016, Pinto-Ledezma et al.
556 2017).

557 Finally, it should be clear that more research would be necessary to
558 establish how extinction affect estimation parameters in state-dependent
559 diversification approaches (e.g., the GeoSSE model). For instance, the inclusion-
560 exclusion of extinct species in simulated phylogenies using birth-death models
561 could substantially affect the geographical inferences of speciation, extinction and
562 dispersal parameters in the GeoSEE model. This kind of approach might provide
563 some lights on how to biased can be the parameter estimates with only molecular
564 phylogenies using the GeoSSE model or any other modeling approach.

565

566 ***Parametric biogeographical approaches in macroecological studies.***

567 The use of parametric biogeographic approaches is an optimal solution to
568 estimate dispersals across time and space (Matzke 2014, Dupin et al. 2017).
569 These methods are promising in identifying the relative roles of cladogenetic and
570 anagenetic processes shaping regional species richness. Recently, Dupin et al.
571 (2017) developed a biogeographical stochastic mapping to infer the number of
572 dispersals, and other biogeographical events, in the evolutionary history of
573 Solanaceae plants across the world. This approach allows the inference from
574 multiple process including sympatric speciation, allopatric speciation, founder-
575 event speciation, range expansion (i.e., dispersal without speciation) and local
576 extinction (i.e., range contractions) based on a time-calibrated phylogenetic tree
577 and the occurrence of species in geographical regions (see also Matzke 2014 for
578 more detailed description of the method). These explicit biogeographical
579 approaches are promising in macroecological studies since they allow to test
580 simultaneously a set of evolutionary process during the diversification of a clade in
581 a region (Velasco 2018). In addition, with these new approaches it is possible to
582 differentiate effectively between macroevolutionary sources and sink areas
583 (Goldberg et al. 2005, Castroviejo-Fisher et al. 2014, Poe et al. 2017). For
584 instance, Poe et al. (2017) used a parametric biogeographical approach to
585 estimate the number of events among regions and distinguish those areas where
586 many cladogenetic events occurred (i.e., *in situ* speciation) and areas where
587 almost all its diversity was build-up from extensive colonization of other regions.

588 The biogeographical stochastic mapping (BSM) method developed by
589 Dupin et al. (2017) is promising to estimate more accurately the number of

590 dispersal events between regions based on a better estimation of the ancestral
591 area for a clade. We evaluated how dispersal rates between regions can affect
592 inferences drawn only from phylogenetic metrics in our three data sets. We
593 implemented GeoSSE and BSM approaches for each data set (Table 3 and 4). For
594 the case of hylid frogs, we counted the inferred number of dispersal events
595 between tropical and extra-tropical regions in the Americas (Table 3; see also
596 (Wiens et al. 2006, Algar et al. 2009). For furnariid birds, we counted the number
597 of dispersal events between open and forest areas (Table 3; see also Pinto-
598 Ledezma et al. 2017). Finally, for anole lizards, we counted the number of
599 dispersal events between insular and mainland areas (Table 3; see also Algar and
600 Losos 2011, Poe et al. 2017, Velasco et al. 2018).

601 Using biogeographical stochastic mapping –BSM–, we inferred the number
602 of dispersal events from one region to another for each one of the three
603 taxonomic groups examined (Table 3). The BSM approach allows us to
604 disentangle which dispersals were only range expansions and which dispersals
605 generated a speciation event (i.e., a founder-event speciation; (Barton and
606 Charlesworth 1984, Templeton 2008). For furnariid birds, we found that range
607 expansions were three times higher from forest areas to open areas than the
608 reverse and founder events were twice higher from forest to open areas than the
609 opposite (Table 3). This result suggests that differences in species richness
610 between forest and open areas are due by recurrent dispersal events along the
611 furnariid diversification history (Pinto-Ledezma et al. 2017). Pinto-Ledezma et al.
612 (2017) found a similar result using the GeoSSE approach, but they conducted a

613 parametric simulation approach to evaluate whether there was a direct effect of
614 the geographic location on the parameter estimates. Their results show that the
615 GeoSSE approach, in this case, had limited power to detect a signature of
616 geographic region on speciation, extinction and dispersal rates. With the
617 implementation of the BSM approach here, we corroborate Pinto-Ledezma et al.'s
618 findings with improved statistical power. In the case of hylid frogs and the
619 transitions between tropical and extra-tropical areas, we found that the BSM
620 approach inferred more dispersal events from tropical to extra-tropical regions
621 (Table 3 and 4). However, the number of founder events was relatively low in
622 comparison with range expansions (Table 3). These results suggest that few
623 dispersal events have occurred across the diversification of hylid frogs and
624 corroborate that the species richness in each region largely originated by *in situ*
625 speciation modulated by climatic factors (Wiens et al. 2006, Algar et al. 2009).
626 Finally, for *Anolis* lizards, we found that dispersal events between insular and
627 mainland regions were relatively low (Table 3 and 4). We did not find evidence of
628 any expansion range events from mainland to island or vice versa. This also
629 corroborates previous findings that evolutionary radiation of anole in insular and
630 mainland settings is due to extensive *in situ* diversification (Poe et al. in press,
631 2017, Algar and Losos 2011).

632 These results point out that the BSM approach (Dupin et al. 2017) is a
633 promising approach when we are interested in testing the role of anagenetic and
634 cladogenetic events on the resulting geographical species richness gradients.
635 Although parametric biogeographic approaches are still in their infancy (Sanmartín

636 2012, Matzke 2014, Dupin et al. 2017), these methods allow us to evaluate
637 macroevolutionary dynamics (i.e., speciation and extinction) in an explicit
638 geographical context. These methods are statistical powerful and make use of a
639 series of explicit geographic range evolution models (Velasco 2018).

640

641 ***TOWARD AN INTEGRATION OF BIOGEOGRAPHICAL AND SPECIES***

642 ***DIVERSIFICATION APPROACHES IN MACROECOLOGICAL STUDIES.***

643 Although different parametric biogeographic methods have been
644 developing at least for the last 20 years (Ronquist 1997, Ree et al. 2005, Landis et
645 al. 2013, Matzke 2014, Dupin et al. 2017), the adoption of these methods to test
646 evolutionary-based hypotheses underlying geographical diversity gradient has
647 been rare. For instance, few studies examined here tested the effect of dispersal
648 events in the generation of regional species richness assemblages. It should clear
649 that the current paradigm in biogeography makes a call for an evaluation of the
650 relative frequency of cladogenetic and anagenetic process during the
651 biogeographical history of lineages. The adoption of parametric approaches in
652 future macroecological studies will contribute to an improvement of the estimation
653 of speciation, extinction and dispersal processes as drivers of the geographical
654 diversity gradients. In addition, we also think that it is necessary to establish which
655 macroevolutionary dynamics govern regional assemblages. Phylogenetic
656 approaches based on fitting diversification models help to test whether regional
657 species richness is due to diversity dependence (i.e., ecological limits), time
658 dependence, or environmental factors (Rabosky and Lovette 2008, Etienne et al.

659 2012, Etienne and Haegeman 2012, Condamine et al. 2013). We also stress that
660 the adoption of many approaches providing multiple lines of evidence will help to
661 disentangle the evolutionary and ecological causes of biodiversity gradients.
662 Some recent studies have pointed toward this strategy and have begun to provide
663 evidence from many lines to understand how evolutionary processes underlying
664 species richness gradients works (Hutter et al. 2017, Pinto-Ledezma et al. 2017).
665

666 ***Conclusions and recommendations***

667 The resulting geographical pattern of several phylogenetic metrics did not
668 provide any robust evidence of a spatially explicit diversification dynamic. As we
669 have shown, these resulting geographical patterns did not differ from that
670 generated by a simple null model. It is hard to untangle causal mechanisms (i.e.,
671 speciation, extinction, and dispersal) from only the geographical signature that
672 these metrics attempt to capture. We recommend that phylogenetic metrics
673 should be used only to visualize geographical patterns of total diversification (e.g.,
674 MRD, residual PD; MDR), phylogenetic structure (e.g., PSV), or mean ages of co-
675 distributed species (e.g., MA) (Table 1). We suggest that conclusions about the
676 role of evolutionary processes in the generation and maintenance of species
677 richness gradients based only in these phylogenetic metrics should be avoided
678 and additional approaches always should be used.

679 Some explicit diversification approaches (e.g., model fitting approaches;
680 (Etienne et al. 2012, Rabosky 2014, Valente et al. 2015) are useful to establish the
681 macroevolutionary dynamics operating at regional scales. Although some

682 approaches (e.g, the GeoSSE model) allow us to evaluate the relative role of the
683 ultimate process that modify the regional species diversity, its statistical power
684 (e.g., high Type I errors) has been challenged by simulation and empirical studies.
685 Furthermore, the extinction and dispersal estimates inferred by the GeoSSE model
686 tend to be unbiased. Parametric biogeographic approaches are becoming a
687 standard tool to evaluate how evolutionary processes can explain the
688 geographical distribution of extant taxa. These approaches are promising and
689 should be extensively used because allow us to estimate the relative frequency of
690 cladogenetic and anagenetic process shaping the regional species richness.

691 It is necessary that macroecological studies use a combination of explicit
692 diversification approaches and parameter biogeographic methods with the aim to
693 clarify how evolutionary process have shaped regional species richness
694 assemblages. As Jablonski et al. (2017) have outlined, one of the main obstacles
695 to generate an appropriate understanding of the causal mechanisms underlying
696 geographical diversity gradients has been that many studies have tested a single
697 hypothesis, either evolutionary or ecological, as an explanatory factor. We suggest
698 that ecological and evolutionary hypotheses should be tested simultaneously to
699 explain the relative contribution of each process to the regional diversity. As
700 shown by our empirical comparison of phylogenetic metrics, explicit
701 diversification models, and historical biogeographic methods have showed, it is
702 necessary to obtain evidence of different approaches to guarantee sound
703 conclusions about the evolutionary causes of these biodiversity gradients.

704

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713

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1011 Diversity Gradient in Hylid Frogs: Treefrog Trees Unearth the Roots of High
1012 Tropical Diversity. - Am. Nat. 168: 579–596.
- 1013

1014 Table 1: Phylogenetic metrics and explicit diversification approaches used in
 1015 macroecological studies to address evolutionary questions related with the
 1016 geographical diversity gradients.

Metric	Author	Description	Software / R package
MRD	Kerr and Currie 1999	MRD is calculated by counting the number of nodes separating each terminal species in a regional assemblage or cell from the tips to root of the phylogenetic tree. This metric does not need that trees be ultrametric or have branch lengths.	metricTester
PD (residual)	Faith 1992	PD is calculated by summing all the branch lengths of species co-occurring in a regional assemblage or cell. Residual PD is obtained from an ordinary least square regression between PD and species richness.	picante, metricTester, pez
PSV	Helmus et al. 2007, Algar et al. 2009	PSV is calculated from a matrix where their diagonal elements provide the evolutionary divergence (based on the branch lengths) of each terminal species from the root to the tips of the tree, and the off-diagonal elements provide the degree of shared evolutionary history among species. Values close to zero indicates that all species in a regional assemblage or cell are very close related whereas values close to one indicate that species are not related.	picante, metricTester, pez
mean DR	Jetz et al. 2012	DR is calculated as the inverse of a measure of evolutionary isolation (Redding & Mooers 2006) which sum all the edge lengths from a species to the root of the tree. The inverse of this evolutionary isolation metric therefore capture the level of splitting rate of each species (i.e., its path to a top).	FISSE
Mean age	Latham and Ricklefs 1993	The mean age of co-occurring species in a regional assemblage or cell simply is calculated tallying the age of each most recent common ancestor (MRCA) for each species and the averaged.	None
GeoSSE	Goldberg et al. 2011	The geographic state speciation and extinction -GeoSSE- model is a trait-dependent diversification method linking geographic occurrence with diversification rates. These method allow to infer both speciation and extinction rates as movement (dispersal) rates among two regions.	Diversitree R package
BAMM	Rabosky 2014	BAMM is a method that attempt to identify whether a phylogeny exhibit a single or various macroevolutionary regimes (i.e., different diversification dynamics). As speciation, extinction and net diversification rates are considered to be heterogeneous across the phylogeny it is possible to estimate a rate for each branch or species in the tree.	BAMM software and BAMMtools R package

1017

1018 Table 2. Description of some evolutionary hypothesis tested in macroecological studies as causal mechanisms of
 1019 regional species richness.

Hypothesis	References	Description	Predictions	Metrics and/or methods used to test	Limitations
Phylogenetic niche conservatism (PNC)	Wiens and Graham 2005	Phylogenetic niche conservatism is the tendency of related species to inherit niche requirements from its the most recent common ancestors (Wiens & Graham 2005).	PNC predicts that regions where a clade originated will accumulate more species simply due to more occupation time and diversification rates tend to be similar between regions. The tropical niche conservatism hypothesis (TNC; Wiens and Donoghue 2004) is based on PNC to explain differences in species richness in tropical and temperate regions.	MRD, Mean age, GeoSSE, BAMM	1) MRD metric fails to capture spatially dynamics of the balance of speciation and extinction and it is very hard to establish whether species richness in a region is only generated by higher speciation rates. Furthermore, MRD does not capture dispersal dynamic across regions and species richness in a given region can be generated from only dispersals from nearby regions (e.g., macroevolutionary sinks; CITA). 2) Mean age provide partial is able to test the role of PNC on geographical species richness because only it is possible to establish which regions have, in average, old clades and this not reflects whether many speciation events occurred there. 3) GeoSSE is potentially the only one approach that allow to disentangle these three process but it is only limited to two regions (e.g., tropical vs. temperate). In addition, GeoSSE has been criticized due its low statistical power (see Rabosky and Goldberg 2015).
Regional diversification (RD)	Buckley et al. 2010	Differences in the balance of speciation and extinction across geography can explain differences in species richness between regions.	RD predicts that regions with striking differences in species richness are due to differences in macroevolutionary dynamics between regions.	residual PD, GeoSSE, BAMM	1) Residual PD can be used to discriminate regions with rapid and slow diversification based on the expected phylogenetic diversity given species richness (Buckley et al. 2010). However, this metric ignores the contribution to dispersal to PD in a given region or cell. 2) GeoSSE can estimate speciation, extinction and dispersal rates between regions but again is limited to two regions. 3) BAMM potentially could be used to estimate speciation rates for regional clades but this method is unable to estimate dispersal rates between regions.

Out of the tropics (OTT)	Jablonski et al. 2006	Species were generated in the tropical regions and dispersed to extratropical regions but maintain its presence in its ancestral areas	High rates of speciation are predicted in tropical regions in contrast with temperate regions. Asymmetric dispersal have occurred along the biogeographical history of a taxa from tropical to temperate areas.	MRD, Mean age, GeoSSE	These metrics are the same used to test the PNC/TNC hypothesis as we discuss above.
Time for speciation effect (TEE)	Stephens and Wiens 2003	Tropical regions accumulated more species because their clades had more time to speciate than temperate regions.	Regions recently colonized had lower species richness than regions where clades colonized very early in the history of a clade.	Mean age	1) Mean age does not provide an accurate description of which lineages colonized first a region. To test this hypothesis, it might be necessary to perform an ancestral range reconstruction of all co-occurring clades and estimate its diversification rates (i.e., total diversification for each independent colonized clade; Rabosky 2009; 2012).

1021 Table 3. Parameter estimates from the GeoSSE model for three taxonomic groups
1022 (Furnariides birds, hylid frogs, and *Anolis* lizards) across two regions. Areas for each
1023 taxonomic group as follows: Furnariides birds: A: Forest; B: Open areas; Hylid frogs: A:
1024 Extra tropics; B: Tropics; *Anolis* lizards: A: Islands; B: Mainland.

Group	Rates	A	B	AB
Furnariides birds	Speciation	0.139 ± 0.020	0.223 ± 0.065	0.041 ± 0.020
	Extinction	0.040 ± 0.025	0.107 ± 0.075	-
	Dispersal	0.021 ± 0.004	0.311 ± 0.114	-
	Net diversification	0.099 ± 0.005	0.116 ± 0.01	-
Hylid frogs	Speciation	0.044 ± 0.003	0.044 ± 0.003	0.041 ± 0.025
	Extinction	0.002 ± 0.002	0.002 ± 0.002	-
	Dispersal	0.001 ± 0.001	0.035 ± 0.010	-
	Net diversification	0.042 ± 0.003	0.042 ± 0.003	-
<i>Anolis</i> lizards	Speciation	0.058 ± 0.003	0.058 ± 0.003	1.245 ± 1.303
	Extinction	0.001 ± 0.001	0.001 ± 0.001	-
	Dispersal	0.002 ± 0.001	0.0003 ± 0.000	-
	Net diversification	0.057 ± 0.002	0.057 ± 0.002	-

1025

1026 Table 4. Frequency of dispersal events inferred using biogeographical stochastic
 1027 mapping (BSM) for three taxonomic groups (Furnariides birds, hylid frogs, and *Anolis*
 1028 lizards) across two regions. Areas for each taxonomic group as follows: Furnariides
 1029 birds: A: Forest; B: Open areas; Hylid frogs: A: Extra tropics; B: Tropics; *Anolis* lizards:
 1030 A: Islands; B: Mainland.

1031

Event	Group	Regions	A	B
Range expansions	Furnariides birds	A	0	92.62 ± 4.39
		B	31.4 ± 4.29	0
	Hylid frogs	A	0	0.64 ± 0.78
		B	12.92 ± 0.88	0
	<i>Anolis</i> lizards	A	0	0
		B	0	0
Founder events	Furnariides birds	A	0	24.46 ± 2.54
		B	10.88 ± 2.22	0
	Hylid frogs	A	0	0.66 ± 0.66
		B	4.3 ± 1.42	0
	<i>Anolis</i> lizards	A	0	2.02 ± 0.14
		B	0.12 ± 0.33	0

1032

1033

1034 FIGURE LEGENDS

1035 Figure 1. Diagram illustrating how differences in speciation, extinction, and dispersal
1036 rates between regions can generate a geographical species richness gradient. The
1037 phylogenetic trees below illustrate how the differences in speciation and extinction
1038 rates between two regional assemblages can shape a gradient of species richness
1039 (degraded blue colour).

1040
1041 Figure 2. Geographical patterns of some phylogenetic metrics used in macroecological
1042 studies to explore evolutionary process underlying geographical diversity gradients
1043 (see also Table 1 for a detailed explanation). Left column *Anolis* lizards; Middle column:
1044 Hylid frogs; Right column: Furnariides birds. (A-C) observed richness patterns; (D-F)
1045 rPD: residual phylogenetic diversity (i.e., after controlling for species richness); (G-I)
1046 PSV: phylogenetic species variability; (J-L) MRD: mean root distance; (M-O) MDR:
1047 mean diversification rate; (P-R) Mean ages: average ages of species.

1048
1049 Figure 3. P-values distribution for each phylogenetic metric obtained through the null
1050 model (see main text for details). The vertical red lines represent the empirical 0.05 cut-
1051 off. Note that for all cases very few cells are below the 0.05 cut-off. (A-C) rPD: residual
1052 phylogenetic diversity (i.e., after controlling for species richness); (D-F) PSV:
1053 phylogenetic species variability; (G-I) MRD: mean root distance; (J-L) MDR: mean
1054 diversification rate; (M-O) Mean ages: average ages of species.

1055
1056 Figure 4. Variation of phylogenetic metric values for Furnariides birds in forest and
1057 open areas. rPD: residual phylogenetic diversity (i.e., after controlling for species
1058 richness); PSV: phylogenetic species variability; MRD: mean root distance; MDR: mean
1059 diversification rate; Mean ages: average ages of species.

1060
1061 Figure 5. Variation of phylogenetic metric values for Hylid frogs in tropics and extra-
1062 tropics regions. rPD: residual phylogenetic diversity (i.e., after controlling for species
1063 richness); PSV: phylogenetic species variability; MRD: mean root distance; MDR: mean
1064 diversification rate; Mean ages: average ages of species.

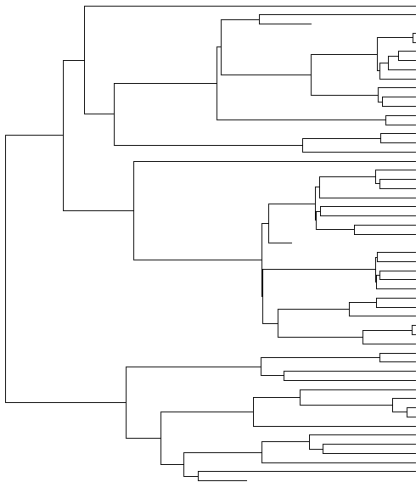
1065
1066 Figure 6. Variation of phylogenetic metric values for *Anolis* lizards in continental and
1067 insular areas. rPD: residual phylogenetic diversity (i.e., after controlling for species
1068 richness); PSV: phylogenetic species variability; MRD: mean root distance; MDR: mean
1069 diversification rate; Mean ages: average ages of species.

1070

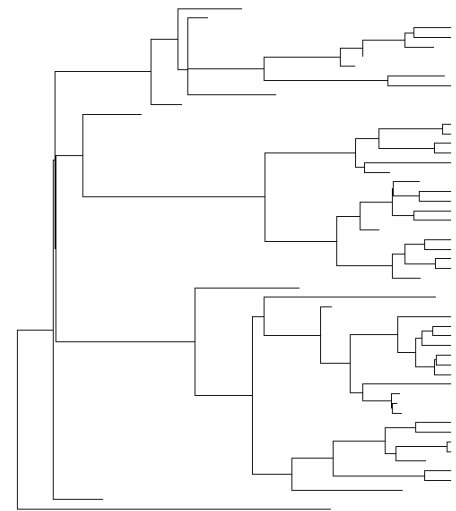
DIVERSITY GRADIENTS

Higher speciation
Lower extinction
Dispersal (out)

Lower speciation
Higher extinction
Dispersal (in)



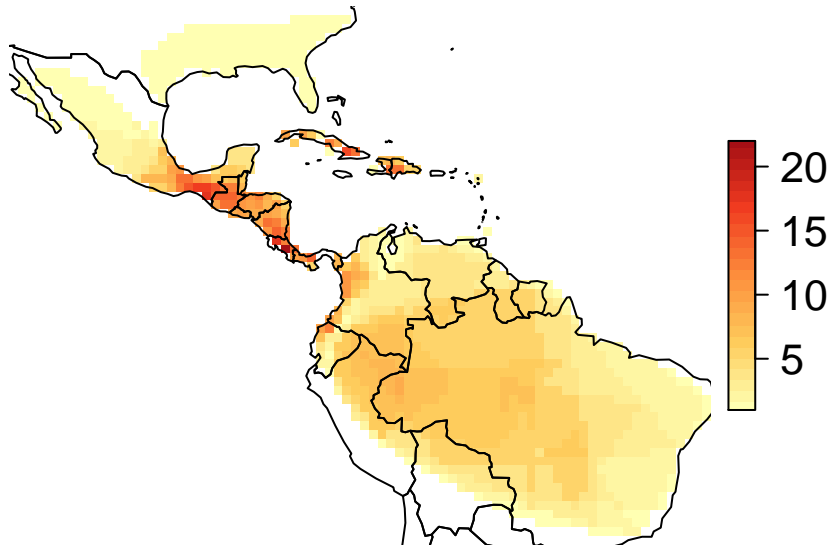
Speciation = 0.5; Extinction = 0.05



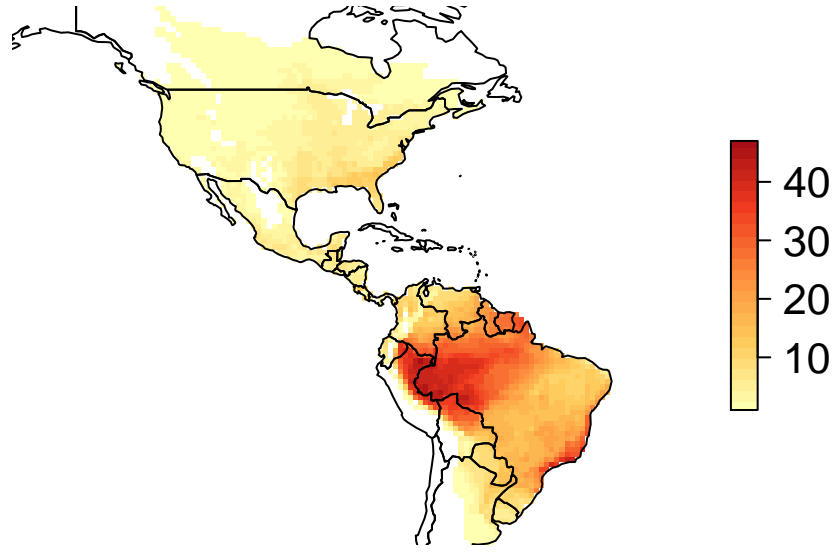
Speciation = 0.3; Extinction = 0.15

Anolis

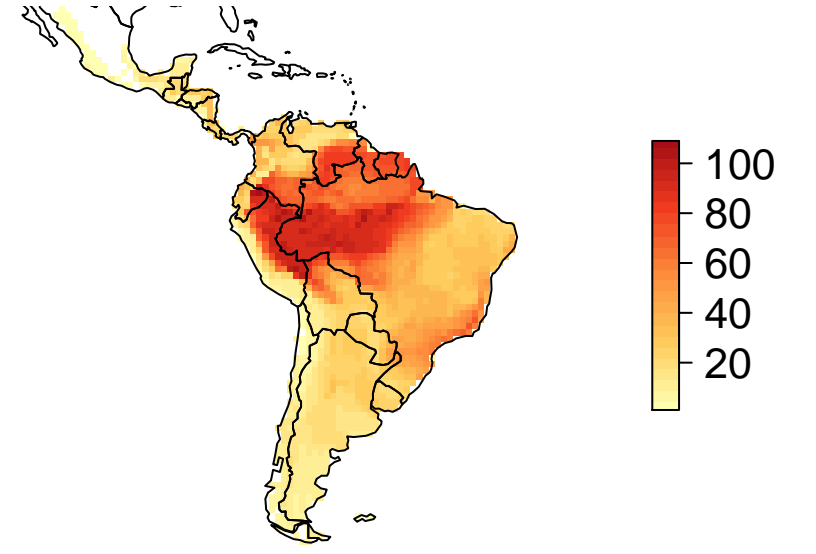
(A)

**Hylids**

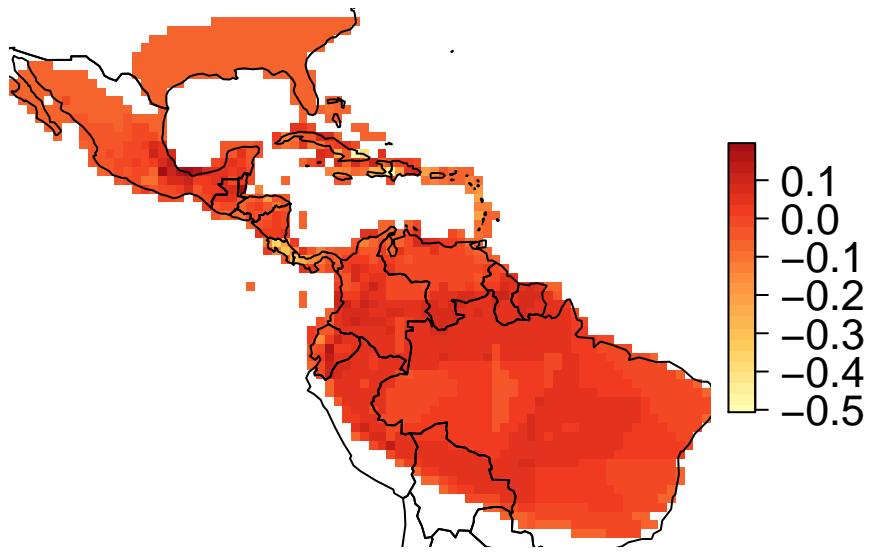
(B)

**Furnariides**

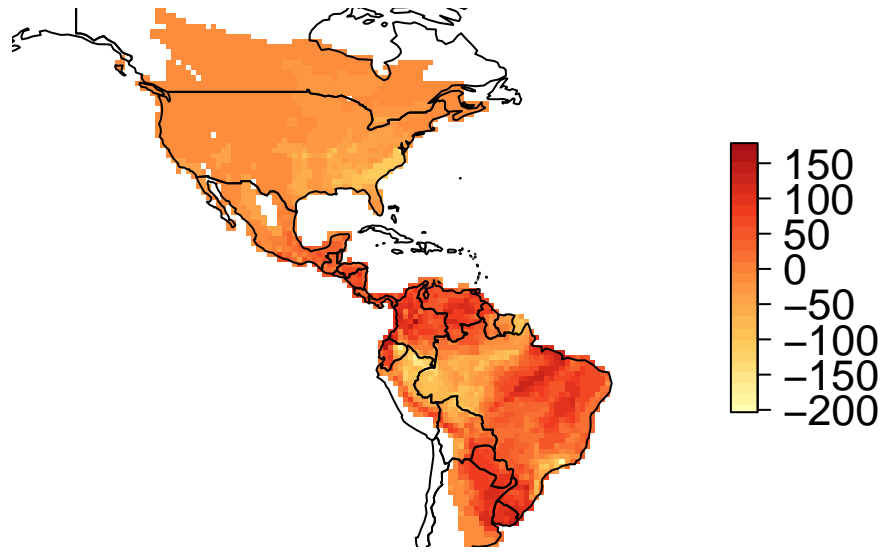
(C)



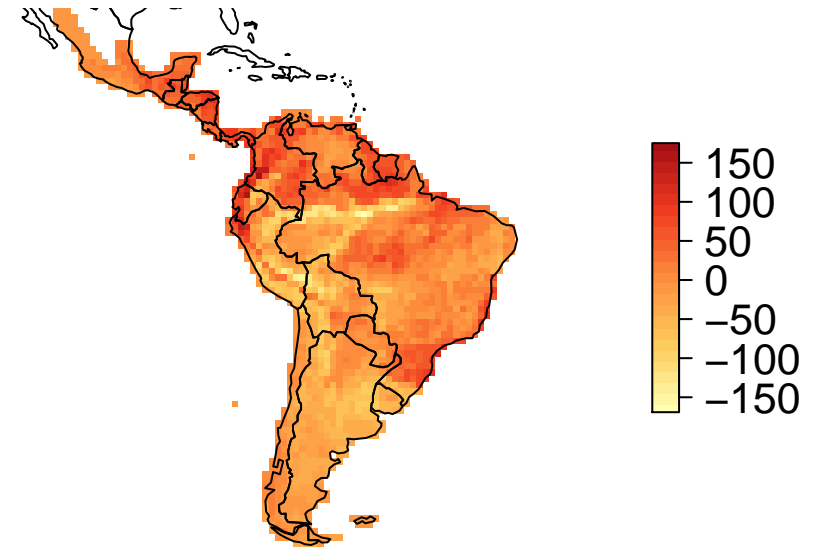
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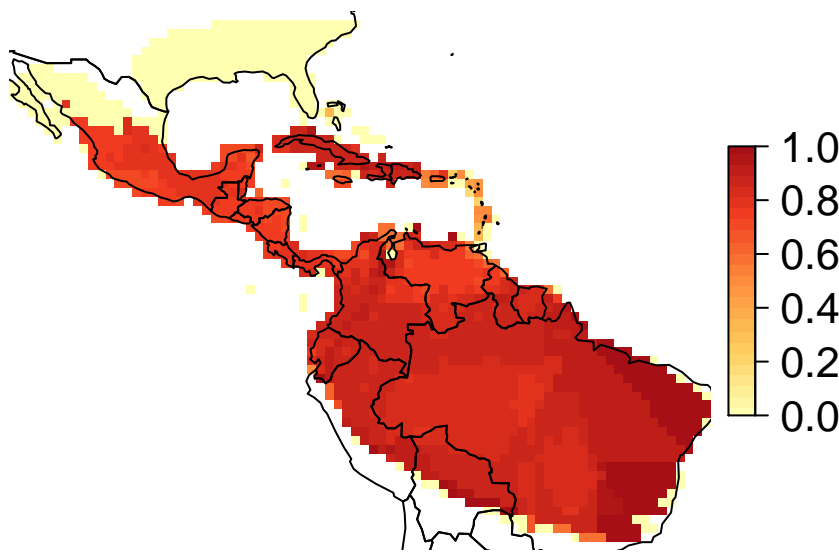
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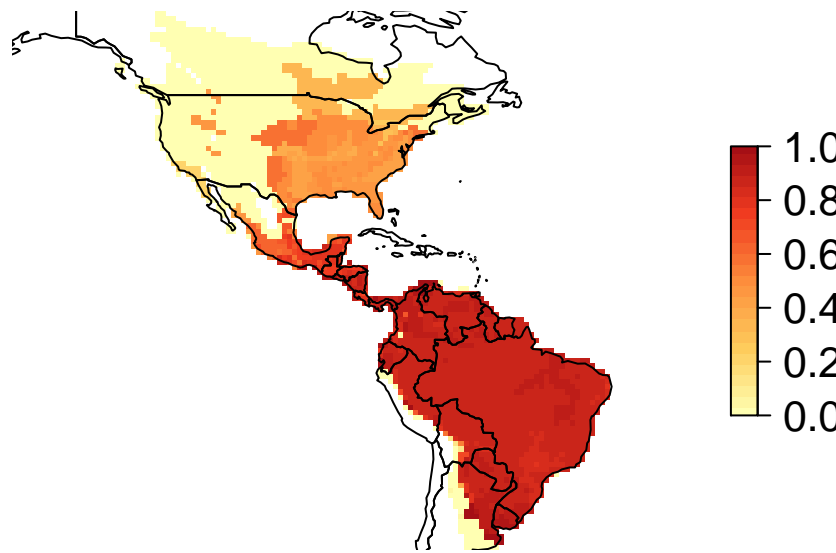
(F)



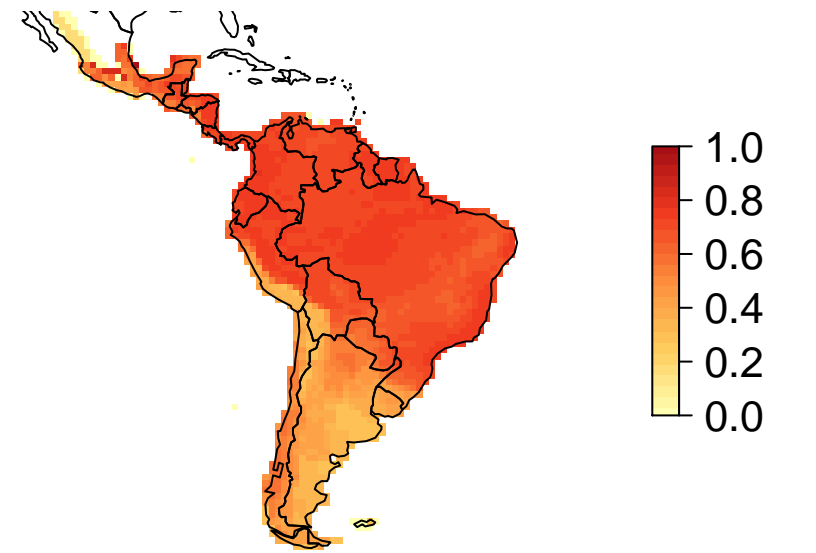
(G)



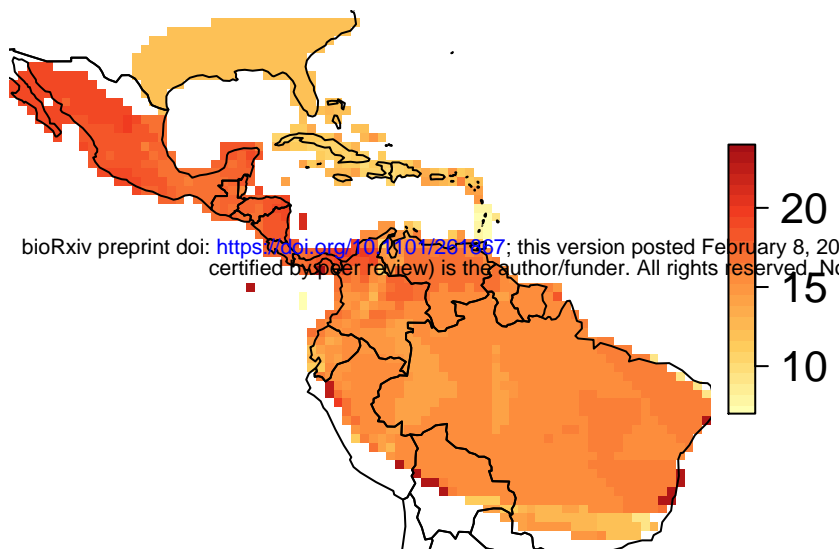
(H)



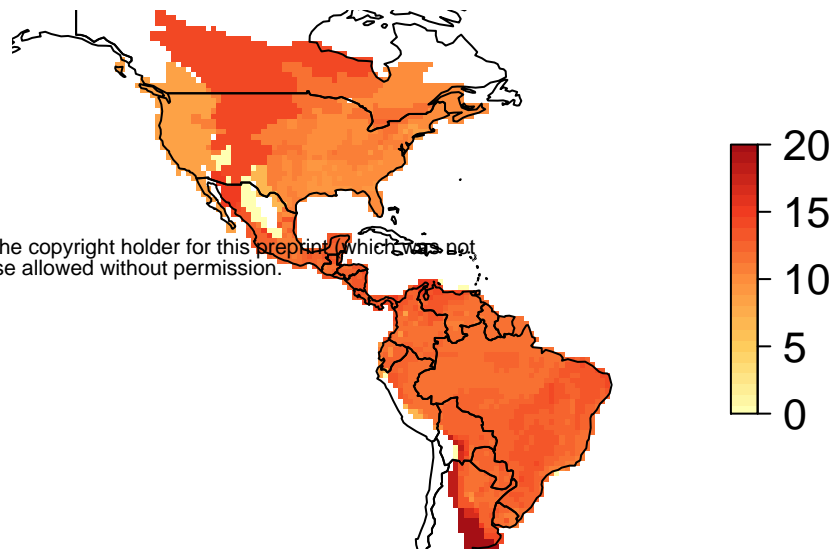
(I)



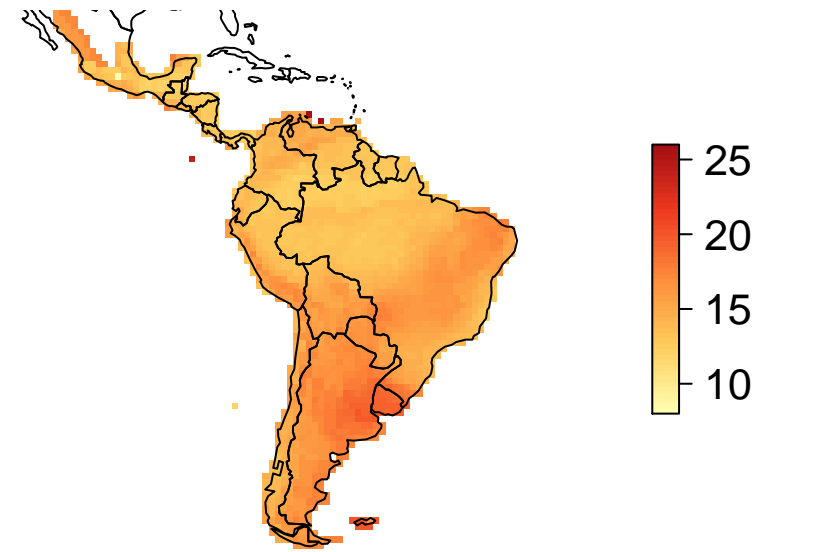
(J)



(K)



(L)

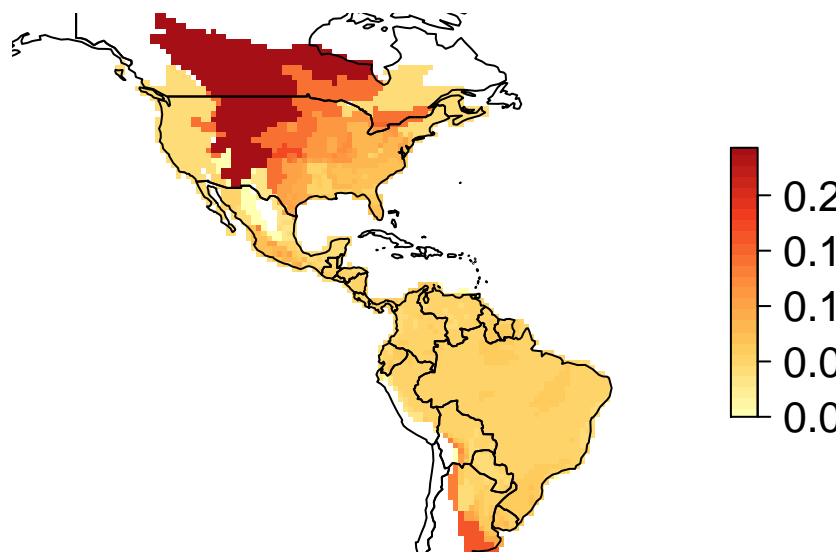


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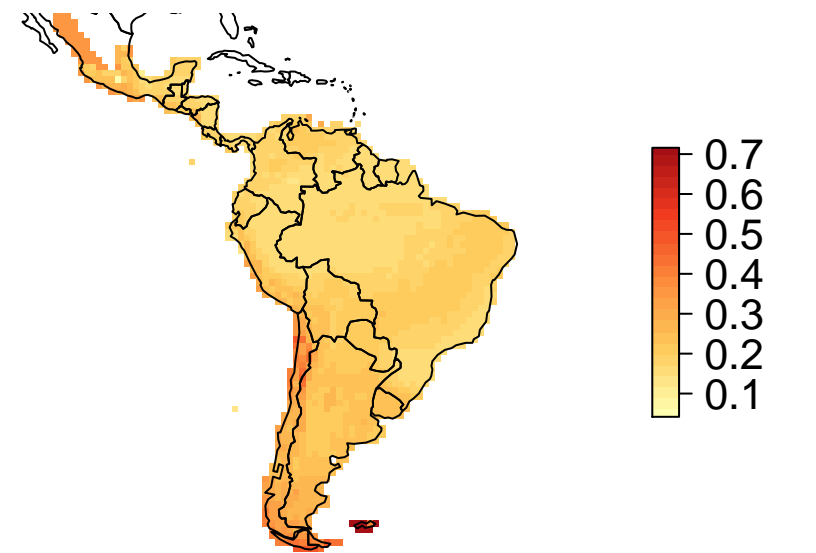
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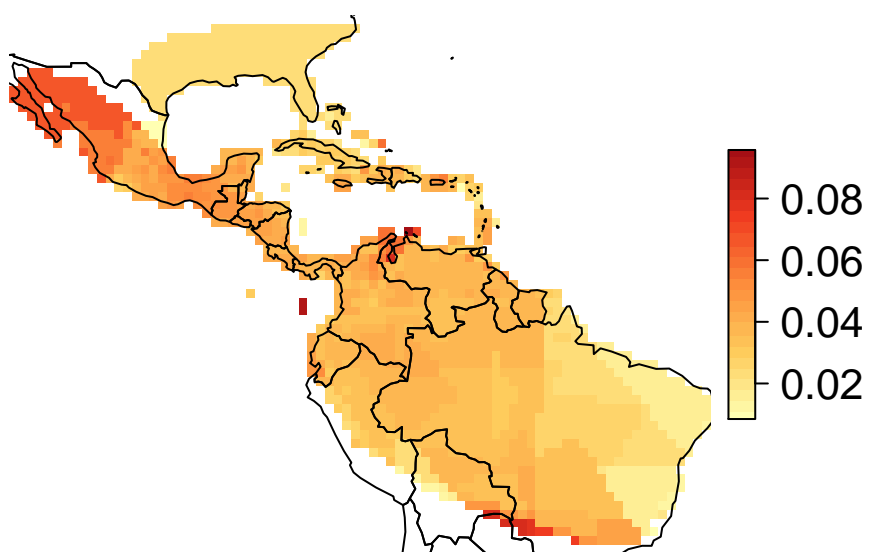
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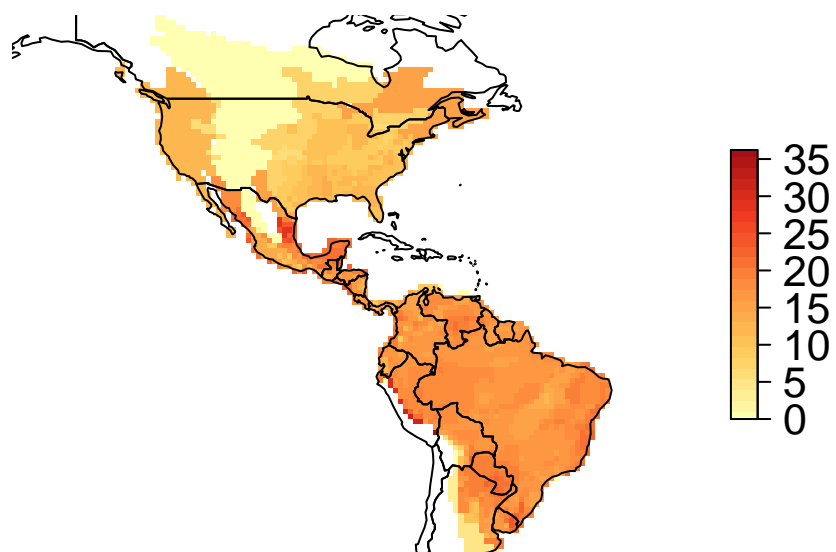
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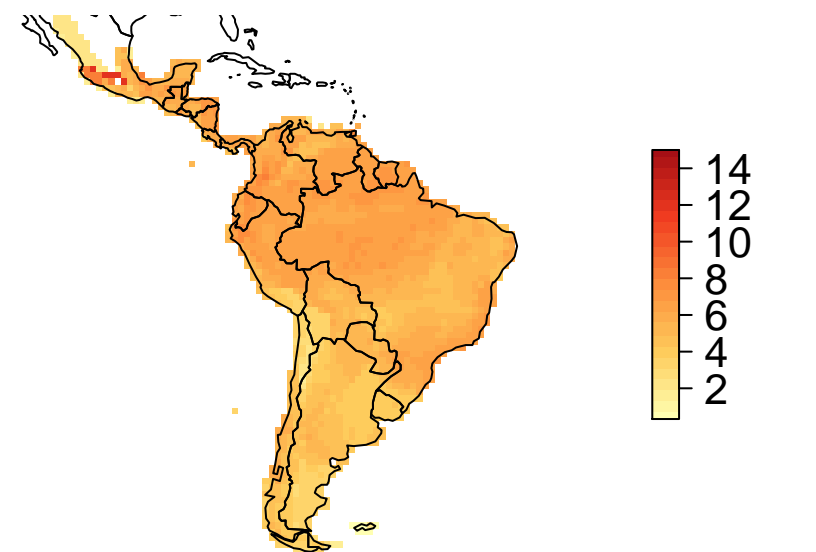
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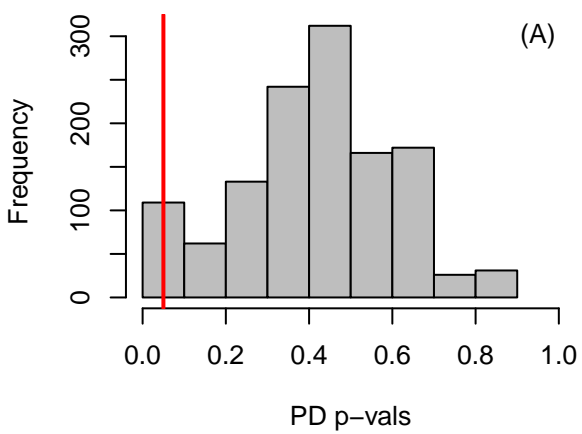
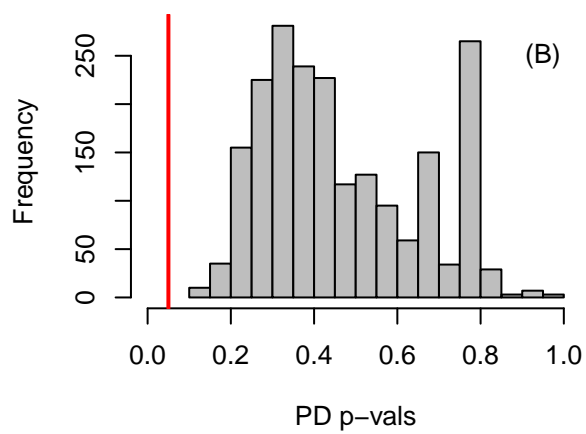
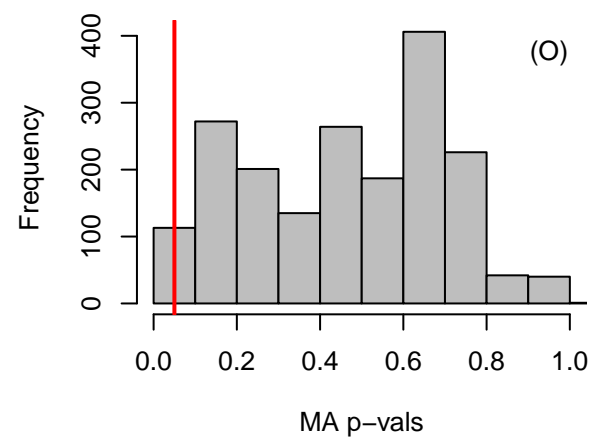
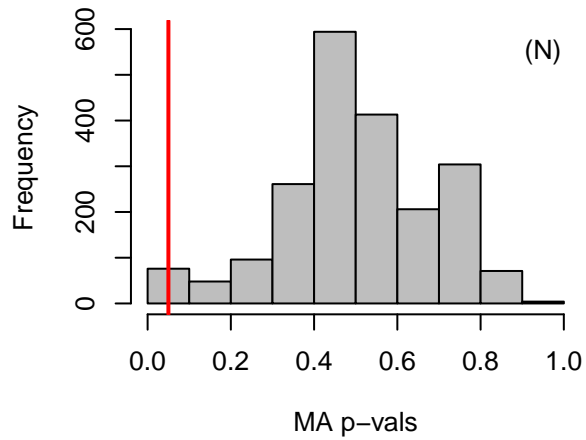
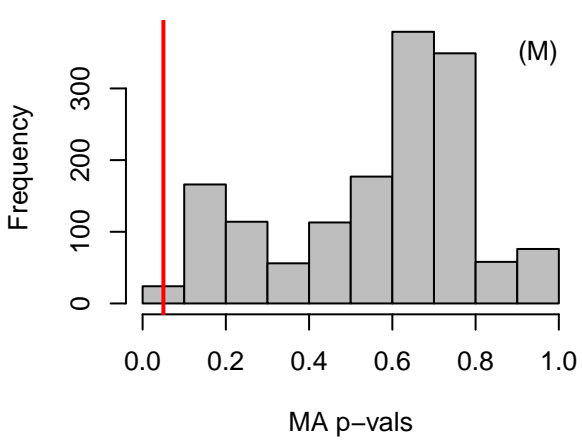
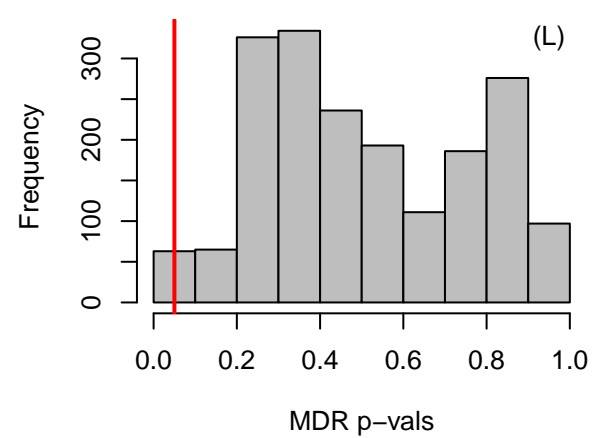
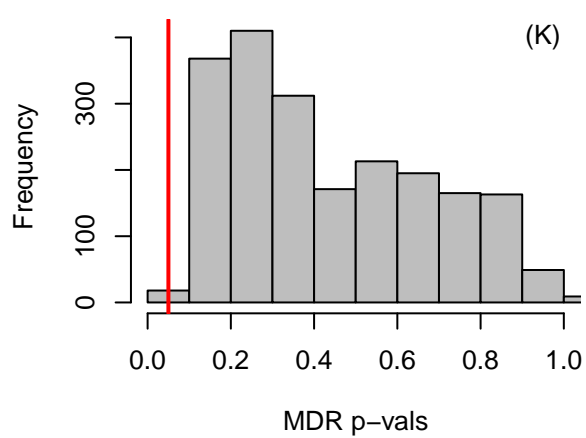
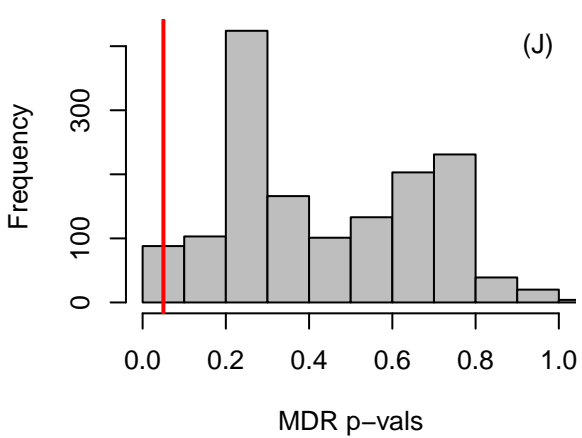
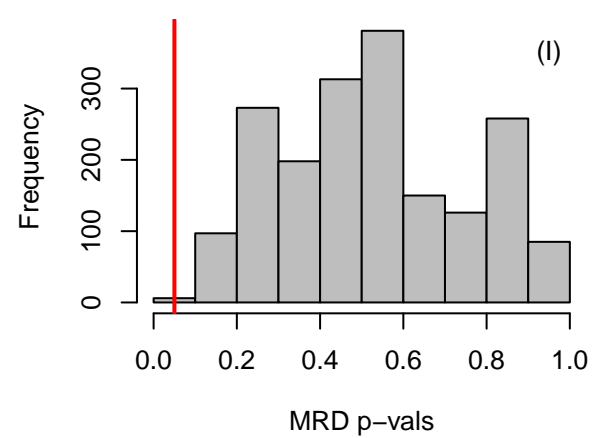
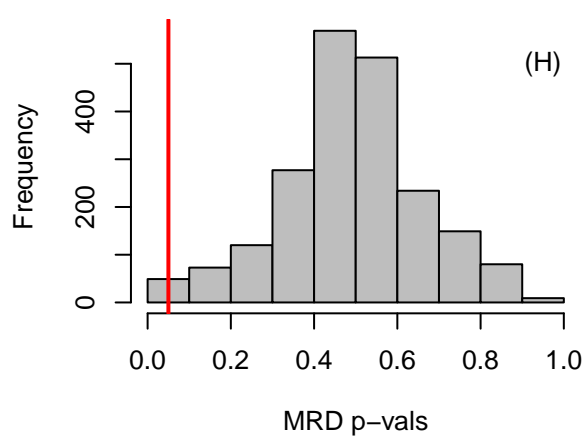
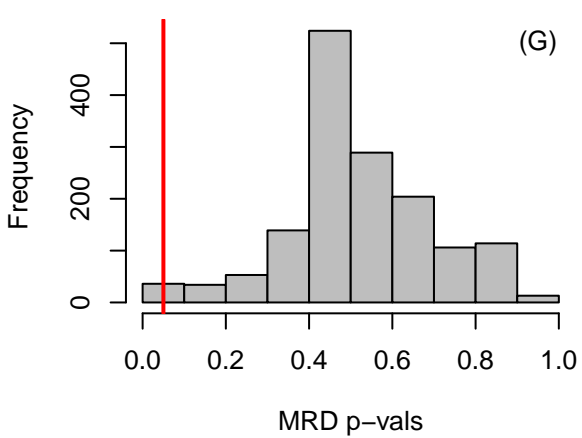
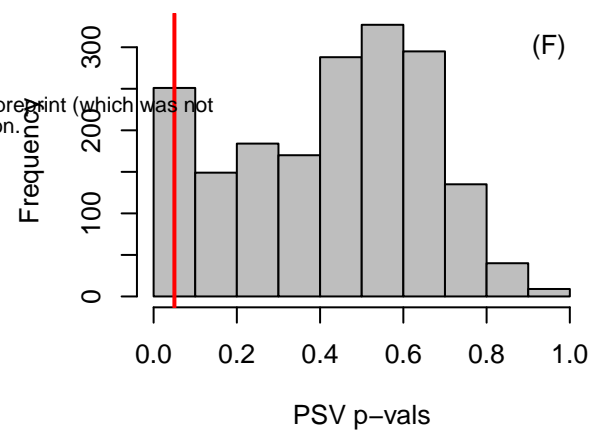
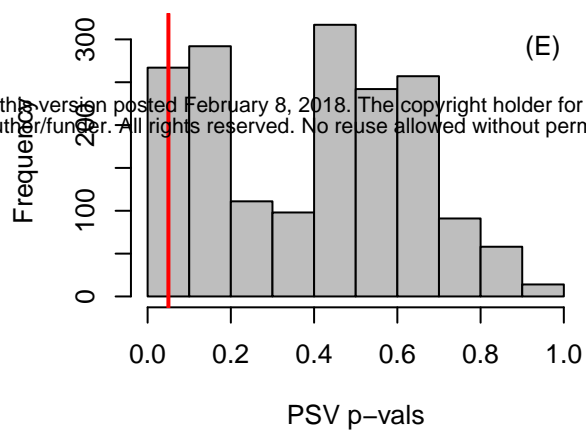
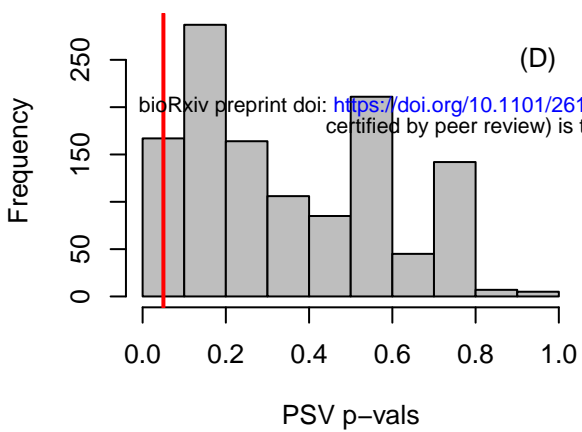
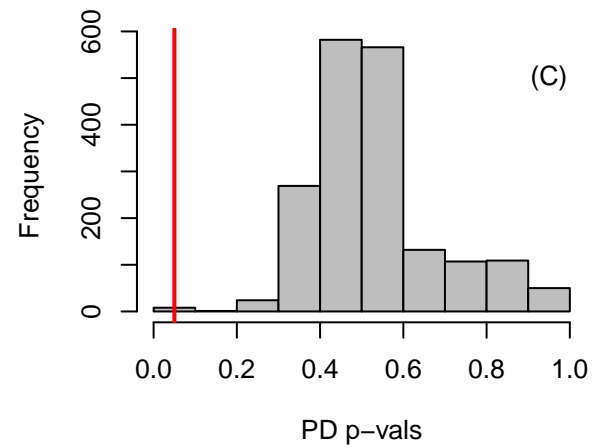


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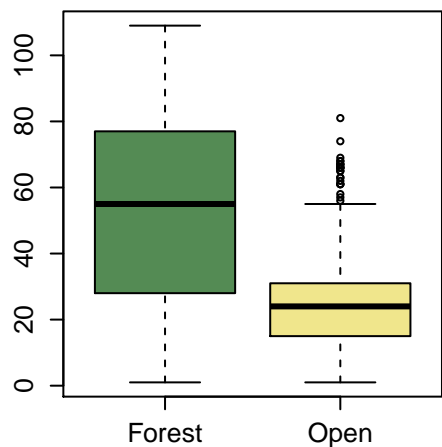
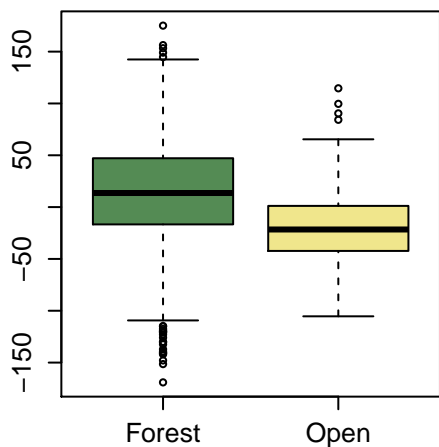
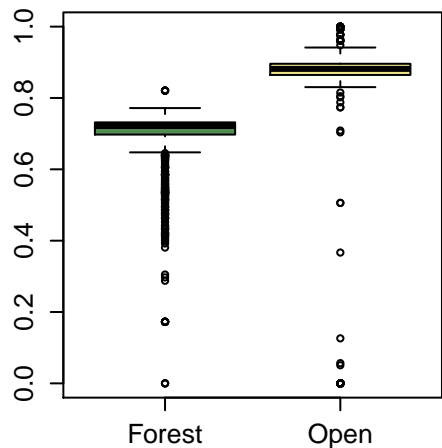
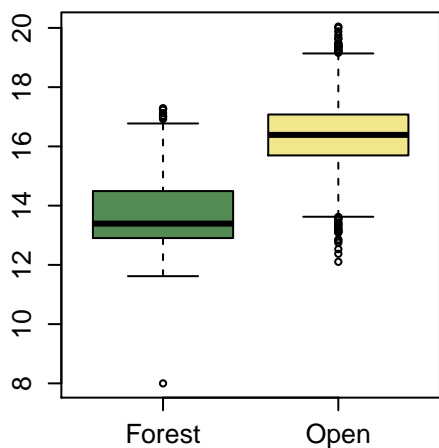
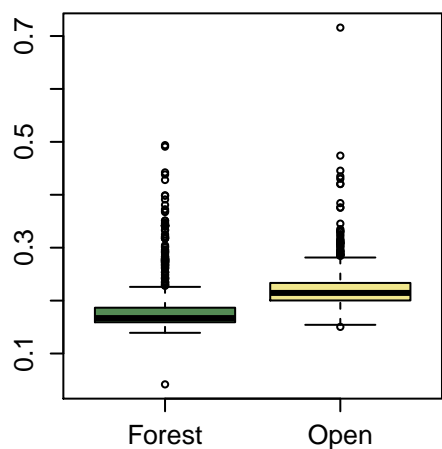
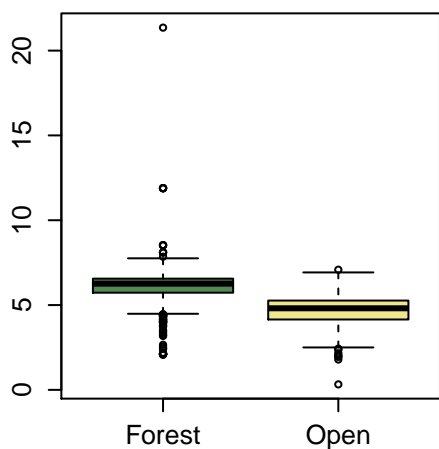


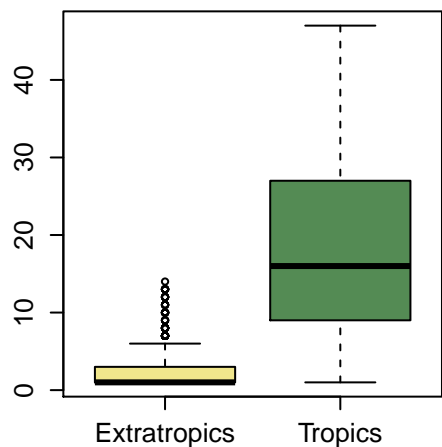
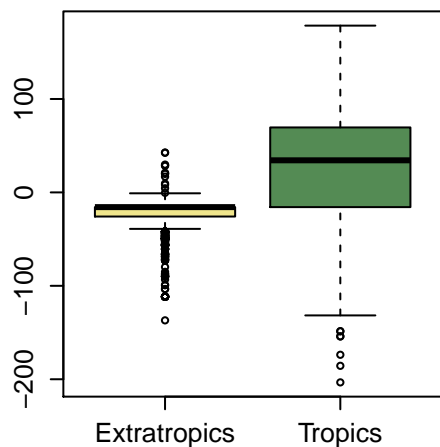
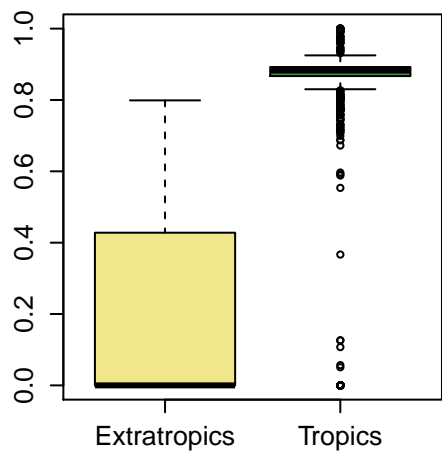
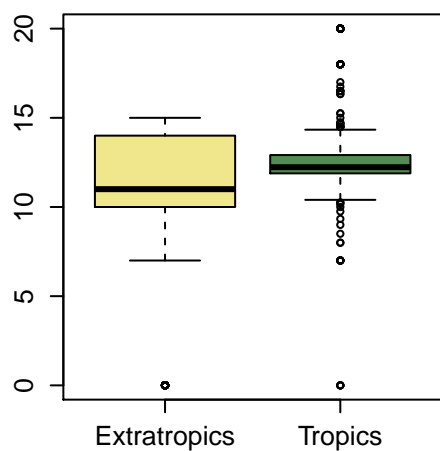
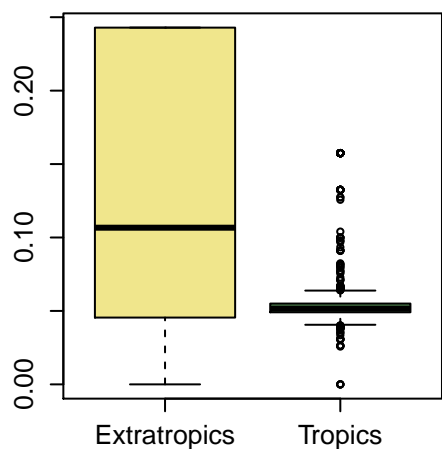
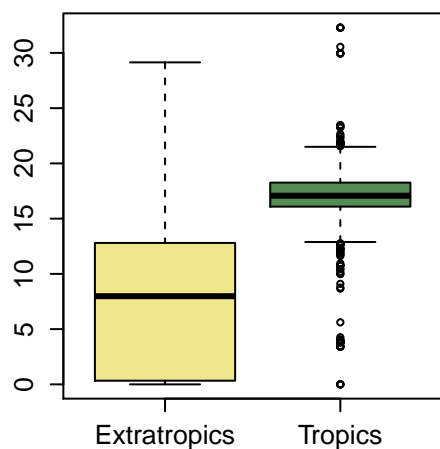
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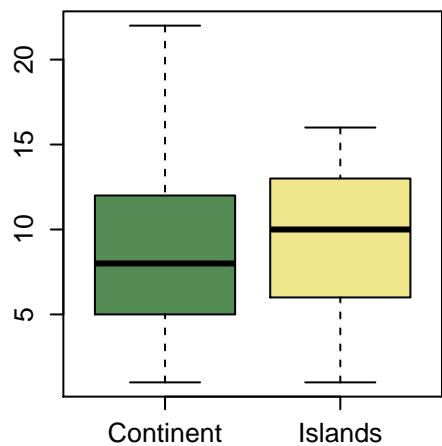
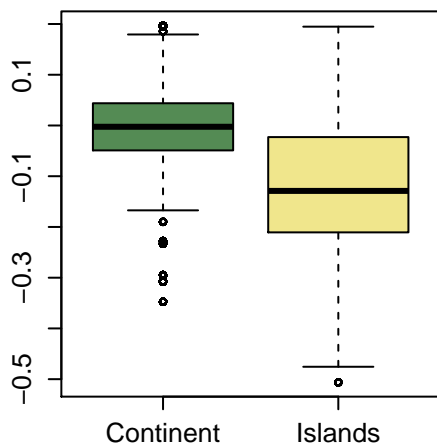
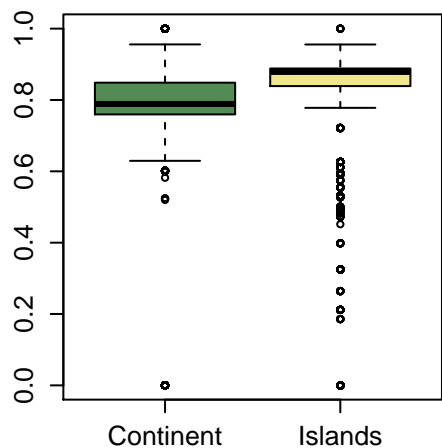
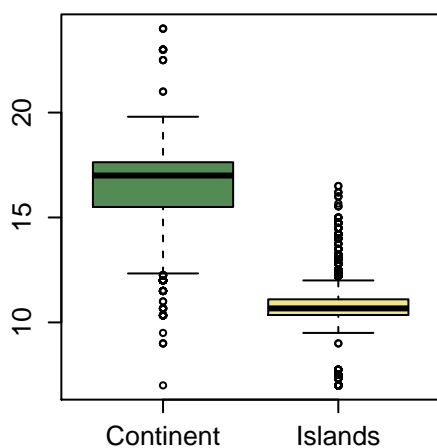
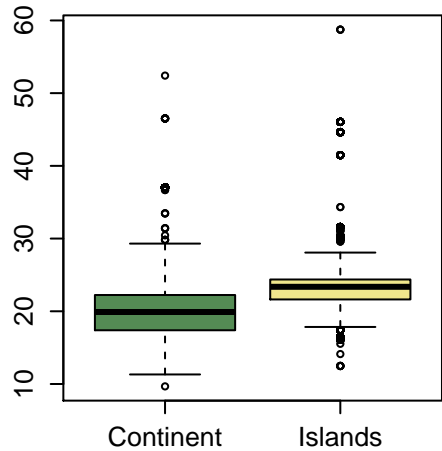
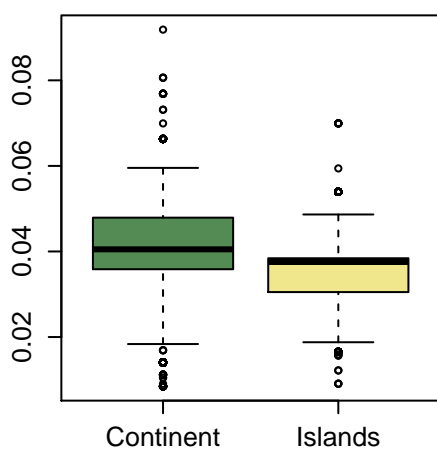


Anolis**Hylids****Furnariides**

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Richness**rPD****PSV****MRD****MDR****MA**

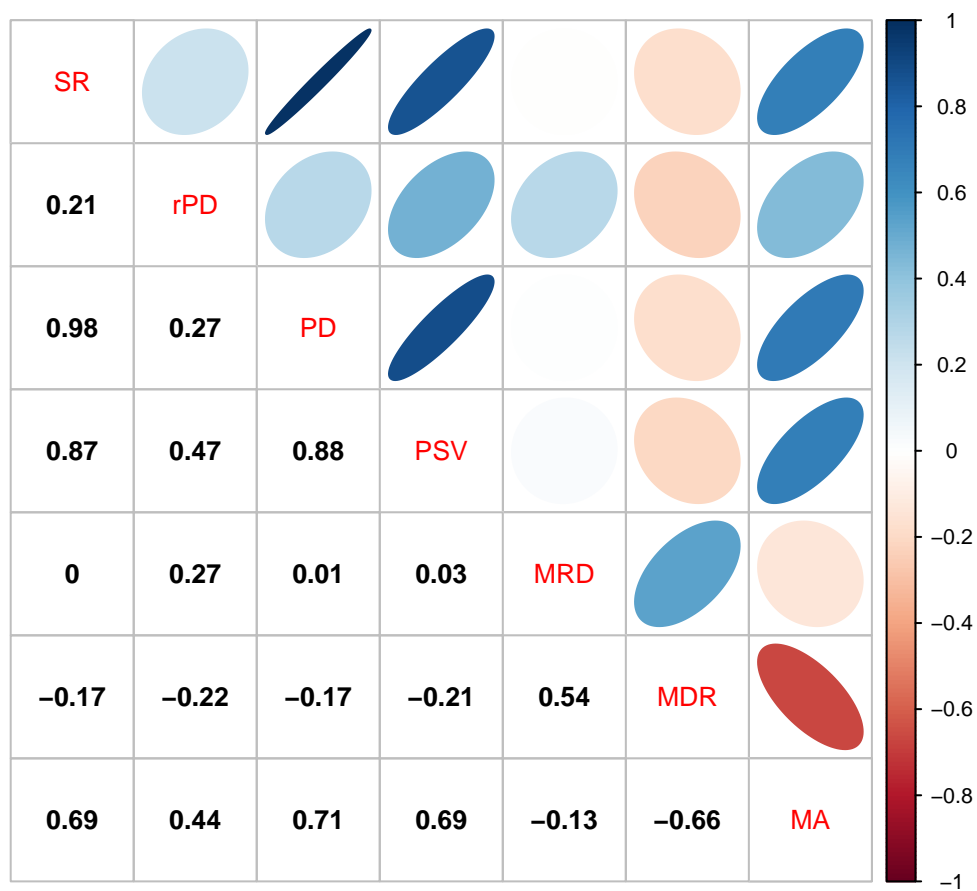
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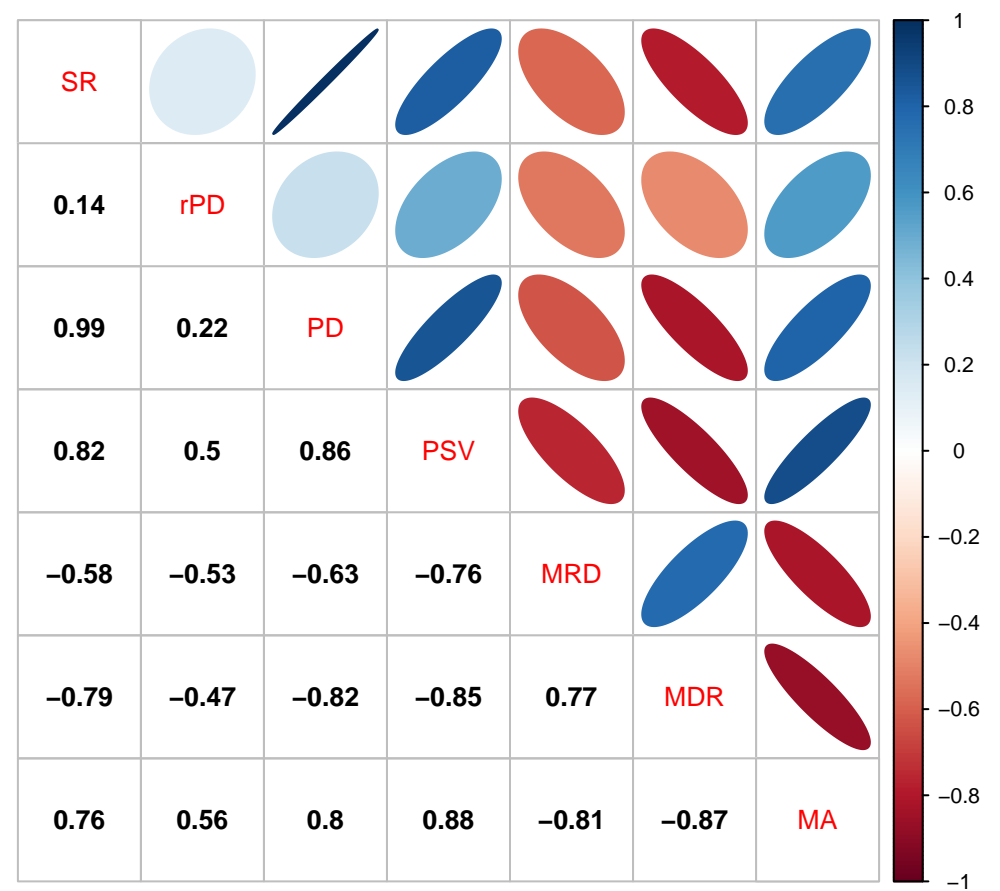
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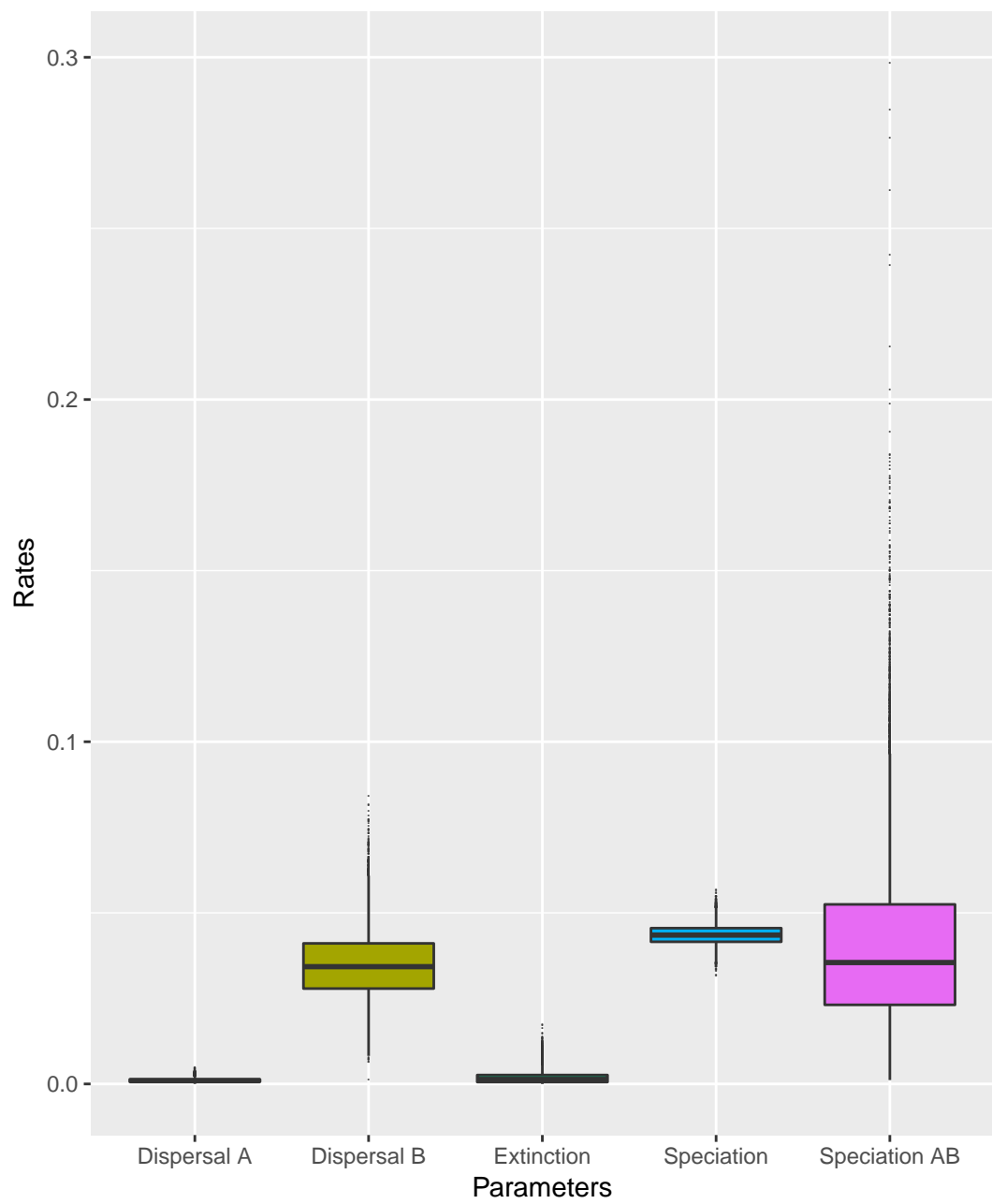
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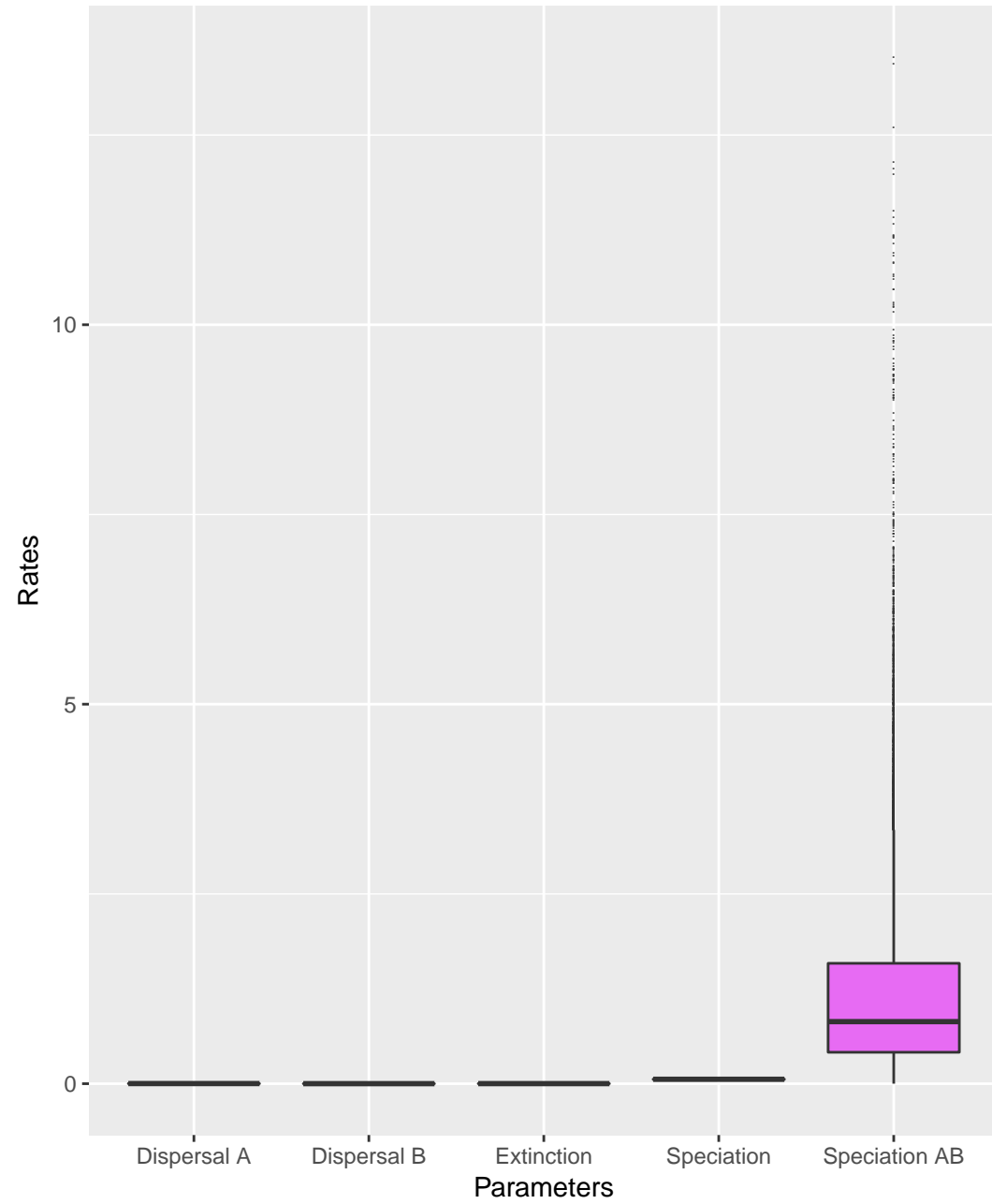
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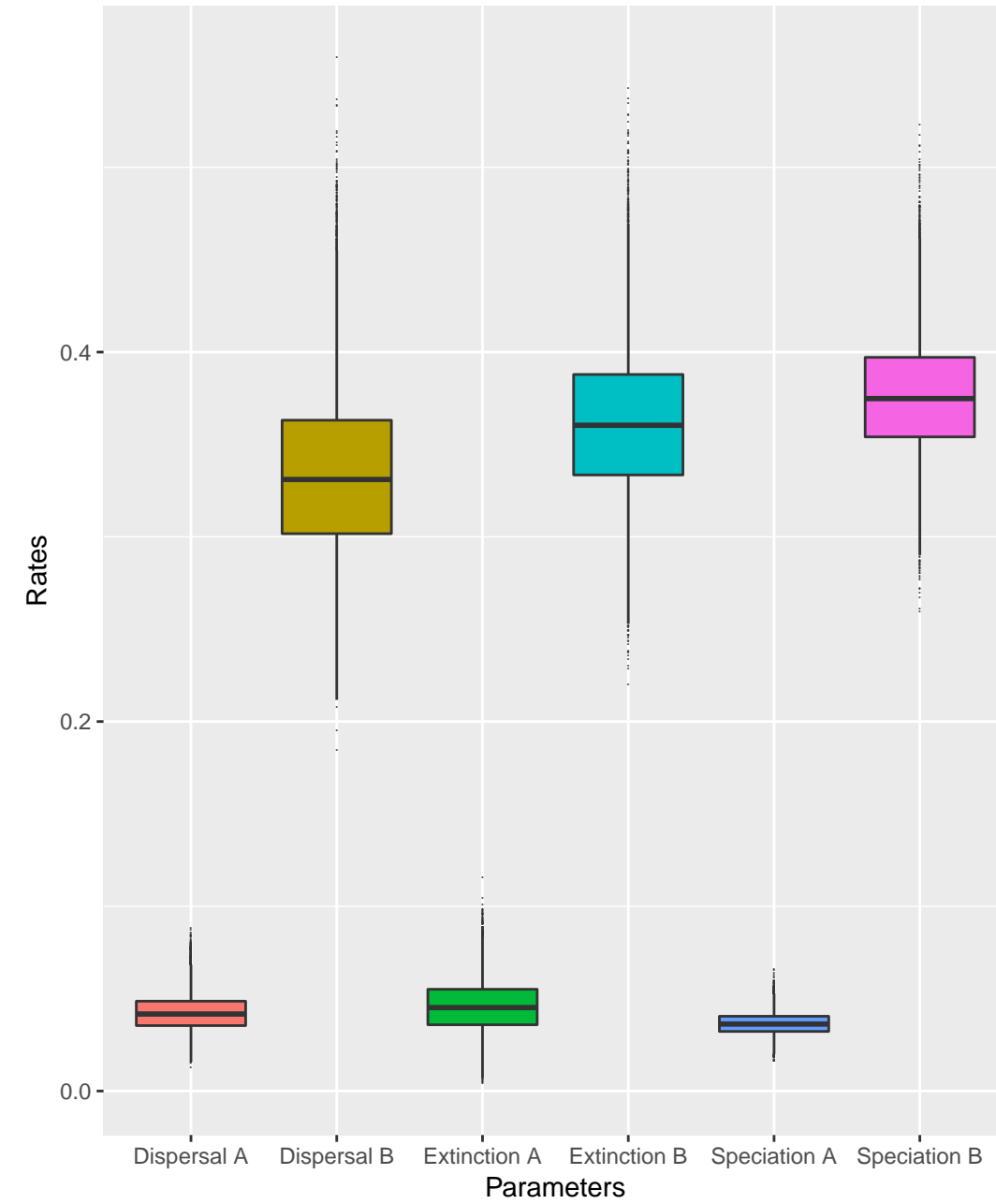
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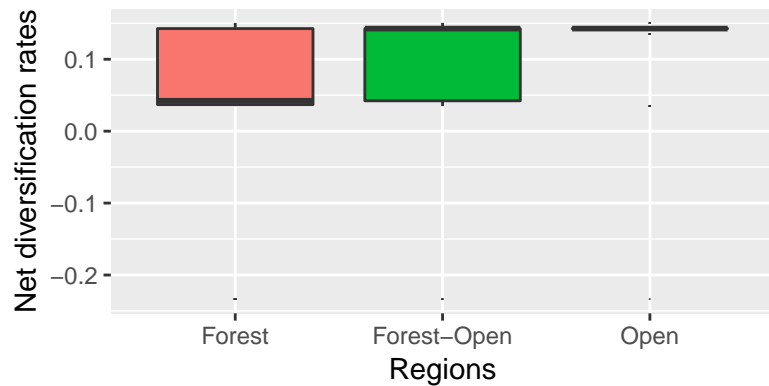
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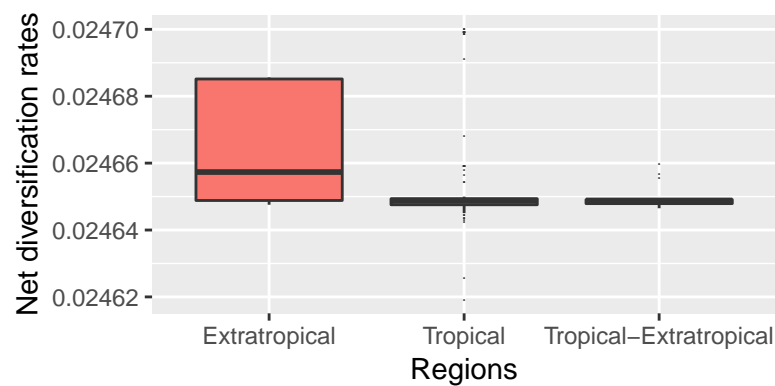
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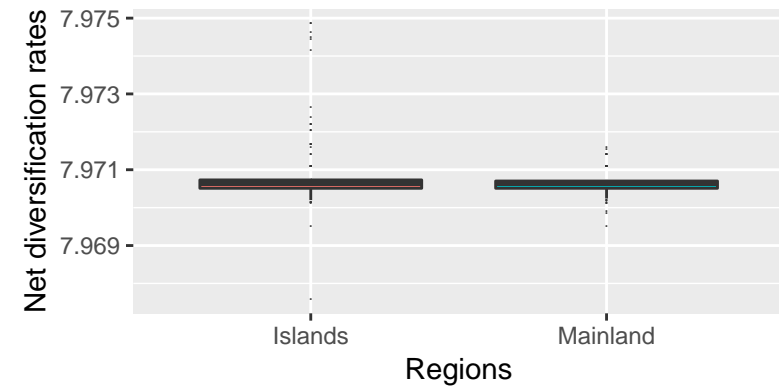
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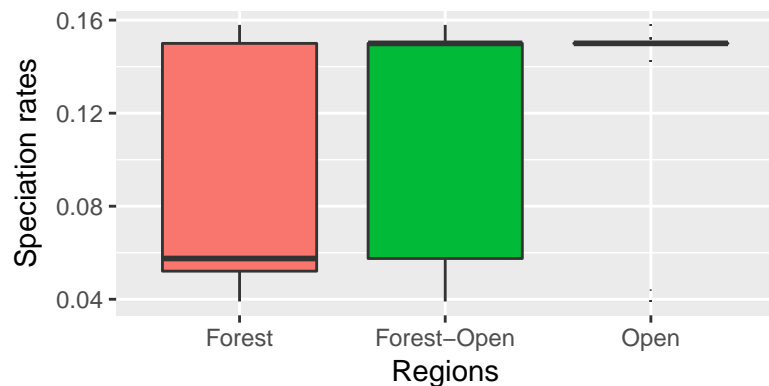
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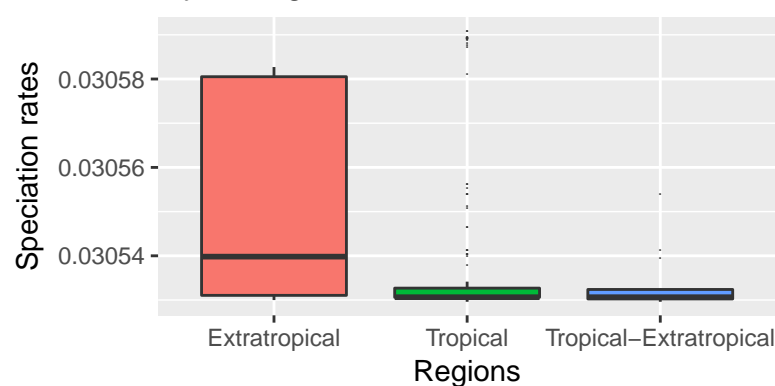
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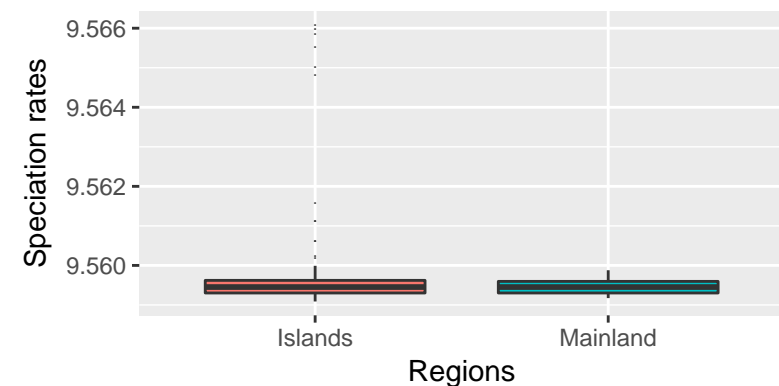
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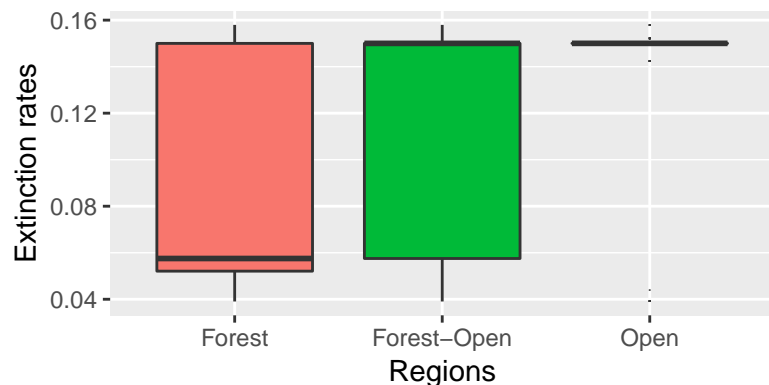
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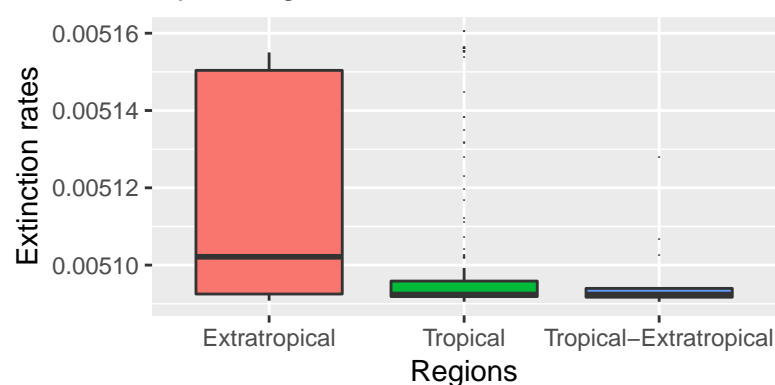
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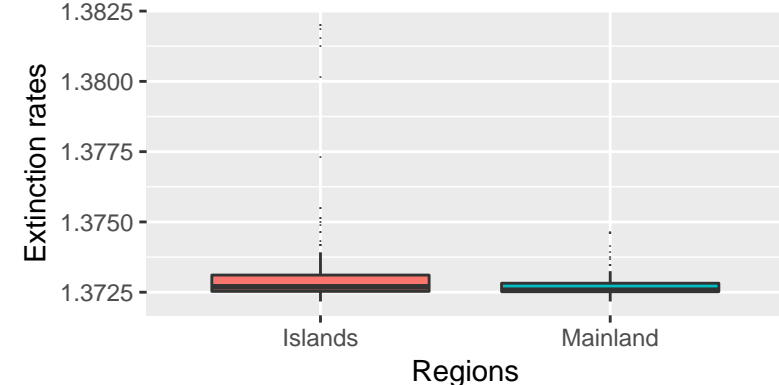
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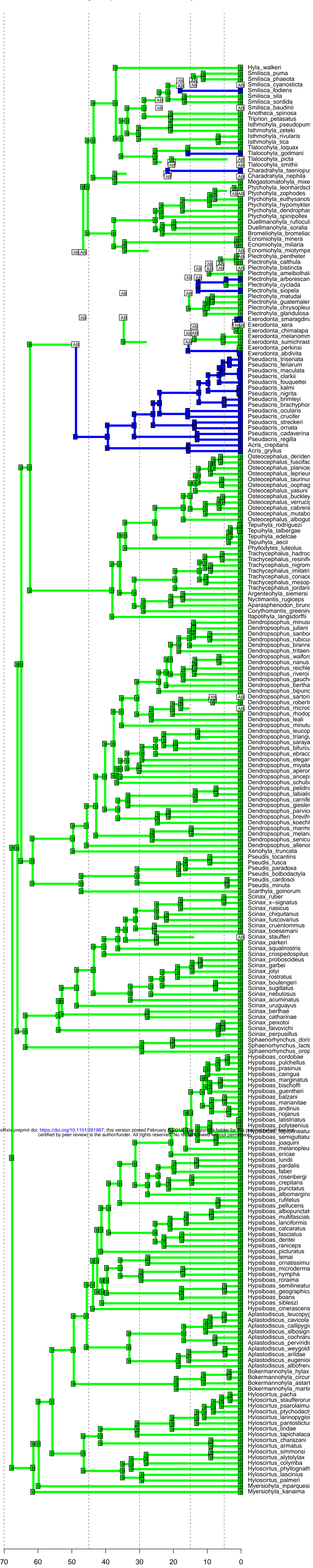


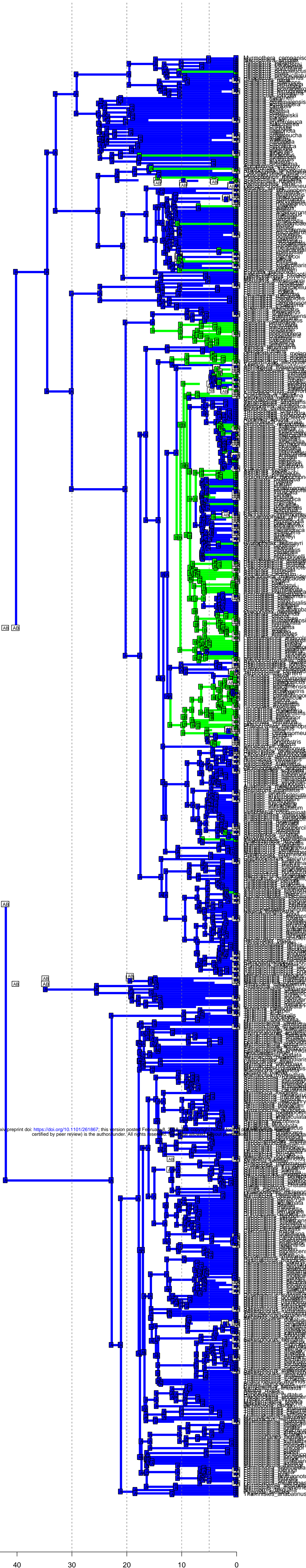
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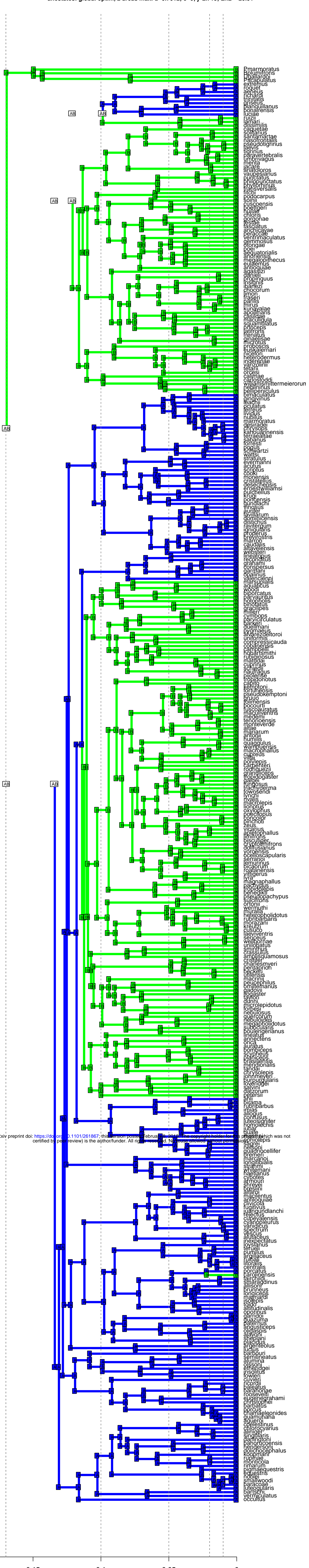
Anole lizards







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