

1 **Biological Sciences/ Evolution**

2

3 **Dissecting historical changes of selective pressures in**  
4 **the evolution of human pigmentation**

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17 **Short title:** Selective pressures during different epochs

18 **Keywords:** complex trait, epistasis, human evolution, natural selection

19

## 20 **Abstract**

21 Human pigmentation is a highly diverse trait among populations, and has drawn  
22 particular attention from both academic and non-academic investigators for thousands  
23 of years. To explain the diversity of human pigmentation, researchers have proposed  
24 that human pigmentation is adapted for ultraviolet radiation and driven by natural  
25 selection. Although studies have detected signals of natural selection in several human  
26 pigmentation genes, none have quantitatively investigated the historical selective  
27 pressures on pigmentation genes during different epochs and thoroughly compared the  
28 differences in selective pressures between different populations. In the present study,  
29 we developed a new approach to dissect historical changes of selective pressures in a  
30 multiple population model by summarizing selective pressures on multiple genes. We  
31 collected genotype data of 16 critical human pigmentation genes from 15 public  
32 datasets, and obtained data for 3399 individuals of five representative populations  
33 from worldwide. Our new approach quantified not only a recent incremental change  
34 of selective pressure ( $0.68 \times 10^{-2}$ /generation) in modern Europeans, but also a  
35 significant historical increase of selective pressure ( $1.78 \times 10^{-2}$ /generation) on light  
36 pigmentation shared by all Eurasians during the out-of-Africa event. We excluded the  
37 relaxation of selective pressures, and favored diversifying selection as the single  
38 explanation for the cause of light pigmentation in Eurasians, a long-standing puzzle in  
39 the evolution of human pigmentation. Our results suggest that epistasis plays  
40 important roles in the evolution of human pigmentation, partially explaining  
41 diversifying selection on human pigmentation among populations.

43 **Significance**

44 The color variation of human skin, hair, and eye is affected by multiple genes with  
45 different roles. This diversity may be shaped by natural selection and adapted for  
46 ultraviolet radiation in different environments around the world, since anatomically  
47 modern human migrated out from Africa to Eurasia. Here, we developed a new  
48 approach and quantified incremental changes of selective pressures on light  
49 pigmentation not only in modern Europeans but also in proto-Eur Asians. Our results  
50 support diversifying selection as the single explanation for the cause of light  
51 pigmentation in Eurasians, and suggest that epistasis might have played important  
52 roles in the evolution of human pigmentation during the out-of-Africa event.

53

## 54 **Introduction**

55 Human pigmentation—the color of the skin, hair, and eye—is one of the most diverse  
56 traits among populations. Its obvious diversity has attracted particular attention from  
57 both academic and non-academic investigators for thousands of years, as noted by  
58 Charles Darwin one century ago (1, p. 192) and as noticed by ancient Egyptians more  
59 than 4000 years ago (2, p. 6). Why human pigmentation diverges, however, remains a  
60 central puzzle in human biology (3). Some researchers have proposed that the  
61 diversity of human pigmentation is adapted for ultraviolet radiation (UVR) and driven  
62 by natural selection (4). Natural selection may favor dark skin for effectively  
63 absorbing sunlight and light skin for efficiently producing vitamin D. Dark skin may  
64 protect individuals against sunburn and skin cancer in low latitude areas with high  
65 UVR (4, 5), while light skin may prevent rickets amongst infants in high latitude areas  
66 with low UVR (6, 7). A better understanding of how natural selection shapes the  
67 diversity of human pigmentation could provide relevant and beneficial information for  
68 public health (4).

69 During the last 10 years, studies have applied statistical tests to detect signals of  
70 natural selection in several human pigmentation genes (8–18). These genes encode  
71 different proteins, including: signal regulators—ASIP, KITLG, MC1R—stimulating  
72 the melanogenic pathway; possible enhancers—BNC2, HERC2—regulating  
73 pigmentation gene expression; important enzymes—TYR, TYRP1—converting  
74 tyrosine into melanin; putative exchangers—OCA2, SLC24A4, SLC24A5, SLC45A2,  
75 TPCN2—controlling the environment within melanosomes; and an exocyst complex

76 unit and molecular motor—EXOC2, MYO5A—conveying vesicles and organelles  
77 within the cytoplasm (19–33). These proteins work at different stages of the  
78 melanogenic pathway, illustrating that human pigmentation is a complex trait affected  
79 by multiple genes with different roles.

80 Previous studies applied two groups of methods for detecting natural selection.  
81 One group of methods detects unusually long extended haplotype homozygosity  
82 (8–12, 14–16). The other group of methods identifies extreme local population  
83 differentiation (8, 9, 11–14, 16). Using both groups of methods, previous studies have  
84 been devoted to understanding the evolution of individual pigmentation genes;  
85 however, few studies have examined how multiple genes contributed to the evolution  
86 of human pigmentation. Moreover, none have quantitatively investigated the historical  
87 selective pressures of pigmentation genes during different epochs, and thoroughly  
88 compared the differences of selective pressures between different populations. To  
89 overcome these weaknesses, it is necessary to perform an extensive investigation with  
90 a creative approach.

91 In the present study, we extended an established method (34) to dissect historical  
92 changes of selective pressures for different epochs of human evolution. Using genetic  
93 variants from worldwide populations, we quantitatively investigated the selective  
94 pressures on human pigmentation during different stages of human evolutionary  
95 history. Our results well explain the current features of human pigmentation among  
96 representative populations. Using individual variants of pigmentation genes, we  
97 thoroughly compared the differences of selective pressures between populations. Our

98 results indicate epistasis plays an important role in the evolution of human  
99 pigmentation, partially leading to diversifying selection on human pigmentation  
100 among populations.

101

## 102 **Results**

103 **Selection model of multiple populations.** We developed a new approach for  
104 dissecting historical changes of selective pressures during different epochs of human  
105 evolutionary history. The evolutionary history of five representative human  
106 populations was simplified as a binary tree (Fig. 1). Based on our previous work (34),  
107 we measured selective pressures by selection coefficients. For a single locus, we can  
108 estimate the selection (coefficient) difference per generation between populations  $i$   
109 and  $j$  by

$$110 \quad \Delta s_{i,j} = \left[ \ln \left( \frac{p_D^{(i)}}{p_A^{(i)}} \right) - \ln \left( \frac{p_D^{(j)}}{p_A^{(j)}} \right) \right] / t_{i,j}, \quad (\text{Eq. 1})$$

111 where  $p_A$  and  $p_D$  are the frequencies of ancestral and derived alleles, and  $t_{i,j}$  is  
112 the divergence time of populations  $i$  and  $j$ . Further, we extended Eq. 1 to estimate  
113 selection differences using multiple loci (*Materials and Methods*).

114 In a scenario with multiple populations, we can determine selection differences in  
115 multiple loci between paired populations by selection (coefficient) changes and  
116 durations of evolutionary stages. Then we can present selection differences between  
117 all the paired populations as an underdetermined system of equations:

$$118 \quad \mathbf{d} = \mathbf{T}\boldsymbol{\delta}, \quad (\text{Eq. 2})$$

119 where  $\mathbf{d}$  and  $\boldsymbol{\delta}$  are vectors that denote the overall selection differences of  
120 population pairs and selection changes in history, and  $\mathbf{T}$  is a matrix containing  
121 durations of evolutionary stages (*Materials and Methods*).

122 To investigate selection changes in our demographic model (Fig. 1), we

123 considered four independent equations from Eq. 2, and assumed that the selection  
124 change happens in only one of the two child processes after branching occurred  
125 (*Materials and Methods*). Therefore, we can dissect the evolutionary history of human  
126 pigmentation by parameterizing and solving the non-linear equations below:

$$\left\{ \begin{array}{l} \Delta s_{2,1}t_3 = -\delta_1t_3 + \delta_2t_3 \\ \Delta s_{3,1}t_1 = -\delta_1t_3 + \delta_3t_2 - \delta_7t_1 + \delta_8t_1 \\ \Delta s_{4,3}t_2 = -\delta_3t_2 + \delta_4t_3 + \delta_6t_2 \\ \Delta s_{5,4}t_3 = -\delta_4t_3 + \delta_5t_3 \\ \delta_7 = 0 \\ \delta_1\delta_2 = 0 \\ \delta_3\delta_6 = 0 \\ \delta_4\delta_5 = 0 \end{array} \right. \quad (\text{Eq. 3})$$

128 **Multistage selection changes of human pigmentation.** To apply our new approach,  
129 we first summarized the overall selection differences (vector  $\mathbf{d}$  in Eq. 2) between all  
130 the population pairs in 15 loci associated with human pigmentation (*Materials and*  
131 *Methods*). Figure 2 plots the overall selection differences, and Table S1 provides more  
132 details. The maximum difference ( $3.059 \times 10^{-2}$ /generation) was observed between  
133 Europeans and West Africans, while the minimum difference ( $0.680 \times 10^{-2}$ /generation)  
134 was observed between Europeans and Siberians. The estimated 95% confidence  
135 intervals (CIs) indicate that these selection differences were significantly deviated  
136 from zero, except for the pair of East and West Africans (Table S2). The large 95% CI  
137 of the pair of East and West Africans (between  $-0.139 \times 10^{-2}$  and  $2.768 \times$   
138  $10^{-2}$ /generation) was likely due to the genetic drift between East and West Africans.  
139 Our results of selection differences are consistent with differences of skin reflectance  
140 between populations (Spearman correlation coefficient  $\rho = 0.95$ ,  $p < 0.001$ , Table S3),



141 suggesting that selective pressures on multiple genes can result in different skin colors  
142 among populations. We also note that selective pressure in Siberians was slightly  
143 stronger than that in East Asians (Fig. 2), which is consistent with the view that  
144 latitude is one of the important factors in the evolution of human pigmentation (6, 35).

145 We next parameterized and solved Eq. 3 to dissect the overall selection  
146 differences into multistage selection changes for different evolutionary processes.  
147 This investigation gave eight possible solutions for the multistage selection changes,  
148 any of which can fully explain the observed selection differences between the  
149 population pairs (Table 1). Further analysis suggests that solution #1 is the optimal  
150 solution in our evolutionary scenario (*Materials and Methods*). We present the  
151 sequential selection changes of solution #1 in Fig. 3. This solution indicates that the  
152 largest selection change may have occurred during the out-of-Africa event ( $\delta_8 = 1.78$   
153  $\times 10^{-2}$ /generation, Table 1), which implies a dramatic environmental change at the  
154 first stage of the great human migration.

155 We then summarized all the selection changes on the evolutionary path of each  
156 population (Table S4). All the eight possible solutions exhibited the same pattern that  
157 the summarized selection changes were positive for all Eurasians since the  
158 out-of-Africa event (in the range between  $0.36 \times 10^{-2}$  and  $3.33 \times 10^{-2}$ /generation,  
159 Table S4), which indicates that light skin is favorable for all Eurasians. All the  
160 solutions suggest that the modern European lineage had the largest selective pressure  
161 on derived alleles, whereas the modern East Asian lineage experienced the smallest  
162 selective pressure of light pigmentation. From the optimal solution, our results

163 suggest that the modern European lineage had an incremental selection change ( $\delta_3 =$   
164  $0.68 \times 10^{-2}$ /generation, Table 1), while the modern East Asian lineage had a  
165 decremental change ( $\delta_5 = -1.42 \times 10^{-2}$ /generation, Table 1), experiencing a possible  
166 relaxation of selective pressure on derived alleles. These observations are consistent  
167 with the lower skin reflectance in East Asians than in Europeans (7, Table S3).

168 **Quantification of selection differences in individual loci.** Finally, we separately  
169 quantified selection differences of the selected 31 single nucleotide polymorphisms  
170 (SNPs) to explore selection patterns of the individual loci (*Materials and Methods*).  
171 Our pilot analysis illustrates that linkage disequilibrium was generally weak between  
172 these SNPs (Fig. S1). Statistical tests suggest that most of the selection differences  
173 between populations were highly significant (Table S5). Therefore, these differences  
174 probably could not be explained by population history or the relaxation of selective  
175 pressures. Based on the selection patterns of the individual loci, we categorized the 31  
176 selected SNPs into four groups (Fig. 4).

177 In the first group, all Eurasians presented directional selection on derived alleles  
178 of the SNPs (Fig. 4A). This result is consistent with the observation that reduced  
179 pigmentation occurred in most populations outside Africa. In this group, rs6119471  
180 (*ASIP*), rs2228479 (*MC1R*), and rs885479 (*MC1R*) showed extreme selection  
181 differences between Africans and Eurasians. Among them, the selection difference of  
182 rs6119471 was ranked the second largest in our study ( $\Delta_{SEAS-WAF} = 2.274 \times$   
183  $10^{-3}$ /generation). We note that these two genes are the major regulators upstream of  
184 the melanogenic pathway (Fig. 5). Conversely, rs4776053 (*MYO5A*) and rs4959270

185 (*EXOC2*) had small selection differences, suggesting their little contribution to the  
186 diversity of human pigmentation.

187 The second and third groups showed European- and Asian-specific selection,  
188 respectively (Fig. 4B and 4C). One notable SNP is rs1426654 (*SLC24A5*), which had  
189 the largest selection difference in our study ( $\Delta s_{\text{EUR-EAS}} = 2.625 \times 10^{-3}/\text{generation}$ ).  
190 Previous studies reported that this SNP is under strong directional selection in  
191 Europeans (8–10, 12–14, 36). Another notable SNP is rs1800414 (*OCA2*), which had  
192 the third largest selection difference in our study ( $\Delta s_{\text{EAS-WAF}} = 2.217 \times$   
193  $10^{-3}/\text{generation}$ ). Several studies have suggested directional selection on this SNP in  
194 East Asians (15, 37, 38). These large selection differences indicate the significant  
195 contributions of these SNPs to light pigmentation in Europeans and East Asians,  
196 respectively. In addition, other SNPs in these groups support the hypothesis that  
197 recent natural selection for light pigmentation independently occurred in Europeans  
198 and Asians since their divergence. In these two groups, most of the genes work  
199 downstream of the melanogenic pathway (Fig. 5).

200 The last group included the remaining four SNPs (Fig. 4D), which exhibited  
201 specific selection differences between limited populations pairs. Among them, the  
202 derived allele of rs1800401 (*OCA2*) is associated with dark pigmentation (20, 21, 39).  
203 Our study indicates a rare case that the variant associated with dark pigmentation  
204 might be favored by natural selection.

## 205 **Discussion**

206 In the present study, we dissected historical changes of selective pressures by  
207 summarizing the selection differences in multiple human pigmentation genes. Our  
208 results quantify not only a recent incremental change of selective pressure ( $\delta_3 = 0.68$   
209  $\times 10^{-2}$ /generation, Table 1) in Europeans, but also a significant historical increase of  
210 selective pressure ( $\delta_8 = 1.78 \times 10^{-2}$ /generation, Table 1) that favored light  
211 pigmentation in all Eurasians during the out-of-Africa event. Recent studies using  
212 ancient DNA support our observation of recent directional selection in Europeans (17,  
213 40). Compared with these ancient DNA studies, our study has the advantage that we  
214 do not need to assume population continuity (17, 41), because our study is based on  
215 genetic data from only present-day populations. Thus, our results could provide more  
216 solid evidence of the recent directional selection in Europeans. Further, our results  
217 demonstrate independent selective pressures on light pigmentation in modern  
218 Europeans and East Asians as previous studies (12, 13, 37, 38), and a shared selective  
219 pressure that favored light pigmentation in proto-Eurasian populations. This shared  
220 selection is consistent with other studies, which revealed that *ASIP*, *BNC2*, and  
221 *KITLG* were under directional selection before the divergence of ancestral Europeans  
222 and Asians (9, 42). Unlike previous studies, we summarized selective pressures on  
223 multiple human pigmentation genes with larger sample size and more representative  
224 populations. Therefore, our results could be more relevant to the evolution of human  
225 pigmentation. Overall, our results suggest that natural selection continuously favors  
226 light pigmentation in Eurasians since the out-of-Africa event, supported by allele age

227 estimation from another study (43).

228 In addition, our results of individual loci strongly suggest that epistasis plays a  
229 critical role in the evolution of human pigmentation. Most of the selected genes  
230 shared by Eurasians are major regulators upstream of the melanogenic pathway (Fig.  
231 5), such as *ASIP*, *KITLG*, and *MC1R* (Fig. 4A). *MC1R* encodes a  
232 seven-transmembrane G-protein coupled receptor that can interact with  
233  $\alpha$ -melanocyte-stimulating hormone ( $\alpha$ -MSH), and switch the melanogenic pathway  
234 from synthesizing the red/yellow pheomelanin to black/brown eumelanin. When  
235 UVR exists, *MC1R* is activated by  $\alpha$ -MSH, resulting in the synthesis of eumelanin  
236 (21);  $\alpha$ -MSH can be blocked or inhibited by agouti signaling protein (*ASIP*), leading  
237 to the production of pheomelanin (19). Most of the remaining genes (Fig. 4B and 4C)  
238 with continental-specific selection patterns work downstream of the melanogenic  
239 pathway (Fig. 5). These upstream genes can regulate the expression of genes  
240 downstream in the melanogenic pathway, such as *TYR*, *TYRP1*, *OCA2*, and *SLC45A2*  
241 (44, 45). Published genetic association studies could support our hypothesis about the  
242 role of epistasis in light pigmentation. The derived allele of rs12913832 (*HERC2*)  
243 significantly increases the risk of light skin, only when individuals also carry derived  
244 alleles of the upstream gene *MC1R* (46, 47). These upstream genes contributed large  
245 selection differences to all Eurasians (Fig. 4A), indicating that epistasis might have  
246 played an important role in the evolution of human pigmentation during the  
247 out-of-Africa event.

248 Finally, our results also solve the long-standing puzzle regarding whether light

249 pigmentation in low UVR areas is caused by diversifying selection or the relaxation  
250 of selective pressures (17, 48, 49). We used the statistical test that we recently  
251 developed (34) to exclude the relaxation of selective pressures, and favor diversifying  
252 selection as the single explanation. Before our study, multiple studies closely  
253 inspected *MC1R* because its coding region has unusually higher diversity in Eurasians  
254 than in Africans (9, 12, 13, 16, 50–52). These studies reached different conclusions,  
255 however, either diversifying selection or the relaxation of selective pressures. The  
256 relaxation of selective pressures (51) would suggest that the diversity of *MC1R*  
257 variants increased in Eurasians due to the lack of selective constraints. In this scenario,  
258 the genetic diversity of *MC1R* variants could be largely attributed by genetic drift. In  
259 contrast, diversifying selection (50) would suggest that *MC1R* variants were under  
260 directional selection in Eurasians. In this scenario, genetic drift cannot explain the  
261 genetic divergence of *MC1R* between Africans and Eurasians. Our results show that  
262 the divergences of rs2228479 (*MC1R*) and rs885479 (*MC1R*) between Africans and  
263 Eurasians are highly significant departure from neutral evolution (Table S5).  
264 Experimental evidence suggests that the derived allele of rs2228479 (*MC1R*) could  
265 cause lower affinity for  $\alpha$ -MSH than the ancestral allele (56). Another study showed  
266 that the derived allele of rs885479 (*MC1R*) carries a lower risk of developing freckles  
267 and severe solar lentigines than the ancestral allele in East Asians (57). These studies  
268 revealed the potential roles of these *MC1R* variants in pigmentation phenotypes. In  
269 addition to our results, one study reported possible diversifying selection in parts of  
270 the promoter sequence of *MC1R* (58). Combining our previous results with

271 aforementioned experimental evidence, we suggest that diversifying selection may  
272 have occurred since the out-of-Africa event, and favored light pigmentation through  
273 epistasis.

274 We note that our investigation has several limitations. First, we assumed that the  
275 selection change occurs in only one of the two branching processes. This assumption  
276 is consistent with the stepping stone model (59), which is a good and simple  
277 approximation to the history of human migration. Second, we obtained eight possible  
278 solutions of historical selection changes (Table 1). Although we chose the most  
279 conservative one as the optimal solution, we cannot exclude the possibility of other  
280 solutions. All the solutions, however, show the same trend of natural selection that  
281 light pigmentation is favored in all Eurasians. This reflects the difficulty of analyzing  
282 historical selective pressures, which is a well-recognized challenge in population  
283 genetics.

284 To summarize, our study provides information of historical selective pressures  
285 during different epochs of the evolution of human pigmentation. The results of our  
286 analysis suggest that epistasis partially explains the diversifying selection on human  
287 pigmentation among worldwide populations. Further studies are in progress to verify  
288 our present views on the evolution of human pigmentation.

289

## 290 **Materials and Methods**

291 **Data collection.** Fifteen publicly available datasets containing genotype data from  
292 worldwide human populations were downloaded from web resources (60–74).

293 **Data preparation.** All the downloaded data were transferred to genomic coordinates  
294 using NCBI dbSNP (build 144) with the Human Reference Genome hg19. A merged  
295 dataset containing 6119 individuals and 81,364 SNPs were obtained after removing  
296 duplicate and related individuals (Table S6, S7). PLINK 1.7 (75) was used to exclude  
297 SNPs for which genotyping rates were lower than 0.99 and major allele frequencies  
298 were lower than 0.05 (--geno 0.01 --maf 0.05). SNPs in strong linkage disequilibrium  
299 were removed by applying a window of 50 SNPs advanced by 5 SNPs and an  $r^2$   
300 threshold of 0.02 (--indep-pairwise 50 5 0.02) in PLINK. The remaining 13,499 SNPs  
301 were used for principal components analysis (PCA). PCA was performed using  
302 SMARTPCA (version: 13050) from EIGENSOFT 6.0.1 (76, 77). After removing  
303 individuals from admixed populations and outliers (Table S8) identified by PCA,  
304 3399 individuals (Table S9) were obtained and divided into five groups according to  
305 their geographic regions for further analysis. PCA plots (Fig. S2) showed these 3399  
306 individuals were properly separated into different population groups.

307 **Data imputation.** Genotypes of 16 human pigmentation genes with 500-kb flanking  
308 sequences on both sides were obtained from the downloaded datasets. Haplotype  
309 inference and genotype imputation were performed on the selected genotypes using  
310 BEAGLE 4.1 with 1000 Genomes phase 3 haplotypes as the reference panel (78, 79,



311 Table S10). During phasing and imputation, the effective population size was assumed  
312 to be 10,000 ( $N_e=10000$ ), and the other parameters were set to the default values.  
313 Forty-two SNPs were selected for analysis due to their strong association with human  
314 pigmentation in published genome-wide association studies or phenotype prediction  
315 models (Table S11). Eleven SNPs (rs1110400, rs11547464, rs12203592, rs12821256,  
316 rs1800407, rs1805005, rs1805006, rs1805007, rs1805008, rs1805009, rs74653330)  
317 were removed because of their low frequencies in our datasets after imputation (Fig.  
318 S3). Because rs12203592 (*IRF4*) was removed, 15 loci with the remaining 31 SNPs  
319 were used for further analysis.

320 **Selection difference estimation in a single locus.** The logarithm odds ratios for the  
321 selected loci were calculated and used for estimating their selection differences  
322 between populations. The estimated CIs were calculated using the imputed genotype  
323 data of 31 SNPs. Variances of genetic drift between populations (Fig. S4, Table S12)  
324 were determined using 13,499 SNPs without strong linkage disequilibrium in the  
325 merged dataset (see *Data preparation*). Details of the calculations are described  
326 elsewhere (34). Results were presented in Table S5, and visualized in heat maps (Fig.  
327 4) using Python 3.5.1 with Matplotlib 1.4.3 through Jupyter Notebook 4.1.0 in  
328 ANACONDA 2.4.0.

329 **Selection difference estimation in multiple loci.** We extended Eq. 1 to estimate the  
330 selection difference in multiple loci. Here, we take two bi-allelic loci as an example.  
331 We can estimate the selection difference of the haplotype carrying two derived alleles  
332 between populations  $i$  and  $j$  by

$$333 \quad \Delta s_{i,j}^{D_1D_2} = \left[ \ln \left( \frac{p_{D_1D_2}^{(i)}}{p_{A_1A_2}^{(i)}} \right) - \ln \left( \frac{p_{D_1D_2}^{(j)}}{p_{A_1A_2}^{(j)}} \right) \right] / t_{i,j},$$

334 where  $p_{A_1A_2}$  is the frequency of the haplotype carrying two ancestral alleles,  $p_{D_1D_2}$   
 335 is the frequency of the haplotype carrying two derived alleles, and  $t_{i,j}$  is the  
 336 divergence time between populations  $i$  and  $j$ . Assuming linkage equilibrium, we have

$$337 \quad \begin{aligned} \Delta s_{i,j}^{D_1D_2} &= \left[ \ln \left( \frac{p_{D_1D_2}^{(i)}}{p_{A_1A_2}^{(i)}} \right) - \ln \left( \frac{p_{D_1D_2}^{(j)}}{p_{A_1A_2}^{(j)}} \right) \right] / t_{i,j} \\ &= \left[ \ln \left( \frac{p_{D_1}^{(i)} p_{D_2}^{(i)}}{p_{A_1}^{(i)} p_{A_2}^{(i)}} \right) - \ln \left( \frac{p_{D_1}^{(j)} p_{D_2}^{(j)}}{p_{A_1}^{(j)} p_{A_2}^{(j)}} \right) \right] / t_{i,j}, \\ &= \left[ \ln \left( \frac{p_{D_1}^{(i)}}{p_{A_1}^{(i)}} \right) - \ln \left( \frac{p_{D_1}^{(j)}}{p_{A_1}^{(j)}} \right) \right] / t_{i,j} + \left[ \ln \left( \frac{p_{D_2}^{(i)}}{p_{A_2}^{(i)}} \right) - \ln \left( \frac{p_{D_2}^{(j)}}{p_{A_2}^{(j)}} \right) \right] / t_{i,j} \\ &= \Delta s_{i,j}^{L_1} + \Delta s_{i,j}^{L_2} \end{aligned}$$

338 where  $p_{A_1}$  and  $p_{A_2}$  are the frequencies of ancestral alleles in the first and second  
 339 loci, respectively;  $p_{D_1}$  and  $p_{D_2}$  are the frequencies of derived alleles in the first  
 340 and second loci, respectively; and  $\Delta s_{i,j}^{L_1}$  are  $\Delta s_{i,j}^{L_2}$  the selection differences between  
 341 populations  $i$  and  $j$  in the first and second loci, respectively. Therefore, we can obtain  
 342 the overall selection differences of multiple loci by summarizing estimations of  
 343 individual loci.

344 **Selection difference dissection in a multiple population model.** We developed an  
 345 approach to calculate the selection differences for different evolutionary stages using  
 346 our demographic model of five populations (Fig. 1). When  $k$  is the most recent  
 347 common ancestral population of  $i$  and  $j$ , we can divide  $\Delta s_{i,j}$  in Eq. 1 into separate  
 348 terms:

$$349 \quad \Delta s_{i,j} t_{i,j} = \left[ \ln \left( \frac{p_D^{(i)}}{p_A^{(i)}} \right) - \ln \left( \frac{p_D^{(k)}}{p_A^{(k)}} \right) \right] - \left[ \ln \left( \frac{p_D^{(j)}}{p_A^{(j)}} \right) - \ln \left( \frac{p_D^{(k)}}{p_A^{(k)}} \right) \right].$$

350 We can further divide the selection difference between paired populations into  
 351 multiple terms, if there are multiple branches between them (Fig. 1). For example,  
 352 using the notations and demographic model in Fig. 1, we can estimate the total  
 353 selection difference between Europeans and West Africans as

$$354 \quad \Delta s_{3,1} t_1 = - \left\{ \left[ \ln \left( \frac{p_D^{(1)}}{p_A^{(1)}} \right) - \ln \left( \frac{p_D^{(x)}}{p_A^{(x)}} \right) \right] + \left[ \ln \left( \frac{p_D^{(x)}}{p_A^{(x)}} \right) - \ln \left( \frac{p_D^{(r)}}{p_A^{(r)}} \right) \right] \right\} \\ + \left\{ \left[ \ln \left( \frac{p_D^{(3)}}{p_A^{(3)}} \right) - \ln \left( \frac{p_D^{(y)}}{p_A^{(y)}} \right) \right] + \left[ \ln \left( \frac{p_D^{(y)}}{p_A^{(y)}} \right) - \ln \left( \frac{p_D^{(r)}}{p_A^{(r)}} \right) \right] \right\}.$$

$$355 \quad \text{Let } s_1 = \left[ \ln \left( \frac{p_D^{(1)}}{p_A^{(1)}} \right) - \ln \left( \frac{p_D^{(x)}}{p_A^{(x)}} \right) \right] / t_3, \quad s_3 = \left[ \ln \left( \frac{p_D^{(3)}}{p_A^{(3)}} \right) - \ln \left( \frac{p_D^{(y)}}{p_A^{(y)}} \right) \right] / t_2,$$

$$356 \quad s_7 = \left[ \ln \left( \frac{p_D^{(x)}}{p_A^{(x)}} \right) - \ln \left( \frac{p_D^{(r)}}{p_A^{(r)}} \right) \right] / (t_1 - t_3), \quad \text{and } s_8 = \left[ \ln \left( \frac{p_D^{(y)}}{p_A^{(y)}} \right) - \ln \left( \frac{p_D^{(r)}}{p_A^{(r)}} \right) \right] / (t_1 - t_2), \text{ then we}$$

357 have

$$358 \quad \Delta s_{3,1} t_1 = -s_1 t_3 + s_3 t_2 - s_7 (t_1 - t_3) + s_8 (t_1 - t_2). \quad (\text{Eq. 4})$$

359 Therefore, we can represent the selection difference between paired populations as a  
 360 combination of multistage selection coefficients. Moreover, if we let  $s_0$  be the  
 361 selection coefficient of the population in the root, we can express the selection  
 362 coefficients of populations in different branches using  $s_0$  plus some selection changes  
 363 (Fig. 1). Using the notations in Fig.1, we can rewrite Eq. 4 into

$$364 \quad \Delta s_{3,1} t_1 = -\delta_1 t_3 + \delta_3 t_2 - \delta_7 t_1 + \delta_8 t_1.$$

365 As a result, we can write down the selection differences of all the paired populations

366 in Fig. 1 as Eq. 2, where

$$367 \quad \mathbf{d} = (\Delta s_{2,1}t_3 \quad \Delta s_{3,1}t_1 \quad \Delta s_{4,1}t_1 \quad \Delta s_{5,1}t_1 \quad \Delta s_{3,2}t_1 \quad \Delta s_{4,2}t_1 \quad \Delta s_{5,2}t_1 \quad \Delta s_{4,3}t_2 \quad \Delta s_{5,3}t_2 \quad \Delta s_{5,4}t_3)^T$$

$$368 \quad \boldsymbol{\delta} = (\delta_1 \quad \delta_2 \quad \delta_3 \quad \delta_4 \quad \delta_5 \quad \delta_6 \quad \delta_7 \quad \delta_8)^T,$$

369 and

$$370 \quad \mathbf{T} = \begin{pmatrix} -t_3 & t_3 & 0 & 0 & 0 & 0 & 0 & 0 \\ -t_3 & 0 & t_2 & 0 & 0 & 0 & -t_1 & t_1 \\ -t_3 & 0 & 0 & t_3 & 0 & t_2 & -t_1 & t_1 \\ -t_3 & 0 & 0 & 0 & t_3 & t_2 & -t_1 & t_1 \\ 0 & -t_3 & t_2 & 0 & 0 & 0 & -t_1 & t_1 \\ 0 & -t_3 & 0 & t_3 & 0 & t_2 & -t_1 & t_1 \\ 0 & -t_3 & 0 & 0 & t_3 & t_2 & -t_1 & t_1 \\ 0 & 0 & -t_2 & t_3 & 0 & t_2 & 0 & 0 \\ 0 & 0 & -t_2 & 0 & t_3 & t_2 & 0 & 0 \\ 0 & 0 & 0 & -t_3 & t_3 & 0 & 0 & 0 \end{pmatrix}.$$

371 Here, the superscript  $T$  denotes the transpose of a vector. Because this system has  
 372 collinearity, we chose four independent equations, and assumed that one population  
 373 majorly contributes to the selection difference between any paired populations  $i$  and  $j$   
 374 sharing the most recent common ancestor  $k$ :

$$375 \quad \delta_i \delta_j = 0.$$

376 We also assumed  $\delta_7 = 0$ , because the ancestral African population before the  
 377 divergence of East and West Africans stayed in the same environment. Its selective  
 378 pressure should be the same as that of the population in the root. In other words,  
 379  $s_7 = s_0$ . Therefore, we can transform Eq. 2 into Eq. 3 and solve it in R 3.2.0 (80) with  
 380 RStudio 1.0.136 (81).

381 **Optimal solution.** Under neutral evolution (NE), we considered each estimated  $\delta$  as  
382 an independent random variable following a normal distribution with zero mean and  
383  $\sigma^2$  variance. For each solution with four variables, the summation below follows a  
384 chi-square distribution with four degrees of freedom:

385 
$$\frac{1}{\sigma^2} \sum_i \delta_i^2 \sim \chi^2(4).$$

386 Therefore, we have  $\Pr(|\delta|^2 > |\delta_a|^2 \mid NE) \geq \Pr(|\delta|^2 > |\delta_b|^2 \mid NE)$ , if  $|\delta_a|^2 \leq |\delta_b|^2$  for  
387 solutions  $a$  and  $b$ . In other words, we can choose the most conservative solution with  
388 the least deviation from neutral evolution using a probabilistic approach.

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## 403 **Author Contributions**

404 Y.H. designed the study. X.H. and Y.H. developed the model, analyzed the data, and  
405 wrote the manuscript. L.J. and S.W. revised the manuscript. L.J. and Y.H. jointly  
406 supervised the study.

407

## 408 **References**

- 409 1. Darwin C (1889) *The Descent of Man, and Selection in Relation to Sex*. (Appleton  
410 and Company, New York).
- 411 2. Norton HL (2005) Human skin pigmentation variation: a phenotypic, genotypic  
412 and evolutionary perspective. Ph.D. Dissertation, Pennsylvania State University.  
413 Retrieved from <https://etda.libraries.psu.edu/catalog/6658>. Last visited date:  
414 2017-02-11.
- 415 3. Rees JL, Harding RM (2012) Understanding the evolution of human pigmentation:  
416 recent contributions from population genetics. *J Invest Dermatol* 132(3):846–853.
- 417 4. Parra EJ (2007) Human pigmentation variation: evolution, genetic basis, and  
418 implications for public health. *Am J Phys Anthropol Suppl* 45:85–105.
- 419 5. Greaves M (2014) Was skin cancer a selective force for black pigmentation in  
420 early hominin evolution? *P R Soc B* 281(1781):20132055.
- 421 6. Loomis WF (1967) Skin-pigment regulation of vitamin-D biosynthesis in man.  
422 *Science* 157(3788):501–506.
- 423 7. Jablonski NG, Chaplin G (2000) The evolution of human skin coloration. *J Hum*  
424 *Evol* 39(1):57–106.
- 425 8. Izagirre N, García I, Junquera C, de la Rúa C, Alonso S (2006) A scan for  
426 signatures of positive selection in candidate loci for skin pigmentation in humans.  
427 *Mol Biol Evol* 23(9):1697–1706.

- 428 9. McEvoy B, Beleza S, Shriver MD. (2006) The genetic architecture of normal  
429 variation in human pigmentation: an evolutionary perspective and model. *Hum*  
430 *Mol Genet* 15(Spec No 2):R176–R181.
- 431 10. Voight BF, Kudravalli S, Wen X, Pritchard JK (2006) A map of recent positive  
432 selection in the human genome. *PLoS Biol* 4(3):e72.
- 433 11. Lao O, de Gruijter JM, van Duijin K, Navarro A, Kayser M (2007) Signatures of  
434 positive selection in genes associated with human skin pigmentation as revealed  
435 from analyses of single nucleotide polymorphisms. *Ann Hum Genet*  
436 71(3):354–369.
- 437 12. Myles S, Somel M, Tang K, Kelso J, Stoneking M (2007) Identifying genes  
438 underlying skin pigmentation differences among human populations. *Hum Genet*  
439 120(5):613–621.
- 440 13. Norton HL, et al. (2007) Genetic evidence for the convergent evolution of light  
441 skin in Europeans and East Asians. *Mol Biol Evol* 24(3):710–722.
- 442 14. Pickrell J, et al. (2009) Signals of recent positive selection in a worldwide sample  
443 of human populations. *Genome Res* 19(5):826–837.
- 444 15. Donnelly MP, et al. (2012) A global view of the *OCA2-HERC2* region and  
445 pigmentation. *Hum Genet* 131(5):683–696.
- 446 16. Hider JL, et al. (2013) Exploring signatures of positive selection in pigmentation  
447 candidate genes in populations of East Asian ancestry. *BMC Evol Biol* 13:150.



- 448 17. Wilde S, et al. (2014) Direct evidence for positive selection of skin, hair, and eye  
449 pigmentation in Europeans during the last 5,000 y. *Proc Natl Acad Sci USA*  
450 111(13):4832–4837.
- 451 18. Field Y, et al. (2016) Detection of human adaptation during the past 2000 years.  
452 *Science* 354(6313):760–764.
- 453 19. Abdel-Malek ZA, et al. (2001) The melanocortin 1 receptor is the principal  
454 mediator of the effects of agouti signaling protein on mammalian melanocytes. *J*  
455 *Cell Biol* 114(5):1019–1024.
- 456 20. Rebbeck TR, et al. (2002) *P* gene as an inherited biomarker of human eye color.  
457 *Cancer Epidem Biomar* 11(8):782–784.
- 458 21. Duffy DL, et al. (2004) Interactive effects of MC1R and OCA2 on melanoma risk  
459 phenotypes. *Hum Mol Genet* 13(4):447–461.
- 460 22. Duffy DL, et al. (2007) A three-single-nucleotide polymorphism haplotype in  
461 intron 1 of *OCA2* explains most human eye-color variation. *Am J Hum Genet*  
462 80(2):241–252.
- 463 23. Graf J, Hodgson R, van Daal A (2005) Single nucleotide polymorphisms in the  
464 *MATP* gene are associated with normal human pigmentation variation. *Hum Mutat*  
465 25(3):278–284.
- 466 24. Sulem P, et al. (2007) Genetic determinants of hair, eye and skin pigmentation in  
467 Europeans. *Nat Genet* 39(12):1443–1452.

- 468 25. Sulem P, et al. (2008) Two newly identified genetic determinants of pigmentation  
469 in Europeans. *Nat Genet* 40(7):835–837.
- 470 26. Anno S, Abe T, Yamamoto T (2008) Interactions between SNP alleles at multiple  
471 loci contribute to skin color differences between Caucasoid and Mongoloid  
472 subjects. *Int J Biol Sci* 4(2):81–86.
- 473 27. Han J, et al. (2008) A genome-wide association study identifies novel alleles  
474 associated with hair color and skin pigmentation. *PLoS Genet* 4(5):e1000074.
- 475 28. Ito S, Wakamatsu K (2008) Chemistry of mixed melanogenesis - pivotal roles of  
476 dopaquinone. *Photochem Photobiol* 84(3):582–592.
- 477 29. Kayser M, et al. (2008) Three genome-wide association studies and a linkage  
478 analysis identify *HERC2* as a human iris color gene. *Am J Hum Genet*  
479 82(2):411–423.
- 480 30. Sturm RA, Duffy DL (2012) Human pigmentation genes under environmental  
481 selection. *Genome Biol* 13(9):248.
- 482 31. Visser M, Kayser M, Palstra R (2012) *HERC2* rs12913832 modulates human  
483 pigmentation by attenuating chromatin-loop formation between a long-range  
484 enhance and the *OCA2* promoter. *Genome Res* 22(3):446–455.
- 485 32. Visser M, Palstra R, Kayser M (2014) Human skin color is influenced by an  
486 intergenic DNA polymorphism regulating transcription of the nearby *BNC2*  
487 pigmentation gene. *Hum Mol Genet* 23(21):5750–5762.

- 488 33. Guenther CA, Tasic B, Luo L, Bedell MA, Kingsley DM (2014) A molecular basis  
489 for classic blond hair color in Europeans. *Nat Genet* 46(7):748–754.
- 490 34. He Y, et al. (2015) A probabilistic method for testing and estimating selection  
491 differences between populations. *Genome Res* 25(12):1903–1909.
- 492 35. Jablonski NG, Chaplin G (2010) Human skin pigmentation as an adaptation to UV  
493 radiation. *Proc Natl Acad Sci USA* 107(Suppl 2):8962–8968.
- 494 36. Lamason RL, et al. (2005) SLC24A5, a putative cation exchanger, affects  
495 pigmentation in zebrafish and humans. *Science* 310(5755):1782–1786.
- 496 37. Edwards M, et al. (2010) Association of the *OCA2* polymorphism His615Arg with  
497 melanin content in East Asian Populations: further evidence of convergent  
498 evolution of skin pigmentation. *PLoS Genet* 6(3):e1000867.
- 499 38. Yang Z, et al. (2016) A genetic mechanism for convergent skin lightening during  
500 recent human evolution. *Mol Biol Evol* 33(5):1177–1187.
- 501 39. Jannot AS, et al. (2005) Allele variations in the *OCA2* gene (pink-eyed-dilution  
502 locus) are associated with genetic susceptibility to melanoma. *Eur J Hum Genet*  
503 13(8):913–920.
- 504 40. Mathieson I, et al. (2015) Genome-wide patterns of selection in 230 ancient  
505 Eurasians. *Nature* 528(7583):499–503.
- 506 41. Slatkin M, Racimo F (2016) Ancient DNA and human history. *Proc Natl Acad Sci*  
507 *USA* 113(23):6380–6387.

- 508 42. Beleza S, et al. (2013) The timing of pigmentation lightening in Europeans. *Mol*  
509 *Biol Evol* 30(1):24–35.
- 510 43. Nakagome S, et al. (2016) Estimating the ages of selection signals from different  
511 epochs in human history. *Mol Biol Evol* 33(3):657–669.
- 512 44. Vachtenheim J, Borovanský J (2010) "Transcription physiology" of pigment  
513 formation in melanocytes: central role of MITF. *Exp Dermatol* 19(7):617–627.
- 514 45. San-Jose LM, et al. (2017) *MC1R* variants affect the expression of melanocortin  
515 and melanogenic genes and the association between melanocortin genes and  
516 coloration. *Mol Ecol* 26(1):259–276.
- 517 46. Branicki W, Brudnik U, Wojas-Pelc A (2009) Interactions between *HERC2*, *OCA2*  
518 and *MC1R* may influence human pigmentation phenotype. *Ann Hum Genet*  
519 73(2):160–170.
- 520 47. Pośpiech E, et al. (2014) The common occurrence of epistasis in the determination  
521 of human pigmentation and its impact on DNA-based pigmentation phenotype  
522 prediction. *Forensic Sci Int Gen* 11:64–72.
- 523 48. Rees JL (2003) Genetics of hair and skin color. *Annu Rev Genet* 37:67–90.
- 524 49. Harris EE, Meyer D (2006) The molecular signature of selection underlying  
525 human adaptations. *Am J Phys Anthropol Suppl* 43:89–130.
- 526 50. Rana BK, et al. (1999) High polymorphism at the human melanocortin 1 receptor  
527 locus. *Genetics* 151(4):1547–1557.

- 528 51. Harding RM, et al. (2000) Evidence for variable selective pressures at MC1R. *Am*  
529 *J Hum Genet* 66(4):1351–1361.
- 530 52. Martínez-Cadenas C, et al. (2013) Simultaneous purifying selection on the  
531 ancestral *MC1R* allele and positive selection on the melanoma-risk allele V60L in  
532 south Europeans. *Mol Biol Evol* 30(12):2654–2665.
- 533 53. Wasmeier C, Hume AN, Bolasco G, Seabra MC (2008) Melanosomes at a glance.  
534 *J Cell Sci* 121(24):3995–3999.
- 535 54. Valencia JC, Hearing VJ (2012) The role of glycosylation in the control of  
536 processing and cellular transport of the functional amyloid PMEL17.  
537 *Glycosylation*, ed. Petrescu (In Tech, Rijeka). Chapter 15.
- 538 55. Wu B, Guo W (2015) The exocyst at a glance. *J Cell Sci* 128(16):2957–2964.
- 539 56. Xu X, Thörnwall M, Lundin L, Chhajlani V (1996) Val92Met variant of the  
540 melanocyte stimulating hormone receptor gene. *Nat Genet* 14(4):384.
- 541 57. Motokawa T, Kato T, Hashimoto Y, Katagiri T (2007) Effect of Val92Met and  
542 Arg163Gln variants of the *MC1R* gene on freckles and solar lentigines in Japanese.  
543 *Pigment Cell Res* 20(2):140–143.
- 544 58. Makova KD, Ramsay M, Jenkins T, Li WH (2001) Human DNA sequence  
545 variation in a 6.6-kb region containing the melanocortin 1 receptor promoter.  
546 *Genetics* 158(3):1253–1268.
- 547 59. Kimura M, Weiss GH (1964) The stepping stone model of population structure

- 548 and the decrease of genetic correlation with distance. *Genetics* 49(4):561–576.
- 549 60. Li JZ, et al. (2008) Worldwide human relationships inferred from genome-wide  
550 patterns of variation. *Science* 319(5866):1100–1104.
- 551 61. Teo YY, et al. (2009) Singapore genome variation project: a haplotype map of  
552 three Southeast Asian populations. *Genome Res* 19(11):2154–2162.
- 553 62. The 1000 Genomes Project Consortium (2010) A map of human genome variation  
554 from population-scale sequencing. *Nature* 467(7319):1061–1073.
- 555 63. The International HapMap 3 Consortium (2010) Integrating common and rare  
556 genetic variation in diverse human populations. *Nature* 467(7311):52–58.
- 557 64. Behar DM, et al. (2010) The genome-wide structure of the Jewish people. *Nature*  
558 466(7303):238–242.
- 559 65. Rasmussen M, et al. (2010) Ancient human genome sequence of an extinct  
560 Palaeo-Eskimo. *Nature* 463(7282):757–762.
- 561 66. Metspalu M, et al. (2011) Shared and unique components of human population  
562 structure and genome-wide signals of positive selection in South Asia. *Am J Hum*  
563 *Genet* 89(6):731–744.
- 564 67. Pagani L, et al. (2012) Ethiopian genetic diversity reveals linguistic stratification  
565 and complex influences on the Ethiopian gene pool. *Am J Hum Genet*  
566 91(1):83–96.
- 567 68. Yunusbayev B, et al. (2012) The Caucasus as an asymmetric semipermeable

- 568 barrier to ancient human migrations. *Mol Biol Evol* 29(1):359–365.
- 569 69. Yunusbayev B, et al. 2015. The genetic legacy of the expansion of  
570 Turkic-speaking nomads across Eurasia. *PLoS Genet* 11(4):e1005068.
- 571 70. Cristofaro JD, et al. (2013) Afghan Hindu Kush: where Eurasian sub-continent  
572 gene flows converge. *PLoS One* 8(10):e76748.
- 573 71. Fedorova SA, et al. (2013) Autosomal and uniparental portraits of the naive  
574 populations of Sakha (Yakutia): implications for the peopling of Northeast Eurasia.  
575 *BMC Evol Biol* 13:127.
- 576 72. Xing J, et al. (2013) Genomic analysis of natural selection and phenotypic  
577 variation in high-altitude Mongolians. *PLoS Genet* 9(7):e1003634.
- 578 73. Kovacevic L, et al. (2014) Standing at the gateway to Europe - the genetic  
579 structure of Western Balkan populations based on autosomal and haploid markers.  
580 *PLoS One* 9(8):e105090.
- 581 74. Raghavan M, et al. (2014) Upper Paleolithic Siberian genome reveals dual  
582 ancestry of Native Americans. *Nature* 505(7481):87–91.
- 583 75. Purcell S, et al. (2007) PLINK: a toolset for whole-genome association and  
584 population-based linkage analysis. *Am J Hum Genet* 81(3):559–575.
- 585 76. Patterson N, Price AL, Reich D (2006) Population structure and eigenanalysis.  
586 *PLoS Genet* 2(12):e190.
- 587 77. Price AL, et al. (2006) Principal components analysis corrects for stratification in

- 588 genome-wide association studies. *Nat Genet* 38(8):904–909.
- 589 78. Browning SR, Browning BL (2007) Rapid and accurate haplotype phasing and  
590 missing data inference for whole genome association studies by use of localized  
591 haplotype clustering. *Am J Hum Genet* 81(5):1084–1097.
- 592 79. Browning BL, Browning SR (2016) Genotype imputation with millions of  
593 reference samples. *Am J Hum Genet* 98(1):116–126.
- 594 80. R Core Team (2015) R: A language and environment for statistical computing. R  
595 Foundation for Statistical Computing, Vienna, Austria. URL:  
596 <http://www.R-project.org/>.
- 597 81. RStudio Team (2016) RStudio: Integrated development for R. RStudio, Inc.,  
598 Boston, MA. URL: <http://www.rstudio.com/>.  
599



## 600 **Figure Legends**

601 **Fig. 1. Modeling selective pressures with multiple population phylogeny.** We  
602 modeled the evolutionary history of five representative human populations as a binary  
603 tree. The details of demographic events were ignored. Here,  $s_i$  ( $i = 0, 1, \dots, 8$ ) denotes  
604 the selection coefficient of the  $i$ -th epoch.  $\delta_i$  ( $i = 1, 2, \dots, 8$ ) denotes the selection  
605 change of the  $i$ -th epoch, and can be obtained by estimating selection differences  
606 between paired populations. The numbers on the branches indicate different epochs.  
607 In the present study, we assumed that the divergence time of separation between  
608 Africans and Eurasians was  $\sim 3600$  generations ago; the divergence time of separation  
609 between Europeans and Asians was  $\sim 3000$  generations ago; the divergence time of  
610 separation between Siberians and East Asians was  $\sim 2000$  generations ago; and the  
611 divergence time of separation between East and West Africans was  $\sim 2000$  generations  
612 ago.

613 **Fig. 2. The overall selection differences in multiple loci between populations.** We  
614 summarized the overall selection differences in 15 loci for paired representative  
615 populations using genotype data from 15 public datasets. Red color (positive numbers)  
616 indicates selective pressures of populations in rows are larger than those in columns;  
617 blue color (negative numbers) indicates selective pressures of populations in rows are  
618 smaller than those in columns. Populations are abbreviated as follows: WAF, West  
619 Africans; EAF, East Africans; EUR, Europeans; SIB, Siberians; EAS, East Asians.

620 **Fig. 3. Historical changes of selective pressures on human pigmentation during**

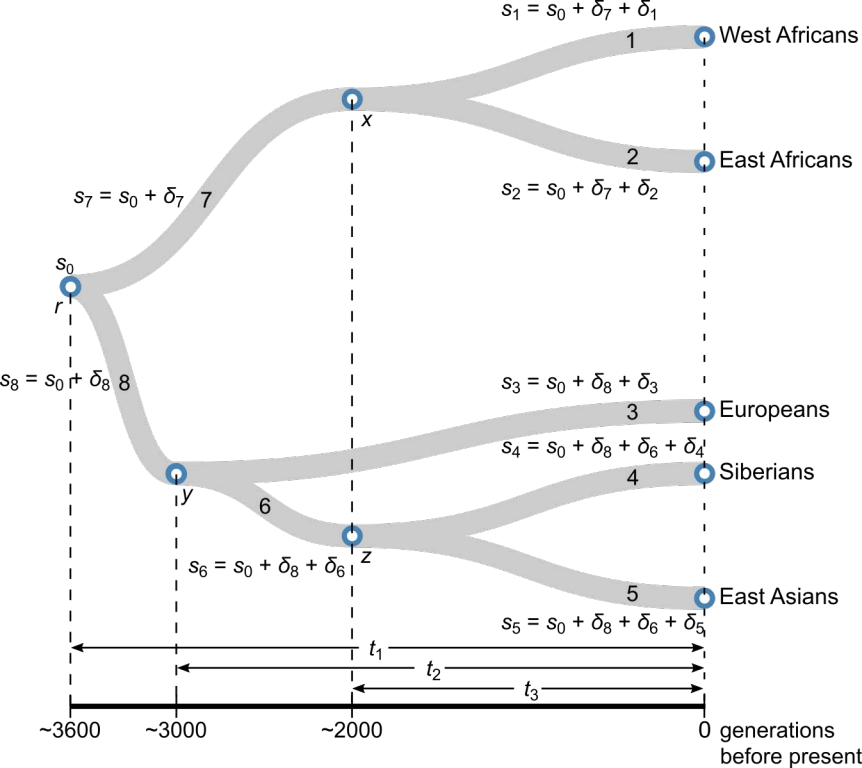
621 **different epochs of human migration.** We determined historical changes of selective  
622 pressures on human pigmentation during different epochs by solving the non-linear  
623 equations developed in this study. The most conservative solution was chosen to  
624 represent the possible historical selection on the world map. We observed not only a  
625 recent incremental change of selective pressure in modern Europeans, but also a  
626 significant historical increase of selective pressure on light pigmentation in all  
627 Eurasians during the out-of-Africa event. Here,  $s_0$  is the selection coefficient of the  
628 ancestral population for all modern human populations. The numbers are the selection  
629 changes ( $\times 10^{-2}$ /generation) during different epochs. Zero changes are ignored. The  
630 arrows indicate the direction of human migration, and their color gradient indicates  
631 the trends of human skin color.

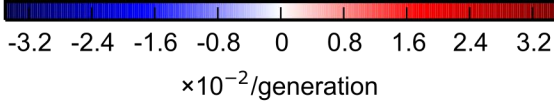
632 **Fig. 4. Selection differences in individual loci between populations.** We used Eq. 1  
633 to quantify the selection differences of 31 SNPs associated with human pigmentation,  
634 and categorized them into four kinds of selection patterns: (A) Eurasian-shared  
635 pattern; (B) European-specific pattern; (C) Asian-specific pattern; and (D) Other. Red  
636 color (positive numbers) indicates selective pressures of populations in rows are  
637 larger than those in columns; blue color (negative numbers) indicates selective  
638 pressures of populations in rows are smaller than those in columns. All alleles are in  
639 the forward strand of the Human Reference Genome, and the arrows indicate  
640 substitutions from ancestral to derived alleles. Populations are abbreviated as follows:  
641 1, West Africans; 2, East Africans; 3, Europeans; 4, Siberians; 5, East Asians.

642 **Fig. 5. Human pigmentation genes under different population-specific selection**

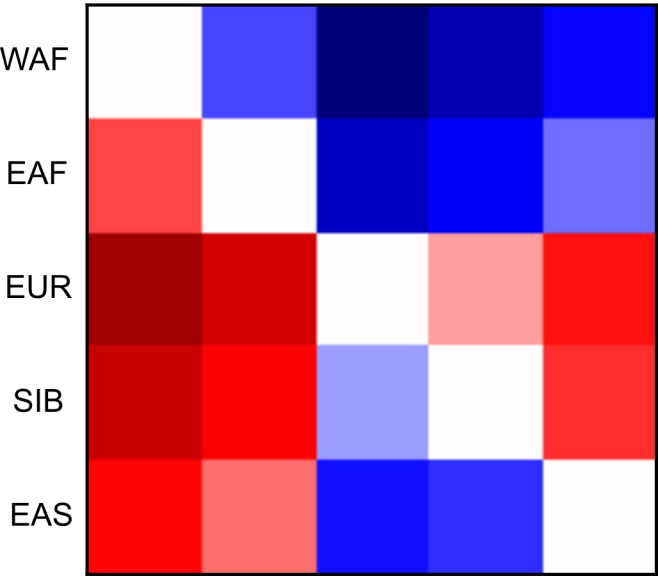
643 **in the melanogenic pathway.** We placed human pigmentation genes affected by  
644 population-specific selection into the melanogenic pathway. Two critical genes, *ASIP*  
645 and *MC1R*, are the major regulators upstream of the melanogenic pathway. The  
646 largest selection differences of these two genes were between Africans and Eurasians,  
647 indicating that epistasis plays important roles in the evolution of human pigmentation.  
648 Most of the remaining genes are downstream of the melanogenic pathway. The  
649 melanogenic pathway is based on previous publications (4, 30–32, 53–55). The  
650 arrows indicate the direction of the melanogenic pathway.

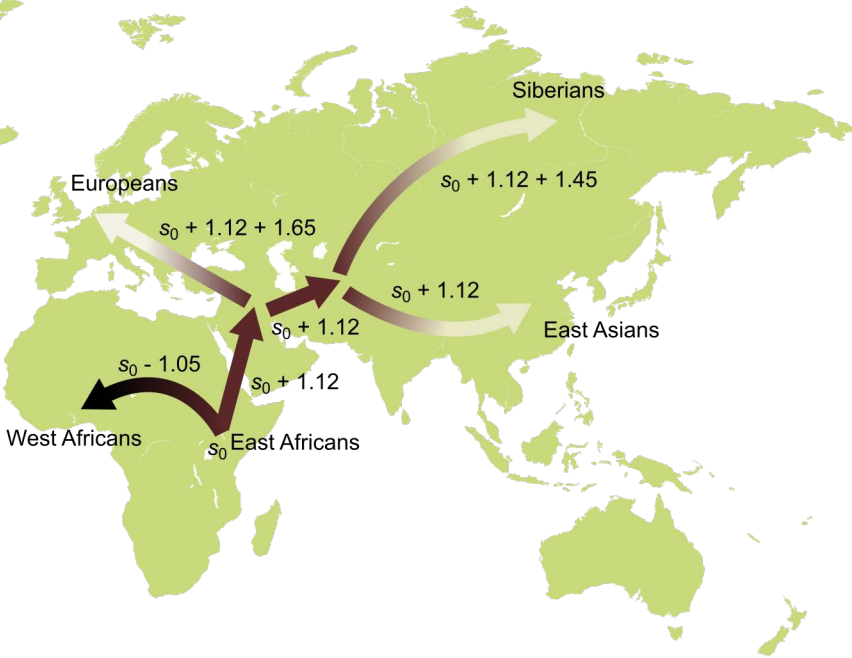
651



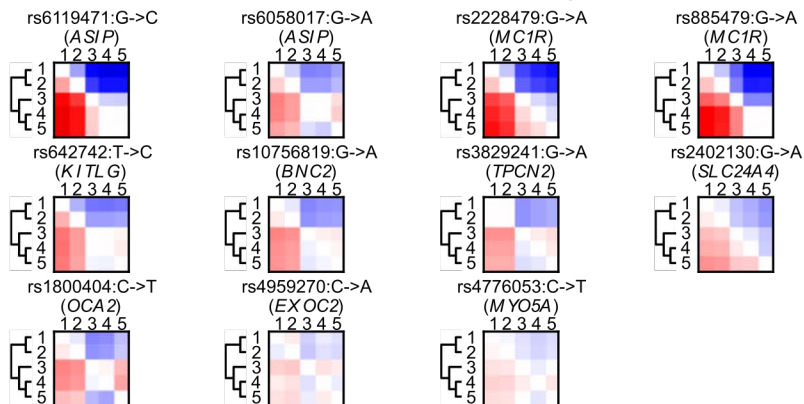
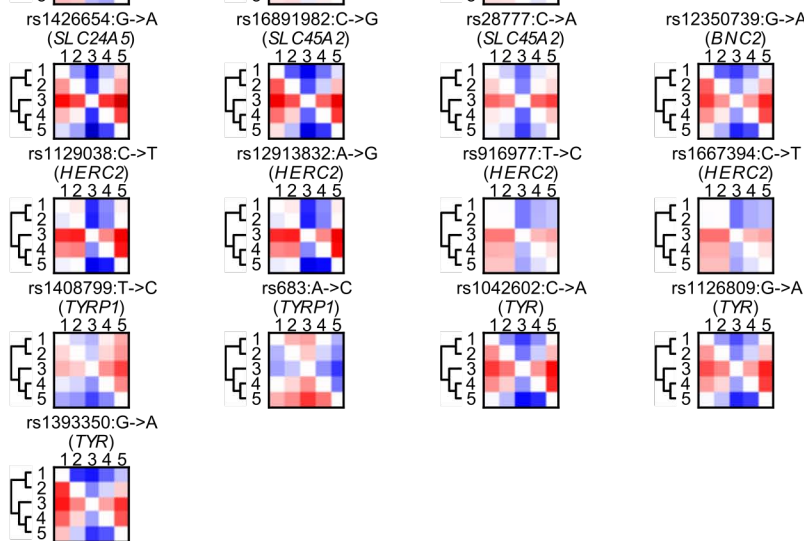
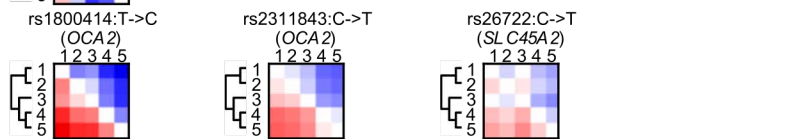
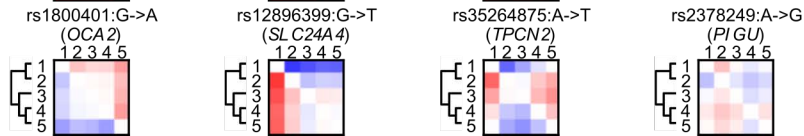


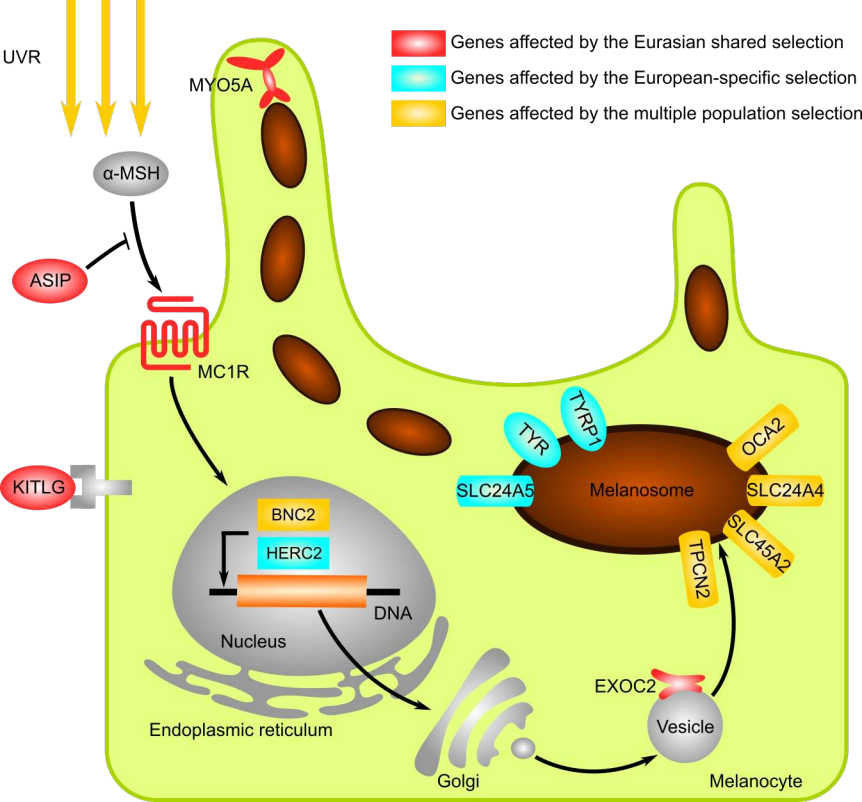
WAF      EAF      EUR      SIB      EAS





-4 -3.2 -2.4 -1.6 -0.8 0 0.8 1.6 2.4 3.2 4

 $\times 10^{-3}/\text{generation}$ **A****B****C****D**





**Table 1. Solutions of historical selection (coefficient) changes during different epochs ( $\times 10^{-2}$ /generation).**

Solution	$\delta_1$	$\delta_2$	$\delta_3$	$\delta_4$	$\delta_5$	$\delta_6$	$\delta_7$	$\delta_8$
#1	-1.28	0	0.68	0	-1.42	0	0	1.78
#2	-1.28	0	1.63	1.42	0	0	0	0.99
#3	-1.28	0	0	0	-1.42	-0.68	0	2.35
#4	-1.28	0	0	1.42	0	-1.63	0	2.35
#5	0	1.28	0	0	-1.42	-0.68	0	3.06
#6	0	1.28	0	1.42	0	-1.63	0	3.06
#7	0	1.28	0.68	0	-1.42	0	0	2.49
#8	0	1.28	1.63	1.42	0	0	0	1.70

We applied our new approach to dissect historical changes of selective pressures in a multiple population model by summarizing selective pressures on multiple human pigmentation genes. Here,  $\delta_i$  ( $i = 1, 2, \dots, 8$ ) denotes the selection change of the  $i$ -th epoch, as shown in Fig. 1. A positive change indicates a stronger directional selection on derived alleles than before; a negative change suggests a weaker positive selection on derived alleles than before. Further, we chose solution #1 as the most conservative solution using a probabilistic approach.