1 Population genomics of pneumococcal carriage in Massachusetts children following PCV-

2 **13 introduction**

3 Mitchell PK¹, Azarian T¹, Croucher NJ, Callendrello A¹, Thompson CM¹, Pelton SI²,

4 Lipsitch M¹, WP Hanage¹

1 Center for Communicable Disease Dynamics, Department of Epidemiology, T.H. Chan School of Public
 Health, Harvard University, Boston, MA; 2 MRC Centre for Outbreak Analysis and Modelling, Department

of Infectious Disease Epidemiology, Imperial College London, London, W2 1PG, UK; 3 Division of

8 Pediatric Infectious Diseases, Maxwell Finland Laboratory for Infectious Diseases, Boston Medical

- 9 Center, Boston, MA
- 10
- 11 Patrick Mitchell mitchell.patrick.k@gmail.com
- 12 Taj Azarian <u>Tazarian@hsph.harvard.edu</u>
- 13 Nick J Croucher <u>n.croucher@imperial.ac.uk</u>
- 14 Alanna Callendrello <u>alcallendrello@gmail.com</u>
- 15 Claudette M Thompson <u>cthompso@hsph.harvard.edu</u>
- 16 Stephen I Pelton <u>spelton@bu.edu</u>
- 17 Marc Lipsitch mlipsitc@hsph.harvard.edu
- 18 Bill Hanage <u>whanage@hsph.harvard.edu</u>
- 19
- 20
- 21 **Corresponding Author:**
- 22 Bill Hanage
- 23 Center for Communicable Disease Dynamics,
- 24 Harvard T.H. Chan School of Public Health,
- 25 677 Huntington Avenue, Suite 506, Boston, MA 02115
- 26 <u>whanage@hsph.harvard.edu</u>

27 Background

28	The 13-valent pneumococcal conjugate vaccine (PCV-13) was introduced in the United States in
29	2010. Using a large pediatric carriage sample collected from shortly after the introduction of
30	PCV-7 to several years after the introduction of PCV-13, we investigate alterations in the
31	composition of the pneumococcal population following the introduction of PCV-13, evaluating
32	the extent to which the post-vaccination non-vaccine type (NVT) population mirrors that from
33	prior to vaccine introduction and the effect of PCV-13 on vaccine type lineages.

34 Methods and Findings

35 Draft genome assemblies from 736 newly sequenced and 616 previously published

36 pneumococcal carriages isolates from children in Massachusetts between 2001 and 2014 were

analyzed. Isolates were classified into one of 22 sequence clusters (SCs) on the basis of their

core genome sequence. We calculated the SC diversity for each sampling period as the

39 probability that any two randomly drawn isolates from that period belong to different SCs. The

40 sampling period immediately after the introduction of PCV-13 (2011) was found to have higher

41 diversity than preceding (2007) or subsequent (2014) sampling periods (Simpson's D 2007:

42 0.915 95% CI [0.901, 0.929]; 2011: 0.935 [0.927, 0.942]; 2014: 0.912 [0.901, 0.923]). Amongst

43 NVT isolates, we found the distribution of SCs in 2011 to be significantly different from that in

44 2007 or 2014 (Fisher's Exact Test p=0.018, 0.0078), but did not find a difference comparing

45 2007 to 2014 (Fisher's Exact Test p=0.24), indicating greater similarity between samples

46 separated by a longer time period than between samples from closer time periods. We also found

47 changes in the accessory gene content of the NVT population between 2007 and 2011 to have

48 been reduced by 2014. Amongst the new serotypes targeted by PCV-13, four were present in our

49 sample. The proportion of our sample composed of PCV-13-only vaccine serotypes 19A, 6C,

and 7F decreased between 2007 and 2014, but no such reduction was seen for serotype 3. We

- 51 did, however, observe differences in the genetic composition of the pre- and post-PCV-13
- 52 serotype 3 population. Our isolates were collected during discrete sampling periods from a small
- 53 geographic area, which may limit the generalizability our findings.

54 Conclusion

- 55 Pneumococcal diversity increased immediately following the introduction of PCV-13, but
- subsequently returned to pre-vaccination levels. This is reflected in the distribution of NVT
- 57 lineages, and, to a lesser extent, their accessory gene frequencies. As such, there may be a period
- 58 during which the population is particularly disrupted by vaccination before returning to a more
- 59 stable distribution. The persistence and shifting genetic composition of serotype 3 is a concern
- 60 and warrants further investigation.

61 INTRODUCTION

62	Streptococcus pneumoniae is a common bacterial colonizer of the human nasopharynx,
63	particularly among children ¹ . In Massachusetts, it has consistently been found in approximately
64	30% of children under the age of 7 between 2001 and 2011^2 . While colonization rarely
65	progresses beyond asymptomatic carriage, the ubiquity of the pneumococcus leads to a
66	substantial burden of disease, causing an estimated 4 million disease episodes, including 445,000
67	hospitalizations and 22,000 deaths in the United States in 2004 ³ .
68	Conjugate vaccination has been a major advance in the reducing pneumococcal disease.
69	The seven-valent pneumococcal conjugate vaccine (PCV-7), introduced in the United States in
70	2000, was highly effective in reducing overall rates of pneumococcal disease, as vaccine type
71	(VT) pneumococci were responsible for the vast majority of cases ^{4–6} . Carriage of vaccine
72	serotypes also declined, though overall carriage prevalence remained roughly constant due to
73	serotype replacement ^{2,7,8} .
73 74	serotype replacement ^{2,7,8} . Despite lower overall rates of pneumococcal disease, increases were seen in the incidence
74	Despite lower overall rates of pneumococcal disease, increases were seen in the incidence
74 75	Despite lower overall rates of pneumococcal disease, increases were seen in the incidence of disease due to the replacement non-vaccine type (NVT) population. Serotype 19A in
74 75 76	Despite lower overall rates of pneumococcal disease, increases were seen in the incidence of disease due to the replacement non-vaccine type (NVT) population. Serotype 19A in particular became a significant cause of invasive disease ^{5,9,10} . The thirteen-valent vaccine (PCV-
74 75 76 77	Despite lower overall rates of pneumococcal disease, increases were seen in the incidence of disease due to the replacement non-vaccine type (NVT) population. Serotype 19A in particular became a significant cause of invasive disease ^{5,9,10} . The thirteen-valent vaccine (PCV- 13), introduced in 2010, extended coverage to six additional serotypes, including 19A, beyond
74 75 76 77 78	Despite lower overall rates of pneumococcal disease, increases were seen in the incidence of disease due to the replacement non-vaccine type (NVT) population. Serotype 19A in particular became a significant cause of invasive disease ^{5,9,10} . The thirteen-valent vaccine (PCV- 13), introduced in 2010, extended coverage to six additional serotypes, including 19A, beyond those included in PCV-7, and has resulted in further reductions in pneumococcal disease ¹¹ . As
74 75 76 77 78 79	Despite lower overall rates of pneumococcal disease, increases were seen in the incidence of disease due to the replacement non-vaccine type (NVT) population. Serotype 19A in particular became a significant cause of invasive disease ^{5,9,10} . The thirteen-valent vaccine (PCV- 13), introduced in 2010, extended coverage to six additional serotypes, including 19A, beyond those included in PCV-7, and has resulted in further reductions in pneumococcal disease ¹¹ . As with PCV-7, however, overall carriage prevalence has not changed substantially ² . Worryingly,
74 75 76 77 78 79 80	Despite lower overall rates of pneumococcal disease, increases were seen in the incidence of disease due to the replacement non-vaccine type (NVT) population. Serotype 19A in particular became a significant cause of invasive disease ^{5,9,10} . The thirteen-valent vaccine (PCV- 13), introduced in 2010, extended coverage to six additional serotypes, including 19A, beyond those included in PCV-7, and has resulted in further reductions in pneumococcal disease ¹¹ . As with PCV-7, however, overall carriage prevalence has not changed substantially ² . Worryingly, serotype 3, a highly invasive serotype included in PCV-13, appears to have not declined as the

84	Pediatric pneumococcal carriage in Massachusetts has been extensively studied since
85	shortly after the introduction of PCV- 7^7 . The effects of vaccination can be seen both in the
86	prevalence of specific lineages as well as in broader population metrics. The apparent effects of
87	vaccination are variable depending on how the population is characterized and the timescale over
88	which it is examined. Serotype diversity was found to have increased then stabilized following
89	the introduction of PCV-7 ¹⁵ , reflecting the selective impact of vaccines and the period while
90	carriage replacement was taking place. Interestingly, minimal changes were found when
91	comparing the presence and absence of specific pneumococcal genes in this population between
92	2001 and 2007, suggesting that the overall genetic composition of the population was not much
93	changed other than in one of the loci conferring vaccine serotype 6B ¹⁶ . Another study
94	considering multilocus sequence type (MLST) profiles found no significant change in diversity
95	or population composition in the immediate aftermath of PCV-13 ¹⁷ . With more time elapsed
96	since PCV-13 introduction, it is possible to evaluate the longer-term effects of this vaccine.
97	Here we examine population-scale genetic changes in carriage pneumococci amongst
98	children in Massachusetts since the introduction of PCV-13. Using genomic sequencing data for
99	isolates collected between 2000 and 2014, we analyze alterations to the clonal composition,
100	defined on the basis of core genome variability, and gene content of the pneumococcal NVT

101 population following the introduction of PCV-13. Additionally, we evaluate whether serotype 3

102 pneumococci have declined and how they have changed through this time period.

103

104 METHODS

105 Sample Collection

106	Pneumococcal isolates were collected from nasopharyngeal swabs of children in
107	Massachusetts between October and April of 2000-01, 2003-04, 2006-07, 2008-09, 2010-11 and
108	2013-14 as previously described ^{2,7,8} . Each sampling season is referred to by the later year.
109	Pneumococcal genomes from the 2001, 2004, and 2007 sampling periods were previously
110	published and read data for these were obtained from ENA ¹⁶ . Isolates from 2009 through 2014
111	were sequenced from NexteraXT genomic libraries analyzed on an Illumina MiSeq to produce
112	paired-end 2x150 bp reads with a minimum depth of coverage of 30X.
113	Genomic Processing
113 114	Genomic Processing Draft assemblies were constructed using SPAdes v3.10 and annotated using Prokka
114	Draft assemblies were constructed using SPAdes v3.10 and annotated using Prokka
114 115	Draft assemblies were constructed using SPAdes v3.10 and annotated using Prokka v1.11 ^{18,19} . Assemblies not between 1.9 and 2.3 Mb were excluded from further analysis, as were
114 115 116	Draft assemblies were constructed using SPAdes v3.10 and annotated using Prokka $v1.11^{18,19}$. Assemblies not between 1.9 and 2.3 Mb were excluded from further analysis, as were those that produced fewer than 1900 annotated coding sequences (CDS). Roary v3.10.0 was then

119 **Typing**

Serotype was identified using the Quellung reaction as previously described and reported
 for all but the 2014 sample^{16,17,21}. Serotypes were checked using SRST2 v0.2.0 and a database
 constructed from 91 published sequences of the pneumococcal capsule biosynthetic locus²²⁻²⁴.

123 Phylogenetic Analysis

The core genome alignment generated by Roary was used to construct a phylogeny using FastTree v2.1.10²⁵. In order to identify clusters of related sequences (Sequence Clusters - SCs), three iterations of hierBAPS were run on the core genome alignment, setting the maximum cluster depth to 1 and maximum number of clusters to 30, 40, and 50²⁶.

128 Sequence Cluster Diversity

In order to determine the potential effect of PCV-13 on diversity in this population, we 129 calculated Simpson's D for each sampling period, for sequence clusters. This value, which 130 131 represents the probability that two randomly drawn isolates from a given sampling period belong to different SCs, was calculated as $D = \frac{N}{N-1} (1 - \sum_{i=1}^{m} x_i^2)$, where $x = \frac{n_i}{N}$, the fraction of isolates 132 in that year belonging to sequence cluster i and $\frac{N}{N-1}$ is a correction for finite sample size²⁷. 133 Following an earlier analysis of serotype diversity in this population, Welch's t-test was used to 134 compare the 2007 and 2011 populations and the 2011 and 2014 populations in order to test 135 whether SC diversity changed following the introduction of PCV-13¹⁵. The polyphyletic SC was 136 excluded from these calculations. 137

An increase in diversity would be expected if common lineages become more rare and rare lineages become more common. To estimate the expected change in diversity we would observe if there were a smooth transition between the 2007 and 2014 population, a series of composite diversities were calculated in which the proportion belonging to each SC was a weighted combination of the 2007 and 2014 value for that SC, with the weights for the two years summing to 1. The sample size correction factor, $\frac{N}{N-1}$, was similarly weighted.

144 The proportion of the population belonging to each SC and their rank order in the 145 population were determined. As diversity increases, the shape of this distribution would be

expected to flatten, with the most common lineages decreasing and the least common lineages
increasing.²⁸ In order to compare this distribution from the 2007 and 2014 sampling periods with
that from 2011, the frequency of each SC was plotted against its rank and overlaid with the
distribution from 2011. In order to determine which SCs became more or less common following
the introduction of PCV13, we conducted a Fisher's exact test for each SC comparing its
frequency between the 2007 and 2014 samples.

152 NVT Composition

To determine the clonal composition of the pre- and post-PCV-13 NVT population, the proportion of the NVT population belonging to each of the SCs identified by hierBAPS was calculated for 2007, 2011, and 2014. For the purpose of these analyses, serotype 6C was considered a PCV-13 type due to its cross-reactivity with serotype $6A^{29,30}$. Fisher's exact test was used to determine whether these proportions varied between each pairwise combination of these three sampling periods.

159 We then sought to determine if the gene content of the NVT population varied between sampling periods before and after the introduction of PCV-13. Logistic models were used 160 evaluate the extent to which individual genes became more or less common between 2007 and 161 2014, as well as between 2007 and 2011. Genes were excluded if they were universally present 162 or absent in either sampling period or present or absent in fewer than 5 total isolates between the 163 three sampling periods. For the set of genes included in both models, we calculated a linear fit 164 comparing the regression coefficients corresponding to the time periods from 2007 to 2011 and 165 2007 to 2014. 166

167 In order to determine whether changes in the gene content of the NVT population from 2007 to 2011 continued, stabilized, or reversed from 2011 to 2014, we compared the observed 168 data to hypothetical scenarios in which the 2014 population was purely reflective of the 169 170 population from either the earlier sampling periods. To do this, we drew with replacement a sample of the same size as the 2014 population from either the 2007 or 2011 population. Twenty 171 resampled populations were generated from each of 2007 and 2011, then used in place of the true 172 2014 population in the previous regression analyses. This process was repeated for an additional 173 twenty resampled populations drawn from the true 2014 population in order to gauge its 174 175 variability. This enabled us to evaluate the gene content of the 2014 population in relation to what would be expected if there was no overall change either from 2007 or from 2011. 176

177 Evaluation of Serotype 3

Previous studies have noted that PCV-13 may not be as effective against serotype 3 as it 178 is against the other serotypes included^{2,11,13,14}. We compared the proportion of the pneumococcal 179 180 population composed of serotype 3 between 2007, 2011, and 2014 in relation to the other three PCV-13 serotypes present in our sample, 19A, 7F, and 6C. We identified MLST profile of 181 serotype 3 isolates using as previously described¹⁶. We then used RAxML to construct a 182 phylogenetic tree based on the core genome of serotype 3 isolates to determine if the pre- and 183 post-PCV-13 populations were genetically distnict³¹. To assess nucleotide and amino acid 184 variation among capsular polysaccharide (CPS) loci, we mapped reads to the S. pneumoniae 185 OXC141 serotype 3 reference strain (NC 017592) using SMALT v0.7.6. Single nucleotide 186 polymorphisms (SNPs) were identified using SAMtools v1.3.1³². The CPS region spanning 187 nucleotides 343,104-356,408 (*dexB* – *aliA*) was abstracted and investigated for mutations. 188 Further, RAxML was used to construct a phylogeny of the CPS region. 189

190 **RESULTS**

191 Sample

A total of 1.352 isolates were included in the final analysis. The core genome consists of 192 1,000 genes found in at least 99% of isolates, producing an alignment 885 kb in length. A total of 193 10,941 genes were identified. Setting the maximum number of hierBAPS clusters to 30 and 40 194 produced identical results, with 21 clusters identified. With the maximum number of clusters set 195 to 50, an additional cluster was identified and another cluster was expanded. This resulted in 22 196 SCs, 21 of which were monophyletic and ranged in size from 14 to 177 isolates. The other, SC1, 197 contained 150 isolates belonging to multiple small clades or individual leaves throughout the tree 198 and should be interpreted as containing all lineages that could not be grouped, other than on the 199 200 basis of their lack of similarity to any other cluster [Fig 1].

201 **Diversity**

Sequence cluster diversity was calculated for each year using Simpson's D, excluding the polyphyletic cluster SC1. Diversity was significantly higher in 2011, the first sampling period following the introduction of PCV13, than it was in either 2007 or 2014, the adjacent periods for which data were available (2007 p=0.018, 2014 p=0.00098) [Fig 2a]. A similar increase was observed after the introduction of PCV-7. The weighted diversity estimate displayed the expected increase over either the 2007 or 2014 values, but was never as high as the diversity calculated for 2011 [Fig 2b].

After Bonferroni correction, only 3 SCs (SC3, SC9, and SC20) changed significantly in their share of the pneumococcal population between 2007 and 2014 (Fisher's exact test $p=0.0021, 0.0022, and 4.5 \times 10^{-6}$, respectively). SC3 became more common, increasing from 5.8%

212	of the 2007 sample to 13.4% of the 2014 sample, with serogroups 23 and 15 coming to
213	predominate over serogroup 6. Both SCs 9 and 20 are primarily composed of serotypes against
214	which PCV-13 afforded protection (7F and 6C, respectively) and were completely absent in the
215	2014 sample.

The overall shape of the frequency distribution was slightly flatter in 2011 as compared to 2007 and 2014, as would be expected from the higher diversity in that sampling period. Relatively rare SCs in particular were more common in the 2011 sample than the adjacent periods [Fig 2c,d].

220 NVT Composition

221 Non-PCV-13 types increased from 66.5% of the pneumococcal population in 2007 222 sampling period to 92.3% in the 2014 sampling period. Fifteen SCs had at least 1 NVT isolate. There was no significant difference between the SC distribution amongst NVTs in 2007 and 223 2014 (Fisher's exact test p=0.24). There was, however, a significant difference between 2007 224 225 and 2011 (p=0.0018) and between 2011 and 2014 (p=0.0078), indicating a bounce-back effect in which the population was disrupted in 2011 but returned to its pre-vaccination state by 2014. 226 Correspondingly, many of the common SCs that showed a distinct increase in there prevalence in 227 the NVT population between 2007 and 2011 decreased from 2011 to 2014 while those that 228 decreased between 2007 and 2011 increased from 2011 to 2014. [Fig 3]. 229

This bounce-back is partially reflected by the trend in gene content over time. The linear fit comparing the 2007-2011 and 2007-2014 regression coefficients for each gene had a slope of 0.62, indicating less overall change between 2007 and 2014 than between 2007 and 2011. This slope fell between those from hypothetical 2014 populations drawn from either 2007 or 2011,

234	which clustered around a slope of 0 and 1, respectively [Fig. 4]. This indicates that while the
235	direction in which genes changed in frequency from 2007 to 2011 was generally preserved
236	through 2014, the trend was partially counteracted between 2011 and 2014 with genes returning
237	closer to their 2007 levels prior to the introduction of PCV-13.

- 238 **Persistence of Serotype 3**
- In order to evaluate whether the whether the new serotypes included in PCV-13

decreased following its introduction, we conducted a Fisher's exact test comparing the 2007 and

241 2014 carriage share of serotypes 19A, 6C, 7F, and 3. While serotypes 19A, 6C and 7F all showed

- significant reductions between the two time periods (p<0.0001, p=0.00014, p=0.0011,
- respectively), serotype 3 had no such change (p=0.46) [Fig 5].

To test if the persistence of serotype 3 may be related to some genetic factor, we assessed 244 population structure and CPS nucleotide variation. All of the serotype 3 isolates clustered in the 245 same SC and were MLST sequence type (ST) 180 belonging to the Netherlands³-31 (PMEN31) 246 clone CC180. While all isolates clustered into the same SC, there was a distinct bifurcation in the 247 phylogeny [Fig 6]. Of the 28 serotype 3 isolates, 16 fell into one subclade and 12 into the other. 248 In the larger subclade, 4 isolates (25%) are from 2011 or 2014, after the introduction of PCV-13. 249 The other subclade contains 11 (92%) post-PCV-13 isolates (γ^2 p=0.0018). Further assessment of 250 CPS showed low nucleotide diversity [mean pairwise SNP distance:1.5 (S.E. 0.7)] and only four 251 polymorphic amino acids, none of which segregated the subclades. However, the CPS phylogeny 252 recapitulated the bifurcation in the core genome phylogeny, with all isolates belonging to the 253 post-PCV-13 subclade displaying as highly clustered. 254

255 **DISCUSSION**

256 Here we have analyzed a sample of carriage pneumococci collected in Massachusetts 257 between the winters of 2000-01 and 2013-14, focusing primarily on changes occurring following 258 the introduction of PCV-13. Using genomic data, we find that the NVT population in the most 259 recent sampling period more closely reflects that of our last full pre-PCV13 sample than our first 260 post PCV-13 sample. This suggests a return to equilibrium following disruption by vaccine, which is consistent with observations made following the introduction of PCV-7 in the same 261 population³³, but now with the added resolution offered by genomic data. We also find that 262 263 serotype 3 CC180 has been more persistent than other serotypes added for PCV-13, but a different subclade of this lineage now predominates. 264

Given the value of being able to predict the composition of the pneumococcal population 265 following PCV use, the pattern observed amongst the NVTs is quite interesting. Our 2014 266 sample appears to be broadly a reflection of the 2007 sample, but 2011 is unlike either. As such, 267 it is possible that the pre-vaccine NVT population may be a good predictor of the post-vaccine 268 population, but that the disruption caused by vaccine introduction can temporarily interrupt this 269 pattern. Some of this could be due to variation in the age of children who have been vaccinated, 270 which should increase over time as vaccinated children age. The observed increase in SC 271 diversity in the immediate post-vaccine period, with the most common lineages making up a 272 smaller proportion of the total population, may provide an enhanced opportunity for rarer 273 lineages to increase. Considering this scenario, lineages such as SCs 3, 14, and 19 (serotypes 274 23A/15BC, 21 and 33F, respectively), may have a similar trajectory to that of serotype 19A 275 ST320 after PCV-7^{5,9,10,34}. It has also recently been suggested that negative frequency dependent 276 selection on elements of the accessory genome could be responsible for structuring the 277

pneumococcal population at both spatial and temporal scales³⁵. Further observation will help
determine the role of this and whether these or other lineages become more substantial
contributors to both carriage and invasive disease.

Previous studies have indicated that PCV-13 may not be as effective against serotype 3 as 281 it is against other included serotypes^{2,11,13,14}. The shift we observed in the serotype 3 CC180 282 283 population following the introduction of PCV-13 may reflect a similar phenomenon to that 284 leading to the recognition of serotype 6C as distinct from 6A following the introduction of PCV- $7^{36,37}$. The dominant lineage pre-PCV-13 was also more homogenous (i.e., less diverse) than the 285 286 post-vaccination population, so it is possible that the immunity generated against serotype 3 by PCV-13 is narrowly tailored to that subset of the population. At present, little genetic variation 287 among the CPS loci was observed, suggesting an alternative explanation for the recent post-288 PCV-13 emergent subclade. Given its propensity for causing disease, the persistence of serotype 289 3 despite its inclusion in PCV-13 warrants further investigation. 290

291 The response of the pneumococcal population to serotype-targeting conjugate vaccines may also provide insights for other pathogens for which vaccines have been targeted at or 292 differentially affect a subset of their population. The efficacy of the RTS.S malaria vaccine 293 appears to be partially dependent on how well the circumsporozoite protein of a given 294 *Plasmodium* type matches that in the vaccine³⁸. There has also been interest in understanding 295 how the strain dynamics and epidemiology of meningococcal disease caused by the bacteria 296 Neisseria meningitidis will be affected by the rollout of vaccinations against a variety of 297 serogroups^{39,40}. While each of these disease systems is different, there is some potential for 298 findings in one to inform hypotheses for how others will behave. 299

- 300 Pneumococcal epidemiology has changed substantially as a result of conjugate
- 301 vaccination. While PCVs have been highly effective in reducing the incidence of pneumococcal
- disease^{4,5,11}, continued vigilance is necessary to monitor for, and respond to, the emergence of
- 303 potentially dangerous lineages not protected against by current vaccine formulations.

304 Competing interests

- M.L. has consulted for Pfizer, Affinivax and Merck and has received grant support not related to
- this paper from Pfizer and PATH Vaccine Solutions. W.P.H., M.L. and N.J.C. have consulted for
- 307 Antigen Discovery Inc. S.I.P. has investigator initiated research funding (through Boston
- 308 Medical Center) from Pfizer and Merck Vaccines. He has also received honorarium from Pfizer,
- 309 GSK bio, Merck Vaccines, and Seqirus.

310 **References**

311	1.	Mehr S, Wood N. S	Streptococcus	pneumoniae – a	a review of	f carriage.	infection.	serotype

- replacement and vaccination. *Paediatr Respir Rev.* January 2012:2-8.
- doi:10.1016/j.prrv.2011.12.001.
- 2. Lee GM, Kleinman K, Pelton SI, et al. Impact of 13-valent pneumococcal conjugate
- vaccination on Streptococcus pneumoniae carriage in young children in Massachusetts. J
- 316 *Pediatric Infect Dis Soc.* 2014;3(1):23-32. doi:10.1093/jpids/pit057.
- 317 3. Huang SS, Johnson KM, Ray GT, et al. Healthcare utilization and cost of pneumococcal
- disease in the United States. *Vaccine*. 2011;29(18):3398-3412.
- doi:10.1016/j.vaccine.2011.02.088.
- 4. Whitney CG, Farley MM, Hadler J, et al. Decline in Invasive Pneumococal Disease after

the Introduction of Protein-Polysachharide Conjugate Vaccine. *N Engl J Med.*

- 322 2003;348(18):1737-1746.
- 5. Pilishvili T, Lexau C, Farley MM, et al. Sustained reductions in invasive pneumococcal

disease in the era of conjugate vaccine. *J Infect Dis*. 2010;201(1):32-41.

- doi:10.1086/648593.
- 326 6. Hausdorff WP, Bryant J, Paradiso PR, Siber GR. Which pneumococcal serogroups cause
- the most invasive disease: implications for conjugate vaccine formulation and use, part I.
 Clin Infect Dis. 2000;30(1):100-121. doi:10.1086/313608.
- 329 7. Huang SS, Platt R, Rifas-Shiman SL, Pelton SI, Goldmann D, Finkelstein J a. Post-PCV7
- changes in colonizing pneumococcal serotypes in 16 Massachusetts communities, 2001
- and 2004. *Pediatrics*. 2005;116(3):e408-13. doi:10.1542/peds.2004-2338.

332	8.	Huang SS, Hinrichsen VL, Stevenson AE, et al. Continued Impact of Pneumococcal
333		Conjugate Vaccine on Carriage in Young Children. <i>Pediatrics</i> . 2009;124(1):e1-e11.
334		doi:10.1542/peds.2008-3099.

- 335 9. Moore MR, Gertz RE, Woodbury RL, et al. Population snapshot of emergent
- 336 Streptococcus pneumoniae serotype 19A in the United States, 2005. *J Infect Dis*.

337 2008;197(7):1016-1027. doi:10.1086/528996.

- 10. Pelton SI, Huot H, Finkelstein J a, et al. Emergence of 19A as virulent and multidrug
- resistant Pneumococcus in Massachusetts following universal immunization of infants
- 340 with pneumococcal conjugate vaccine. *Pediatr Infect Dis J.* 2007;26(6):468-472.
- doi:10.1097/INF.0b013e31803df9ca.
- 11. Moore MR, Link-Gelles R, Schaffner W, et al. Effect of use of 13-valent pneumococcal

343 conjugate vaccine in children on invasive pneumococcal disease in children and adults in

the USA: analysis of multisite, population-based surveillance. *Lancet Infect Dis.*

345 2015;15(3):301-309. doi:10.1016/S1473-3099(14)71081-3.

- 12. Yildirim I, Hanage WP, Lipsitch M, et al. Serotype specific invasive capacity and
- persistent reduction in invasive pneumococcal disease. *Vaccine*. 2010;29(2):283-288.

doi:10.1016/j.vaccine.2010.10.032.

- 34913.Andrews NJ, Waight PA, Burbidge P, et al. Serotype-specific effectiveness and correlates
- of protection for the 13-valent pneumococcal conjugate vaccine: a postlicensure indirect
- 351 cohort study. *Lancet Infect Dis*. 2014;14(9):839-846. doi:10.1016/S1473-3099(14)70822-
- 352

9.

14. Harboe Z, Dalby T. Impact of 13-Valent Pneumococcal Conjugate Vaccination in

354	Invasive Pneumococcal	Disease Incidence a	nd Mortality.	Clin Infect Dis.	2014;59:1066-

- 1073. doi:10.1093/cid/ciu524. 355
- 356 15. Hanage WP, Finkelstein JA, Huang SS, et al. Evidence that pneumococcal serotype
- replacement in Massachusetts following conjugate vaccination is now complete. 357
- Epidemics. 2010;2(2):80-84. doi:10.1016/j.epidem.2010.03.005. 358
- 16. Croucher NJ, Finkelstein JA, Pelton SI, et al. Population genomics of post-vaccine 359 changes in pneumococcal epidemiology. Nat Genet. 2013;45(6):656-663.
- 360
- doi:10.1038/ng.2625. 361
- 362 17. Chang Q, Stevenson AE, Croucher NJ, et al. Stability of the pneumococcal population
- structure in Massachusetts as PCV13 was introduced. BMC Infect Dis. 2015;15:68. 363
- doi:10.1186/s12879-015-0797-z. 364
- Bankevich A, Nurk S, Antipov D, et al. SPAdes: a new genome assembly algorithm and 18. 365
- its applications to single-cell sequencing. J Comput Biol. 2012;19(5):455-477. 366
- 367 doi:10.1089/cmb.2012.0021.
- 19. Seemann T. Prokka: rapid prokaryotic genome annotation. *Bioinformatics*. 368
- 2014;30(14):2068-2069. doi:10.1093/bioinformatics/btu153. 369
- 370 20. Page AJ, Cummins CA, Hunt M, et al. Roary: Rapid large-scale prokaryote pan genome
- analysis. Bioinformatics. 2015;31(22):btv421. doi:10.1093/bioinformatics/btv421. 371
- 21. Hanage WP, Bishop CJ, Huang SS, et al. Carried pneumococci in Massachusetts children: 372
- the contribution of clonal expansion and serotype switching. Pediatr Infect Dis J. 373
- 2011;30(4):302-308. doi:10.1097/INF.0b013e318201a154. 374
- 375 22. Inouye M, Dashnow H, Raven L-A, et al. SRST2: Rapid genomic surveillance for public

376		health and hospital microbiology labs. Genome Med. 2014;6(11):90. doi:10.1186/s13073-
377		014-0090-6.
378	23.	Bentley SD, Aanensen DM, Mavroidi A, et al. Genetic analysis of the capsular
379		biosynthetic locus from all 90 pneumococcal serotypes. PLoS Genet. 2006;2(3):e31.
380		doi:10.1371/journal.pgen.0020031.
381	24.	Park IH, Park S, Hollingshead SK, Nahm MH. Genetic basis for the new pneumococcal
382		serotype, 6C. Infect Immun. 2007;75(9):4482-4489. doi:10.1128/IAI.00510-07.
383	25.	Price MN, Dehal PS, Arkin AP. FastTree: Computing Large Minimum Evolution Trees
384		with Profiles instead of a Distance Matrix. Mol Biol Evol. 2009;26(7):1641-1650.
385		doi:10.1093/molbev/msp077.
386	26.	Cheng L, Connor TR, Sirén J, Aanensen DM, Corander J. Hierarchical and spatially
387		explicit clustering of DNA sequences with BAPS software. Mol Biol Evol.
388		2013;30(5):1224-1228. doi:10.1093/molbev/mst028.
389	27.	Simpson EH. Measurement of Diversity. Nature. 1949;163:688-688.
390		doi:10.1038/163688a0.
391	28.	Hanage WP, Finkelstein JA, Huang SS, et al. Evidence that pneumococcal serotype
392		replacement in Massachusetts following conjugate vaccination is now complete.
393		Epidemics. 2010;2(2):80-84. doi:10.1016/j.epidem.2010.03.005.
394	29.	Dagan R, Patterson S, Juergens C, et al. Comparative Immunogenicity and Efficacy of 13-
395		Valent and 7-Valent Pneumococcal Conjugate Vaccines in Reducing Nasopharyngeal
396		Colonization: A Randomized Double-Blind Trial. Clin Infect Dis. 2013;57(7):952-962.
397		doi:10.1093/cid/cit428.

398 30. Cooper D, Yu X, Sidhu M, Nahm MH, Fernsten P, Jansen KU. The 13-val	398	30.	Cooper D, Y	Yu X, Sidhu M	, Nahm MH,	Fernsten P.	Jansen KU. The 13-valer
---	-----	-----	-------------	---------------	------------	-------------	-------------------------

- 399 pneumococcal conjugate vaccine (PCV13) elicits cross-functional opsonophagocytic
- 400 killing responses in humans to Streptococcus pneumoniae serotypes 6C and 7A. *Vaccine*.
- 401 2011;29(41):7207-7211. doi:10.1016/j.vaccine.2011.06.056.
- 402 31. Stamatakis A. RAxML-VI-HPC: Maximum likelihood-based phylogenetic analyses with
- thousands of taxa and mixed models. *Bioinformatics*. 2006;22(21):2688-2690.
- doi:10.1093/bioinformatics/btl446.
- 405 32. Li H, Handsaker B, Wysoker A, et al. The Sequence Alignment/Map format and

406 SAMtools. *Bioinformatics*. 2009;25(16):2078-2079. doi:10.1093/bioinformatics/btp352.

- 407 33. Hanage WP, Fraser C, Tang J, Connor TR, Corander J. Hyper-Recombination, Diversity,
- and Antibiotic Resistance in Pneumococcus. *Science (80-)*. 2009;324(5933):1454-1457.
 doi:10.1126/science.1171908.
- 410 34. Hanage WP, Bishop CJ, Lee GM, et al. Clonal replacement among 19A Streptococcus
- 411 pneumoniae in Massachusetts, prior to 13 valent conjugate vaccination. *Vaccine*.

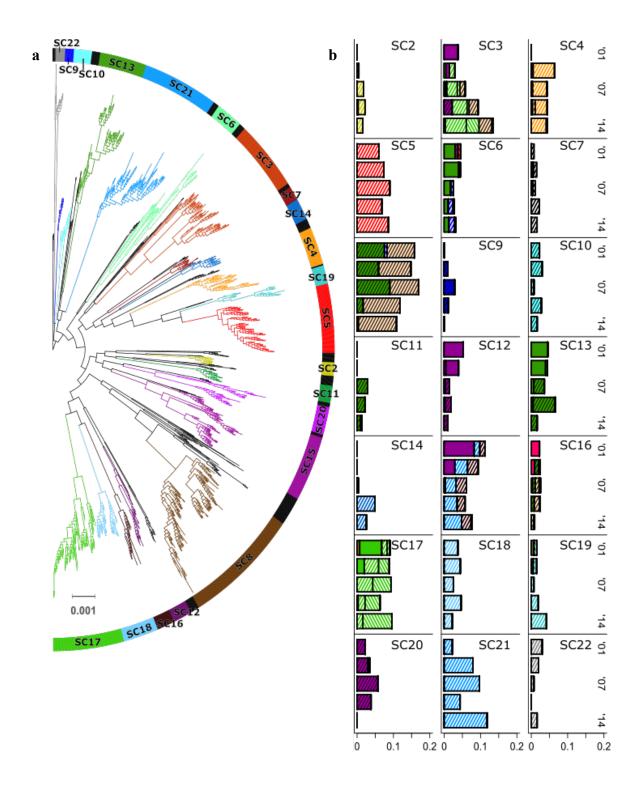
412 2011;29(48):8877-8881. doi:10.1016/j.vaccine.2011.09.075.

413 35. Jukka Corander, Christophe Fraser, Michael U. Gutmann, Brian Arnold, William P.

414 Hanage, Stephen D. Bentley, Marc Lipsitch NJC. Frequency-dependent selection in

- 415 vaccine-associated pneumococcal population dynamics. *Nat Ecol Evol*. October 2017:In
- 416 press. doi:10.1038/s41559-017-0337-x.
- 417 36. Park IH, Moore MR, Treanor JJ, et al. Differential effects of pneumococcal vaccines
 418 against serotypes 6A and 6C. *J Infect Dis*. 2008;198:1818-1822. doi:10.1086/593339.
- 419 37. Park IH, Pritchard DG, Cartee R, Brandao A, Brandileone MCC, Nahm MH. Discovery of

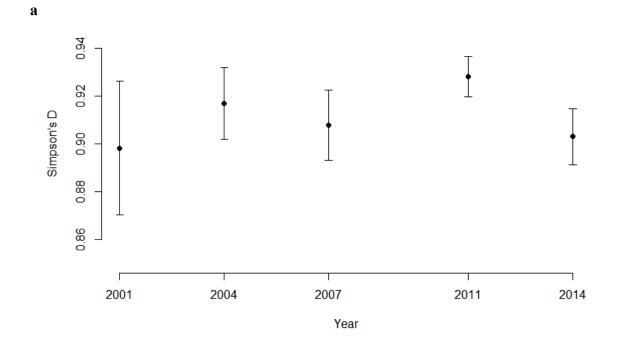
420		a new capsular serotype (6C) within serogroup 6 of Streptococcus pneumoniae. J Clin
421		Microbiol. 2007;45(4):1225-1233. doi:10.1128/JCM.02199-06.
422	38.	Neafsey DE, Juraska M, Bedford T, et al. Genetic Diversity and Protective Efficacy of the
423		RTS,S/AS01 Malaria Vaccine. N Engl J Med. 2015;373(21):2025-2037.
424		doi:10.1056/NEJMoa1505819.
425	39.	Halperin SA, Bettinger JA, Greenwood B, et al. The changing and dynamic epidemiology
426		of meningococcal disease. Vaccine. 2012;30:26-36. doi:10.1016/j.vaccine.2011.12.032.
427	40.	Ali O, Aseffa A, Bedru A, et al. The diversity of meningococcal carriage across the
428		African meningitis belt and the impact of vaccination with a group a meningococcal
429		conjugate vaccine. J Infect Dis. 2015;212:1298-1307. doi:10.1093/infdis/jiv211.
130		



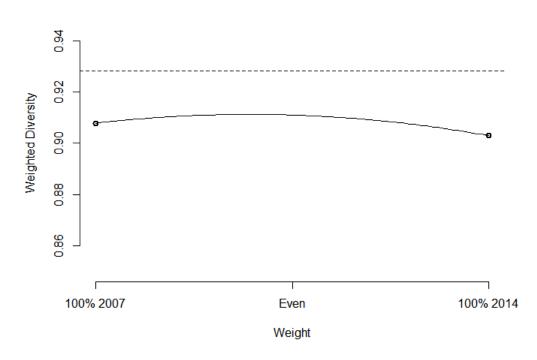
431

Figure 1: (a) Core genome phylogeny with SCs denoted by color. (b) Proportion of population
in each sampling period composed of each SC, with shading indicating serotype. Solid colors are

- 435 PCV-7 type, solid colors with black hatching are PCV-13, and white with colored hatching are
- 436 not covered by either. Serotype 6A is dotted as it is cross-reactive with 6B, a PCV-7 type, but is
- 437 itself included in PCV-13.



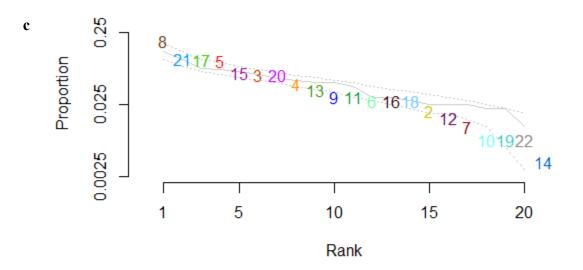




440 Figure 2 (continued)

441 (Continued)

442



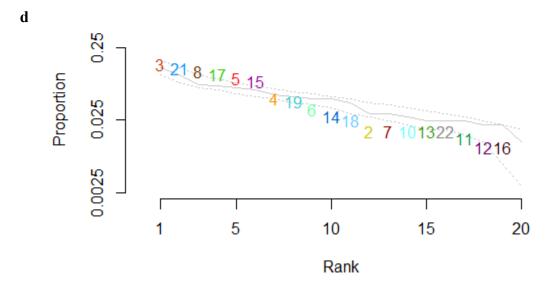
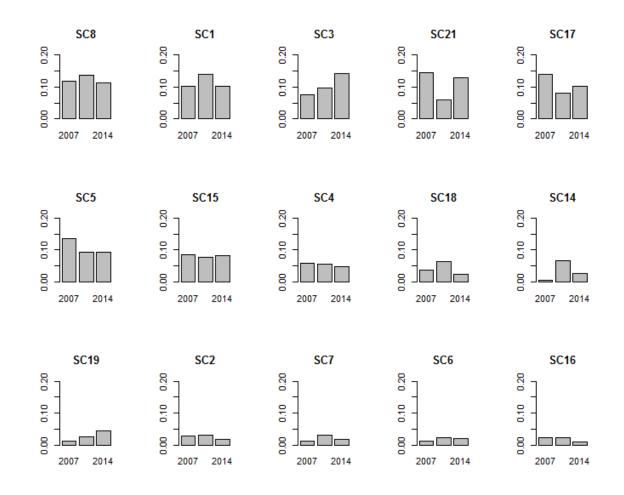


Figure 2: (a) Simpson's diversity of SCs, excluding the polyphyletic cluster, for each sampling
period. (b) Diversity calculated for hypothetical composites of 2007 and 2014 populations, with

- 446 2011 diversity shown as dashed line. (c-d) Proportion of population in (c) 2007 and (d) 2014
- 447 composed of each SC, ordered by frequency. Gray line is the corresponding distribution from
- 448 2011, with dotted lines representing 95% of values from 10000 random samples drawn from the
- 449 2011 population.



450

Figure 3: Proportion of the NVT population (i.e., those serotypes not included in PCV-13)
comprised of each SC. Two additional SCs, SC11 and SC20, had a single NVT isolate and were
excluded from this plot.

454

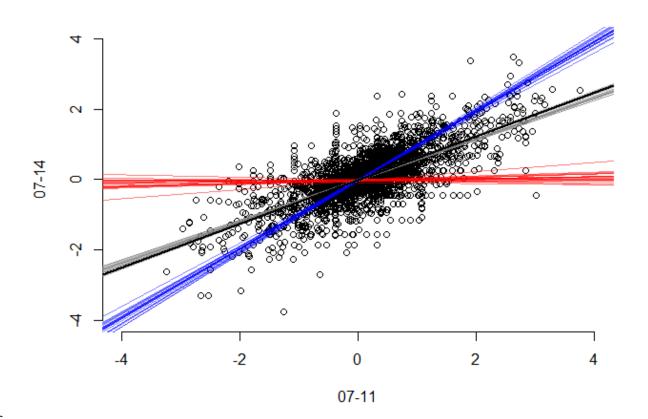
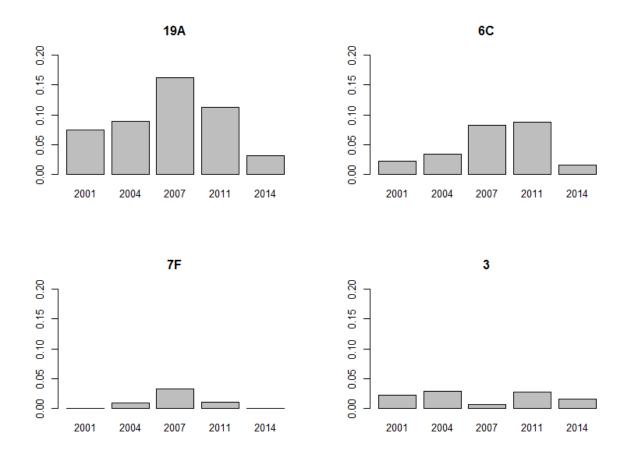
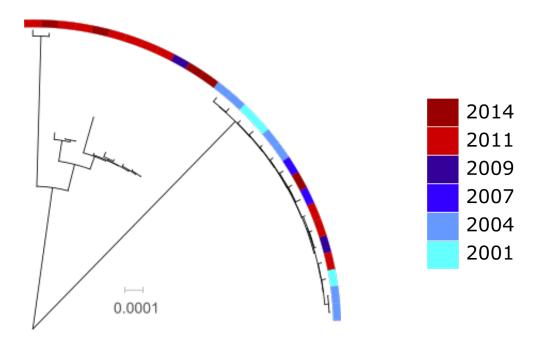


Figure 4: Regression coefficients comparing gene content of the NVT population from 2007 to
2011 and 2014. Black circles correspond to the coefficients with individual genes, with a linear
fit to the data shown in black. Fits in which a hypothetical 2014 population was drawn from
either the 2007, 2011, or 2014 population are shown in red, blue, and gray, respectively.



462

Figure 5: Proportion of the population in each sampling period comprised of the serotypes
included in PCV-13 but not PCV-7. As a note, PCV-7 was introduced in the United States in
2000 and PCV-13 was introduced in 2010.



467

Figure 6: Serotype 3 phylogeny, with sampling period shown by color. Isolates collected before
the introduction of PCV13, shown in blue, are found primarily on one monophyletic clade of the
tree, while post-introduction isolates, indicated by red, are primarily on the other.