2 worldwide Arabidopsis thaliana p

- 4 Yanjun Zan¹, Örjan Carlborg^{1*}
- 5 Affiliation: ¹Department of Medical Biochemistry and Microbiology, Uppsala University,
- 6 Uppsala, Sweden

7

10

11

- 8 * Corresponding author
- 9 Email: orjan.carlborg@imbim.uu.se

Abstract

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

As a locally adapted complex trait, flowering time in *Arabidopsis thaliana* has attracted much attention in genetics. Most studies have, however, focused on contributions by individual loci rather than the joint contributions by the large number of loci in the genetic architecture of flowering time to local and global adaptation. In an earlier study, we reported 46 loci associated with flowering time variation during growth at 10°C, 16°C or both in the 1,001genomes collection of Arabidopsis thaliana accessions. Here, we explore how these loci together contribute to differences among genetically defined, and geographically divided, subpopulations across the native range of this species. Our approach was to define flowering time as a trait, and the measurements at 10 and 16 °C as two independent measurements of it. This facilitated explorations of the dynamics in the genetic architecture –which loci contribute and their effects— of flowering time across growth temperatures and their potential roles in local and global adaptation. The overall flowering time differences between populations could be explained by subtle changes in allele-frequencies and gradual changes in phenotype due to globally present alleles. More extreme local adaptations were on several occasions due to contributions by regional alleles with relatively large effects. About 2/3 of the 48 evaluated flowering time loci had similar effects on flowering time at 10°C and 16°C, while the remaining 1/3 had different effects in the two temperatures, suggesting an important contribution of gene by temperature interactions to this trait. There are also indications that co-evolution of functionally connected alleles in local populations has been important for local adaptation. Overall, this study provides deeper insights to the polygenic genetic basis of flowering time variation in *Arabidopsis thaliana* across a wide range of ecological habitats.

Author Summary

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

Many genes can affect flowering time in Arabidopsis thaliana, but their contribution to natural flowering time variation in the worldwide population is largely unknown. We explored how 48 loci associated with flowering time, measured at 10°C and 16°C, or their difference, for the same wild collected 1.001-genomes Arabidopsis thaliana accessions together contribute to differences among the genetically defined and geographically divided subpopulations from the native species range. The overall flowering time differences among these subpopulations could be explained by the joint small effects of globally present alleles. suggesting an important contribution by polygenic adaptation for this trait. Most alleles with large effects on flowering were present only in some populations, facilitating more extreme local adaptations. Long-range LD was observed between genes in several biological pathways, indicating possible local adaptation via co-evolution of functionally connected polymorphisms. The genetic architecture of flowering time was also found to depend on the growth temperature. Most flowering time loci had similar effects on flowering time measured at 10°C and 16°C, but the effects of about 1/3 of them had effects that varied with temperature. Overall, new insights are provided to how the polygenic architecture of flowering time has facilitated its colonisation of a wide range of ecological habitats.

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

Introduction Understanding the genetic basis of adaptation is a central theme in evolutionary genetics [1,2]. Adaptation in nature can rely on a few loci with large individual effects [1,3], and such loci have been identified in natural populations of for example Arabidopsis thaliana [4–7] and the Darwin finch [8]. In other cases, populations adapt via selection on many loci with small effect [9], with examples reported for fitness traits in a range of species including mice, drosophila, human and maize[10–12]. In depth characterizations of the genetic mechanisms contributing to adaptation for polygenic quantitative traits is, however, a difficult [13] and despite some successes little is overall known. Adaptations for polygenic traits could be rapid due to selection on available standing variation [14-16] and new adaptive optima could be reached via gradual allele frequency changes across multiple loci [17,18]. The individual effects of the contributing loci might be too low for detection in genome wide scans based on stringent multiple testing thresholds, but they might despite this be necessary for reaching fitness optima [19–21]. Arabidopsis thaliana, a widely used model species in plant biology, has colonised a broad range of ecological habitats around the world. One of the most well studied adaptive traits is flowering time due to its role in ecological adaptation and potential impact on agronomic production in related species. Molecular studies in the laboratory, primarily using the reference accession Col-0, have identified many genes with the potential to alter flowering time. Association studies in natural populations, however, rarely reveal more than a handful of the loci contributing to the genetic variation of this trait [22–25]. Hence, even though it is

likely that flowering time adaptations across the native range of A. thaliana would involve

many loci, the polygenic basis of these differences is still largely unexplored.

Previously, we developed and used a new statistical approach to dissect the genetic basis of polygenetic traits in natural populations. The properties of the method was illustrated by exploring the polygenic basis of flowering time in Arabidopsis thaliana, using public data from the 1,001 genomes project[26]. In total, 33/29 loci were there mapped for flowering times in 1,004 wild collected Arabidopsis thaliana accessions from the 1,001-genomes project grown at 10/16°C[26]. Here we explored how the genetic effects of these loci i) vary within and across loci and growth temperatures and ii) how these variations in allelic effects contribute to the flowering time variation between genetically defined subpopulations that have colonized the native range of the species [22]. Loci with globally segregating alleles made, in general, small individual contributions to flowering but together captured the overall pattern of flowering time differences between the subpopulations, suggesting a role of polygenic adaptation[14–18]. The alleles with more restricted geographic distributions had in several instances large effects and made important contributions to local adaptations of some subpopulations. The genetic effects of many loci varied across temperatures, suggesting that genotype by temperature interactions might be important for flowering time adaptation. The long-range LD between polymorphisms in genes of the same biological pathways suggests that co-evolution of different pathways has contributed to local adaptation in different geographic areas. Overall this reanalysis of public data provides new insights to the polygenetic basis of flowering time adaptation in A. thaliana.

Results

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

100

101

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

Flowering time differentiation in A. thaliana worldwide population Flowering time variation of Arabidopsis thaliana accessions sampled from its native range is strongly associated with geographic variation in climate [27–29]. To study the genetics of the this geographical differentiation in greater detail, we reanalysed a publicly available dataset where flowering time was measured at 10°C (FT10) and 16°C (FT16) under greenhouse conditions on 1,004 whole-genome re-sequenced Arabidopsis thaliana accessions[22]. The genetic population structure and geographic sampling locations for the analysed accessions are illustrated in S1 Fig. We approached flowering time in this population as a single complex trait measured at two temperatures to reveal the dynamics in its underlying genetic architecture. At these temperatures, flowering times displayed a continuous variation with earlier average flowering times (9.3 days) at 16 °C than at 10 °C (Figure 1A). The variations in flowering times at both growth temperatures were correlated with the location and climate at the sampling sites of the accessions, suggesting its potential role in global, and local, adaptation. For example, there was a significant latitudinal cline of flowering times (Fig 1B/S2A Fig for FT10/FT16; Pearson correlation = 0.37; P < 2.2×10^{-16} ; df = 963) and a significant correlation between flowering times and mean temperature at the sampling locations (Fig 1C/S2B Fig for FT10/FT16; Pearson correlation = -0.31; P < 2.2×10^{-16} ; df = 963). The phenotypic variance increased with temperature, being larger at 16°C than at 10°C (Fig. 1A), while the proportion of additive variance estimated by fitting the IBS (Identity By State) kinship matrix were similar at both temperatures (82/83% at 10/16°C). Additive genetic variance is thus released as the temperature increases, which will be explored in more detail in

the following section. Regardless of whether flowering times were measured at 10°C or 16°C, there were substantial phenotypic differences between the eight genetically defined and geographically separated subpopulations defined by *Alonso et al* [22] in their admixture analysis of this population (Fig 1D). Increasing the growth temperature leads to earlier flowering times in most of the sub-populations, with the North Swedish population being the only exception with an on average later (4.9 days) flowering time at 16°C.

[Fig 1 about here]

The genetic architecture of flowering time is highly polygenic in the worldwide

A.thaliana population

By utilizing prior molecular knowledge of genes regulating flowering time variation in the reference accession (*Col-0*), and results from an expression QTL analysis of those genes in this 1001-genomes collection of natural accessions, we earlier mapped 46 flowering time loci where 33/29 loci were associated with FT10/FT16 using a multi-locus association analysis[26]. Together these mapped loci explain 55/48% of the total genetic variance for FT10/FT16[26]. Here, rather than focusing on the results from the two temperatures separately, the complete genetic architecture of flowering time revealed across them was evaluated. Overall, 16 loci were mapped at both temperatures, and 2 pairs of loci were located within 20 kb from each other, suggesting that they contribute to flowering time regardless of temperature in a relatively stable manner. The remaining 30 loci were significantly associated with flowering times only at one temperature in the earlier analysis[26], and their possible involvement in genotype-by-temperature interactions are further explored here.

Two additional analyses were performed to explore the possible role of genotype-by-

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

temperature interactions in this population. First a GWA mapping analysis to the flowering time difference at the two evaluated temperatures (FT10-FT16). Two genome-wide significant associations, one at Chr1: 23,453,207 bp and one at Chr4: 1,338,596 bp, were detected in this analysis after Bonferroni correction for ~1.4 million SNPs (S3 Fig). Second, the polygenic contribution of the loci that were significant in one temperature, but not the other, to flowering times was evaluated. This by testing if loci associated with FT16, but not FT10, together with the two additional loci affecting FT10-FT16 jointly contributed a significant amount of genetic variation in FT10, and vice versa. Models with or without nonoverlapping loci were fitted, and significant increases in the adjusted R² were detected at both temperatures (4/8% for FT10/FT16; Likelihood ratio test; P-value= 3.2 x 10⁻¹⁴/1.3 x 10⁻³³; S4 Fig). Linear models including the genotypes of the 48 mapped flowering time loci, but excluding their kinship, explained the majority of the additive genetic variance at both temperatures (79/75% for FT10/FT16). In the following sections, we will explore how these 48 loci together contribute to flowering time across the different genetically and geographically divided sub-populations in this population, to potentially gain deeper insights to how polygenic and genotype-by-temperature interactions might contribute to local and global adaptation in nature. The role of genotype-by-temperature interactions for flowering times in A. thaliana It is known that genotype-by-temperature interactions are important for flowering times in Arabidopsis thaliana [30–32]. The genetic basis of this phenomenon is, however, largely unknown. Here, we studied this by evaluating the temperature dependence of the genetic

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

effects of 48 mapped flowering time loci. As shown in Fig 1A, increased growth temperature leads to a release of additive genetic variance in flowering time. To identify the genetic basis of this, the genetic effects of the 48 mapped flowering time loci on FT10 and FT16 were estimated by fitting them jointly in a mixed model (Fig 2). The alleles at 42 of the 48 loci have effects in the same direction on FT10 and FT16, i.e they consistently increase or decrease flowering time regardless of temperature. The alleles at the remaining six loci have opposite effects in 10°C and 16°C, but as all these estimates in at least one temperature are close to zero (Fig 2A). This result is likely to at least in part be the outcome of the uncertainty in the estimation of the allelic effects. For $\sim 2/3$ of the loci, the sizes of the genetic effects on flowering time are in good general agreement (Fig 2A), suggesting that they affect flowering time in similar way regardless of whether the plants are grown at 10°C or 16°C. The remaining $\sim 1/3$ loci, however, have larger effects on flowering time when the plants are grown at 16°C than when they are grown at 10°C (Fig 2A). This is consistent with the observation of an increased additive genetic variance at 16°C, but at the same time unexpected as an increase in temperature generally leads to an overall decrease in flowering time. To evaluate whether these putative genotype-by-temperature were statistically significant, we compared the fits of two linear models for every locus: one including all 48 loci and an indicator variable for the two growth temperatures as fixed effects, and another also including an interaction term between growth temperature and each tested locus. In total, 18 loci displayed significant genotype-by-temperature interactions after correction for multiple testing (highlighted in Fig 2A; names of loci & P-values from the corresponding Likelihood Ratio Test in Table S1).

[Fig 2 about here]

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

Distribution and effects of flowering time alleles in the worldwide A. thaliana population To study the putative role of the 48 mapped flowering time loci in global and local adaptation, we evaluated how the alleles at these were distributed across the subpopulations identified in this dataset by Alonso et al [22]. The assumption in this analysis is that allele frequency differences between subpopulations across these loci, in its most extreme form the complete presence/absence of alleles, at least in part reflects past and ongoing selection leading to global adaptation along latitudinal or geographical clines, and local adaptations in individual subpopulations [33–35]. All 48 mapped loci were identified by having effects on flowering time at 10°C, 16°C, their difference or all, and we therefore consider all to be candidate adaptive loci as the temperature will vary both within and across the sampling sites of the accessions. Consistent with the observation that the 48 mapped loci explained the majority of the additive genetic variance in flowering time, the overall pattern of early to late flowering times between the subpopulations in this dataset is captured well by the combined additive effects of these (Fig 3A/S4 Fig). Further, late flowering populations have accumulated both major and minor effect late flowering alleles across many loci (Fig 3B; late flowering alleles in red; effect sizes sorted from top to bottom). The late flowering north Swedish accessions are most extreme, with the highest frequencies at many loci. A visual inspection of the allele frequency patterns of the 48 loci across the populations (Fig 3B) suggests a greater overall similarity between populations from nearby geographic locations. This is analytically supported by a clustering analysis across the subpopulations based on the allele frequencies across these 48 loci (Fig. 3C), and the pairwise correlations of the allele-frequencies between the populations (S5 Fig).

For example, the accessions from Germany and Central Europe are closely related (Figs 3B and 3C), with an exception being the relatively late flowering southern Swedish accessions that overall were more similar to the earlier flowering Central European/German accessions than the late flowering north Swedish accessions.

[Fig 3 about here]

For some of the 48 loci, both alleles are present (here defined as having MAF > 0.05) in all subpopulations, for example the loci on chromosomes 2 (16,387,379bp) and 4 (1,961,868bp) (Fig 2B; S6A/S6B Fig). For others, one of the alleles is either rare (MAF < 0.05) or absent in one or more of the subpopulations, for example chromosomes 2 (9,312,968bp), 4 (1,728,204bp) and 5 (22,979,827bp) (Fig 2B; S6C/S6E Fig). Both global and local allelic variations are thus likely to contribute to the natural flowering time variation across the worldwide range of *A. thaliana*. To explore this further, the 48 mapped flowering time loci were divided into two groups containing either those that are present across all populations (MAF > 0.05 in all populations; n = 23), or those that are rare or absent in at least one subpopulation (MAF < 0.05 in at least one population; n = 25). The choice of 0.05 as MAF cut-off is arbitrary but nevertheless serves the purpose of representing the spatial distribution of global vs primarily local alleles (Fig 3; Fig 5), and the results are overall similar also for MAF < 0.03 and MAF < 0.10 (results not shown).

Next, the contributions by global and local alleles to flowering time variation in the 8 subpopulations was evaluated. A mixed model, including the IBS kinship and the genotype of all 48 loci, was first fitted to FT10 and FT16 separately. Then, the estimated additive effects

of these 48 alleles were used to calculate the contributions by local and global alleles, as defined above, to flowering time variation.

Contributions by global alleles to flowering time variation: For FT10, the IBS kinship captures the overall early to late flowering time pattern for subpopulations well and explicitly modelling the global alleles improves it further (Fig 4A). This illustrates that the polygenetic effect, together with the gradual allele frequency shifts across the loci with global alleles, are important for the overall pattern of flowering time differentiation between the subpopulations. The majority of the global alleles have moderate to small effects on flowering time (Fig 4A/B), relative to the local alleles (discussed further in section below), but together they contribute 21% of the phenotypic variance of the trait measured at both 10°C and 16°C. This analysis also shows that the global alleles have a larger contribution at 16°C than that to 10°C, again suggesting a role of genotype-by-temperature interaction for this trait (Fig 4A/B). The kinship is estimated to make a negative contribution to flowering times measured at 16°C in some subpopulations (Fig 4B), while the joint contribution by the global alleles is larger than at 10°C. One likely interpretation of this is that some of the global alleles have large overall effects at this temperature. They seems to also differ in effects across subpopulations and this genotype-by-environment interaction is captured by the kinship.

[Fig 4 about here]

Contribution by local alleles to flowering time variation. We next explored the loci with alleles that are rare or absent (MAF < 0.05) in at least one of the populations in more detail (Fig 5). Across these loci, the most apparent enrichment is for late flowering alleles in the

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

Swedish populations (Fig 3B), where one or both of these populations contain near private late flowering alleles at high frequencies. These include the strongest late-flowering allele of all (chromosome 5 at 23 Mb; Fig 5), suggesting this large effect allele having emerged locally and been under strong selection here. A few other loci with relatively large effects on flowering were also predominantly present in one or a few subpopulations, but none were as striking as the aforementioned locus (Figs 3B and 5; S4 Fig). The local alleles appears to primarily be contributing more extreme phenotypes in individual subpopulations (Fig 4A). In contrast to the global alleles, there is not only generally larger allele frequency differences between the populations for the local alleles, this group is also enriched for alleles with large effects that individually explain larger proportions of the phenotypic variance (Fig 4B). Together the local alleles contribute 45/42% of the phenotypic variance for FT10/FT16 in this population. The flowering time differences between the subpopulations thus result from shifts in the allele frequencies of global alleles (Fig 3B), with more extreme (for example northern Sweden) or variable (for example Spain) local phenotypes being due to private, or near private, local alleles (Fig 5). [Fig 5 about here] Long-range LD between flowering time loci Long-range LD between loci is a possible sign of co-selection of them. To screen for such signatures, we calculated the pairwise LD (D') between the 48 associated loci (Fig 6). In total, 67 pairs were in significant nominal long-range LD (P-value < 0.05; overall FDR for this set = 12%), where a few loci were in long-range LD with several other loci on either the same or

different chromosomes (Fig 6). When a locus was in long-range LD with more than 4 other loci, the loci were defined as belonging to an LD-cluster.

[Fig 6 about here]

We explored the five identified clusters in more detail. A literature review suggested functional connections between candidate genes in three of the five clusters. One cluster contained several genes in a miRNA mediated flowering time regulation pathway, suggesting a functional connection to the FLC-CO/FT module [36–41] (Table 1). Two clusters contained genes whose function are connected to the *FRI-FLC* module, one of these possibly through vernalization [42–44]. The minor alleles of the five loci that via their long-range LD defined these clusters were enriched in specific geographic regions. The one allele on chromosome 2 (15,044,483bp) was found primarily in Sweden, the one on chromosome 1 (16,471,394bp) in Italy Caucasus, the one on chromosome 3 (7,240,079bp) in Asia, the one on chromosome 4 (17,282,047bp) in Spain and the one on chromosome 5 (22,979,827bp) in north Sweden (Fig 4B). Together this suggests possible contributions from these pathways to local adaptation in these geographic regions.

Table 1. Long-range LD defined clusters of genes with assigned functional classification

Cluster ^a	Locus ^b	Gene	Alias	Functional classification ^c
	1:16471394*	AT1G03970	GBF4	
	1:23453207	NA	Unknown	
1 2:1 3:1	2:15044483	AT2G42590	GRF9	Unclear
	2:19204119	AT5G08190	NF-YB12	
	3:16321974	AT3G03450	RGL2	
	3:16325481	AT3G33520	SUF3	

	4:8272818	AT2G23080	CKA3	
	5:16541207	AT5G17490	RGL3	
	5:22979827	AT5G57380	VIN3	
	5:26805231	AT5G67180	TOE3	
	2:15044483*	AT2G42590	GRF9	
	2:16387379	AT2G39250	SNZ	
	4:259252	AT4G00650	FRI	
2	4:1338596 NA Unk	Unknown	FLC-CO/FT	
4:7172638	4:7172638	AT2G28190	SOD2	TLC-CO/FT
	4:10999188	NA	TSF	
	4:17282047	NA	Unknown	
	5:3274505	AT4G00650	FLA	
	3:7240079 [*]	AT3G20740	FIS3	
	3:16325481	AT3G33520	SUF3	FRI-FLC
3	4:1338596	NA	Unknown	
	4:8272818	AT2G23080	CKA3	
	4:17282047	NA	Unknown	
	5:5345341 4:17282047*	AT5G16320 NA	FRL1	
	2:9048513	AT3G22380	TIC	
4	2:15044483	AT2G42590	GRF9	Unclear
	3:7240079	AT3G20740	FIS3	
	4:259252	AT4G00650	FRI	
	4:1338596	NA	Unknown	
	4:6559815	AT4G10180	DET1	
5	5:22979827 [*]	AT5G57380	VIN3	
	1:16471394	AT1G03970	GBF4	
	2:14312485	AT2G33835	FES1	
	4:259252	AT4G00650	FRI	Vin-FRI-FLC
	4:1338596	NA	Unknown	
	4:7172638	AT2G28190	SOD2	
	5:3274505	AT4G00650	FLA	

5:5345341 AT5G16320 FRL1

^aCluster as defined by the high long-range LD to the first locus (underlined) in the group; ^bLocus (chromosome: position in bp) associated with flowering time at 10°C, 16°C or both in the multi-locus analysis; ^cFunctional classification of the cluster based on the likely pathway affected by the polymorphisms based on a literature study of the included genes [36–43,45–47].

Discussion

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

In an earlier study, we mapped the polygenetic basis of flowering time variation in the latest public release of data from the 1,001 genomes A. thaliana project [22,26]. In this analysis, 33/29 loci were reported to be associated with flowering time variation measured at 10/16°C and they together explained 66/58% of the additive genetic variation[26]. These loci were mapped using a two-step polygenic analysis method. First, loci with individual genome-wide significant associations either directly to measured flowering times, or to the expression of experimentally known flowering time genes, were identified as candidate genes for the polygenic analysis in this population. Then, a joint analysis across all these candidate loci was performed for each trait separately using a backward elimination analysis, to infer a polygenic architecture of the trait at a 15% false discovery rate significance threshold. At this significance threshold, a few of the 33/29 loci associated with FT10/FT16 are expected to be false positives. Here, we continued this work by treating flowering time as one trait measured independently for the same genotypes at 10°C and 16 °C. The focus was to evaluate the joint contributions by all mapped flowering time loci to the genetics of this trait and their possible role in global and local adaptation to the different ecological conditions across the worldwide range of this plant. We found that the groups of loci that were individually significant in one growth temperature, but not the other, made significant joint contributions to flowering time

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

in both conditions. Fitting all 48 loci together in a multi-locus linear model, they explain most (79%/75% for FT10/FT16) of the additive genetic variance in flowering times at these growth temperatures. As most of the loci are likely to be true associations, we performed an in-depth evaluation of how this set of loci contribute to the global flowering time variation in A. thaliana to provide valuable complementary information to earlier studies that have focused primarily on loci with strong individual associations [23,25,48,49]. It is generally difficult to study the genetic basis of highly polygenic traits in natural populations due to, for example, the small contributions made by individual loci to trait variation and the confounding of adaptive alleles with population structure. This is likely one of the reasons why standard genome-wide association mapping approaches have only been able to reveal a small number of flowering time loci in this or related populations [22–24]. By approaching flowering time as a polygenic complex trait, and making use of its measures in two temperatures, this study could shed lights on how allele frequency changes across many loci, as well as genotype by temperature interactions, contribute to the genetic basis of global and local trait variation. The role of global and local alleles in shaping worldwide flowering time variation For many of the polymorphic flowering time loci, both alleles were present in all subpopulations of the worldwide population, albeit at different frequencies. Together these contributed to the overall pattern of early to late flowering in the geographically divided subpopulations. Polygenic global adaptation via small allele frequency shifts across many loci

thus appears to be a possible contributor to the variation of this trait in A. thaliana.

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

382

383

384

For a number of the associated loci, one of the alleles was absent or present at low frequency (MAF < 0.05) in one or more of the subpopulations. Several of these local alleles had strong effects on flowering time, and these loci made important contributions to the more extreme flowering time phenotypes of individual subpopulations. Alleles with large effects on adaptive traits have earlier been reported to be present in local populations of *Arabidopsis* thaliana [5,7,23,50]. Our finding that many such alleles are present in locally in the worldwide population, is coherent with earlier speculations that locally adaptive alleles for flowering time variation in Arabidopsis thaliana are likely to have large effects [51]. Those local populations appear to reach their respective flowering times via the effects of alleles that are primarily present in some populations suggests that different combinations of alleles are likely explaining adaptation to different local environments. We evaluated the distribution of many flowering time alleles across the native range of the species. The results are interpreted under the assumption that the alleles associated with flowering time are very likely to have an adaptive value and that the observed patterns of allele frequency differences between populations, to large extent, represent local and global adaptations. Other explanations are, however, also possible as similar patterns could emerge due to, for example, an interplay of drift, historical contingency and selection. A few observations are particularly valuable when interpreting the results. First, the consistency in which loci are mapped, and their patterns of effects when the plants are grown in two different growth temperatures, suggests that the genetic architecture of flowering time is overall stable. Hence, even though the individual effects of many loci are small, making it difficult to assess the selective pressure on each locus in nature, polygenic adaptation acting over time on the many alleles that are present globally in A.thaliana appears like a possible

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

route to adaptation. The finding that stronger local alleles complements the global polygenic adaptation, likely through a more rapid increase in allele-frequencies due to stronger selection for locally favourable phenotypes, is also consistent with earlier reports in which a few flowering time QTL were found to be correlation with latitude [48,52], indicating a potential role of spatial adaptation. That the same process might have been important also for other traits is suggested by the results of another recent study on drought adaptation reporting a polygenic genetic architecture and accompanying geographical enrichment of OTL[53]. Another possible explanation for the finding of, more or less, local alleles in several subpopulations is that the plants are naturally restricted to its native range over their lifespan and that self-pollination in Arabidopsis thaliana slows down the long distance spreading of new genetic variants. Hence, newly emerged alleles with large beneficial adaptive effects can quickly sweep to high frequency in local populations, but long distance spreading of those alleles will take longer in this selfing plant, although it is known to happen due to for example human or animal activities [54]. A possible example of such long distance spreading of genetic variants observed in this data, is the large effect allele identified on chromosome 5, 23 Mb. Here, the strong allele is present at high frequency in northern Sweden and in a relict population in Spain (S4F Fig). These populations are geographically distant, and it appears a likely result from lineage mixing between relict and northern populations as indicated in an earlier study of this dataset [22]. Two late-flowering alleles, with intermediate to high effect, are private (or nearly so) to the Spanish population (Fig 4). We note that this population has a significantly higher withinpopulation variation in flowering time ($P = 1.1 \times 10^{-6}$; Brown-Forsythe test) than the other populations (Fig 2A). The reason for this is not known, but a possible explanation could be

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

430

431

432

that these strong alleles have contributed to specific local adaptations in this subpopulation. Further studies of these effects would be valuable to evaluate the possible contributions of these alleles to local adaptations. Possible contribution by co-evolution of functionally related loci in local populations Four of the flowering time associated loci were in long-range linkage-disequilibrium (D') with several other mapped loci. These alleles were enriched in relatively distinct geographic regions, suggesting that they have emerged and spread locally making a confounding with population structure a possible explanation for this observation. We, however, consider this unlikely as they were detected in a statistical multi-locus analysis accounting for populationstructure and hence each of them captures flowering time variation that could not be explained by the other loci. Further, as in three of the five cases the loci involve candidate genes in related biological pathways. An intriguing alternative interpretation is that the LD instead result from co-selection of multiple functionally connected loci. Further work is, however, needed to confirm this hypothesis. Genotype-by-temperature interactions and their possible implications for adaptation An earlier study reported genotype by environment interactions in the Swedish A. thaliana populations that were part of the dataset analysed in this study [30]. Here, we explored the possible role of genotype-by-temperature interactions for in total 48 flowering time loci associated with this trait in the entire 1,001-genomes collection of worldwide accessions grown at both 10°C and 16°C. Our conceptual approach to study these interactions was slightly different to the one used before. First, flowering time was considered as a single complex trait and all loci associated with this trait, regardless of growth temperature, was

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

452

453

454

455

456

considered as possible loci involved in genotype-by-temperature interactions. Next, the effects of all the 48 associated loci on the flowering times measured at 10/16°C were estimated. The allelic effects in the two growth temperatures were compared, and possible genotype-by-temperature interactions inferred based on observed differences in effects between the two temperatures. Given the high phenotypic correlation between flowering times of accessions grown in these different temperatures (Pearson correlation=0.88), we expected the effects to be overall similar between environments. However, for $\sim 1/3$ of the mapped loci (18 of 48 loci; Fig 6), the effects on flowering time changed with temperature. These results suggest that the genetic effects of many loci in the genetic architecture of this polygenic trait are dependent on one or more environmental factors, as exemplified by temperature here. As studies aiming to detect complex trait loci, for example using a GWAS approach, are generally limited in their ability to assess the effects of loci across multiple environments a significant fraction of loci important in the variable conditions under which plants live in nature might be missed. More studies, for example to identify seasonally sensitive QTL in the field using GWA analysis[48], will therefore be of great value for our understanding of the genetic of such traits and their potential role in adaptation. The potential adaptive values of the genotype-by-temperature interactions identified here is difficult to assess. We observed a geographical enrichment of loci involved in such interactions, that have larger effects at 16°C, in the northern Swedish populations, where flowering time is increased at higher temperature (S7 Fig). Further investigations uncovered significant pairwise correlations between the joint effects of these loci on flower time differences between temperatures (FT10-FT16) and within year temperature fluctuation (temperature difference between hottest and coldest day; S8 Fig). It is therefore possible that these loci are of adaptive value in environments where the temperature fluctuates drastically, and where having alleles that can shorten flowering time in response to cold temperatures would be beneficial. However, further studies are needed to evaluate this hypothesis thoroughly.

Concluding remarks

457

458

459

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

478

479

480

Our study demonstrates that the variation in flowering times between earlier defined geographically and genetically separated A. thaliana subpopulations is likely due to allelic variations in many loci. At some of these loci both alleles are present across the entire range of the species, whereas others are only present in some of the subpopulations. Across these loci, the allele frequency shifts were often more gradual at loci with global present alleles. Larger differences in allele frequencies were observed for several loci with alleles of relatively large effects, and higher allele frequencies were then found in local populations with more extreme phenotypes. This suggests that flowering time adaptation results via a combination of global polygenic adaptation across many loci, complemented by local adaptation via the effects of stronger alleles with a more regional distribution. The results also indicate that local phenotypes might also result from co-evolution of multiple polymorphisms in biological pathways. We identified genotype-by-temperature interactions involving 18 flowering time loci, and propose a hypothesis that these might be adaptive in environments with large temperature fluctuations. However, further investigations are needed to validate this hypothesis. Overall, this study provides new insights to the genetic basis of flowering time in A. thaliana and propose possible explanations for how this self-pollinating plant has been able to colonize and adapt to such a wide range of ecological habitats around the world.

Materials and Methods

481

482

483

484

485

486

487

488

489

490

491

492

493

494

495

496

497

498

499

500

501

502

503

504

Data

All phenotype, genome-sequence and subpopulation information are publicly available as part

of the Arabidopsis thaliana 1,001-genomes project [22,55]. Flowering times measured at

10°C and 16°C in the greenhouse were downloaded from [56,57]. The subpopulation

classifications of the accessions were downloaded from

http://1001genomes.org/tables/1001genomes-accessions.html. The imputed whole genome

SNP data matrix was downloaded from

http://1001genomes.org/data/GMIMPI/releases/v3.1/SNP_matrix_imputed_hdf5/1001_SNP_

MATRIX.tar.gz. We filtered the SNP data for minor allele frequency and only retained loci

with MAF > 0.03. SNP markers were pruned to remove loci in pairwise LD of $r^2 > 0.99$. In

total, 1,396,438 SNPs on 1,004 individuals remained. The 33/29 loci associated with

flowering times at 10/16°C were identified in a previous study[26]. These loci were inferred

using a two-step polygenic mapping method. First, candidate flowering-time loci were

identified as those with either i) individual genome-wide significant associations to flowering

time or ii) that altered the expression of genes that are experimentally known to affect

flowering. Then, these candidate loci were analysed jointly for associations to flowering-

times measured at 10°C and 16°C using a backward elimination based multi-locus mapping

approach to infer the polygenic architecture of flowering time at a 15% false discovery rate

significance level [26].

Detecting loci involved in genotype by environment interactions

Two methods were used to test for QTL by environment interaction. The first was a genome

wide scan using the mean flowering time difference between 10°C and 16°C as phenotype. In

the analysis, a forward-selection based approach was employed where a linear mixed model $\bar{y}_{FT10} - \bar{y}_{FT16} = X\beta + Zu + e$ (1) was iteratively fitted to the SNPs across the genome with significant SNPs from earlier rounds added as covariates in the model. This model has earlier been shown to have equivalent power with a bi-variate analysis for detecting loci with increased/decreased effects across environments[58]. Here, $\bar{y}_{FT10} - \bar{y}_{FT16}$ is the mean flowering time difference between 10°C and 16°C for an individual accession. X is the design matrix including a column vector of 1's for the population mean and one or more additional column vectors with the indicator regression variables for the homozygous genotypes at the SNP or SNPs included in the model (coded as 0/2, respectively). Z is matrix obtained by singular value decomposition of the G matrix ($ZZ^T = G$), where G is the genomic kinship matrix estimated from the whole genome marker set using the ibs function in the GenABEL R-package [59]. The significance-threshold used in the analysis was Bonferroni corrected for ~ 1.4 million tested SNPs (-log10(p-value)=7.33).

- 519 Second, the loci associated with flowering-time in the earlier polygenic analysis were
- explored for genotype by environment interactions. Two models were fitted to the data:

521
$$\bar{y}_{ij} = u + E_j + a_1 b_1 + a_2 b_2 + \cdots + a_n b_n + e (2)$$

522
$$\bar{y}_{ij} = u + E_j + a_1b_1 + a_2b_2 + \cdots + a_nb_n + E_ja_xB_x + e(3)$$

In both models, \bar{y}_{ij} is the mean flowering time among replicates of individual i phenotyped in environment j where j here take the values 1 and 2 for FT10 and FT16, respectively; a_x is the genotype at QTL x coded as 0 and 2 for the homozygous minor and major allele genotypes; b are the corresponding estimated effect sizes; u is the population mean and E_j is the effect of environment on flowering time. Model 3 includes an interaction term between one of the QTL and the environment E, and was fitted for each of the QTL in turn. The significance of

530

531

532

533

534

535

536

537

538

539

540

541

542

543

544

545

546

547

548

549

550

551

552

each QTL by environment interaction was assessed using a likelihood ratio test between model 2 and model 3. Population structure was accounted for by the simultaneous fitting of the 48 loci. The analyses were performed using customised R scripts [60]. Evaluating how groups of loci contribute to flowering time variation Several of the mapped loci are only associated with flowering-time in one of the growth temperatures. We tested whether the flowering-time loci that were not significantly associated individually in one of the temperatures made a significant joint contribution by comparing the fit of the following models to the data (models 4 and 5) using a likelihood ratio test. $\overline{y} = X_1 \beta_1 + e(4)$ $\bar{y} = X_1 \beta_1 + X_2 \beta_2 + e(5)$ Here, \bar{y} , and e are defined as for model 1. Contributions from the individually significant/nonsignificant loci in the tested temperature were modelled as $X_1\beta_1/X_2\beta_2$, respectively. X_I is a similar design matrix as X in model 1, which includes a column vector of 1's for the population mean and one or more additional column vectors with the indicator regression variables for the homozygous genotypes at the SNP or SNPs included in the model (coded as 0/2, respectively). X_2 is a second design matrix only including the genotype for additional loci unique to FT10 or FT16. β_1/β_2 are the corresponding effect sizes. A likelihood ratio test was used to compare the fit of the two models using the *lrtest* function in R package *lmtest* [61]. Clustering of subpopulations based on the allele frequencies of flowering time associated loci Euclidean distances between the 8 subpopulations in the 1,001-genomes dataset defined in [22] were calculated using the dist function in R based on the allele frequencies at the 48 loci

554

555

556

557

558

559

560

561

562

563

564

565

566

567

568

569

570

571

572

573

574

575

576

associated with the flowering time at 10°C or 16°C in this dataset. The subpopulations were then clustered using the hierarchical clustering function *hclust* in R. Estimating the joint, local and global effects of all associated flowering-time loci The effects of all loci associated with flowering-time in 10°C, 16°C or both temperatures (as in model 4) were estimated simultaneously using the hglm function in hglm R-package [62]. $\overline{y} = X\beta + Zu + e (6)$ All parameters are the same as in model 1, except X in which a column vector of 1s is included to model the population mean and then n genotype columns (where the genotype for an individual at a locus is coded as 0 and 2 as above) for each of the *n* mapped flowering-time loci. The fitted values from model 6 were then used to model the contributions by global and local alleles to the flowering time. Contributions from the local alleles were estimated by $X_1\beta_1$, where X_I is the genotype matrix of the local alleles and β_1 is the corresponding estimated effects from model (6). Contributions from the global alleles were estimated by $X_2\beta_2$, where X_2 is the genotype matrix of the local alleles and β_2 is the corresponding estimated effects from model (6). Contributions from the kinship were then estimated by subtracting $X_1\beta_1$ and $X_2\beta_2$, from the modelled phenotype. Then, estimates for each subpopulation were obtained by averaging the individual estimates obtained for the accessions in it. Testing for significant long-range Linkage Disequilibrium An empirical significance threshold was derived to test for significant long-range LD between the flowering-time loci associated with flowering time at 10°C, 16°C or both. An empirical null distribution was obtained using 1,000 simulations, where in each the same number loci as

578

579

580

581

582

583

584

585

586

587

588

589

590

591

592

593

594

595

596

597

598

599

600

detected in the association analysis were simulated with allele frequencies being the same as those of the associated loci. In each simulated dataset, all pairwise LD (D') values were calculated and saved to generate a null distribution for the significance test. We used the 0.99 quantile (corresponding to a D' value of 0.96) as cut-off in our analyses. The FDR for the result was calculated as the ratio between the expected number and observed numbers of significant pairs (chose (n,2)*0.01), where n is the number of associated loci. Visualization of the results Figs 3 and 5 were created using the *heatmap.2* function in *gplots* package in R [63]. Fig 6 was created using the *LDheatmap* function in the *LDheatmap* package in R [64]. Fig S3 was made using the R package maptools [65]. All other figures were created using custom R-scripts [60]. References 1. Pardo-Diaz C, Salazar C, Jiggins CD. Towards the identification of the loci of adaptive evolution. Methods Ecol Evol. 2015;6: 445–464. doi:10.1111/2041-210X.12324 2. Olson-Manning CF, Wagner MR, Mitchell-Olds T. Adaptive evolution: evaluating empirical support for theoretical predictions. Nat Rev Genet. 2012;13: 867–77. doi:10.1038/nrg3322 3. Rockman M V. The OTN program and the alleles that matter for evolution: all that's gold does not glitter. Evolution. 2012;66: 1–17. doi:10.1111/j.1558-5646.2011.01486.x Fournier-Level a., Korte A, Cooper MD, Nordborg M, Schmitt J, Wilczek a. M. A 4. map of local adaptation in Arabidopsis thaliana. Science. 2011;334: 86–89. doi:10.1126/science.1209271

601 5. Shen X, De Jonge J, Forsberg SKG, Pettersson ME, Sheng Z, Hennig L, et al. Natural 602 CMT2 variation is associated with genome-wide methylation changes and temperature 603 seasonality. PLoS Genet. 2014;10: e1004842. doi:10.1371/journal.pgen.1004842 604 Forsberg SKG, Andreatta ME, Huang XY, Danku J, Salt DE, Carlborg Ö. The multi-6. 605 allelic genetic architecture of a variance-heterogeneity locus for molybdenum 606 concentration in leaves acts as a source of unexplained additive genetic variance. PLoS 607 Genet. 2015;11: e1005648. doi:10.1371/journal.pgen.1005648 608 7. Barboza L, Effgen S, Alonso-Blanco C, Kooke R, Keurentjes JJB, Koornneef M, et al. 609 Arabidopsis semidwarfs evolved from independent mutations in GA20ox1, ortholog to 610 green revolution dwarf alleles in rice and barley. Proc Natl Acad Sci. 2013;110: 611 15818-23. doi:10.1073/pnas.1314979110 612 8. Lamichhaney S, Berglund J, Almén MS, Maqbool K, Grabherr M, Martinez-Barrio A, 613 et al. Evolution of Darwin's finches and their beaks revealed by genome sequencing. 614 Nature. 2015;518: 371–375. doi:10.1038/nature14181 615 9. Nadeau NJ, Jiggins CD. A golden age for evolutionary genetics? Genomic studies of 616 adaptation in natural populations. Trends Genet. 2010;26: 484–92. 617 doi:10.1016/j.tig.2010.08.004 618 Flint J, Mackay TFC. Genetic architecture of quantitative traits in mice, flies, and 10. 619 humans. Genome Research. 2009. pp. 723–733. doi:10.1101/gr.086660.108 620 11. Buckler ES, Holland JB, Bradbury PJ, Acharya CB, Brown PJ, Browne C, et al. The genetic architecture of maize flowering time. Science. 2009;325: 714-718. 621 622 doi:10.1126/science.1174276 623 12. Turchin MC, Chiang CWK, Palmer CD, Sankararaman S, Reich D, Hirschhorn JN. 624 Evidence of widespread selection on standing variation in Europe at height-associated

- SNPs. Nat Genet. 2012;44: 1015–1019. doi:10.1038/ng.2368
- 626 13. Savolainen O, Lascoux M, Merilä J. Ecological genomics of local adaptation. Nat Rev
- Genet. 2013;14: 807–20. doi:10.1038/nrg3522
- 628 14. Teotónio H, Chelo IM, Bradić M, Rose MR, Long AD. Experimental evolution reveals
- natural selection on standing genetic variation. Nat Genet. 2009;41: 251–257.
- doi:10.1038/ng.289
- 631 15. Orozco-terWengel P, Kapun M, Nolte V, Kofler R, Flatt T, Schlötterer C. Adaptation
- of Drosophila to a novel laboratory environment reveals temporally heterogeneous
- trajectories of selected alleles. Mol Ecol. 2012;21: 4931–41. doi:10.1111/j.1365-
- 634 294X.2012.05673.x
- 635 16. Burke MK, Liti G, Long AD. Standing genetic variation drives repeatable experimental
- evolution in outcrossing populations of saccharomyces cerevisiae. Mol Biol Evol.
- 637 2014;31: 3228–3239. doi:10.1093/molbev/msu256
- 638 17. Pritchard JK, Di Rienzo A. Adaptation not by sweeps alone. Nat Rev Genet. 2010;11:
- 639 665–667. doi:10.1038/nrg2880
- 640 18. Graves JL, Hertweck KL, Phillips MA, Han MV, Cabral LG, Barter TT, et al.
- Genomics of parallel experimental evolution in Drosophila. Mol Biol Evol. 2017;34:
- 642 831–842. doi:10.1093/molbev/msw282
- 643 19. Collins S, De Meaux J, Acquisti C. Adaptive walks toward a moving optimum.
- Genetics. 2007;176: 1089–1099. doi:10.1534/genetics.107.072926
- 645 20. Kopp M, Hermisson J. The genetic basis of phenotypic adaptation I: fixation of
- beneficial mutations in the moving optimum model. Genetics. 2009;182: 233–49.
- doi:10.1534/genetics.108.099820
- Yeaman S. Local adaptation by alleles of small effect. Am Nat. 2015;186: S74–S89.

649 doi:10.1086/682405 650 22. Alonso-Blanco C, Andrade J, Becker C, Bemm F, Bergelson J, Borgwardt KMM, et al. 651 1,135 Genomes Reveal the Global Pattern of Polymorphism in Arabidopsis thaliana. 652 Cell. 2016;166: 481–491. doi:10.1016/j.cell.2016.05.063 653 23. Shindo C, Aranzana MJ, Lister C, Baxter C, Nicholls C, Nordborg M, et al. Role of 654 FRIGIDA and FLOWERING LOCUS C in determining variation in flowering time of 655 Arabidopsis, Plant Physiol. 2005;138: 1163–1173. doi:10.1104/pp.105.061309.1 656 24. Atwell S, Huang YS, Vilhjálmsson BJ, Willems G, Horton M, Li Y, et al. Genome-657 wide association study of 107 phenotypes in Arabidopsis thaliana inbred lines. Nature. 658 2010;465: 627–631. doi:10.1038/nature08800 659 25. Brachi B, Faure N, Horton M, Flahauw E, Vazquez A, Nordborg M, et al. Linkage and 660 association mapping of Arabidopsis thaliana flowering time in nature. PLoS Genet. 661 2010;6: 40. doi:10.1371/journal.pgen.1000940 662 Zan Y, Carlborg Ö. A multilocus association analysis method integrating phenotype 26. 663 and expression data reveals multiple novel associations to flowering time variation in 664 wild-collected Arabidopsis thaliana. Mol Ecol Resour. 2018; 1–11. doi:10.1111/1755-665 0998.12757 666 Caicedo AL, Stinchcombe JR, Olsen KM, Schmitt J, Purugganan MD. Epistatic 27. 667 interaction between Arabidopsis FRI and FLC flowering time genes generates a 668 latitudinal cline in a life history trait. Proc Natl Acad Sci. 2004;101: 15670–15675. 669 doi:10.1073/pnas.0406232101 670 28. Stinchcombe JR, Weinig C, Ungerer M, Olsen KM, Mays C, Halldorsdottir SS, et al. A 671 latitudinal cline in flowering time in Arabidopsis thaliana modulated by the flowering 672 time gene FRIGIDA. Proc Natl Acad Sci. 2004;101: 4712–4717.

- doi:10.1073/pnas.0306401101
- 674 29. Samis KE, Heath KD, Stinchcombe JR. Discordant longitudinal clines in flowering
- time and Phytochrome C in Arabidopsis thaliana. Evolution. 2008;62: 2971–2983.
- doi:10.1111/j.1558-5646.2008.00484.x
- 677 30. Sasaki E, Zhang P, Atwell S, Meng D, Nordborg M. Missing G x E Variation Controls
- Flowering Time in Arabidopsis thaliana. PLOS Genet. 2015;11: e1005597.
- doi:10.1371/journal.pgen.1005597
- 680 31. Li Y, Cheng R, Spokas KA, Palmer AA, Borevitz JO. Genetic variation for life history
- sensitivity to seasonal warming in Arabidopsis thaliana. Genetics. 2014;196: 569–577.
- doi:10.1534/genetics.113.157628
- 683 32. Stratton DA. Reaction norm functions and QTL-environment interactions for flowering
- time in Arabidopsis thaliana. Heredity. 1998;81: 144–155. doi:10.1038/sj.hdy.6883690
- 685 33. Allard RW, Babbel GR, Clegg MT, Kahler a L. Evidence for coadaptation in Avena
- barbata. Proc Natl Acad Sci. 1972;69: 3043–8. doi:10.1073/pnas.69.10.3043
- 687 34. Mitton JB, Linhart YB, Hamrick JL, Beckman JS. Observations on the genetic
- structure and mating system of ponderosa pine in the Colorado front range. Theor Appl
- 689 Genet. 1977;51: 5–13. doi:10.1007/BF00306055
- 690 35. Barker JSF, Frydenberg J, Sarup P, Loeschcke V. Altitudinal and seasonal variation in
- microsatellite allele frequencies of Drosophila buzzatii. J Evol Biol. 2011;24: 430–439.
- 692 doi:10.1111/j.1420-9101.2010.02180.x
- 693 36. Liang G, He H, Li Y, Wang F, Yu D. Molecular Mechanism of microRNA396
- Mediating Pistil Development in Arabidopsis. Plant Physiol. 2014;164: 249–258.
- doi:10.1104/pp.113.225144
- 696 37. Golembeski GS, Imaizumi T. Photoperiodic Regulation of Florigen Function in

697 Arabidopsis thaliana. Arab B. 2015;13: e0178. doi:10.1199/tab.0178 698 38. Hewezi T, Maier TR, Nettleton D, Baum TJ. The Arabidopsis MicroRNA396-699 GRF1/GRF3 Regulatory Module Acts as a Developmental Regulator in the 700 Reprogramming of Root Cells during Cyst Nematode Infection. PLANT Physiol. 701 2012;159: 321–335. doi:10.1104/pp.112.193649 702 39. Lee H, Yoo SJ, Lee JH, Kim W, Yoo SK, Fitzgerald H, et al. Genetic framework for 703 flowering-time regulation by ambient temperature-responsive miRNAs in Arabidopsis. 704 Nucleic Acids Res. 2010;38: 3081–3093. doi:10.1093/nar/gkp1240 705 40. Teotia S, Tang G. To bloom or not to bloom: Role of micrornas in plant flowering. Mol 706 Plant. 2015;8: 359–377. doi:10.1016/j.molp.2014.12.018 707 41. Wang JW. Regulation of flowering time by the miR156-mediated age pathway. Journal 708 of Experimental Botany. 2014. pp. 4723–4730. doi:10.1093/jxb/eru246 709 42. Choi K, Kim J, Hwang H-J, Kim S, Park C, Kim SY, et al. The FRIGIDA Complex 710 Activates Transcription of FLC, a Strong Flowering Repressor in Arabidopsis, by 711 Recruiting Chromatin Modification Factors. Plant Cell. 2011;23: 289–303. 712 doi:10.1105/tpc.110.075911 713 43. Martin-Trillo M. EARLY IN SHORT DAYS 1 (ESD1) encodes ACTIN-RELATED 714 PROTEIN 6 (AtARP6), a putative component of chromatin remodelling complexes 715 that positively regulates FLC accumulation in Arabidopsis. Development. 2006;133: 716 1241-1252. doi:10.1242/dev.02301 717 Bond DM, Wilson IW, Dennis ES, Pogson BJ, Jean Finnegan E. Vernalization 44. 718 insensitive 3 (VIN3) is required for the response of Arabidopsis thaliana seedlings 719 exposed to low oxygen conditions. Plant J. 2009;59: 576–587. doi:10.1111/j.1365-720 313X.2009.03891.x

- 721 45. Hall A. The TIME FOR COFFEE Gene Maintains the Amplitude and Timing of
- 722 Arabidopsis Circadian Clocks. PLANT CELL ONLINE. 2003;15: 2719–2729.
- 723 doi:10.1105/tpc.013730
- 724 46. Kim J, Kim Y, Yeom M, Kim J-H, Nam HG. FIONA1 Is Essential for Regulating
- Period Length in the Arabidopsis Circadian Clock. Plant Cell. 2008;20: 307–319.
- 726 doi:10.1105/tpc.107.055715
- 727 47. Zhang B, Wang L, Zeng L, Zhang C, Ma H. Arabidopsis TOE proteins convey a
- 728 photoperiodic signal to antagonize CONSTANS and regulate flowering time. Genes
- 729 Dev. 2015;29: 975–987. doi:10.1101/gad.251520.114.
- 48. Li Y, Huang Y, Bergelson J, Nordborg M, Borevitz JO. Association mapping of local
- climate-sensitive quantitative trait loci in Arabidopsis thaliana. Proc Natl Acad Sci.
- 732 2010;107: 21199–21204. doi:10.1073/pnas.1007431107
- 733 49. Dittmar EL, Oakley CG, Ågren J, Schemske DW. Flowering time QTL in natural
- populations of *Arabidopsis thaliana* and implications for their adaptive value. Mol
- 735 Ecol. 2014;23: 4291–4303. doi:10.1111/mec.12857
- 736 50. Rus A, Baxter I, Muthukumar B, Gustin J, Lahner B, Yakubova E, et al. Natural
- variants of AtHKT1 enhance Na+ accumulation in two wild populations of
- 738 Arabidopsis. PLoS Genet. 2006;2: 1964–1973. doi:10.1371/journal.pgen.0020210
- 739 51. Salomé PA, Bomblies K, Laitinen RAE, Yant L, Mott R, Weigel D. Genetic
- architecture of flowering-time variation in Arabidopsis thaliana. Genetics. 2011;188:
- 741 421–433. doi:10.1534/genetics.111.126607
- 742 52. Gould BA, Stinchcombe JR. Population genomic scans suggest novel genes underlie
- convergent flowering time evolution in the introduced range of *Arabidopsis thaliana*.
- 744 Mol Ecol. 2017;26: 92–106. doi:10.1111/mec.13643

- 53. Exposito-Alonso M, Vasseur F, Ding W, Wang G, Burbano HA, Weigel D. Genomic
- basis and evolutionary potential for extreme drought adaptation in Arabidopsis
- 747 thaliana. Nature Ecology and Evolution. 2017: 1–7. doi:10.1038/s41559-017-0423-0
- 748 54. Tilman D, Lehman C. Human-caused environmental change: Impacts on plant
- diversity and evolution. Proc Natl Acad Sci. 1987;98: 5433–5440.
- 750 doi:10.1073/pnas.091093198
- 751 55. Kawakatsu T, Huang S shan??Carol, Jupe F, Sasaki E, Schmitz RJ, Urich MA, et al.
- Epigenomic Diversity in a Global Collection of Arabidopsis thaliana Accessions. Cell.
- 753 2016;166: 492–506. doi:10.1016/j.cell.2016.06.044
- 754 56. Consortium 1001genomes. FT10. In: 1,135 Genomes Reveal the Global Pattern of
- Polymorphism in Arabidopsis thaliana. [Internet]. 1 Jan 2016.
- 756 doi:10.21958/PHENOTYPE:261
- 757 57. Consortium 1001genomes. FT16. In: 1,135 Genomes Reveal the Global Pattern of
- Polymorphism in Arabidopsis thaliana. [Internet]. 1 Jan 2016.
- 759 doi:10.21958/PHENOTYPE:262
- 760 58. Korte A, Vilhjálmsson BJ, Segura V, Platt A, Long Q, Nordborg M. A mixed-model
- approach for genome-wide association studies of correlated traits in structured
- 762 populations. Nat Genet. 2012;44: 1066–1071. doi:10.1038/ng.2376
- 763 59. Aulchenko YS, Ripke S, Isaacs A, van Duijn CM. GenABEL: An R library for
- genome-wide association analysis. Bioinformatics. 2007;23: 1294–1296.
- doi:10.1093/bioinformatics/btm108
- 766 60. R Core Team. R Core Team, 2015 R: A Language and Environment for Statistical
- Computing. R Found. Stat. Comput. Vienna Austria URL: https://www.R-project.org/.
- 768 [Internet]. R Foundation for Statistical Computing Vienna Austria. 2015. doi:ISBN 3-

769 900051-07-0 770 Zeileis A, Hothorn T. Diagnostic checking in regression relationships. R News. 2002;2: 61. 771 7–10. Available: http://cran.r-project.org/doc/Rnews/ 772 62. Rönnegård L, Shen X, Alam M. hglm: A Package for Fitting Hierarchical Generalized 773 Linear Models. R J. 2010;2: 20–28. Available: http://journal.r-774 project.org/archive/2010-2/RJournal 2010-2 Roennegaard~et~al.pdf 775 Warnes GR, Bolker B, Bonebakker L, Gentleman R, Liaw WHA, Lumley T, et al. 63. 776 gplots: Various R Programming Tools for Plotting Data. In: R package version 2.17.0. 777 [Internet]. 2015 p. 2015. doi:10.1111/j.0022-3646.1997.00569.x 778 Shin J-H, Blay S, Graham J, McNeney B. LDheatmap: An R Function for Graphical 64. 779 Display of Pairwise Linkage Disequilibria Between Single Nucleotide Polymorphisms. 780 J Stat Softw. 2006;16. doi:10.18637/jss.v016.c03 781 65. Lewin-Koh N, Biyand R. Maptools: Tools for reading and handling spatial objects 782 [Internet]. R package version 2011. p. 74. Available: http://cran.r-783 project.org/package=maptools 784 785

787

788

789

790

791

792

793

794

795

796

797

798

799

800

801

802

803

804

805

806

807

808

809

Figures and figure captions Fig 1. Flowering time measured at 10 and 16 °C, correlation with latitude/temperature, and the subpopulation differentiation. (A). Histogram of flowering time measured at 10°C (blue) and 16 °C (tomato). Flowering time under controlled green house condition is significantly correlated both with latitude (B) and temperature (C) at the sampling sites, and vary amongst the earlier defined subpopulations (**D**), of the 1,004 analysed accessions from the 1,001-genomes A. thaliana collection [22]. Fig 2. Additive effects of 48 loci on flowering time measured at 10 and 16 °C. (A) Additive genetic effects of 48 flowering time loci on FT10 (blue) and FT16 (tomato). The yaxis indicates the additive effects on flowering time and the x-axis the respective loci (SNPs) sorted by the differences in additive effects (FT16-FT10) from left to right. The estimates were obtained by fitting all 48 loci jointly in a mixed model. The bars represent standard errors of the effect estimates. The 18 loci with significant genotype by temperature interactions are indicated with a black star. (B) Correlation between the additive genetic effects for the 48 loci estimated for accessions grown at 10 and 16 °C. The colour of the dots indicates whether the locus is significant i) only for FT10 (blue), only for FT16 (tomato), for both FT10 and FT16 (purple), or the difference between the two (FT10-FT16; black). Fig 3. Flowering times, allele frequencies of flowering time associated loci and similarities of these between eight subpopulations of the A. thaliana 1,001-genomes collection. (A) Flowering times for the 8 subpopulations in this data defined by [22] are modelled well by the additive effects of the 48 flowering time loci and the kinship. (B) Distribution of allele frequencies for the 48 loci in the 8 subpopulations. Intensity of 810

811

812

813

814

815

816

817

818

819

820

821

822

823

824

825

826

827

828

829

830

831

832

833

illustrates frequency of early/late-flowering alleles, respectively. subpopulations are sorted from left to right by increasing average flowering time (columns) and the loci from top to bottom by decreasing additive effects on flowering time (rows). X:YYYYY-ZZZ represent a locus as Chromosome:Position-Type, where Position is in bp and Type is either "QTL", "Diff" or "locus-id" indicating its detection by direct association to FT10, FT16 or (FT10-FT16) in a GWAS ("QTL"/"Diff") or via a genetically regulated expression of the locus "locus-id" in the eQTL analysis. (C) Dendrogram showing how the 8 subpopulations cluster based on the allele frequency spectrum across the 48 associated loci. Fig 4. Modelling of the flowering times of the eight subpopulations in the 1,001-genomes A. thaliana collection and how the individual associated loci contribute. Experimentally measured and modelled flowering times in the 8 genetically defined subpopulations of the A. thaliana 1,001-genomes collection of accessions [22] grown at 10°C (A) and 16 °C (B). The height of the bars represents the experimentally observed mean flowering times for the accessions belonging to the sub population. The kinship (purple) and additive effects of globally segregating alleles (grey) capture the overall pattern of early to late flowering times between the sub populations well. The additive effects of the local alleles (blue) improve the fit to more extreme adaptations of some sub populations, in particular in Spain and Sweden. The differences between measured and modelled flowering times (tomato) are small at both temperatures. The modelled contributions by the global alleles are larger at 16°C than at 10°C due to their genotype-by-temperature interactions (Fig. 2A). The relationship between the variance contributed by each individual locus and its allele frequency/additive effect is illustrated for FT10 (C) and FT16 (D). The y-axis gives the minor allele frequency and the xaxis the estimated additive effect on flowering time in days. The size of each dot is

834

835

836

837

838

839

840

841

842

843

844

845

846

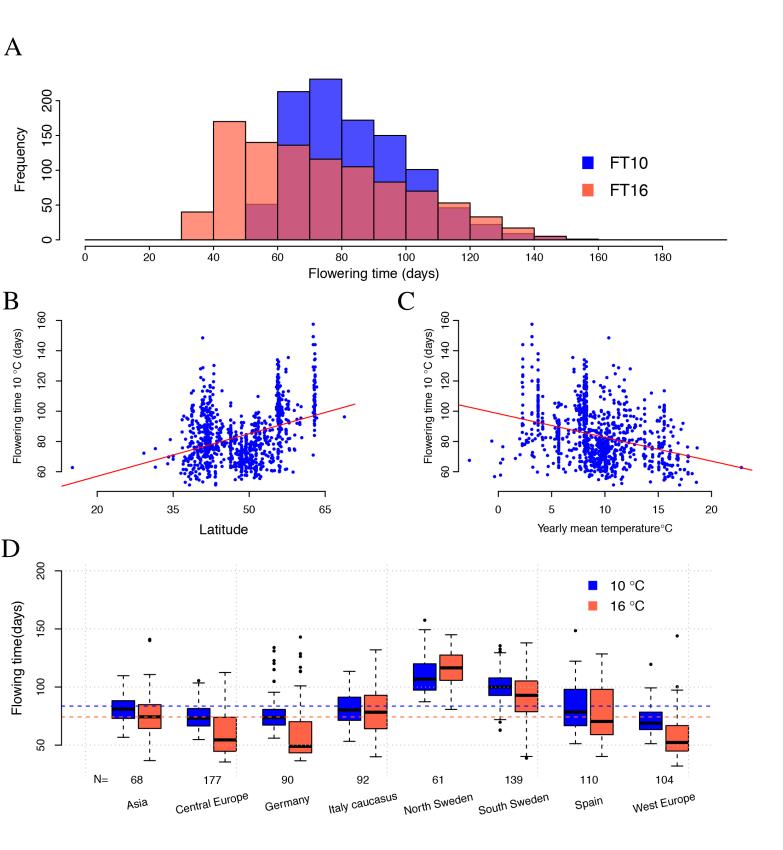
847

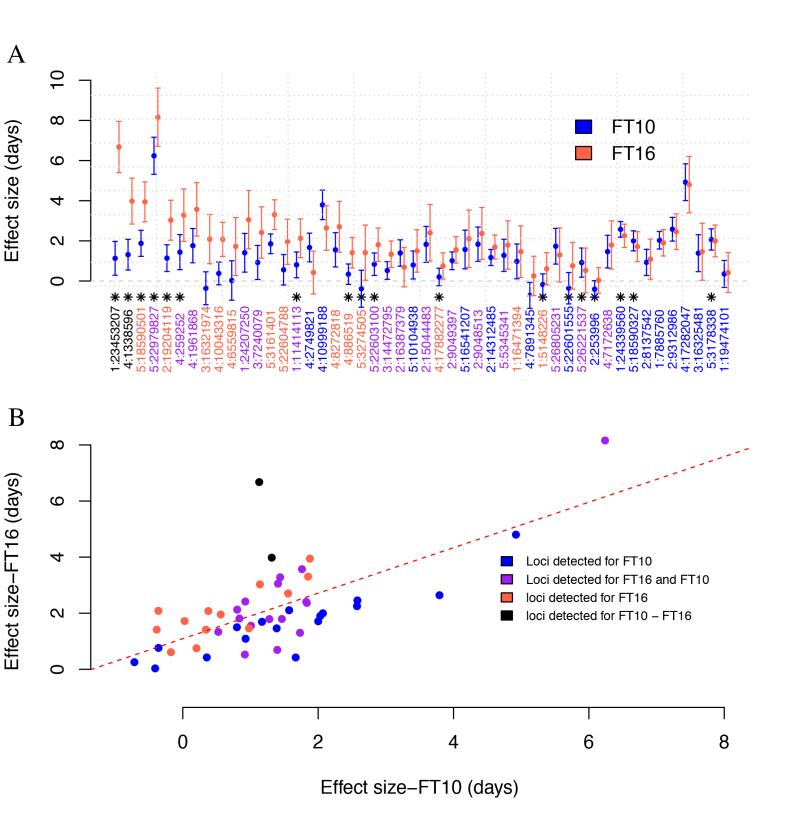
848

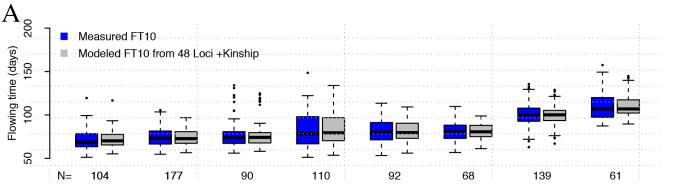
849

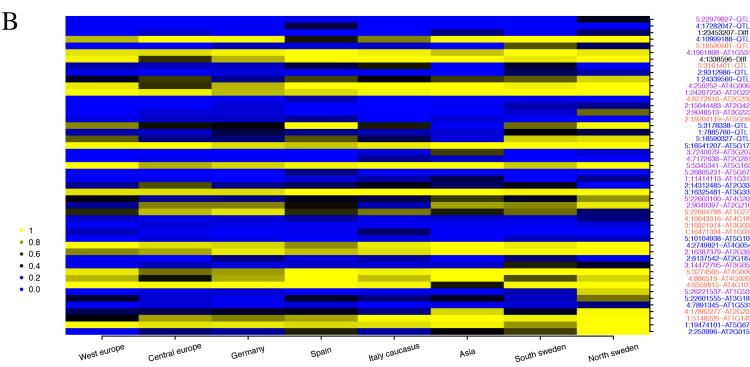
850

proportional to the phenotypic variance explained by the locus. The colour of the dot indicates whether the minor allele is defined as a global (tomato) or local (blue) allele in the population. Fig 5. Allele frequencies for loci where one variant is rare (MAF < 0.05) in at least one of the eight 1,001-genomes A. thaliana subpopulations. Blue/yellow indicates high allele frequency for the early/late flowering alleles, respectively. The populations are clustered based on the overall similarity of the allele frequency pattern (left to right) and the loci based on the similarity of the allele frequencies in the subpopulations (top to bottom). X:YYYYY-ZZZ represent a locus as chromosome: position-type, where Position is in bp and Type is either "QTL", "Diff" or "locus-id" indicating its detection by direct association to FT10, FT16 or (FT10-FT16) in a GWAS ("QTL"/"Diff") or via a genetically regulated expression of the locus "locus-id" in the eQTL analysis. Fig 6. Pairwise linkage disequilibrium (LD) between the 48 flowering time-associated loci across the genome (D'). Values above the 0.99 quantile of a simulated null distribution are highlighted as yellow. The loci are labelled as chromosome: position on chromosome in bp.

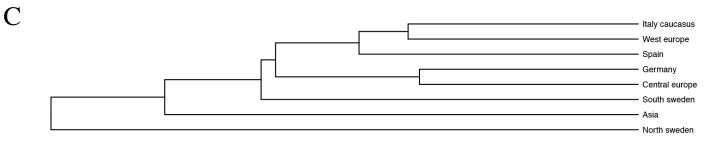


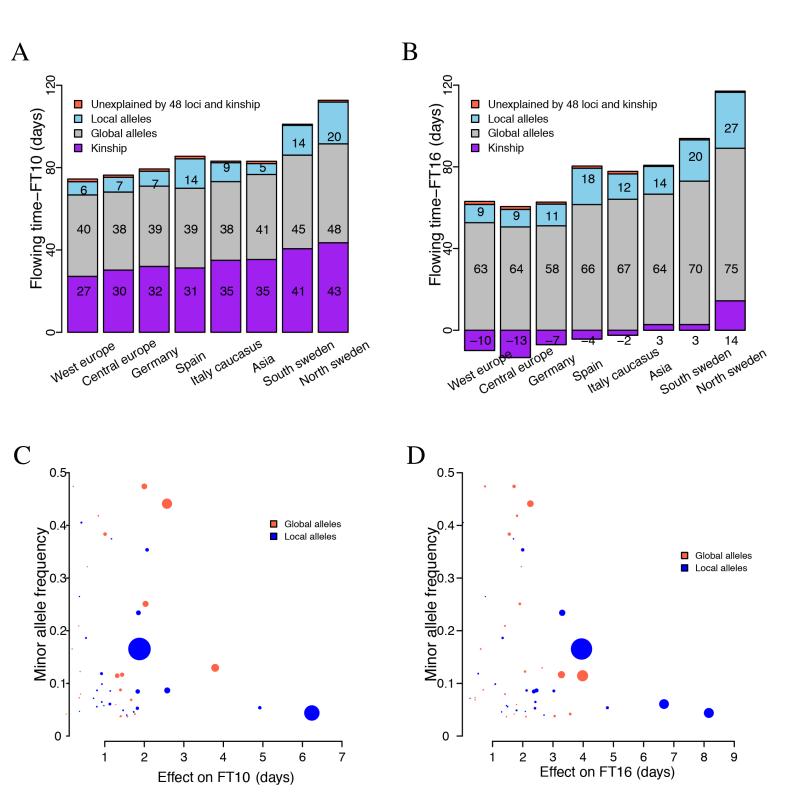


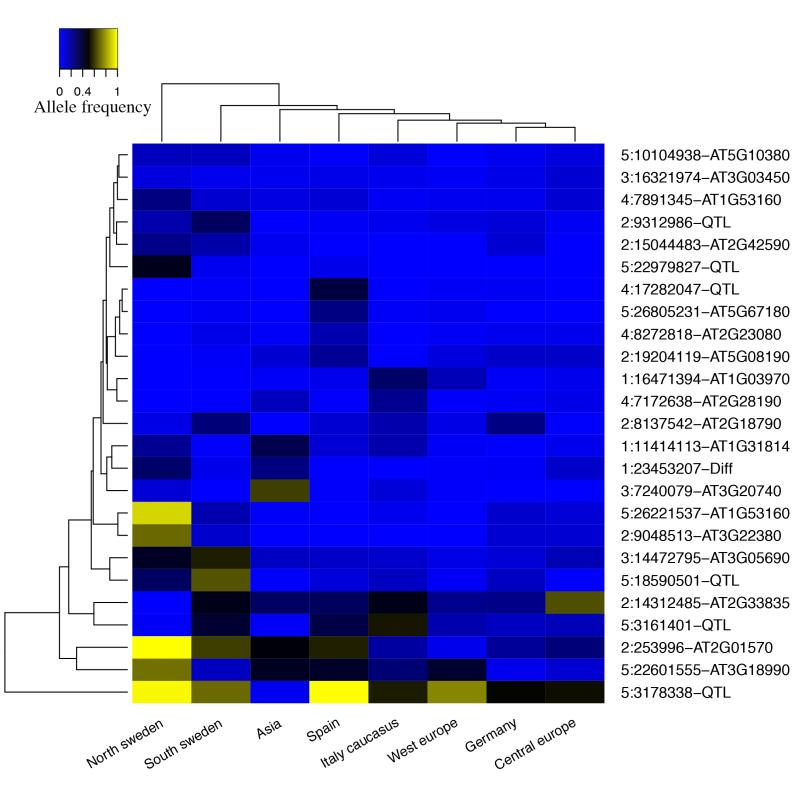




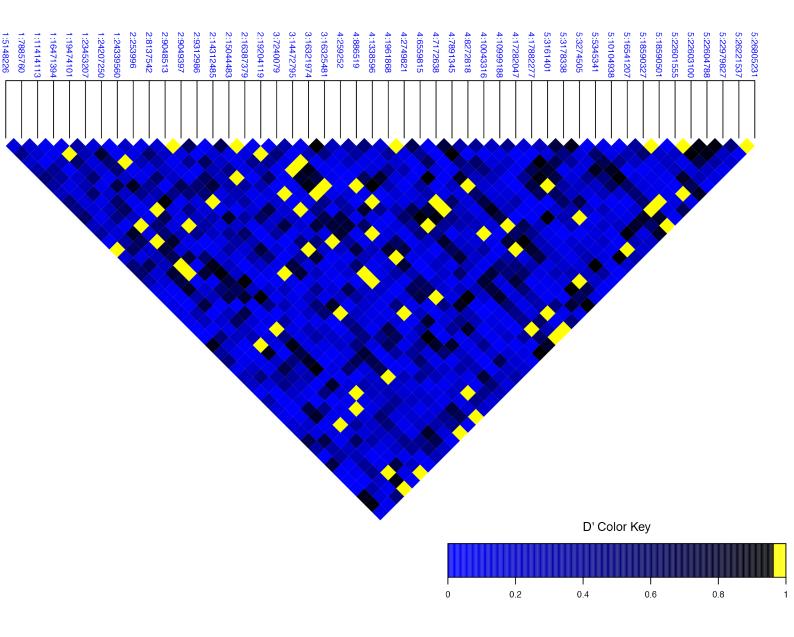
2:9048513-A13G22380 2:19204119-AT5G08190 5:3178338-QTL 1:7885760-QTL 5:16590327-QTL 5:16541207-AT5G17490 3:7240079-AT3G20740 4:7172638-AT2G28190 5:5345341-AT5G16320



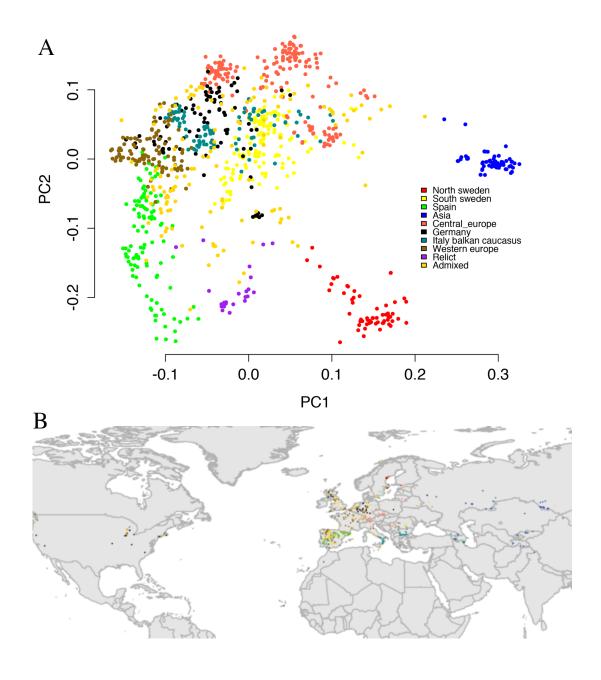




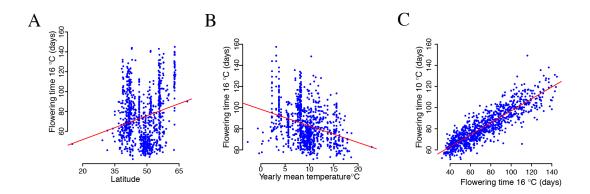
Pairwise LD measured by D'



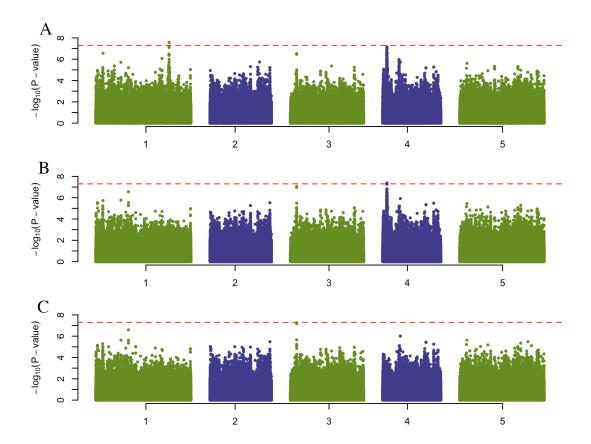
Supplementary information for "Explorations of the polygenic genetic architecture of flowering time in the worldwide *Arabidopsis thaliana* population" by Zan and Carlborg.



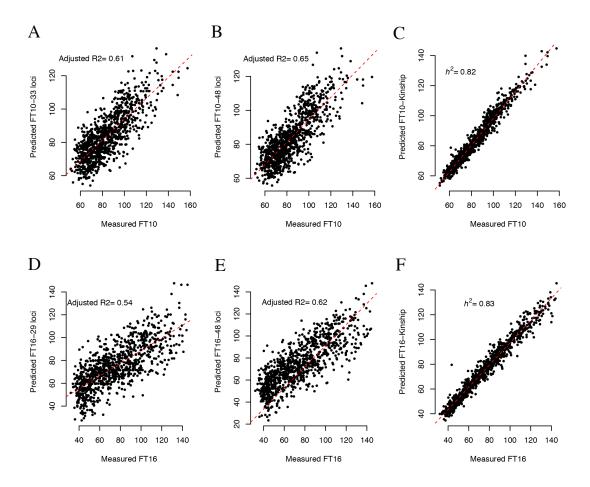
S1 Fig. Population structure and sampling locations for the accessions in the **1,001-genomes** *A. thaliana* collection. (A) Population structure indicated by the first two principal components of the IBS (Identity By State) kinship matrix with the subpopulations defined by [22] highlighted in colour. (B) Sampling locations of the analysed accessions.



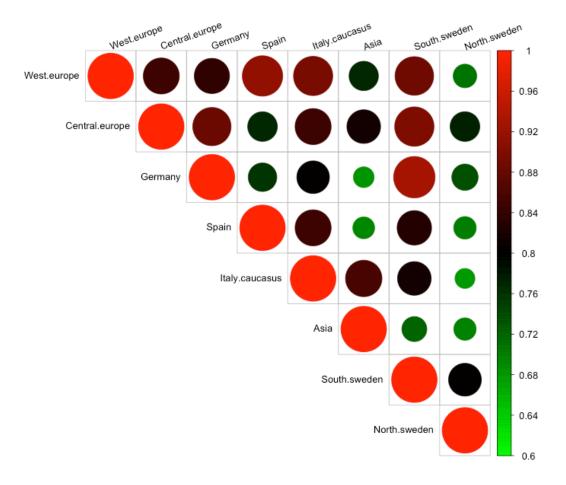
S2 Fig. Geographical differentiation, and correlations, of the flowering times for the accessions of the 1,001-genomes *A. thaliana* collection grown at 10 and 16°C. Flowering times measured for accessions grown at 16°C in the green house are significantly correlated both with latitude (**A**) and temperature (**B**) at the sampling sites for the 1,004 analysed accessions from the 1,001-genomes *A. thaliana* collection [22]. (**C**) There is a high correlation between flowing times measured for accessions grown at 10 and 16°C.



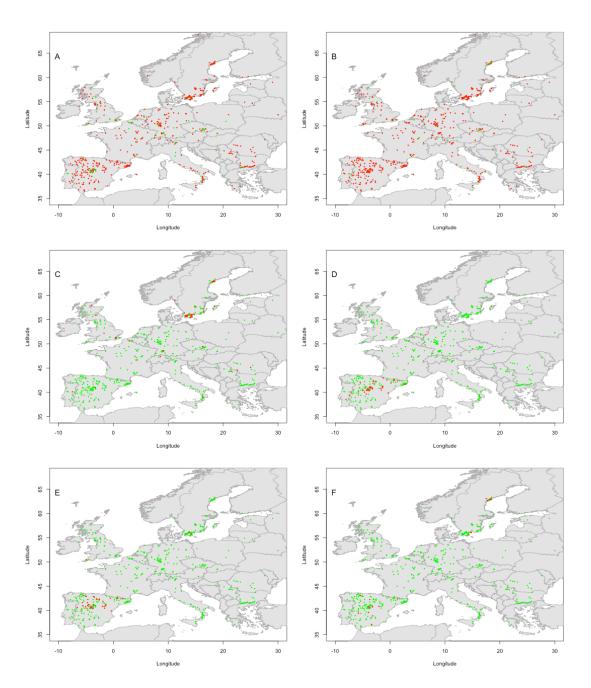
S3 Fig. Results from the GWAS analysis using the differences in flowering times at 10 and 16°C for the accessions of the 1,001-genomes *A. thaliana* collection. (A) Manhattan plot for a GWAS using FT10-FT16 as phenotype where one locus on Chromosome 1 at 23,453,207 bp reached genome wide significance. (B) Manhattan plot for a GWAS using FT10-FT16 as phenotype with the locus identified in (A) as covariate, where a second locus on Chromosome 4 at 1,338,596 bp reached genome wide significance. (C) Manhattan plot for a GWAS using FT10-FT16 as phenotype with the 2 loci identified in (A) and (B) as covariates, where no additional locus was detected.



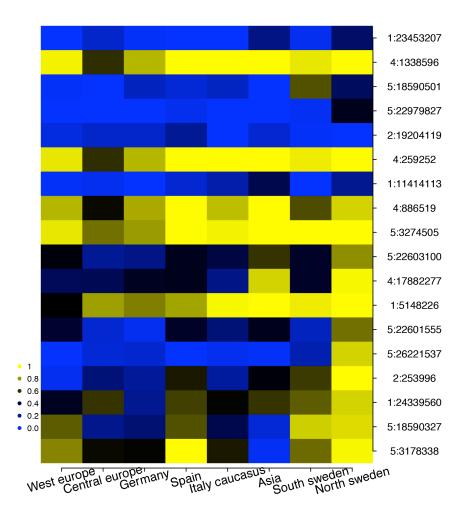
S4 Fig. Modelled and experimentally measured flowering times for the accessions of the 1,001-genomes *A. thaliana* **collection.** Modelled phenotypes for the 1,001-genomes accessions using the 33 (29) loci detected in the independent analyses of FT10/FT16 and the experimentally measured phenotypes are illustrated in (A) and (D). Modelled flowering times using all 48 flowering time loci and measured phenotypes are illustrated in (B) and (E). Modelled flowering times from kinship and measured phenotypes for FT10/FT16 are illustrated in (C) and (F).



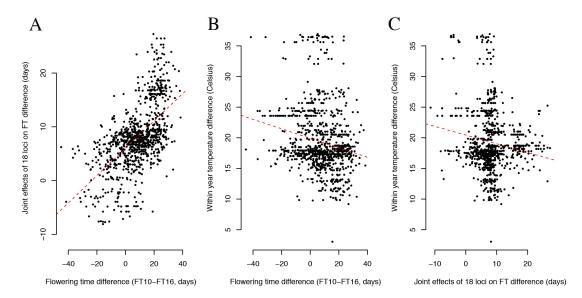
S5 Fig. Illustrations of the overall similarities in the allele frequency spectrum across the sub-populations as Spearman correlation between the allele-frequencies across the 48 associated loci



S6 Fig. The geographical distribution of alleles at 6 selected loci associated with flowering-time at 10°C (FT10). The five panels illustrates the geographical distributions for the six associated loci at Chromosome 2:16,387,379 bp (**A**), Chromosome 4:1,961,868 bp (**B**), Chromosome 2: 9,312,968 bp (**C**), Chromosome 4:1,728,204 bp (**D**), Chromosome 5:26,805,231bp (**E**) and Chromosome 5:2,297,9827 bp (**F**). Red/green colours indicate the allele that increases/decreases FT10, respectively.



S7 Fig. Allele frequencies for the 18 loci showing significant genotype by temperature interactions across the 8 sub populations of the 1,001-genomes collection. Blue/yellow indicates high allele frequency for early/late flowering alleles, respectively. The populations are sorted based on early to late flowering (left to right) and large to small additive effects on the flowering time difference (FT10-FT16; top to bottom). Genotype by temperature interacting alleles that have a larger effects on FT16 than on FT10 are enriched in the North Sweden sub population.



S8 Fig. Correlations between observed and modelled flowering time differences for accessions grown at 10°C and 16°C and the within year temperature fluctuations at their sampling locations. (A) The correlation between the measured flowering time difference (FT10-FT16) and the flowering time difference modelled by the joint effects the 18 loci with significant genotype by temperature interactions (Adjusted R-square = 0.34; P-value = 1.3×10^{-90} ; Fig 2A). (B) There is a significant correlation (Adjusted R-squared = 0.04; P-value = 1.7×10^{-10}) between the within year temperature fluctuation (temperature difference between hottest and coldest day) at the sampling locations of the accessions [22] and the measured flowering time difference (FT10-FT16). (C) There is also a significant correlation (Adjusted R-squared = 0.03; P-value = 1.2×10^{-7}) between the within year temperature fluctuation (temperature difference between hottest and coldest day) [22] and the modelled flowering time differences for the accessions from the joint effects of the 18 loci with significant genotype by temperature interactions (Fig 2A).