

In utero exposure to cannabidiol disrupts select early-life behaviors in a sex-specific manner

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Abstract

Cannabidiol (CBD), one of the main components of cannabis, is generally considered safe, despite the lack of studies on the possible consequences of its consumption during critical periods of neurodevelopment, including prenatal life. Although CBD crosses the placenta and its use during pregnancy is steadily increasing, the impact of gestational CBD exposure on early life is unknown. Here, we combined behavioral exploration and deep learning to assess how *in utero* exposure to low doses of CBD alters pre-weaning behaviors in mouse pups of both sexes. The data reveal that pups from CBD-treated dams exhibit sex-specific alterations in weight growth, homing behavior, and the syllabic repertoire of ultrasonic vocalizations. Thus, prenatal CBD is associated with alterations in innate behavioral responses and communication skills.

Introduction

While Δ^9 -THC is the component of most concern in *Cannabis sativa* L. in terms of prenatal exposure, the plant contains over 300 compounds, including cannabidiol (CBD). Although structurally similar, CBD does not induce the psychotropic effects classically associated with Δ^9 -THC (1,2). Consequently, CBD is globally perceived as safe and free of harmful side effects. Its clinical interest is due to its potential benefits as a natural antipsychotic, anti-nociceptive, anticonvulsant, antiemetic, anxiolytic, anti-inflammatory, antioxidant, and neuroprotective agent (3–6). Despite the lack of scientific evidence regarding safety of CBD during gestation, pregnant women use CBD for a plethora of pregnancy-related symptoms including nausea, insomnia, anxiety, and chronic pain (7). CBD crosses the placenta and alters its very structure, both of which can have a significant impact on pregnancy outcomes (8–10). Moreover, a recent study showed that extended exposure of CD1 mice to CBD spanning from gestation through the first week after birth alters repetitive and hedonic behaviors in the adult progeny (11). The paucity of preclinical data on the impact of *in utero* CBD exposure prompted us to investigate the postnatal impact of gestational CBD exposure to assess potential risks associated with CBD use during this period. The data reveal that pups from CBD-treated dams exhibit previously unknown sex-specific cognitive alterations in early-life.

Materials and Methods

Animals

Male and female C57BL6/J (8-10 weeks age) were purchased from Janvier Lab and housed in standard wire-topped Plexiglas cages (42 cmx27 cmx14 cm), in a temperature and humidity-controlled condition (i.e., temperature $21 \pm 1^\circ\text{C}$, $60 \pm 10\%$ relative humidity and 12 h light/dark cycles). Food and water were available *ad libitum*. After one week of acclimation, the female pairs were placed with a single male mouse in the late afternoon. The morning the vaginal plug was found was designated as day 0 of gestation (GD0) and pregnant mice were housed individually. From GD5 to GD18, dams were injected subcutaneously (s.c.) daily with vehicle or 3 mg/Kg of CBD (Nida Drug Supply Program), dissolved in a vehicle consisting of Cremophor EL (Sigma-Aldrich), ethanol, and saline at 1:1:18 ratios, and administered at volume of 4 mL/Kg. Control dams (SHAM) were injected the same volume of vehicle solution. This dose of CBD reaches the embryonic brain and cause some behavioral changes in the offspring (11). For each litter, the date of birth was designated as postnatal day (PND) 0. The behavioral tests were performed in male and female offspring during perinatal period (PNDs 10 and 13). Body weight of SHAM and CBD pups was measured every 3 days until one day after weaning (PND 22).

All procedures were performed in conformity with the European Communities Council Directive (86/609/EEC) and the United States NIH Guide for the Care and Use of Laboratory Animals. The French Ethical committee authorized the project APAFIS#3279.

Behavioral tests

Ultrasound vocalizations (USVs)

USVs were induced by quick maternal separation in male and female pups at PND 10 as previously described (12). Each tested mouse was placed into an empty plastic container (11 x 7 x 3.5 cm), located inside a sound-attenuating isolation box (32 x 21 x 14 cm). USVs were recorded using an ultrasonic microphone (Ultravox Noldus), connected via the Ultravox device (Noldus, Netherlands) and placed 20 cm above the pup in its plastic container. At the end of 4-minute recording session, each pup was weighed, and the body temperature checked.

Acoustic analysis was performed using DeepSqueak (Version 2.6.2), a deep learning-based software for the detection and analysis of USV (for more details see, (13)). The audio file was individually transferred into DeepSqueak, converted in the corresponding sonograms, and analyzed using a Faster-RCNN object detector. The lower and higher cut-off frequency, 20 kHz and 100 kHz respectively, were applied in order to reduce the background noise outside the relevant frequency. Once detected, each sonogram was converted in the corresponding spectrogram and the calls identified as noise were manually removed. Call classification, transitional probabilities and syntax analysis were performed automatically with DeepSqueak's built in mouse call classification neural network. USV parameters were classified based on a quantitative and qualitative analysis. The *quantitative* analysis included the percentage of vocalizing and non-vocalizing mice in each group, the number of calls, the latency to the first vocalization (in sec.), the mean call duration (in msec), and the mean dominant frequency (in kHz). Whereas the *qualitative* analysis focused on the study of vocal repertoire based on syntax analysis (i.e., different categories of calls) and the transitional probabilities for each group. The latter was expressed as the probability that one type of call followed the previous one and the following calls were indicated on the x-axis.

Homing test

The homing test was performed as published (14), in SHAM and CBD mice previously tested for the USVs. At PND 13 both male and female pups were separated from the dam, and kept for 30 min in a different cage on a heating pad set at the temperature of 35°C. Each tested mouse was placed in the Plexiglas cage (21x 15 cm) which had one-third of the litter from the pup's original cage and two-thirds of clean litter. The latter was considered as the unfamiliar area, while the one with the old litter was the nest area. The pup was located at the edge of the clean bedding and its behavior was videorecorded for the following 4 min. Homing performance was performed using Ethovision and considering the locomotory activity (in cm.), the velocity (in cm/sec), the moving time (in cm), the latency to reach the nest (in sec.) and the distance moved (in cm), the time spent (in sec.) and the entries in the nest and in the unfamiliar area.

Statistical analysis

Statistics were performed with GraphPad Prism 9 and DeepSqueak 2.6.2. Behavioral data were tested for the Normality and statistical analysis was performed with Multiple Mann-Whitney *U* test. N values corresponds to the number of animals tested for each group. When achieved, the significance was expressed as exactly *p*-value in the figures. The ROUT test was applied to all data sets to identify outliers, which were then excluded from the data sets.

Results

Gestational CBD affect postnatal growth in a sex specific manner.

Prenatal cannabis exposure influences neonatal outcome in multiple ways (15) and preclinical data indicate that gestational THC reduces body weight in early life. In contrast, the effects of *in utero* CBD exposure are unknown. Dams were given a low dose of CBD (3 mg/Kg, s.c.) or a vehicle (SHAM) once daily from GD5 to GD18 and their pups' body weights were monitored throughout the perinatal period until weaning (PND 10-22; for more details see Table 1). Body weights of pups from CBD-exposed dams were consistently higher than those of SHAM dams (Fig. 1). Remarkably, the increase in body weight was observed exclusively in male offspring (Fig. 1A), whereas the weight of *in utero* exposed females was indistinguishable from that of SHAM females (Fig. 1B). Thus, gestational CBD alters the growth trajectory in a sex-specific manner.

Gestational CBD modifies the coarse characteristics of ultrasonic communications in a sex-specific manner.

Offspring-mother communication is necessary for mouse pups, who emit ultrasonic vocalizations (USV) to convey their emotional conditions (16). Thus, upon separation from their mother and nest, rodent pups vocalize to engage maternal care (17). Perinatal cannabinoids (i.e., Δ^9 -THC or cannabimimetics) negatively impact neurodevelopment (18–21) and strongly alter USV emissions (22,23). We first quantified the coarse characteristics (i.e., number, latency, duration, and frequency) of separation-induced calls emitted by pups in our different groups (Fig. 2 and Table 2). Prenatal CBD altered the proportion of vocalizing and non-vocalizing pups: more males in the CBD-exposed groups did not vocalize at all during the recording session compared with all other groups (Fig. 2A-B). The total number and latency of first vocalization were similar across treatments and sex (Fig. 2C-D). However, marked differences in the sex-specific effects of prenatal CBD were evident in the mean duration and mean frequency of USVs (Fig. 2E-F).

CBD males made shorter calls (Fig. 2E), whereas CBD females made more high frequency calls (Fig. 2F) than their SHAM counterparts. The probability distribution of USV frequency followed a bi-modal distribution in CBD-exposed males but not in females (Fig. 2G-H). Finally, a minor mode corresponding to high frequency calls was specifically observed in CBD-exposed males (Fig. 2G).

Prenatal CBD modifies the syllabic repertoire of ultrasonic communication in a sex-specific manner.

Does prenatal CBD alter vocalization patterns in pups? To address this question, we analyzed the amount and spectral characteristics of USVs detected in our different groups with DeepSqueak (13). We first compared the vocal repertoire of SHAM and CBD pups of both sexes. Call categorization (Fig. 3 and Table 3) showed that, while the number of USVs was similar, CBD gestation largely affected the vocal repertoire (Fig. 3). Thus, the proportion of each type of call made during the 5-min test was compared in male and female CBD pups and their control counterparts (Fig. 3B-C-E-F). Notably, male and female CBD pups showed a significant reduction in Complex Trill, Downward Ramp, and Inverted-U calls (Fig. 3D, G) compared with SHAM pups. Furthermore, only CBD male pups showed a significant increase in Short, Trill, and Step-up call (Fig. 3D). In addition, we observed that CBD females emitted significantly less Flat calls compared to SHAM females (Fig. 3G). Thus, call syntax analysis showed multiple sex-specific differences in the vocal repertoire of CBD-exposed pups.

CBD *in utero* exposure changes the complexity of communication in a sex-specific manner.

To test whether the differences found in the syllable repertoire reflect a different development of communication complexity in CBD-exposed pups, we next examined the "call order probability." Thus, we analyzed the most frequently occurring call combinations (Fig. 4 and Table 4). Certain call sequences were similar in CBD and SHAM males. Indeed, the Inverted-U, Upward ramp, and Complex calls were followed by another Downward Ramp, Upward ramp, and Complex call, respectively, with a similar probability in both CBD and SHAM males (Fig. 4A-B). CBD male showed a significantly lower probability towards the use of Downward Ramp and Complex Trill calls compared to SHAM male pups (Fig. 4C). In the contrast, Trill call was more often used by CBD than SHAM males (Fig. 4C).

Finally, we observed a significantly lower probability for the use of Complex, Downward Ramp, and Short calls in CBD females compared to SHAM females (Fig. 4F).

CBD prenatal exposure impact the motor and discriminative skills during early development in a sex-specific manner.

The Homing Test allows the investigation of complex abilities, such as sensory, motor, and odor-detection skills. Thus, we examined homing behavior in PND13, CBD- and SHAM pups (Fig. 5 and Table 5). Gestational CBD significantly reduced the total distance moved (Fig. 5A) only in CBD female pups. Interestingly, these CBD-exposed pups moved slower compared to SHAM female pups (Fig. 5B) and spent less time moving (Fig. 5C) during the 4-min homing test. Moreover, we observed a significant reduction in the distance moved inside the Nest (D) in CBD female pups compared to SHAM pups. On the contrary, no differences were found in the latency to reach, entrances to, and cumulative time spent in the nest, nor in the distance moved in the unfamiliar territory (Fig. 5E-H). Finally, CBD female pups spent more time in the unfamiliar zone, entering more than CBD male pups (Fig 5I-J). These homing test data therefore unveiled an additional sex-specific effect of gestational CBD.

Discussion

The consumption of CBD during pregnancy is increasing, but the developmental consequences are still largely unknown. Here, we investigated the sex-specific consequences of prenatal CBD exposure on pre-weaning behaviors. Fetal exposure to a low dose of CBD was associated with increased body weight in male pups during the perinatal period. In addition, the offspring of dams exposed to CBD during gestation showed sex-specific disturbances in their communication, motor skills, and discrimination abilities. Overall, these data indicate that gestational CBD is deleterious to early life behaviors in a sex-specific manner.

In human, gestational cannabis negatively impacts neonatal outcomes (24) and, at birth, the weight of infants exposed to cannabis *in utero* is lower (15,25). Animal models of prenatal THC or synthetic cannabinoid exposure confirm these observations (26). The present results extend these findings to another abundant phytocannabinoid, CBD. We found that maternal exposure to low CBD increases body weight, an effect observed only in males. This sex difference may be linked to differential levels of brain CBD in embryos exposed to CBD (11). In the sole other study that tested *in utero* CBD, no change in the body weight was found in the progeny after weaning or during adulthood (11). This discrepancy may be due to different dam strains (C57BL6/J vs CD1), the time of observation of pups' growth, or both.

USVs represent one of the earliest markers of neurobehavioral development, allowing quantification of affect, motivation, and social behavior (27–29). USV have an important communicative role in mother-offspring interactions, notably to elicit parents' attention and care. Thus, understanding mother-pup communication will ameliorate the comprehension and allow the early identification of neurodevelopmental diseases. Pre- and perinatal exposure to psychoactive cannabinoids (e.g., THC) impacts rats' USVs during infancy in a sex-specific way (23,30). The current results show that *in utero* CBD lowers the number of vocalizing males (not females). CBD-exposed males emitted shorter calls, while CBD-exposed females emitted calls at higher frequency compared to other groups. We found that prenatal CBD also reduced the complexity of the vocal repertoire. Thus, compared to SHAM pups, CBD-exposed pups of both sexes emitted fewer composite calls such as Complex Trill, Downward Ramp, and Inverted-U, when compared to their SHAM littermates. In addition, male CBD-exposed pups employed monosyllabic calls (e.g., "Short" calls) more often than other groups. Interestingly, we found that the same categories of calls found altered in CBD-treated animals reflected lower probabilities to use those calls, suggesting a call-specific effect of gestational CBD. The complexity of the vocal repertoire increases during life (31) and though the precise meaning of these vocalizations remains unclear, one could hypothesize that prenatal exposure to CBD changes early communication skills. Our observation is reminiscent of altered ultrasonic communication reported in several murine models of autism (i.e., *fmr1*^{Y/-}, BTBR, *Shank1*^{-/-}) (32–35) and is in line with human studies showing abnormal cry characteristics in sick toddlers with diseases affecting the central nervous system, including autism spectrum disorders (36). Thus, it is tempting to conclude that communication deficit is a common and early marker of neurodevelopmental diseases. Homing behavior requires sensory and cognitive skills (e.g., to differentiate the scent of the original cage) as well as motor skills (e.g., to navigate to the original litter). CBD had sex-specific effects on homing; only CBD-treated females showed an overall reduction in motor activities (i.e., distance traveled, speed, and total time spent moving). In addition, CBD-treated females entered the unfamiliar area more often and spent more time in the unfamiliar area than CBD-treated male pups, suggesting differential development of sensory and cognitive abilities.

Taken together, this study reveals sex-specific cognitive impairments in early life associated with fetal CBD. This work challenges the view that CBD is a universally safe compound and warrants further study of the developmental consequences of prenatal CBD.

Author contributions:

Daniela Iezzi: Conceptualization, Data curation, Formal analysis, Validation, Writing— original draft, review and editing.

Alba Caceres: Data curation, Formal analysis.

Pascale Chavis: Conceptualization, Supervision, Methodology, Writing—, review and editing.

Olivier JJ Manzoni: Conceptualization, Supervision, Funding acquisition, Methodology, Project administration, Writing— original draft, review, and editing.

The authors declare no conflict of interest.

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References

1. Bloomfield MAP, Hindocha C, Green SF, Wall MB, Lees R, Petrilli K, et al. The neuropsychopharmacology of cannabis: A review of human imaging studies. *Pharmacol Ther.* 2019 Mar 1;195:132–61.
2. Grof CPL. Cannabis, from plant to pill. *British Journal of Clinical Pharmacology.* 2018 Nov 1;84(11):2463–7.
3. Machado Bergamaschi M, Helena Costa Queiroz R, Waldo Zuardi A, Alexandre S. Crippa J. Safety and Side Effects of Cannabidiol, a Cannabis sativa Constituent. *Current Drug Safety.* 2011 Nov 28;6(4):237–49.
4. Calpe-López C, García-Pardo MP, Aguilar MA. Cannabidiol Treatment Might Promote Resilience to Cocaine and Methamphetamine Use Disorders: A Review of Possible Mechanisms. *Molecules.* 2019 Jul 16;24(14):2583.
5. Razavi Y, Shabani R, Mehdizadeh M, Haghparast A. Neuroprotective effect of chronic administration of cannabidiol during the abstinence period on methamphetamine-induced impairment of recognition memory in the rats. *Behavioural Pharmacology.* 2020;31(4):385–96.
6. Karimi-Haghighi S, Razavi Y, Iezzi D, Scheyer AF, Manzoni O, Haghparast A. Cannabidiol and substance use disorder: Dream or reality. *Neuropharmacology.* 2022;207(January):108948.
7. Abuhasira R, Shbiro L, Landschaft Y. Medical use of cannabis and cannabinoids containing products - Regulations in Europe and North America. *Eur J Intern Med.* 2018 Mar 1;49:2–6.
8. Alves P, Amaral C, Teixeira N, Correia-da-Silva G. Cannabidiol disrupts apoptosis, autophagy and invasion processes of placental trophoblasts. *Archives of Toxicology.* 2021;95(10):3393–406.
9. Sarrafpour S, Urits I, Powell J, Nguyen D, Callan J, Orhurhu V, et al. Considerations and Implications of Cannabidiol Use During Pregnancy. *Current Pain and Headache Reports.* 2020;24(7).
10. Ochiai W, Kitaoka S, Kawamura T, Hatogai J, Harada S, Iizuka M, et al. Maternal and fetal pharmacokinetic analysis of cannabidiol during pregnancy in mice. *Drug Metabolism and Disposition.* 2021;49(4):337–43.
11. Maciel I de S, de Abreu GHD, Johnson CT, Bonday R, Bradshaw HB, Mackie K, et al. Perinatal CBD or THC Exposure Results in Lasting Resistance to Fluoxetine in the Forced Swim Test: Reversal by Fatty Acid Amide Hydrolase Inhibition. *Cannabis and Cannabinoid Research.* 2021;X(X):1–10.
12. Scheyer AF, Borsoi M, Pelissier-Alicot AL, Manzoni OJJ. Maternal exposure to the cannabinoid agonist WIN 55,12,2 during lactation induces lasting behavioral and synaptic alterations in the rat adult offspring of both sexes. *eNeuro.* 2020;7(5):1–11.
13. Coffey KR, Marx RG, Neumaier JF. DeepSqueak: a deep learning-based system for detection and analysis of ultrasonic vocalizations. *Neuropsychopharmacology* [Internet]. 2019 [cited 2021 Sep 23]; Available from: <https://doi.org/10.1038/s41386-018-0303-6>
14. Servadio M, Melancia F, Manduca A, Di Masi A, Schiavi S, Cartocci V, et al. Targeting anandamide metabolism rescues core and associated autistic-like symptoms in rats prenatally exposed to valproic acid. *Translational Psychiatry.* 2016;6(9):1–11.
15. Paul SE, Hatoum AS, Fine JD, Johnson EC, Hansen I, Karcher NR, et al. Associations Between Prenatal Cannabis Exposure and Childhood Outcomes: Results From the ABCD Study. *JAMA Psychiatry* [Internet]. 2021 Jan 1 [cited 2022 Apr 28];78(1):64–76. Available from: <https://pubmed.ncbi.nlm.nih.gov/32965490/>
16. Simola N, Granon S. Ultrasonic vocalizations as a tool in studying emotional states in rodent models of social behavior and brain disease. *Neuropharmacology* [Internet]. 2019 Nov 15 [cited 2022 Apr 27];159. Available from: <https://pubmed.ncbi.nlm.nih.gov/30445100/>
17. Brudzynski SM. Biological Functions of Rat Ultrasonic Vocalizations, Arousal Mechanisms, and Call Initiation. *Brain Sciences* [Internet]. 2021 May 1 [cited 2022 Jun 29];11(5). Available from: <https://pmc/articles/PMC8150717/>

18. Bara A, Manduca A, Bernabeu A, Borsoi M, Serviado M, Lassalle O, et al. Sex-dependent effects of in utero cannabinoid exposure on cortical function. *Elife*. 2018;7:1–31.
19. Harkany T, Guzmán M, Galve-Roperh I, Berghuis P, Devi LA, Mackie K. The emerging functions of endocannabinoid signaling during CNS development. *Trends Pharmacol Sci* [Internet]. 2007 Feb [cited 2022 Jun 22];28(2):83–92. Available from: <https://pubmed.ncbi.nlm.nih.gov/17222464/>
20. Scheyer AF, Melis M, Trezza V, Manzoni OJJ. Consequences of Perinatal Cannabis Exposure. *Trends Neurosci* [Internet]. 2019 Dec 1 [cited 2022 Jun 22];42(12):871–84. Available from: <https://pubmed.ncbi.nlm.nih.gov/31604585/>
21. Hurd YL, Manzoni OJ, Pletnikov M v., Lee FS, Bhattacharyya S, Melis M. Cannabis and the Developing Brain: Insights into Its Long-Lasting Effects. *J Neurosci* [Internet]. 2019 Oct 16 [cited 2022 Jun 22];39(42):8250–8. Available from: <https://pubmed.ncbi.nlm.nih.gov/31619494/>
22. Trezza V, Campolongo P, Cassano T, Macheda T, Dipasquale P, Carratù MR, et al. Effects of perinatal exposure to delta-9-tetrahydrocannabinol on the emotional reactivity of the offspring: a longitudinal behavioral study in Wistar rats. *Psychopharmacology (Berl)* [Internet]. 2008 Jul [cited 2022 Jun 27];198(4):529–37. Available from: <https://pubmed.ncbi.nlm.nih.gov/18452035/>
23. Manduca A, Servadio M, Melancia F, Schiavi S, Manzoni OJ, Trezza V. Sex-specific behavioural deficits induced at early life by prenatal exposure to the cannabinoid receptor agonist WIN55, 212-2 depend on mGlu5 receptor signalling. *British Journal of Pharmacology*. 2020;177(2):449–63.
24. Fergusson DM, Horwood LJ, Northstone K. Maternal use of cannabis and pregnancy outcome. *BJOG* [Internet]. 2002 Jan [cited 2022 Jun 27];109(1):21–7. Available from: <https://pubmed.ncbi.nlm.nih.gov/11843371/>
25. Gunn JKL, Rosales CB, Center KE, Nuñez A, Gibson SJ, Christ C, et al. Prenatal exposure to cannabis and maternal and child health outcomes: a systematic review and meta-analysis. *BMJ Open* [Internet]. 2016 [cited 2022 May 23];6(4). Available from: <https://pubmed.ncbi.nlm.nih.gov/27048634/>
26. Scheyer AF, Borsoi M, Wager-Miller J, Pelissier-Alicot AL, Murphy MN, Mackie K, et al. Cannabinoid Exposure via Lactation in Rats Disrupts Perinatal Programming of the Gamma-Aminobutyric Acid Trajectory and Select Early-Life Behaviors. *Biological Psychiatry* [Internet]. 2020;87(7):666–77. Available from: <https://doi.org/10.1016/j.biopsych.2019.08.023>
27. Branchi I, Santucci D, Alleva E. Ultrasonic vocalisation emitted by infant rodents: a tool for assessment of neurobehavioural development. *Behavioural brain research* [Internet]. 2001 Nov 8 [cited 2022 Jun 27];125(1–2):49–56. Available from: <https://pubmed.ncbi.nlm.nih.gov/11682093/>
28. Branchi I, Santucci D, Alleva E. Analysis of Ultrasonic Vocalizations Emitted by Infant Rodents. *Current Protocols in Toxicology* [Internet]. 2006 Nov 1 [cited 2022 Jun 27];30(1):13.12.1–13.12.14. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1002/0471140856.tx1312s30>
29. Wöhr M, Schwarting RKW. Affective communication in rodents: ultrasonic vocalizations as a tool for research on emotion and motivation. *Cell Tissue Res* [Internet]. 2013 Oct [cited 2022 May 2];354(1):81–97. Available from: <https://pubmed.ncbi.nlm.nih.gov/23576070/>
30. Antonelli T, Tomasini MC, Tattoli M, Cassano T, Tanganelli S, Finetti S, et al. Prenatal exposure to the CB1 receptor agonist WIN 55,212-2 causes learning disruption associated with impaired cortical NMDA receptor function and emotional reactivity changes in rat offspring. *Cereb Cortex* [Internet]. 2005 Dec [cited 2022 Jun 27];15(12):2013–20. Available from: <https://pubmed.ncbi.nlm.nih.gov/15788701/>
31. Hepbasli D, Gredy S, Ullrich M, Reigl A, Abeßer M, Raabe T, et al. Genotype- and Age-Dependent Differences in Ultrasound Vocalizations of SPRED2 Mutant Mice Revealed by Machine Deep Learning. *Brain Sci* [Internet]. 2021 Oct 1 [cited 2022 Jun 27];11(10). Available from: <https://pubmed.ncbi.nlm.nih.gov/34679429/>

32. Scattoni ML, Gandhi SU, Ricceri L, Crawley JN. Unusual repertoire of vocalizations in the BTBR T+tf/J mouse model of autism. *PLoS One* [Internet]. 2008 Aug 27 [cited 2022 Jun 27];3(8). Available from: <https://pubmed.ncbi.nlm.nih.gov/18728777/>
33. Roy S, Watkins N, Heck D. Comprehensive analysis of ultrasonic vocalizations in a mouse model of fragile X syndrome reveals limited, call type specific deficits. *PLoS One* [Internet]. 2012 Sep 11 [cited 2022 Jun 27];7(9). Available from: <https://pubmed.ncbi.nlm.nih.gov/22984567/>
34. Sungur AÖ, Schwarting RKW, Wöhr M. Early communication deficits in the Shank1 knockout mouse model for autism spectrum disorder: Developmental aspects and effects of social context. *Autism Res* [Internet]. 2016 Jun 1 [cited 2022 Jun 27];9(6):696–709. Available from: <https://pubmed.ncbi.nlm.nih.gov/26419918/>
35. Wöhr M, Rouillet FI, Hung AY, Sheng M, Crawley JN. Communication impairments in mice lacking Shank1: reduced levels of ultrasonic vocalizations and scent marking behavior. *PLoS One* [Internet]. 2011 [cited 2022 Jun 27];6(6). Available from: <https://pubmed.ncbi.nlm.nih.gov/21695253/>
36. Michelsson K, Michelsson O. Phonation in the newborn, infant cry. *Int J Pediatr Otorhinolaryngol* [Internet]. 1999 Oct 5 [cited 2022 Jun 27];49 Suppl 1(SUPPL. 1):S297–301. Available from: <https://pubmed.ncbi.nlm.nih.gov/10577825/>

Figure Legends

Figure 1. Prenatal exposure to CBD specifically increases the body weight of male offspring. (A-B) Starting at PND 10, SHAM and CBD pups of both sexes were weighed every 3 days until one day after weaning (i.e., 22). Fetal CBD exposure was associated to increase of bodyweight in male (A) but not female progeny (B), (SHAM MALE N = 18 light green, CBD MALE N = 14 dark green, SHAM FEMALE N = 14 light orange, CBD FEMALE N = 12 dark orange). Data are represented as Box and whisker plots (first quartile, median, third quartile). Individual data point represents a single animal. Multiple Mann-Whitney *U* test.

Figure 2. Fetal CBD modifies the frequency and the duration of perinatal USVs. (A-B) Pie graphs depicting the percentages of vocalizing (C: call) and non-vocalizing (NC: no call) mice, in SHAM and CBD male (left) or female (right) pups. Percentages were calculated as number of animals vocalizing or not / total number of tested animals. Quantitative analysis of USVs shows that following CBD prenatal exposure, the total number of USVs (C) and the latency to the first vocalization (D) were similar to those of sham, in both sexes. In contrast, mean duration (E) and mean frequency (F) of emitted calls were modified in a sex-specific manner in the CBD progeny. The mean call duration was shorter in CBD male, while the mean frequency was higher in CBD females, compared to their respective SHAM group, (SHAM MALE N = 14 light green, CBD MALE N = 10 dark green, SHAM FEMALE N = 11 light orange, CBD FEMALE N = 11 dark orange). Data are represented as Box and whisker plots (first quartile, median, third quartile). Individual data points represent a single animal. Multiple Mann-Whitney *U* test, **p*<0.05. (G) The average distribution of call frequency was monomodal (i.e., fitted with a single Gaussian function, dark green) in SHAM male, but bimodal (i.e., fitted by the sum of two Gaussians for CBD male, light green). (H) In contrast, the call frequency was monomodal in SHAM (light orange) and CBD exposed female pups (dark orange). Data are represented as curve fit (\pm CI) of the principal frequency's average distribution.

Figure 3. Call type profile is altered in a sex-specific manner in CBD-exposed pups. (A) Representative USV calls classified into ten distinct categories based on Supervised-call classification neural network (DeepSqueak). Color maps of USVs showing differential distributions of call types emitted by SHAM (B) vs CBD (C) males and CBD-exposed females (F) vs SHAM (E) pups. Each call category was expressed as (number of calls in each category for each subject / total number of calls analyzed in each subject) and represented as the average of each group (SHAM MALE N = 14, CBD MALE N = 7, SHAM FEMALE N = 11, CBD FEMALE N = 11). (D-G) CBD modified the vocal repertoire. (D) CBD-exposed male emitted more often Short, Trill and Step-up calls, and less Complex Trill and Downward Ramp calls than their SHAM counterparts (G). CBD females emitted significantly less Flat, Complex Trill, Downward Ramp, and Inverted-U vocalizations than SHAM (G), (SHAM MALE N = 14 light green, CBD MALE N = 7 dark green, SHAM FEMALE N = 11 light orange, CBD FEMALE N = 11 dark orange). Data are represented as Box and whisker plots (first quartile, median, third quartile). Individual data points represent a single animal. Multiple Mann-Whitney *U* test, **p*<0.05.

Figure 4. Transitional probabilities for call type transitions within USV bouts in SHAM and CBD pups. Heat maps of transition probabilities in SHAM (A) and CBD male (B) as well as in SHAM (D) and CBD female (E) pups. The values in the individual boxes indicate the probability of one call to follow the previous (A-B-D-E). The transitional probability was expressed as the mean probability of each transition for each subject. (C) The comparison of transitional probabilities

shows that CBD-exposed male transitioned significantly less to Downward Ramp and Complex Trill calls than SHAM male pups and that CBD male transitioned more to Trill than SHAM. (F) CBD females showed a lower probability to transition to Complex, Downward Ramp and Short calls. Data are represented as Box and whisker plots (first quartile, median, third quartile). Individual data points represent the transitional probabilities of calls emitted by each group. SHAM MALE N = 14, CBD MALE N = 7, SHAM FEMALE N = 11, CBD FEMALE N = 11. Multiple Mann-Whitney *U* test, **p*<0.05.

Figure 5. Fetal CBD modifies homing behavior selectively in female pups. (A-D) In the female progeny exposed to CBD *in utero*, the total distance moved, the distance moved in the nest, the velocity, and the total time spent moving were diminished. In contrast, these parameters were normal in the male progeny. **(E-H)** Fetal CBD had no discernable effects on the latency to reach the nest, cumulative time spent in the latter, the distance moved in the unfamiliar area or entries into the Nest **(I-J)** CBD females entered more often and spent more time in the unfamiliar zone than the CBD male pups (SHAM MALE N = 15 light green boxplot, CBD MALE N = 13 dark green boxplot, SHAM FEMALE N = 15 light orange boxplot, CBD FEMALE N = 13 dark orange boxplot). Data are represented as Box and whisker plots. Individual data points represent one animal while the line the median. Multiple Mann-Whitney *U* test, **p*<0.05.

Tables

Postnatal day weights (g)	TREATMENT	Median	Max	Min	N	<i>p</i> value Multiple Mann-Whitney Unpaired t test
PND 10	SHAM MALE	3.4	6.7	3.1	18	0.003
	CBD MALE	5.5	5.8	4	14	
	SHAM FEMALE	3.5	5.9	3.1	14	0.06
	CBD FEMALE	5.1	5.8	3.9	12	
PND 13	SHAM MALE	4.9	8	3.6	18	0.003
	CBD MALE	6.6	7.4	5.7	14	
	SHAM FEMALE	5	8.6	3.5	14	0.08
	CBD FEMALE	6.4	7.3	5.6	12	
PND 16	SHAM MALE	6	9.4	4.2	18	0.04
	CBD MALE	7.2	8.2	6.3	14	
	SHAM FEMALE	6	9.6	3.9	14	0.27
	CBD FEMALE	7	8.2	6.1	12	
PND 19	SHAM MALE	6.9	9.7	5	18	0.01
	CBD MALE	8.45	9.9	6.6	14	
	SHAM FEMALE	7.3	10	4.2	14	0.09
	CBD FEMALE	8.2	9.8	6.2	12	
PND 22	SHAM MALE	8.2	11.8	5.7	18	0.002
	CBD MALE	10.5	11.8	7.5	14	
	SHAM FEMALE	7.9	11	5.6	14	0.20
	CBD FEMALE	8.6	10.6	6.8	12	

Table 1: Cannabidiol prenatal exposure increases pup weights in a sex-specific way.

Male and female pup weights were collected every three days from postnatal day 10 to postnatal day 22. Values are expressed as median, maximum and minimum. The *p* values are given for each day as compared with pups from sham-treated dams on the same postnatal day, as determined by Multiple Mann-Whitney *U* test.

Parameter	PND	TREATMENT	Median	Max	Min	N	<i>p</i> value Multiple Mann-Whitney Unpaired t test
Number of USVs	10	SHAM MALE	67	258	3	14	0.1052
		CBD MALE	31	96	1	10	
		SHAM FEMALE	77	320	4	11	0.0785
		CBD FEMALE	18	130	2	11	
Latency (sec)	10	SHAM MALE	4.42	163.38	0.01	14	0.0821
		CBD MALE	32.56	256.39	1.34	10	
		SHAM FEMALE	20.29	56.35	0.32	11	0.26
		CBD FEMALE	38.64	212.51	0.07	11	
Mean USVs duration (sec)	10	SHAM MALE	0.05	0.09	0.02	14	0.03
		CBD MALE	0.03	0.06	0.02	10	
		SHAM FEMALE	0.05	0.08	0.02	11	0.6396
		CBD FEMALE	0.04	0.08	0.01	11	
Mean Frequency (KHz)	10	SHAM MALE	56.05	65.00	51.81	14	>0.9999
		CBD MALE	56.15	67.11	47.08	10	
		SHAM FEMALE	56.47	63.46	53.39	11	0.004
		CBD FEMALE	59.76	64.12	56.21	11	

Table 2: Fetal CBD modifies the frequency and the duration of perinatal USVs.

Data were collected from litters for each condition as described in Methods and Materials. Values are expressed as median, maximum and minimum. Significant difference was observed in the Mean USVs duration in CBD compared to SHAM male pups and in the Mean frequency duration in CBD compared to SHAM female pups as determined by Multiple Mann-Whitney *U* test.

Call Type	PND	TREATMENT	Median	Max	Min	N	p value Multiple Mann-Whitney Unpaired t test
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Flat	10	SHAM MALE	6.53	16.67	0.00	14	0.85
		CBD MALE	7.02	14.55	0.00	10	
		SHAM FEMALE	8.51	11.11	0.00	11	0.003
		CBD FEMALE	0.00	18.03	0.00	11	
Short	10	SHAM MALE	2.88	12.50	0.00	14	<0.0001
		CBD MALE	17.54	33.33	13.54	10	
		SHAM FEMALE	2.13	5.45	0.00	11	0.13
		CBD FEMALE	7.69	80.00	0.00	11	
Trill	10	SHAM MALE	1.29	16.47	0.00	14	0.0002
		CBD MALE	23.40	35.09	0.00	10	
		SHAM FEMALE	3.19	10.66	0.00	11	0.08
		CBD FEMALE	0.21	34.23	0.00	11	
Complex	10	SHAM MALE	13.34	25.00	0.00	14	0.89
		CBD MALE	9.09	33.33	5.26	10	
		SHAM FEMALE	16.36	25.53	0.00	11	0.33
		CBD FEMALE	11.29	38.46	0.00	11	
Complex Trill	10	SHAM MALE	12.88	41.67	5.88	14	0.002
		CBD MALE	2.22	5.45	0.00	10	
		SHAM FEMALE	9.72	75.00	7.27	11	0.0003
		CBD FEMALE	0.00	44.44	0.00	11	
Downward Ramp	10	SHAM MALE	32.72	42.24	0.00	14	<0.0001
		CBD MALE	4.44	17.54	0.00	10	
		SHAM FEMALE	28.57	51.06	0.00	11	0.0002
		CBD FEMALE	0.00	11.48	0.00	11	
Upward Ramp	10	SHAM MALE	3.60	11.11	0.00	14	0.07
		CBD MALE	6.97	14.58	0.00	10	
		SHAM FEMALE	3.19	11.43	0.00	11	0.75
		CBD FEMALE	0.18	27.34	0.00	11	
Step Up	10	SHAM MALE	0.00	11.11	0.00	14	0.004
		CBD MALE	7.29	42.86	0.00	10	
		SHAM FEMALE	1.06	25.00	0.00	11	0.19
		CBD FEMALE	3.60	55.56	0.00	11	
Step Down	10	SHAM MALE	2.35	16.67	0.00	14	0.58
		CBD MALE	3.51	4.44	0.00	10	
		SHAM FEMALE	2.13	3.49	0.00	11	0.07
		CBD FEMALE	0.00	3.85	0.00	11	
Inverted-U	10	SHAM MALE	13.13	33.33	0.00	14	0.003
		CBD MALE	0.00	4.26	0.00	7	
		SHAM FEMALE	17.14	23.64	0.00	11	0.0002
		CBD FEMALE	0.00	3.85	0.00	11	

Table 3: Call type profile is altered in a sex-specific manner in CBD-exposed pups.

Data were collected from litters for each condition as described in Methods and Materials. Values are expressed as median, maximum and minimum.

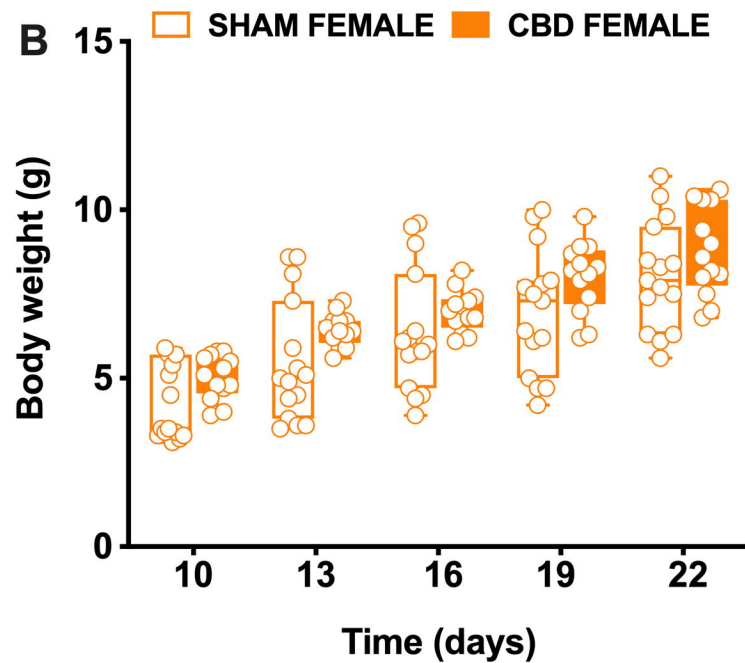
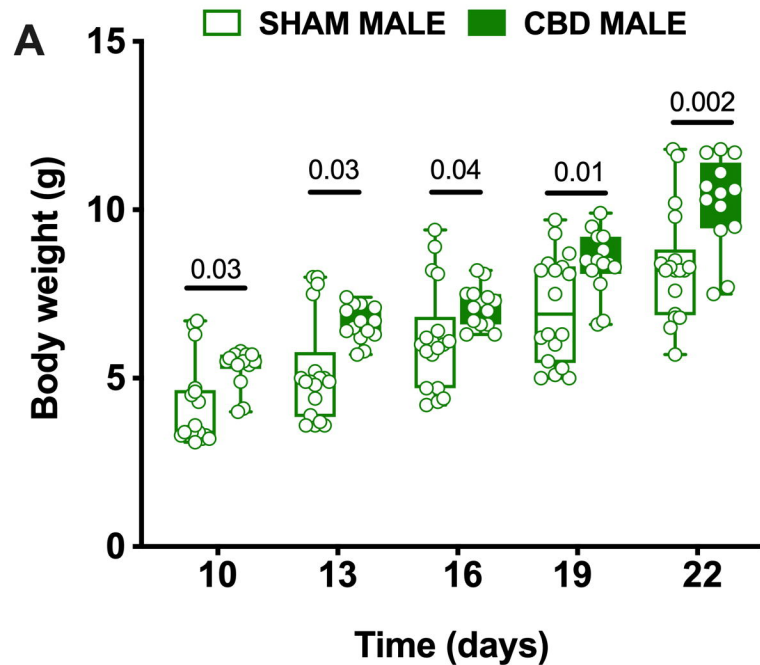
Trantitional	PND	TREATMENT	Median	Max	Min	N	p value
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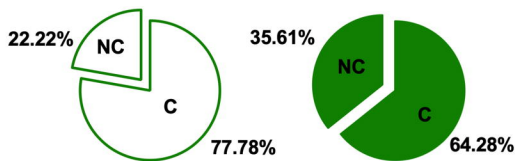
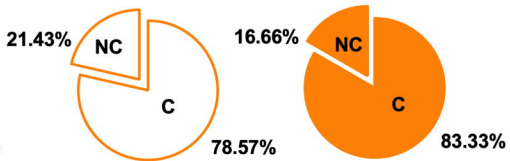
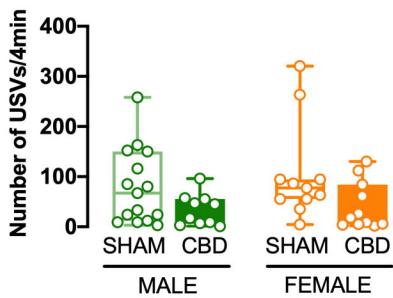
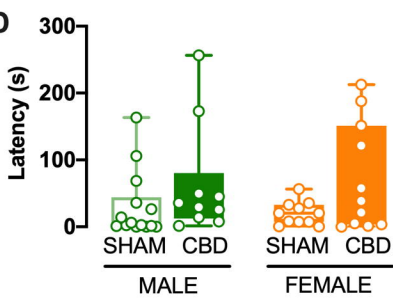
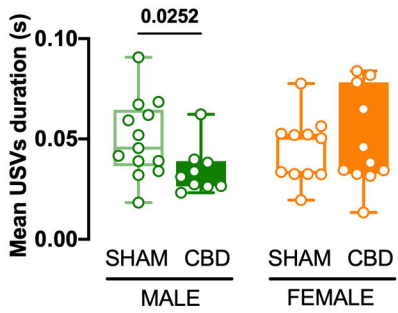
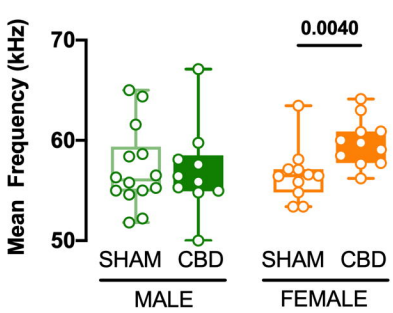
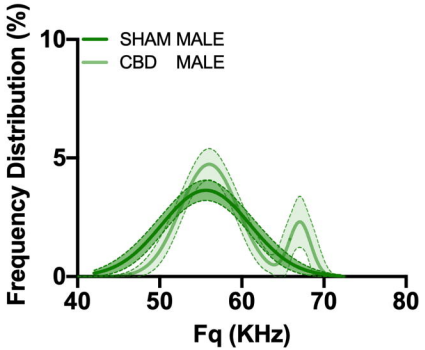
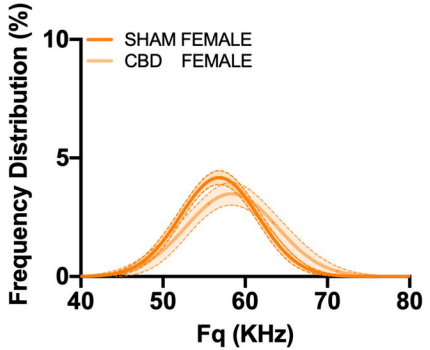
Parameter	PND	TREATMENT	Median	Max	Min	N	p value
Probability							Multiple Mann-Whitney Unpaired t test
Flat	10	SHAM MALE	0.03	0.09	0.00	14	0.27
		CBD MALE	0.00	0.09	0.00	7	
		SHAM FEMALE	0.03	0.13	0.00	11	0.22
		CBD FEMALE	0.01	0.07	0.00	11	
Complex	10	SHAM MALE	0.02	0.04	0.00	14	0.45
		CBD MALE	0.01	0.02	0.00	7	
		SHAM FEMALE	0.13	0.25	0.04	11	0.01
		CBD FEMALE	0.05	0.12	0.00	11	
Downward Ramp	10	SHAM MALE	0.03	0.15	0.00	14	0.01
		CBD MALE	0.10	0.21	0.00	7	
		SHAM FEMALE	0.16	0.26	0.10	11	0.01
		CBD FEMALE	0.09	0.21	0.00	11	
Complex Trill	10	SHAM MALE	0.08	0.11	0.00	14	0.01
		CBD MALE	0.09	0.20	0.00	7	
		SHAM FEMALE	0.06	0.16	0.01	11	0.69
		CBD FEMALE	0.06	0.20	0.00	11	
Step Up	10	SHAM MALE	0.09	0.18	0.04	14	0.96
		CBD MALE	0.05	0.10	0.02	7	
		SHAM FEMALE	0.00	0.03	0.00	11	0.46
		CBD FEMALE	0.00	0.04	0.00	11	
Upward Ramp	10	SHAM MALE	0.14	0.29	0.09	14	0.70
		CBD MALE	0.08	0.16	0.03	7	
		SHAM FEMALE	0.02	0.10	0.00	11	0.45
		CBD FEMALE	0.01	0.07	0.00	11	
Step Down	10	SHAM MALE	0.02	0.09	0.00	14	0.70
		CBD MALE	0.03	0.07	0.00	7	
		SHAM FEMALE	0.00	0.02	0.00	11	0.78
		CBD FEMALE	0.00	0.02	0.00	11	
Inverted-U	10	SHAM MALE	0.00	0.07	0.00	14	0.99
		CBD MALE	0.00	0.09	0.00	7	
		SHAM FEMALE	0.07	0.18	0.00	11	0.25
		CBD FEMALE	0.05	0.10	0.00	11	
Trill	10	SHAM MALE	0.00	0.02	0.00	14	0.01
		CBD MALE	0.00	0.05	0.00	7	
		SHAM FEMALE	0.02	0.06	0.00	11	0.18
		CBD FEMALE	0.04	0.20	0.00	11	
Short	10	SHAM MALE	0.09	0.20	0.00	14	0.19
		CBD MALE	0.08	0.25	0.00	7	
		SHAM FEMALE	0.01	0.05	0.00	11	0.02
		CBD FEMALE	0.00	0.06	0.00	11	

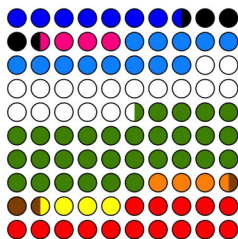
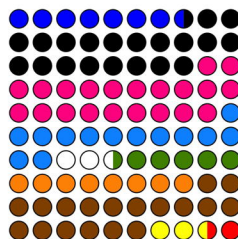
Table 4: Transitional probabilities for call type transitions within USV bouts in SHAM and CBD pups. Data were collected from litters for each condition as described in Methods and Materials. Values are expressed as median, maximum and minimum.

							Multiple Mann-Whitney Unpaired t test
Distance moved (cm)	13	SHAM MALE	430.00	1114.00	76.29	15	0.06
		CBD MALE	268.70	897.50	61.09	13	
		SHAM FEMALE	489.40	1111.00	300.70	15	0.003
		CBD FEMALE	261.80	839.80	105.40	13	
Velocity (cm/s)	13	SHAM MALE	1.79	4.64	0.32	15	0.13
		CBD MALE	1.22	3.83	0.25	13	
		SHAM FEMALE	2.04	4.63	1.25	15	0.004
		CBD FEMALE	1.12	3.60	0.44	13	
Moving (sec)	13	SHAM MALE	60.60	149.50	1.92	15	0.12
		CBD MALE	43.44	116.50	1.70	13	
		SHAM FEMALE	85.60	156.00	42.76	15	0.002
		CBD FEMALE	44.68	112.40	7.54	13	
Distance moved Nest (cm)	13	SHAM MALE	226.10	353.90	27.91	15	0.52
		CBD MALE	170.80	432.70	89.30	13	
		SHAM FEMALE	244.00	747.90	109.90	15	0.009
		CBD FEMALE	98.74	428.60	5.04	13	
Latency (sec)	13	SHAM MALE	12.20	49.31	0.00	15	<0.9999
		CBD MALE	7.24	87.95	0.93	13	
		SHAM FEMALE	15.16	65.80	1.50	15	<0.9999
		CBD FEMALE	16.78	69.10	0.00	13	
Entries in Nest	13	SHAM MALE	5	15	0	15	0.54
		CBD MALE	3.5	12	0	13	
		SHAM FEMALE	5	13	1	15	0.08
		CBD FEMALE	8	14	0	13	
Nest Time (s)	13	SHAM MALE	186.30	219.70	0.00	15	0.78
		CBD MALE	168.70	236.60	0.00	13	
		SHAM FEMALE	180.00	224.40	66.52	15	0.09
		CBD FEMALE	139.40	200.10	0.00	13	
Distance moved Un. Area (cm)	13	SHAM MALE	115.3	835.2	48.37	15	0.43
		CBD MALE	133.2	464.8	10.49	13	
		SHAM FEMALE	161.6	842.3	70.79	15	0.79
		CBD FEMALE	209.2	411.1	6.695	13	
Entrie Un. Area	13	SHAM MALE	3	11	1	15	0.77
		CBD MALE	3	13	1	13	
		SHAM FEMALE	4	17	1	15	0.15
		CBD FEMALE	7	15	1	13	
Unfamiliar Area (s)	13	SHAM MALE	43.28	221.2	13.21	15	0.21
		CBD MALE	43.54	125.7	1.501	13	
		SHAM FEMALE	52.58	218.4	11.68	15	0.18
		CBD FEMALE	100	240	38.5	13	

Table 5: Fetal CBD modifies homing behavior selectively in female pups. Data were collected from litters for each condition as described in Methods and Materials. Values are expressed as median, maximum and minimum.

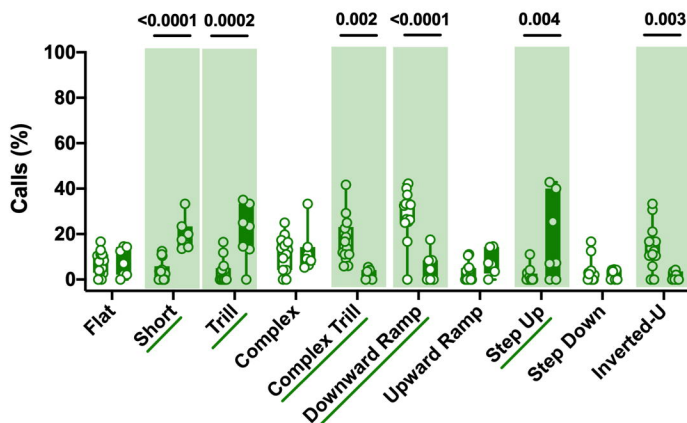
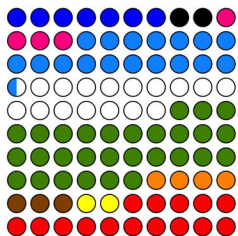
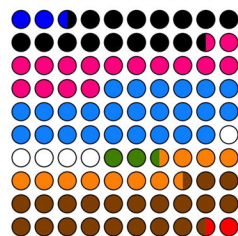


A**B****C****D****E****F****G****H**

B**SHAM MALE****C****CBD MALE****D**

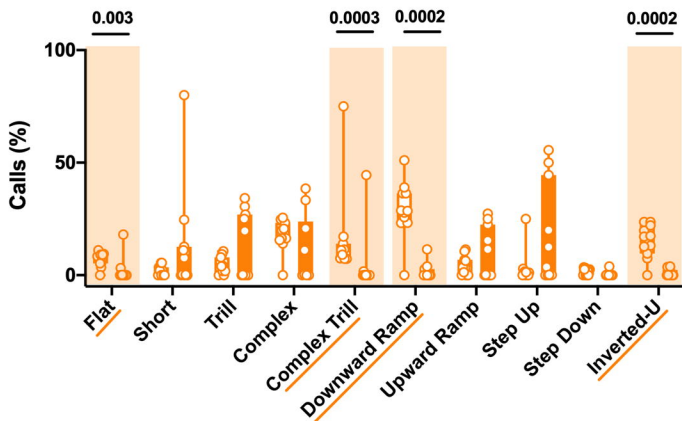
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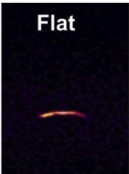
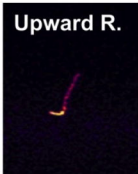
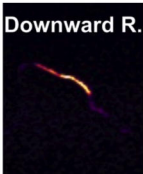
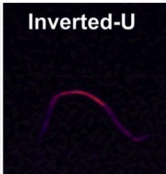
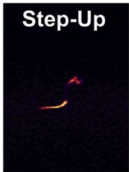
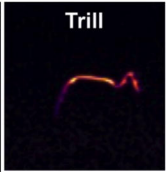
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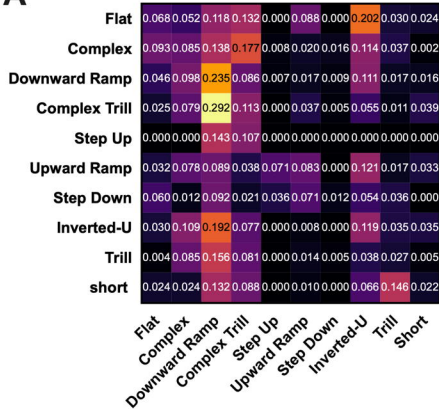
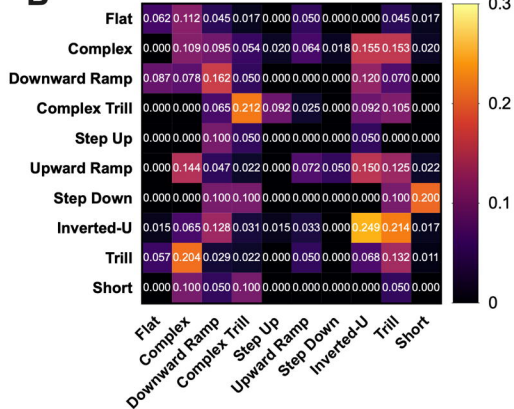
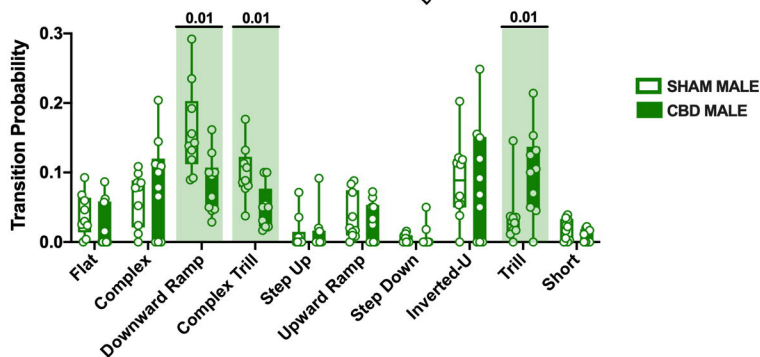
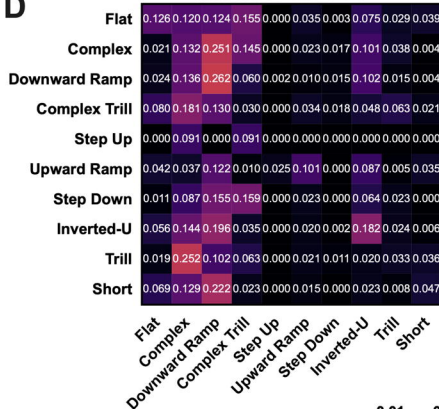
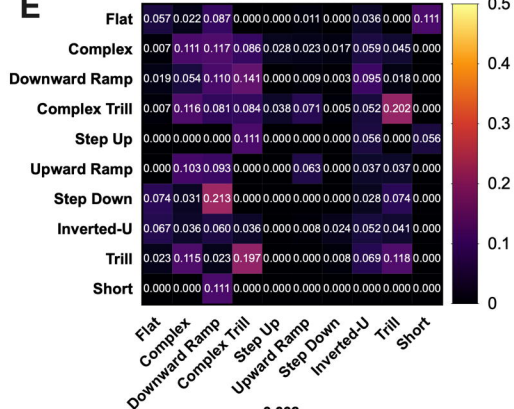
**E****SHAM FEMALE****F****CBD FEMALE****G**

SHAM FEMALE

CBD FEMALE



A**Short****Flat****Upward R.****Downward R.****Inverted-U****Step-Down****Step-Up****Complex****Complex Trill****Trill**

A**SHAM MALE****B****CBD MALE****C****D****SHAM FEMALE****E****CBD FEMALE****F**