1	Assessing risk for butterflies in the context of climate change,
2	demographic uncertainty, and heterogenous data sources
3	
4	Matthew L. Forister ¹ , Eliza M. Grames ¹ , Christopher A. Halsch ¹ , Kevin J. Burls ² ,
5	Cas F. Carroll ¹ , Katherine L. Bell ¹ , Joshua P. Jahner ³ , Taylor Bradford ¹ , Jing Zhang ⁴ ,
6	Qian Cong ⁵ , Nick V. Grishin ^{4,6} , Jeffrey Glassberg ^{7,8} , Arthur M. Shapiro ⁹ , Thomas V. Riecke ¹⁰
7	
8	¹ Ecology, Evolution and Conservation Biology, Department of Biology, University of Nevada,
9	Reno, Nevada, USA
10	² Xerces Society for Invertebrate Conservation, Portland, Oregon, USA
11	³ Department of Botany, University of Wyoming, Laramie, Wyoming, USA
12	⁴ Howard Hughes Medical Institute, University of Texas Southwest Medical Center, Dallas,
13	Texas, USA
14	⁵ Institute for Protein Design and Department of Biochemistry, University of Washington,
15	Seattle, Washington, USA
16	⁶ Departments of Biophysics and Biochemistry, University of Texas Southwest Medical Center,
17	Dallas, Texas, USA
18	⁷ North American Butterfly Association, Morristown, New Jersey, USA
19	⁸ Department of BioSciences, Rice University, Houston, Texas, USA
20	⁹ Center for Population Biology, University of California, Davis, California, USA
21	¹⁰ Swiss Ornithological Institute, CH-6204 Sempach, Switzerland
22	Correspondence: Matthew L. Forister, mforister@unr.edu
23	Open Research Statement: all data will be archived on Dryad upon acceptance for publication.

Forister et al. p. 2

24 Abstract

Ongoing declines in insect populations have led to substantial concern and calls for conservation 25 action. However, even for relatively well-studied groups, like butterflies, information relevant to 26 species-specific status and risk is scattered across field guides, the scientific literature, and 27 agency reports. Consequently, attention and resources have been spent on a miniscule fraction of 28 insect diversity, including a few well-studied butterflies. Here we bring together heterogenous 29 sources of information for 396 butterfly species and 1,004 subspecies to provide the first regional 30 assessment of butterflies for the 11 western US states. For 184 species, we use monitoring and 31 other observational data to characterize historical and projected trends in population abundance; 32 for another 212 species (for which sufficient observational data are not available), we use 33 exposure to climate change, development, geographic range, host breadth and other factors to 34 rank species for conservation concern. We also organize information relevant to subspecific risk 35 and prioritize a top 50 subspecies for further attention. A phylogenetic signal is apparent, with 36 concentrations of declining and at-risk species in the families Lycaenidae and Hesperiidae. A 37 geographic bias exists in that many species that lack monitoring data occur in more southern 38 states where we expect that impacts of warming and drying trends will be most severe. Legal 39 protection is relatively uncommon among the taxa with the highest risk values: of the top 100 40 species, one is listed as threatened under the US Endangered Species Act and one is a candidate 41 for listing; of the top 50 subspecies, 15 have federal legal protection and one is under review for 42 protected status. Among the many taxa not currently protected, we highlight a short list of 43 species in decline, including Vanessa annabella, Thorybes mexicanus, Euchloe ausonides, and 44 *Pholisora catullus*. Notably, many of these species have broad geographic ranges, which perhaps 45

Forister et al. p. 3

highlights a new era of insect conservation in which small or fragmented ranges will not be the
only red flags that attract conservation attention.

48 **KEYWORDS**

⁴⁹ Anthropocene, butterfly, climate change, Lepidoptera, population viability analysis,

- ⁵⁰ demographic uncertainty, extinction, heterogeneous data, hierarchical Bayesian model
- 51
- 52

53 INTRODUCTION

Reductions in abundance, contractions in geographic range, extirpation, and extinction have 54 become common features of wild plant and animal populations impacted by the various stressors 55 of the Anthropocene (Dirzo et al. 2014, Turvey and Crees 2019). Effects on individual 56 populations translate into depauperate assemblages of species in remaining natural lands, even 57 those far removed from the most immediate effects of habitat destruction and degradation 58 (McLaughlin et al. 2002, Brook et al. 2008). To the extent that the loss of evolutionary lineages 59 (populations, species and higher taxonomic groups) is a part of life on earth and always has been, 60 the current mass extinction crisis affords ecologists the chance to study extinction as an 61 important earth-system process (Benton 2003). However, the need to maintain functioning 62 natural ecosystems is increasingly generating motivation among scientists and the general public 63 to reverse or slow whatever biotic losses might still be addressed (Naeem et al. 2016). Concern 64 for functioning ecosystems has been elevated in recent years by a steady pulse of papers 65 reporting declines in insect abundance and diversity (Eisenhauer et al. 2019, Wagner 2019) that 66 have inspired calls for new conservation attention focused on "the little things that run the world" 67 (Wilson 1987, Goulson 2019, Cardoso et al. 2020). 68

Forister et al. p. 4

69	For certain charismatic and well-studied organisms, like the greater sage-grouse
70	(Centrocercus urophasianus) or the desert tortoise (Gopherus agassizii), governmental agencies
71	have been mobilized on a regional scale to monitor populations and management efforts, often in
72	a proactive rather than reactive way (Pilliod et al. 2020). That kind of conservation and
73	management depends on the synthesis of multiple lines of information including population
74	monitoring, natural history studies, and geographic surveys. For insects, the taxonomic diversity
75	is so great and the available information is so sparse (Cardoso et al. 2011), that proactive
76	conservation informed by diverse data types has rarely been an option. As a consequence, insect
77	conservation has often been motivated largely by fragmentation and small geographic ranges
78	(Samways 2007, Diniz-Filho et al. 2010). Exceptions to that pattern include a few European
79	countries where studies of butterflies have been sufficiently thorough in terms of natural history
80	and monitoring that researchers have been able to prioritize species for conservation attention in
81	a way that follows the International Union for Conservation of Nature (IUCN) and the Red List
82	framework (Fox et al. 2011, van Swaay et al. 2011, Maes et al. 2012, Bonelli et al. 2018). That
83	depth of species-specific information for insects is unusual, even for butterflies, and most
84	countries will have a more complex mix of some monitoring or observational data, natural
85	history observations, and expert opinion (New et al. 1995, Edge and Mecenero 2015, Geyle et al.
86	2021).

Butterflies in the western United States provide an excellent case study for the challenge of conservation prioritization that involves a mixture of heterogenous data types and sources of information. The region does include butterfly monitoring programs, but also expansive areas that are sparsely populated and understudied, including in particular the Intermountain West with hundreds of mountain ranges in the nearly 500k square kilometers of the Great Basin Desert. The

Forister et al. p. 5

most temporally-intensive butterfly monitoring program in the western US is the Shapiro transect
of ten permanent sites across Northern California that have been monitored biweekly during the
flight season for between 35 and 51 years (Shapiro 2022). Many years before the entomological
world made a collective pivot to the problem of insect declines (Hallmann et al. 2017), work
with the Shapiro data documented shifting spring phenologies (Forister and Shapiro 2003), and
the influence of land use and warming temperatures on extensive declines in abundance and
species richness (Forister et al. 2010, Casner et al. 2014b).

Within the last couple of years, the issue has been raised of geographic and taxonomic 99 heterogeneity or consistency of declines (Crossley et al. 2020, van Klink et al. 2020). This was 100 the impetus for asking if results from the temporally-intensive Shapiro dataset would be 101 consistent with geographically-extensive monitoring data from the North American Butterfly 102 Association (NABA) and iNaturalist observations across the 11 western states (Forister et al. 103 2021). That effort quantified a compounding loss of 1.6% fewer butterflies observed per year and 104 highlighted the negative influence of warming and drying conditions on butterfly populations in 105 natural areas. However, the species included in Forister et al. (2021) were only those common 106 and widespread enough to be present with sufficient frequency in monitoring databases to allow 107 for inclusion in statistical models. Moreover, an attempt was not made to combine different lines 108 of information into a ranking of species for conservation concern. 109

Here we address that need by taking a multi-faceted approach to conservation
prioritization that utilizes observational data when available (for approximately half the species)
and a combination of data types for other species, including natural history traits and quantitative
estimates of exposure to climate change and development. We also include an assessment of
subspecies using a combination of conventional conservation rankings and data on historical

Forister et al. p. 6

occurrences. The different data types are detailed below and are used (1) to produce a 115 quantitative ranking that highlights the taxa most severely declining and most likely to face 116 regional extirpation or extinction in coming decades; and (2) to identify geographic and 117 taxonomic knowledge gaps in our understanding of western butterflies. It is our hope that these 118 results will be used by conservation practitioners and land managers to guide restoration and 119 protection efforts, and will also motivate additional monitoring and the development of new 120 population models that take maximum advantage of the heterogenous data types we have 121 brought together. Throughout this paper, we use the word "risk" (and related terms, like "risk 122 index") in a flexible way that encompasses evidence of past decline, projected declines, and 123 combinations of traits that could predispose species to ongoing and future declines. This 124 flexibility is necessary given the nature of our project encompassing species for which different 125 kinds and quantities of information are available, but in all cases we intend the concept of high 126 risk to flag species that could profitably receive careful attention from ecologists, conservation 127 biologists, and the general public. 128

129

130 MATERIALS AND METHODS

A schematic overview of our methods is shown in Figure 1, emphasizing the flow of information
from external data sources through analyses to the generation of quantitative risk assessment. All
parts of the process are discussed in detail here. Starting with the 875 taxa on the North
American Butterfly Association's 2nd edition checklist of butterflies occurring north of Mexico
(NABA 2018), we retained 396 species with resident (non-vagrant) status in the eleven western
states (Washington, Oregon, California, Idaho, Montana, Nevada, Wyoming, Colorado, Utah,
New Mexico, and Arizona) based on range maps in Glassberg (2017), and collapsed 18

138	subspecies into full species. For clarity and in order to facilitate wide use of our results, we also
139	reference a second checklist by Pelham (2022) in places where names differ. The Pelham list
140	includes a larger number of subspecies and is thus important in particular for our organization of
141	subspecific risk information, discussed in detail below.
142	Of the 396 species from the NABA list, 184 were present in monitoring databases (either
143	the Shapiro transect or the NABA count circles) with sufficient frequency to be used in
144	population models. For those species, which we will refer to in text and in figures as the "A
145	group", our approach is to rank species based on observed and forecast population trajectories.
146	Acknowledging the great uncertainty inherent to insect time series analyses, we present the
147	ranking of A group species in a way that risk associated with other variables (e.g., geographic
148	range size) can be evaluated by the reader. As will be discussed below, we use iNaturalist
149	observations for A group species as a third source of historical information, but give it
150	proportionally less weight than the Shapiro or NABA data.
151	For the other 212 species (the "B group", not present in monitoring schemes in sufficient
152	frequency for inclusion in core population models), we have accumulated seven variables that
153	form a composite picture of risk: geographic range, exposure to developed land, exposure to
154	climate change, average (range-wide) precipitation, voltinism (number of generations per year),
155	wingspan, and host breadth (or "host range"). We combine those seven variables into a single
156	risk index as a weighted sum, where the weights are determined in part by our previous work
157	with western butterflies, but also by analyses of the A group (described in detail below). The
158	weighting scheme and other steps in data processing involve informed but partly subjective
159	judgements with respect to threats to butterflies and natural history traits that predispose
160	butterflies to risk. We have presented all data decisions in a transparent way, so that the reader

Forister et al. p. 8

can judge for themselves the consequences of our methods and decisions, and alternative weights
can be assigned by researchers using an online tool (see supplementary material). In the sections
below, we describe first the three observational datasets (Shapiro, NABA, and iNaturalist) and
associated analyses, then the seven other variables and how they are combined into composite
risk indices and are visualized geographically and in a phylogenetic context.

166

167 North American Butterfly Association (NABA) counts and models

The NABA butterfly count program is a suite of hundreds of individual locations throughout the 168 country that are monitored during midsummer (typically once, but in some cases more than 169 once) by a group of at least four observers recording counts of all individual butterflies seen and 170 identified to species, in a 15-mile (24.14 km) diameter circle. Observations from count circles in 171 the 11 western states encompass different numbers of years at different sites from the 1970s to 172 the present, with the final year in the dataset we examined being 2018 (the data were compiled 173 for analysis in 2019). For the current project, we filtered the observations so that we only 174 included sites that had been monitored for at least ten years, and with the final year being 2017 175 or 2018 (we did this so as not to generate forecasts for species with a substantial recent gap in 176 observations). More than one monitoring day has been reported per year at a small number of 177 sites, and for those locations we retained only the survey closest to the 4th of July, which is the 178 traditional target date for these censuses. We then excluded any site-by-species combinations in 179 which a species was not present for at least ten years (not necessarily consecutive years); finally, 180 only species meeting the latter criterion for at least three locations were retained. Those filters 181 resulted in a dataset with 162 species from 44 locations used in the core model and associated 182 population forecasts (we experimented with less stringent filters but found that model 183

Forister et al. p. 9

184	performance suffered). For species with less data, we ran a second set of models with lower
185	thresholds, as described after the core model below.

Previous work with the NABA data used hierarchical Bayesian linear Poisson regressions run separately for each species (Forister et al. 2021). Here we advance that approach using a single, multi-species model that shares information about heterogeneity in the observation process across species observed at each site (Riecke et al. 2021). The components of the model (each described in turn below) include an observation sub-model, an abundance sub-model, and a forecast or simulation process that projects occupancy (the fraction of sites with non-zero presence by species) for various intervals of years in the future.

For the observational component, we modeled the counts of individual butterflies (*y*) using a Poisson distribution given the expected count of each species at each location during each year ($\mu_{t,l,s}$), where *t*, *l*, and *s* identify the year, location, and species respectively:

196
$$y_{t,l,s} \sim Poisson(\mu_{t,l,s}).$$

¹⁹⁷ We modeled the expected count $(\mu_{t,l,s})$ as a function of an abundance index $(N_{t,l,s})$, year- and ¹⁹⁸ site-specific survey effort (β) , and a year- and location-specific random effect $(\delta_{t,l})$ shared ¹⁹⁹ among species:

200
$$\mu_{t,l,s} = \exp(\ln(N_{t,l,s}) + \beta * effort_{t,l} + \delta_{t,l}),$$

with a vague prior for the effect of survey effort:

$$\beta \sim \operatorname{normal}(0,10)$$

The empirical variable for effort is the z-standardized total hours searched by all survey groups at a site on a day. After accounting for the effect of survey effort, we modeled additional variation in detection probability for each survey or monitoring day as a random effect shared among species. This random effect can be thought of as the combined effects of survey-specific

Forister et al. p. 10

variation in detection due to processes such as variation in observer experience and local weather 207 conditions (Riecke et al. 2021): 208 $\delta_{\rm t,l} \sim {\rm normal}(0, \varsigma^2),$ 209 $\varsigma \sim \text{gamma}(1,1).$ 210 For the abundance sub-model, we assigned priors for initial population abundance indices 211 for each species at their first encounter $(f_{site_i,species_i})$ at a study site as a function of initial survey 212 effort and the initial count: 213 $N_{f_{1,s},l,s} \sim \text{gamma}(\exp[\ln(y_{f_{1,s},l,s}) + (\text{effort}_{f_{1,s},l,s} * -0.1)], 1).$ 214 We modeled changes in population size (N) from one year to the next for each species at each 215 site as a function of year (t), location (l), and a species(s)-specific population growth rate (λ): 216 $N_{t+1,l,s} = N_{t,l,s} * \lambda_{t,l,s}$ 217 Variation in population growth rate was in turn modeled as a function of a species-specific mean 218 population growth rate (γ_s), and species-specific random variance in population growth rate: 219 $\lambda_{t,l,s} \sim \text{lognormal}(\gamma_s, \sigma_s^2),$ 220 $\gamma_s \sim \text{normal}(0,1),$ 221 $\sigma_{\rm s}^2 \sim \text{gamma}(1,1).$ 222 Finally, for each species at each location, we projected the abundance index into the 223 future using Monte Carlo simulation from the posterior distributions of species-specific 224 population growth rate $(\lambda_{t,l,s})$, and species-specific population growth rate variance (σ_s^2) : 225 $N_{t+1,l,s} = N_{t,l,s} * \lambda_{t,l,s}$ 226 $\lambda_{t,l,s} \sim \text{lognormal}(\gamma_s, \sigma_s^2).$ 227 We defined local 'extirpations' as locations at which the expected count of a species given mean 228 effort was less than 0.1 individuals, and calculated extirpation probability for each species at 10, 229

Forister et al. p. 11

20, and 50 years into the future. Thus, one minus the extirpation probability is the probability of 230 population persistence, and it is that value (probability of persistence) for each species from the 231 core NABA model that moves forward (represented by 1k samples from the final year of the 232 simulations) into the calculation of the risk index for the A group species. 233 The above model and 50-year projections were used for 162 species (in the A group) with 234 sufficient data (passing filters described above). For another 105 species (with a median presence 235 of 2 sites per species), we used a less complex model. These species are part of the B group with 236 minimal observational data that were too sparse to be included in the core NABA model 237 described above. However, in the interest of presenting maximal information on all species, we 238 estimated trends through time for this subset of the B group (albeit at many fewer sites per 239 species); the results are reported but not incorporated into the risk index calculation for these 240 species. In this model, the counts (v) were also modeled with a Poisson distribution given the 241 expected count for each location and year $(\mu_{t,l})$, where t is the year and l is the location: 242 $y_{t1} \sim \text{Poisson}(\mu_{t1}).$ 243 The expected count (μ_{tl}) was then modelled as a linear function of a site-specific intercept (α_l), a 244 site-specific (s) year effect (β_1), and site-specific effect of effort (β_2): 245 $\ln(\mu_{t,l}) = \alpha_l + \beta_{1,s} * \text{year}_t + \beta_{2,s} * \text{effort}_{t,l}.$ 246 The intercept and both beta coefficients were drawn from normal priors, with the normal 247

truncated at zero to be positive for effort (β_2); the means and variances of those distributions were in turn drawn from hyperpriors (thus estimating effects across sites) with means drawn from normal distributions (with mean of zero and variance of 100) and variances drawn from gamma(1,1) as in the core model above. For 35 species present at only a single site, the model was run without the hierarchical (across sites) structure. The output of these secondary models

253	(for the 105 species) was retained as a directional probability (the fraction of the posterior
254	distribution above zero for species with a positive year coefficient, and below zero for species
255	with a negative year coefficient).
256	All Bayesian models were implemented using JAGS (version 4.3) and the jagsUI
257	package (Kellner 2017) in R (R Core Team 2020). The core model (for A group species) was run
258	with three chains for 500k iterations, with a 250k iteration burn-in. The secondary models (for
259	the 105 B group species with some presence in the NABA data) were run with two chains for 2k
260	steps and a 1k burn-in. Model diagnostics included inspection of plots of chain histories (all
261	chains converged; $\hat{R} < 1.01$), and effective samples sizes.
262	
263	Shapiro transect data and models
264	Ten long-term study sites across northern California have been monitored for between 35 and 51
265	years (depending on the site), with the presence of all butterflies noted along fixed routes every
266	two weeks during the flight season. Data used here were compiled in 2021, including
267	observations through 2020; earlier years were truncated so the dataset starts at 1985, except for
268	three sites where data collection began in 1988. Species by site combinations of at least eight
269	years were retained for analyses of 133 species. Additional details on sites, butterflies and field
270	methods have been described elsewhere (Forister et al. 2010, Halsch et al. 2021, Shapiro 2022).
271	In brief, data from the Shapiro sites have been analyzed using hierarchical Bayesian linear
272	models in which the response variable (the number of days a species is observed in a year) is
273	modeled as a binomial process, with a beta coefficient from the year term in the linear model
274	representing change through time in the probability that a species is observed (Nice et al. 2014,
275	Halsch et al. 2021). Here we use the version of this model and implementation as described in

Forister et al. p. 13

276	Forister et al. (2021) in which the model was run separately for each species and beta
277	coefficients for years are estimated within and across sites; the higher level coefficients (across
278	sites) are used as indices of population change for each species across the northern California
279	sites. As with the NABA models, model diagnostics included inspection of convergence and
280	effective sample sizes. For downstream analyses (the creation of the risk index for A group
281	species), 1k samples were retained from the posterior distributions of the year coefficients
282	estimated across sites for each species. For two species, Lycaena rubidus and Agraulis vanillae,
283	the year coefficients were extreme outliers (in the negative and positive direction, respectively)
284	and were not used in the creation of the risk index values (described below) but we do include
285	those coefficients in visual summaries of patterns across species.
286	The year coefficients from this modeling approach have been shown to be effective
287	indices of change in total abundance as reflected in total counts of individuals which are
288	available from a subset of years and sites (Casner et al. 2014a). Unlike the main NABA model,
289	described in the previous section, we have not taken a forecasting approach with the Shapiro
290	data. The two datasets have different strengths and weaknesses. The strengths of the Shapiro data
291	are intensity and consistency of observation, which lend precision to estimates of species-
292	specific change through time. In contrast, the NABA observations are only once per year, but the
293	geographically distributed nature of the NABA sites (with greater independence among
294	locations) lends value to the forecasting of population occupancy with our simulation approach

²⁹⁵ (which does not account for dispersal or demographic connections among locations).

296

297 iNaturalist observations and expected ranges

298	Observations recorded on the iNaturalist platform are a wealth of geographic and phenological
299	information, which researchers are increasingly using to answer ecological questions (Prudic et
300	al. 2018, Kirchhoff et al. 2021), even for rare insects (Wilson et al. 2020). In our previous work
301	with western butterflies, we used iNaturalist records in time series models, which revealed trends
302	that were generally consistent with temporal patterns in the NABA and Shapiro data (Forister et
303	al. 2021). Here we take a different approach, with the goal of using the broad geographic
304	coverage of iNaturalist records to generate information on species status that is complementary
305	to the detailed time series information from the other two observational datasets. We used
306	iNaturalist records from the last 15 years (2007-2021) to generate a community scientist-derived
307	estimate of area of occupancy. Those area of occupancy estimates were then compared to expert-
308	derived range area estimates (described below) in a simple linear regression, and residuals from
309	that relationship were saved. In other words, we asked which species have been seen more or less
310	frequently in the last 15 years relative to the expected area based on the expert-derived range.
311	To generate the citizen scientist-derived area of occupancy estimates, we downloaded
312	iNaturalist research grade observations from the Global Biodiversity Information Facility
313	(GBIF.org 2021a, 2021b, 2021c, 2021d, 2021e, 2021f) for all butterfly species in the 11 western
314	states. We retained observations from 2007 onwards for species that were observed at least 100
315	times (with very few exceptions, these were all A group species, present in Shapiro or NABA
316	datasets, thus the analysis of iNaturalist records was confined to the A group). We calculated an
317	unweighted Gaussian Kernel Density estimate using the function sp.kde from the R package
318	spatialEco v1.3-7 (Evans and Ram 2021) based on the iNaturalist observations with a
319	distance bandwidth of 2 (four examples are shown in Figure S1). The resulting raster was

Forister et al. p. 15

320	converted to a disjoint spatial polygon (i.e. not all parts of the range needed to be connected) that
321	encompassed all values > 0.00001 and calculated the area of the resulting range map.

322

323 Geographic ranges and voltinism

Expert-derived range estimates from Glassberg (2017) were generated from Keyhole Markup 324 Language (.kml) files for each species. The range of each species was separated by voltinism 325 (the number of generations per year in different portions of the range), with spatial polygons 326 retained separately for uni-, bi-, and multivoltine regions. Quantitative areal estimates were then 327 derived for all range portions within 11 western states using the area function in the R package 328 raster v3.5-11(Hijmans et al. 2021), which estimates area based on the size of raster cells. 329 This estimate is biased closer to the poles; however, we only generated range areas within the 11 330 western states (excluding portions of ranges that extended farther east, north into Canada, or 331 south into Mexico) and thus the bias is expected to be minimal. 332

The expert-derived geographic ranges were used for multiple purposes (see Figure 1), 333 including comparison with iNaturalist ranges (described in the previous section). The total 334 expert-derived range estimates for each species were also used as a variable that contributes to 335 the composite risk index, as did the fraction of the range that was univoltine (i.e., for simplicity, 336 we focus on a univoltine vs bi- plus multivoltine comparison, rather than considering bivoltinism 337 as a distinct category). The outlines of the expert-derived ranges were also used to calculate 338 exposure to land use and climate (as described in the next section) and to calculate weighted 339 latitudinal midpoints as another geographic descriptor. We used the function rasterToPoints from 340 the raster package v3.5-11 (Hijmans et al. 2021) to convert each species range map to 341

Forister et al. p. 16

342	coordinates; weighted latitudinal midpoints were then calculated that account for the longitudinal
343	width of the range (i.e. the mean latitude across all cells in the raster).

As yet another line of expert-derived geographic information, we assigned each species a 344 qualitative biogeographic designation of North, South, East, or West to reflect where the 345 majority of the range area is found. For example, species labelled as western have the majority of 346 their range in the 11 western states, with a minor presence north or south of the US borders or in 347 the eastern states. Northern species have most of their range in Canada with only outlier (and 348 often isolated) locations in the western US; similarly, southern species are those with ranges in 349 Mexico often extending only dozens or hundreds of square kilometers into Arizona or New 350 Mexico. Finally, species were labelled as eastern if they either had a transcontinental range (e.g., 351 Pieris rapae, Vanessa cardui) or had a range almost entirely in the eastern states with only a 352 minority of the range area in the 11 western states. These assignments were made by visual 353 inspection of range maps in field guides (Scott 1986, Glassberg 2017). 354

355

Land use and climate change

Previous work with butterflies in our region has revealed effects of land use and climate change 357 that are complex, potentially interacting, and dependent on both the species involved and the 358 landscape context (Casner et al. 2014b, Forister et al. 2018, Halsch et al. 2021). Summarizing 359 exposure to land use and climate change is not a simple task, but we have taken the relatively 360 straightforward option of using the range outline (described in previous section) to quantify these 361 stressors within the range of each species. Note that this differs from the use of point locations to 362 quantify proximity to, for example, urban development (Jamwal et al. 2021). The range-outline 363 approach is a better fit for our goals simply because all species have the same starting data (the 364

Forister et al. p. 17

365	expert-derived ranges), which would not be true of 396 species using available point-occurrence
366	records in, for example, iNaturalist. For highly mobile animals, like butterflies, the range-outline
367	method has another advantage in that we do not have to assume that point locations of
368	observations represent the only or most relevant habitats.
369	To quantify land use change, we reclassified the 2020 Cropland Data Layer (USDA
370	2020) into land cover types of agriculture, development, or natural and semi-natural habitats
371	using the associated Cropland Data Layer scheme; all crops were classified as agriculture,
372	development of any intensity level as development, and remaining land cover types (including
373	pastureland) as natural or semi-natural habitat. For each species, we used the spatial polygon
374	generated from the range map to clip the rasterized land cover types and calculated the
375	proportion that was agriculture or development. This was done separately for regions of different
376	voltinism, but these were summed to a single value for each species (see Figure S2 for examples
377	of range-wide exposure to land use).
378	To estimate climate change exposure, we used TerraClimate data for minimum
379	temperature, maximum temperature, and precipitation (Abatzoglou et al. 2018), which we
380	resampled from ~4km spatial resolution to ~40km for computational efficiency. Using
381	multivariate Mahalanobis distance as a measure of departure (Farber and Kadmon 2003,
382	Abatzoglou et al. 2020), we calculated departure from baseline conditions (1958-1987) for the
383	most recent thirty years (1991-2020) for each cell. To estimate exposure to climate change, we
384	calculated rate of change in departure over time using Theil-Sen slopes (Theil 1950, Sen 1968)
385	which estimate the median slope between each pairwise set of observations and are relatively

departures for the eleven western states. For each species, we then clipped the climate departure

robust to outliers near the start or end of a series. We generated a raster of these trends in

386

Forister et al. p. 18

388	raster layer using the species range maps as spatial polygons and calculated the mean climate
389	change exposure across that portion of the range (as with land use, this was done separately by
390	voltinism, but then added for a single value per species for further analyses; see Figure S2 for
391	examples). We also calculated 30-year climate normals (1991-2020) for minimum temperature,
392	maximum temperature, and precipitation annually and within each season across the entire range
393	for each species. Among those three variables, precipitation was recently found to be predictive
394	of changes in butterfly abundance across the west (Forister et al. 2021), thus it was used as a
395	static description of climate for inclusion in the composite risk index (described below).

396

397 Wingspan and host range

Among the many morphological and natural history traits that could be informative of status and 398 risk, body size and ecological specialization are widely studied, and thus relevant data are 399 available for many species. More narrow diets are often associated with greater sensitivity to 400 habitat loss and other disturbance (Hughes et al. 2000), and dispersal ability is a key determinant 401 of metapopulation resilience in the face of fragmentation or other stressors. Wingspan has been 402 shown to be a proxy for dispersal ability in butterflies (Sekar 2012). For most of the butterflies 403 studied here, wingspan was previously estimated (in Forister et al. 2021) with data derived from 404 Opler (1999). For a small number of species included in the present study for which a 405 measurement was not available from that source, we supplemented with ad hoc online searches. 406 Similarly with diet breadth (or host range), we used a single source for the vast majority of 407 species (Scott 1986), and supplemented from other field guides and other online resources for the 408 few species with missing data. 409

Forister et al. p. 19

410	We gathered both the number of plant genera and plant families reported as caterpillar
411	hosts for each species, and then calculated a combined index of diet breadth as the number of
412	taxonomic families plus the natural log of the number of genera. That calculation of taxonomic
413	diet breadth puts most weight on the number of families but allows for some influence of the
414	number of genera eaten. For example: a species that uses hosts in two genera in two families
415	would have a diet breadth of 2.69 $(2 + \ln(2))$, while a species that uses plants in three genera in
416	two families would have a diet breadth of $3.10 (2 + \ln(3))$. We did not attempt to gather species-
417	level host records, for which too much data would be missing or unreliable.

418

419 **Transformations**

In total, we compiled ten variables that contribute to the prioritization of A and B group species 420 in different ways: (1) 50-year occupancy projections (probabilities of population persistence) 421 based on NABA data; (2) historical rates of change from the Shapiro data; (3) recent change in 422 range based on the difference between community scientist-derived and expert-derived ranges; 423 (4) geographic range based on expert assessment; (5) exposure to agricultural and other 424 developed lands; (6) exposure to climate change; (7) average precipitation throughout the range; 425 (8) the fraction of the range with one generation per year; (9) wingspan; and (10) an index of diet 426 or host breadth (Figure 1). Prior to their use in assigning a risk value to each species (discussed 427 in the next section), each variable was subjected to a specific set of transformations that resulted 428 in a variable with a range of 0 to 1 where larger values represent greater risk. Depending on the 429 nature of the variable (when larger values do or do not naturally represent higher risk), the 430 transformations included inversion, and (for all variables) standardization between 0 and 1 (by 431 dividing by the largest value). In some cases, for highly skewed variables, a natural log 432

Forister et al. p. 20

433	transformation was applied as the first step. For example, wingspan was first log transformed,
434	then multiplied by -1, such that all values become negative and the larger wingspans become
435	larger negative numbers; the distribution was then shifted to the positive by adding the absolute
436	value of the smallest (most negative) value to all of the numbers; finally, the distribution was
437	divided by the largest value, thus scaling the numbers between 0 and 1, where the smallest
438	wingspans (representing the greatest risk because of less dispersal ability) are now closest to 1.
439	All transformations and scaling steps are illustrated in Figure S3.
440	For visualization of the transformed and scaled variables, we divided the distributions
441	(Figure S3) into quantiles and assigned circles of different sizes to the different intervals, with
442	larger circles indicating larger values and greater assumed risk. For most of the variables, we

found that the following breakpoints provided a useful assignment of circles for visualization:

0.15, 0.5, and 0.85; in other words, the interval from 0 to 0.15 was assigned the smallest circle
(the least risk), from 0.15 to 0.5 the next largest, etc. Breakpoints differed for some of the more
skewed variables (e.g., host range), but the results are interpreted in the same way (larger circles
represent larger assumed risk).

448

449 Calculation of risk index for A and B group species

The A group species are those species for which data were available from at least one of the monitoring programs (Shapiro or NABA), and many of these species also had enough iNaturalist observations for analysis. For these species, we calculated a weighted sum based on those three lines of information with weights as follows: 47.5% NABA, 47.5% Shapiro, and 5% iNaturalist. The small weight given to the iNaturalist data reflects the fact that the data are heterogeneous (in space and among species) and rapidly accumulating; these data are thus complex and potentially

Forister et al. p. 21

456	important but still only barely explored from an analytical perspective. Alternative weighting
457	schemes among all variables (including the three observational variables) can be explored using
458	an interactive, online tool; see supplementary material.

The variables used in our A group weighting scheme (multiplied by the three 459 percentages, 47.5%, 47.5%, and 5%) were the 50-year probabilities of persistence (from NABA), 460 historical rates of change (from Shapiro), and observed range changes (from iNaturalist) that had 461 been transformed (see previous section) such that larger values represent greater evidence for 462 decline or (in the case of NABA) projected decline. Thus, a species with the most severe 463 declining values (historical or projected) for each dataset would receive a composite risk score of 464 1. To incorporate uncertainty retained from Bayesian analyses of the NABA and Shapiro data, 465 the composite risk index was recalculated 1k times using 1k samples of the relevant posterior 466 distributions; we then calculated a mean and 85% highest density interval of risk for each 467 species. 468

The B group species are those lacking observational data. Thus, we used a composite of 469 the other seven variables to estimate risk. We experimented with a number of weighting schemes 470 for those seven variables and settled on an approach that was partly influenced by previous 471 research (e.g., Forister et al. 2021) but also informed by an additional analysis of the species in 472 the monitoring data. Specifically (for that additional analysis), we took the composite risk index 473 for the A group species (based on NABA, Shapiro, and iNaturalist data) and used linear 474 regression models to determine which of the other seven variables were most predictive of that 475 risk index (following general protocols with other Bayesian models as described above). The 476 exact weighting scheme for B group species (influenced partly by results of the analysis of the A 477 group) is described fully in results below. Clearly many schemes are possible for a weighted sum 478

Forister et al. p. 22

of seven variables, and we report correlations among outcomes from different schemes. Finally,
many of the B group species had some data from the NABA dataset that were not sufficient for
inclusion in our main model and occupancy forecasts. For those species, we ran a less complex
model (described above as the secondary set of NABA models) and report the results along with
other B group results, but we do not incorporate those values into the B group risk index to
maintain consistency in risk index calculations.

The calculation of the risk index for both the A and B groups relied on a complete data 485 matrix. For most of the variables used for the B group, there were no missing values, specifically 486 for all of the variables deriving in part from the expert geographic ranges: range area, voltinism, 487 precipitation, development, and climate departure (Figure 1). A few species lacked data for host 488 range, and these we filled with interpolation of the median value calculated across all species. A 489 more consequential decision was to similarly use median interpolation with the observation data 490 and the A group species. In other words, a species without sufficient iNaturalist observations for 491 analysis was given the median value associated with that variable prior to the calculation of the 492 risk index. The same was true for species not represented in the NABA or Shapiro data: lacking 493 any other information we assume those species are simply following the central tendency (for 494 historical and projected change) as estimated across other species. 495

496

497 **Risk index for subspecies**

A list of subspecies present in the western United States (Arizona, California, Colorado, Idaho,
 Montana, New Mexico, Nevada, Oregon, Utah, Washington, and Wyoming) was gathered from
 Pelham (2022). Geographic inclusion for subspecies was based on described ranges in original
 descriptions and records from public databases (n = 1,004 subspecies). Two categories of

Forister et al. p. 23

502	conservation need were considered in ranking subspecies: 1) global, national, and subnational
503	(state) ranks assigned by the organization NatureServe; and 2) the last year a subspecies had a
504	publicly available recorded observation based on our inspection of available databases.
505	A full description of NatureServe ranking methods and rank descriptions is available in
506	Faber-Langendoen et al. (2012). Briefly, ranks are assigned a 1-5 number where 1 is Critically
507	Imperiled, 2 is Imperiled, 3 is Vulnerable, 4 is Apparently Secure, and 5 is Secure. Rankings are
508	assessed using a Rank Calculator that includes aspects of rarity, threats, and population trends.
509	Ranks are used to assess imperilment over the entire (global) range of subspecies as well as at
510	the national and state levels. For subspecies, global rankings are indicated by a "T" rank
511	following the global rank. Thus, an Imperiled subspecies (rank of 2) of an Apparently Secure
512	species (rank of 4) would have the rank G4T2. National ("N") and state ("S") rankings are
513	assessed separately for species and subspecies. Additional ranks are "X" for taxa that are
514	presumed extirpated, "H" if a taxon is possibly extirpated with records in the last 20-40 years
515	and might be rediscovered, as well as others including "NR" for taxa that have not yet been
516	assessed (Faber-Langendoen et al. 2012).
517	First, we created a summary measure of subspecies imperilment at various geographic
518	scales based on the NatureServe evaluations by creating a quantitative scale for global, national,
519	and state rankings. Points increased with imperilment, such that a rank of G1 is worth 4 points

⁵²¹ ("G") ranks, global subspecies ("T") ranks, national ("N") ranks, and each state with a S1-S5

520

and G5 is worth 0 points. This scoring was completed similarly for the nominate species global

ranking. State scores for each species were averaged to create a single state-level score. Any
 other rankings including "X" and "H" were scored as a 0 as these are equally uninformative with

Forister et al. p. 24

524	respect to realized conservation need. A total score for each taxon was calculated as the sum of
525	the global, national, and state scores such that a taxon could have a score from 0-16.
526	Second, the most recent year of observation for each subspecies was collected from
527	various accessible databases, websites, photographic collections, and peer-reviewed literature,
528	with a goal of finding one observation for a taxon from 2001 or more recent as evidence of
529	recent presence. Searches began with all specimen records by family in the Symbiota Collections
530	of Arthropods Network for the 11 western states (SCAN 2022). For any taxon that did not have a
531	post-2001 observation, progressive searches through the literature were made until one was
532	found or until all references had been searched for the most recent record available; a full list of
533	resources used is available Table S1. Any taxon with a record from 2001 or more recent received
534	a score of 0, and each year previous to this increased the score by 1 point. Scores for this
535	category ranged from 0 to 34 (for Megathymus yuccae harbisoni). While we have done our best
536	to collect available observations, identification to the subspecies level was challenging at times
537	due to either recent taxonomic changes or difficulty identifying individuals using only
538	photographs. We do not believe these issues significantly affect our overall ranking of
539	subspecies.
540	As with the species-level risk assessment, subspecies values for each of the three

As with the species-level fisk assessment, subspecies values for each of the three categories were normalized to be between 0 and 1 before the calculation of a composite index. The NatureServe score was given 75% weight towards a total score as it includes the most information regarding total threat or risk, and the year of observation scores were given 25% weight. Weighted scores for the two categories were added together to create a single comprehensive score for each subspecies between 0 and 1. Those scores were used to rank the subspecies, and we also asked if subspecies risk values were correlated with risk calculated

Forister et al. p. 25

independently at the species level (calculations described in previous section). For species with
multiple subspecies evaluated for risk, the subspecies values were averaged (within each nominal
taxon), and then a simple Pearson correlation was calculated between the two sets of risk values
(at the species and subspecies level).

551

552 Geographic and Phylogenetic visualization of risk

Finally, we asked how the composite risk indices were distributed across the landscape and 553 across the phylogeny of western butterflies. From a spatial perspective, we calculated both 554 species richness (separately for each cell in a raster covering the extent of the eleven western 555 states) and average risk among species present in a cell. We did this separately for the A and B 556 group species, and we restricted analyses to only species with higher risk values by subsetting to 557 the upper 75th quantile of risk values separately for each list (A and B). Within those higher-risk 558 groups, we converted each species range map from a spatial polygon to a raster layer where 559 values within the range were set to 1 and values outside the range to 0. We summed these values 560 across all rasters to produce a new raster of species richness. To calculate mean risk for each cell, 561 we divided the cumulative risk index raster by the species richness raster. 562

For the evolutionary perspective, we used the phylogeny from Zhang et al. (2019) for all 845 butterfly species from the United States and Canada. Briefly, this tree was based on 756 universal single-copy orthologs we identified from 36 reference genomes using OrthoMCL (Li et al. 2003). Sequences of these orthologs were aligned using both local (BLAST [Altschul et al. 1997]) and global (MAFFT [Katoh et al. 2002]) alignment methods, and only positions that were consistently aligned by both methods were used. Sequences of non-reference species were derived by mapping the Illumina reads to the exon sequences of the reference species and

Forister et al. p. 26

570	performing reference-guided assembly. Multiple sequence alignments (MSA) of different
571	orthologs were concatenated to a single MSA. This MSA was partitioned by codon position and
572	used to build a tree by IQ-TREE (version 1.6.12) (Nguyen et al. 2015) with the most suited
573	evolutionary model automatically found by IQ-TREE.
574	The phylogeny was imported as a time-calibrated .tre file into R and pruned to our focal
575	western butterflies (the combined A and B group lists). The package ggtree (Yu et al. 2017)
576	was used to plot a phylogeny with tips labelled by risk categories assigned based on the quantiles
577	of the risk distributions separately for the A and B group species. Specifically, species in the
578	upper 90th quantile were labelled as "high risk," species between the 75th and 90th quantiles
579	were labelled as "medium risk", and species below the 75th were "low risk." Finally, the
580	phylosig function from phytools (Revell 2012) was used to calculate lambda and K (with
581	1000 simulations for the permutation test) as measures of phylogenetic signal for the continuous
582	risk index across all species, which in this context is informative with respect to the extent to
583	which closely related species share similar levels of risk.

584

585 **RESULTS**

We calculated an index of risk for 396 species, which includes two groups: 184 species in the A group with extensive monitoring or observational data, and 212 species in the B group without observational data (or without enough to be used in our primary population models). Not surprisingly, the B group species tend to have smaller geographic ranges (Figure 2a), which in part explains their reduced presence (just by geographic chance) in monitoring groups, but the two groups differ in other ways (Figure 2). The B group species have slightly lower exposure to development (Figure 2b) and moderately higher exposure to climate change (Figure 2c). The

Forister et al. p. 27

		orist
593	higher climate change exposure is explained in part by the greater presence of more south	ern

species in the B group, as seen by latitudinal midpoints (Figure 2g) and qualitative
 characterization of range (Figure 2h).

For the A group species, we modeled historical and projected population trajectories 596 using different sources of observational data. Consistent with previous work with NABA data, 597 our new model with shared (across-species) observation heterogeneity found a majority of 598 species (71%) with annual growth rates below replacement (Figure S4). We used those estimated 599 annual growth rates and the most recent year of observed counts to simulate 50 years into the 600 future. The median fraction of extant locations (or probability of local persistence) per species at 601 50 years was 0.60, and that fraction was positively related to historical population growth rates 602 (Figure S4). Results from analyses of Shapiro data also find a majority of species with downward 603 trends through time of varying magnitude (84.5% of species have negative year coefficients). We 604 combined the 50-year persistence estimates (from the NABA model) with historical rates of 605 change (from the Shapiro data) and an estimate of shift in range size based on community 606 scientist observations (from iNaturalist) relative to expert range sizes to generate a composite 607 risk index for the A group species. Note that the A group species are shown in Figure 3 with risk 608 information associated with the other seven variables (geographic range, exposure to 609 development, etc.), even though the actual ranking of the A group is based solely on the 610 observational data (NABA, Shapiro, iNaturalist). We present the information in this way because 611 we acknowledge the imperfect geographic coverage of monitoring programs and the inherent 612 uncertainty in population models. Thus, the reader or conservation practitioner can easily see if 613 two species with similar risk values in the A group (based on NABA, Shapiro and iNaturalist 614 results) potentially have similar risk based on other variables like range size. We also generated 615

616	the risk values with an even split in weights between NABA and Shapiro data (leaving out
617	iNaturalist), and found that the resulting risk values were correlated with our primary risk values
618	at $r = 0.997$ (which reflects primarily the low weight assigned to iNaturalist observations but also
619	the fact that they are correlated with results from Shapiro data, Figure S5).
620	Without observational data, the ranking of B group species required a partitioning of
621	weights among the other lines of information. To partly inform that process, we used the A group
622	species to estimate the effects of other variables on risk index (based on NABA, Shapiro, and
623	iNaturalist data). The model explained a relatively small proportion of variance in the risk index
624	(Table S2), but did demonstrate that smaller wingspans (99% probability of effect) and lower
625	range-wide precipitation (86% probability of effect) are associated with risk for the A group
626	species. In addition, we also suspected climate change would be important based on our previous
627	work with western butterflies (Forister et al. 2021, Halsch et al. 2021). This is especially true
628	given the large presence of B group species with ranges in the desert southwest (Figure 2h), a
629	region heavily impacted by warming and drying trends. We adopted the following weighting
630	scheme to calculate a single risk value for each species in the B group: 20% precipitation, 20%
631	wingspan, 20% climate change, 10% development, 10% range size, 10% voltinism, and 10%
632	host range; correlations among the seven variables as well as the three observational variables
633	(for the A group) are shown in Figure S5. As a comparison to that scheme, we also ranked the B
634	group species with equal weights among the seven variables (14.3%); the resulting risk values
635	were correlated at $r = 0.90$ ($t = 29.32$, df = 210, $P < 0.001$) with the values from the primary
636	scheme. With a third weighting scheme based on 50% from each of average range-wide
637	precipitation and wingspan (the two variables identified by the analyses of A group risk), the
638	correlation with the main scheme was $r = 0.55$ ($t = 9.66$, df = 210, $P < 0.001$).

639	The top fifty species with the highest risk values from each of the A and B groups are
640	shown in Figure 3 (the other species with lower risk values are in Figures S6, S7 and S8). For the
641	highest-ranked A group species, agreement between the two monitoring schemes is apparent
642	with large "risk circles" in both the NABA and Shapiro columns (Figure 3a). In some cases,
643	these top-ranked A group species have also been seen less frequently over the last 15 years
644	relative to expectation based on the expert-derived range maps (see the iNaturalist column in
645	Figure 3a). Time series plots for two of those top species are shown in Figure 4 (Vanessa
646	annabella) and Figure 5 (Euchloe ausonides); in Figure 6, neutral or upward trajectories can be
647	seen for Poanes melane, the species with the lowest risk index among the A group species
648	(Figure S8). Similar plots for all other A group species are available through an online tool (see
649	supplementary material). The rankings for the A group species are shown with 85% credible
650	intervals (Figure 2a), which are broad; this uncertainty reflects the high inter-annual variability
651	inherent to the time series data being modelled (from both NABA and Shapiro) and should be
652	kept in mind when interpreting the position of species on the A group list.
653	We compiled data for 1,004 subspecies, and ranked them using criteria that were largely
654	based on NatureServe ratings, but also included the last year in which an observation was
655	reported for a particular taxon. The 50 subspecies with the highest priority for conservation are
656	shown in Table 1, where the high frequency of butterflies in the family Lycaenidae is notable,
657	with almost half (22 out of 50) in the top 50 list in that family. Another 15 taxa are in the
658	Nymphalidae family, 8 of which are subspecies of Speyeria [Argynnis], a charismatic group of
659	subspecifically diverse species. It is interesting to note that the split between A and B group
660	species in the top 50 subspecies list is 31 A group and 19 B group, which at least suggests that
661	the evaluation of subspecies is not necessarily biased towards species with the smallest ranges

Forister et al. p. 30

(which tend to be the B group species; Figure 2a). Additional information on the 1,004
subspecies that we evaluated is available in archived data for this project (see data availability
statement), and in an online tool where output similar to Table 1 can be filtered by state and by
family (see supplementary material).

We also asked if the risk index calculated at the subspecies level could be predicted by the risk index calculated at the species level (Figure 3). An overall correlation was detected between the two indices at r = 0.17 (t = 2.05, df = 140, P = 0.04), and the relationship was driven by the B group species. With the data split into the A and B groups, a correlation was not detected for the former (r = 0.10, t = 0.95, df = 99, P = 0.34) but was for the latter: for the species without monitoring data (the B group) the risk index calculated at the species level is correlated at r = 0.28 (t = 2.15, df = 55, P = 0.0036) with the risk index at the subspecies level.

Finally, we examined the distribution of the species-level risk index geographically and 673 phylogenetically. Considering the species with the highest risk index values (above the 75th 674 quantile of risk values) for the A group, across the 11 western states the spread of average risk 675 shows a partially inverted relationship with richness of the most at-risk species in some parts of 676 the region (Figure 7). For example, average risk is high in the northern Central Valley of 677 California and in the northwestern region of Oregon (Figure 7a), while total richness of at-risk 678 species is lower in those areas (Figure 7b). Similarly, richness of at-risk species is high in the 679 Sierra Nevada, but average risk is low. The distributions of risk for the B group species highlight 680 the bias of that group towards the most southern areas, with high average risk along the southern 681 California coast (Figure 7c) and a concentration of at-risk species along the border between 682 Mexico and New Mexico (Figure 7d). 683

Forister et al. p. 31

684	The phylogenetic picture of risk shows multiple clusters of at-risk species, and some
685	lineages with notably lower risk, like the Papilionidae and much of the Nymphalidae (Figure 8).
686	The families sharing the disproportionate amount of risk are the Hesperiidae (with 16% of
687	species in the high risk category, above the 90th quantile of risk) and the Lycaenidae (with 14%
688	of species at high risk); these are followed by the Riodinidae (with 13% of species at high risk,
689	albeit based on a small sample size with the family represented by only 8 species) and the
690	Pieridae (with 12% of species at high risk). The percentages of high risk species in the
691	Papilionidae and Nymphalidae are just 8% and 1%, respectively (Figure 8). Tests of
692	phylogenetic inertia are consistent with the observation of phylogenetically clustered risk
693	(Pagel's $\lambda = 0.39$, $P < 0.001$; Blomberg's $K = 0.052$, $P = 0.001$ based on 1k randomizations).
694	

695 **DISCUSSION**

Our primary goal in this paper has not been to document butterfly declines or to identify traits 696 that make insects more or less sensitive to the stressors of the Anthropocene, as these topics have 697 been addressed elsewhere for North America (Schultz et al. 2019, Wepprich et al. 2019, Crossley 698 et al. 2021, Forister et al. 2021), the Neotropics (Janzen and Hallwachs 2019, Salcido et al. 699 2020), and numerous other parts of the world (Nakamura 2011, Fox 2013, Wagner 2019). 700 Instead, our goal has been to organize and analyze heterogenous data sources in a way that 701 allows conservation biologists to identify the butterflies in the 11 western US states that are most 702 likely to suffer serious reductions in range or population size in coming years. We hope that our 703 work advances the issue of the prioritizing of species for conservation given mixed data types, 704 uneven spatial coverage and uncertainty in historical trends. Although some parts of the world 705

706	(notably countries in western Europe) have dense coverage with standardized monitoring,
707	prioritization in most of the world will involve some mix of monitoring and trait-based inference.
708	The western states have been our region of study, rather than the entire US, because the
709	impacts of climate change are severe and distinct in this arid region (Gonzalez et al. 2018), and
710	the butterfly fauna is similarly shaped by a unique topography and climatic history (Shapiro
711	1996, Hawkins 2010). At the continental scale, butterflies in the west also appear to be
712	experiencing the most severe declines (Crossley et al. 2021). As a consequence of expansive
713	areas with low human population density, about half of the butterfly species in the region are not
714	included in the monitoring datasets used here, yet we have brought together information on the
715	entire fauna (with the exception of a few species with rare occurrences, mostly strays across the
716	US-Mexico border). Because of this, our study has an apples-and-oranges structure (species with
717	and without monitoring data) that extends to the interpretation of the risk index values and
718	engenders certain ironies. Chief among the ironies of our work is the fact that we rank B group
719	species in part by certain variables (geographic range, exposure to climate change, etc.) that are
720	not evidently associated with declines in the species for which we have historical records (the A
721	group). In other words, considering Figure 3, the A group species near the top of the list do not
722	necessarily have the smallest ranges, and the same can be said of other variables. Even for the
723	two variables (wingspan and average precipitation) which do predict risk in the A group, the
724	variance explained is low (Table S2) yet we still emphasize these variables in ranking the B
725	group species. We discuss these apparently counterintuitive decisions below, and then discuss
726	phylogenetic and geographic hotspots of risk. Finally, we end with a consideration of individual
727	taxa most deserving of attention given available evidence.

728	Among the complexities of variables potentially associated with risk, an understanding of
729	geographic range starts by noting that the A group species have broader geographic ranges
730	(Figure 2a), which is indeed why they are present at enough NABA sites to be included in our
731	core population model. Thus the fact that many of the most severely declining species are
732	widespread (e.g., Vanessa annabella in all 11 states) does not diminish the logic of prioritizing B
733	group species based in part on small range size, which is a well-known determinant of risk
734	(Staude et al. 2020). Similarly, the effects of voltinism and ecological host specialization are
735	relatively straightforward: everything else being equal, we expect a species with multiple
736	generations per year and an ability to utilize many hosts to be more resilient (to any number of
737	stressors) than another species without those traits (Eskildsen et al. 2015).
738	The interpretation of other variables is less straightforward, chief among them being
739	exposure to climate change. Previous work with western butterflies has identified warming and
740	drying conditions as stressors, based in particular on analyses of geographic variation among
741	study sites in climate change effects and changes in aggregate butterfly density (Forister et al.
742	2021). At the species level (rather than the level of individual study sites), the same signal is not
743	as apparent in the present study for the A group species (in other words, the species towards the
744	top of the A group list do not have particularly high exposures to climate change). This is
745	because most of these species have large enough ranges that their exposure to climate change
746	(when quantified across the entire range) includes areas with both more and less severe warming
747	and drying that tend to cancel each other out at the scale of broadly-distributed species.
748	However, the B group species have smaller and more southern ranges (Figure 2), which is the
749	part of the west most impacted by climate change (Gonzalez et al. 2018). Thus, we believe

Forister et al. p. 34

exposure to climate change is well justified as a contributing factor to risk specifically for these
species for which we lack monitoring data.

Exposure to development (urban, suburban and agricultural lands) requires similarly 752 careful interpretation. This is chiefly because the data most well suited to understanding the 753 effects of habitat destruction on insects will rarely be collected: places that have already been 754 developed will not be monitored, and existing monitoring efforts will often be located in more 755 pristine locations even when relatively proximate to human habitation. The Shapiro dataset is an 756 exception, as it encompasses a severe land use gradient from the agricultural and urban Central 757 Valley to the undeveloped high elevations of the Sierra Nevada. From that program, we know 758 that land conversion and contamination (with pesticides) have effects of similar magnitude at 759 low elevations (Forister et al. 2016). Though similar information does not exist across the west, 760 we included exposure to development in our rankings here for the B group species for the simple 761 reason that common sense suggests that a range that encompasses more development is likely to 762 experience increasing fragmentation and contamination in coming years relative to a species with 763 less exposure. 764

Geographic projections of risk for B group species emphasize the southern areas of the 765 west (Figure 7), but also point to specific hotpots of average risk that include the southern 766 California coast. Like A group species in the Central Valley of California, that coastal region has 767 low richness of B group species, but on average the species that are there in the vicinity of the 768 Los Angeles basin score high for our risk factors. Arizona and southwestern New Mexico have a 769 high concentration of B group species with high risk factors, thus this area should be prioritized 770 for future monitoring efforts. For A group species, the Sierra Nevada Mountains (especially the 771 northern Sierra), the Colorado Plateau and the southern Rocky Mountains are hotspots of 772

Forister et al. p. 35

773	declining species (Figure 7). These same places have been recently identified as hotspots of
774	imperiled species in analyses that included plants, vertebrates, freshwater invertebrates and some
775	terrestrial insects (Hamilton et al. 2022). For both A and B group species, iNaturalist records
776	(and other distributed, community-scientist platforms such as eButterfly) hold great promise for
777	understanding population trajectories in coming years.
778	We have used iNaturalist records to ask if species have been seen across smaller or larger
779	areas relative to expectation based on the areal extent of expert derived range estimates. We
780	consider that approach to be exploratory and gave it a corresponding low weight in our ranking.
781	Although we used research grade observations from iNaturalist (Hochmair et al. 2020),
782	misidentifications are still possible and (more generally) complexities in taxonomic usage and
783	metadata associated with GBIF (the Global Biodiversity Information Facility from which we
784	accessed the records) produce challenges when merging with other datasets. We have been
785	conservative in our vetting of that process but acknowledge that tool development in this area is
786	needed, and we offer our results in the hope of encouraging other researchers to explore creative
787	uses of iNaturalist and other publicly-sourced records. Despite the potential issues, we note that
788	the variable for change in range size that we derived from the iNaturalist-to-expert comparison
789	was positively correlated ($r = 0.21$) with historical trajectories derived from Shapiro data (Figure
790	S5) but not with 50-year projections based on NABA data ($r = 0.04$).

Phylogenetically, risk values are strongly clustered within and among families, with notable concentrations in the Lycaenidae and Hesperiidae, with the latter in part due to both species with small southern ranges (B group species) and species in monitoring programs with observed declines. Of the high-risk category species (with risk index values above the 90th quantile), 53% are Hesperiidae. The family Nymphalidae has the lowest concentration of at-risk

Forister et al. p. 36

species, although one of the most notably-declining species is in this family. Despite being large
and dispersive and able to use a number of exotic plants as larval hosts, *Vanessa annabella* is
becoming hard to find across locations that include urban centers, high mountains, and southern
deserts (Figure 4).

Although V. annabella is deservedly at the top of the risk list (Figures 3 - 4), we stress the 800 uncertainty in the actual risk values that we have generated and we do not place much weight on 801 the exact position of species on that list. In other words, we believe that the top species in the A 802 group are indeed in historical declines that will likely continue in coming years, but the fact that 803 one species is in the 4th position vs the 10th or even the 25th position on the list is not 804 necessarily important. Small differences in, for example, the projected 50-year probability of 805 population persistence affect the positions for those top species which have mostly similar risk 806 values (and broadly overlapping credible intervals). This is why we conservatively suggest that 807 all of the top 50 species in the A group (Figure 3) deserve closer scrutiny and in some cases 808 likely deserve protection. The fact that rankings should be treated as approximate is also why we 809 have presented other lines of information (geographic range, host specialization, etc.) for the A 810 group, even though the risk index ranking is based solely on the observational data (NABA, 811 Shapiro and iNaturalist) for those species. For example, *Pontia protodice* and *Lycaena* 812 xanthoides have nearly identical risk indices, but the latter (L. xanthoides) is univoltine with a 813 smaller geographic range, greater exposure to development and a more specialized diet (Figure 814 3); these are all factors that could be considered by conservation biologists and ecologists 815 interested in declining insects. With respect to current protections, only two of the species that 816 we have studied have status at the federal level: one of the A group species (the monarch 817 butterfly, *Danaus plexippus*) is currently a candidate for protection under the US Endangered 818

Forister et al. p. 37

819	Species Act (ESA), and one of the B group species, Lycaena hermes, is currently listed as
820	threatened. Note that Boloria acrocnema is treated as a full species under the ESA, but we have
821	followed both NABA (2018) and Pelham (2022) in counting it among the protected subspecies
822	(Boloria improba acrocnema) in Table 1.

Our presentation of the top 50 species in the A group (Figure 3) includes sample sizes 823 (for NABA and Shapiro datasets) which should also be considered when judging the evidence 824 for risk. For example, the 2nd and 3rd species on the A group list (Figure 3) are represented by 825 data from 3 or fewer sites for the NABA and Shapiro datasets, and are not represented in 826 iNaturalist analyses. The small samples for those species are reflected in broad intervals around 827 the risk values, and it can be noted that other species in the top 10 for the A list are known to be 828 in decline based on evidence from two to three times as many sites (e.g., *Pholisora catullus*, 829 Atalopedes campestris, and Euchloe ausonides). The number of sites for individual species is a 830 reflection not just of information available for analysis, but it should be remembered that risk 831 associated with the NABA data derives from a multi-species population viability analysis, and 832 species with fewer sites are more likely by chance to have lower occupancy in forecasts than 833 species known from a greater number of sites. This is both a methodological feature of stochastic 834 simulations but also reflects a biological reality in that more widespread species are known from 835 a greater number of NABA sites (thus geographic range is indirectly involved in the contribution 836 that the NABA analyses make to our estimate of risk). 837

Yet another important aspect of sample size involves the great many A group species not represented in all three of the observational datasets (Shapiro, NABA and iNaturalist); for these species, we used median interpolation. In other words, when calculating the risk index for a species present in, for example, the Shapiro and iNaturalist datasets but not NABA, we assigned

Forister et al. p. 38

a 50-year projection value based on the median across all other species represented in the NABA
dataset. For the present effort, we consider this to be at least a relatively simple assumption,
although we acknowledge that future analyses could use more sophisticated interpolation
perhaps including information from closely related species. The phylogenetic signal observed
here suggests that genetic relatedness could be a tool for dealing with uncertainty and missing
data in conservation ranking.

The weight of missing data and uncertainty of course becomes greater when we turn to 848 the top 50 species in the B group (Figure 3) for which monitoring data is either absent or 849 insufficient for robust models. Not only is robust observational data lacking, but so many of the 850 B group species are similar in having small ranges in hot and dry parts of the region that the 851 overall spread of risk values is smaller than for the A group. Thus, rankings in the top 50 for the 852 B group should be taken with an even more substantial serving of salt. Indeed, there are certainly 853 species beyond the top 50 that merit careful scrutiny. For example, Strymon avalona is restricted 854 entirely to Catalina Island (less than 200 square kilometers) off the coast of southern California. 855 The partly wild nature of the island gives the species a low development score and the area 856 happens to be characterized by only moderate departure from climatic baseline. Thus S. avalona 857 ranks outside of the top 50 for the B group (Figure S6), even though that small geographic range 858 of course puts it at risk of stochastic loss. Similarly, many of the B group species below the top 859 50 have red lambda symbols (to the right of the panel) which indicate negative annual trends 860 (Figures S6-S8), albeit based on very few NABA sites (which is why we have shown those 861 results but did not use them in the calculation of the B group risk index). In general we hope that 862 the data organized here for the B group species is an inspiration for greater monitoring of these 863

Forister et al. p. 39

taxa with small ranges in regions vulnerable to threats that include ongoing climate change and
the loss of natural disturbance regimes (Haddad 2018).

Even greater uncertainty underlies the prioritization of subspecies for conservation, 866 which we have done using a composite of rankings published elsewhere (NatureServe) and a 867 survey of the last year of a publicly reported observation. Despite the uncertainty and different 868 approaches involved in ranking species and subspecies, it is noteworthy that the subspecific risk 869 values are correlated (r = 0.28) with risk values for the associated B group species. Not 870 surprisingly given their well-known propensity for subspecific differentiation and localized 871 population dynamics, 44% of the top 50 subspecies (Table 1) are in the family Lycaenidae. Our 872 ranking of subspecies also highlights two states with high numbers of at risk subspecies. First, 25 873 of the top 50 taxa have a range that includes California, reflecting the long-standing risks to 874 butterfly populations and endemic subspecies from various types of habitat loss and degradation 875 in that state (Forister et al. 2016). Second, 21 of the top 50 taxa have a range that includes 876 Nevada, a region of high subspecific diversity and endemism for many butterfly species across 877 families. In particular, the extreme subspecific diversity of Euphilotes species in the western US 878 is apparent and should be a target for future investigation with resurveys, conservation genetics 879 and targeted monitoring; more than 50 *Euphilotes* subspecies are listed in Pelham (2022). 880

Finally, we can also note that several subspecies in our top list are either protected federally or currently in review for protection under the US Endangered Species Act, but those are intermixed with many taxa in the top 50, and especially the top 25, not receiving federal conservation attention. In addition, while some state agencies in the west manage proactive conservation efforts that prevent species from needing federal protection (e.g., through a list of Species of Greatest Conservation Need [FWS 2001]), other state wildlife agencies do not have

Forister et al. p. 40

887	regulatory authority over terrestrial invertebrates. Perhaps our most important finding for
888	subspecies is not reflected in Table 1: of the more than 1k taxa that we reviewed, approximately
889	400 are not included in NatureServe assessments. Thus the need for broader evaluation is great,
890	and is also urgent as there are many examples of subspecies that have not been seen in many
891	years; these include Philotiella speciosa bohartorum with no sightings since the 1970s despite
892	extensive searches (Davenport 2007), Plebejus [Icaricia] saepiolus aureolus presumed extinct
893	from development, and Euchloe ausonides andrewsi, threatened by fires and drought and with its
894	last available observation from 1983 (Davenport 2018, SCAN 2022).

895

896 CAVEATS & CONCLUSIONS

Our synthesis of status and trends for a diverse fauna faced many challenges. Chief among these 897 is the fact that even for species that are relatively well represented in monitoring schemes, the 898 information is still clustered around areas of human population density. Thus, broad ranges (e.g., 899 Figure 4) and more narrow ranges (e.g., Figure 6) alike are not particularly well sampled in terms 900 of spread of monitored locations in space. We can hope that coming years will see greater 901 investment in monitoring and participation by the general public, and we hope that our use of 902 iNaturalist data in particular encourages both increasing contributions by the general public and 903 the development of new models that can take advantage of mixed data types (e.g., Strebel et al. 904 2022). Another major data issue that we faced was at the US-Mexico border; although ranges are 905 more recently available for species in Mexico (Glassberg 2018), we have limited our studies for 906 now to north of the border (and south of the Canadian border). We did this partly because of our 907 previous focus on butterflies of the 11 western US states (Forister et al. 2021), but also because 908 one has to bite off a manageable problem which in this case involved stopping at political 909

Forister et al. p. 41

borders. We note, however, that the political border especially in the south created many 910 apparently small ranges for those species just crossing that line. Most seriously, those very small 911 ranges are subject to stochasticity in our assessment of exposure to development because a pixel 912 of development can by chance be included or not in small ranges and thus have an outsized 913 influence (in terms of the fraction of the range exposed to development). Better integration of 914 data across southern and northern US borders is an important area for future work, especially 915 since threats involving development or pesticide use could be different in different countries. In 916 the meantime, it is for these reasons that we have included our qualitative range labels (N, S, E, 917 and W) with our rankings (Figure 3). For the B group species in particular, those labels can be 918 used to focus on western species where the political boundaries are considerably less of an issue. 919

The traditional focus for butterfly conservation in the United States has been at the 920 taxonomic level of subspecies, which is partly a consequence of the fact that population 921 segments cannot be listed for invertebrates (thus leaving subspecies as the next unit below full 922 species that can be protected). We have organized subspecific information and present a list of 923 subspecies that could be profitable targets of conservation attention (Table 1), though most of 924 our effort has been at the level of full species. Thus, we acknowledge that our results fall partly 925 outside of the traditional scope of conservation work for butterflies in the United States. It is, 926 however, entirely likely that compounding population losses across the wild spaces of the region 927 have pushed many full species to the point where range-wide research and conservation attention 928 are warranted. A notable example of this is recent effort focused on conservation of the monarch 929 butterfly, Danaus plexippus (Pelton et al. 2019), which is indeed in our list of the 50 most at-risk 930 species (Figure 3), but a number of species are higher on the list and are equally deserving of 931 attention. It is our chief hope that the work presented here is a framework that will facilitate 932

Forister et al. p. 42

such work in coming decades, acknowledging the many assumptions that have been made alongthe way.

935

936 ACKNOWLEDGEMENTS

- 937 MLF thanks the National Science Foundation (DEB-2114793), and CAH was supported by a
- ⁹³⁸ National Institute of Food and Agriculture fellowship (NEVW-2021-09427). EMG, KLB, and
- JPJ were supported by the Modelscape Consortium with funding from NSF (OIA-2019528).
- ⁹⁴⁰ Thanks to the Plant Insect Group at UNR for much thoughtful feedback, and Lee Dyer in
- particular for various key ideas, as well as Sarina Jepsen who provided important feedback on
- subspecies risk. We thank Texas State University for the use of the LEAP computing cluster,
- ⁹⁴³ and thanks also to authors of our previous analysis of western butterflies (Forister et al. 2021)

without which the current paper would not have gotten off the ground.

945

946 CONFLICT OF INTEREST

⁹⁴⁷ The authors declare no conflict of interest.

948

949 AUTHOR CONTRIBUTIONS

MLF conceived the project and the overall organization of data and the presentation of results.
TVR wrote the Bayesian population models for the NABA data and contributed to the design of
analyses. EMG contributed to numerous components, especially the analysis of spatial data. KJB
organized information on subspecies. CAH, CFC, KLB, and TB contributed to component
analyses and assisted with data collation or organization. JZ, QC, and NVG generated the
phylogeny which was analyzed here by JPJ. JG manages the collection of NABA data and

956	contributed the expert-derived range outlines. AMS has collected the vast majority of the Shapiro
957	transect data (with a few recent years of collection contributed in part by MLF and CAH). All
958	authors contributed to interpretation of results and writing of the manuscript.
959	
960	DATA AVAILABILITY STATEMENT
961	Data and code will be made available through Dryad at the time the manuscript is accepted for
962	publication.
963	
964	REFERENCES
965	Abatzoglou, J. T., S. Z. Dobrowski, and S. A. Parks. 2020. Multivariate climate departures have
966	outpaced univariate changes across global lands. Scientific Reports 10:1-9.
967	Abatzoglou, J. T., S. Z. Dobrowski, S. A. Parks, and K. C. Hegewisch. 2018. TerraClimate, a
968	high-resolution global dataset of monthly climate and climatic water balance from 1958-
969	2015. Scientific Data 5:170191.
970	Altschul, S. F., T. L. Madden, A. A. Schäffer, J. Zhang, Z. Zhang, W. Miller, and D. J. Lipman.
971	1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search
972	programs. Nucleic acids research 25:3389–3402.
973	Benton, T. G. 2003. Understanding the ecology of extinction: are we close to the critical
974	threshold? Annales Zoologici Fennici 40:71-80.
975	Bonelli, S., L. P. Casacci, F. Barbero, C. Cerrato, L. Dapporto, V. Sbordoni, S. Scalercio, A.
976	Zilli, A. Battistoni, and C. Teofili. 2018. The first red list of Italian butterflies. Insect
977	Conservation and Diversity 11:506–521.
978	Brook, B. W., N. S. Sodhi, and C. J. A. Bradshaw. 2008. Synergies among extinction drivers

979	under global change. Trends in Ecology & Evolution 23:453–460.
980	Cardoso, P., P. S. Barton, K. Birkhofer, F. Chichorro, C. Deacon, T. Fartmann, C. S. Fukushima,
981	R. Gaigher, J. C. Habel, and C. A. Hallmann. 2020. Scientists' warning to humanity on
982	insect extinctions. Biological Conservation 242:108426.
983	Cardoso, P., T. L. Erwin, P. A. V Borges, and T. R. New. 2011. The seven impediments in
984	invertebrate conservation and how to overcome them. Biological Conservation 144:2647-
985	2655.
986	Casner, K., M. L. Forister, K. Ram, and A. M. Shapiro. 2014a. The utility of repeated presence-
987	absence data as a surrogate for counts: a case study using butterflies. Ecological
988	Applications 18:13–27.
989	Casner, K. L., M. L. Forister, J. M. O'Brien, J. H. Thorne, D. P. Waetjen, and A. M. Shapiro.
990	2014b. Loss of agricultural land and a changing climate contribute to decline of an urban
991	butterfly fauna. Conservation Biology 28:773–782.
992	Crossley, M. S., A. R. Meier, E. M. Baldwin, L. L. Berry, L. C. Crenshaw, G. L. Hartman, D.
993	Lagos-Kutz, D. H. Nichols, K. Patel, and S. Varriano. 2020. No net insect abundance and
994	diversity declines across US Long Term Ecological Research sites. Nature Ecology &
995	Evolution 4:1368–1367.
996	Crossley, M. S., O. M. Smith, L. L. Berry, R. Phillips-Cosio, J. Glassberg, K. M. Holman, J. G.
997	Holmquest, A. R. Meier, S. A. Varriano, and M. R. McClung. 2021. Recent climate change
998	is creating hotspots of butterfly increase and decline across North America. Global Change
999	Biology 27:2702–2714.
1000	Davenport, K. 2007. Yosemite Butterflies. The Taxonomic Report 5.
1001	Davenport, K. 2018. Butterflies of southern California in 2018: updating Emmel and Emmel's

- 1002 1973 Butterflies of southern California. Lepidoptera of North America 15. Contributions of
- the CP Gillette Museum of Arthropod Diversity, Colorado State University. Fort Collins,
- 1004 CO.
- Diniz-Filho, J. A. F., P. De Marco Jr, and B. A. Hawkins. 2010. Defying the curse of ignorance:
- perspectives in insect macroecology and conservation biogeography. Insect Conservationand Diversity 3:172–179.
- Dirzo, R., H. S. Young, M. Galetti, G. Ceballos, N. J. B. Isaac, and B. Collen. 2014. Defaunation
 in the Anthropocene. Science 345:401–406.
- Edge, D. A., and S. Mecenero. 2015. Butterfly conservation in southern Africa. Journal of Insect
 Conservation 19:325–339.
- Eisenhauer, N., A. Bonn, and C. A Guerra. 2019. Recognizing the quiet extinction of
 invertebrates. Nature Communications 10:1–3.
- 1014 Eskildsen, A., L. G. Carvalheiro, W. D. Kissling, J. C. Biesmeijer, O. Schweiger, and T. T.
- 1015 Høye. 2015. Ecological specialization matters: long-term trends in butterfly species richness
- and assemblage composition depend on multiple functional traits. Diversity and
- 1017 Distributions 21:792–802.
- 1018 Evans, J. S., and K. Ram. 2021. Package 'spatialEco.' R CRAN Project.
- ¹⁰¹⁹ Faber-Langendoen, D., J. Nichols, L. Master, K. Snow, A. Tomaino, R. Bittman, G. Hammerson,
- B. Heidel, L. Ramsay, and A. Teucher. 2012. NatureServe conservation status assessments:
- 1021 methodology for assigning ranks, https://www.natureserve.org/sites/ default/files/
- natureserveconservationstatusmethodology_jun12.pdf. NatureServe, Arlington, VA.
- ¹⁰²³ Farber, O., and R. Kadmon. 2003. Assessment of alternative approaches for bioclimatic
- ¹⁰²⁴ modeling with special emphasis on the Mahalanobis distance. Ecological Modelling

1025	160:115-130.
1025	100.113-130

- ¹⁰²⁶ Forister, M. L., B. Cousens, J. G. Harrison, K. Anderson, J. H. Thorne, D. Waetjen, C. C. Nice,
- M. De Parsia, M. L. Hladik, and R. Meese. 2016. Increasing neonicotinoid use and the
 declining butterfly fauna of lowland California. Biology Letters 12:20160475.
- ¹⁰²⁹ Forister, M. L., J. A. Fordyce, C. C. Nice, J. H. Thorne, D. P. Waetjen, and A. M. Shapiro. 2018.
- Impacts of a millennium drought on butterfly faunal dynamics. Climate Change Responses5:3.
- 1032 Forister, M. L., C. A. Halsch, C. C. Nice, J. A. Fordyce, T. E. Dilts, J. C. Oliver, K. L. Prudic, A.
- M. Shapiro, J. K. Wilson, and J. Glassberg. 2021. Fewer butterflies seen by community
- scientists across the warming and drying landscapes of the American West. Science371:1042–1045.
- ¹⁰³⁶ Forister, M. L., A. C. McCall, N. J. Sanders, J. A. Fordyce, J. H. Thorne, J. O'Brien, D. P.
- ¹⁰³⁷ Waetjen, and A. M. Shapiro. 2010. Compounded effects of climate change and habitat
- alteration shift patterns of butterfly diversity. Proceedings of the National Academy of
- Sciences of the United States of America 107:2088–2092.
- Forister, M. L., and A. M. Shapiro. 2003. Climatic trends and advancing spring flight of
 butterflies in lowland California. Global Change Biology 9:1130–1135.
- Fox, R. 2013. The decline of moths in Great Britain: a review of possible causes. Insect
 Conservation and Diversity 6:5–19.
- Fox, R., M. S. Warren, T. M. Brereton, D. B. Roy, and A. Robinson. 2011. A new Red List of
 British butterflies. Insect Conservation and Diversity 4:159–172.
- 1046 FWS. 2001. Fish and Wildlife Service, Department of the Interior. 2001. 50 Million FY 2001
- 1047 Wildlife Conservation and Restoration Account, 50 Million FY 2001 State Wildlife Grants

- Program. Federal Register 66:7657–7660.
- 1049 GBIF.org. 2021a. GBIF Occurrence Download (Papilionidae)
- 1050 https://doi.org/10.15468/dl.m652hd.
- 1051 GBIF.org. 2021b. GBIF Occurrence Download (Hesperiidae) https://doi.org/10.15468/dl.dcthdd.
- 1052 GBIF.org. 2021c. GBIF Occurrence Download (Nymphalidae)
- 1053 https://doi.org/10.15468/dl.qzgtcr.
- 1054 GBIF.org. 2021d. GBIF Occurrence Download (Lycaenidae) https://doi.org/10.15468/dl.2j2ay4.
- 1055 GBIF.org. 2021e. GBIF Occurrence Download (Pieridae) https://doi.org/10.15468/dl.vquqtz.
- 1056 GBIF.org. 2021f. GBIF Occurrence Download (Riodinidae) https://doi.org/10.15468/dl.bbtkq2.
- 1057 Geyle, H. M., M. F. Braby, M. Andren, E. P. Beaver, P. Bell, C. Byrne, M. Castles, F. Douglas,
- R. V Glatz, and B. Haywood. 2021. Butterflies on the brink: identifying the Australian
 butterflies (Lepidoptera) most at risk of extinction. Austral Entomology 60:98–110.
- 1060 Glassberg, J. 2017. A Swift guide to butterflies of North America. Princeton University Press.
- 1061 Glassberg, J. 2018. A Swift guide to butterflies of Mexico and Central America. Princeton
- 1062 University Press.
- Gonzalez, P., F. Wang, M. Notaro, D. J. Vimont, and J. W. Williams. 2018. Disproportionate
 magnitude of climate change in United States national parks. Environmental Research
 Letters 13:104001.
- 1066 Goulson, D. 2019. The insect apocalypse, and why it matters. Current Biology 29:R967–R971.
- Haddad, N. M. 2018. Resurrection and resilience of the rarest butterflies. PLoS Biology
 16:e2003488.
- Hallmann, C. A., M. Sorg, E. Jongejans, H. Siepel, N. Hofland, H. Schwan, W. Stenmans, A.
- Müller, H. Sumser, T. Hörren, and others. 2017. More than 75 percent decline over 27 years

1071	in total flying insect biomass in protected areas. PIOS ONE 12:e0185809.
1072	Halsch, C. A., A. M. Shapiro, J. A. Fordyce, C. C. Nice, J. H. Thorne, D. P. Waetjen, and M. L.
1073	Forister. 2021. Insects and recent climate change. Proceedings of the National Academy of
1074	Sciences:10.1073/pnas.2002543117.
1075	Hamilton, H., R. L. Smyth, B. E. Young, T. G. Howard, C. Tracey, S. Breyer, D. R. Cameron, A.
1076	Chazal, A. K. Conley, and C. Frye. 2022. Increasing taxonomic diversity and spatial
1077	resolution clarifies opportunities for protecting imperiled species in the US. Ecological
1078	Applications:e2534.
1079	Hawkins, B. A. 2010. Multiregional comparison of the ecological and phylogenetic structure of
1080	butterfly species richness gradients. Journal of Biogeography 37:647-656.
1081	Hijmans, R. J., J. van Etten, M. Mattiuzzi, M. Sumner, J. A. Greenberg, O. P. Lamingueiro, A.
1082	Bevan, E. B. Racine, and A. Shortridge. 2021. raster: Geographic Data Analysis and
1083	Modeling, Version 2.9-23, R package.
1084	Hochmair, H. H., R. H. Scheffrahn, M. Basille, and M. Boone. 2020. Evaluating the data quality
1085	of iNaturalist termite records. PLOS ONE 15:e0226534.
1086	Hughes, J. B., G. C. Daily, and P. R. Ehrlich. 2000. Conservation of insect diversity: a habitat
1087	approach. Conservation Biology 14:1788–1797.
1088	Jamwal, P. S., M. Di Febbraro, M. L. Carranza, M. Savage, and A. Loy. 2021. Global change on
1089	the roof of the world: Vulnerability of Himalayan otter species to land use and climate
1090	alterations. Diversity and Distributions.
1091	Janzen, D. H., and W. Hallwachs. 2019. Perspective: Where might be many tropical insects?
1092	Biological Conservation 233:102–108.
1093	Katoh, K., K. Misawa, K. Kuma, and T. Miyata. 2002. MAFFT: a novel method for rapid

Forister et al. p. 49

1094	multiple sequence alignment based on fast Fourier transform. Nucleic Acids Research
1095	30:3059–3066.

- Kellner, K. 2017. R Package 'jagsUI': a wrapper around 'rjags' to streamline 'JAGS' Analyses, 1096 v.1.4.9. 1097
- Kirchhoff, C., C. T. Callaghan, D. A. Keith, D. Indiarto, G. Taseski, M. K. J. Ooi, T. D. Le 1098
- Breton, T. Mesaglio, R. T. Kingsford, and W. K. Cornwell. 2021. Rapidly mapping fire 1099
- effects on biodiversity at a large-scale using citizen science. Science of The Total 1100 Environment 755:142348. 1101
- van Klink, R., D. E. Bowler, K. B. Gongalsky, A. B. Swengel, A. Gentile, and J. M. Chase. 1102
- 2020. Meta-analysis reveals declines in terrestrial but increases in freshwater insect 1103 abundances. Science 368:417-420. 1104
- Li, L., C. J. Stoeckert, and D. S. Roos. 2003. OrthoMCL: identification of ortholog groups for 1105 eukaryotic genomes. Genome Research 13:2178–2189. 1106
- Maes, D., W. Vanreusel, I. Jacobs, K. Berwaerts, and H. Van Dyck. 2012. Applying IUCN Red 1107
- List criteria at a small regional level: a test case with butterflies in Flanders (north 1108 Belgium). Biological Conservation 145:258–266. 1109
- McLaughlin, J. F., J. J. Hellmann, C. L. Boggs, and P. R. Ehrlich. 2002. Climate change hastens
- population extinctions. Proceedings of the National Academy of Sciences of the United 1111 States of America 99:6070-6074. 1112
- NABA. 2018. Checklist of North American Butterflies Occurring North of Mexico Edition 2.4. 1113
- Naeem, S., R. Chazdon, J. E. Duffy, C. Prager, and B. Worm. 2016. Biodiversity and human 1114
- well-being: an essential link for sustainable development. Proceedings of the Royal Society 1115
- B: Biological Sciences 283:20162091. 1116

1110

- Nakamura, Y. 2011. Conservation of butterflies in Japan: status, actions and strategy. Journal of
 Insect Conservation 15:5–22.
- New, T. R., R. M. Pyle, J. A. Thomas, C. D. Thomas, and P. C. Hammond. 1995. Butterfly
- 1120 Conservation Management. Annual Review of Entomology 40:57–83.
- 1121 Nguyen, L.-T., H. A. Schmidt, A. Von Haeseler, and B. Q. Minh. 2015. IQ-TREE: a fast and
- effective stochastic algorithm for estimating maximum-likelihood phylogenies. MolecularBiology and Evolution 32:268–274.
- Nice, C. C., M. L. Forister, Z. Gompert, J. A. Fordyce, and A. M. Shapiro. 2014. A hierarchical
- perspective on the diversity of butterfly species' responses to weather in the Sierra Nevada
- 1126 Mountains. Ecology 95:2155–2168.
- ¹¹²⁷ Opler, P. A. 1999. A field guide to western butterflies. Houghton Mifflin Harcourt.
- Pelham, J. P. 2022. A catalogue of the butterflies of the United States and Canada.
- Pelton, E. M., C. B. Schultz, S. J. Jepsen, S. H. Black, and E. E. Crone. 2019. Western monarch
- population plummets: status, probable causes, and recommended conservation actions.
- 1131 Frontiers in Ecology and Evolution 7:258.
- Pilliod, D. S., M. I. Jeffries, R. S. Arkle, and D. H. Olson. 2020. Reptiles under the conservation
 umbrella of the greater sage-grouse. The Journal of Wildlife Management 84:478–491.
- Prudic, K. L., J. C. Oliver, B. V Brown, and E. C. Long. 2018. Comparisons of citizen science
- data-gathering approaches to evaluate urban butterfly diversity. Insects 9:186.
- 1136 RCoreTeam. 2020. R: A language and environment for statistical computing R Foundation for
- 1137 Statistical Computing, Vienna, Austria. URL https://www.R-project.org/.
- Revell, L. J. 2012. phytools: an R package for phylogenetic comparative biology (and other
- things). Methods in Ecology and Evolution:217–223.

	Forister
1140	Riecke, T. V, D. Gibson, M. Kéry, and M. Schaub. 2021. Sharing detection heterogeneity
1141	information among species in community models of occupancy and abundance can
1142	strengthen inference. Ecology and evolution 11:18125–18135.
1143	Salcido, D. M., M. L. Forister, H. G. Lopez, and L. A. Dyer. 2020. Ecosystem services at risk
1144	from declining taxonomic and interaction diversity in a tropical forest. Scientific Reports
1145	10:1–10.
1146	Samways, M. J. 2007. Insect conservation: a synthetic management approach. Annu. Rev.
1147	Entomol. 52:465–487.
1148	SCAN. 2022. Symbiota Collections of Arthropods Network https://scan-bugs.org/portal/.
1149	Schultz, C. B., N. M. Haddad, E. H. Henry, and E. E. Crone. 2019. Movement and demography
1150	of at-risk butterflies: building blocks for conservation. Annual Review of Entomology
1151	64:167–184.
1152	Scott, J. A. 1986. The butterflies of North America. Stanford University Press, Stanford,
1153	California.
1154	Sekar, S. 2012. A meta-analysis of the traits affecting dispersal ability in butterflies: can
1155	wingspan be used as a proxy? Journal of Animal Ecology 81:174–184.
1156	Sen, P. K. 1968. Estimates of the regression coefficient based on Kendall's tau. Journal of the
1157	American Statistical Association 63:1379–1389.
1158	Shapiro, A. M. 1996. Status of butterflies. Pages 743–757 Sierra Nevada Ecosystem Project:

- 1159 Final Report to Congress Vol. II. Center for Water and Wildland Resources, University of
- 1160 California, Davis, Davis, California.
- 1161 Shapiro, A. M. 2022. Art Shapiro's Butterfly Site https://butterfly.ucdavis.edu/.
- Staude, I. R., L. M. Navarro, and H. M. Pereira. 2020. Range size predicts the risk of local

1163	extinction from habitat loss. Global Ecology and Biogeography 29:16-25.
1164	Strebel, N., M. Kéry, J. Guélat, and T. Sattler. 2022. Spatiotemporal modelling of abundance
1165	from multiple data sources in an integrated spatial distribution model. Journal of
1166	Biogeography 49:563–575.
1167	van Swaay, C., D. Maes, S. Collins, M. L. Munguira, M. Šašić, J. Settele, R. Verovnik, M.
1168	Warren, M. Wiemers, and I. Wynhoff. 2011. Applying IUCN criteria to invertebrates: How
1169	red is the Red List of European butterflies? Biological Conservation 144:470-478.
1170	Theil, H. 1950. A rank-invariant method of linear and polynomial regression analysis.
1171	Indagationes Mathematicae 12:173.
1172	Turvey, S. T., and J. J. Crees. 2019. Extinction in the Anthropocene. Current Biology 29:R982-
1173	R986.
1174	USDA. 2020. National Agricultural Statistics Service Cropland Data Layer. Published crop-
1175	specific data layer. https://nassgeodata.gmu.edu/CropScape/.
1176	Wagner, D. L. 2019. Insect declines in the Anthropocene. Annual Review of Entomology 65.
1177	Wepprich, T., J. R. Adrion, L. Ries, J. Wiedmann, and N. M. Haddad. 2019. Butterfly abundance
1178	declines over 20 years of systematic monitoring in Ohio, USA. PLOS ONE 14:e0216270.
1179	Wilson, E. O. 1987. The little things that run the world (the importance and conservation of
1180	invertebrates). Conservation Biology 1:344-346.
1181	Wilson, J. S., A. D. Pan, D. E. M. General, and J. B. Koch. 2020. More eyes on the prize: an
1182	observation of a very rare, threatened species of Philippine Bumble bee, Bombus
1183	irisanensis, on iNaturalist and the importance of citizen science in conservation biology.
1184	Journal of Insect Conservation 24:727–729.
1185	Yu, G., D. K. Smith, H. Zhu, Y. Guan, and T. T. Lam. 2017. ggtree: an R package for

Forister et al. p. 53

- visualization and annotation of phylogenetic trees with their covariates and other associated
- data. Methods in Ecology and Evolution 8:28–36.
- ¹¹⁸⁸ Zhang, J., Q. Cong, J. Shen, P. A. Opler, and N. V Grishin. 2019. Genomics of a complete
- butterfly continent. BioRxiv:829887.

1190

1191

TABLE 1. Top 50 subspecies ranked by quantitative risk index, including taxonomic name and family, states in which subspecies are found, as well as the species group (A or B) to which the nominal species belongs. The taxonomy used here is based on Pelham (2022) because of the emphasis on subspecific distinctions; where the generic or specific names differ from NABA (2018), the alternative name is in brackets (exceptions to that are the *Euphilotes* taxa, for which taxonomy and differences in usage are complex; in those cases we have not listed synonymies between Pelham and NABA); common names are also given here as they are more stable for some subspecies than trinomials. Species marked with an asterisk (*) are protected by the US Endangered Species Act; ** = currently under review for protection.

D' 1		<u></u>	<u>г 1</u>	C
KISK	Subspecies	States	Family	Group
0.837	Argumia [Sugaria] adiasta adiasta ungiluared fritillary	CA	Lycaenidae	B
0.730	<i>Exchlor humatic and youri</i> Androwic morble	CA	Diaridaa	
0.725	<i>Eucnice nyanus anarewsi</i> , Andrews marches See Cabriel Marc groupich blue	CA	Fielidae	A
0.720	<i>Icaricia</i> [<i>Piebejus</i>] <i>saepiolus aureolus</i> , San Gabriel Miths greenish blue	CA	Lycaenidae	A
0./12	Euphilotes pallescens mattonii, Mattoni's blue	NV	Lycaenidae	В
0.692	Cercyonis oetus alkalorum, small wood-nymph	NV	Nymphalidae	A
0.677	Argynnis [Speyeria] nokomis nokomis, Great Basin nokomis fritillary **	AZ, CO, NM, UT	Nymphalidae	В
0.677	Euphilotes pallescens arenamontana, San Mountain blue	NV	Lycaenidae	В
0.677	Euphilotes pallescens calneva, Honey Lake blue	CA, NV	Lycaenidae	В
0.677	Hesperia miriamae longaevicola, White Mountains skipper	CA, NV	Hesperiidae	В
0.677	Satyrium polingi organensis, Organ Mountains Poling's hairstreak	NM	Lycaenidae	В
0.677	Euphilotes pallescens ricei, Rice's blue	NV	Lycaenidae	В
0.677	Philotiella speciosa septentrionalis, Great Basin small blue	NV	Lycaenidae	В
0.664	Euphilotes enoptes primavera, dotted blue	NV	Lycaenidae	А
0.654	Megathymus ursus deserti, desert yucca borer	AZ	Hesperiidae	В
0.653	Argynnis [Speyeria] nokomis carsonensis, Carson Valley nokomis fritillary	CA, NV	Nymphalidae	В
0.650	Colias skinneri [pelidne] hinchliffi, Skinner's pelidne sulphur	OR	Pieridae	В
0.629	Callophrys mossii bayensis, San Bruno elfin *	CA	Lycaenidae	А
0.629	Callophrys mossii marinensis, Moss' elfin *	CA	Lycaenidae	А
0.629	Euphilotes bernardino minuta, Baking Powder Flat blue	NV	Lycaenidae	А
0.629	Hesperia leonardus montana, Leonard's skipper *	СО	Hesperiidae	В
0.629	Euphilotes mojave virginensis, Virgin Mountains Mojave blue	AZ, NV, UT	Lycaenidae	А
0.626	Hesperia uncas fulvapalla, Uncas skipper	NV	Hesperiidae	В
0.609	Argynnis [Speveria] egleis volaboli, Great Basin fritillary	CA	Nymphalidae	А
0.605	Pseudocopaeodes eunus alinea. Eunus skipper	CA. NV	Hesperiidae	В
0.605	Hesperia uncas giulianii, Railroad Valley skipper	ĊA	Hesperiidae	В
0.605	Hesperia uncas grandiosa. Big Smoky Valley skipper	NV	Hesperiidae	B
0.605	Callonhrys loki [grvneus] thornei. Thorne's hairstreak	CA	Lycaenidae	Ā
0.600	Cercyonis pegala carsonensis. Carson Valley wood nymph	CA NV	Nymphalidae	A
0.591	<i>Funkilotes battoides fusimaculata</i> square dotted blue	NV	Lycaenidae	Δ
0.591	Roloria improba acrocnema Uncompanyre fritillary *	CO	Nymphalidae	B
0.581	Phyciodes cocyta [selenis] arenacolor. Stentoe Valley crescent	NV	Nymphalidae	Δ
0.501	Funkilotes enontes aridorum dotted blue	CANV	Lycaenidae	Δ
0.581	Arounnis [Spavaria] zarana bahransii Behren's silverspot *		Nymphalidae	Δ
0.581	Argynnis [Speyeria] callinna callinna Callinna silverspot *		Nymphalidae	Λ
0.581	Leavicia [Plaboius] shasta charlestonensis Mt Charleston blue *		Lycoonidoo	л л
0.581	<i>Funhydryas anicia</i> [chalcadona] clouderoffi. Sacramento Mtrs checkersnot	NM	Nymphalidae	A
0.581	Euphyaryas anteia [chaiceaona] ciouacroju, Sacramento Mitis eneckerspor	AZ NIV LIT	Lyoponidoo	A
0.501	Laguicia [Dloboius] isquisidas fordari. Fondario hluo *	AL, NV, UI	Lycaenidae	A
0.381	<i>Icaricia</i> [<i>Fiebejus</i>] <i>icarioides Jenderi</i> , Fender's blue	CA OR WA	Nycaenidae	A
0.581	Argynnis [Speyeria] zerene nippolyla, Oregon silverspol	CA, OK, WA	Nymphalidae	A
0.581	Eucnice ausonides insulanus, island marble *	WA	Pleridae	A
0.581	Pyrgus ruraiis lagunae, Laguna Mountains skipper *	CA	Hesperiidae	A
0.581	Apodemia mormo langei, Lange's metalmark *	CA	Riodinidae	A
0.581	<i>Icaricia</i> [<i>Plebejus</i>] <i>icarioides missionensis</i> , Mission blue *	CA	Lycaenidae	A
0.581	Ceryconis oetus pallescens, small wood-nymph	NV	Nymphalidae	A
0.581	Glaucopsyche lygdamus palosverdesensis, Palos Verde blue *	CA	Lycaenidae	A
0.581	Polites sabuleti sinemaculata, bleached sandhill skipper	NV	Hesperiidae	Α
0.581	Argynnis [Speyeria] zerene sonomensis, Zerene fritillary	CA	Nymphalidae	Α
0.581	Euphydryas editha taylori, Taylor's checkerspot *	OR, WA	Nymphalidae	А
0.568	Glauconsyche piasus gabrieling. San Gabriel Mtns arrowhead blue	CA	Lvcaenidae	А

Forister et al. p. 55

1193 Figure legends

1194

FIGURE 1 Schematic overview of main inputs, processes and products associated with the 1195 generation of risk index values for species (subspecies are treated separately). As noted in the 1196 key, data sources are in brown, analyses (and other calculations) are in blue boxes, variables 1197 (used in the creation of the risk index) are in red, and the primary products are in green. The 1198 central branching path illustrates the division of species into the A and B groups, with 1199 observational data contributing to the A group risk assessment on the left, and other data types 1200 contributing to B group assessment on the right. The 10 variables (in red) are identical to the 1201 columns in Figure 3, although labelled slightly different here, especially for the observational 1202 variables: "B year" is the year coefficient from analyses of Shapiro data summarizing change 1203 through time; "P(persistence)" is the probability of population persistence from 50-year 1204 forecasts, and Δ range is an index of change in geographic range based on the relationship 1205 between the last 15 years of iNaturalist observations while controlling for the size of the expert-1206 derived range. Variables on the right ("range area", "precipitation," etc.) are more self-1207 explanatory. Also note that the expert-derived geographic ranges contribute to the risk index 1208 calculations both directly ("range area" and "voltinism") and indirectly as indicated with 1209 connecting arrows. Finally, the "Risk analysis" process box (towards the lower left) illustrates 1210 the analysis of A group risk that was used to partly inform the weighting scheme for the B group 1211 species. 1212

1213

FIGURE 2 Summary of differences between species in the A and B groups. The 184 A group species are those with observational data from either the Shapiro monitoring program or the

Forister et al. p. 56

1216	NABA annual counts; the 212 B group species are not included in those data sources (at least not
1217	with sufficient abundance to be used in our primary models). Comparisons in panels (a) through
1218	(g) are shown as violin plots with kernel density estimates and horizontal lines marking medians
1219	inside rectangles spanning interquartile ranges; vertical lines are upper and lower fences
1220	computed as the third quartile plus one and a half times the interquartile range, and the first
1221	quartile minus one and a half times the interquartile range, respectively. Colors in panels (a)
1222	through (f) match those used in Figure 3 for the same variables. Area-weighted latitudinal
1223	midpoints are shown in panel (g), and the mosaic plot in (h) shows the biogeographical
1224	breakdown of qualitative range positions for A and B group species (e.g., species with ranges in
1225	the South category have a majority of their range south of the US-Mexico border, with only a
1226	small presence north of the border in the western US).

1227

FIGURE 3 The top 50 species with the highest risk rankings in the A group (on the left) and the 1228 B group (on the right). The two panels have some features in common, and some unique 1229 elements. In common they both show the extent to which different variables are associated with 1230 higher or lower risk for each species: a large circle under NABA occupancy, for example, marks 1231 a species that we infer as being at risk because of low forecast occupancy (probability of 1232 population persistence) across currently-extant locations; similarly, a large circle under 1233 development indicates a species at risk because of high exposure to developed lands, and a large 1234 circle under geographic range indicates corresponding risk associated with a relatively small 1235 range. The sizes of the circles were assigned separately within the two lists, A and B group 1236 species, and thus indicate relative differences within those lists. Although all variables are shown 1237 for comparison, the overall risk ranking for the A group species is based solely on the first three 1238

Forister et al. p. 57

1239	variables (NABA occupancy, Shapiro monitoring, and iNaturalist, to the left of the vertical gray
1240	line), while the ranking for the B group species is based entirely on the other seven variables (see
1241	main text for details, and Figure 1). Both panels also have in common the quantitative risk values
1242	shown to the right (e.g., the risk index for Vanessa annabella in panel A is 0.675); note that the
1243	risk values for the A group species include 85% credible intervals (in parentheses),
1244	encompassing uncertainty derived from Bayesian analyses of both NABA and Shapiro data. The
1245	capital letters (N, S, E and W) running down the left side of each panel are qualitative
1246	biogeographical descriptions (see main text for details), and the asterisks next to species names
1247	flag taxonomic issues (see Table S3). A unique element of the panel on the left is the sample size
1248	in parentheses, e.g. "(14,10)" for Vanessa annabella, which is the number of locations from
1249	which data were included from the NABA and Shapiro datasets, respectively. Finally, on the far
1250	right of panel (b), the lambda symbols represent the results of individual time series models run
1251	for the species present in the NABA program but without enough sites and years to be included
1252	in the main model (and thus not a part of the A list); a blue symbol indicates a species with an
1253	80% or greater probability of increasing in recent years, while a red symbol indicates an 80%
1254	chance of decreasing, and black is neither increasing nor decreasing. The other species (beyond
1255	the top 50 highest ranked shown here) are included in Figures S6, S7, and S8.

1256

FIGURE 4 Overview of site-specific trends through time for *Vanessa annabella* at Shapiro sites (on the left) and NABA sites (on the right and along the bottom). Plots for Shapiro sites are shown with decreasing elevation (cooler colors are montane sites) and colored to match the elevational profile of Northern California shown below the map of the western US. The y-axes for Shapiro plots are the fraction of days a species was seen at a site in a year (Shapiro data were

Forister et al. p. 58

truncated at 1984 for analyses, but earlier years are shown here and in Figures 5 and 6). Plots for 1262 NABA sites are shown with decreasing latitude (starting with the most northern sites), with 1263 symbols matching the locations shown in the central map. Values shown in NABA plots have 1264 been adjusted for variation in sampling effort, and values plotted are total counts of individuals 1265 on a natural log scale. Finally, the light gray triangles on the central map are locations of 1266 iNaturalist records within the last 15 years that were used to estimate the difference between 1267 expert-derived geographic range and community scientist-derived area of occupancy (based on 1268 the iNaturalist records). Adult and caterpillar images by Camryn Maher, copyright 2022. 1269 1270

FIGURE 5 Overview of site-specific trends through time for *Euchloe ausonides* at Shapiro sites 1271 (on the left) and NABA sites (on the right). Plots for Shapiro sites are shown with decreasing 1272 elevation (cooler colors are montane sites) and colored to match the elevational profile of 1273 Northern California shown below the map of the western US. The y-axes for Shapiro plots are 1274 the fraction of days a species was seen at a site in a year. Plots for NABA sites are shown with 1275 decreasing latitude (starting with the most northern sites), with symbols matching the locations 1276 shown in the central map. Values shown in NABA plots have been adjusted for variation in 1277 sampling effort, and values plotted are total counts of individuals on a natural log scale. Finally, 1278 the light gray triangles on the central map are locations of iNaturalist records within the last 15 1279 years that were used to estimate the difference between expert-derived geographic range and 1280 community scientist-derived area of occupancy (based on the iNaturalist records). Adult and 1281 caterpillar images by Camryn Maher, copyright 2022. 1282

1283

Forister et al. p. 59

FIGURE 6 Overview of site-specific trends through time for *Poanes melane* at Shapiro sites 1284 (on the left) and NABA sites (on the right). Plots for Shapiro sites are shown with decreasing 1285 elevation (cooler colors are montane sites) and colored to match the elevational profile of 1286 Northern California shown below the map of the western US. The y-axes for Shapiro plots are 1287 the fraction of days a species was seen at a site in a year. Plots for NABA sites are shown with 1288 decreasing latitude (starting with the most northern sites), with symbols matching the locations 1289 shown in the central map. Values shown in NABA plots have been adjusted for variation in 1290 sampling effort, and values plotted are total counts of individuals on a natural log scale. Finally, 1291 the light gray triangles on the central map are locations of iNaturalist records within the last 15 1292 years that were used to estimate the difference between expert-derived geographic range and 1293 community scientist-derived area of occupancy (based on the iNaturalist records). Adult and 1294 caterpillar images by Camryn Maher, copyright 2022. 1295

1296

FIGURE 7 The geography of risk for species with values in the upper 75th quantile of risk
indices as shown in Figure 3 (i.e., combining "medium" and "high" risk categories treated
separately in Figure 8). Panels (a) and (b) show average risk values among those high risk
species, separately for the A and B group species, while panels (c) and (d) show species richness
again for the A group and B group species.

1302

FIGURE 8 The phylogenetic distribution of risk, here shown as three categories: high risk
(upper 90th quantile), medium risk (75th to 90th quantiles), and low risk (below the 75th
quantile). Species names in black are the A group species, other are B group. Butterfly images as
follows: (A) *Apodemia mormo* (Riodinidae); (B) *Euphilotes pallescens arenamontana*

- 1307 (Lycaenidae); (C) Euchloe ausonides (Pieridae); (D) Polites sabuleti (Hesperiidae); (E) Adelpha
- 1308 bredowii (Nymphalidae); (F) Papilio rutulus (Papilionidae). Photo credits go to CAH (panels A,
- 1309 C, E, and F); MLF (panels B and D). Bootstrap support is not shown but the vast majority of
- nodes have support above 0.95; see Zhang et al. (2019) for additional details.



Figure 1



Figure 2

(a) ^A group Sp. 1–50	NABA occupancy	snapiro monitoring	iNaturalist	Geographic range	Development	Climate departure	Precipitation	Voltinism	wingspan	Host range		(b) ^B group Sp. 1–50		Geographic range	Development	Climate departure Precipitation	Voltinism	Wingspan Host range	
Vanessa annabella (14,10) W –	•			•	•	•	•	-	•	•	- 0.675 (0.54, 0.81)	Lycaena hermes*	W -	Ð	•	•••	•		0.753
Thorybes mexicanus* (3,2) W-		•		•	•	•					- 0.666 (0.41, 0.86)	Nastra julia	s -	•	•	• •	•	••	- 0.735
Amblyscirtes vialis (3,3) E-	•	•	-	•	•	•					- 0.634 (0.35, 0.82)	Amblyscirtes elissa*	s -	•	•	• •	•		- 0.734 <mark>λ</mark>
Euchloe ausonides (8,8) W-	•			•	•	•	•		•	•	- 0.63 (0.43, 0.81)	Copaeodes minimus*	s -		•	••	•		- 0.728
Pholisora catullus (10,5) E-	•	D		•	•	•	•	• • •			- 0.628 (0.46, 0.81)	Panoquina errans	s -	•	•	• •	•	•••	- 0.694
Atalopedes campestris (6,7) S-	•	•	•	•	0-	•	•	•	•		- 0.611 (0.44, 0.77)	Piruna cingo*	s -	•	•	• •	•	•	- 0.693λ
Lycaena arota* (5,6) W-	•	•		•	•	•	•				- 0.609 (0.43, 0.79)	Emesis ares*	s-	•			•	•••	- 0.691λ
Papilio indra (4,2) W-	•	•		•	•	•	•		•	•	- 0.598 (0.35, 0.82)	Satyrium ilavia	w -		•	• •	•	•••	- 0.686
Habrodais grunus (3,2) W-		•	•	•	•	•					- 0.588 (0.31, 0.8)	Eurema lisa*	Е-	•	•		•	•••	- 0.686
Paratrytone snowi (3,0) W-		_	-		•	•	•				- 0.586 (0.35, 0.77)	Amblyscirtes nereus	s -	•				-	- 0.684λ
Polites sabuleti (6,8) W-			-	•	•	•	•	-			- 0.581 (0.38, 0.79)	Agathymus evansi	s -	•	•		•	•••	- 0.68
Anthocharis sara (10,6) W-		-		•	:	•	-			•	- 0.578 (0.42, 0.77)	Eurema nise*	s -		-		-		- 0.673
Pontia protodice (20,10) E-		2	•	-	-	•	•	_		-	- 0.565 (0.43, 0.71)	Amblyscirtes fimbriata	s -						- 0.672λ
Lycaena xanthoides* (7,5) E-				•	•	•					- 0.558 (0.36, 0.75)	Strymon istapa	s -		-				- 0.672
Glaucopsyche plasus (4,1) W-	-			•							- 0.546 (0.23, 0.78)	Agathymus polingi	s-						- 0.668
Lycaeides meiissa" (12,2) w-				•		•					- 0.542 (0.33, 0.73)	Chiomara asychis"	S -						- 0.664
Everes amyntula" (9,4) W-											- 0.537 (0.37, 0.72)	Pyrrnopyge araxes"	5-						- 0.654 A
Lycaena rubidus" (3,1) W-				-							- 0.537 (0.27, 0.77)	Agatnymus mariae	5-						- 0.654
											- 0.536 (0.4, 0.68)	All ylonopsis pillacus	о- с						0.654
											0.532 (0.34, 0.75)	Coualiacius anzonensis	о- с						0.055 1
Bloboius lupipi* (5.2) W											-0.532(0.26, 0.75)	Salynum polingi Europilotoo rito	 						0.052
Hosporia comma (12.5) W											- 0.510 (0.20, 0.74)	Daramaaara alluni	s -						0.05
Apodemia nais (3.0) W	<u> </u>	<u> </u>		ě.							- 0.513 (0.26, 0.75)	Callophrye fotie	w -				-		0.0407
Nymphalis milberti* (6.4)											- 0.512 (0.33, 0.75)	Chlosyne californica	w -				-		- 0.6452
Ministrymon leda (3.0) W-					-	Š.					- 0 509 (0 24 0 74)	Furema dina*	s -	à	-		-		- 0.643
Nathalis iole (8.0) E	ě.		-								- 0.506 (0.35, 0.71)	Eurema daira	s -	ŏ			ă		- 0.643
Hesperia pabaska (4.0) W-	-		_			-	-		-		- 0 504 (0 27 0 74)	Phaeostrymon alcestis*	E -	•			ŏ		- 0.641
l erodea eufala (5,5) S-	- • -	-	•		<u> </u>	ŏ	<u> </u>	_			- 0.502 (0.27, 0.71)	Atrytononsis lunus	- s -			.	ŏ		- 0.638 Å
Vanessa virginiensis (18.10) E-		-	Ó	-	ě.	-	<u> </u>	•	-	•	- 0.499 (0.35, 0.65)	Agathymus arvxna	s -			ŏ-O	ŏ		- 0.636
Danaus plexippus (21,10) E-		Ď	ŏ	•	•	<u> </u>	•	•	•		- 0.497 (0.35, 0.64)	Eurema proterpia*	s-			ĎŎ	ŏ	•••	- 0.636 λ
Satyrium tetra (3,2) W-		ŏ-	•	•	Ó-	•					- 0.495 (0.18, 0.73)	Kricogonia lyside	s-	•		Ŏ.	ŏ		- 0.632 λ
Colias cesonia* (5,0) E-	•			•	•			-	•	ŏ	- 0.491 (0.27, 0.7)	Euphyes bimacula	Е-	•	•	•••	ŏ		- 0.631λ
Cercyonis sthenele (3,3) W-	Õ-	•	•	•	•	•	Õ-()-	•		- 0.49 (0.24, 0.76)	Calephelis wrighti	w -	•	- • -	• •	-	- • -•	- 0.628λ
Euphydryas editha (5,2) W-	•	•	•	•	•	•	•-(•	•	- 0.487 (0.26, 0.74)	Agathymus stephensi	s -		•	••	•		- 0.627
Phyciodes selenis* (5,0) W-	•		_	•	•	•	-(•		- 0.487 (0.27, 0.71)	Phyciodes batesii	N -	•	•	• •	•		- 0.626
Phyciodes campestris* (20,6) W-			-	•	•	•	•	•(- 0.486 (0.36, 0.66)	Atrytonopsis deva	s-	•		••	•	•	- 0.625
Hemiargus isola* (8,0) S-	•		-	•	•	•-(0-	•(- 0.485 (0.3, 0.68)	Atrytone arogos	Е-	•	•	• •	•		- 0.624 <mark>λ</mark>
Atlides halesus (3,8) E-	•	•	•	•	•	•	•	•	•(- 0.483 (0.22, 0.73)	Pyrgus xanthus	w -	•	•	• •	•		- 0.623
Pyrgus albescens* (5,0) S-	•		_	•	•	•	0-	•(- 0.481 (0.27, 0.69)	Eurema boisduvaliana*	s -	•			•	• •	- 0.623λ
Satyrium sylvinus (8,8) W-	•	•	•	•	•	•	•-(- 0.481 (0.29, 0.67)	Callophrys henrici	Е-	•		• •	•	•••	- 0.62
Colias philodice (10,1) E-	•	•	•	•	•	•	•	-	•		- 0.48 (0.25, 0.71)	Microtia elva	s -			••	•	•	- 0.619
Satyrium californica (10,7) w-	•	•	•	•	•	•	•		-	•	- 0.478 (0.3, 0.66)	Gyrocheilus patrobas	s -	•	•	••	•	-••	- 0.619
Phyciodes tharos (3,0) E-	•	_	+	•	•	•	•	•	-		- 0.478 (0.25, 0.75)	Euphilotes spaldingi	w -	•	•	• •	•		- 0.618
Pyrgus scriptura (0,3) W-			-	•	•	•			Η		- 0.476 (0.37, 0.6)	Callophrys mcfarlandi	w -	•	•	• •	•	•••	- 0.618
Pontia beckerii* (10,2) W-	•	•	•	•	•	•	•	•	-		- 0.476 (0.26, 0.67)	Notamblyscirtes simius	w -	•	•	• •	•		- 0.617 λ
Brephidium exile* (10,6) W-	•	•	+	•	•	•-(-	•(•	- 0.476 (0.3, 0.63)	Oarisma edwardsii*	s-	•	-	• •	•	•	- 0.615 <mark>λ</mark>
Callophrys spinetorum (7,0) W-	•	_	•	•	•	•	•	•			- 0.473 (0.28, 0.68)	Amblyscirtes oslari	w -		•	• •		•••	- 0.615λ
Vanessa atalanta (24,9) E−	•		•	•	•	•	•	-	•		- 0.473 (0.33, 0.6)	Agathymus neumoegeni	s -	•	•	•••	•	•••	- 0.612
Eurema mexicana* (4,0) S⊐	•		1	•	• ()—	•(- 0.471 (0.22, 0.68)	Megathymus ursus	s -	•		• •	•		L 0.609

Figure 3



Figure 4



Figure 5



Figure 6



Figure 7



Figure 8